SUPPLEMENTARY MATERIAL

Indigodole E from Strobilanthes cusia exhibits anti-IL-17A effect

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ABSTRACT

One new indazole alkaloid, indigodole E (1), was isolated from a traditional Chinese medicine Qing Dai prepared from the aerial parts of *Strobilanthes cusia*. The structure of **1** was elucidated by NMR, MS, UV, and IR spectra as well as optical rotation. Additionally, compound **1** could obviously inhibit not only IL-17A protein production at concentrations from 1.25 to 2.5 μ g/mL, but also IL-17 gene expression at concentrations from 5.0 to 10.0 μ g/mL without cytotoxicity toward Th17 and Jukat cells, respectively. Overall, indazole analogue **1** could be the anti-IL 17A contributor of Qing Dai in this investigation.

KEYWORDS: Qing Dai; *Strobilanthes cusia*; Acanthaceae; indigodole E; indazole alkaloid; IL-17A inhibition

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Experimental in detail

1.1. Generals

NMR spectra were taken on Bruker Avance III 500 MHz. The chemical shift (δ) values are in ppm (part per million) with CDCl₃ as the solvent, and coupling constants (*J*) are in Hz. Low- and high-resolution ESIMS were measured on a Bruker Daltonics Esquire HCT ultra high capacity trap mass spectrometer, and an Orbitrap mass spectrometer (LTQ Orbitrap XL and Q Exactive Plus, Thermo Fisher Scientific), respectively. The optical rotation was recorded on a JASCO-P-2000 polarimeter (cell length 10 mm). UV and IR spectra were measured on a Perkin Elmer#Lambda265 and a Shimadzu model IR Prestige-21 Fourier- transform infrared spectrophotometer, respectively. TLC was performed on Kieselgel 60 F₂₅₄ (0.25 mm, Merck) and RP-18 F₂₅₄S (0.25 mm, Merck) coated alumina plates, spots were viewed under ultraviolet light at 254 nm, and then stained by spraying with 5% H₂SO₄ in MeOH before heating on a hot plate. Silica gel (Silicycle 70–230 and 230–400 mesh, Sigma-Aldrich) and RP-18 (LiChroprep[®] 40–63 µm, Merck) were used for column chromatography.

1.2. Thelper 17 (Th17) cell polarization and intracellular staining

Naïve CD4 T cells were isolated from spleens and peripheral lymph nodes harvested from BALB/c mice via EasySepTM Mouse T Cell Isolation Kit (STEMCELL Technologies Inc., Vancouver, Canada) according to the manufacturer's protocol as previously described (Ben-Shaanan et al., 2018). The kit is designed to isolate T cells from single-cell suspensions of splenocytes and lymphocytes by negative selection. Unwanted cells are targeted for removal with biotinylated antibodies directed against non-T cells and streptavidin-coated magnetic particles. Desired CD4+ T cells are isolated. 1×10^5 cells/well CD4+ T cells were cultured in 96 well plate coated with 5 µg/mL anti-CD3 Ab for 5 days in the following polarization condition: 1 µg/mL anti-CD28 Ab (Biolegend), 20 ng/mL IL-6 (PeroTech), 1.25 ng/mL TGF- β (PeroTech), 20 ng/mL IL-1 β (PeroTech), 20 ng/mL IL-23 (R&D, USA), 20 µg/mL anti-IFN- γ Ab (Biolegend), and 20 µg/mL anti-IL-4 Ab (Biolegend). Culture medium used was IMDM (Gibco) supplemented with 1.0 mM sodium pyruvate (Gibco), 0.1 mM nonessential amino acids (Gibco), 100 IU/mL penicillin, 100 µg/mL streptomycin (Gibco), and 5% heat-inactivated FBS (Hyclone) (Harris et al., 2007; Yen et al., 2009).

For investigation of the level of IL-17A production after polarization, the polarized Th17 cells were treated with or without indicated compounds for 24 h. After incubation, cells were restimulated for 5 h in the presence of PMA (50 ng/mL), ionomycin (500 ng/mL), and GolgiStop (BD Biosciences). The IL-17A secreting cells were stained with anti-CD4 Ab (BD Biosciences) and anti-IL-17A Ab (BioLegend) and analyzed by a BD FACSVerse flow cytometer.

1.3. IL-17 luciferase reporter assay

The resultant IL-17 promoter-luciferase gene was cloned and constructed as described previously (Lee et al., 2019). Jurkat cells (4×10^6) were transfected with 10 µg IL-17 promoter-luciferase gene construct (pGL4-hIL-17prom) or control vector (pGL4.18) by electroporation according to the manufacturer's protocol (Neon transfection system, Invitrogen). Stable expression of IL-17 luciferase reporter from a clone of Jurkat cells were selected with G418 (800 µg/mL, GIBCO) for neomycin resistance. IL-17 luciferase reporter stably transfected cells (IL-17Luc cells) were seeded at the density of 1×10^5 cells/well in 96-well plate followed by treatment with PMA (50 ng/mL), ionomycin (500 ng/mL) and indicating compounds for 5 h. Assay medium was renewed and added a volume of Steady-Glo® Reagent equal to the volume of culture medium in the well for 30 min. Then the activity of luciferase in the transfected cells was measured with the BioTek Synergy microplate reader.

1.4. HPLC analysis of indigodole E

A SHIMADZU LC-20AT (Kyoto, Japan) equipped with a degasser, a binary pump, a DAD detector SPD–M20A, and a liquid handler SIL–20A autosampler was applied to analyze the chemical. The mobile phase consisted of water containing 0.1% formic acid (eluent A) and acetonitrile (eluent B). The gradient program was used as following: 10% to 76% B (22 min), 76% to 85% (10 min), 85% isocratic elution (5 min), 85% to 100% (5 min), 100% isocratic elution (8 min) with a HPLC column, COSMOSIL[®] 5C18–MS–II (5 μ m, 4.6 × 250 mm I.D.) performed at room temperature. The flow rate was 1.0 mL/min and the injection volume was 20 μ L. The concentration of indigodole E was 0.1 mg/mL. UV/Vis spectra were recorded in the range of 200–800 nm.

Reference

- Ben-Shaanan TL, Schiller M, Azulay-Debby H, Korin B, Boshnak N, Koren T, Krot M, Shakya J, Rahat MA, Hakim F, Rolls A. 2018. Modulation of anti-tumor immunity by the brain's reward system. Nat Commun. 9:2723.
- Harris TJ, Grosso JF, Yen HR, Xin H, Kortylewski M, Albesiano E, Hipkiss EL, Getnet D, Goldberg MV, Maris CH, Housseau F, Yu H, Pardoll DM, Drake CG. 2007. Cutting edge: An in vivo requirement for STAT3 signaling in TH17 development and TH17-dependent autoimmunity. J Immunol. 179:4313–4317.
- Lee CL, Wang CM, Hu HC, Yen HR, Song YC, Yu SJ, Chen CJ, Li WC, Wu YC. 2019. Indole alkaloids indigodoles A–C from aerial parts of *Strobilanthes cusia* in the traditional Chinese medicine Qing Dai have anti-IL-17 properties. Phytochemistry. 162:39–46.

Yen HR, Harris TJ, Wada S, Grosso JF, Getnet D, Goldberg MV, Liang KL, Bruno TC, Pyle KJ, Chan

SL, Anders RA, Trimble CL, Adler AJ, Lin TY, Pardoll DM, Huang CT, Drake CG. 2009. Tc17 CD8 T cells: functional plasticity and subset diversity. J Immunol. 183:7161–7168.



Figure S1. ¹H NMR (500 MHz, CDCl₃) of indigodole E.



Figure S2. ¹³C NMR (125 MHz, CDCl₃) of indigodole E.



10 9 8 7 6 5 4 3 2

Figure S4. HMBC spectrum (500 MHz, CDCl₃) of indigodole E.

100

- 120

-140

- 160

- 180

200

0 ppm

1



Figure S5. UV spectrum of indigodole E.



Figure S6. IR spectrum of indigodole E.



Figure S7. The HPLC retention time of indigodole E at 254 nm (Rt = 26.949 min).



Figure S8. The key HMBC correlations of indigodole E.

	Indigodole E	
position	$\delta_{ m H} \left(J \text{ in Hz} \right)$	$\delta_{ m C}$
1	7.89 (d, 7.5)	125.0
2	7.47 (t, 7.5)	128.5
3	7.80 (t, 7.5)	136.5
4	8.87 (d, 7.5)	119.0
4a		145.1
5a		140.9
6		158.7
6a		121.4
7	8.18 (d, 7.5)	123.9
8	7.28 (t, 7.5)	122.5
9	7.66 (t, 7.5)	134.9
10	7.73 (d, 7.5)	119.5
10a		161.0
11		139.9
11a		118.2
12		184.2
12a		124.5
13	3.05 (3H, s)	18.9

Table S1 ¹H and ¹³C NMR spectroscopic data (500 and 125 MHz, CDCl₃) for indigodole E.



Figure S9. Cytotoxicity toward Th17 cells (A) and IL-17A protein expression (B) data of compound **1**. The polarized Th17 cells were treated with or without compound **1** for 24 h. After incubation, cells were restimulated with PMA (phorbol 12-myristate 13-acetate) and ionomycin for 5 h, and then cell viability was measured by staining with 1 μ g/mL propidium iodide. The IL-17 secreting cells were stained with anti-CD4 Ab and anti-IL-17 Ab and then analyzed by flow cytometry. The data shown are representative of three independent experiments. ## *p* compared with the untreated group. **p*, ***p*, ****p* compared with the DMSO group. The concentration of dexamethasone (Dex) was 5 μ g/mL.



Figure S10. Compound **1** inhibited the IL-17 gene expression in the IL-17 luciferase reporter assay. IL-17Luc cells were stimulated with PMA and ionomycin, and co-treated with DMSO or compound **1** for 5 h. Then the activity of luciferase in the transfected cells was measured with the microplate reader. Vector means control vector (pGL4.18) transfected cells. IL 17 means IL-17 promoter-luciferase gene (pGL4-hIL-17prom) transfected cells. P+I means PMA and ionomycin. The data shown are representative of three independent experiments. ****p* compared with the DMSO group.