



REVIEW

REVISED Individuality, phenotypic differentiation, dormancy and 'persistence' in culturable bacterial systems: commonalities shared by environmental, laboratory, and clinical microbiology [version 2; peer review: 2 approved, 1 approved with reservations]

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Abstract

For bacteria, replication mainly involves growth by binary fission. However, in a very great many natural environments there are examples of phenotypically dormant, non-growing cells that do not replicate immediately and that are phenotypically 'nonculturable' on media that normally admit their growth. They thereby evade detection by conventional culture-based methods. Such dormant cells may also be observed in laboratory cultures and in clinical microbiology. They are usually more tolerant to stresses such as antibiotics, and in clinical microbiology they are typically referred to as 'persisters'. Bacterial cultures necessarily share a great deal of relatedness, and inclusive fitness theory implies that there are conceptual evolutionary advantages in trading a variation in growth rate against its mean, equivalent to hedging one's bets. There is much evidence that bacteria exploit this strategy widely. We here bring together data that show the commonality of these phenomena across environmental, laboratory and clinical microbiology. Considerable evidence, using methods similar to those common in environmental microbiology, now suggests that many supposedly non-communicable, chronic and inflammatory diseases are exacerbated (if not indeed largely caused) by the presence of dormant or persistent bacteria (the ability of whose components to cause inflammation is well known). This dormancy (and resuscitation therefrom) often reflects the extent of the availability of free iron. Together, these phenomena can provide a ready explanation for the continuing inflammation common to such chronic diseases and its correlation with iron dysregulation. This

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Any reports and responses or comments on the article can be found at the end of the article.

implies that measures designed to assess and to inhibit or remove such organisms (or their access to iron) might be of much therapeutic benefit.

Keywords

Dormancy, persisters, sepsis, microbiome, inflammation, culturability , iron dysregulation

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REVISED Amendments from Version 1

This summary of the differences between versions 2 and 1 is very short, since we simply made modifications as described in our response to the referees' very helpful comments, particularly around recognising the semantic issues (persistence 'vs' dormancy). We rehearsed a little more some of our areas of ignorance of the detailed physiological states that these high-level, replicatorily observable phenotypes represent. We clarified the meaning of "reversibility" (of growth/non-growth) in terms of states vs mechanisms. We added references to some work that we had missed, e.g. that of McKinney. We amplified slightly the points about how a 'standing crop' of mainly non-growing bacteria (else it would be sepsis) must reflect a balance between resuscitation, growth and clearance, and how these and related questions (e.g. how cells evade the innate and adaptive immune systems) represent a future 'to do' list. We stressed further that the observation of bacterial sequences in the absence of immediately culturable microbes always implies their potential for resuscitation/regrowth, although it cannot, of course, discriminate dormant from moribund, injured or irreversibly nonculturable ('dead') cells. We added a paragraph on the more philosophical reasoning behind our approach, which takes the idea that a self-consistent narrative is more persuasive intellectually than one lacking elements of join-up, a principle known in Philosophy of Science circles as 'coherence'. Finally, we entirely redid Table 3 to make its layout much more logical, and streamlined it so as to add more emphasis on the nature of the evidence of bacterial involvement in the various classes of diseases.

See referee reports

Introduction

"It is now well established that some micro-organisms can, under certain conditions, be deprived of all visible signs of life and yet these organisms are not dead, for, when their original conditions are restored, they can return to normal life and activity."¹

"Bacterial populations in both batch and continuous culture are much more heterogeneous than is normally assumed, and such cultures may consist of several types of subpopulations simultaneously differing in viability, activity and integrity of the cells"².

Consider a typical axenic flask or broth culture of bacteria (Figure 1), arguably the staple of modern laboratory microbiology. We seed a suitable growth medium with an appropriate inoculum of cells known to be capable of replicating in that growth medium. After a lag phase the number of culturable cells (the 'viable count'^{3,4}, as judged by plate counts of the number of colony-forming units observable on the same medium solidified by agar or a similar material) is observed to increase, typically exponentially, for a number of generations (the growth phase or exponential phase). Apart from the changes in nutrient concentration, and for non-synchronised cultures, it is generally taken that cells pass smoothly through their cell cycles *en route* to doubling their numbers by binary fission. The population distribution of organisms in different parts of their cell cycle during the exponential phase is thereby unchanged and thus in a steady state (from which the cell cycle parameters can even be inferred⁵). In time this increase in cell numbers ceases, usually because of the exhaustion of a nutrient in a closed system, or sometimes in part or whole because of the build-up of toxins. Again, after a further period, the viable or colony count decreases (often to

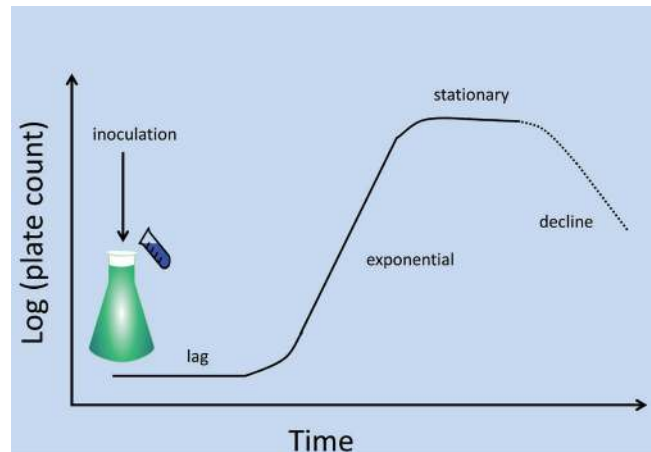


Figure 1. A typical laboratory bacterial culture. After the end of stationary phase the viable count decreases over time, but very rarely to precisely zero. Some authors recognise an extended "period of prolonged decrease"⁸⁵² during which some of the survivors undergo significant dynamics, and in which mutants are selected. Our interest here is largely in cells that have not mutated.

quite low levels if such starvation is carried out for extended periods). Inoculation of a new broth culture with a similar number of viable cells from this culture usually provides a simple repeat of the previous culture⁶, and in the absence of mutation may reasonably be anticipated, for organisms proliferating asexually, to be played out indefinitely.

The development of continuous⁷, nutrient-limited ('chemostat'⁸) or feedback-controlled ('turbidostat'⁹⁻¹¹) cultures was and is entirely consistent with this view of steady-state microbial doubling via homogeneous cell cycles that are common, within statistical fluctuations, to each cell. The same is true for cultures undergoing serial transfer (where there is slightly more of a focus on selection for genotypic variants that grow faster – see e.g. 12–14).

There should be nothing controversial in the above passage, but in fact it hides a variety of assumptions that themselves conceal a considerable feast of very interesting physiology. The chief one here is that – given that all cells in the culture are genetically homogeneous and see the same 'environment', and *modulo* where they are in their cell cycles – all such cells are indeed supposed to represent a single population (as per Figure 2). If they do not, and as we shall see they never do¹⁵⁻¹⁸, we are dealing with differentiated systems. It turns out that a particular subset of typical cell cultures – a phenotypically dormant or non-growing sub-population, occurring even in non-sporulating bacteria² – is widespread to the point of ubiquity. This leads to an exceptionally important biology with significant consequences both for our understanding of microorganisms and our ability to harness and domesticate them. Although the relevant literatures rarely cite each other or overlap, it is clear that similar phenomena are common to bacterial behaviour in the natural environment, the laboratory, and in a variety of samples of clinical interest. This theory or hypothesis that we develop here comes about from the synthesis¹⁹ of a large amount of data, and is summarised in Figure 3 and Figure 4.

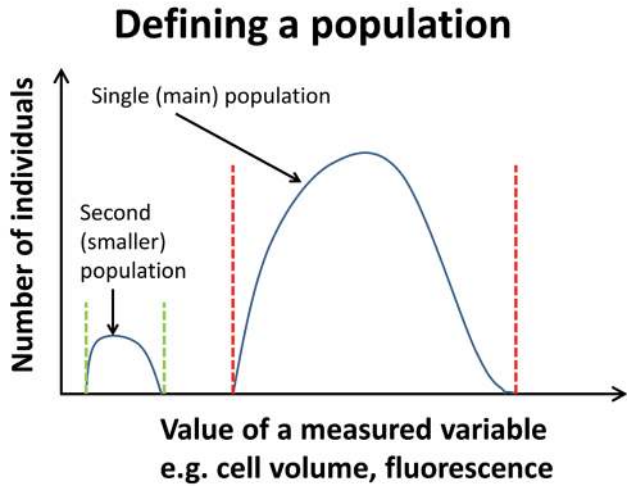


Figure 2. To clarify the general concept of a population as used here, a population of individuals involves those who share certain properties (between stated values). One main population is shown. A second, smaller population is also shown; these might represent dormant cells.

Phenotypic differentiation to dormancy or persistence – some early indications

While dormancy and resuscitation of rotifers had been observed by Leeuwenhoek himself in 1702¹, some of the earliest modern indications for a physiologically significant ‘phenotypic heterogeneity’²⁰ or differentiation of microbial cultures came in the 1940s. In a conceptually simple experiment (illustrated in Figure 5), Bigger²¹ exposed staphylococcal cultures to concentrations of penicillin that would normally be sufficient to kill them completely (and they did kill all but 1 in a million). However, these (10⁻⁶) survivors, that Bigger²¹ and McDermott²² (and many modern commentators have) referred to as ‘persisters’, were not genetic mutations selected for resistance to penicillin, since when they were inoculated into fresh broth they were just as susceptible as were those in the first culture. Bigger recognised (correctly) that the only explanation that made any kind of sense was that despite being exposed to nominally the same conditions, these cells were operationally dormant in the sense of not replicating in a medium that, apart from the penicillin, would normally admit their growth (even if they were metabolically active^{23,24}) and thus phenotypically resistant to the penicillin (that anyway kills only dividing cells^{25–27}). Similarly, Luria and Latarjet²⁸

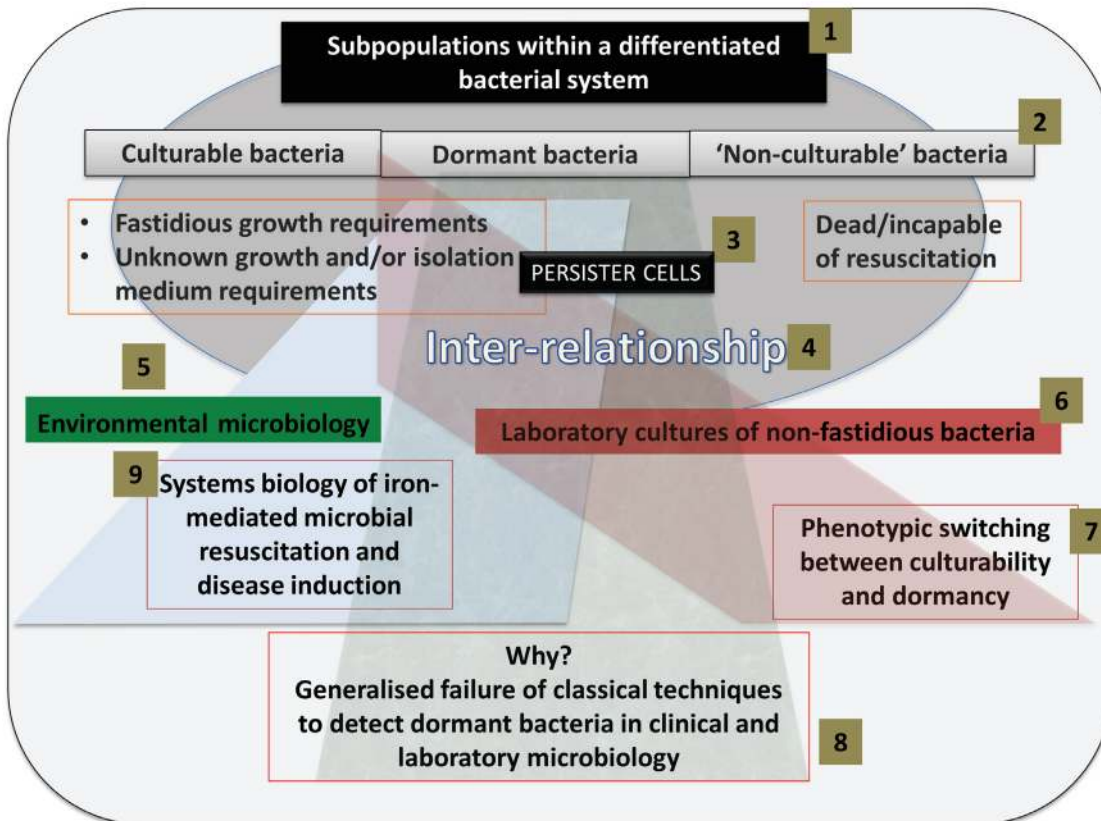


Figure 3. Infographic summary of the review. (1) A bacterial system contains distinct subpopulations, that we classify as culturable, dormant and non-culturable (2). Specific attention is given to persister cells (3), and the inter-relationship (4) between the subpopulations. Subpopulations within environmental biology are discussed (5), followed by subpopulations within laboratory cultures (6). Particular emphasis is placed on phenotypic switching between the culturable and dormant subpopulation of laboratory cultures (7). Generalized detection techniques typically fail to detect dormant cells, and we review the various reasons for this failure and discuss alternatives (8). Resuscitation of and endotoxin production by such dormant cells underpins many diseases not normally seen as having a microbial component.

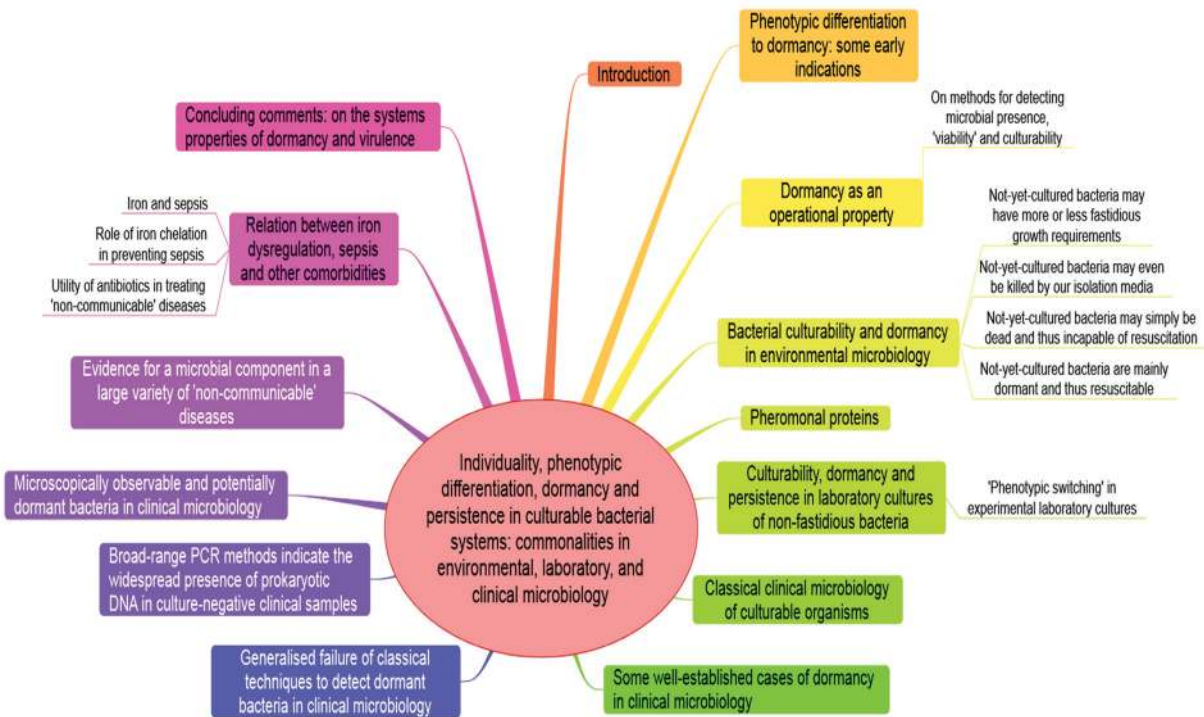


Figure 4. Summary of the review in the form of a 'mind map'⁸⁵³ of the article.

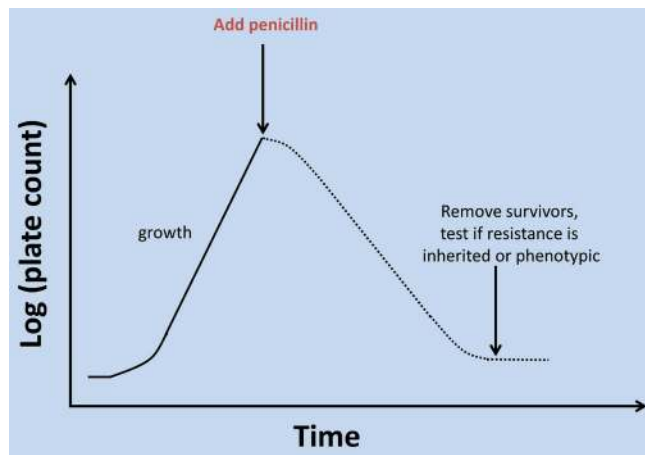


Figure 5. Assessment of phenotypic differentiation of a dormant subpopulation via antibiotic challenge. This kind of protocol can be used to determine if the resistant subpopulation has accumulated genetic mutations that encoded resistance or whether, as focused on here, the resistance is purely phenotypic. A detailed analysis of the shape of the time-survivor curves may also be informative⁸⁵⁴.

noted that approximately 1% of the cells in a culture of *Escherichia coli* displayed a phenotypic resistance to normally sterilising doses of ultraviolet irradiation. Many similar experiments since (e.g. 29–32), discussed in more detail below, have recapitulated this basic phenomenon. (We note here that high-frequency antigenic ‘phase’ variation can occur due e.g. to changes in microsatellite DNA³³; detailed discussions of such genotypic changes³⁴, including those that can affect the extent of dormancy in persistent bacteria³⁵, are outwith the scope of the present, purely phenotypic analyses.)

Dormancy as an operational property, and semantic issues

For the avoidance of doubt, and in accordance with Keilin’s description with which we opened, we shall define dormancy as:

“a reversible state of {often} low metabolic activity, in which cells can persist for extended periods without division; we shall see that this often corresponds to a state in which cells are not ‘alive’ in the sense of being able to form a colony when plated on a suitable solid medium, but one in which they are not ‘dead’ in that when conditions are more favourable they can revert to a state of ‘aliveness’ as so defined”².

We thus stress³⁶ the recognition that dormancy is not solely an innate property of a bacterial cell; it is a property assessed by one or more experiments, so whether a cell appears to be dormant depends on both the cell and the experiment used to assess that dormancy. (This principle shares a similar philosophical foundation to the independence from any specific experiment, or otherwise, of the perceived state of objects within the quantum theory^{36–38}). As do Postgate^{3,4,39} and Barer^{40–44}, we take the hallmark of a viable or living bacterial cell to be its ability to replicate or its ‘culturability’. This means that we cannot tell via culturability that a cell is alive, only (after a cell division) that it was alive^{36,45}. Dormant cells – even if ‘not immediately culturable’ – must by definition be resuscitable to form culturable cells. We also recognise (as does Michael Barer⁸⁸⁹) that it may be hard to discriminate the resuscitation of dormant cells from the recovery of injured cells. Although the term ‘nonculturable’ is quite commonly used to describe not-immediately-culturable cells it is best avoided, as we cannot try every possible combination⁴⁶ of incubation conditions that might serve to resuscitate a cell in a sample. ‘Non-cultured’, ‘as-yet-uncultured’ or ‘operationally nonculturable’ are better terms. Culturable, (operationally) non-culturable and (operationally) dormant bacteria in the differentiated bacterial (cellular) system can therefore be seen as distinct subpopulations of the system, and culturable and dormant bacteria as reversible states of the same population. A culture containing several subpopulations, whether distinct (as in Figure 2, or part of a single population characterised by a particular value from a range of an extensive variable) may be said to be differentiated (and of course may de-differentiate) in terms of physiological macrostates, that may or may not be able to interconvert. However, we recognise (thanks to Michael Barer⁸⁸⁹) that such interconversion does not imply a mechanistic reversibility. The same kinds of issues attach to cells described as having any other physiological property with regard to the ability to replicate. We note (with thanks again to Michael Barer⁸⁸⁹) that it is easy to conflate dormancy and ‘persistence’, since they do share some similarities (e.g. such cells are not immediately replicable); however, there is not much in the way of evidence as to how different say their expression profiles are, since it would require, for instance, single cell omics measurements, that are only just becoming available (e.g. 47,48), more typically⁴⁹ for the much

larger eukaryotic cells. Certainly there can be extensive changes in gross biochemical composition as cultures are starved⁵⁰. One strategy would be to separate sub-populations^{51,52}, acquire ‘averaged’ values of say their transcriptome, proteome or metabolome, and see how much they differed. In a similar vein, whether states such as dormancy are adaptive is a matter for experiment.

The general relationships between various subpopulations of the bacteria within a differentiated cellular system are shown in Figure 6.

On methods for detecting microbial presence, ‘viability’ and culturability

Given our operational definition of dormancy as including reversible culturability, we note that different kinds of assays for the presence or activity of bacteria necessarily reflect cells in different kinds of physiological states (and can thereby be used to discriminate them). Thus direct counts with stains such as acridine orange (a list of these and other methods is given in Table 1 of 36) do not determine

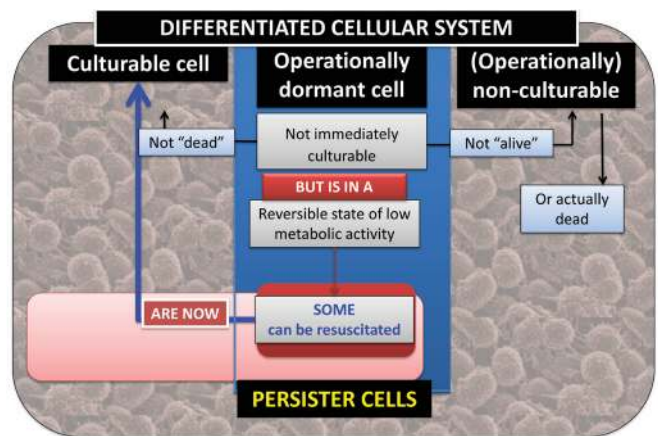


Figure 6. The relationships between culturable, dormant and non-culturable bacteria within a differentiated cellular system.

Table 1. Some bacterial infections for which an intracellular, reversibly non-replicating, persistent or dormant state is well established as part of the cells’ lifestyle. Examples are given for both low- and high-GC Gram positives, as well as a number of Gram-negative organisms.

Organism	Comments	Selected references
<i>Bartonella</i> spp.	Persists inside erythrocytes	347–350
<i>Brucella</i> spp.	Environmental and intracellular persistence and immune evasion	351–354
<i>Listeria monocytogenes</i>	Well-established low-GC Gram-positive intracellular saprophyte and non-sporulating persister	355,356
<i>Mycobacterium tuberculosis</i>	Often seen as the ‘classical’ dormant bacterium, a high-GC Gram-positive; probably one third of humans carry it in a latent or potentially dormant state; other forms may be metabolically active	357–366
<i>Salmonella typhimurium</i>	Gram-negative; non-replicating forms common in macrophages and elsewhere	367–370
<i>Staphylococcus aureus</i>	Low-GC Gram-positive; can escape antibiotics by hiding inside various phagocytes	371–374

culturability, only presence or activity. Similarly, macromolecular sequencing methods such as those based on rDNA and its amplification (e.g. 53–58), or that of other housekeeping genes (e.g. 59–61), almost certainly reflect mainly dormant cells plus any actively dividing ones (in that ‘naked’ DNA is usually degraded fairly rapidly in serum or the environment). The difference between culturable counts and total sequence-based counts probably provides one of the best methods for detecting and enumerating potentially dormant cells when they cannot yet be brought back into culture, although (as recognised by referee 1) such differences may also reflect dead, injured or moribund cells. It is particularly noteworthy (and see also 62 and below) that the amount of prokaryotic DNA in whole blood exceeds by 10–100-fold that detectable in serum⁶³, implying adsorption onto or sequestration within blood cells.

We shall return to clinical and laboratory microbiology later, but it is to environmental microbiology that we now turn to discuss the culturability of typical microbes. While the same general truths undoubtedly pertain in viruses (e.g. 64,65), and in yeasts, fungi, archaea, mycoplasmas and other unicellular organisms, our focus will be on prokaryotes.

Bacterial culturability and dormancy in environmental microbiology

It has long been known that the number of bacteria observable microscopically exceeds, typically 100-fold, those that can readily be grown axenically in standard isolation media (i.e. to proliferate in liquid culture or to form colonies on solid media). The latter has been referred to as ‘the great plate count anomaly’⁶⁶, and has been amply confirmed by more modern, culture-independent sequencing methods. A selection of papers and reviews serve to document both the numerical anomaly and the much greater biodiversity detectable by sequencing (e.g. 67–86). It is thus useful to discriminate (1) bacteria that have been cultured, that are typically available in culture collections, and whose growth requirements are known, from (2) bacteria that may be recognised as novel via macromolecular sequencing (typically of ribosomal DNA^{80,87–90}) but that have not yet been cultured and whose growth requirements may not yet even be known. Much (sequencing) evidence indicates that the bulk of the ‘missing microbes’ or ‘dark matter’^{91–93} in natural ecosystems falls into this second category⁹⁴, and that ‘single cell’ methods may be required to culture them⁹⁵.

There are at least four general reasons of principle why these organisms have not yet been cultured. We consider each in turn (although more than one may contribute in individual cases).

Not-yet-cultured bacteria may have more-or-less fastidious growth requirements

It is an elementary observation in microbiology, and the basis for selective isolation media, that not all bacteria grow on all media and in all conditions. Leaving aside truly syntrophic bacteria (that for thermodynamic or unknown nutritional reasons require another organism for growth (e.g. 96–102)), some organisms may have quite fastidious growth requirements. A number of bacteria determined as causative of disease, whose role had originally been inferred only through microscopic observation, were later cultured and could be shown to fulfil Koch’s postulates. These include *Helicobacter pylori*^{103,104} (with an unusually high requirement for urea to fuel its

alkalinogenic urease activity¹⁰⁵) and *Legionella pneumophila*^{106–109} (with an unusually high requirement for cysteine). Note that even the supposedly rich LB medium¹¹⁰ (Lysogeny Broth, often erroneously called Luria-Bertani medium, see <http://schaechter.asmblog.org/schaechter/2009/11/the-limitations-of-lb-medium.html>) is not in fact a particularly rich medium^{111–113}. An especially nice example^{114,115} is provided by *Tropheryma whipplei*, the causative organism of Whipple’s disease^{116,117}. It resisted attempts (over many decades) to bring it into axenic culture until systematic genome sequencing^{118,119} showed its requirements for a variety of common amino acids that it was unable to synthesise itself, the provision of which permitted its growth. The MetaGrowth database¹²⁰ is now available for similar purposes. Another good example is *Coxiella burnetii*, the causative agent of Q fever, for which a genome-derived growth medium (‘acidified citrate cysteine medium’) permitting axenic culture has now been developed^{121,122}. Other examples are given by Stewart¹²³ and by Singh and colleagues¹¹⁴, and include marine bacteria of the highly common SAR11 clade^{83,124,125}. Of course these kinds of phenomena are not absolute; much evidence indicates that host stress hormones may act as growth or virulence factors for a variety of Gram-negative organisms, representing a kind of ‘microbial endocrinology’ (e.g. 126–128).

Not-yet-cultured bacteria may even be killed by our isolation media

Organisms in nature are often living in low-nutrient conditions^{129–133}. It is thus reasonable (and unsurprising) that the isolation of microbes from starved, oligotrophic environments benefits from the use of low-nutrient conditions^{75,123,134–136}; some manifest this ‘starvation’ through their size, as ‘ultramicrobacteria’ (see e.g. 137–143). In a similar vein, taking cells from low-nutrient natural environments directly onto, say, a highly aerobic agar plate may produce stresses that effectively kill them, so that afterwards they would not even grow on the kinds of media (as in the previous section) that would support their growth. Thus, Tanaka and colleagues¹⁴⁴ showed interactions between phosphate and agar when autoclaved together that led to the production of compounds inimical to bacterial growth. Gellan may be a better solidifying agent here⁹⁶. However, we recognise that it may be hard to discriminate cells that we kill in the act of trying to isolate and grow them from ‘already dead’ bacteria.

Not-yet-cultured bacteria may simply be dead and thus incapable of resuscitation

While this possibility certainly exists, and is included for completeness, it is actually the least likely for a number of conceptual and empirical reasons. The first is that if an organism is present in a particular environment it must have been able to grow and divide in it at some point in the more or less recent past, even if the result of such growth was its utilisation of a finite amount of necessary nutrients or growth factors whose exhaustion caused replication to cease. (Interestingly, in soil it seems that sequestration, rather than complete exhaustion, of nutrients is the more significant phenomenon^{145–147}.) Secondly, it is highly unlikely that evolution could select for unicellular organisms that cannot replicate. Thirdly, environmental organisms can be shown to metabolise even when they cannot be shown to divide (e.g. in the ‘Direct Viable Count’ method¹⁴⁸ and in any number of other tests that detect metabolic activity^{36,149}). And finally, as we shall see in the next section, careful methods of resuscitation/cultivation do indeed allow a very significant fraction

of organisms that can be isolated from a variety of environments (e.g. the gut^{150–153}) to be resuscitated and to grow very effectively.

Not-yet-cultured bacteria are mainly dormant and thus resuscitable

As indicated in the introduction, it is now well established that even laboratory cultures, that from a macroscopic point of view are growing exponentially, contain subpopulations of non-growing cells. These cells are dormant by definition, because they may later be resuscitated and grow. It is easy to ascribe an evolutionary advantage of this culture differentiation from the perspective of the benefits of having a sub-population that by not growing is more resistant to environmental stresses (e.g. 154–156). Indeed, this general kind of phenotypic differentiation strategy, in which the variance in reproductive rate is traded off at the expense of the mean, has been referred to as bet hedging^{78,156–167} and is actually adaptive^{168,169}. An important point here¹⁶⁸ is that in many natural environments, asexually reproducing organisms such as bacteria are likely to be (spatially) close to their ancestors and descendants, such that inclusive fitness theory^{170,171} implies that it is entirely reasonable for them to behave altruistically, e.g. by ‘bet hedging’. This is also discussed further below.

It is also reasonable that in isolated (closed) natural environments, nutrients and thus sources of energy must be exhausted at some point, and thus for simple energetic reasons multiplication becomes impossible and a dormant state likely (if later resuscitation proves it to be so). Similarly, it is likely that in the absence of energy, nutrients and/or signalling molecules, and based on more ecological or community considerations (e.g. 172–175), it is necessary to add any or each of them to ‘prime’ bacteria to resuscitate. This has indeed been shown^{70,174,176–179}, including for sources of energy^{180,181}, iron-acquiring compounds¹⁸² (siderophores^{183–185}), cell wall muropeptides¹⁸⁶, and various signalling molecules^{187,188} (especially pheromones^{168,169,189,190}) that exist in natural environments^{70,174,191}. We note too that ‘kick starting’ dormant cells may require the synthesis of transporters (a neglected clade¹⁹²) necessary for the uptake of all kinds of molecules^{193–197}. Overall, the idea that most bacteria that may be observed in the natural environment are ‘unculturable’ is incorrect.

Finally here, and though this is obvious it is well worth rehearsing, the simple fact that we can store non-growing microbes under desiccated or frozen conditions or as agar ‘stabs’ in culture collections for extended periods means that most microbes are certainly well adapted to entering and leaving dormancy.

Pheromonal proteins

A related and unexpected discovery came from analyses of starved laboratory cultures of the actinobacterium *Micrococcus luteus*, in which almost all cells lost culturability^{2,198–200}. However, they were not dead but dormant, as they could be resuscitated by using a combination of weak nutrient media and a signalling molecule found in spent culture supernatants^{201–206}. The original studies used flow cytometry to discriminate the physiological state of individual cells^{51,207–210} (see also 211,212). By using another ‘single cell’ assay based on dilution to extinction (that avoids artefacts connected with the regrowth of ‘initially viable’ bacteria³⁶), we were able to purify the signalling molecule. It turned out to be a protein, named Rpf

(for ‘resuscitation-promoting factor’)²¹³. In *M. luteus* there is only one homologue²¹⁴, and the gene (product) is essential for both resuscitation and multiplication^{213,215}. Rpf contains a highly conserved 70 amino acid ‘Rpf domain’ and is widely (and probably ubiquitously) distributed throughout the actinobacteria^{216–219}, but with examples elsewhere^{220,221}. Most organisms that have a homologue have more than one. Thus *M. tuberculosis* has five homologues^{222–224}. Rpfs can have peptidoglycanase and muralytic activity^{225–230} and known crystal structures are consistent with this^{231–236}. These activities can certainly account for at least some²³⁷ of the resuscitation-promoting properties. As an extracellular protein that may be required for growth, and with a high level of immunogenicity²³⁸, it is obviously an excellent candidate target for inclusion in appropriate vaccines against pathogenic actinobacteria^{213,225,239–246}. It is also more directly of potential utility in stimulating bacterial communication and resuscitation in a variety of cultures in both samples taken from Nature^{247–257} and in the laboratory^{258–271}.

Culturability, dormancy and persistence in laboratory cultures of non-fastidious bacteria

Having established the frequency of occurrence of microbial dormancy in the natural environment, it is of interest to understand better the mechanisms by which microbes might effect this dormancy and potential resuscitation. Unsurprisingly, microbiologists have turned to *E. coli*, and considerable progress has been made^{24,272–279}.

The starting position is as in Figure 1 and Figure 6, to the effect that at any given moment in a typical culture a small fraction of the population is non-growing, and thus potentially dormant. Since clearly the same fraction cannot (or is wise not to) remain in dormancy indefinitely in the presence of suitable nutrients that permit the growth of its siblings, we must invoke at least one mechanism that can cause the bacteria to ‘oscillate’ between growing and dormant states. Many simple gene expression network topologies admit this behaviour^{159,280–284}, including a simple feedback loop with delay^{285,286}, and we note that even whole cultures can exhibit oscillations and deterministic chaos²⁸⁷. While flow cytometric observations (e.g. 51,288) show that even ‘homogeneous’ laboratory cultures show highly heterogeneous distributions in cellular volume (not just between X and 2X) and expression profiles (and see 289), our particular focus will be on ‘binary’ or ‘bistable’ systems in which individual cells either are or are not operationally culturable.

Experimentally, it is also common to assess the phenotypic ability of subpopulations of cells to tolerate normally inhibitory concentrations of bactericidal drugs^{290,291}, this being a marker for that fraction of cells that is ‘persistent’ (and maybe dormant) at the stage in question. Note that the persistence phenotype is not induced by the drugs²⁷⁵. Changes or transitions in the state of a particular cell in a population between the various phenotypic states is a phenomenon that may be (and is commonly) referred to as ‘phenotypic switching’.

‘Phenotypic switching’ in experimental laboratory cultures

A particularly well-developed example of this ‘bet hedging’ or phenotypic switching between physiologically dormant and growing states may be observed in laboratory cultures of organisms such as *E. coli* demonstrating ‘persistence’^{161,164,166,292–298}. In general, any scheme in which both a first gene product inhibits cellular proliferation and in which this first gene product may be titrated

out potently²⁹⁹ by a second gene product that thereby undoes the inhibition of proliferation, can have the effect of phenotypically switching cells between dormancy and growth. This seems to be precisely what is going on, and such pairs of gene products have been referred to (somewhat misleadingly³⁰⁰) as toxin-antitoxin (TA) pairs^{300–307}. One such involves the well-known pp(p)Gpp metabolic system that can serve to inhibit DNA gyrase^{24,308–311}, and points to the fact that in these circumstances, persisters may be quite metabolically active^{23,24,309,312}, even if transiently incapable of reproduction. Another phenotype switching mechanism, underlying colony phenotype switching, comes from metabolic bifurcations driven by the levels of a particular metabolite³¹³.

Any mechanisms that permit cells to communicate with each other can amplify switching effects by cell synchronisation, and by definition such ‘social’ signals act as pheromones, whose apparent ‘altruism’ can be explained on the basis of kin selection theory¹⁶⁸. There is considerable interest, largely outwith our scope here, in these evolutionary aspects (e.g. 314–321). Such systems are commonly, but far too broadly relative to the term’s origin³²², referred to as ‘quorum-sensing’. However, they do offer opportunities for limiting bacterial virulence (e.g. 323–330).

Classical clinical microbiology of culturable organisms

Until relatively recently, almost all of clinical microbiology^{331,332} was based on rather classical methods of plate counting³³³, coupled to assessment of antibiotic sensitivity. Various means of automated blood culture that assess metabolism exist (although they require typically 48–72h to show a ‘positive’)³³⁴. Positive tests, often implicitly involving culture (and not just metabolism) within the assay, would be followed by other tests seeking to identify the organisms detected, nowadays typically by nucleic acid sequence-based methods^{58,335–338}. However, these and other tests for the presence of antigens or even antibodies³³⁹ cannot speak to the question of culturability (and of course antigens such as lipopolysaccharide (LPS) are shed by dying cells). This said, it makes little sense to try to culture microbes from samples that molecular sequencing methods indicate lack them, so the molecular methods always provide a useful starting point for seeking to resuscitate any resuscitable (hence operationally dormant) microbes that might be present.

The existence of bacterial DNA in even ‘healthy’ blood has long been known³⁴⁰, and since naked DNA would be degraded and living cells would soon kill the host, the (seemingly) obvious conclusion that the prokaryotic DNA must reflect occult, and potentially dormant, cells seems neither to have been drawn nor acted upon.

Some well-established cases of dormancy in clinical microbiology

The idea that (typically intracellular) dormancy is a major component in some infectious diseases (including in the absence of antibiotics that may serve to light up ‘persisters’) is of course well-established, and the main purpose of this brief section is simply to remind readers of this. Such a reminder serves as a prelude to a longer discussion of the very many clinical circumstances where we consider that the role of dormant microbes is not widely appreciated, and where they are not really considered to involve a communicable or microbial component at all. Thus [Table 1](#) shows a few organisms (and references) for which we consider that most readers

would regard the idea of and evidence for dormancy as more or less uncontroversial. We do not include disease-causing infectious agents where they are better known for their ability to persist in the natural environment. Organisms such as *Legionella pneumophila* that represent significant public health issues, fall into this category, and *Legionella* and other persisters (in environments such as water system biofilms) are indeed well known (e.g. 341–345), although they too have special adaptations to an intracellular lifestyle (e.g. 346).

Generalised failure of classical techniques to detect dormant bacteria in clinical microbiology

As noted above for environmental microbiology, dormant bacteria can represent as much as 99% of the organisms that may be observed microscopically or by macromolecular sequencing, but classically (and by definition) they are not enumerated by culture-based methods that determine ‘immediate culturability’³⁶. Such culture-based methods are also widely used in clinical microbiology. However, if we were to plate out 100 μL of a culture containing 200 bacteria. mL^{-1} , of which 99% were dormant at any instant, we would expect (based on a Poisson distribution) to see fewer than 1 propagule or colony-forming unit per sample. We have noted above that it can be determined by sequencing that many of the non-cultured environmental organisms largely differ from those in standard culture collections. Certainly the examples given above in clinical microbiology, such as *Tropheryma whippelii*, were both observed microscopically and were sequenced prior to being brought into axenic culture.

The PCR method is exquisitely sensitive (down to one cell or propagule per sample), and we note that contamination artefacts from the PCR reagents represent a real issue that must always be checked (e.g. 375–379), albeit this is no less true of blood cultures³⁸⁰. We have rehearsed elsewhere⁶² five classes of argument that collectively make it implausible that these are all contamination artefacts; probably the most persuasive is simply the sheer number of prokaryotic DNA molecules that can be measured in blood and serum (e.g. 381–383). While some of the most recent nucleic acid sequencing methods (e.g. 384–389) do operate on single molecules, and genome-wide sequencing may soon be routine (e.g. 390,391), the analysis of prokaryotes usually used a broad-range PCR step to amplify small-subunit rDNA to assess their presence, whether in environmental^{74,80,88,392} or clinical^{388,393–405} samples. Using this, and while these methods alone cannot tell whether they were operationally dormant or dead, a very considerable number of studies have been performed in which ‘culture-negative’ clinical samples showed the presence of prokaryotes (at least as judged by sequence-based methods). This has some profound consequences.

We note that in a steady state such cells must be supplied at a rate equal to that of their clearance, and that the fact that clearance is lower than probably expected implies a significant ability of such cells to evade the innate and adaptive immune systems. We also take it that at least for common organisms (not very slow growers such as certain mycobacteria) the former rates must be much lower than those typically attainable in laboratory cultures, else we would have classical sepsis, and we do not. Most likely the observable facts are best accounted for by a combination of a periodic resupply of resuscitating cells, coupled to physiological changes in

non-growing cells (especially including of cell wall antigens) that help them evade natural clearance mechanisms.

Broad-range PCR methods indicate the widespread presence of prokaryotic DNA in culture-negative clinical samples

While PCR-based methods have long been used to assess the species involved in culture-positive samples⁴⁰⁶, e.g. from blood, our interest here is in samples that are culture-negative⁴⁰⁷ that may yet (and indeed likely do) contain dormant cells. Among the first such indications of this was the study by Relman's group³⁴⁰, who showed that the blood of even healthy controls contained significant amounts of prokaryotic DNA. **Table 2** lists some studies in which broad-range PCR has been used to amplify and detect prokaryotic rDNA in culture-negative samples.

In environmental microbiology, as mentioned above, there were many early indications (as observed microscopically or flow cytometrically) for the presence of bacteria that did not (or not easily) prove resuscitable or culturable. In a similar vein, many studies have shown microscopically observable organisms in

culture-negative but disease-positive samples. This is true both for diseases considered to be due to microbial pathogens and, in fact, for many others normally considered non-communicable⁶².

Microscopically observable and potentially dormant bacteria in clinical disease

Microscopic observations in tissues have been a major part of the discovery process by which certain bacteria were indeed identified as the cause of various diseases. Billings⁴³¹, Price⁴³², Domingue^{413,433–435}, Mattman⁴³⁶, Ewald⁴³⁷ and Onwuamaegbu and colleagues⁴³⁸ review the extensive and largely forgotten early literature. Domingue and Schlegel⁴³⁹ also mentioned that they could recover culturable bacteria, probably mainly from L forms (see [62,436,440](#)), from lysates of normal and diseased blood. It was to be assumed that these cells were not replicating at significant rates in the blood itself. However, we can find no evidence that this was ever followed up. Our own work^{441,442}, summarised in [62](#), showed that both bacillary and coccoid cells could be found attached to and within the erythrocytes of patients with Parkinson's disease and Alzheimer's disease, at rather greater concentrations than in samples taken from nominally healthy controls.

Table 2. Some examples of blood culture-negative but PCR-positive systems, implying the presence of dormant bacteria. Note that we have sought to exclude examples where anaerobic bacteria could be detected by PCR but not cultured simply because cultures were not anaerobic, and also cases (e.g. [408,409](#)) where high antibiotic concentrations might have prevented culture.

Aims	Culture-negative but PCR-positive	References
Assessment of endocarditis	6 out of 29	410
Development of broad-range PCR	71 out of 382	406
Development of broad-range PCR; limit of detection 5000 cfu.mL ⁻¹	10 out of 103	411
Improved broad-range PCR method	20 out of 24	53
Review	Many examples	412
Interstitial cystitis	14 out of 14	413
Endocarditis	270 (36.5%) of 740	414 (and see 415)
Endophthalmitis	116 out of 116 (selected)	416
General study	18 out of 394 (271 also culture-positive, PCR-positive)	417
Bacteraemia in intensive care	48 out of 197 45 out of 94	418 419
Sepsis/SIRS	29 out of 59 38 out of 72 culture-positive 14.6% vs 10.3% (no antibiotics) 123 vs 95	420 421 422 423
Osteoarticular samples	141 out of 1667	424
Review	Many examples	425
Various, including antibiotic-treated	34 out of 240	426
Meningitis	26 out of 274 19 out of 21	427 428
Orthopaedic samples	9% out of 125	398
Thoracic empyema	14 out of 22	429
Trauma	28 out of 35	430

In a similar way, our preliminary data show that bacteria are visible in plasma, as well as in whole blood smears in various inflammatory conditions. Here we show bacteria in platelet-rich plasma (PRP) taken from a patient with systemic lupus erythematosus and smeared onto a glass cover slip (Figure 7A and Figure 7B). We also show the same from patients with hereditary hemochromatosis (Figure 7C) and type 2 diabetes (Figure 7D). We also noted microbiota associated with erythrocytes in thromboembolic ischemic stroke (Figure 8A and Figure 8B). (Our microscopy methods are as published previously (e.g. 442–451), but fuller publications will appear elsewhere). The ultramicroscopic evidence that these are indeed small bacteria and not say, cellular debris or microparticles (see 452) is presently mainly morphological, though we note the considerable evidence for the presence of bacterial DNA in blood (see previous sections and e.g. 63,340,453).

It is worth rehearsing the very great significance of this. With erythrocytes being present at some $5 \times 10^9 \cdot \text{mL}^{-1}$ in human blood, even if

only one erythrocyte in a thousand harboured just a single dormant bacterium (that would be hard to detect microscopically, but see 453–457), the dormant bacterial load would still be $5,10^6 \cdot \text{mL}^{-1}$. This is both far from negligible, and serves to exclude the (always potentially worrisome) claim that ‘it is all contaminants’.

A culturable blood microbiome

A recent and highly significant paper by Damgaard and colleagues⁴⁵⁸ bears discussion. These workers note⁴⁵⁸ that while bacterial growth can normally be elicited during sterility testing *in vitro* from fewer than 1 in a 1000 blood units^{459–461}, transfusion-transmitted infections occur with a very much higher frequency (more like 10–12%^{462,463}, or even more⁴⁶⁴), and are responsible for a high fraction of transfusion-associated deaths^{465–467}. Although it was acknowledged that venepuncture-associated contamination or an effect of transfusion in suppressing the immune system might contribute, it was also recognised⁴⁵⁸ that one means by which to account for this would be that ‘normal blood’, and in particular its erythrocyte components, might

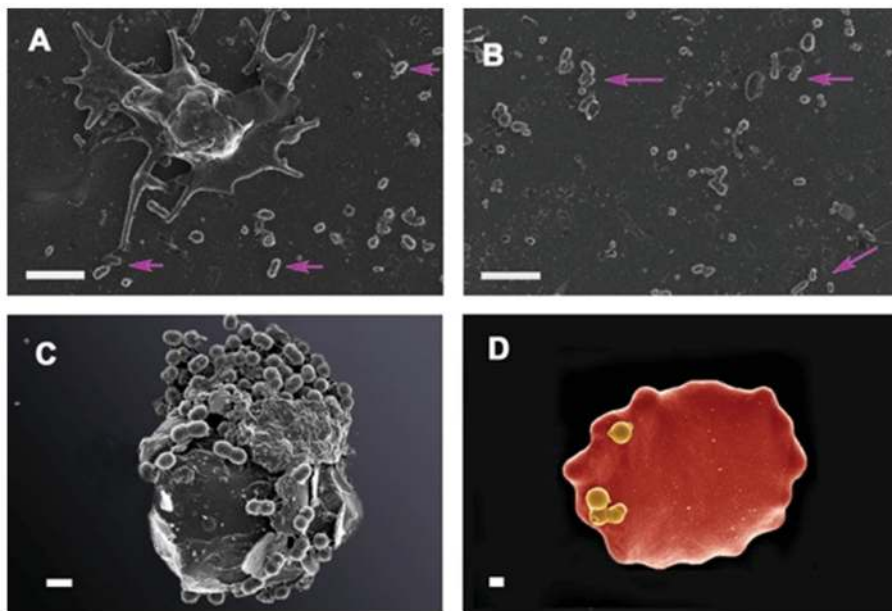


Figure 7. **A** and **B**) Platelet rich plasma (PRP) from a patient with systemic lupus erythematosus (SLE). **A**) Platelet with bacteria visible in the surrounding smear (pink arrows); **B**) areas in smear with bacteria (pink arrows); **C**) Erythrocyte with associated bacteria from patient with confirmed hereditary hemochromatosis **D**) Erythrocytes with bacteria from patients with diagnosed type II diabetes. **A–C** Scale bar: 1 μm and **D** 400 nm.

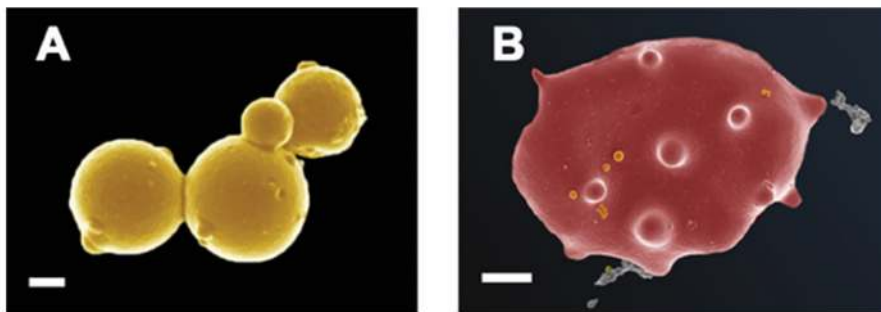


Figure 8. Bacteria in whole blood from a patient with thromboembolic ischemic stroke **A**) Microbiota in whole blood; scale bar: 200 nm. **B**) Erythrocyte with bacteria; scale bar: 1 μm .

also contain infectious agents that might be able to grow post-transfusion. Indeed, these authors found⁴⁵⁸ that under anaerobic conditions a small number of colony-forming units (ca 4–5.mL⁻¹) could be recovered by direct plating from fully 62% of blood units, with ‘controls’ producing an average of just 1 cfu.mL⁻¹. More of the bacteria were associated with red blood cells than with plasma, and rDNA was used to identify them. These data are entirely consistent with the idea that dormant bacteria are present in the blood of even ‘normal’ individuals (note that periodontitis was not a criterion for donor exclusion here⁴⁵⁸), that they are probably lurking in or on erythrocytes^{468,469}, and that they can be resuscitated and grow under the correct conditions.

Evidence for a microbial component in a very large variety of ‘non-communicable’ diseases

We have surveyed the literature for evidence in which a microbial component has indeed been observed to be an accompaniment of, and probably a major contributory factor to, a variety of (typically inflammatory) diseases that are normally considered ‘non-communicable’. Rarely has the physiological state of these microbes been considered, but since it would be obvious if they were growing, it is most likely that they are indeed dormant. **Table 3** summarises these highly extensive associations. While some are just associations, and we could have extended this table considerably, some studies (e.g. 470) contain very detailed aetiological arguments that

Table 3. Evidence for infectious agents in non-communicable diseases. We purposely largely confine ourselves to bacteria here, but include the occasional parasite, fungus, mycoplasma and virus. While obesity is usually seen as a cause of other diseases, rather than a disease itself, we note the influence of endotoxaemia on obesity^{471–476}. We note too the extensive evidence for the role of LPS in inflammation^{477–479}, and the experimental models (e.g. for Parkinson’s⁴⁸⁰) where it can induce disease directly. We do not much discuss diseases such as Crohn’s disease where the extensive uncertainty over the extent of involvement of mycobacteria (e.g. 481–483) needs no extra rehearsal (albeit it serves to illustrate the difficulties of identifying the role of hard-to-cultivate bacteria in chronic diseases). Further, while similar phenomena may be observed in a variety of cancers (e.g. 484–489), for reasons of space we have determined that this must be the subject of a separate work.

Disease	Class of bacteria	Nature of the evidence of involvement	Selected References
AUTOIMMUNE DISEASES			
Ankylosing spondylitis	<i>Klebsiella pneumoniae</i>	LPS antibodies found in various patient populations	490–493
Multiple sclerosis	<i>Clostridium perfringens</i>	Single case isolation: Immunoreactivity to ETX, fecal culture and PCR analysis, lysogenic bacteriophage footprint analysis (to exclude the possibility of laboratory contamination), sequencing of the patient-derived ETX gene	494
	<i>Chlamydia (Chlamydophila) pneumoniae</i>	17 patients with relapsing-remitting MS, 20 patients with progressive MS, and 27 patients with other neurological conditions. Bacterial present in the cerebrospinal fluid.	495–501
	<i>Chlamydia (Chlamydophila) pneumoniae</i>	PCR, Serology Many patients studied: cerebrospinal fluid	496–498, 500,501
	Infectious causes of multiple sclerosis – discussion in The Lancet Neurology		499
Rheumatoid arthritis/ Osteoarthritis/reactive arthritis	<i>Porphyromonas gingivalis</i>	Periodontal bacterial DNA in serum and synovial fluid of many patient groups Anaerobic cultures (from subgingival samples), PCR, ELISA	502–506
	<i>Porphyromonas gingivalis</i>	Antibody responses found in many patients	503,505
	<i>Proteus mirabilis</i> , <i>Escherichia coli</i>	ELISA and indirect immunofluorescence techniques Anti-LPS antibodies and human serum Elevated levels of IgM and IgA specific to bacteria Studies involving many patients	470,507–515
	<i>Mycoplasma (arthritidis mitogen, hominis and fermentans (MAM))</i>	PCR, Western Blot Elevation of antibodies to MAM in RA sera: stuies involve many patients	520–522
		Mycoplasma in 209 synovial fluid samples	520
	<i>Staphylococcus aureus</i>	Microbiology reports from patient records	523,524
	<i>Salmonella</i> <i>Shigella</i> <i>Yersinia</i> <i>Campylobacter</i> <i>Clostridium difficile</i>	Review discussing the involvement of these bacteria in arthritis	525
	<i>Propionibacterium acnes</i>	In 23 of 55 patients, undergoing primary shoulder joint replacement, <i>P. acnes</i> was found in the joint fluid	526
	<i>Chlamydia trachomatis</i>	Synovial tissues of patients: review of literature	528
<i>Chlamydia</i> from synovial fluid in single case		527	

Disease	Class of bacteria	Nature of the evidence of involvement	Selected References
Systemic Lupus Erythromatosus	Cell wall-deficient form	Histologic observations of coccoid forms suggestive of cell wall deficient bacteria in cutaneous and systemic lupus erythematosis in 7 patients	529
	<i>Streptococcus pneumoniae</i> , <i>Haemophilus influenzae</i> , <i>Mycobacterium tuberculosis</i> , <i>Listeria monocytogenes</i> , <i>Klebsiella pneumoniae</i> , <i>Staphylococcus aureus</i> ; <i>Cryptococcus neoformans</i> , <i>Aspergillus fumigatus</i>	Blood & tissue culture, patient records Hypocomplementaemia and infection with encapsulated bacteria	530–534
Vasculitis	Possibly mainly viral, but bacteria include <i>Staphylococcus aureus</i> , <i>Treponema pallidum</i> , Rickettsiaceae, <i>Borrelia burgdorferi</i> , <i>M. tuberculosis</i>	Various reviews that suggest bacterial involvement	535–541
CARDIOVASCULAR DISEASES			
General	Comprehensive reviews		383,542,543
Atherosclerosis	<i>Aggregatibacter actinomycetemcomitans</i>	This was an animal (mice) study	544
	<i>Chlamydia (Chlamydophila) pneumoniae</i>	Antigens, PCR and treatment of patients with antibiotics with good results	545–549
	<i>Helicobacter cinaedi</i>	This was an animal study. H. cinaedi infection significantly enhanced atherosclerosis in hyperlipidaemic mice	550
	<i>Helicobacter pylori</i> <i>Chlamydia pneumoniae</i>	Bacteria in atherosclerotic plaques of carotid arteries: PCR detection: study comprised 52 patients	547
	<i>Porphyromonas gingivalis</i>	PCR: periodontopathic bacteria were detected in atherosclerotic arterial wall specimens of large patient group	551–556
		PCR, IgG Titers Against <i>P.gingivalis</i> Measurement	553
		Comprehensive reviews	554,556
		PCR in a murine models	551,555
	Periodontopathic bacteria <i>Prevotella intermedia</i> <i>Treponema denticola</i>	PCR: large patient based study	552
<i>Streptococcus pneumoniae</i>	Inoculated animals	557	
<i>Toxoplasma gondii</i>	Animal (mouse) model	558	
Endocarditis	Many cell-wall-deficient forms	Comprehensive review	559 See Table 2
		Benefit of antibiotic prophylaxis: review of literature	560
Hereditary haemochromatosis	<i>Chryseomonas</i> , <i>Veillonella</i> , <i>Streptococcus</i>	qPCR: 454 pyrosequencing of 16S rRNA genes to survey the bacterial diversity of atherosclerotic plaque, oral, and gut samples of 15 patients with atherosclerosis	561
	<i>Gemella haemolysans</i>	Blood culture (Gram stain, catalase activity and biochemical characteristics)	562
	<i>Listeria monocytogenes</i>	Letter to the editor regarding infection	563,564
		Case study	564
	<i>Plesiomonas shigelloides</i>	Case study: Blood culture; API20E system	565
	<i>Vibrio vulnificus</i>	Case study: wound infection	566,567
		Infected wild-type and hepcidin-deficient mice	567
	<i>Vibrio cholerae</i>	Case studies: Blood culture; PASCO and API20E	568
	<i>Yersinia enterocolitica</i>	Case studies: Microbial cultures, serotype O:3, serotype 9	569–572
<i>Yersinia pseudotuberculosis</i>	Case studies: Mobility test and API	573,574	

Disease	Class of bacteria	Nature of the evidence of involvement	Selected References
Hypertension	Periodontal infection with <i>A. actinomycetemcomitans</i> , <i>P. gingivalis</i> , <i>T. forsythia</i> , and <i>T. denticola</i>	Large study: DNA-DNA checkerboard hybridization	575,576
	Periodontal infection	Review: Strong positive association between periodontal infection and prevalent hypertension	576
Myocardial infarction	Chronic dental infection correlated positively with MI	Association between dental chronic inflammatory diseases and the occurrence of acute myocardial infarction was studied	577–579
	<i>Chlamydia pneumoniae</i> , <i>Helicobacter pylori</i>	Large study: 3315 case patients aged 75 years or younger	580
	Enterobacteria & influenza-like illness	Immunohistochemistry: Association study	582
Stroke (and TIA)	Comprehensive papers reviewing infection and stroke		585–594
	Many bacterial species	84 different species detected in 77 patients	595,596
	Community-acquired bacteremia	Population-based cohort study	597
	Bacterial endocarditis (Organisms found included <i>S. pneumoniae</i> , <i>N. meningitidis</i> and other)	Culture of cerebrospinal fluid: Observational cross-sectional study	598
	<i>Borrelia burgdorferi</i>	ELISA	599
	<i>Brucella</i> spp.	<i>Brucella</i> agglutination and Coombs' tests in blood	600
	<i>Chlamydia pneumoniae</i>	Serology	601–603
	<i>Haemophilus influenzae</i>	Multivariate time series analysis to assess an association between infections and stroke using the established '3h-algorithm'	604
	<i>Mycobacterium tuberculosis</i>	Cox proportional hazard regressions	605
	<i>Mycoplasma pneumoniae</i>	Association between MP infection and risk of ischemic stroke; ELISA; serology	606–608
	<i>Neisseria meningitidis</i>	Latex agglutination test and counterimmunoelectrophoresis	609
	<i>Staphylococcus aureus</i>	Prospective observational cohort study and retrospective review	610,611
	<i>Streptococcus bovis</i>	Blood culture	612
	<i>Streptococcus mutans</i>	PCR	613
	<i>Streptococcus pneumoniae</i>	Cox proportional hazard model	614
<i>Streptococcus viridans</i>	Blood culture	615	
<i>Treponema pallidum</i>	Neurosyphilis also present Serology and <i>Treponema pallidum</i> haem agglutination test; rapid plasma regain test, and fluorescent treponemal antibody-absorption test <i>Serum and cerebrospinal fluid profiles for syphilis in Thai patients</i>	616,617	
	<i>Treponema pallidum</i>	Case study: Serology and haem agglutination test	616
Vascular disease (aneurysmal and lesions and atherosclerotic plaques)	Numerous bacterial species found in atheromas	Seven nonseptic patients: 6S rDNA analysis, biochemical tests, random amplification of polymorphic DNA PCR analysis, quantitative polymerase chain reaction (qPCR) and immunohistofluorescence	618

Disease	Class of bacteria	Nature of the evidence of involvement	Selected References
ENDOCRINE DISEASES			
Diabetes	Overview papers		624,625
	Pseudomonads, <i>Stenotrophomonas maltophilia</i> and <i>Ps. aeruginosa</i>	PCR and antibodies from blood samples	626
type 1	<i>E. coli</i> , <i>Candida albicans</i> , enterovirus	Urine and blood culture: from patients with urinary tract infection	627–629
	Various proteobacteria	PCR: 16SRNA from human blood	630
	Decreased bacteroidetes	Review paper	631
type 2	Systemic antibiotics improved diabetes control	Measured as a reduction in glycated hemoglobin or reduction in insulin requirements	632
	Many Gram-positives	qPCR: blood from patients	633
NEUROLOGICAL DISORDERS			
General	Comprehensive reviews		634–636
Alzheimer's Disease	Comprehensive reviews		637,638
	<i>Porphyromonas gingivalis</i> <i>Chlamydia pneumoniae</i>	Immunolabeling and immunoblotting of brain tissue for the presence of LPS from <i>P. gingivalis</i> LPS will activate innate immune system in CNS and initiate pro-inflammatory cascades.	639
	Spirochetal bacteria	Comprehensive overview papers: Immunohistochemistry, Statistical correlation of a meta-analysis	640–653
	<i>Helicobacter pylori</i>	Histology for diagnosis of Hp-I from AD patients	654–656
		Population studies: eradication of bacteria versus state of dementia	655
		Animal (Rat) model	656
	<i>Actinomyces naeslundii</i>	Serum IgG levels in patients	657
Amyotrophic Lateral Sclerosis	<i>Mycoplasma</i> infections (<i>M. fermentans</i> , <i>M. genitalium</i> , <i>M. penetrans</i> , <i>M. fermentans</i> , <i>M. hominis</i> , <i>M. pneumoniae</i>), <i>Chlamydia pneumoniae</i> , <i>Borrelia burgdorferi</i>	PCR, serology, microscopic observation: patient blood antibody analysis	436,658–660
Autism spectrum disorders	<i>Mycoplasma</i> infections (<i>M. fermentans</i> , <i>M. genitalium</i> , <i>M. penetrans</i> , <i>M. fermentans</i> , <i>M. hominis</i> , <i>M. pneumoniae</i>)	PCR	661
	<i>Chlamydia pneumoniae</i> (co-infection with mycoplasma and human herpes virus-6), or wall-less bacteria	PCR: detected in blood of patients	663
		Critical review: amyotrophic lateral sclerosis (ALS)	662
Chronic depression	Numerous Gram-negatives from gut, e.g. <i>Hafnia alvei</i> , <i>Pseudomonas aeruginosa</i> , <i>Morganella morganii</i> , <i>Pseudomonas putida</i> , <i>Citrobacter koseri</i> , <i>Klebsiella pneumoniae</i>	IgA and IgM responses in patients	665
Parkinson's Disease	<i>Helicobacter pylori</i>	¹³ C urea breath test, odd ratios for the association between treatment for HP and risk of PD using logistic regression	666–669
	<i>Toxoplasma gondii</i>	Serology, ELISA (IgG antibodies) patient-based study	670
	<i>Helicobacter suis</i>	DNA evidence: gastric biopsies of patients	671

Disease	Class of bacteria	Nature of the evidence of involvement	Selected References
Schizophrenia	<i>Toxoplasma gondii</i> (and Herpes simplex virus type 2)	A correlation between contact with house cats in early life and the development of schizophrenia exist	672–676
	Prenatal exposure to bacterial infection in the first trimester increased risk of schizophrenia in the offspring	Prospective association study	677
	<i>Toxoplasma</i> , <i>Mycoplasma</i> and <i>Chlamydia trachomatis/pneumoniae</i>	Hypothesis paper Antibodies against bacteria in blood of patients	679 678,679
OTHER INFLAMMATORY CONDITIONS			
Preeclampsia	<i>Tannerella forsythensis</i> , <i>Porphyromonas gingivalis</i> , <i>Actinobacillus actinomycetemcomitans</i> , <i>Prevotella intermedia</i> , <i>Fusobacterium nucleatum</i> <i>Treponema denticola</i> Significantly lowered risk following antibiotic treatment	PCR: placentas of 16 women	689
		Hypothesis and review	690
	Significant association with periodontal disease and UTI	Review papers	691–694
	<i>Chlamydia pneumoniae</i>	ELISA and qPCR of genomic DNA of bacteria from studies using many patients	695 (but cf. 696)
	<i>Chlamydia trachomatis</i>	Serology: Antibodies were analyzed at a first prenatal visit (mean 14.2 weeks) and at delivery	697
	<i>Helicobacter pylori</i> <i>Chlamydia pneumoniae</i>	Review paper discussing hypothesis of bacterial involvement in condition	698,699
		Serology C-reactive protein (CRP), tumor necrosis factor alpha (TNFalpha), <i>Chlamydia pneumoniae</i> IgG, IgM and plasma <i>Helicobacter pylori</i> IgA levels between 40 preeclamptic and 40 normal pregnant women	698
Chronic fatigue syndrome	Comprehensive reviews		701,702
	<i>Hafnia alvei</i> , <i>Pseudomonas aeruginosa</i> , <i>Morganella morganii</i> , <i>Proteus mirabilis</i> , <i>Pseudomonas putida</i> , <i>Citrobacter koseri</i> , <i>Klebsiella pneumoniae</i>	Serum IgA and IgM against LPS Serology	700–703
	Mycoplasmal infections (<i>M. pneumoniae</i> , <i>M. fermentans</i> , <i>M. hominis</i> , <i>M. penetrans</i>), <i>Chlamydia pneumoniae</i> , Human herpes virus-6	PCR: <i>Conference proceedings</i>	704
	Various enterbacteria and others	IgG in patient blood	705
Vitamin D receptor (VDR) dysregulation	Cell wall deficient bacteria	Evade immune destruction by invading nucleated cells where they persist in the cytoplasm. From here they down-regulated the vitamin D receptor	706
	Multiple organisms, including mycobacteria, <i>Borrelia</i>	Paper discusses a model describing how multiple species-bacterial, viral, and fungal-can cumulatively dysregulate expression by the VDR nuclear receptor	705
Antiphospholipid syndrome	<i>S. aureus</i>	A review paper: Cross-reacting antibodies	707
	Various viral and bacterial triggers	General review paper reviewing co-infections	708–710
	<i>Toxoplasma</i>	Anti- <i>Toxoplasma</i> antibody screening in 98 patients with antiphospholipid syndrome	711

Disease	Class of bacteria	Nature of the evidence of involvement	Selected References
Sudden Infant Death Syndrome	<i>S. aureus</i>	Review papers: seasonality, bacteriology	712–714
		Papers discuss markers of infection and inflammation are often found on autopsy along with microbial isolates	715,716
		Toxaemic shock indicators in serum	717,718
Other Inflammatory Bowel Diseases	Papers discussing dysbiosis of gut microbiota		719–727
Sarcoidosis	<i>P. acnes</i>	<i>P. acnes</i> antibodies and antigens	728–730
Migraine	<i>H. pylori</i>	A randomized, double blind, controlled trial	731,732
		A meta-analysis of research between 2000 and 2013	732

leave little room for doubt. Overall, the sheer size of the Table does strongly indicate the commonality of many of the microbially based mechanisms underpinning or accompanying various autoimmune and inflammatory diseases. In conditions such as atherosclerosis, transient ischemic attacks (TIAs), and stroke, it is very easy to conceive how resuscitating bacteria might serve to block the flow of blood, for instance. At all events, our main point here is that the evidence for a microbial contribution to many diseases supposedly lacking a microbial component is both multi-factorial and very considerable. Indeed, the purpose of a synthetic review such as this is to provide such pointers for more detailed studies in individual cases. Our specific interest is with the chief mechanisms by which these supposedly dormant bacteria might resuscitate and act as triggers of disease.

Relation between iron dysregulation, sepsis and other comorbidities

Many of the diseases in Table 3 are precisely those inflammatory diseases that we have listed before as coupled to iron dysregulation^{183,184,449,452,733}. A consequence of our analysis is that iron dysregulation and sepsis (as judged either by genuine infection by culturable bacteria or their inflammatory products such as LPS) should be associated causally with these various other diseases.

This leads to a variety of predictions and postdictions that we rehearse. A purposely simple (and simplistic) indication of a plausible chain of events (for which each step is underpinned by substantial evidence) is given in Figure 9, both in general terms (for unspecified diseases) and for a couple of steps to type 2 diabetes. Figure 9 aims specifically to highlight the relationship between the ability of available iron to stimulate bacterial growth and the potential disease sequelae thereof.

Iron and sepsis

First of all, it is well established that free iron may be raised in sepsis and related conditions^{734–742}, as may serum ferritin^{743–747} (that has mainly dumped its iron⁴⁵²). We have here argued that this is likely to be a significant contributor to the relationship between overt or

cryptic infection and the many iron-related inflammatory diseases discussed here and elsewhere^{183,184,452,733}. Note that patients suffering from iron overload diseases such as hereditary haemochromatosis are especially susceptible to infection (see e.g. 748–750 and Table 3). Certainly the idea that iron-related metabolism and siderophores are virulence factors (e.g. 751–763) is established unequivocally. In many diseases (e.g. lupus^{764,765} or type 1 diabetes⁷⁶⁶) it is considered that patients with the disease are more prone to sepsis, but we suggest here that (as with stroke^{581,585,586,588–590,767–775}) it may more likely be the converse that is truer: patients suffering from latent infections are in fact more prone to acquiring, having, or exacerbating the state of these other conditions, in a vicious cycle (see Figure 9).

Role of iron chelation in preventing sepsis

This was discussed at considerable length previously¹⁸⁴, and that discussion is not repeated here (though a few more recent and pertinent references include^{776–779}). However, while (perhaps surprisingly, given what we see as the evidence) it does not even appear in the guidelines⁷⁸⁰, there is considerable evidence¹⁸⁴ that appropriate iron chelation slows, inhibits or overcomes sepsis. We note, however, that some chelators are in fact known iron siderophores, and such molecules may assist the pathogen (e.g. 781–783) and are to be avoided. On this basis, iron chelation may be a suitable alternative to antibiotics in preventing multiple inflammatory diseases (and such chelation may be nutritional rather than pharmacological in nature, e.g. 183). However, it is clear that we also need to learn to kill ‘dormant’ bacteria, and this usually requires that they are growing.

Utility of antibiotics in treating non-communicable diseases

It is well established that the re-use of protein motifs in natural (and directed⁷⁸⁴) evolution means that most drugs, especially the more lipophilic ones, are promiscuous in the sense that they bind to multiple targets^{194,785} (on average six known ones for marketed drugs⁷⁸⁶). This said (and while we are very far from wishing to encourage the unnecessary use of antibiotics), the prediction here is that appropriate antibiotics will prove to have clinical benefit in diseases commonly seen as non-communicable. This is certainly known to be the

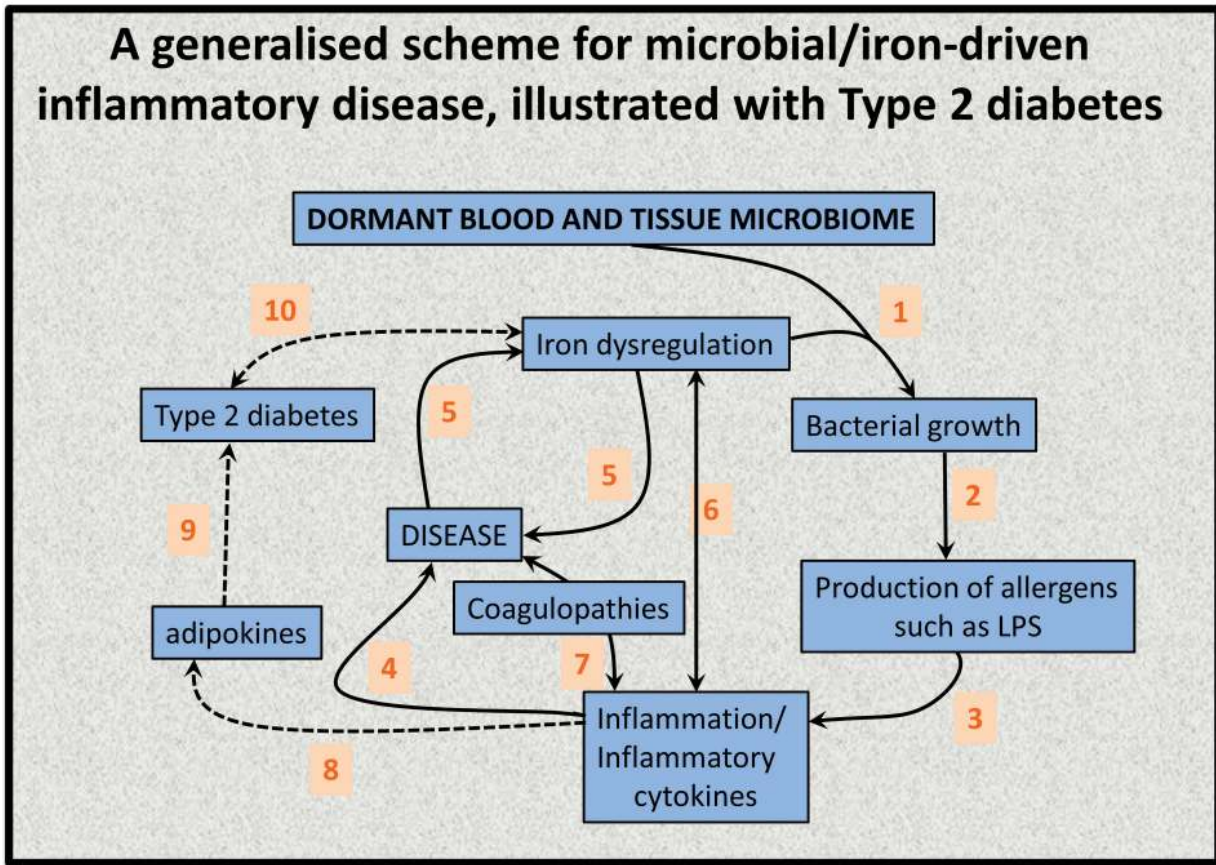


Figure 9. An elementary systems biology model of how iron dysregulation can stimulate dormant bacterial growth that can in turn lead to antigen production (e.g. of LPS) that can then trigger inflammation leading to cell death¹⁸⁴ and to a variety of diseases. While it is recognised that this simple diagram is very far from capturing the richness of these phenomena, there is abundant evidence for each of these steps, but sample references for the numbered interactions are (1)⁸⁵⁵⁻⁸⁵⁸ (especially including the release of free iron from ferritin⁴⁵²), (2)⁸⁵⁹⁻⁸⁶¹, (3)^{285,473,475,862-869}, (4)^{476,733,870-873}, (5)^{183, 184,452}, (6)^{974,875}, (7)⁸⁷⁶⁻⁸⁸², (8)⁸⁸³, (9)⁸⁸⁴⁻⁸⁸⁶, (10)^{887,888}.

case for a number of autoimmune diseases⁷⁸⁷ such as rheumatoid arthritis⁷⁸⁸⁻⁷⁹³, multiple sclerosis⁷⁹⁴⁻⁸⁰⁰ and psoriasis⁸⁰¹⁻⁸⁰³. Vaccination may prove equally effective^{804,805}.

Concluding comments: on the systems properties of dormancy and virulence

We have here brought together some of the relevant elements of environmental, laboratory, and clinical microbiology. We have argued that while their languages may differ (e.g. ‘dormancy’ vs ‘persistence’), very similar phenomena have been observed in each of these spheres (plausibly underlying a commonality of mechanism). Certainly the ability to culture microbes, and not merely to observe them (whether microscopically or via their macromolecular sequences or chemical products), remains an important goal of basic microbiology. This is likely to have significant payoffs in bioprospecting (e.g. 179,806). However, we are sure that improved methods of detecting and identifying these dormant bacteria, whether this is done via chemical imaging, through macromolecular amplification and/or sequencing, or through resuscitation and culturing, will have a major role to play in increasing the awareness of their existence and importance.

Clearly dormant and/or persistent bacteria are likely to be relatively avirulent when they are in such dormant states, and able to bypass the attentions of the innate immune system (albeit the production of superantigens by at least some microorganisms^{807,808} may be what triggers autoimmune diseases). This ‘stealth’ antigenicity is probably why they have been largely unnoticed by us too⁸⁰⁹, and their routine estimation via molecular methods⁸¹⁰ seems highly desirable. Indeed, virulence varies widely between individual strains (e.g. 811,812). Modern molecular microbiology places much emphasis on the virulence of the pathogen, with concepts such as ‘pathogenicity islands’⁸¹³⁻⁸¹⁸, ‘virulence genes’^{819,820}, and the ‘virulome’⁸²¹ being commonplace. However, if dormant microbes resuscitate (or are to be resuscitated) *in vivo* we shall need to pay much more attention to the environmental triggers that can cause this to happen than we probably have so far⁸²² (given that the pathogen genotype is fixed^{823,824}). In other words, virulence, like dormancy, is a phenotypic as well as a genotypic property. We remain largely ignorant of the means by which an optimal immune system has been selected for (or against) by longer-term evolution on the basis of microbial exposures in early life, and how this may have changed with more recent changes in human lifestyle⁸²⁵⁻⁸²⁸. Nor do we understand

how such microbes might enter and exit blood cells (and see 62,347, 829–833) (albeit the known endosymbiotic origins^{834,835} of eukaryotic organelles must have presaged such mechanisms). Similarly, we do not yet know what may cause these dormant microbes to resuscitate (and/or to exit their intracellular niches). However, the potential for iron-associated replication and (e.g.) LPS production and shedding does provide a very straightforward explanation for the continuing low- or medium-grade inflammation characteristic of the many inflammatory diseases we have considered here and elsewhere^{183,184,449,452,733,890} (Figure 9).

Recognising that correlation does not at all equate to causality (e.g. 195,836), one approach to Science is based on varying independently something considered a cause and observing its predicted effects (e.g. 195,837,838). Temporal covariation of measurands can also be performed. The levels of free iron are clearly one such possibility. To assess causality in microbiology it is usual (e.g. 815, 839–841) to invoke what are (variously⁸⁴² referred to the Henle-Koch or Koch's postulates. These are based on the nature and presence, but not the physiological state, of an agent that might be believed to 'cause' (or at least contribute to) an infectious disease. Consequently, dormancy poses something of a challenge to the full completion of the required tests. Indeed a number of authors^{437,815,842–845} have recognised that these tests may need revision in the light of the ability to identify disease-causing microbes by sequence alone. We suspect that a key element here will be the ability to resuscitate dormant organisms *in vivo* and to see the effects of that on clinical disease.

From a 'philosophy of science' point of view (e.g. 841), one strategy taken to develop the evidence for a particular point of view

hinges on the idea that if a series of ostensibly unrelated findings are brought together into a self-consistent narrative, that narrative is thereby strengthened. This is the strategy pursued here, and it is known as 'coherence'^{846–848}.

As phrased by Silvers⁸⁴⁹, "Several of our contributors showed how discoveries and insights could emerge with what seemed great promise, and yet be pushed aside, discarded, and forgotten – only to re-emerge once again, sometimes many years later, and become, in their new formulation, accepted as important". In this sense, and as presaged in the opening quotation¹, it seems that ideas, as well as bacteria, can remain dormant for extended periods^{850,851}.

Author contributions

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No competing interests were disclosed.

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Gerald Domingue

Professor Emeritus, Tulane University, New Orleans, LA, USA

This review paper is an important modern perspective on bacterial persistence and expression of disease. The role of 'stressed', atypical, cell wall-defective, cryptic, pleomorphic forms in chronic inflammatory diseases in clinical medicine and especially in diagnostic pathology and clinical microbiology is grossly neglected and overlooked. This paper warrants publication for its timely approach to a vastly important overlooked topic in science and medicine and its relevant, useful 890 cited references. I see no point in further elaboration on semantics: persistence vs. dormancy. In my opinion, regardless of terminology preferred (or debated) the relevant and important fact is identity of atypical forms in tissues, their basic biology and their relationship to disease.

Although there is provocative circumstantial evidence linking pleomorphic forms suspected of being bacterial in origin in a wide array of chronic diseases, many were categorized as autoimmune, unrelated to microbes (unambiguous proof lacking). While the persistence of stainable forms (various structures) are often seen in tissue specimens utilizing histopathologic and bacteriologic stains, most are discarded as insignificant staining artifacts and debris, and especially in the absence of non-cultivable bacteria from accompanying specimens. In my opinion, this is a primary reason the significance of such stainable findings has been ignored in clinical medicine and has stymied their identity as causative agents of disease. Furthermore, even when there may be growth of atypical bacterial forms on artificial culture media, bizarre (non-standard) morphologic, biochemical, physiological characteristics of the isolated organism *in vitro*, the findings are most often disregarded as "contaminants".

Permit me to digress and cite such an example of an unidentified pleomorphic form isolated from patients with interstitial cystitis (a chronic, debilitating disease of unknown origin). These atypical isolates were subjected to elaborate, microbiological, immunological, biochemical, physiologic and electron microscopic characterizations. The findings (data) were presented at the American Urological Association annual meeting, only to have a well-known academic urologist and interstitial cystitis specialist congratulate the researchers for the elaborate experimental description of an 'artifact'. Obviously with that type of unexpected and disappointing comment,

there was nothing more to be said, other than, thank you! So there you have it: The regrettable dismissal of a potentially important finding in a disease of unknown origin by an individual who could not see the forest through the trees and without evidence to substantiate the claim that the finding was an 'artifact'.

The 14 topics and subsections outlined in Figure 4 of the review set the stage for the "dramatics" – 'mind map' – that follows. In itself this graphic may have been enough when accompanied by germane references instead of lengthy written discussions for each topic since much is *déjà vu*, gleaned over a period of many decades from published findings. On the other hand, it is often useful to repeat, for emphasis, and especially to call attention to neglected topics, which I suspect was the intent. Although the tables and graphics are worthwhile, it does take time to digest it all, meaning it may have been possible to shorten the paper.

Two important publications (not cited in list of 890 references): companion papers by Green *et al* in *Infection and Immunity*, 1974, Oct; 10 (4): 889-914 and 915-927; demonstrated the phenomena of microbial persistence and reversion with *Streptococcus faecalis* L-forms in human embryonic kidney cells, followed by a proposed reproductive cycle for a relatively stable L-phase variant of *Streptococcus faecalis*. I call these publications to the attention of the authors because of their possible application to the fundamental basis of persistence by 'stressed', atypical bacteria in chronically diseased human subjects. Essential to the thesis of Green *et al* is that small, electron dense, non-vesiculated L-forms were shown to be the central (core) element in bacterial persistence in these experimental studies. The researchers concluded that depending on the stimulus received, these dense forms might be considered as undifferentiated cells, with the capacity to develop along several different routes. *In vitro*, the dense form was observed to divide and bud rapidly. In addition, the dense forms appeared to be capable of growth and development within vesicles of mature mother forms. When these forms were released from the vesicles into the surrounding fluid medium, further growth occurred, resulting in the development of immature and ultimately mature mother forms. Under conditions unfavorable for L-form growth, these dense forms developed first into transitional forms and then into the bacterial form. These dense forms might therefore be considered as undifferentiated 'stem cells' with the capacity to develop along several different routes, depending upon the stimulus received. Hence, in applying these findings to altered forms created *in vivo* (humans) these may take up intracellular and/or extracellular residence; possibly establishing a sort of immune protected parasitic relationship persisting/surviving phagocytic action, and creating subtle pathologic changes in the host during a prolonged period of tissue persistence. This might translate into an etiology for chronic inflammatory diseases, when the 'stressed' bacteria increase in numbers and overwhelm the normal biological functions of the host. I further propose that *in vivo* persistence of these bacterial elements escape immune surveillance partially, completely, or may integrate with host cell organelles to create bacteria-host cell-antigen complexes which could provoke immunopathologic consequences. Highly relevant, recently published data on modifications of gene expression, modes of division for stressed bacteria, and the paradoxical finding of peptidoglycan in L-forms are pertinent to the hypothesis that atypical, pleomorphic bacteria are the organisms responsible for persistence and expression of disease. Finally, it is hoped that the Kell, Potgieter, Pretorius timely, interesting and provocative review will call attention to this highly significant, too often overlooked subject. In my opinion, this review calls for a scientific/medical challenge: 1) to motivate visionary scientists and clinicians to investigate the fundamental origins of bacterial persistence in chronic diseases; 2) to unambiguously identify tissue persisting forms utilizing modern molecular technology, and 3) to design elegant experiments to provide

convincing scientific proof (or disprove) that extracellular and or intracellular stainable bodies observed in histopathologic specimens and dense bodies at the electron microscopic level (culture negative) are bacteria existing as 'stressed' altered forms in tissues and not tissue or staining artifacts. Proof of the above hypothesis would open new arenas in clinical diagnosis, management and treatment of numerous chronic inflammatory human diseases of unknown etiology and might even extend to a bacterial cause for certain malignancies (as previously proposed many decades ago).

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Version 1

Reviewer Report 11 August 2015

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Vanya Gant

Department of Medical Microbiology, University College London Hospitals NHS Foundation Trust, London, UK

I review Kell *et al's* review relating to individuality, phenotypic differentiation, dormancy and "persistence" as a clinical microbiologist, infectious diseases doctor, with an interest in developing and assessing the impact of rapid sequence-based molecular blood and lung diagnostics in the critically ill.

This review reminded me of Mussorsky's *Pictures at an Exhibition*, a collection of hastily composed pieces whose theme was to take an interested individual through an art gallery, and to tarry awhile in front of 10 *Tableaux*, interspersed with musical elements referring to the "Promenade" through the gallery.

And so it is with Kell *et al's* review. After an introductory *Promenade* relating to matters of bacterial dormancy and its relationship with just about any other conceivable physical state between life and death, exhaustively referenced together with the thought provoking *Postgate*-ian concept of the difficulties inherent in differentiating bacterial life from death if you only have an instant in time to measure it – we are then presented with several pictures, garlanded for us in extensively referenced detail by the authors. Were mindmaps not enough to capture the reader's curiosity as to this *magnum opus* of a kind, we are invited to walk through Kell *et al's* gallery of mental pictures depicting scenes of the Yet to be Cultured, Those bacteria that aren't culturable yet but are

certainly not dead, the biological importance of bacterial pheromones, the evils of Iron - thence to the Clinical Microbiology Room of Pictures with a liberal helping of systems biology throughout.

I am a proponent of, and believer in, the present and future potential of Nucleic Acid Technology (NAT) for pathogen detection in Clinical microbiology and I use such techniques on a daily basis. When appropriately deployed, it allows me to find those “unculturable” pathogens as drivers for individual clinical cases of infection. Perhaps strangely, this is a relatively new paradigm for most practising clinicians, and one which likely will generate fundamental discoveries highly relevant to human disease, and for all we know as equally important as *Helicobacter*. That such sequences should be found in blood is hardly surprising, given that human beings have between 10 and 100 times more bacterial cells than their own, living (or persisting, or dormant) on and in them. This groups’ demonstration of bacteria adhering to red cells (also *in* red cells) is certainly very intriguing, and such suggested “atopobiosis” is more expansively dealt with in another publication and prompts far more questions than it answers – in a good way. Another obvious question relates to how these adherent bacteria may remain undetected and intact in the presence of numerous moieties central to both innate and acquired immunity (complement and antibody to name but two) as well as escaping phagocytosis in the liver and spleen. It would certainly be interesting to look at red cells in the grave condition of erythrophagocytosis, a condition whose mechanism is in most cases obscure –it might even be that adherent bacteria “opsonize” the red cells in these cases. This reader, however, does balk at the very serious work to be done as regards untangling the mechanistic nature of an “association” with several diseases, and certainly at this stage it would be very unwise to suggest it’s anything more than that. Further work of this nature should be approached and undertaken with extreme caution and rigor in view of the myriad possible explanations other than causative ones; the Measles vaccine/autism saga comes to mind here.

It is likely therefore that such technologies will perforce “lift the lid” on what might lie beyond the Culturable, and its relationship to human disease. This is explored in Table 3, which represents a *tour de force* as concerns the sheer volume of references relating to all that appears to associate human Disease and organisms, mostly bacteria.

Unfortunately, this Table doesn’t work for me. Whilst it will serve me as a unique and accessible resource of information in this space, it is anarchic. Correctly described as “Evidence for agents in non-communicable diseases”, it lists, in no particular order, and with no apparently critical eye, references 470 to 712 as relevant to the Table subject stated above. This list’s breadth as concerns both organisms and clinical diseases is extraordinary; and the literature quoted in a table described as “effect of bacterial involvement” ranges from unusual cases, to mechanistic assumptions of what LPS might do, to the concept of “dysbiosis” amongst many others. I was left rather dizzy from the mental exercise needed to constantly adjust to the sheer scale and variation of why a particular organism, or something it produces, might either directly causally relate to a particular disease, or perhaps through the individuals’ immune response to it; especially now we know how outbred we are as concerns immune responsiveness.

This review finishes with an impressive and lyrical chiding for Scientists, whereby those who research this field should wake up from their intellectual slumber, as might and indeed do bacteria.

This review is additionally peppered with tantalizing if perhaps sometimes unfounded

assumptions, some arguable and some bordering on plain unreasonable. Certainly my eyebrow raising went into overdrive when considering Kell's conviction as concerns a Catholic Grand Unifying Theory based around the Evils of Iron, the subject of a previous equally grand *Magnum Opus*.

This review has to be one of the most undisciplined I have read in a long time, on occasions associating seemingly disparate observations and conflating "scientifically" determined facts with clinical issues.

Having said this, I should finish by applauding Kell *et al's* review as a thumping good read. It's fast paced, edgy, a real treasure trove of papers for me to read at leisure, and goes way outside the usual, expected and conventional boundaries of style of prose and rigor we "normally expect" of such scientific publications. And (warts and all, and there are many) it left this reader thinking that there indeed is Life beyond dormancy within the review's style itself, beyond the doubtless very important but less imaginative run-of-the-mill, tightly written yet dreary "Scientific Publication". It is almost as if this review in all its unconventionality were particularly well aligned to the current state of the Art for the Uncultured in Clinical Medicine (bacteria, not Doctors) and its potential to release significant Paradigm shifts. No doubt this reviews' readers are made up of those who have the capacity to appreciate Kells' latest brand of emergent, imaginative systems biology style of thinking underneath what some might consider a publication of inadequate scientific rigor.

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 18 Aug 2015

Douglas Kell, The University of Manchester, Manchester, UK

- *"I review Kell et al's review relating to individuality, phenotypic differentiation, dormancy and "persistence" as a clinical microbiologist, infectious diseases doctor, with an interest in developing and assessing the impact of rapid sequence-based molecular blood and lung diagnostics in the critically ill.*

This review reminded me of Mussorsky's Pictures at an Exhibition, a collection of hastily composed pieces whose theme was to take an interested individual through an art gallery, and to tarry awhile in front of 10 Tableaux, interspersed with musical elements referring to the "Promenade" through the gallery.

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be Cultured, Those bacteria that aren't culturable yet but are certainly not dead, the biological importance of bacterial pheromones, the evils of Iron - thence to the Clinical Microbiology Room of Pictures with a liberal helping of systems biology throughout."

This is a lovely analogy, which we shall let readers enjoy in the open referee's report; we are probably not capable of recasting the review in Mussorgskian style anyway! In this regard, readers might also enjoy a little known and whimsical piece on bioinformatics that takes just such an approach: Goble C, Wroe C: [The Montagues and the Capulets](#). *Comp Func Genomics* 2004; 5:623-632.

- *"I am a proponent of, and believer in, the present and future potential of Nucleic Acid Technology (NAT) for pathogen detection in Clinical microbiology and I use such techniques on a daily basis. When appropriately deployed, it allows me to find those "unculturable" pathogens as drivers for individual clinical cases of infection. Perhaps strangely, this is a relatively new paradigm for most practising clinicians, and one which likely will generate fundamental discoveries highly relevant to human disease, and for all we know as equally important as Helicobacter. That such sequences should be found in blood is hardly surprising, given that human beings have between 10 and 100 times more bacterial cells than their own, living (or persisting, or dormant) on and in them. This groups' demonstration of bacteria adhering to red cells (also in red cells) is certainly very intriguing, and such suggested "atopobiosis" is more expansively dealt with in another publication and prompts far more questions than it answers – in a good way. Another obvious question relates to how these adherent bacteria may remain undetected and intact in the presence of numerous moieties central to both innate and acquired immunity (complement and antibody to name but two) as well as escaping phagocytosis in the liver and spleen. It would certainly be interesting to look at red cells in the grave condition of erythrophagocytosis, a condition whose mechanism is in most cases obscure –it might even be that adherent bacteria "opsonize" the red cells in these cases. This reader, however, does balk at the very serious work to be done as regards untangling the mechanistic nature of an "association" with several diseases, and certainly at this stage it would be very unwise to suggest it's anything more than that. Further work of this nature should be approached and undertaken with extreme caution and rigor in view of the myriad possible explanations other than causative ones; the Measles vaccine/autism saga comes to mind here."*

These are excellent points, and we have covered some of them in the forward-looking concluding section. While they might be seen as 'premature' (in the sense that it requires acceptance of the basic 'dormancy' hypothesis in the first place) they do point to important areas where we would seek a mechanistic understanding of what is going on.

- *"It is likely therefore that such technologies will performe "lift the lid" on what might lie beyond the Culturable, and its relationship to human disease. This is explored in Table 3, which represents a tour de force as concerns the sheer volume of references relating to all that appears to associate human Disease and organisms, mostly bacteria.*

Unfortunately, this Table doesn't work for me. Whilst it will serve me as a unique and

accessible resource of information in this space, it is anarchic. Correctly described as "Evidence for agents in non-communicable diseases", it lists, in no particular order, and with no apparently critical eye, references 470 to 712 as relevant to the Table subject stated above. This list's breadth as concerns both organisms and clinical diseases is extraordinary; and the literature quoted in a table described as "effect of bacterial involvement" ranges from unusual cases, to mechanistic assumptions of what LPS might do, to the concept of "dysbiosis" amongst many others. I was left rather dizzy from the mental exercise needed to constantly adjust to the sheer scale and variation of why a particular organism, or something it produces, might either directly causally relate to a particular disease, or perhaps through the individuals' immune response to it; especially now we know how outbred we are as concerns immune responsiveness."

We very much accept the point that the table could be improved with regard to ordering, and we have done so accordingly. However, we think that readers will recognise it for what it is (as does the referee), viz. as a useful resource and/or pointer to a large literature in which specialists in disease X may wish to read at least those papers we suggest as relevant to 'their' disease, while others will simply see it as a recognition of the widespread evidence for our more general claims.

- *"This review finishes with an impressive and lyrical chiding for Scientists, whereby those who research this field should wake up from their intellectual slumber, as might and indeed do bacteria.*

This review is additionally peppered with tantalizing if perhaps sometimes unfounded assumptions, some arguable and some bordering on plain unreasonable. Certainly my eyebrow raising went into overdrive when considering Kell's conviction as concerns a Catholic Grand Unifying Theory based around the Evils of Iron, the subject of a previous equally grand Magnum Opus."

As mentioned in the comments on the review of referee 1, the basis for this is the desire to produce a coherent story (in the sense used by Philosophers of Science), and (as referee 1 also states) it is well known that microbial growth in vivo is normally limited by iron availability. That iron dysregulation is also a hallmark of just those chronic inflammatory diseases that we highlight here is consistent with this view, and indeed serves to provide a simple explanation for this. Of course, as the referee indicates (and referee 1 does too), further demonstrations will benefit from varying iron levels as an independent variable.

- *"This review has to be one of the most undisciplined I have read in a long time, on occasions associating seemingly disparate observations and conflating "scientifically" determined facts with clinical issues.*

Having said this, I should finish by applauding Kell et al's review as a thumping good read. It's fast paced, edgy, a real treasure trove of papers for me to read at leisure, and goes way outside the usual, expected and conventional boundaries of style of prose and rigor we "normally expect" of such scientific publications. And (warts and all, and there are many) it left this reader thinking that there indeed is Life beyond dormancy within the

review's style itself, beyond the doubtless very important but less imaginative run-of-the-mill, tightly written yet dreary "Scientific Publication". It is almost as if this review in all its unconventionality were particularly well aligned to the current state of the Art for the Uncultured in Clinical Medicine (bacteria, not Doctors) and its potential to release significant Paradigm shifts. No doubt this reviews' readers are made up of those who have the capacity to appreciate Kells' latest brand of emergent, imaginative systems biology style of thinking underneath what some might consider a publication of inadequate scientific rigor."

Many thanks for these last comments; we have nothing further to add here.

Competing Interests: No competing interests were disclosed.

Reviewer Report 23 July 2015

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Michael R Barer

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Kell, Potgieter and Pretorius present a stimulating and argumentative review ranging from the interrelationships between the culturability of bacteria and their viability and any links these descriptions may have to defined physiological states, through a discussion of environmental bacteria and ultimately focusing on the human-associated microbiota, particularly those found in blood (without associated symptoms of sepsis) and their proposed roles in disease. Two central themes are developed beyond those that have been discussed extensively elsewhere: 1) the proposal that failure to culture bacteria from many samples often reflects dormancy and 2) that such dormant bacteria interact with host iron regulation to contribute to or directly cause a panoply of chronic diseases largely labelled as non-communicable.

At a general level I support the provocative stance taken by the authors. With 861 cited references, at the very least they provide a valuable resource for anyone wishing to consider the potential microbial contribution to diseases traditionally considered free of this aetiological component. Of course *Helicobacter* infection stands as a monument to the stupidity of dismissing this possibility in the face of carefully assembled evidence. Indeed this reviewer, who many years ago, was presented with a case of duodenal ulcer in his final medical exams, would probably have experienced quite a different career had he claimed a role for infection in causing his patient's pathology.

In considering the specific points presented I have multiple concerns, the most significant of

which I will indulge in outlining below.

Semantics present a central problem in considering bacterial viability and physiology and I broadly support the approach taken here. The authors do try to define their terms but some problems remain. In particular I take issue with the very broad application of term "Persisters" which should be reserved for cells that survive (have the potential to replicate) after exposure to an antimicrobial stress to which kills most cells in an actively growing culture of the organism concerned. Conflation of this term with "Dormancy" implies on the one hand that the persisting cells must have been dormant and on the other that dormancy and persistence represent the same physiological state in bacteria. This difficulty resurfaces later when they define dormancy but other problems emerge before then.

I was next concerned by the extensive use of the term "Differentiation". I completely agree that what we used to think of as uniform bacterial populations are probably never so but the degree to which subpopulations may be considered differentiated rather than reflecting a range of adaptive responses or indeed, some degree of injury, is not considered here and again I think this leads to problems in considering their hypotheses under a unitary banner downstream. I consider differentiation to require phenotypic changes that are not directly reversible, as in the case of sporulation, whereas adaptation can involve expression of a single gene that can be reversed by its subsequent repression. I do agree that cell cycle contributes to the range of phenotypes in a pure bacterial culture and that this is not the only reason for their diversity (but was not enlightened by use of the term "*modulo*" in this regard).

The operational definition of dormancy given deliberately leaves open the possibility of metabolic activity and seems only to require that the cell so defined should not divide; this did not allow me to recognise which operational tests might be applied to enumerate or detect dormant cells. Subsequently the detection of molecular signals indicative of bacterial presence in samples from which they were not isolated in culture is taken as evidence of dormancy. In the first case do we accept any non-dividing cell as dormant and in the second I can (and will) offer multiple alternate explanations other than dormancy. Moreover, returning briefly to the conflation between dormancy and persisters, the recent work of John McKinney and colleagues shows that antibiotic exposed persisting cells are not necessarily non-dividing cells in the mycobacterial system he studied.

Alternative interpretations of the presence of bacterial 16S rDNA sequences in blood when culture fails to detect the organisms from which they derive, include the presence of dead, injured or moribund cells. If they are shown to be repeatedly present then they must either be able to persist in the face of clearance mechanisms or be supplied at a rate equal to their clearance; both seem equally plausible to the dormancy explanation to me. Moreover, why the first three explanations offered for "Not-yet-cultured" should apply to environmental bacteriology but not to clinical samples escapes me.

I am led to the conclusion that the authors have chosen to label evidence for discrepancies between culture and nucleic acid detection of bacteria in blood to give their hypotheses a simple headline. I have no problem with the proposal that human blood and tissues classically considered sterile in the absence of overt symptoms of infection are frequently exposed to bacteria and bacterial products that in many cases contribute to serious chronic disease. However, I consider the burden of available evidence currently provides many potential explanations within the field of

microbiomics/metagenomics in contrast to the dormancy hypothesis offered here. Further, I feel this broad application of dormancy to bacterial phenotypes which, even in the case of Rpf dependency, have not been shown to result from a programme of gene expression that could be considered as differentiation, diminishes the value of the term. Indeed there remains no direct proof that dormancy of *Mycobacterium tuberculosis* underpins what we call latent tuberculosis infection and it is not essential to the observed clinical or pathological pattern, notwithstanding the widespread acceptance of this view by most researches, including me.

I am not fundamentally opposed to the ideas presented by Kell and colleagues but I do not think they are assisted by lack of attention to the contradictions I have identified above.

Finally I come to the iron dysregulation hypothesis and its pro-inflammatory consequences. It is beyond my expertise to comment on the plausibility of the inorganic chemistry deployed here or to review the evidence relating to more than a fraction of the conditions listed. The importance of the struggle between pathogens and host for access to iron is beyond question. When I entered the medical field of infectious disease it was fully recognised that depriving bacteria from iron was a potential therapeutic angle and indeed iron chelation was studied. Desferioxamine, a widely used agent in iron overload, was investigated and found to effectively deliver iron to the pathogen and the approach was set aside. More recently this agent has been identified as a major risk factor in serious fungal infection and guidance specifically recommends its avoidance. Newer agents seem not to suffer from this problem and the approach deserves renewed attention. However, I would not underestimate the ability of pathogens to outwit our pharmaceutical industry in the battle to sequester iron. While there are reasons beyond the host-pathogen tug-of-war for iron to consider chelation as a therapeutic option, the potential for adverse effects is significant and I think the suggestion that omission of iron chelation from recent guidance on sepsis management is "shocking" is not justified.

Focussing briefly on the specific diseases cited and their relation to bacterial exposure in one form or another, I find that evidence cited frequently rests on what can be considered "fringe" hypotheses that have little currency in their respective fields. This is not to discourage their continued pursuit but it does weaken the strength of the authors' argument when investigation of the supporting literature frequently leads to papers that are given little credence in the specialist field. Of course "cave *Helicobacter*" must remain on the table. But there, an accidental technical breakthrough led to an avalanche of convincing laboratory and clinical data.

In summary Kell, Potgieter and Pretorius have produced an interesting read which bring many important ideas to our attention. I am not convinced of the breadth of conditions to which they argue their ideas are applicable and I await with interest, demonstration of of how they may be practically pursued and some selected definitive proofs that iron-driven inflammatory disease is as important as they claim.

Competing Interests: No competing interests were disclosed.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 18 Aug 2015

Douglas Kell, The University of Manchester, Manchester, UK

- *"Kell, Potgieter and Pretorius present a stimulating and argumentative review ranging from the interrelationships between the culturability of bacteria and their viability and any links these descriptions may have to defined physiological states, through a discussion of environmental bacteria and ultimately focusing on the human-associated microbiota, particularly those found in blood (without associated symptoms of sepsis) and their proposed roles in disease. Two central themes are developed beyond those that have been discussed extensively elsewhere: 1) the proposal that failure to culture bacteria from many samples often reflects dormancy and 2) that such dormant bacteria interact with host iron regulation to contribute to or directly cause a panoply of chronic diseases largely labelled as non-communicable.*

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In considering the specific points presented I have multiple concerns, the most significant of which I will indulge in outlining below."

Many thanks for the above; it is perfectly accurate and we have nothing to add here.

- *"Semantics present a central problem in considering bacterial viability and physiology and I broadly support the approach taken here. The authors do try to define their terms but some problems remain. In particular I take issue with the very broad application of term "Persisters" which should be reserved for cells that survive (have the potential to replicate) after exposure to an antimicrobial stress to which kills most cells in an actively growing culture of the organism concerned. Conflation of this term with "Dormancy" implies on the one hand that the persisting cells must have been dormant and on the other that dormancy and persistence represent the same physiological state in bacteria. This difficulty resurfaces later when they define dormancy but other problems emerge before then."*

This is entirely fair; we see that we occasionally elided the terms 'dormancy' and 'persistence' to imply synonymy, when either there is none or at least there is no evidence for it. We think the best solution is to add a little section pointing out the semantic difficulties, repeating the operational nature of the definitions, and specifying that in very few cases do we actually know the true physiological state of individual cells – which is what matters with regard to replicatory potential. This material mainly appears in the section defining dormancy, and its title has been extended to note the semantic issues.

- *"I was next concerned by the extensive use of the term "Differentiation". I completely agree*

that what we used to think of as uniform bacterial populations are probably never so but the degree to which subpopulations may be considered differentiated rather than reflecting a range of adaptive responses or indeed, some degree of injury, is not considered here and again I think this leads to problems in considering their hypotheses under a unitary banner downstream. I consider differentiation to require phenotypic changes that are not directly reversible, as in the case of sporulation, whereas adaptation can involve expression of a single gene that can be reversed by its subsequent repression. I do agree that cell cycle contributes to the range of phenotypes in a pure bacterial culture and that this is not the only reason for their diversity (but was not enlightened by use of the term "modulo" in this regard)."

We mainly agree, and suggest what we think is a useful clarification or extension. We note again that "reversibility" is established post hoc, but there are at least two meanings involved. At one level we are discussing a reversibility of states. Let us take a spore and a vegetative cell, which obviously, for sporulating bacteria, can indeed interconvert ("reversibly"). However, another level or meaning implies a mechanistic reversibility, i.e. the path from A to B is simply traversed in the opposite direction when B reverts or interconverts to A. Not only is this not what we mean but (also for thermodynamic reasons) it is certainly not what is done (sporulation and germination in *B. subtilis* are definitely quite separate processes, as indicated by the referee, and one is not at all the reverse of the other). We have added clarificatory comments accordingly. (One might also have added, but we have not in the ms as it would distract, that similar issues apply to the 'reversibility' of enzymes and of biochemical pathways (gluconeogenesis is not mechanistically a reversal of glycolysis, even if the "start" and "end" states are the same molecules.)

- *"The operational definition of dormancy given deliberately leaves open the possibility of metabolic activity and seems only to require that the cell so defined should not divide; this did not allow me to recognise which operational tests might be applied to enumerate or detect dormant cells. Subsequently the detection of molecular signals indicative of bacterial presence in samples from which they were not isolated in culture is taken as evidence of dormancy. In the first case do we accept any non-dividing cell as dormant and in the second I can (and will) offer multiple alternate explanations other than dormancy. Moreover, returning briefly to the conflation between dormancy and persisters, the recent work of John McKinney and colleagues shows that antibiotic exposed persisting cells are not necessarily non-dividing cells in the mycobacterial system he studied."*

The hallmark of the dormant macrostate, stated in quotation marks in the second paragraph of the 'dormancy' section, is indeed that the cells in question do not immediately grow when attempts to culture them under "suitable" conditions (that normally admit their growth), are often (but not necessarily) of low metabolic activity, but are not operationally dead since they can be resuscitated. On this basis we think that this should allow the referee or anyone else to determine the operational tests. It follows that we do not accept 'any' non-dividing cell as dormant since only resuscitable cells can – *post hoc* – be considered dormant, and certainly a non-dividing cell it may be irreversibly injured or operationally dead. However, the presence of molecular signals (e.g. 16S) in samples from which nothing (or many fewer colonies or OTUs)

may be recovered by culture is certainly an indication of the possibility of resuscitation, and hence dormancy.

The referee is entirely correct that we had missed John McKinney's recent and very relevant work, and we mention it accordingly.

- *"Alternative interpretations of the presence of bacterial 16SrDNA sequences in blood when culture fails to detect the organisms from which they derive, include the presence of dead, injured or moribund cells. If they are shown to be repeatedly present then they must either be able to persist in the face of clearance mechanisms or be supplied at a rate equal to their clearance; both seem equally plausible to the dormancy explanation to me. Moreover, why the first three explanations offered for "Not-yet-cultured" should apply to environmental bacteriology but not to clinical samples escapes me."*

The referee is entirely correct with regard to the last sentence, and the whole point (or at least a major theme) of our review is precisely that what is well established in environmental microbiology has had much less impact in clinical microbiology (referee 2 makes this exact point, even more explicitly). We agree that in a steady state such cells must be supplied at a rate equal to that of their clearance, and that the fact that clearance is lower than probably expected implies a significant ability to evade the innate and adaptive immune systems. We also take it that for common organisms (not very slow growers such as certain mycobacteria) the former rates must be much lower than those typically attainable in laboratory cultures, else we would have classical sepsis. We have added a few comments on these issues accordingly, in the section entitled 'Generalised failure of classical techniques to detect dormant bacteria in clinical microbiology'.

- *"I am led to the conclusion that the authors have chosen to label evidence for discrepancies between culture and nucleic acid detection of bacteria in blood to give their hypotheses a simple headline. I have no problem with the proposal that human blood and tissues classically considered sterile in the absence of overt symptoms of infection are frequently exposed to bacteria and bacterial products that in many cases contribute to serious chronic disease. However, I consider the burden of available evidence currently provides many potential explanations within the field of microbiomics/metagenomics in contrast to the dormancy hypothesis offered here. Further, I feel this broad application of dormancy to bacterial phenotypes which, even in the case of Rpf dependency, have not been shown to result from a programme of gene expression that could be considered as differentiation, diminishes the value of the term. Indeed there remains no direct proof that dormancy of *Mycobacterium tuberculosis* underpins what we call latent tuberculosis infection and it is not essential to the observed clinical or pathological pattern, notwithstanding the widespread acceptance of this view by most researches, including me."*

I am not fundamentally opposed to the ideas presented by Kell and colleagues but I do not think they are assisted by lack of attention to the contradictions I have identified above."

All of the above is entirely fair, and we do not disagree. We hope that the changes we have now made to the ms to weaken the ostensible claims (and misplaced synonymies) now meet the referee's approval. For instance we have stressed that

while the presence of suitable molecular sequences (e.g. 16S) implies that it is worth seeking to resuscitate the organisms from which it came, an absence would imply that it is not. A success in resuscitating organisms from a sample that initially appeared sterile would from our operational definition imply that those ones were indeed dormant, and we'd like to think that this had now been clarified.

- *"Finally I come to the iron dysregulation hypothesis and its pro-inflammatory consequences. It is beyond my expertise to comment on the plausibility of the inorganic chemistry deployed here or to review the evidence relating to more than a fraction of the conditions listed. The importance of the struggle between pathogens and host for access to iron is beyond question. When I entered the medical field of infectious disease it was fully recognised that depriving bacteria from iron was a potential therapeutic angle and indeed iron chelation was studied. Desferrioxamine, a widely used agent in iron overload, was investigated and found to effectively deliver iron to the pathogen and the approach was set aside. More recently this agent has been identified as a major risk factor in serious fungal infection and guidance specifically recommends its avoidance. Newer agents seem not to suffer from this problem and the approach deserves renewed attention. However, I would not underestimate the ability of pathogens to outwit our pharmaceutical industry in the battle to sequester iron. While there are reasons beyond the host-pathogen tug-of-war for iron to consider chelation as a therapeutic option, the potential for adverse effects is significant and I think the suggestion that omission of iron chelation from recent guidance on sepsis management is "shocking" is not justified."*

The point about desferrioxamine is well made (and we mention it, with citations), but the molecule is of course in fact a natural prokaryotic siderophore, from *Streptomyces pilosus*. We have replaced the term 'shocking' with something more suitable.

- *"Focussing briefly on the specific diseases cited and their relation to bacterial exposure in one form or another, I find that evidence cited frequently rests on what can be considered "fringe" hypotheses that have little currency in their respective fields. This is not to discourage their continued pursuit but it does weaken the strength of the authors' argument when investigation of the supporting literature frequently leads to papers that are given little credence in the specialist field. Of course "cave Helicobacter" must remain on the table. But there, an accidental technical breakthrough led to an avalanche of convincing laboratory and clinical data."*

It is probably a philosophical distraction to rehearse how often in science something outside the mainstream is blocked for many years by 'vested interests'. However, we may as well mention Peyton Rous, whose discovery of a viral cause of certain cancers was sidelined for decades (he received a Nobel prize when he was 87, 40 years after first being nominated https://en.wikipedia.org/wiki/Francis_Peyton_Rous!). Closer to (prokaryotic) home, Barry Marshall has edited a book (Marshall BJ (ed.): *Helicobacter pioneers: firsthand accounts from the scientists who discovered helicobacters*. Melbourne: Blackwell, 2002.) whose invited contributors had all long recognised a bacterial cause of ulcers and treated their patients accordingly, on the simple grounds that the antibiotics worked! Of course Marshall and Warren (and the wider world) knew nothing of this at the time of their discovery of *H. pylori*. Under these

circumstances (as here) we rely on the overall weight of evidence (as much as its place of publication) to support our views. In Philosophy of Science circles this bolstering of a view via overlapping circles of self-consistent reasoning and data is referred to as 'coherence'. Accordingly, in this sense, we have tried to make this a coherent story, and rehearse this point in the concluding section.

- *"In summary Kell, Potgieter and Pretorius have produced an interesting read which bring many important ideas to our attention. I am not convinced of the breadth of conditions to which they argue their ideas are applicable and I await with interest, demonstration of how they may be practically pursued and some selected definitive proofs that iron-driven inflammatory disease is as important as they claim."*

We have no further comments at this stage. Many thanks again for a very thoughtful review.

Competing Interests: No competing interests were disclosed.

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