

RESEARCH

Open Access



Induction of targeted, heritable mutations in barley and *Brassica oleracea* using RNA-guided Cas9 nuclease

Tom Lawrenson^{1†}, Oluwaseyi Shorinola^{1†}, Nicola Stacey¹, Chengdao Li², Lars Østergaard¹, Nicola Patron³, Cristobal Uauy^{1*} and Wendy Harwood^{1*}

Abstract

Background: The RNA-guided Cas9 system represents a flexible approach for genome editing in plants. This method can create specific mutations that knock-out or alter target gene function. It provides a valuable tool for plant research and offers opportunities for crop improvement.

Results: We investigate the use and target specificity requirements of RNA-guided Cas9 genome editing in barley (*Hordeum vulgare*) and *Brassica oleracea* by targeting multicopy genes. In barley, we target two copies of *HvPM19* and observe Cas9-induced mutations in the first generation of 23 % and 10 % of the lines, respectively. In *B. oleracea*, targeting of *BoI.C.GA4.a* leads to Cas9-induced mutations in 10 % of first generation plants screened. In addition, a phenotypic screen identifies T₀ plants with the expected dwarf phenotype associated with knock-out of the target gene. In both barley and *B. oleracea* stable Cas9-induced mutations are transmitted to T₂ plants independently of the T-DNA construct. We observe off-target activity in both species, despite the presence of at least one mismatch between the single guide RNA and the non-target gene sequences. In barley, a transgene-free plant has concurrent mutations in the target and non-target copies of *HvPM19*.

Conclusions: We demonstrate the use of RNA-guided Cas9 to generate mutations in target genes of both barley and *B. oleracea* and show stable transmission of these mutations thus establishing the potential for rapid characterisation of gene function in these species. In addition, the off-target effects reported offer both potential difficulties and specific opportunities to target members of multigene families in crops.

Keywords: Genome editing, CRISPR/Cas9, Barley, *Brassica*, *PM19*, *GA4*, Crops, Mutations, Breeding, Off-target

Background

Genetic modification is a key research tool for advancing knowledge of gene function as well as allowing the development of crops with valuable traits. Genetic modification enables the introduction of genes of interest or the reduction in expression of endogenous genes (RNAi approaches) through the insertion of transgenic sequences at random locations within the plant genome. Genetic modification technologies have advanced substantially over the past 30 years, but more recently, a series of exciting developments offer significant opportunities for the

analyses of plant genomes, as well as having applications in crop improvement [1]. These approaches, collectively called genome editing, provide the opportunity to make precise changes at specific genomic locations. Genome editing may be used to induce gene insertions, gene replacements, or insertions or deletions that disrupt the function of a specific gene [2]. This latter application, leading to knock-out of target genes, has enormous benefits for research in plants, especially in crops that lack genetic resources such as knock-out libraries.

Genome editing requires a site-directed nuclease to introduce one or more breaks in the DNA at the target locus. The cell's endogenous DNA repair mechanisms repair these breaks; imperfect repair can produce mutations or deletions in the genes of interest. To generate

* Correspondence: cristobal.uauy@jic.ac.uk; wendy.harwood@jic.ac.uk

†Equal contributors

¹John Innes Centre, Norwich Research Park, Colney NR4 7UH, UK
Full list of author information is available at the end of the article

site-specific breaks, different approaches have employed different combinations of nucleases fused to programmable DNA binding domains including Zinc Finger Nucleases (ZFNs) and Transcription-Activator Like Effector Nucleases (TALENs). More recently, the Cas9 protein associated with Type II Clustered Regulatory Interspaced Short Palindromic Repeats (CRISPR) found in bacteria has been repurposed for genome editing in eukaryotes [3, 4]. The RNA-guided Cas9 system uses a small non-coding RNA, known as the single guide RNA (sgRNA), to direct the Cas9 nuclease to the DNA target of interest. Being small and easy to reprogram, this offers a flexible, easy-to-implement and relatively cheap method for genome editing [5]. The first applications of RNA-guided Cas9 in plants were described in 2013 [6–8] using transient systems. Inheritance of induced mutations in progeny plants was demonstrated for the first time in *Arabidopsis* by Feng *et al.* [9] and heritable changes have also been shown in rice [10, 11]. In wheat (*Triticum aestivum*), RNA-guided Cas9 has been used to mutate a single homoeologue of the mildew resistance locus *MLO* in stable T₀ transgenic plants although no information was provided regarding the inheritance of the mutant alleles [12].

Very few studies have described the inheritance of RNA-guided Cas9-induced mutations and questions remain regarding its efficiency, especially in crop plants. In addition, the frequency with which the nuclease induces mutations in unintended targets (known as off-targets) has yet to be extensively investigated across plant species. The aim of this study was therefore to use RNA-guided Cas9 for targeted mutagenesis in both monocotyledonous and dicotyledonous crop species, demonstrating for the first time its application in both barley (*Hordeum vulgare*) and *Brassica oleracea*. In addition, we aimed to assess the efficiency of mutagenesis and test whether off-target effects occurred.

Arabidopsis GA4 is involved in the gibberellin biosynthesis pathway and *GA4* loss-of-function mutants have dwarf stature and reduced fruit dehiscence [13, 14]. Since plant architecture and seed dispersal are important targets for crop improvement in Brassicas, we tested the effect of mutating *GA4* orthologues in *B. oleracea*. In barley, we chose *HvPM19* as our target. *HvPM19* encodes an ABA-inducible plasma membrane protein [15], which in wheat acts as a positive regulator of grain dormancy [16], an important agronomic trait in cereals.

Here we demonstrate the successful use of RNA-guided Cas9 genome editing to knockout the function of target genes in both barley and *B. oleracea*. We show transmission of the mutation to progeny plants in both species and we demonstrate the segregation of the transgenic locus (encoding the nuclease and sgRNA) from the mutation, resulting in transgene-free plants that contain the desired mutation.

Results and discussion

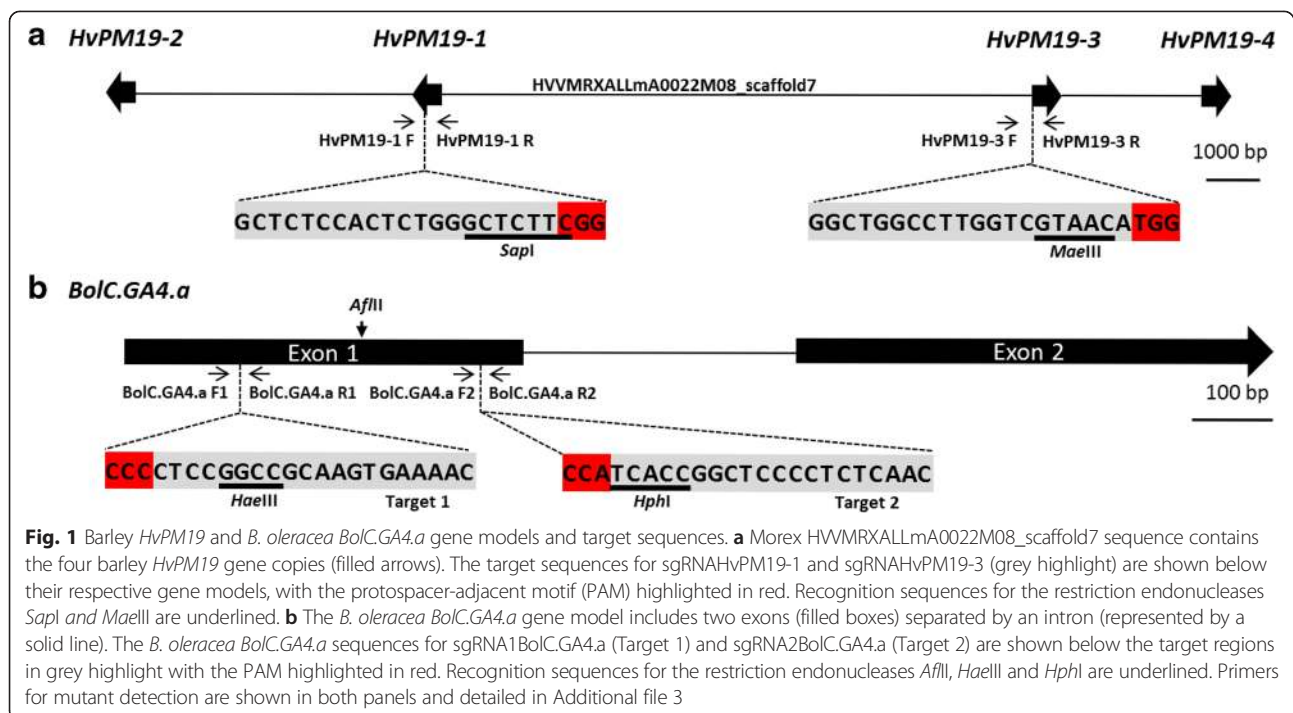
RNA-guided Cas9-induced genome editing in barley

We investigated the use and target specificity requirements of RNA-guided Cas9 genome editing in barley by focusing on a multi-copy gene. We selected *HvPM19*, which is present as four copies within a single barley BAC clone from the cultivar ‘Morex’ (*HvPM19-1* to *HvPM19-4*; Fig. 1a). Relative to *HvPM19-1*, the *HvPM19-2*, *HvPM19-3* and *HvPM19-4* loci have sequence identities of 89.8 %, 89.5 % and 88.6 %, respectively, whereas *HvPM19-3* and *HvPM19-4* have greater sequence identity to *HvPM19-2* (98.4 % and 99.6 %). This suggests that *HvPM19-1* was involved in the more ancestral duplication event and that there was a series of very recent duplication events between *HvPM19-2*, *HvPM19-3* and *HvPM19-4*.

We independently targeted two ancestral *HvPM19* gene copies (*HvPM19-1* and *HvPM19-3*) in the spring barley cultivar ‘Golden Promise’ which is amenable to *Agrobacterium*-mediated transformation. We were able to amplify *HvPM19-4* from Golden Promise, but unable to amplify *HvPM19-2* suggesting that this cultivar lacks this copy of *HvPM19*. Two binary constructs, sgRNA_{HvPM19-1}, referred to as pPM19-1 and sgRNA_{HvPM19-3}, referred to as pPM19-3 (Fig. 2a), were designed to independently target *HvPM19-1* and *HvPM19-3*, respectively. The 20 base-pair target sequence in pPM19-1 has a single nucleotide mismatch with each of the corresponding sequences in *HvPM19-3* and *HvPM19-4*, while the target sequence in pPM19-3 has three mismatches with *HvPM19-1* and one mismatch with *HvPM19-4* (Fig. 3a).

The two constructs were independently transformed into immature barley embryos to generate 28 and 20 independent transgenic lines for pPM19-1 and pPM19-3, respectively. T₀ regenerated plantlets were screened for mutations using a restriction digest/PCR assay. We detected deletions in *HvPM19-1* in three out of 13 pPM19-1 T₀ lines screened (T₀-181, T₀-122 and T₀-191). Similarly, out of the 10 pPM19-3 T₀ plantlets screened, one line (T₀-211) showed an insertion in *HvPM19-3*. Therefore, the frequency of Cas9-induced mutations in the first generation was 23 % for pPM19-1 and 10 % for pPM19-3. These mutation frequencies are comparable to those reported in stable T₀ transformants from other monocotyledonous species such as wheat [12], rice (*Oryza sativa*; reviewed in [17]) and sorghum (*Sorghum bicolor*; [18]).

As is characteristic of Cas9-induced mutations [3, 4, 19], all the insertions or deletions (in-dels) detected were at the 3' end of the target region, 3 or 4 bp upstream of the Protospacer Adjacent Motif (PAM; Additional file 1). In the T₀ plants, we detected in-dels only after enriching for the mutation by the restriction digest/PCR assay presumably because only a small proportion of the cells had

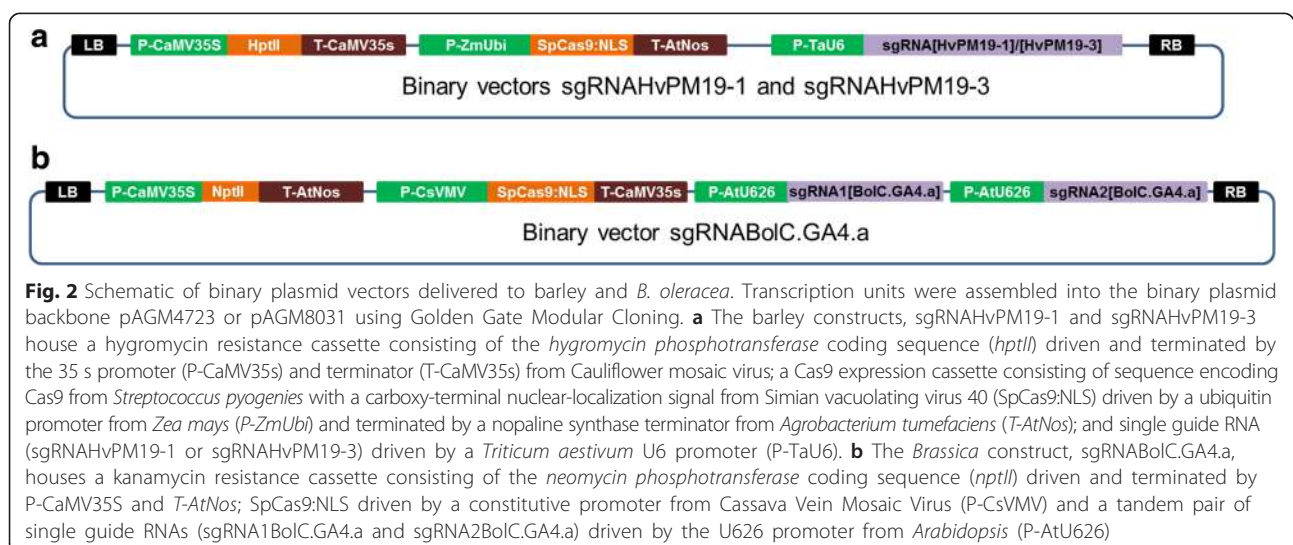


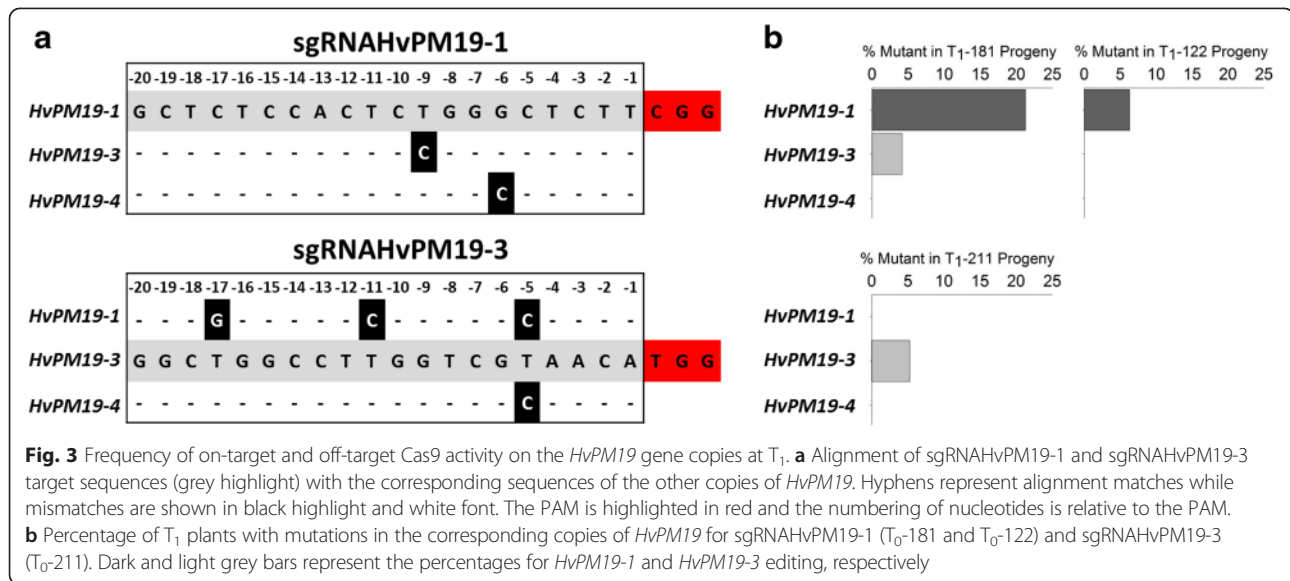
been mutated at the time of sampling. To further characterise the events, we measured T-DNA copy number in the T₀ lines and found that T₀-181, T₀-191 and T₀-211 each contained a single copy of the T-DNA whereas T₀-122 contained two copies.

On-target and off-target activity of RNA-guided Cas9 in T₁ transgenic barley plants

Next, we examined the T₁ progenies of T₀-181, T₀-122, T₀-191 and T₀-211. Twenty out of 93 T₁ progenies of T₀-181 contained in-dels in the target *HvPM19-1*

sequence as determined by Sanger sequencing. Of these, two were homozygous and 18 were heterozygous mutants determined by the presence of double peaks in the sequencing chromatogram beginning from the site of the in-del. For T₀-122, only six out of 95 T₁ progenies had in-dels in the target *HvPM19-1* sequence, with all being heterozygous mutations. This represents mutation frequencies of 22 % in T₀-181 and 6 % in T₀-122 progenies. For line T₀-211, which showed Cas9 activity for *HvPM19-3*, we detected four mutant plants out of 76 T₁ progenies tested, all of which had heterozygous





mutations (mutation frequency of 5 %). Line T₀-191 showed mutations in seven out of 90 T₁ plants, but was not analysed further. As in the T₀ generation, all the indels were observed to occur in the 3–4 bp adjacent to the PAM. All the T₁ plants with Cas9-induced mutations retained their corresponding T-DNA construct, while there was segregation in the non-mutated T₁ plants. This indicated that the mutations could still be the product of sgRNA/Cas9 expression in somatic cells rather than due to germline inheritance.

To assess the specificity of the T-DNA constructs, we sequenced *HvPM19-3* and *HvPM19-4* in the progenies of T₀-181 and T₀-122 (designed to target *HvPM19-1*). We found no off-target activity in the T₁ progenies of T₀-122; whereas three T₀-181 progeny from 72 tested (4.2 %) had off-target activity on *HvPM19-3* (Fig. 3b). By contrast, we observed no off-target activity on *HvPM19-1* and *HvPM19-4* in the 73 T₁ progenies of T₀-211 that contained the T-DNA designed to target *HvPM19-3*.

Cas9-induced mutations are stably transmitted to T₂ barley plants independently of the T-DNA construct

The mutation in the target gene theoretically should segregate independently of the T-DNA that encodes the nuclease and sgRNA. We observed complete co-segregation of the Cas9-induced mutations with the T-DNA construct in the T₁ transgenic lines. We therefore tested the T₂ progenies of several T₁ lines to determine if the mutations could be stably inherited and segregate independently from the T-DNA construct. We screened for the presence of the T-DNA through PCR and qPCR assays and determined the mutation status in the T₂ progenies of T₁-181, T₁-122 and T₁-211 lines (T₁ mutant lines originating from the corresponding T₀ lines; Table 1). The T-DNA

segregated in the progeny of some, but not all, of these T₁ lines. Segregation of pPM19-1 was detected in 11 out of 19 T₁-181 lines, whereas pPM19-1 segregated in the progeny of four out of six T₁-122 lines. However, only one out of three T₁-211 lines tested showed segregation of the pPM19-3 construct. A 3:1 presence:absence ratio was confirmed using a χ^2 test in all progenies in which the T-DNA segregated ($P > 0.44$ or higher).

We next sequenced *HvPM19-1* and *HvPM19-3* from all 45 T₂ plants that had not inherited the T-DNA. In these plants, we detected mutations in *HvPM19-1* in 15 T₂ progenies originating from seven independent T₁-181 lines and seven T₂ progenies originating from three independent T₁-122 lines (Table 1). A single progeny of T₁-181_H2 showed an off-target mutation in *HvPM19-3* in addition to the on-target *HvPM19-1* mutation. Interestingly, T₁-181_H2 is one of the three T₀-181 lines that showed off-target activity of pPM19-1 in the T₁ generation. For pPM19-3, we did not detect mutations in *HvPM19-3* (and *HvPM19-1*) in the absence of the T-DNA in the progenies of any of the T₁-211 lines.

We found that the mutations detected in the T₂ progenies matched those observed in the corresponding T₁ parent in all cases examined. For instance, the homozygous 1-bp deletion observed in line T₁-181_E1 was also present in all its T₂ progenies that segregated away from the T-DNA construct (Table 1; Fig. 4a). Likewise, a range of mutations found in heterozygous T₁ plants were also identified in homozygous T₂ individuals in the absence of the T-DNA construct (Fig. 4b). Taken together, the T₁ and T₂ sequence data from six lines, originating from two independent T₀ events (T₀-181 and T₀-122), provide strong evidence of stable germline transmission

Table 1 Summary of transgenerational RNA-guided Cas9 activity and segregation in barley

T-DNA Construct	T ₀ line	T ₁ mutation type	T ₁ line	Number of T ₂ plants screened for T-DNA	Plants without T-DNA	Plants without T-DNA and with (homozygous/heterozygous) mutations ^a
pPM19-1	T ₀ -181	Homozygous	T ₁ -181_B5	4	0	-
			T ₁ -181_E1	12	5	5/0
			T ₁ -181_A11	12	3	1/1
		Heterozygous	T ₁ -181_B1	8	2	0/0
			T ₁ -181_B8	11	0	-
			T ₁ -181_C1	9	0	-
			T ₁ -181_C12	1	0	-
			T ₁ -181_C3	12	0	-
			T ₁ -181_C4	12	1	0/0
			T ₁ -181_C9	11	1	0/1
			T ₁ -181_D11	12	4	1/1
			T ₁ -181_D2	12	3	2/0
			T ₁ -181_D9	2	0	-
	T ₁ -181_E4	12	0	-		
	T ₁ -181_G4	11	3	0/1		
	T ₁ -181_G5	12	1	0/0		
	T ₁ -181_H2	12	3	1/1		
	T ₁ -181_H5	12	0	-		
	T ₁ -181_H9	12	4	0/0		
	T ₀ -122	Heterozygous	T ₁ -122_B11	12	4	2/1
T ₁ -122_C6			12	1	0/1	
T ₁ -122_F12			12	4	0/3	
T ₁ -122_H2			12	0	-	
T ₁ -122_H4			12	3	0/0	
pPM19-3	T ₀ -211	Heterozygous	T ₁ -211_B11	12	3	0/0
			T ₁ -211_D10	12	0	-
			T ₁ -211_G4	7	0	-

^aHyphens (-) indicate that all plants had presence of the T-DNA construct, and thus were not tested

of Cas9-induced mutations in barley in the absence of the T-DNA.

The ability to develop transgene-free and stable germline mutations is of considerable interest in crop species given the current regulatory framework for deployment of transgenic crops in the field. Although regulation of edited crops is still being debated [20], crops free of transgenes may not be subject to existing regulations on genetic modification. Here, we demonstrate that in several instances there was stable germline-transmitted inheritance of Cas9-induced mutations in barley from the T₁ to the T₂ generation in the absence of the T-DNA construct. This supports previous studies in plants (*Arabidopsis*, tomato, tobacco and rice) that have documented transgene-free inheritance of Cas9-induced mutations in the T₁ and T₂ generations. No description of

germline inheritance has been previously reported for *Triticeae* [12]. We also identified a single plant with an off-target mutation in the T₂ generation in the absence of the T-DNA construct. This plant had mutations in both *HvPM19-1* and *HvPM19-3*, suggesting that tandemly duplicated genes can be knocked-out with a single sgRNA, although we have yet to establish if these mutations are in *cis* or on homologous chromosomes. Previous work in rice had identified off-target mutations only in the T₁ generation and in the presence of the sgRNA/Cas9 construct [21].

RNA-guided Cas9 induced genome editing in *Brassica oleracea*

RNA-guided Cas9-induced genome editing was performed in *B. oleracea* DH1012 [22] by targeting *BolC.GA4.a*

a		
<i>HvPm19-1</i> Wild type Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCTCTTTCGAAACCAGTTTGGTAA	
T ₁ -181-E1 Mutant Allele:	CTGTACGTGCTTGCCTCCACTCTGGGC-CTTCGAAACCAGTTTGGTAA	
T ₂ -181-E1-p3, p5, p6, p9 and p10:	CTGTACGTGCTTGCCTCCACTCTGGGC-CTTCGAAACCAGTTTGGTAA	
b		
T ₁ -181-A11 WT Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCT-CTTTCGAAACCAGTTTGGTAA	x13
T ₁ -181-A11 Mutant Allele 1:	CTGTACGTGCTTGCCTCCACTCTGGGCTTCTTCGAAACCAGTTTGGTAA	x2
T ₁ -181-A11 Mutant Allele 2:	CTGTACGTGCTTGCCTCCACTCTGGGC---TTCGAAACCAGTTTGGTAA	x2
T ₂ -181-A11-p11:	CTGTACGTGCTTGCCTCCACTCTGGGC---TTCGAAACCAGTTTGGTAA	
T ₁ -181-D2 WT Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCTCTTTCGAAACCAGTTTGGTAA	x10
T ₁ -181-D2 Mutant Allele:	CTGTACGTGCTTGCCTCCACTCTGGGC-CTTCGAAACCAGTTTGGTAA	x5
T ₂ -181-D2-p11 and p12:	CTGTACGTGCTTGCCTCCACTCTGGGC-CTTCGAAACCAGTTTGGTAA	
T ₁ -181-D11 WT Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCT-CTTTCGAAACCAGTTTGGTAA	x7
T ₁ -181-D11 Mutant Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCTTCTTCGAAACCAGTTTGGTAA	x8
T ₂ -181-D11-p7:	CTGTACGTGCTTGCCTCCACTCTGGGCTTCTTCGAAACCAGTTTGGTAA	
T ₁ -181-H2 WT Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCT-CTTTCGAAACCAGTTTGGTAA	x8
T ₁ -181-H2 Mutant Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCTACTTCGAAACCAGTTTGGTAA	x5
T ₂ -181-H2-p9:	CTGTACGTGCTTGCCTCCACTCTGGGCTACTTCGAAACCAGTTTGGTAA	
T ₁ -122-B11 WT Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCT-CTTTCGAAACCAGTTTGGTAA	x10
T ₁ -122-B11 Mutant Allele:	CTGTACGTGCTTGCCTCCACTCTGGGCTTCTTCGAAACCAGTTTGGTAA	x5
T ₂ -122-B11-p8 and p12:	CTGTACGTGCTTGCCTCCACTCTGGGCTTCTTCGAAACCAGTTTGGTAA	
c		
<i>BolC.GA4.a</i> Wild type Allele:	GTGGTCCGAAGGTTTTCACCCGGCTCCCCTCTCAACGACTTCCGTAA	
T ₁ _L2F1_E_B6:	GTGGTCCGAAGGTTTACCATCA-----CCCCTCTCAACGACTTCCGTAA	
T ₂ -L2F1-E_B6_p6 and p7:	GTGGTCCGAAGGTTTACCATCA-----CCCCTCTCAACGACTTCCGTAA	
T ₁ _L2F1_E_C7:	GTGGTCCGAAGGTTTACCATCA-----CCCCTCTCAACGACTTCCGTAA	
T ₂ -L2F1-E_C7_p1, p4, p9 and p10:	GTGGTCCGAAGGTTTACCATCA-----CCCCTCTCAACGACTTCCGTAA	
T ₁ _L2F1_E_D8:	GTGGTCCGAAGGTTTACCATCA-----CCCCTCTCAACGACTTCCGTAA	
T ₂ -L2F1-E_D8_p2, p7 and p9:	GTGGTCCGAAGGTTTACCATCA-----CCCCTCTCAACGACTTCCGTAA	

Fig. 4 Germline transmission of Cas9 induced mutations from T₁ to T₂ plants in barley and *B. oleracea* in the absence of the T-DNA construct. **a** Sequence alignment of T₁-181_E1 and five homozygous T₂ progeny with homozygous 1-bp deletion in *HvPM19_1*. **b** Sequence alignment from representative clones of T₁ heterozygote mutants and direct Sanger sequencing of their T₂ progeny with homozygous mutations in the absence of the T-DNA. The numbers of clones supporting T₁ mutant alleles are indicated on the right. **c** Sequence alignments of *BolC.GA4.a* Target 2 in homozygous T₁ and T-DNA free T₂ plants. Across panels the target sequences for sgRNAHvPM19-1 and sgRNABolC.GA4.a (grey) and PAM (red) are highlighted and Cas9 induced insertions and deletions are indicated by red font or red hyphens, respectively. Names of homozygous T₂ plants that lack the presence of the T-DNA construct are indicated in blue; individual homozygous plants with the same allele are shown on the same row and are labelled with a 'p' prefix

(*Bol038154*) located on chromosome 5. This gene is an orthologue of *Arabidopsis GA4* which encodes AtGA3OX1, the last enzyme in the biosynthesis of bioactive gibberellins. In *Arabidopsis*, *ga4* loss-of-function mutants show a semi-dwarf phenotype [23] and this gene is required for efficient seed dispersal as it promotes specification of the dehiscence zone in siliques [13]. *BolC.GA4.a* has a paralog on chromosome 8, designated *BolC.GA4.b* (*Bol031570*), which shares 90 % DNA sequence identity. To generate Cas9 induced

mutations in *BolC.GA4.a*, we designed a binary construct containing two sgRNAs (sgRNA1BolC.GA4.a and sgRNA2-BolC.GA4.a) that target separate regions (Target 1 and Target 2, respectively) in the first exon of *BolC.GA4.a* (Figs. 1b and 2b).

Eighty independent transgenic lines were generated by *Agrobacterium*-mediated transformation, and 20 of these T₀ plantlets were screened by the restriction digest/PCR assay to detect mutations in the target sequences. We

identified in-dels at the target sites in *BolC.GA4.a* in two out of 20 T₀ lines (L2F1_8.2 and L2E1_17.1). Mutations in L2E1_17.1 were confirmed by TA cloning and Sanger sequencing of the PCR products (Fig. 5a). Line L2F1_8.2 showed a 282-bp deletion that corresponds to re-joining of the DNA at exactly 3 bp from the PAM in both target regions. As in barley, the detection of the mutations required an enrichment of the target by restriction digest prior to PCR.

We also hypothesised that plants with homozygous mutations in *BolC.GA4.a* would show a dwarf phenotype similar to that observed in *Arabidopsis ga4* mutants. Therefore, we performed a phenotypic screen of the 80 T₀ *B. oleracea* plants. All 80 T₀ lines were grown to maturity, and at flowering two lines not previously characterised by the restriction digest/PCR assay were observed to be dwarf in stature (L2F1_A and L2F1_E; Fig. 6a). The *BolC.GA4.a* sequences from both dwarf plants were found to contain a series of mutant alleles in Target 1 and Target 2, in two independent leaf samples from each plant (Fig. 5b, Additional file 2). In addition, the mutation was restricted to *BolC.GA4.a*, as we were unable to detect any mutation in *BolC.GA4.b*. The identification of T₀ plants with a visible knockout phenotype has also been reported in rice, tomato, and *Arabidopsis* [8, 10, 24].

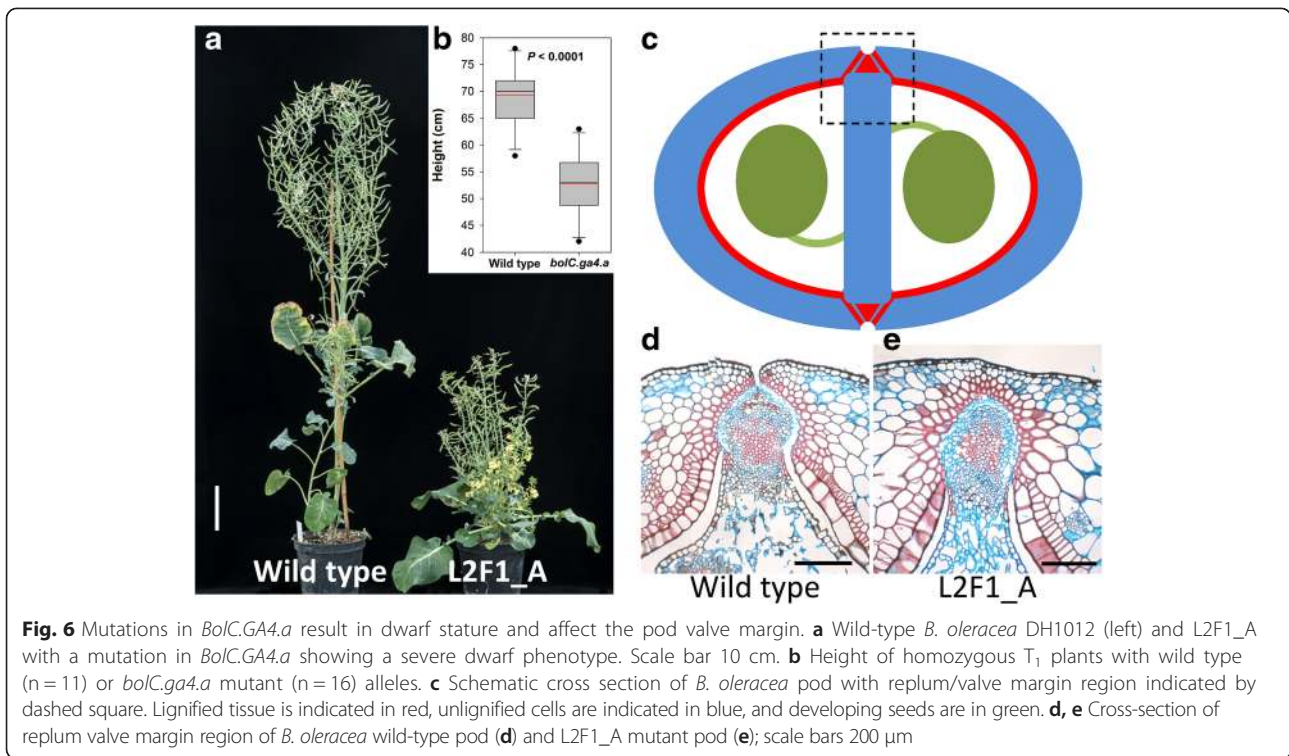
The 80 T₀ *B. oleracea* plants described above originated from the same transformation experiment, but differed in their culture period. A first batch of 41 T₀ shoots was isolated four weeks after *Agrobacterium*

inoculation, whereas a second batch of 39 T₀ shoots was isolated 7 weeks after inoculation. Both dwarf lines were derived from the 7-week batch, supporting a recent report in rice [25] that obtained an increased proportion and variety of mutated cells by extending the culture period of rice calli by 4 weeks. Across different target genes, Mikami *et al.* [25] found a 3.7-fold increase in mutation frequencies between rice calli cultured for 1 month compared to 2 months. They hypothesize that this is due to a greater chance of inducing novel mutations in non-mutated cells [25]. Our results are consistent with this hypothesis which also implies that shorter selection periods during culture of calli could reduce the number of off-target mutations.

Cas9-induced mutations are stably transmitted to T₂ *B. oleracea* plants independently of the T-DNA construct

To examine the mutation frequency of the target locus *BolC.GA4.a*, the T₁ progenies of lines L2F1_8.2 and L2E1_17.1 were screened for Cas9-induced mutations in Target 1 and 2 by PCR amplification of *BolC.GA4.a* followed by direct sequencing. Using the sequencing chromatograms it was possible to identify homozygous and heterozygous mutations. We detected mutations in the T₁ progenies of L2F1_8.2, but not in L2E1_17.1. Heterozygous in-dels were observed in 68 of 90 L2F1_8.2 T₁ progenies; however, no homozygous mutations were identified. Of these 68 T₁ plants, 35 had mutations in Target 1, whereas Target 2 was mutated in 67 lines, suggesting a higher efficiency of the Target 2

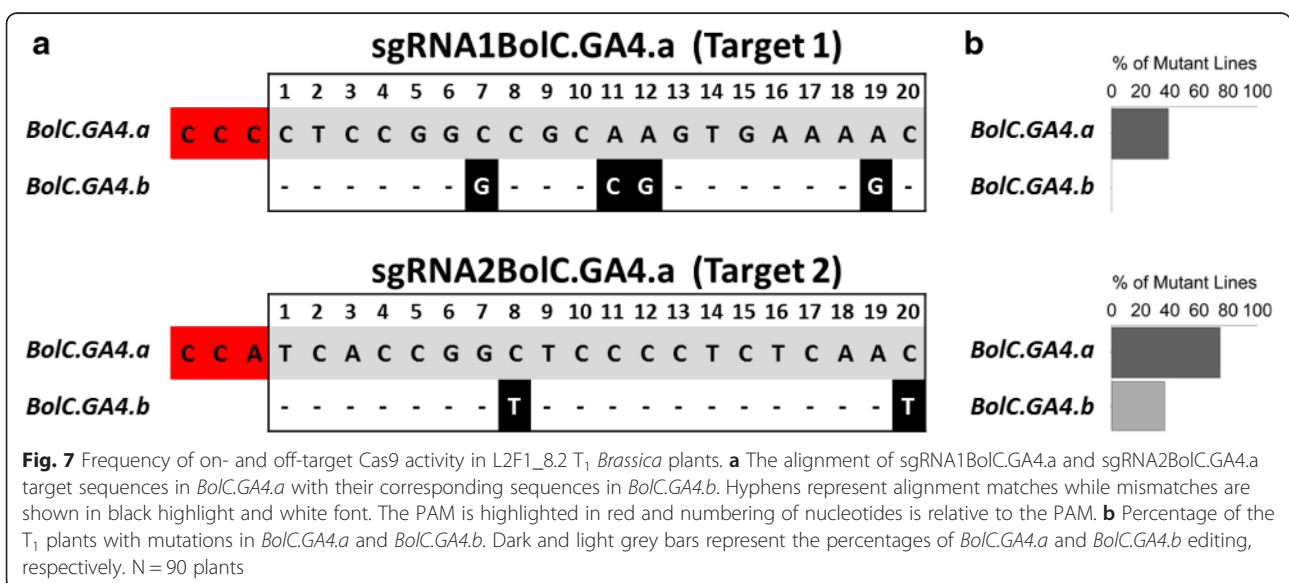




sgRNA (Fig. 7b). None of the 90 T₁ progenies inherited the complete 282-bp deletion between the two *BolC.GA4.a* target regions that was observed in the T₀ generation.

The sgRNA targets were also sequenced in the T₁ progenies of lines L2F1_A and L2F1_E that showed a dwarf phenotype. T₁ plants from each of L2F1_A and L2F1_E were screened and found to carry a range of either homozygous or heterozygous mutations across the target regions in

BolC.GA4.a (Additional file 2). Of the 39 plants screened, 20 had either homozygous mutations or a combination of two mutant alleles previously identified in the T₀ plants; all of these plants displayed the dwarf phenotype at maturity (Fig. 6b). In the remaining 19 heterozygous lines we also identified the same mutations as in the T₀ plants, including the large 68-bp deletion across Target 1 (Fig. 5b; Additional file 2). These results are consistent with stable transmission of Cas9-induced mutant alleles in *B. oleracea*.



To determine germline inheritance, we screened for the presence of the T-DNA construct in 12 individual T_2 plants derived from eight homozygous mutant T_1 lines (96 plants in total). Nine T_2 plants which lacked the T-DNA construct were recovered (Fig. 4c). These plants all carried the same 6-bp deletion in Target 2, and wild-type allele in Target 1, as in the parental T_1 plants (L2F1_E_B6, L2F1_E_C7, and L2F1_E_D8). The fact that the mutations in T_1 plants were stably transmitted to the T_2 generation in the absence of the T-DNA construct supports the germline inheritance of the Cas9-induced mutations in *B. oleracea*.

Off-target activity of RNA-guided Cas9 in T_1 transgenic *B. oleracea* plants

Sequencing of *BolC.GA4.b* in the T_1 progenies of T_0 -L2F1_8.2 revealed off-target activity in 32 out of 88 plants (36.4 %; Fig. 7b). This was restricted to Target 2 where the sgRNA contained two mismatches with *BolC.GA4.b* (Fig. 7a). No mutations were observed across Target 1 where the sgRNA contained four mismatches in the target region compared to *BolC.GA4.b*. In T_0 lines with the dwarf phenotype (T_0 lines L2F1_A and L2F1_E), we detected no off-target activity in *BolC.GA4.b*, indicating that the dwarf phenotype was due to mutations in *BolC.GA4.a* only.

In this study, we show that a single sgRNA (sgRNA2-BolC.GA4.a) can simultaneously target two copies of *GA4* despite the presence of a mismatch between the sgRNA and the *BolC.GA4.b* off-target sequence 8 bp from the PAM (Fig. 7a). This observation mirrors our results in barley, in which off-target activity was detected in *HvPM19-3* due to the sgRNA in pPM19-1, designed to target *HvPM19-1* and which has a mismatch 9 bp from the PAM (Fig. 3). This off-target activity was detected only in the progeny of the *B. oleracea* and barley lines with high on-target mutation frequencies. Given these results and the idea that on-target mutations may precede off-target mutations [26], it is tempting to speculate that higher on-target Cas9 activity positively correlates with higher off-target mutation frequencies.

These results differ from reports in wheat, where a single mismatch 3 bp from the PAM between *MLO* homoeologues limited off-target activity, although on-target mutation frequencies were relatively low (5.6 %; [12]). Previous studies found that a single mismatch within the 12 bp adjacent to the PAM could confer specificity in humans and other systems [3, 27]. However, others [28, 29] have shown that up to two mismatches, as well as small insertions and deletions, are tolerated within this sequence. Taken together, these results suggest that additional work is needed to decipher the key design rules and experimental parameters relating to on- and off-target mutations using the Cas9 system.

The presence of off-target activity can be considered a negative feature of the Cas9 system when specificity is sought. Several approaches have been suggested for the reduction of off-target activity. These include using truncated sgRNAs [30], a pair of Cas9 nickase mutants directed to opposing strands that require a pair of correctly positioned 20 bp DNA targets to produce a DSB [31, 32], and also the fusion of catalytically dead Cas9 (dCas9) to homodomains of a FokI nuclease dimer that will also only produce a DSB when both targets are in correct proximity [33, 34]. However, off-target activity can also be beneficial for targeting gene families [26] or closely related sequences. Our results suggest that a single sgRNA can simultaneously target multiple gene copies facilitating gene functional analysis by overcoming possible redundancy between the closely related sequences [35]. Importantly, we identified an individual transgene-free barley plant that had concurrent heterozygous mutations in the target (*HvPM19-1*) and off-target (*HvPM19-3*) genes. Many crop species are polyploid (for example, wheat, potato), have undergone recent whole-genome duplication events (for example, Brassica, maize; [36]), or have a high number of tandemly duplicated genes [37], such as the *HvPM19* locus investigated in this study. Therefore, the potential to generate progeny with mutations limited to on-target sites, as well as progeny with both on- and off-target mutations, makes the RNA-guided Cas9 system especially relevant for functional analyses in crops.

Mutations in *BolC.GA4.a* affect the pod valve margin

Tissue patterning in the fruits of *Arabidopsis* and members of the Brassica genus is highly similar reflecting their close evolutionary relationship [38]. Seed dispersal in these species depends on formation of valve margin cells along the valve and replum borders that mediate fruit opening [39]. Since valve margins from *Arabidopsis ga4* mutants fail to mediate efficient seed dispersal [13], we tested if the *B. oleracea* Cas9 lines presented here suffered from similar defects. Cross-sections stained with a combination of Safranin O and Alcian Blue revealed that in comparison to wild-type fruits, fruits from L2F1_A failed to pattern the valve margin region properly, such that valve cells replaced the valve margin cells in this line (Fig. 6c-e). As a result, these fruits would disperse their seeds less efficiently than wild type. Although less severe, this phenotype resembled the phenotype observed when another regulator of valve margin formation, *BolC.IND.a*, was downregulated by RNAi [38]. These data therefore demonstrate that the *BolC.GA4.a* function is conserved between *B. oleracea* and *Arabidopsis* and likely regulated in a similar fashion. They also demonstrate the potential for the use of RNA-guided Cas9 to target important traits in *Brassica* crops based on knowledge of gene function from model plants.

Conclusions

In this study, we demonstrate the use of RNA-guided Cas9 to induce targeted mutations in two crop species, *B. oleracea* and barley, and report stable transmission of the mutations across generations. We show that knock-out phenotypes can be recovered as early as the primary T₀ generation, exemplifying the use of this technology for rapid analyses of gene function. We produced transgene-free barley and *B. oleracea* plants with stably-inherited mutations in the target gene, supporting the potential for downstream biotechnological applications. Both species showed off-target activity, despite the presence of at least one mismatch between the sgRNA and the paralogous gene. This led to the identification of a single barley plant with concurrent mutations in the target and off-target gene in the absence of the T-DNA construct. Our results suggest that experimental parameters relating to on- and off-target mutations need to be carefully considered and monitored, and that a single sgRNA has the potential to generate progeny with simultaneous knock-out mutations in paralogous genes. Given that crop genomes commonly contain multiple closely related sequences, the features described herein make RNA-guided Cas9 especially relevant for functional analyses in these species.

Materials and methods

Target locus selection and sgRNA design

Gene sequences for *B. oleracea* *BolC.GA4.a* (*Bol038154*) and barley *HvPm19* (AF218627.1; [15]) were obtained from The Brassica Database [40] and the International Barley Sequencing Consortium [41] databases. For barley, sequence of the BAC clone HVVMRXALL-mA0022M08 from the cultivar 'Morex' was kindly provided pre-publication by Dr Nils Stein (IPK). This BAC was annotated and four copies of *HvPm19* were identified (*HvPM19-1* to *HvPM19-4*). Target sequences that conformed to G(N)₂₀GG were identified on sense and anti-sense strands in the coding sequence for *BolC.GA4.a* and for *HvPM19-1* and *HvPM19-3* and potential off-target sequences were detected via BLAST searches [40, 41]. Potential targets were also evaluated for the presence of non-CpG sensitive restriction site sequences predicted to be disrupted by Cas9 induced in-dels, which also had to be unique within a PCR amplicon. Final target sequences were chosen to be as specific as possible to the intended target sequence (that is, keeping the number of mismatches to off-target sequences high), close to the start codon, and to include an appropriate restriction site (Fig. 1). These targets were checked by PCR and Sanger sequencing (Additional file 3) in the varieties to be transformed (spring barley cultivar 'Golden Promise' and *Brassica oleracea* DH1012) to ensure that no polymorphisms existed between the sgRNA and the target G(N)₂₀GG

sequences. Single sgRNAs were used for barley *HvPM19-1* and *HvPM19-3*, whereas two independent sgRNAs were targeted to the first exon of *Brassica BolC.GA4.a*. Barley 'Golden Promise' sequences for the three *HvPM19* genes were deposited in GenBank (accession numbers KT336449-KT336451).

Construct assembly

The binary plasmid vector constructs were assembled using Golden Gate Modular Cloning (MoClo) [42]. We used Level 0 parts from the Golden Gate MoClo Plant Parts Kit (Addgene kit # 1000000047) and plasmids from Golden Gate MoClo Plant Toolkit (Addgene kit # 1000000044) described in Engler *et al.* [43]. Level 1 transcriptional units were assembled from Level 0 parts and these were subsequently assembled to make the plasmids vectors shown in Fig. 2. A detailed protocol for the assembly of binary vectors with multiple sgRNAs using the Golden Gate MoClo ToolKit and the identity of all plasmids used are given in Additional file 4. Annotated sequences of the plasmids made in this study are provided in Additional file 5 and are available at the non-profit plasmid depository AddGene (<https://www.addgene.org/browse/article/14759/>).

Plant transformation and screening of transgenic material

Barley (*cv.* 'Golden Promise') was transformed by *Agrobacterium tumefaciens*-mediated transformation of immature embryos as described by Harwood [44]. *Brassica oleracea* (DH1012) was transformed by *Agrobacterium tumefaciens* infection of 4-day-old cotyledonary petioles according to Hundleby and Irwin [45].

Primary transgenic T₀ materials were screened using a modified restriction enzyme site loss method [46]. Briefly, for single sgRNA targets, genomic DNA was digested prior to PCR with a CpG-insensitive enzyme to remove wild-type template and thus favour the PCR amplification of mutant DNA where the restriction site had been lost. For *Brassica*, where a pair of sgRNAs was used, an additional screen was implemented; a CpG-insensitive restriction enzyme (*AflIII*) was used prior to PCR to enrich for mutant DNA where the fragment between the two guides had been removed. PCR amplification across the region thus led to shorter PCR products than expected from a wild-type individual.

DNA was extracted according to Edwards *et al.* [47] from rooted shoots of less than 10 cm in height and quantified using a Nanodrop 8000 (Thermo Scientific). Genomic DNA (100 ng) was digested overnight with 20 units of the appropriate restriction enzyme shown in Fig. 1 (*SapI*, *HaeIII*, *HphI*, *AflIII* (NEB); *MaeIII* (Roche)) and then purified using a Qiagen QIAquick Gel Extraction Kit (final elution with 25 μ L of water). Purified digested DNA (5 μ L) was used as PCR template to amplify across the target regions using gene-specific primers (Additional

file 3). PCR products were confirmed by agarose gel electrophoresis, purified using the QIAquick Gel Extraction Kit, and Sanger sequenced (Eurofins MWG) to confirm the presence of in-dels. Where the amplicon was too short for direct sequencing, the PCR product was first cloned using the pGEMT-Easy kit (Promega) according to the manufacturer's instructions and then sequenced with M13 universal primers.

The detection of mutations in T_1 and T_2 transgenic lines was performed through Sanger sequencing of PCR amplicons produced using genomic DNA template that was not digested prior to PCR (Additional file 3). Sequences were compared to wild type to detect the presence of homozygous in-dels. Chromatograms were also examined to identify overlapping traces in the region surrounding the PAM, indicative of the presence of mutations. The presence of the T-DNA construct was assessed in progenies of active lines by PCR amplification of the *nptII* CDS in *Brassica* and *hptII* CDS in barley (Additional file 3).

Phenotyping of *B. oleracea* transgenic lines

The 80 primary T_0 transgenic lines and corresponding controls were grown in a controlled environment room with 16 h light (high-pressure sodium lamps with an average bench reading of 200 $\mu\text{mol}/\text{m}^2/\text{s}$) at 12 °C and 8 h dark at 12 °C and constant 65–75 % humidity. Plant height was measured at final maturity. Seed pods at developmental stage 17 [48] were collected from dwarf line L2F1_A and the wild-type DH1012 control. Pods were fixed for 16 h in FAA solution (3.7 % formaldehyde, 5 % acetic acid, 50 % ethanol) and subsequently dehydrated through an ethanol series consisting of 50 %, 60 %, 70 %, 80 %, 90 %, 95 %, and 100 % ethanol for 30 min each at room temperature. The tissues were cleared with Histo-clear (National Diagnostics,) and embedded in paraffin wax. Transverse sections 8 μm thick were cut using an RM 2055 rotary microtome (Leica) and mounted on Polysine™ slides (VWR International). The wax was removed using Histo-clear and sections stained using an Alcian Blue/Safranin-O solution (0.05 % Alcian Blue and 0.01 % Safranin-O in 0.1 M acetate buffer (pH5.0)) as described by Østergaard *et al.* [49]. Sections were examined by light microscopy using a Zeiss Axioplan microscope and images captured using a Leica DFC 320 camera with Leica Application Suite software.

T-DNA copy number and presence/absence determination in transgenic barley

Quantitative real-time PCR was used to determine copy number (T_0) and presence/absence (T_2) of the T-DNA in transgenic barley and *B. oleracea* lines. The reaction compared the C_q values of an *HptII* (Fig. 2a) amplicon to a single-copy barley gene *CO2* (*Constans-like*, AF490469)

amplicon and the C_q values of an *NptII* amplicon to a single-copy *B. oleracea* gene *GL2-like* (*Bol021421*) within a single multiplexed assay (Additional file 3). The reactions used Thermo ABGene Absolute QPCR Rox Mix (Cat number AB1139) with the probes and primers at a final concentration of 200 nM (*HptII* and *NptII*) and 100nM (*CO2* and *GL2*). The assay contained 5 μL DNA solution, and was optimised for final DNA concentrations of 1 to 10 ng/ μL (5 to 50 ng DNA in the assay). PCRs were carried out in a Bio-Rad CFX96 machine (C1000 Touch). The detectors used were FAM-TAMRA and VIC-TAMRA for barley and HEX-BHQ1 and FAM-BHQ1 for *B. oleracea*. The PCR cycling conditions were 95 °C for 15 min (enzyme activation), 40 cycles of 95 °C for 15 s, and 60 °C for 60 s. Each sample was analysed twice and for presence/absence determinations, two independent DNA extractions of the T_2 transgenic plants were used. C_q values were determined using the accompanying CFX96 software (version 3.1), with C_q determination set to regression mode. Values obtained were used to calculate T-DNA copy number according to published methods [50].

Additional files

Additional file 1: Chromatogram traces of T_0 -181 and T_0 -191 plants. (JPG 439 kb)

Additional file 2: Analysis of *BolC.GA4.a* sequences in L2F1_A and L2F1_E T_0 and T_1 plants. (DOCX 25 kb)

Additional file 3: Primers and probes used in this study. (XLSX 12 kb)

Additional file 4: Detailed methods for assembly of binary vectors with multiple sgRNAs using the Golden Gate MoCloToolKit. (DOCX 28 kb)

Additional file 5: Genbank files (.gb) of the 14 plasmids submitted to AddGene. (ZIP 41 kb)

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

TL, OS and NP designed and generated the constructs used in this study; TL generated the transgenic lines; TL and OS performed the molecular characterisation of the transgenic lines; NS performed the phenotypic characterisation of *B. oleracea* lines; CL provided pre-publication access to barley BAC sequence; WH and CU designed the experiments. TL, OS, NS, LO, NP, CU and WH contributed to the writing of the manuscript and all authors reviewed the final version. All authors read and approved the final manuscript.

Acknowledgments

The authors would like to thank Dr Nils Stein (IPK), Xiao-Qi Zhang (Murdoch University), Guoping Zhang (Zhejiang University) and Mr Simon Lee (BGI-Shenzhen) for providing pre-publication access to barley Morex BAC sequences.

Funding

This work was supported by the Institute Strategic Programme grant BB/J004553/1 from the UK Biotechnology and Biological Sciences Research Council (BBSRC), the BBSRC and Engineering and Physics Research Council (EPSRC) Synthetic Biology Research Centre award (BB/L014130/1), by the John Innes Centre Innovation fund, the John Innes Foundation and the Gatsby Charitable Foundation.

Author details

¹John Innes Centre, Norwich Research Park, Colney NR4 7UH, UK. ²Western Barley Genetics Alliance, Murdoch University, Murdoch WA6150, Australia. ³The Sainsbury Laboratory, Norwich Research Park, Colney NR4 7UH, UK.

Received: 30 July 2015 Accepted: 5 November 2015

Published online: 30 November 2015

References

- Voytas DF, Gao C. Precision genome engineering and agriculture: opportunities and regulatory challenges. *PLoS Biol.* 2014;12:e1001877.
- Hsu Patrick D, Lander Eric S, Zhang F. Development and applications of CRISPR-Cas9 for genome engineering. *Cell.* 2014;157:1262–78.
- Cong L, Ran FA, Cox D, Lin S, Barretto R, Habib N, et al. Multiplex genome engineering using CRISPR/Cas systems. *Science.* 2013;339:819–23.
- Mali P, Yang L, Esvelt KM, Aach J, Guell M, DiCarlo JE, et al. RNA-guided human genome engineering via Cas9. *Science.* 2013;339:823–6.
- Belhaj K, Chaparro-Garcia A, Kamoun S, Patron NJ, Nekrasov V. Editing plant genomes with CRISPR/Cas9. *Curr Opin Biotechnol.* 2015;32:76–84.
- Li J-F, Norville JE, Aach J, McCormack M, Zhang D, Bush J, et al. Multiplex and homologous recombination-mediated genome editing in *Arabidopsis* and *Nicotiana benthamiana* using guide RNA and Cas9. *Nat Biotech.* 2013;31:688–91.
- Nekrasov V, Staskawicz B, Weigel D, Jones JDG, Kamoun S. Targeted mutagenesis in the model plant *Nicotiana benthamiana* using Cas9 RNA-guided endonuclease. *Nat Biotech.* 2013;31:691–3.
- Shan Q, Wang Y, Li J, Zhang Y, Chen K, Liang Z, et al. Targeted genome modification of crop plants using a CRISPR-Cas system. *Nat Biotech.* 2013;31:686–8.
- Feng Z, Mao Y, Xu N, Zhang B, Wei P, Yang D-L, et al. Multigeneration analysis reveals the inheritance, specificity, and patterns of CRISPR/Cas-induced gene modifications in *Arabidopsis*. *Proc Natl Acad Sci.* 2014;111:4632–7.
- Zhang H, Zhang J, Wei P, Zhang B, Gou F, Feng Z, et al. The CRISPR/Cas9 system produces specific and homozygous targeted gene editing in rice in one generation. *Plant Biotechnol J.* 2014;12:797–807.
- Zhou H, Liu B, Weeks DP, Spalding MH, Yang B. Large chromosomal deletions and heritable small genetic changes induced by CRISPR/Cas9 in rice. *Nucleic Acids Res.* 2014;42:10903–14.
- Wang Y, Cheng X, Shan Q, Zhang Y, Liu J, Gao C, et al. Simultaneous editing of three homoeoalleles in hexaploid bread wheat confers heritable resistance to powdery mildew. *Nat Biotech.* 2014;32:947–51.
- Arnaud N, Girin T, Sorefan K, Fuentes S, Wood TA, Lawrenson T, et al. Gibberellins control fruit patterning in *Arabidopsis thaliana*. *Genes Dev.* 2010;24:2127–32.
- Chiang H-H, Hwang I, Goodman HM. Isolation of the *Arabidopsis* *GA4* locus. *Plant Cell.* 1995;7:195–201.
- Ranford JC, Bryce JH, Morris PC. *PM19*, a barley (*Hordeum vulgare* L.) gene encoding a putative plasma membrane protein, is expressed during embryo development and dormancy. *J Exp Bot.* 2002;53:147–8.
- Barrera JM, Cavanagh C, Verbyla KL, Tibbits JFG, Verbyla AP, Huang BE, et al. Transcriptomic analysis of wheat near-isogenic lines identifies *PM19-A1* and *A2* as candidates for a major dormancy QTL. *Genome Biol.* 2015;16:93.
- Bortesi L, Fischer R. The CRISPR/Cas9 system for plant genome editing and beyond. *Biotechnol Adv.* 2015;33:41–52.
- Jiang W, Zhou H, Bi H, Fromm M, Yang B, Weeks DP. Demonstration of CRISPR/Cas9/sgRNA-mediated targeted gene modification in *Arabidopsis*, tobacco, sorghum and rice. *Nucleic Acids Res.* 2013;41(20):e188.
- Jinek M, Chylinski K, Fonfara I, Hauer M, Doudna JA, Charpentier E. A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science.* 2012;337:816–21.
- Hartung F, Schiemann J. Precise plant breeding using new genome editing techniques: opportunities, safety and regulation in the EU. *Plant J.* 2014;78:742–52.
- Xu R-F, Li H, Qin R-Y, Li J, Qiu C-H, Yang Y-C, et al. Generation of inheritable and “transgene clean” targeted genome-modified rice in later generations using the CRISPR/Cas9 system. *Sci Rep.* 2015;5:11491.
- Sparrow P, Dale P, Irwin J. *Brassica oleracea*. In: Wang K, editor. *Agrobacterium Protocols*, vol. 343. New York: Humana Press; 2006. p. 417–26.
- Mitchum MG, Yamaguchi S, Hanada A, Kuwahara A, Yoshioka Y, Kato T, et al. Distinct and overlapping roles of two gibberellin 3-oxidases in *Arabidopsis* development. *Plant J.* 2006;45:804–18.
- Brooks C, Nekrasov V, Lippman ZB, Van Eck J. Efficient gene editing in tomato in the first generation using the clustered regularly interspaced short palindromic repeats/CRISPR-associated9 system. *Plant Physiol.* 2014;166:1292–7.
- Mikami M, Toki S, Endo M. Parameters affecting frequency of CRISPR/Cas9 mediated targeted mutagenesis in rice. *Plant Cell Rep.* 2015;34:1807–15.
- Endo M, Mikami M, Toki S. Multi-gene knockout utilizing off-target mutations of the CRISPR/Cas9 system in rice. *Plant Cell Physiol.* 2015;56:41–7.
- Jiang W, Bikard D, Cox D, Zhang F, Marraffini LA. RNA-guided editing of bacterial genomes using CRISPR-Cas systems. *Nat Biotech.* 2013;31:233–9.
- Cradick TJ, Fine EJ, Antico CJ, Bao G. CRISPR/Cas9 systems targeting β -globin and CCR5 genes have substantial off-target activity. *Nucleic Acids Res.* 2013;41:9584–92.
- Lin Y, Cradick TJ, Brown MT, Deshmukh H, Ranjan P, Sarode N, et al. CRISPR/Cas9 systems have off-target activity with insertions or deletions between target DNA and guide RNA sequences. *Nucleic Acids Res.* 2014;42:7473–85.
- Fu Y, Sander JD, Reyon D, Cascio VM, Joung JK. Improving CRISPR-Cas nuclease specificity using truncated guide RNAs. *Nat Biotechnol.* 2014;32:279–84.
- Ran FA, Hsu Patrick D, Lin C-Y, Gootenberg Jonathan S, Konermann S, Trevino AE, et al. Double nicking by RNA-guided CRISPR Cas9 for enhanced genome editing specificity. *Cell.* 2013;154:1380–9.
- Shen B, Zhang W, Zhang J, Zhou J, Wang J, Chen L, et al. Efficient genome modification by CRISPR-Cas9 nickase with minimal off-target effects. *Nat Meth.* 2014;11:399–402.
- Guilinger JP, Thompson DB, Liu DR. Fusion of catalytically inactive Cas9 to FokI nuclease improves the specificity of genome modification. *Nat Biotech.* 2014;32:577–82.
- Tsai SQ, Wyvekens N, Khayter C, Foden JA, Thapar V, Reyon D, et al. Dimeric CRISPR RNA-guided FokI nucleases for highly specific genome editing. *Nat Biotech.* 2014;32:569–76.
- Borrill P, Adamski N, Uauy C. Genomics as the key to unlocking the polyploid potential of wheat. *New Phytol.* 2015;208:1008–22.
- Adams KL, Wendel JF. Polyploidy and genome evolution in plants. *Curr Opin Plant Biol.* 2005;8:135–41.
- The International Wheat Genome Sequencing Consortium. A chromosome-based draft sequence of the hexaploid bread wheat (*Triticum aestivum*) genome. *Science.* 2014;345:1251788.
- Girin T, Stephenson P, Goldsack CMP, Kempin SA, Perez A, Pires N, et al. Brassicaceae *INDEHISCENT* genes specify valve margin cell fate and repress replum formation. *Plant J.* 2010;63:329–38.
- Spence J, Vercher Y, Gates P, Harris N. ‘Pod shatter’ in *Arabidopsis thaliana*, *Brassica napus* and *B. juncea*. *J Microsc.* 1996;181:195–203.
- The Brassica Database. [<http://brassicadb.org/brad/blastPage.php>].
- International Barley Sequencing Consortium Database [<http://webblast.ipk-gatersleben.de/barley/>].
- Weber E, Engler C, Gruetzner R, Werner S, Marillonnet S. A modular cloning system for standardized assembly of multigene constructs. *PLoS One.* 2011;6:e16765.
- Engler C, Youles M, Gruetzner R, Ehnert T-M, Werner S, Jones JDG, et al. A golden gate modular cloning toolbox for plants. *ACS Synthetic Biol.* 2014;3:839–43.
- Harwood W. A Protocol for High-Throughput Agrobacterium-Mediated Barley Transformation. In: Henry RJ, Furtado A, editors. *Cereal Genomics*, vol. 1099. New York: Humana Press; 2014. p. 251–60.
- Hundleby PAC, Irwin J. *Brassica oleracea* and *B. napus*. In: Wang K, editor. *Agrobacterium Protocols*, vol. 1223. New York: Springer; 2015. p. 287–97.
- Voytas DF. Plant genome engineering with sequence-specific nucleases. *Annu Rev Plant Biol.* 2013;64:327–50.
- Edwards K, Johnstone C, Thompson C. A simple and rapid method for the preparation of plant genomic DNA for PCR analysis. *Nucleic Acids Res.* 1991;19:1349.
- Smyth DR, Bowman JL, Meyerowitz EM. Early flower development in *Arabidopsis*. *Plant Cell.* 1990;2:755–67.

49. Østergaard L, Kempin SA, Bies D, Klee HJ, Yanofsky MF. Pod shatter-resistant Brassica fruit produced by ectopic expression of the *FRUITFULL* gene. *Plant Biotechnol J*. 2006;4:45–51.
50. Weng H, Pan A, Yang L, Zhang C, Liu Z, Zhang D. Estimating number of transgene copies in transgenic rapeseed by real-time PCR assay with HMG 1/Y as an endogenous reference gene. *Plant Mol Biol Rep*. 2004;22:289–300.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

