JOURNAL OF BACTERIOLOGY, Nov. 2007, p. 7932–7936 Vol. 189, No. 21
0021-9193/07/$08.00+0 doi:10.1128/JB.00693-07

Inferring the Evolutionary History of Vibrios by Means of Multilocus Sequence Analysis▽†

Tomoo Sawabe,[1]* Kumiko Kita-Tsukamoto,[2] and Fabiano L. Thompson[3]

Laboratory of Microbiology, Faculty of Fisheries, Hokkaido University, 3-1-1 Minato-cho, Hakodate 041-8611, Japan[1]; Ocean Research Institute, University of Tokyo, 1-15 Minamindai, Nakano-ku, Tokyo 164-8639, Japan[2]; and Department of Genetics, Institute of Biology, Federal University of Rio de Janeiro, Ilha do Fundão, Caixa Postal 68011, CEP 21944-970, Rio de Janeiro, Brazil[3]

Received 1 May 2007/Accepted 9 August 2007

We performed the first broad study aiming at the reconstruction of the evolutionary history of vibrios by means of multilocus sequence analysis of nine genes. Overall, 14 distinct clades were recognized using the SplitsTree decomposition method. Some of these clades may correspond to families, e.g., the clades Salinivibrio and Photobacteria, while other clades, e.g., Splendidus and Harveyi, correspond to genera. The common ancestor of all vibrios was estimated to have been present 600 million years ago. We can define species of vibrios as groups of strains that share >95% gene sequence similarity and >99.4% amino acid identity based on the eight protein-coding housekeeping genes. The gene sequence data were used to refine the standard online electronic taxonomic scheme for vibrios (http://www.taxvibrio.lncc.br).

Vibrios are widespread in the aquatic environment, occupying a variety of ecological niches, such as the human and animal gut, the surface of chitinous organisms, most notably copepods, and the coral mucus layer. A better understanding of the ecology and the patterns of distribution of vibrios relies on the online electronic taxonomy. Polyphasic taxonomic studies of vibrios performed in recent years have underpinned this new paradigm in studies of the biodiversity and systematics of this group (16, 17, 19). Currently, we recognize 78 species of vibrios distributed into five phylogenetic robust clades corresponding to the genera *Vibrio*, *Photobacterium*, *Salinivibrio*, *Enterovibrio*, and *Grimontia* based on 16S rRNA gene sequences (16, 17, 19). Both genome content and architecture indicate that these genera share a common ancestor (12). In addition, the genera within vibrios are defined on the basis of their shared sequence similarities in different loci. Species within the genus *Vibrio* share at least 85% gene sequence similarity in *recA*, *rpoA*, and *pyrH* (18).

Species of vibrios are defined as clusters of strains with high phenotypic and genotypic similarities. Clusters comprise strains with highly similar genomes as determined by multilocus sequence analysis (MLSA), amplified fragment length polymorphism analysis, and DNA-DNA hybridization (DDH) (16, 17, 19). Formal delineation of bacterial species still relies on DDH, with a cutoff level of >70% DDH similarity, but this technique is time-consuming and can be performed in relatively few laboratories and, more importantly, the DDH data are not cumulative in online databases. Clearly, a reliable and straightforward alternative is the use of MLSA. The usefulness of MLSA in the taxonomy of vibrios was described in previous papers (e.g., references 15 and 18). Overall, species form discrete clusters on the basis of *recA*, *rpoA*, and *pyrH*, with a species cutoff level of >94% gene sequence similarity (18). However, some groups of species, e.g., the *Vibrio splendidus* and *Vibrio harveyi* species groups, were somewhat fuzzy on the basis of *recA*, *gyrB*, and *gapA* (15, 18). Thus, it is very important to evaluate additional genetic markers that can distinguish closely related species of vibrios.

DNA sequences may also be useful in unraveling the nature of the speciation processes in vibrios. Some studies suggest that recombination might have occurred between different sister species, such as between *V. cholerae* and *V. mimicus* and between *V. harveyi* and *V. campbellii*, but it is not clear how prevalent and widespread this process is when all vibrio species groups are analyzed simultaneously. The rationale of this study is that by analyzing partial sequences of nine genes (i.e., *ftsZ*, *gapA*, *gyrB*, *mreB*, *pyrH*, *recA*, *rpoA*, *topA*, and 16S rRNA), we will be able to establish a more robust inference of the evolutionary history of vibrios. Clearly, we will also enhance and refine the framework of the online electronic taxonomy (5–19). This framework will allow prompt identification and classification of vibrios through the Internet.

The GenBank accession numbers for the *gapA*, *ftsZ*, *mreB*, *topA*, and *gyrB* gene sequences are listed in Table S1 of the supplemental material (see also references 6 and 21).

The sequences of the *ftsZ*, *gapA*, *gyrB*, *mreB*, *pyrH*, *recA*, *rpoA*, *topA*, and 16S rRNA genes of 78 type strains were concatenated and analyzed by the ClustalX program, MEGA version 3.0, and split decomposition (1, 7, 11, 20) (Fig. 1). Clearly, there were at least 14 monophyletic clades showing split signals with significant bootstrap support (Fig. 1; see also Fig. S2 in the supplemental material [a bifurcating tree]). These clades were always retained in the analysis even when the number of genes was reduced to five loci, i.e., *ftsZ*, *gapA*, *gyrB*, *mreB*, and *topA* (see Fig. S3 and S4 in the supplemental material). The species within each clade shared >20% DDH, <5% GC variation

* Corresponding author. Mailing address: Laboratory of Microbiology, Faculty of Fisheries, Hokkaido University, 3-1-1 Minato-cho, Hakodate 041-8611, Japan. Phone and fax: 81-138-40-5570. E-mail: sawabe@fish.hokudai.ac.jp.

† Supplemental material for this article may be found at http://jb.asm.org/.

▽ Published ahead of print on 17 August 2007.

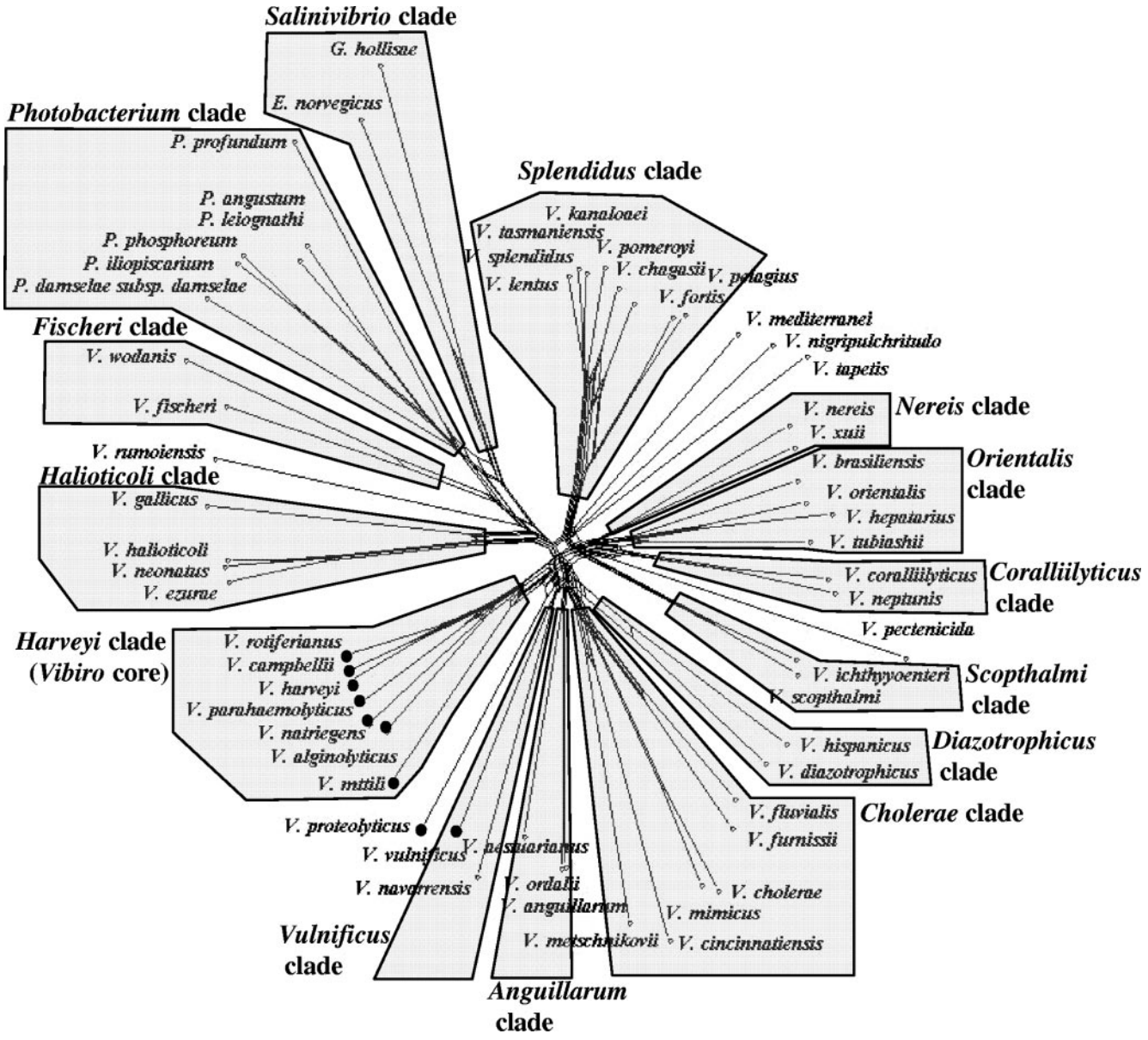

FIG. 1. Concatenated split network tree based on nine gene loci. The *ftsZ*, *gapA*, *gyrB*, *mreB*, *pyrH*, *recA*, *topA*, and 16S rRNA gene sequences (6,050 bp) from 58 taxa were concatenated and reconstructed using the SplitsTree4 program. Vibrios defined as a vibrio core group (3, 9) are marked with a closed circle. Other clades, which were not included in the analysis, were determined based on five to six gene loci (see Fig. S3 and S4 in the supplemental material). Domains used to construct the phylogenetic tree were regions of *ftsZ*, *gapA*, *gyrB*, *mreB*, *pyrH*, *recA*, *rpoA*, *topA*, and 16S rRNA genes of *Vibrionaceae*, positions 133 to 625, 226 to 893, 441 to 1030, 387 to 892, 89 to 533, 304 to 915, 70 to 903, 155 to 1209 and 451 to 1076 (*V. cholerae* O1 El tor N16961 [AE003852] numbering), respectively. Sequence similarities were corrected using the Jukes-Cantor correction. All nodes were supported by 100 bootstrap replications.

(mol%), >85% MLSA sequence similarity, and >89% average amino acid identity (AAI) (Table 1).

Radiation times were estimated for different sister species based on the rate of amino acid substitutions in the eight housekeeping protein genes. These rates were normalized with the known radiation time between *Escherichia coli* and *Salmonella enterica* (ca. 1 million years) (8, 13). The lowest radiation time value was calculated for the pair *V. splendidus* and *V. tasmaniensis* (Table 1; see also Table S3 in the supplemental material). These species have highly related genomes, with 61% DDH similarity, and may occupy very similar niches (19). The time span of speciation in the well-known closely related species pairs (e.g., *V. anguillarum* and *V. ordalii*, *V. cholerae* and *V. mimicus*, *V. halioticoli* and *V. neonatus*, and *V. harveyi* and *V. campbellii*) was estimated as 23 to 56 million years. A common ancestor in some *Vibrio* clades might have occurred at 360 to 390 million years (e.g., *V. anguillarum* and *V. aestuarianus*, *V. fischeri* and *V. logei*, or *V. halioticoli* and *V. gallicus*) (Table 1; see also Table S3 in the supplemental material), corresponding the Devonian era of vigorous diversification of marine fish. The common ancestor of *Salinivibrio*, *Enterovibrio*, and *Grimontia* might have occurred 580 to 620 million years ago (see

TABLE 1. Clades and subclades proposed by means of MLSA for vibrios

Clade or subclade	Described species included	No. of species	DDH value (%)[a]	GC (mol%)[b]	MLSA concatenated similarity (%)	AAI (%)[d]	Phi test[e] (*P* value)	Estimated radiation time(s) for representative pair(s) (10^8 yrs)[f]	Habitat
Vibrio clades		57			85.4–98.6	89.0–99.8			
Anguillarum	*V. anguillarum*, *V. aestuarianus*, and *V. ordalii*	3	>30	43–46	88.9–98.6	95.7–99.6		0.56 (*V. anguillarum*/*V. ordalii*), 3.76 (*V. anguillarum*/*V. aestuarianus*)	Brackish water, seawater, and fish
Cholerae	*V. cholerae*, *V. cincinnatiensis*, *V. furnissii*, *V. fluvialis*, *V. metschnikovii*, and *V. mimicus*	6	NA	44–50	85.4–94.7	92.8–99.6	<0.1	0.30 (*V. cholerae*/*V. mimicus*), 1.79 (*V. fluvialis*/*V. furnissii*), 3.59 (*V. cincinnatiensis*/*V. metschnikovii*)	Brackish water and seawater
Coralliilyticus	*V. coralliilyticus* and *V. neptunis*	2	>64	45–46	95.6	99.6		NT	Seawater, bivalves, and rotifers
Diazotrophicus	*V. diazotrophicus* and *V. hispanicus*	2	NA	43–47	91.2	97.0		NT	Brackish water and seawater
Gazogenes	*V. aerogenes*, *V. gazogenes*, and *V. ruber*	3	>32	46–47	NT	NT		NT	Estuary and salt marsh mud
Fischeri	*V. fischeri*, *V. logei*, *V. salmonicida*, and *V. wodanis*	4	>36	39–42	89.8–94.4[b]	95.2	<0.1[b]	3.90 (*V. fischeri*/*V. logei*)	Seawater, squid, and fish
Halioticoli	*V. halioticoli*, *V. ezurae*, *V. gallicus*, *V. neonatus*, and *V. superstes*	5	>22	39–42	88.0–97.7	94.7–99.5	<0.1	0.23 (*V. halioticoli*/*V. neonatus*), 1.11 (*V. halioticoli*/*V. ezurae*), 3.65 (*V. halioticoli*/*V. gallicus*)	Gut of abalone
Harveyi	*V. harveyi*, *V. alginolyticus*, *V. campbellii*, *V. mytili*, *V. natriegens*, *V. parahaemolyticus*, and *V. rotiferianus*	8	>25	42–48	90.1–96.2	97.2–99.4	<0.1	0.39 (*V. harveyi*/*V. campbellii*), 1.05 (*V. harveyi*/*V. parahaemolyticus*), 1.81 (*V. harveyi*/*V. mytili*)	Seawater, salt marsh mud, and marine animals
Nereis	*V. nereis* and *V. xuii*	2	>30	39–47	91.2	96.8		NT	Seawater and shrimp
Nigripulchritudo	*V. nigripulchritudo* and *V. penaeicida*	2	>36	46–47	89.0[c]	NT		NT	Seawater and shrimp
Orientalis	*V. orientalis*, *V. brasiliensis*, *V. hepatarius and V. tubiashii*	4	>24	43–46	91.2–94.2	97–97.9	<0.1	3.64 (*V. orientalis*/*V. tubiashii*)	Brackish water and seawater
Scopthalmi	*V. scophthalmi* and *V. ichthyoenteri*	2	>32	43–44	95.5	99.4		NT	Gut of flat fish
Splendidus	*V. splendidus*, *V. chagasii*, *V. crassostrea*, *V. cyclitrophicus*, *V. fortis*, *V. gigantis*, *V. kanaloaei*, *V. lentus*, *V. pelagius*, *V. pomeroyi*, and *V. tasmaniensis*	12	>30	39–47	90.6–96.5	96.5–99.8	<0.1	0.17 (*V. splendidus*/*V. tasmaniensis*), 1.61 (*V. splendidus*/*V. chagasii*), 2.59 (*V. splendidus*/*V. pelagius*)	Seawater and marine animals
Vulnificus	*V. vulnificus* and *V. navarrensis*	2	>30	45–48	88.6	96.4		NT	Sewage, seawater, eel, and oyster
Photobacterium subclades		10			87.5–95.8	98.6–99.4			
Damselae	*P. damselae*	1		42					Seawater and fish
Leiognathi	*P. leiognathi* and *P. angustum*	2	>44	40–44	94.0	98.6		1.07	Seawater and luminous organs
Phosphoreum	*P. phosphoreum*, *P. frigidiphilum*, and *P. iliopiscarius*	3	NA	38–43.8	94.24–95.8[b]	99.4		0.61 (*P. phosphoreum*/*P. iliopiscarius*)	Seawater and fish
Profundum	*P. profundum*, *P. indicum*, and *P. lipolyticum*	3	NA	40–42	87.5–92.3[b]			NT	Deep sea
Rosenbergii	*P. rosenbergii*	1		47.6–47.9					Seawater and coral

[a] Data are from references 4 and 5. NA, not available.
[b] Calculated based on six genes.
[c] Calculated based on five genes.
[d] Raw average amino acid identity (10).
[e] The phi test was conducted for clades that included at least four species.
[f] The radiation time was calculated based on average amino acid substitution results (see also Table S3 in the supplemental material). NT, not tested.

Table S3 in the supplemental material), corresponding to the era of Cambrian explosion. Diversification of vibrios may have occurred during this period. Major branches showing distinct split signals represent species groups, some of which (e.g., halioticoli, splendidus, and cholerae) may share ecological niches.

All *Photobacterium* species formed a single clade that may well correspond to a family on its own. Some structuring was observed within this clade though, with at least five subclades. Split decomposition clearly separated *Salinivibrio costicola*, *Enterovibrio norvegicus*, *Enterovibrio corallii*, *Grimontia hollisae*, and *Vibrio calviensis* from the other vibrios. *Salinivibrio* seems to be the ancestor of the vibrios. The clades Salinivibrio and Photobacterium may correspond to families, while other clades, e.g., Splendidus and Harveyi, correspond to genera. The Fischeri clade appeared in an intermediate position between the Photobacterium and Halioticoli clades, suggesting that the *V. fischeri* species group may represent a genus on its own.

Overall, the species found in each clade have related genomes. The clades disclosed in this study are congruent with former polyphasic taxonomic work (Table 1). For instance, the species in the Anguillarum clade have a GC content ranging between 43 and 46 mol%. *V. anguillarum* and *V. ordalii* have at least 58% mutual DDH similarity and around 30% DDH similarity with *V. aestuarianus* (5). The Cholerae clade comprises six species which show a broad GC content range. Most of the species within this clade cause diarrhea, but only *V. cholerae* harbors epidemic and pandemic strains. High DDH values (>65%) between the pair *V. cholerae* and *V. mimicus* and between the pair *V. fluvialis* and *V. furnisii* were reported, suggesting that these species have closely related genomes. The Cholerae clade includes species with lower Na^+ requirements. For instance, the Na^+ requirements of *V. cholerae*, *V. metchinikovii*, and *V. fluvialis* range between 5 and 40 mM (2, 4, 5). The so-called *Vibrio* core group (3, 9) forms the Harveyi clade. Distinguishing species and strains within this clade remains a hard task in taxonomy (20). Recombination between closely related species may partially explain this fact.

Recombination was detected in the Cholerae, Fischeri, Halioticoli, Harveyi, Orientalis, and Splendidus clades (Table 1). The phi test implemented in SplitsTree4 pointed to recombination within the concatenated sequences of vibrios ($P = 5.0 \times 10^{-5}$). Recombination was detected in *gyrB* ($P = 6.4 \times 10^{-3}$), *rrn* ($P = 7.3 \times 10^{-12}$), *gapA* ($P = 6.8 \times 10^{-7}$), and *topA* ($P = 4.5 \times 10^{-2}$), in agreement with the conflicting phylogenetic splits (parallelograms) observed on the basis of the SplitsTree program (Fig. 1). Recombination was observed in the Cholerae, Fischeri, Halioticoli, Harveyi, Orientalis, and Splendidus clades, at least (Table 1). The recombination analysis suggests that genes responsible for different essential functions in the cell may be targets of recombination, but we cannot rule out the possibility that the recombination tests are providing false positive results. Detecting recombination is basically a statistical endeavor and ideally in vitro experimental work should be carried out in order to confirm the ability of vibrios to carry out recombination in the loci analyzed in this study.

We can define species of vibrios as groups of strains that share >95% gene sequence similarity and >99.4% AAI based on the eight protein-coding housekeeping genes. This definition is an alternative to the standard definition of *Vibrio* species based on DDH values. We used the gene sequences generated in this study to refine the current standard online electronic taxonomic scheme for vibrios (http://www.taxvibrio.lncc.br) (15). This work will underpin further analyses of fresh isolates of vibrios. In one test case, we analyzed several presumptive *V. harveyi* strains isolated from diseased animals in Tasmania and found that, in fact, these strains formed a tight new cluster that represents a new species within the Harveyi clade (J. Carson et al., unpublished data).

We are grateful to Shinichi Koizumi and Yasuhiro Tsuruya for technical assistance. We thank C. Vereecke (LMG Culture Collection, Ghent University, Ghent, Belgium) for providing bacterial strains. We also thank K. Kogure and M. Wada (Tokyo University) for helpful comments.

This work was supported by the Institute of Fermentation Osaka. This work was partly supported by an invitation program of the Goho Life Science Foundation and by the JSPS program to F. L. Thompson (S0614). F. L. Thompson acknowledges grants from IFS and CNPq.

REFERENCES

1. **Bandelt, H.-J., and A. W. M. Dress.** 1992. Split decomposition: a new and useful approach to phylogenetic analysis of distance data. Mol. Phylogenet. Evol. **1:**242–252.
2. **Baumann, P., L. Baumann, S. S. Bang, and M. J. Woolkalis.** 1980. Reevaluation of the taxonomy of *Vibrio*, *Beneckea*, and *Photobacterium*: abolition of the genus *Beneckea*. Curr. Microbiol. **4:**127–132.
3. **Dorsch, M., D. Lane, and E. Stackebrandt.** 1992. Toward a phylogeny of the genus *Vibrio* based on 16S rRNA sequences. Int. J. Syst. Bacteriol. **42:**58–63.
4. **Farmer, J. J., III, and J. M. Janda.** 2005. *Vibrionaceae* Veron 1965, p. 491–494. *In* D. J. Brenner, N. R. Krieg, J. T. Staley, and G. M. Garrity (ed.), Bergey's manual of systematic bacteriology, 2nd ed., vol. 2, part B. Springer, New York, NY.
5. **Farmer, J. J., III, J. M. Janda, F. W. Brenner, D. N. Cameron, and K. M. Birkhed.** 2005. *Vibrio* Pacini 1854, 411^{AL}, p. 494–546. *In* D. J. Brenner, N. R. Krieg, J. T. Staley, and G. M. Garrity (ed.), Bergey's manual of systematic bacteriology, 2nd ed., vol. 2, part B. Springer, New York, NY.
6. **Gil, R., F. J. Silva, J. Pereto, and A. Moya.** 2004. Determination of the core of a minimal bacterial gene set. Microbiol. Mol. Biol. Rev. **68:**518–537.
7. **Huson, D. H., and D. Bryant.** 2005. Application of phylogenetic networks in evolutionary studies. Mol. Biol. Evol. **23:**254–267.
8. **Hyma, K. E., D. W. Lacher, A. M. Nelson, A. C. Bumbaugh, J. M. Janda, N. A. Strocjbine, V. B. Young, and T. S. Whittam.** 2005. Evolutionary genetics of a new pathogenic *Escherichia* species: *Escherichia albertii* and related *Shigella boydii* strains. J. Bacteriol. **187:**619–628.
9. **Kita-Tsukamoto, K., H. Oyaizu, K. Nanba, and U. Shimizu.** 1993. Phylogenetic relationships of marine bacteria, mainly members of the family *Vibrionaceae*, determined on the basis of 16S rRNA sequences. Int. J. Syst. Bacteriol. **43:**8–19.
10. **Konstantinidis, K. T., and J. M. Tiedje.** 2005. Towards a genome-based taxonomy for prokaryotes. J. Bacteriol. **187:**6258–6264.
11. **Kumar, S., K. Tamura, and M. Nei.** 2004. MEGA3: integrated software for molecular evolutionary genetics analysis and sequence alignment. Brief. Bioinform. **5:**150–163.
12. **Okada, K., T. Iida, K. Kita-Tsukamoto, and T. Honda.** 2005. Vibrios commonly possess two chromosomes. J. Bacteriol. **187:**752–757.
13. **Reid, S. D., C. J. Herbelin, A. C. Bumbaugh, R. K. Selander, and T. S. Whitam.** 2000. Parallel evolution of virulence in pathogenic *Escherichia coli*. Nature **406:**64–67.
14. **Scola, B. L., Z. Zeaiter, A. Khamis, and D. Raoult.** 2003. Gene-sequence-based criteria for species definition in bacteriology: the *Bartonella* paradigm. Trends Microbiol. **11:**318–321.
15. **Thompson, F. L., B. Gomez-Gil, A. T. R. Vasconcelos, and T. Sawabe.** 2007. Multilocus sequence analysis reveals that *Vibrio harveyi* and *V. campbellii* form distinct species. Appl. Environ. Microbiol. **73:**4279–4285.
16. **Thompson, F. L., B. Hoste, K. Vandemeulebroecke, and J. Swings.** 2001. Genomic diversity amongst *Vibrio* isolates from different sources determined by fluorescent amplified fragment length polymorphism. Syst. Appl. Microbiol. **24:**520–538.
17. **Thompson, F. L., T. Iida, and J. Swings.** 2004. Biodiversity of vibrios. Microbiol. Mol. Biol. Rev. **68:**403–431.
18. **Thompson, F. L., D. Gevers, C. C. Thompson, P. Dawyndt, S. Naser,**

B. Hoste, C. B. Munn, and J. Swings. 2005. Phylogeny and molecular identification of vibrios on the basis of multilocus sequence analysis. Appl. Environ. Microbiol. **71:**5107–5115.
19. **Thompson, F. L., and J. Swings.** 2006. Taxonomy of the vibrios, p. 29–43. *In* F. L. Thompson, B. Austin, and J. Swings (ed.), The biology of vibrios. ASM Press, Washington, DC.
20. **Thompson, J. D., D. G. Higgins, and T. J. Gibson.** 1994. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. Nucleic Acids Res. **22:**4673–4680.
21. **Zeigler, D. R.** 2003. Gene sequences useful for predicting relatedness of whole genomes in bacteria. Int. J. Syst. Evol. Microbiol. **53:**1893–1900.