

RESEARCH ARTICLE

Open Access

Inflammatory mediators in breast cancer: Coordinated expression of TNF α & IL-1 β with CCL2 & CCL5 and effects on epithelial-to-mesenchymal transition

Gali Soria^{1†}, Maya Ofri-Shahak^{1†}, Ilana Haas², Neora Yaal-Hahoshen³, Leonor Leider-Trejo⁴, Tal Leibovich-Rivkin¹, Polina Weitzenfeld¹, Tsipi Meshel¹, Esther Shabtai⁵, Mordechai Gutman^{2,6} and Adit Ben-Baruch^{1*}

Abstract

Background: The inflammatory chemokines CCL2 (MCP-1) & CCL5 (RANTES) and the inflammatory cytokines TNF α & IL-1 β were shown to contribute to breast cancer development and metastasis. In this study, we wished to determine whether there are associations between these factors along stages of breast cancer progression, and to identify the possible implications of these factors to disease course.

Methods: The expression of CCL2, CCL5, TNF α and IL-1 β was determined by immunohistochemistry in patients diagnosed with: (1) Benign breast disorders (=healthy individuals); (2) Ductal Carcinoma *In Situ* (DCIS); (3) Invasive Ductal Carcinoma without relapse (IDC-no-relapse); (4) IDC-with-relapse. Based on the results obtained, breast tumor cells were stimulated by the inflammatory cytokines, and epithelial-to-mesenchymal transition (EMT) was determined by flow cytometry, confocal analyses and adhesion, migration and invasion experiments.

Results: CCL2, CCL5, TNF α and IL-1 β were expressed at very low incidence in normal breast epithelial cells, but their incidence was significantly elevated in tumor cells of the three groups of cancer patients. Significant associations were found between CCL2 & CCL5 and TNF α & IL-1 β in the tumor cells in DCIS and IDC-no-relapse patients. In the IDC-with-relapse group, the expression of CCL2 & CCL5 was accompanied by further elevated incidence of TNF α & IL-1 β expression. These results suggest progression-related roles for TNF α and IL-1 β in breast cancer, as indeed indicated by the following: (1) Tumors of the IDC-with-relapse group had significantly higher persistence of TNF α and IL-1 β compared to tumors of DCIS or IDC-no-relapse; (2) Continuous stimulation of the tumor cells by TNF α (and to some extent IL-1 β) has led to EMT in the tumor cells; (3) Combined analyses with relevant clinical parameters suggested that IL-1 β acts jointly with other pro-malignancy factors to promote disease relapse.

Conclusions: Our findings suggest that the coordinated expression of CCL2 & CCL5 and TNF α & IL-1 β may be important for disease course, and that TNF α & IL-1 β may promote disease relapse. Further *in vitro* and *in vivo* studies are needed for determination of the joint powers of the four factors in breast cancer, as well as analyses of their combined targeting in breast cancer.

* Correspondence: aditbb@tauex.tau.ac.il

† Contributed equally

¹Department of Cell Research and Immunology, George S. Wise Faculty of Life Sciences, Tel Aviv University, Israel

Full list of author information is available at the end of the article

Background

Breast cancer provides a typical example of an inflammation-linked malignant disease. Breast tumors are enriched with inflammatory constituents, including cells that are polarized to the tumor-promoting phenotype, and soluble factors. Cumulative findings of a large number of studies indicate that many of the inflammatory components present in the tumor microenvironment actively support breast cancer development and progression [1-4].

Recently, much attention has been given to the roles of inflammatory chemokines and cytokines in breast cancer, with emphasis on the chemokines CCL2 and CCL5, and the cytokines tumor necrosis factor α (TNF α) and interleukin 1 β (IL-1 β). While each component has been studied meticulously, too little consideration has been given to possible associations and interactions between the inflammatory mediators, and to their joint presence, as part of the inflammatory microenvironment, in breast cancer. The identification of the inflammatory setup prevailing in breast cancer may be instrumental in providing us with a better basis for the future design of improved therapeutic modalities, and of advanced diagnostic and prognostic tools.

In this study, we asked if associations exist between the inflammatory chemokines CCL2 & CCL5 and the inflammatory cytokines TNF α & IL-1 β in breast cancer. All four factors are expressed in breast tumors, they exert a diverse array of activities that support malignancy and they were shown to be directly involved in promoting tumor growth and metastasis in animal model systems of breast cancer (Refs [3,5-24] for CCL2 & CCL5, [25-59] for TNF α and IL-1 β). Out of the four factors, TNF α is of special interest because of reports showing that under specific circumstances it may have cytotoxic and anti-tumor effects in several malignant diseases [25,60,61].

The chemokines CCL2 and CCL5 are categorized as "inflammatory chemokines", and as such they are usually not constitutively expressed by normal tissue cells. Rather, their expression is induced by inflammatory insults that prevail in the course of immune functions, including by stimulation with TNF α and IL-1 β . The inter-connection between CCL2 & CCL5 and TNF α & IL-1 β in the immune setting suggests that similar interactions exist between these chemokines and cytokines also within the inflammatory context of breast tumors. It is possible that TNF α & IL-1 β and the two chemokines are interrelated in breast tumor cells, and that these four factors establish associations that may eventually contribute to tumor growth and metastasis.

To determine if this is indeed the case, we have initiated this study by determining the expression patterns of CCL2, CCL5, TNF α and IL-1 β in biopsy

sections of healthy individuals and in breast cancer patients at different progression stages of disease. While only negligible staining of the factors was detected in infiltrating leukocytes in all groups, there was substantial expression of all four factors in breast tumor cells. Therefore, we focused in this study on the identification of expression patterns of CCL2, CCL5, TNF α and IL-1 β in normal and malignant breast epithelial cells, and their potential contribution to disease course.

The results of our study indicate that significant associations exist between CCL2 & CCL5 and TNF α & IL-1 β in breast cancer, along different stages of disease course. Furthermore, the results of this study support the possibility that TNF α & IL-1 β play an important role in disease progression. These two cytokines were found to be highly persistent in tumors of patients suffering from relapsed disease. In addition, we found that they promoted processes that are required for disease progression and local recurrence, such as EMT which was induced mainly by TNF α .

The tumor-promoting activities of the inflammatory mediators CCL2 & CCL5 and TNF α & IL-1 β in breast cancer are not fully overlapping (references above), therefore their coordinated expression may eventually provide advantage to the developing and metastasizing tumor. This possibility emphasizes the need to determine the joint powers of the four factors together in breast cancer, by using *in vitro* and *in vivo* model systems. Moreover, our results suggest an important role for TNF α in breast malignancy, because the cytokine has substantial ability to promote progression-related events by inducing EMT processes in the tumor cells.

Methods

Patients

This study included 38 healthy individuals who were diagnosed with benign breast disorders that included fibrocystic changes, hyperplastic changes and benign tumors (this group is termed herein "Benign"). The study also included 88 breast cancer patients, who were divided to three groups: (1) 30 patients with DCIS. Since DCIS is a less malignant form of disease that rarely develops to IDC [62-64], the DCIS group included patients in whom disease has not progressed to IDC in the course of 5-10 years of follow up; (2) 23 patients with IDC, who remained disease free in the course of 5-10 years of follow up (IDC-no-relapse); (3) 35 patients with IDC who relapsed with metastases or local tumor/s, or who died of breast cancer, in 5-10 years of follow up (IDC-with-relapse).

The characteristics of the study patients who were diagnosed with DCIS, IDC-no-relapse and IDC-with-relapse are presented in Table 1. Tumor status (T1, etc.)

Table 1 The characteristics of the DCIS, IDC-no-relapse and IDC-with-relapse patients included in the study

Patient characteristics		DCIS		IDC - No Relapse		IDC - With Relapse	
		n = 30	(%)	n = 23	(%)	n = 35	(%)
Menopause status	Pre-menopause	3	(10.0%)	0	(0.0%)	7	(20.0%)
	Menopause	7	(23.3%)	0	(0.0%)	0	(0.0%)
	Post-menopause	8	(26.7%)	0	(0.0%)	25	(71.4%)
	Unknown	12	(40.0%)	23	(100.0%)	3	(8.6%)
Node status	N0	30	(100.0%)	14	(60.9%)	17	(48.6%)
	N1	0	(0.0%)	4	(17.4%)	14	(40.0%)
	N2	0	(0.0%)	1	(4.4%)	0	(0.0%)
	Unknown	0	(0.0%)	4	(17.4%)	4	(11.4%)
Tumor	T0	26	(86.7%)	0	(0.0%)	1	(2.9%)
	T1	0	(0.0%)	2	(8.7%)	14	(40.0%)
	T \geq 2	0	(0.0%)	19	(82.6%)	17	(48.6%)
	Unknown	4	(13.3%)	2	(8.7%)	3	(8.5%)
Grade	1	5	(16.7%)	6	(26.1%)	5	(14.3%)
	2	12	(40.0%)	9	(39.1%)	10	(28.6%)
	3	6	(20.0%)	5	(21.7%)	7	(20.0%)
	Unknown	7	(23.3%)	3	(13.1%)	13	(37.1%)
Radiotherapy	Yes	19	(63.4%)	17	(73.9%)	28	(80.0%)
	No	10	(33.3%)	5	(21.7%)	5	(14.3%)
	Unknown	1	(3.3%)	1	(4.4%)	2	(5.7%)
Chemotherapy	Yes	0	(0.0%)	17	(73.9%)	15	(42.9%)
	No	18	(60.0%)	5	(21.7%)	17	(48.6%)
	Unknown	12	(40.0%)	1	(4.3%)	3	(8.5%)
Her2-neu	Yes	3	(10.0%)	1	(4.3%)	6	(17.1%)
	No	1	(3.3%)	15	(65.3%)	26	(74.3%)
	Unknown	26	(86.7%)	7	(30.4%)	3	(8.6%)
ER	Yes	23	(76.7%)	20	(87.0%)	23	(65.7%)
	No	2	(6.7%)	2	(8.7%)	10	(28.6%)
	Unknown	5	(16.6%)	1	(4.3%)	2	(5.7%)
PR	Yes	17	(56.7%)	19	(82.6%)	19	(54.3%)
	No	7	(23.3%)	3	(3.0%)	13	(37.1%)
	Unknown	6	(20.0%)	1	(4.4%)	3	(8.6%)

was determined by the guidelines of the Cancer Staging Manual of the American Joint Committee on Cancer. The patients were treated and followed in Tel Aviv Sourasky Medical Center and in Meir Medical Center. The study was approved by the Helsinki Committees of both Centers as a retrospective study based on information obtained from files of unidentified patients (included in the category of studies which do not require informed consent from study patients).

Immunohistochemistry

In this study, we have used archived paraffin blocks of patients, all obtained at the time of diagnosis. Serial sections (5 μ m thick) were prepared from the blocks and processed for immunohistochemistry (IHC) by the

Pathology Departments of Tel Aviv Sourasky Medical Center and Meir Medical Center.

The biopsy sections were deparaffinized, dehydrated in xylene and graded alcohols, rinsed in PBS and stained by primary antibodies. All antibodies used in IHC were commercial monoclonal antibodies, and their ability to recognize their respective antigens was confirmed in ELISA assays (data not shown). Staining by primary monoclonal antibodies for human CCL5 (PeproTech, Rocky Hill, NJ; Cat# 500-M75) and to human CCL2 (R&D Systems, Minneapolis, MN; Cat# MAB679) was performed by two protocols that showed similar sensitivity and specificity: (1) Sections were incubated with hyaluronidase at 37°C for 1 hr, then treated with 3% H₂O₂ for 10 min at room temperature (RT). After

rinsing in PBS, non-specific binding was blocked by incubating the sections with normal goat serum at 37°C for 30 min. Then, the sections were stained over night with the primary antibodies at 4°C, except for sections of DCIS patients in which the staining was performed for 4 hr (the time was reduced in this case due to some background staining that was observed only in the DCIS biopsies stained by the different antibodies). (2) Sections were immersed in a staining dish containing citrate buffer at 95-100°C for 22 min, then cooled for 20 min. After washing with PBS, the sections were blocked with blocking solution (0.05% sodium azide and 0.05% Tween 20 in PBS). Next, the sections were incubated with the different primary antibodies for 1 hr at RT, blocked by 3% H₂O₂ for 10 min at RT, and washed thoroughly in PBS.

Staining for TNF α and IL-1 β was performed according to the second procedure detailed above, using monoclonal antibodies to human TNF α (PeproTech; Cat# 500-M26) and to human IL-1 β (Exalpa, Watertown, MA; Cat# L140M).

For all four factors (CCL2, CCL5, TNF α and IL-1 β), the staining by secondary antibodies was performed as follows: sections were washed thoroughly in PBS, stained with biotinylated anti broad spectrum secondary antibody, or alternatively, by anti mouse secondary antibody (both having similar sensitivity and specificity) for 10 min at RT. After additional washing in PBS, the sections were stained by HRP-streptavidin for 10 min at RT. Then, the sections underwent additional washings in PBS and were incubated with Diaminobenzidine for 10 min at RT, washed and counterstained by incubation with hematoxylin for 5 sec at RT. After washing in H₂O, the sections were dehydrated in graded alcohols and mounted to cover slides.

In all cases, negative controls included substitution of the primary antibodies by PBS, or by non-relevant isotype controls. The specificity of the antibodies against TNF α and IL-1 β was verified using mouse IgG1 as an isotype control for the antibodies (that shared the same isotype). Also, since the antibodies against CCL5 share the same isotype with the antibodies against CCL2, but there were biopsies that stained with one but not with the other, they served as internal controls for each other's specificity (In the 88 breast cancer patients: 9% of the biopsies stained for CCL2 but not for CCL5, and 25% of the biopsies stained for CCL5 but not for CCL2; In all other cases, the expression of CCL2 was coordinated with that of CCL5).

All slides were submitted to light microscopy, and the staining pattern in the cells was evaluated in a blind manner by a pathologist having expertise in breast cancer, and by a pathology-experienced researcher. The tumors were evaluated for %

chemokine/cytokine-positive cells in the tumors (0-100%) and intensity of expression of the chemokines/cytokines (1 = low intensity, 2 = medium intensity, 3 = high intensity). TNF α and IL-1 β were also evaluated for score of expression, determined as "% cytokine-positive cells in the tumors \times the intensity of cytokine expression in each of the tumors". In line with the evaluations of many other markers in IHC, a threshold for positive staining was set. Based on our past experience, non-specific staining (exhibited as rare/sporadic/random stainings) could account for up to 5% of the epithelial cells in the biopsy, therefore the cut-off for positivity was set on 5%.

The data on expression of Her2-neu, estrogen receptor α (ER) and progesterone receptor (PR) that are included in Table 1 were provided by Tel Aviv Sourasky Medical Center and by Meir Medical Center, where the patients were treated and followed.

Statistical analyses of immunohistochemistry

In this study, a comparison was made between the incidences of expression the different factors in normal breast epithelial duct cells of the Benign group and the tumor cells in DCIS, IDC-no-relapse and IDC-with-relapse groups (Fig. 2). This analysis was performed by Chi-square or two-tailed Fisher's exact test as applicable. Hochberg's GT2 method for multiple comparisons was applied for pair-wise comparisons between groups of patients.

The differences between the incidences of expression of the different factors in normal breast epithelial cells that were adjacent to tumor cells in the cancer biopsies, as compared to the tumor cells in the DCIS, IDC-no-relapse and IDC-with-relapse groups (Table 2), were determined by McNemar test (except for TNF α expression in "IDC-no-relapse" group, that because of limitations of the McNemar test was analyzed by Wilcoxon).

Chi-square or Fisher's exact test were used in order to determine the significance of associations between factors that belonged to "Group 1" (TNF α & IL-1 β) and factors that belonged to "Group 2" (CCL2 & CCL5) (Fig. 3).

Group means were compared using a one-way Analysis of Variance (Fig. 7 and Table 3). Whenever a significant effect of group was observed, pair-wise comparisons between groups were performed using the Ryan-Einot-Gabriel-Welsch Multiple Range Test. Levene's Test for Homogeneity of Variances was applied to compare dispersion between groups.

Logistic regression model was applied to assess the potential use of Her2-neu, ER and PR with cytokines as risk factors for disease relapse.

All statistical analyses were performed using the SAS for Windows version 9.1.3.

Table 2 The incidence of expression of CCL2, CCL5, TNF α and IL-1 β in tumor cells and in adjacent normal breast epithelial cells in DCIS and IDC patients

Expression of chemokines/cytokines in	Number of patients (%)			
	With CCL2 expression	With CCL5 expression	With TNF α expression	With IL-1 β expression
DCIS				
Breast malignant cells	11/27 (40.7%)	16/27 (59.2%)	13/27 (48.1%)	15/27 (55.5%)
Adjacent normal breast epithelial duct cells	5/27 (18.5%)	7/27 (25.9%)	3/27 (11.1%)	6/27 (22.2%)
p value	0.03	0.01	0.002	0.003
IDC-no-relapse				
Breast malignant cells	10/19 (52.6%)	15/19 (78.9%)	14/19 (73.7%)	13/19 (68.4%)
Adjacent normal breast epithelial duct cells	1/19 (5.3%)	1/19 (5.3%)	0/19 (0.0%)	1/19 (5.3%)
p value	0.003	0.0002	0.0001	0.0005
IDC-with-relapse				
Breast malignant cells	15/29 (51.7%)	17/29 (58.6%)	24/29 (82.7%)	25/29 (86.2%)
Adjacent normal breast epithelial duct cells	3/29 (10.3%)	2/29 (6.9%)	6/29 (20.7%)	8/29 (27.6%)
p value	0.001	0.0001	<0.0001	<0.0001

The expression CCL2, CCL5, TNF α and IL-1 β was determined by IHC in serial biopsy sections of primary tumors of the breast taken at the time of diagnosis, from patients diagnosed with DCIS, IDC-no-relapse, or IDC-with-relapse. In this analysis, the incidences of expression of the four factors were determined in the malignant cells and in adjacent normal breast epithelial cells, in the same biopsy. Therefore, this analysis included only patients in whom normal ducts and breast tumor cells were detected in the same biopsy: DCIS - n = 27; IDC-no-relapse - n = 19; IDC-with-relapse - n = 29. The incidence of CCL2, CCL5, TNF α and IL-1 β expression in the normal epithelium was not significantly different between DCIS and IDC-no-relapse, and also did not differ significantly between DCIS and IDC-with-relapse patients. The expression of the proteins was determined by antibodies whose binding specificity in IHC was verified.

Determination of EMT properties

The T47D and MCF-7 human breast carcinoma cell lines were grown in DMEM medium as described previously [20]. The growth medium was replaced by serum-deficient medium, and the cells were grown to confluency in the absence or in the presence of TNF α or IL-1 β for 72 hr.

E-cadherin expression was determined in live cells by mouse antibodies against human E-cadherin (Santa Cruz Biotechnology, Santa Cruz, CA), followed by FITC-conjugated antibodies against mouse IgG (Jackson ImmunoResearch Laboratories, West Grove, Pennsylvania). Baseline staining was obtained by adding the appropriate buffer to the cells instead of primary antibody, and by staining with non-relevant isotype-treated antibodies. Determination of vimentin expression was performed in methanol-treated cells by mouse antibodies against human vimentin (Santa Cruz Biotechnology), followed by FITC-conjugated antibodies against mouse IgG (Jackson ImmunoResearch Laboratories). Baseline staining was obtained by non-relevant isotype-matched antibodies. Stainings were determined with a Becton Dickinson FACSsort (Mountain View, CA) using the CellQuest software.

The expression of β -catenin was determined by specific antibodies, along with nuclei staining by DAPI. The cells were fixed by 8% paraformaldehyde for 15 min. Following treatment by 0.2% triton for 10 min, blocking was performed with PBS containing 2% BSA for 30 min. Primary mouse antibodies to human β -catenin were

then used (Santa Cruz Biotechnologies), followed by FITC-conjugated mouse antibodies against mouse IgG. Baseline staining was obtained by adding the appropriate buffer to the cells instead of primary antibody. Similarly fixed and permeabilized cells were incubated with phalloidin-Alexa 488 (Molecular Probes, PoortGebouw, The Netherlands) for detection of actin organization. Stained cells were detected by LSM 510 Meta confocal microscope (Carl Zeiss AG, Oberkochen, Germany).

In adhesion assays, TNF α -stimulated cells were trypsinized, and then plated in 96 well non-tissue culture plates for 2 hr at 37°C. Non-adherent cells were removed by washing in PBS. Detection of adherent cells was performed by an alkaline phosphatase assay, by the addition of alkaline phosphatase substrate buffer, containing 3 mg/ml p-nitrophenyl phosphate disodium (Sigma-Aldrich, St. Louis, MO). Following the addition of 1M NaOH, optical density was measured at 405 nm. Statistical analysis was performed by Student's *t* test.

Migration and invasion assays were performed in transwell migration chambers with 8- μ m pore size (Costar, Cambridge, MA). In both assays, MCF-7 cells were stimulated by TNF α for 48 hr, then added to the upper wells of the chamber with or without TNF α (in serum-free DMEM). In invasion assays, the upper wells were pre-coated with matrigel (20 μ g/ml in serum-free cold DMEM; BD Biosciences, Bedford, MA) for 1 hr at 37°C, then blocking was performed for additional 1 hr with 0.1% heat inactivated BSA (diluted in PBS). In both assays, the lower wells of the chamber included 500 μ l

Table 3 TNF α and IL-1 β prevalence in tumors of breast cancer patients

Cytokine	Group No.	Group name	% Positive cells in tumors (X \pm SD)	Score (X \pm SD)
TNF α	I	DCIS	67.3 \pm 24.7	134.8 \pm 82.4
	II	IDC-no-relapse	65.9 \pm 23.7	141.9 \pm 81.1
	III	IDC-with-relapse	85.0 \pm 6.8	191.0 \pm 70.3
		p value	0.0008	0.0331
IL-1 β	I	DCIS	69.3 \pm 21.6	117.6 \pm 80.1
	II	IDC-no-relapse	55.7 \pm 29.3	121.2 \pm 96.5
	III	IDC-with-relapse	84.8 \pm 11.5	206.2 \pm 76.2
		p value	<0.0001	0.0004

The table provides information on the prevalence of TNF α and IL-1 β expression in tumors of breast cancer patients, with analyses that were performed only on patients whose tumors expressed the cytokines: DCIS group = Group I: TNF α n = 15, IL-1 β n = 18; IDC-no-relapse group = Group II: TNF α n = 17, IL-1 β n = 15; IDC-with-relapse group = Group III: TNF α n = 29, IL-1 β n = 31. Each of the TNF α - and IL-1 β -positive tumors were analyzed for % positive cells in the tumors, intensity of expression of the cytokines, and score (as described in "Methods"). The table provides the X \pm SD of % cells positive for TNF α and IL-1 β , and the X \pm SD of the expression scores of TNF α and IL-1 β in the three groups of patients.

For each cytokine, group means were compared by one-way analysis of variance. Whenever a significant effect of group was observed, pair-wise comparisons between groups were performed. The results of the statistical analyses have shown that: (1) For % TNF α -positive cells, p = 0.0008: Group III was statistically different from Groups I and II. (2) For TNF α score, p = 0.0331: Paired comparisons did not identify specific significant differences between the groups of patients. (3) For % IL-1 β positive cells, p < 0.0001: All three groups were statistically different from each other. (4) For IL-1 β score, p = 0.0004: Group III was statistically different from Groups I and II.

of DMEM supplemented with 10% FCS. Following 21-23 hr of incubation, the cells on the upper surface of the filter were completely removed by wiping with a cotton swab. The filters were fixed (by methanol in migration assays and by 4% paraformaldehyde in invasion assays) and were stained with Diff-Quik (Dade Behring, Newark, DE). The cells were counted in high power fields (HPF) by light microscopy. Statistical analysis was performed by Student's *t* test.

Results

Patterns of expression of inflammatory chemokines and cytokines, determined by immunohistochemistry

Biopsy sections were obtained from the total of 126 individuals (as indicated in "Methods") at the time of diagnosis, and were stained by specific antibodies that recognized human CCL2, CCL5, TNF α or IL-1 β . Figure 1 displays representative examples of the staining patterns of the four proteins in biopsy sections of healthy individuals (the "Benign" group), and of DCIS, IDC-no-relapse and IDC-with-relapse patients. While the

expression of CCL2, CCL5, TNF α or IL-1 β was only minimally detected in infiltrating leukocytes of all groups of patients, the four factors were clearly observed in breast tumor cells, with mainly a cytoplasmic staining pattern. Based on the above, we have performed detailed analysis of the expression patterns of the four factors in breast epithelial cells, normal and malignant, in biopsy sections of healthy individuals and of breast cancer patients, as indicated below.

Protein expression in normal and malignant epithelial breast cells

Figure 2 provides information on the incidences of expression of CCL2, CCL5, TNF α and IL-1 β in normal breast epithelial cells of Benign patients, and in the tumor cells in patients diagnosed with DCIS, IDC-no-relapse and IDC-with-relapse. All four factors were minimally detected in normal breast epithelial duct cells in the healthy individuals of the Benign group (expression incidences ranging between 5.3% and 10.5%). A significant elevation was denoted in the expression of CCL2, CCL5, TNF α and IL-1 β in the tumor cells in all groups of cancer patients (DCIS, IDC-no-relapse and IDC-with-relapse), when compared to their expression in the normal breast cells in biopsies of the Benign patients, with significance value of p < 0.001 in all cases.

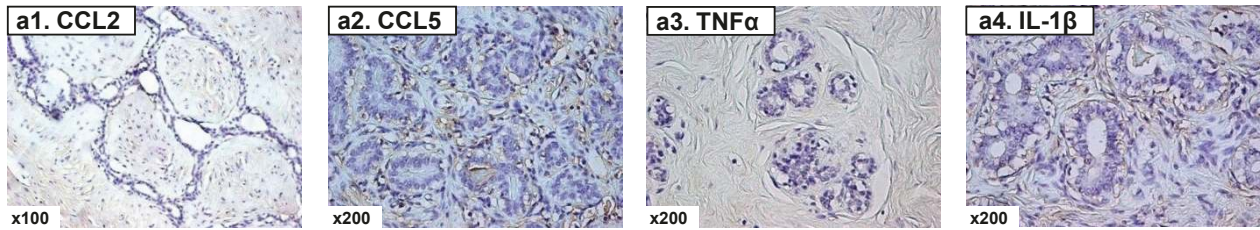
In parallel, we determined the expression of CCL2, CCL5, TNF α and IL-1 β in normal breast epithelial duct cells that were in proximity to the tumor cells in biopsies of patients with DCIS, IDC-no-relapse and IDC-with-relapse (Table 2). In these three groups of cancer patients, we found low incidence of expression of the inflammatory chemokines and inflammatory cytokines in the normal cells adjacent to the tumor cells, while they were expressed at significantly higher incidence in the tumor cells (incidences and p values are presented in Table 2). The incidence of CCL2, CCL5, TNF α and IL-1 β expression in the normal epithelium was not significantly different between DCIS and IDC-no-relapse, and also did not differ significantly between DCIS and IDC-with-relapse patients.

Taken together, these findings indicate that the expression of the inflammatory chemokines and inflammatory cytokines was acquired by breast epithelial cells upon their malignant transformation.

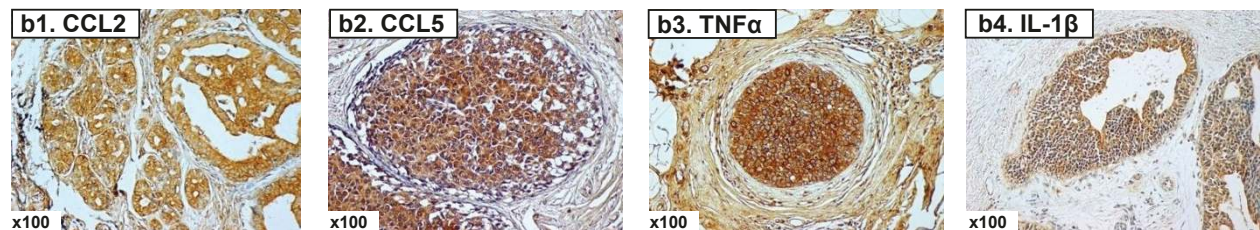
Protein expression along stages of disease progression

Next, we compared the three groups of breast cancer patients (DCIS, IDC-no-relapse, IDC-with-relapse) with respect to the incidence of expression of CCL2, CCL5, TNF α and IL-1 β in the tumor cells. The results of Figure 2 clearly show that the expression of the inflammatory chemokines CCL2 and CCL5 remained high and constant in the tumor cells of DCIS, IDC-no-relapse

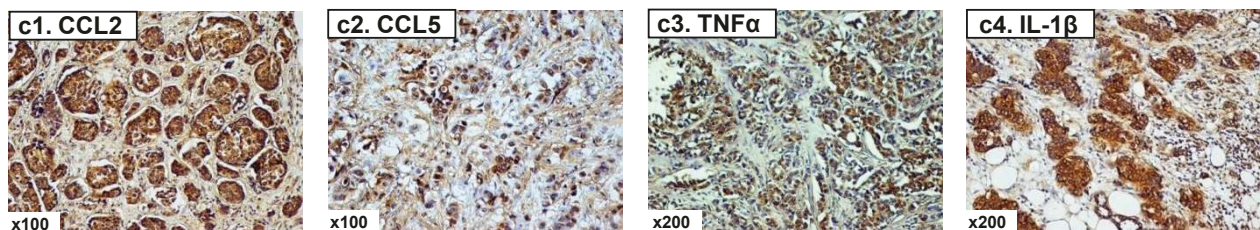
a. Benign Breast Diseases



b. Ductal Carcinoma *In Situ*



c. Invasive Ductal Carcinoma – No Relapse



d. Invasive Ductal Carcinoma – With Relapse

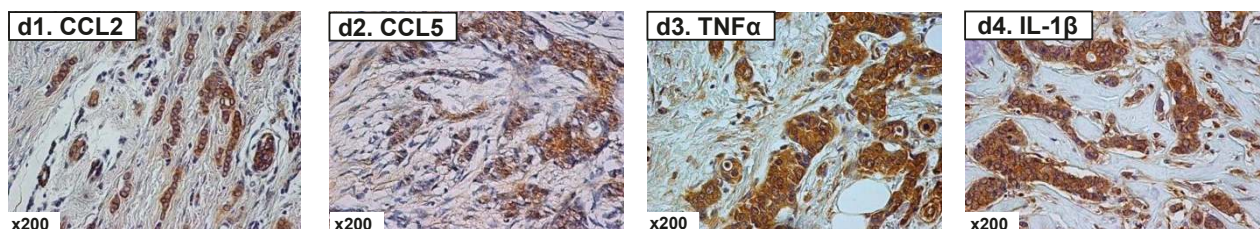
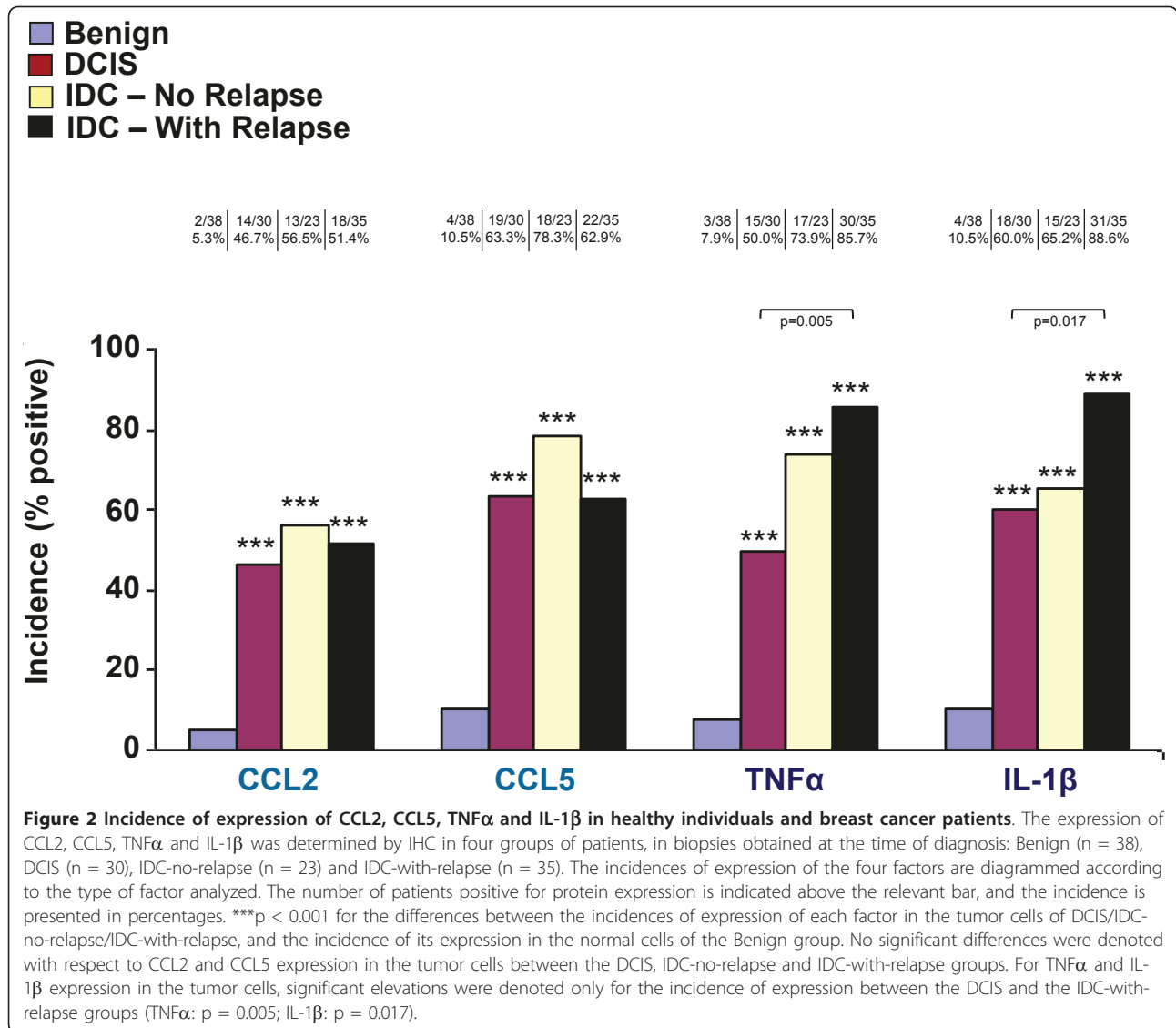


Figure 1 Expression patterns of CCL2, CCL5, TNF α and IL-1 β in healthy individuals and breast cancer patients. Representative examples of the expression of CCL2, CCL5, TNF α and IL-1 β in the different groups of patients included in the study, in biopsies obtained at the time of diagnosis. **(a1-a4)** Patients diagnosed with benign breast disorders. The pictures demonstrate the lack of staining of the four factors in the normal breast epithelial cells, as denoted in the majority of patients included in this group. **(b1-b4)** DCIS patients. The pictures demonstrate positive staining of the four factors in the malignant lesions, as denoted in the majority of patients included in this group. **(c1-c4)** IDC-no-relapse patients. The pictures demonstrate positive staining of the four factors in the tumor cells, as denoted in the majority of patients included in this group. **(d1-d4)** IDC-with-relapse patients. The pictures demonstrate positive staining of the four factors in the tumor cells, as denoted in the majority of patients included in this group. (a1, b1, c1, d1) CCL2 staining; (a2, b2, c2, d2) CCL5 staining; (a3, b3, c3, d3) TNF α staining; (a4, b4, c4, d4) IL-1 β staining. The expression of the proteins was determined by IHC using specific antibodies, whose specificity in IHC was verified. The values of photo magnification are indicated in the left bottom corner of each of the pictures.



and IDC-with-relapse patients. No significant differences were denoted between DCIS, IDC-no-relapse and IDC-with-relapse patients, with respect to the incidence of CCL2 and CCL5 expression in the tumor cells.

A high incidence of expression in the tumor cells was also denoted for TNFα and IL-1β in the three groups of cancer patients: DCIS, IDC-no-relapse and IDC-with-relapse. The results of Figure 2 indicate that the incidence of TNFα expression was 50% in DCIS patients and was significantly increased to 85.7% in IDC-with-relapse patients (p = 0.005); Similarly, the incidence of IL-1β was significantly raised from 60% in DCIS to 88.6% in IDC-with-relapse patients (p = 0.017). No other significant differences were found between the different groups of breast cancer patients with respect to TNFα and IL-1β expression in the tumor cells (namely: IDC-no-relapse vs. DCIS; IDC-no-relapse vs. IDC-with-relapse).

Together, these findings indicate that there was high incidence of CCL2, CCL5, TNFα and IL-1β expression in the tumor cells in the three groups of cancer patients, and that the expression of TNFα and IL-1β was further elevated in the IDC-with-relapse group. The rare expression of all four factors in normal breast epithelial cells, and the high incidence of their expression in tumor cells in biopsies of all groups of cancer patients, indicate that the expression of the inflammatory chemokines CCL2 & CCL5 is coordinated with that of the inflammatory cytokines TNFα & IL-1β along stages of tumor development and progression in breast cancer.

Dependence between the inflammatory chemokines and the inflammatory cytokines along disease progression

Based on the above results, we asked whether there are associations between the inflammatory chemokines

CCL2 & CCL5 and the inflammatory cytokines TNF α & IL-1 β in patients representing different stages along the progression process of breast cancer. To this end, we divided the four factors to two groups, based on their published functional interactions in breast tumor cells: Group 1 - The inflammatory cytokines TNF α & IL-1 β , which promote the release of CCL2 & CCL5 by breast tumor cells; Group 2 - The inflammatory chemokines CCL2 & CCL5, whose release by malignant breast cells is increased by TNF α and IL-1 β (Please refer to Additional Files 1 and 2, as well as to references [65-70]). These analyses were performed in the tumor cells in the three groups of cancer patients: DCIS, IDC-no-relapse and IDC-with-relapse.

When we analyzed the DCIS group, we found that factors belonging to Group 1 (TNF α & IL-1 β) "tended" to be expressed in the same biopsies in which the factors of group 2 were expressed (CCL2 & CCL5) (Figure 3a). For example, the study of CCL2 showed that CCL2 was co-expressed with TNF α and IL-1 β in 33.3% of the DCIS biopsies. In contrast, in only 10% of the DCIS patients CCL2 was detected without co-expression of TNF α and IL-1 β . This finding indicates that there was a preferential expression of CCL2, as a representative of Group 2, with factors that belonged to Group 1, namely TNF α & IL-1 β .

Similar analyses that were performed for the other three factors in DCIS patients - CCL5, TNF α and IL-1 β - have shown similar trends to those obtained for CCL2, namely that the inflammatory chemokines "favored" co-expression with the inflammatory cytokines, and *vice versa*. Accordingly, the statistical analyses that were performed in DCIS patients demonstrated that the factors of Group 1 were significantly associated with the factors of Group 2 ($p = 0.002$), indicating that in this group of patients there were defined associations between the inflammatory chemokines and the inflammatory cytokines.

Thereafter, analyses that were performed for patients diagnosed with IDC-no-relapse have shown that the expression of CCL2 & CCL5 was coordinated with that of TNF α & IL-1 β (Figure 3b). This was substantiated by significant associations that were detected between factors of Group 1 (TNF α & IL-1 β) and Group 2 (CCL2 & CCL5), with $p = 0.036$.

A somewhat different pattern of expression was obtained for IDC-with-relapse patients (Figure 3c). As in the DCIS and IDC-no-relapse groups, the incidence of CCL2 and CCL5 in the IDC-with-relapse group remained at the level of 50-65%; however, the incidence of TNF α and IL-1 β expression in the IDC-with-relapse group was increased to 85-90% (Figure 2). Therefore, in many of the patients of this group, the expression of TNF α & IL-1 β was coordinated with

that of CCL2 & CCL5, but in others the expression of TNF α & IL-1 β was not accompanied by expression of CCL2 & CCL5. This has led to insignificant associations ($p = 0.627$) between Group 1 (the inflammatory cytokines TNF α & IL-1 β) and Group 2 (the inflammatory chemokines CCL2 & CCL5) in the IDC-with-relapse group.

To conclude, in patients that belonged to the DCIS and IDC-no-relapse groups, the expression of the inflammatory chemokines was coordinated and significantly associated with that of the inflammatory cytokines. In contrast, in the IDC-with-relapse group there was sub-population of patients in which the expression of CCL2 & CCL5 did not accompany the expression of TNF α & IL-1 β . In this group of IDC-with-relapse patients, the expression of TNF α & IL-1 β was further elevated and was highly prevalent, supporting an important role for TNF α and IL-1 β in disease progression and recurrence. Furthermore, these results suggest that in this specific sub-group of IDC-with-relapse patients, the two cytokines are coordinated with other tumor-supporting factors that replace CCL2 & CCL5 in promoting disease relapse.

The persistence of inflammatory cytokines in breast tumors may promote disease progression and recurrence, e.g. via epithelial-to-mesenchymal transition

The findings on the elevated expression of TNF α and IL-1 β in the IDC-with-relapse group suggest that these two cytokines support disease progression. In addition to the tumor-promoting activities of TNF α and IL-1 β that have already been described, it is possible that these two cytokines also have functional interactions with CCL2 and CCL5, by that promoting disease course. In line with this possibility, our findings have shown that TNF α and IL-1 β up-regulated the release of CCL2 and CCL5 by breast tumor cells (Additional Files 1 and 2; also suggested by findings of references [65-70]). Such a cross-talk may lead to up-regulated expression of CCL2 and CCL5 in tumors expressing TNF α and/or IL-1 β , thus possibly leading to increased tumor-supporting activities of the two chemokines in breast tumors (see Discussion, below).

In addition to their ability to promote the release of CCL2 and CCL5 by the tumor cells, it is possible that TNF α and IL-1 β act directly to elevate processes that are required for local recurrence or metastasis formation, which identify the IDC-with-relapse group of patients. Recent studies indicate that acquisition of self-renewal properties that are required for formation of recurrent local tumors, as well as metastasis formation, are promoted by Epithelial-to-Mesenchymal Transition (EMT) processes [71-77]. Based on the above, we determined the possibility that TNF α and IL-1 β promote

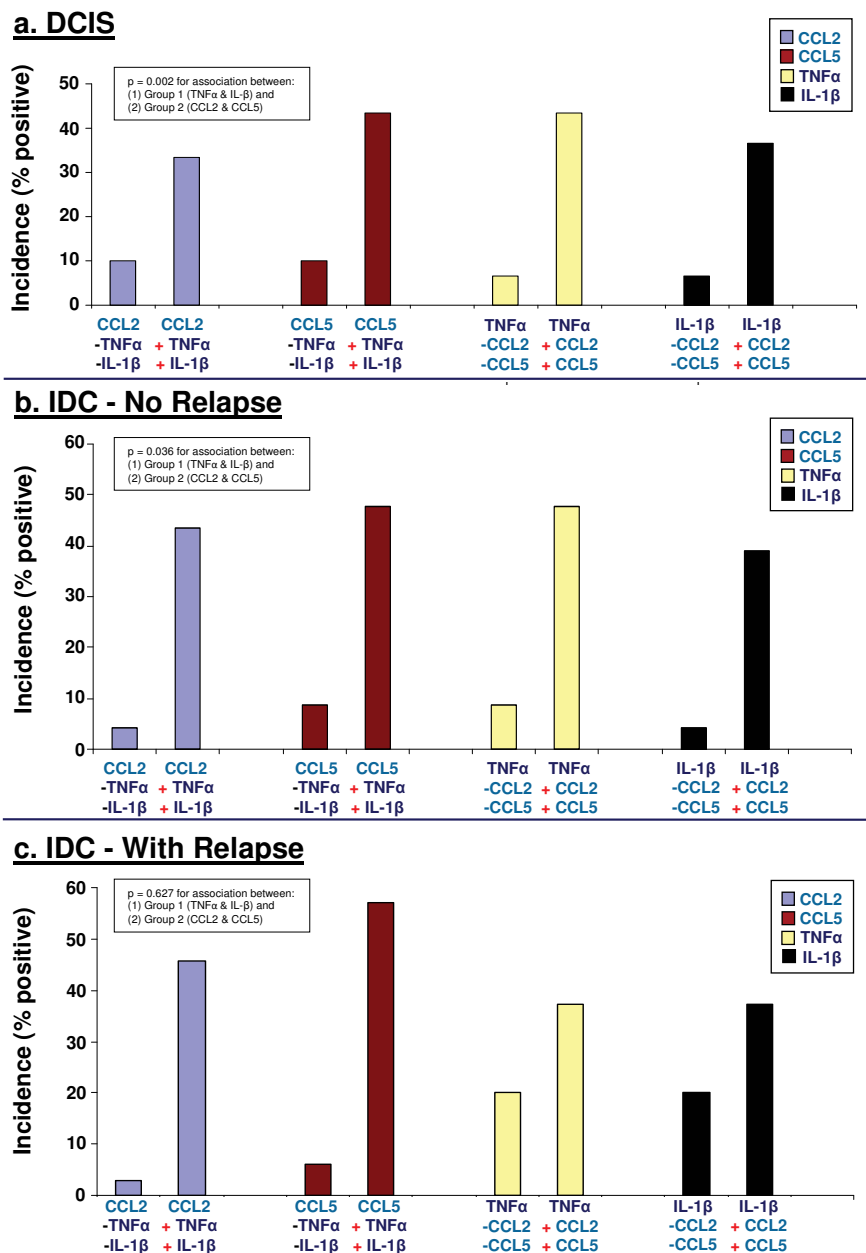


Figure 3 The associations between the inflammatory chemokines CCL2 & CCL5 and the inflammatory cytokines TNFα & IL-1β, in breast cancer patients. The analyzed factors were sub-divided to two groups: Group 1 - The inflammatory cytokines TNFα & IL-1β; Group 2 - The inflammatory chemokines CCL2 & CCL5; Detailed explanation of graphs a, b and c is provided further below. **(a)** In DCIS patients, p = 0.002 for associations between Group 1 and Group 2. **(b)** In IDC-no-relapse patients, p = 0.036 for associations between Group 1 and Group 2. **(c)** In IDC-with-relapse patients, p = 0.627 for associations between Group 1 and Group 2. Explanation: In each group of breast cancer patients (DCIS, IDC-no-relapse, IDC-with-relapse), the graphs show the following: (1) CCL2: The graph shows the incidence of CCL2 expression when it was co-expressed with TNFα and IL-1β in the same biopsy [presented in the graph as “CCL2 (+ TNFα) (+ IL-1β)”), as compared to the incidence of CCL2 expression when it was NOT co-expressed with TNFα and IL-1β in the same biopsy [presented in the graph as “CCL2 (- TNFα) (- IL-1β)”]; (2) CCL5: The graph shows the incidence of CCL5 expression when it was co-expressed with TNFα and IL-1β in the same biopsy [presented in the graph as “CCL5 (+ TNFα) (+ IL-1β)”), as compared to the incidence of CCL5 expression when it was NOT co-expressed with TNFα and IL-1β in the same biopsy [presented in the graph as “CCL5 (- TNFα) (- IL-1β)”]; (3) TNFα: The graph shows the incidence of TNFα expression when it was co-expressed with CCL2 and CCL5 in the same biopsy [presented in the graph as “TNFα (+ CCL2) (+ CCL5)”), as compared to the incidence of TNFα expression when it was NOT co-expressed with CCL2 and CCL5 in the same biopsy [presented in the graph as “TNFα (- CCL2) (- CCL5)”]; (4) IL-1β: The graph shows the incidence of IL-1β expression when it was co-expressed with CCL2 and CCL5 in the same biopsy [presented in the graph as “IL-1β (+ CCL2) (+ CCL5)”), as compared to the incidence of IL-1β expression when it was NOT co-expressed with CCL2 and CCL5 in the same biopsy [presented in the graph as “IL-1β (- CCL2) (- CCL5)”];

EMT processes in the tumor cells, thus pushing forward disease recurrence and progression.

To investigate this possibility, we tested the ability of TNF α and IL-1 β to induce in the tumor cells properties that are typical of EMT [71-77]. In these experiments, we used the T47D and MCF-7 cells, which represent a non-advanced stage of breast malignancy [78-82], and thus can serve as an appropriate platform for induction of EMT processes that are associated with more progressed disease.

First, we stimulated the cells by TNF α and IL-1 β (please see "Note" in legend to Figure 4 on cytokine concentrations used in these analyses), and determined the ability of the cytokines to induce reduction in the membranous expression of E-cadherin, an adhesion molecule which is important in establishing cell-to-cell contacts. The results of Figures 4a and 4b demonstrate that TNF α potently induced a very typical property of EMT, namely reduction in E-cadherin expression at the cell membrane of the tumor cells. This activity of TNF α was induced in both the T47D and the MCF-7 cells in a dose-dependent manner. In parallel, stimulation by IL-1 β has also led to reduced expression of E-cadherin at the plasma membrane, but only in the T47D cells (Figure 4c).

Based on these results and on additional preliminary findings (data not shown), we suspected that TNF α was a more potent inducer of EMT than IL-1 β , and that its effects were more prominent on MCF-7 cells than on T47D cells. Therefore in depth analyses were continued with TNF α , analyzed on MCF-7 cells only. Further analyses indicated that TNF α has led to substantial decrease in the expression of β -catenin at the cell membrane of the tumor cells (Figure 4d), an event which is typical of EMT. Also, the stimulation by the cytokine has promoted the expression of vimentin, a mesenchymal feature whose expression is elevated in cells undergoing EMT (Figure 5a). In parallel, the tumor cells have acquired cellular protrusions, accompanied by extensive re-organization of the actin cytoskeleton (Figures 5b and 5c, respectively).

In addition to the above, TNF α has induced in the tumor cells elevated adhesion to substrate, a step necessary for establishment of productive metastases at remote organs (Figure 6a). Importantly, following TNF α stimulation, the cells have acquired the most critical and metastasis-relevant property of EMT, namely increased migratory and invasive properties (Figures 6b and 6c), by that substantiating the ability of the cytokine to induce EMT in breast tumor cells.

Taken together, the above results indicate that TNF α , and possibly also IL-1 β - although to a lower extent - induced EMT properties in the tumor cells. Of note, similar activities of the two cytokines were obtained in

an unrelated ongoing study in our laboratory, performed on a different set of breast cells (data not shown; manuscript in preparation), further substantiating the EMT-promoting activities of TNF α and IL-1 β . Also, preliminary analyses that we have performed indicated that CCL2 and CCL5 were not capable of inducing EMT-related processes in the breast tumor cells (data not shown).

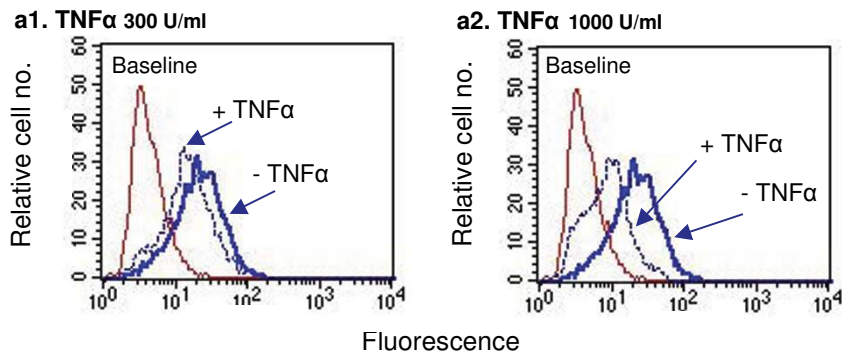
Since the potent EMT-promoting activities of TNF α may push forward processes of increased metastasis formation and disease recurrence, we further addressed the characteristics of TNF α activity. In view of reports showing that the EMT phenotype may be transient and needs continuous exposure to EMT-inducing factors [72], we asked if induction of EMT by TNF α required constant stimulation by the cytokine. To determine this issue, we have taken cells in which EMT was induced by TNF α (as indicated by reduced expression of E-cadherin at the cell membrane), and extended their growth for additional 72 hr with or without TNF α . The results of Figure 7 clearly indicate that the EMT phenotype was reversible if the cells were deprived of TNF α , while in cells that were grown continuously with the cytokine the EMT phenotype was preserved.

The above findings suggest that breast tumor cells benefit from high and constant presence of TNF α , because the persistence of the cytokine may induce EMT processes that lead to progressed and recurred disease. In such a case, we would expect that TNF α would be more persistent in tumors of IDC patients who relapsed with metastases or local tumors, compared to IDC-no-relapse or to DCIS patients.

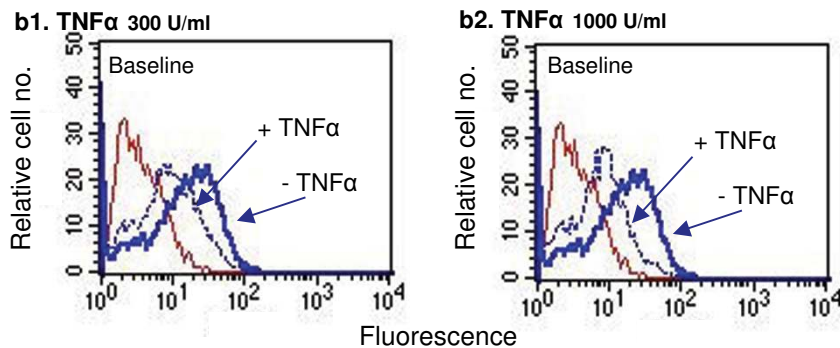
To find out if this was indeed the case, we determined in each of the three groups of breast cancer patients (DCIS, IDC-no-relapse, IDC-with-relapse) the parameters of % TNF α -positive cells in the tumors, and the score of TNF α expression. The results presented in Figure 8a and in Table 3 demonstrate that there was a significant elevation in % TNF α -positive cells/patient in the IDC-with-relapse group when it was compared to DCIS and to the IDC-no-relapse groups, where $p = 0.0008$ for % TNF α -positive cells, and $p = 0.0331$ for TNF α score (as indicated in the legend to Table 3).

Of note, similar analyses that were performed for IL-1 β have also shown highly significant persistence of this cytokine in tumors of IDC-with-relapse patients, with $p < 0.0001$ for % IL-1 β -positive cells, and $p = 0.0004$ for IL-1 β score (Figure 8b and Table 3). These results suggest that persistence of IL-1 β in the tumors is also important for driving forward processes of progression and recurrence. Based on our data, these activities of IL-1 β are not necessarily related to EMT, because this cytokine was not as effective in this function as TNF α . Rather, IL-1 β may acquire activities that are related for

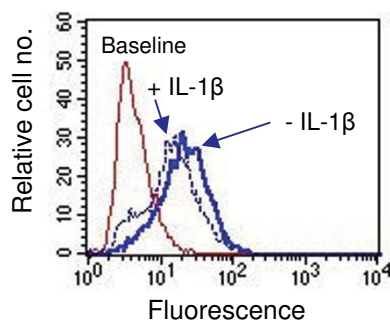
a. E-cadherin in T47D cells - TNF α



b. E-cadherin in MCF-7 cells - TNF α



c. E-cadherin – IL-1 β – 500 pg/ml



d. β -catenin

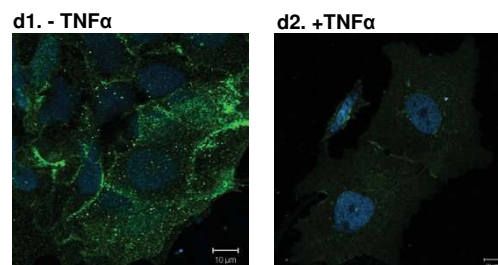
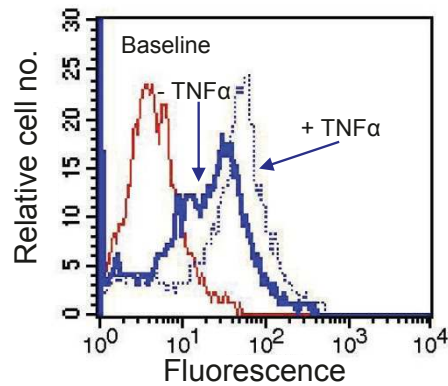
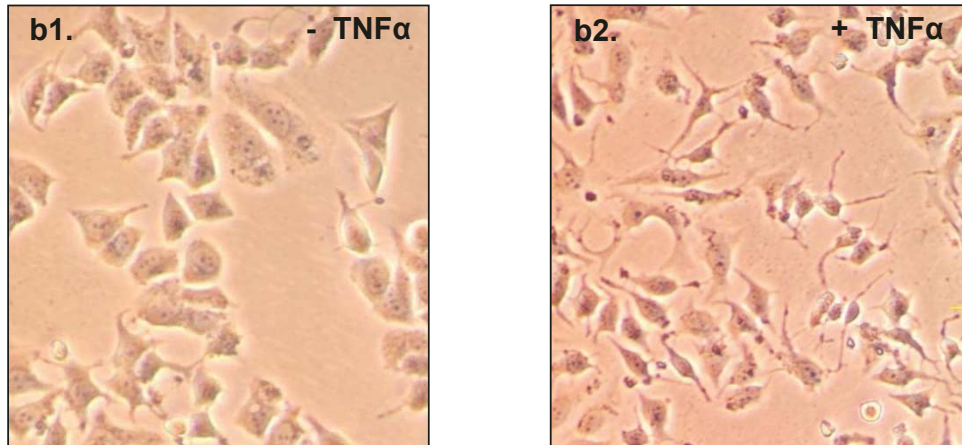


Figure 4 Following stimulation by the inflammatory cytokines (mainly TNF α), the tumor cells show typical characteristics of EMT. The T47D and MCF-7 human breast carcinoma cells were stimulated by TNF α and IL-1 β for 72 hr. Note: The cytokine concentrations were chosen based on analyses in which their effects on the tumor cells were tested in a dose-dependent manner (data not shown). **(a)** Stimulation of T47D cells with 300 U/ml TNF α (a1) or with 1000 U/ml TNF α (a2). The membranous expression of E-cadherin was determined in live cells by flow cytometry. The figure presents the results of a representative experiment of $n = 3$, all showing similar results. **(b)** Stimulation of MCF-7 cells with 300 U/ml TNF α (b1) or 1000 U/ml TNF α (b2). The membranous expression of E-cadherin was determined in live cells by flow cytometry. The figure presents the results of a representative experiment of $n > 3$, all showing similar results. **(c)** T47D human breast carcinoma cells were stimulated by IL-1 β (500 pg/ml). The membranous expression of E-cadherin was determined in live cells by flow cytometry. The figure presents the results of a representative experiment of $n = 3$, all showing similar results. **(d)** MCF-7 cells were stimulated by 1000 U/ml TNF α , and β -catenin expression (green) was determined by confocal analyses, in fixed cells. Nuclei are shown by blue DAPI staining. The figure presents the results of a representative experiment of $n = 3$, all showing similar results.

a. Vimentin



b. Cellular protrusions



c. Actin re-organization

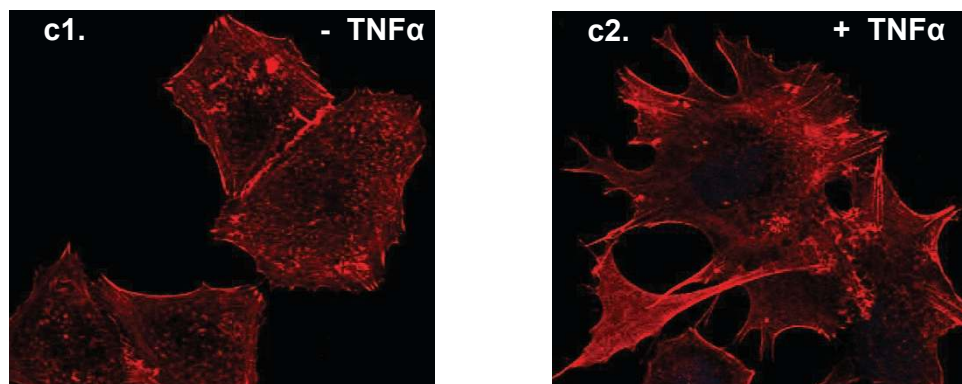


Figure 5 Following stimulation by TNF α , breast tumor cells acquire a mesenchymal phenotype which is accompanied by protrusive characteristics. MCF-7 breast carcinoma cells were stimulated by TNF α (1000 U/ml) for 72 hr. Thereafter, the following analyses were performed: **(a)** Vimentin expression, determined by specific antibodies in methanol-treated cells and analyzed by flow cytometry. The figure presents the results of a representative experiment of $n = 3$, all showing similar results. **(b)** Formation of cellular protrusions, determined by light microscopy. The figure presents the results of a representative experiment of $n > 3$, all showing similar results. **(c)** Actin polymerization, determined by phalloidin staining, analyzed by confocal microscopy. The figure presents the results of a representative experiment of $n = 3$, all showing similar results.

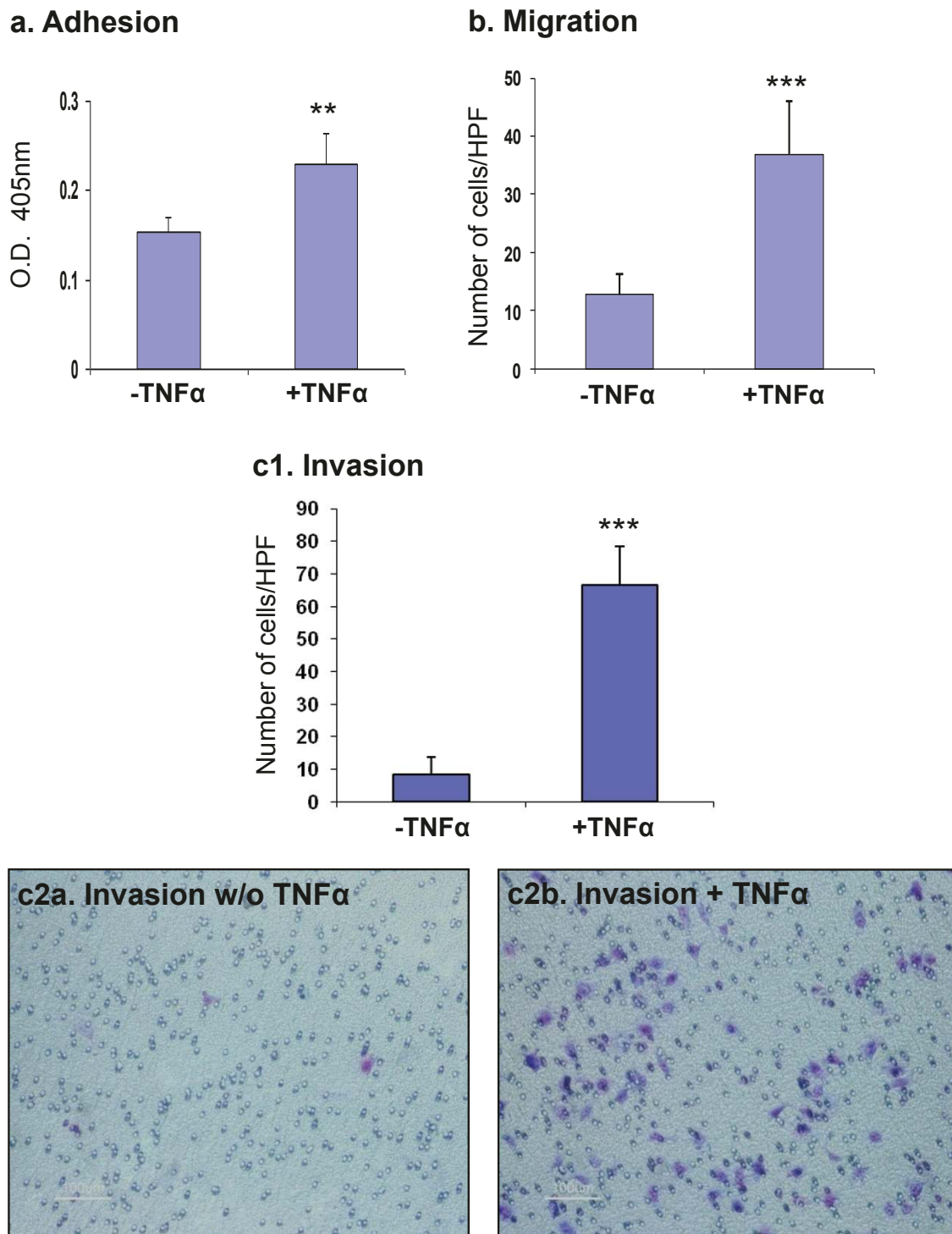


Figure 6 Following stimulation by TNF α , breast tumor cells acquire increased adhesive, migratory and invasive properties. MCF-7 breast carcinoma cells were stimulated by TNF α (1000 U/ml). Thereafter, the following analyses were performed: **(a)** Following 72 hr stimulation by TNF α , adhesion to substrate was determined by alkaline phosphatase assay. ** $p = 0.005$ for the difference between cells that were stimulated or not stimulated by TNF α . **(b, c)** Determination of the ability of the cells to perform migratory and invasive activities, in response to serum. Following 48 hr stimulation by TNF α (1000 U/ml), migration and invasion assays were performed in transwells for 21-23 hr with or without TNF α stimulation. **(b)** Migration. *** $p < 0.001$ for TNF α -stimulated cells vs. non-stimulated cells. **(c1)** Invasion counts. *** $p < 0.001$ for TNF α -stimulated cells vs. non-stimulated cells. **(c2)** Invasion as demonstrated in light microscopy. HPF = High Power Field. In all parts of the figure, experiments are representatives of $n = 3$.

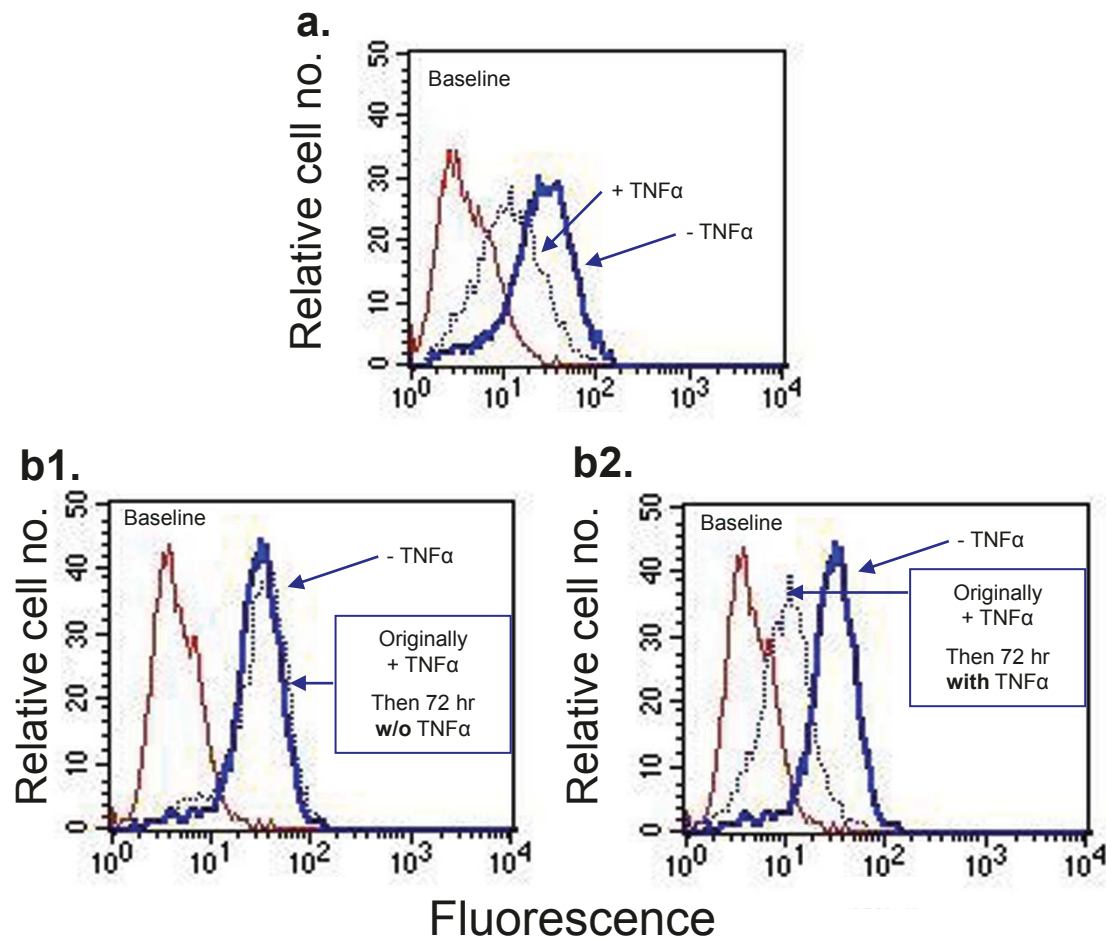


Figure 7 In order to induce EMT, TNF α needs to be constantly present in vicinity to the tumor cells. Determination of the reversibility of the TNF α -induced EMT effects. **(a)** MCF-7 cells were stimulated by TNF α (1000 U/ml) for 72 hr, followed by determination of membranous expression of E-cadherin in live cells, using flow cytometry. **(b)** The non-stimulated cells of part (A) were grown for additional 72 hr without TNF α . In contrast, the previously-stimulated cells (by TNF α) were grown for additional 72 hr without (b1) or with (b2) TNF α (1000 U/ml). Thereafter, the membranous expression of E-cadherin was determined in live cells by flow cytometry. The experiment is a representative of n = 3.

example to elevated angiogenesis, as previously suggested [28,31,83]. In addition, it is possible that IL-1 β acts jointly with other pro-malignancy factors to promote disease progression and recurrence. Such additional factors could be related to the response of the tumor cells to growth factors that are found at the tumor microenvironment, or to hormonal stimulation.

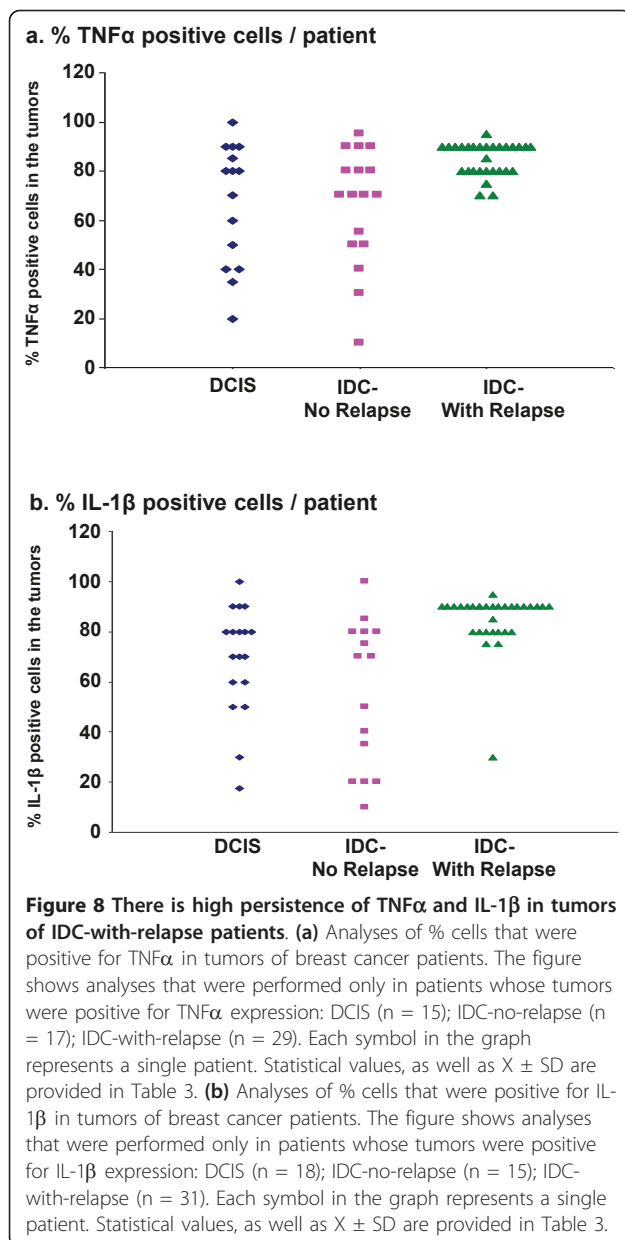
Supporting this possibility are analyses that we have performed, using the data obtained on the expression of Her2-neu, ER and PR in the patients. In this analysis, we have compared between the IDC-no-relapse and the IDC-with-relapse groups. Statistical analysis that was performed for IL-1 β , taking into account the status of Her2-neu, ER and PR, all combined (currently each of these three factors is used in the clinic as a predictor of disease progression and recurrence in breast cancer), indicated that IL-1 β was a significant risk factor for

disease relapse: p = 0.0402, with 95% confidence limits of 1.098 and 59.93 and odds ratio 1 value of 8.112 (also, of the different clinical markers, PR was statistically different between the IDC-no-relapse and IDC-with-relapse groups, with p = 0.0391). Similar analyses performed for TNF α , CCL2 and CCL5 did not reveal significant positive correlation with disease progression.

The findings on IL-1 β strongly support the possibility that when the tumor-derived and/or microenvironment-derived characteristics have higher cancer-promoting properties, IL-1 β can further promote progression-related processes in breast cancer.

Discussion

In this study, we have identified the relationships between inflammatory chemokines and cytokines in breast cancer along different stages of disease, and have



analyzed the potential roles of TNF α and IL-1 β in promoting breast cancer progression. The results of this study shed light on the inflammatory setup in breast cancer, and provide three novel findings:

(1) The expression of the inflammatory chemokines CCL2 & CCL5 and of the inflammatory cytokines TNF α & IL-1 β is coordinated in breast cancer

To date, the relationships between inflammatory chemokines and inflammatory cytokines in breast cancer were not elucidated, and the associations between them during the process of breast cancer development and progression were not analyzed. At this setting, the findings

of our study are important because they indicate that the expression of the inflammatory chemokines CCL2 & CCL5 and of the inflammatory cytokines TNF α & IL-1 β is coordinated in crucial stages along the process of disease progression. Moreover, the expression of all four inflammatory factors is minimal in normal breast epithelial cells, and is simultaneously acquired by the cells once malignant transformation has taken place, namely from the DCIS stage and on. These results suggest that events dictated by genetic/epigenetic alterations in the tumor cells, or by the microenvironment, lead to a synchronized up-regulation in the expression of several inflammatory mediators together, by transformed breast epithelial cells.

The coordinated expression of CCL2 & CCL5 and of TNF α & IL-1 β along stages of breast cancer development and progression is important because the activities of the four factors are not fully overlapping (Refs [3,5-24] for CCL2 & CCL5, [25-59] for TNF α and IL-1 β). Therefore, it is possible that each of these factors contributes its own share to disease course, alongside with the others. Together, the coordinated presence of CCL2 & CCL5 and of TNF α & IL-1 β may support malignancy, possibly also due to spatio-temporal cross-interactions between them. Indeed, our findings (Additional Files 1 and 2) and published studies [65-70] indicate that TNF α and IL-1 β up-regulate the release of CCL2 and CCL5 by breast tumor cells. Furthermore, recent findings obtained in our studies suggest that TNF α and IL-1 β exert tumor-promoting activities that are connected to the ability of CCL2 and CCL5 to function as cancer-supporting factors. Consequently, it is possible that the pro-tumorigenic activities of the inflammatory cytokines and of the inflammatory chemokines depend on each other, and/or are complementary to one another. This possibility emphasizes the need to determine whether combined inhibition of all factors together will lead to improved limitation of tumor growth.

Moreover, since our findings indicate that the coordinated array of cytokines and chemokines takes action in about 50-70% of the patients (depending on the group), it is possible that these factors can act along with other inflammatory and tumor-supporting factors to promote tumor growth and metastasis, as further discussed below.

(2) The expression of TNF α & IL-1 β is further increased in the IDC-with-relapse group of patients

In the group of IDC-with-relapse patients, the incidence of CCL2 & CCL5 expression was similarly high to that of DCIS and IDC-no-relapse group (50-65%), while that of TNF α & IL-1 β has reached about 85-90%. Therefore, while the expression of CCL2 & CCL5 was coordinated with TNF α & IL-1 β in many of the patients belonging to this group, there was a sub-population of patients in

which the expression of TNF α & IL-1 β was not accompanied by CCL2 & CCL5. While we need to understand the reasons for this lack of coordination in this specific subgroup of patients (please see discussion below), this observation indicates that TNF α & IL-1 β are required, and play important roles in disease progression, and suggests that they cooperate with other factors at the tumor microenvironment that substitute for the lack of CCL2 & CCL5.

Of interest in this respect are the reasons for the lack of expression of CCL2 & CCL5 in the specific sub-population of IDC-with-relapse patients who do express TNF α & IL-1 β . This lack of coordination between CCL2 & CCL5 and TNF α & IL-1 β in this sub-population is intriguing mainly because TNF α and IL-1 β were found to stimulate the release of CCL2 and CCL5 by breast tumor cells. The explanation could be given, at least partly, by consideration of the extent to which receptors for TNF α and IL-1 β are expressed by the tumor cells, and of which type. Although the expression of TNF α & IL-1 β receptors by breast tumor cells was already documented [29,31,37,43,44], it is possible that in this specific sub-group of the IDC-with-recurrence patients, the tumor cells do not express the required receptors for TNF α & IL-1 β , or express non-signaling receptors. Under such conditions, TNF α & IL-1 β would not act on the tumor cells to promote the release of CCL2 & CCL5, leading to lack of associations between the two cytokines and CCL2 & CCL5.

Accordingly, in our future investigations we will perform detailed analysis of the expression of receptors for the two cytokines (e.g. TNFR1, TNFR2, IL-1RI) in the course of disease progression in breast cancer. Such an analysis may enable us to better identify the basis for the patterns of expression of the inflammatory cytokines and chemokines in different stages of disease.

(3) Tumors of IDC-with-relapse patients express persistent and high levels of TNF α & IL-1 β , which may contribute to disease recurrence and progression, e.g. via EMT

As indicated above, the incidence of TNF α & IL-1 β was further increased in IDC patients in whom disease has relapsed, suggesting that these two cytokines push forward processes of tumor progression and recurrence. Our findings support this possibility because they indicate that TNF α , and to a lower extent also IL-1 β , induce EMT properties in the tumor cells.

In addition, our findings demonstrate that in order to undergo EMT, the tumor cells had to be constantly stimulated by TNF α . These results suggest that *in situ*, EMT processes would be facilitated by high persistence of TNF α at the tumor site. Under such conditions, we would expect high prevalence of TNF α in tumors of patients belonging to the IDC-with-relapse group,

compared to IDC-no-relapse or to DCIS patients. Indeed, here we have obtained an important indication to the roles of TNF α in disease recurrence and progression, because there was significantly higher prevalence of TNF α in tumors of the IDC-with-relapse group than in the other two groups of cancer patients. In addition, significantly elevated persistence of IL-1 β was also observed in the IDC-with-relapse patients. Although IL-1 β was not a strong stimulant of EMT, it is possible that this cytokine supports tumor recurrence or metastasis by other means, such as increased angiogenesis, as indicated by several published studies [28,31,83], or by having joint activity with other factors. Supporting this possibility is the finding that in a specific setting of Her2-neu, ER and PR expression in the tumors, IL-1 β was identified as a risk factor for disease recurrence, suggesting that it can act jointly with other pro-malignancy factors to promote disease progression in breast cancer.

The observations that were obtained in this part of the study support the possibility that high prevalence of TNF α and IL-1 β expression in the IDC-with-relapse patients contributes to re-growth of the tumors and to metastasis formation by inducing processes that push forward these events, e.g. *via* EMT and/or cooperative activity with other pro-malignancy factors that are expressed by the tumor cells, or by cells of the tumor microenvironment.

Conclusions

Our study is the first to provide detailed analysis of the expression of inflammatory chemokines, alongside with inflammatory cytokines, in different progression stages of breast cancer. We provide evidence to a coordinated expression of the inflammatory chemokines CCL2 & CCL5 and the inflammatory cytokines TNF α & IL-1 β in breast cancer, all having causative roles as tumor-promoting factors in this disease.

Our study also emphasizes the importance of TNF α and IL-1 β in promoting disease metastasis and recurrence, gaining support by their high prevalence in patients in whom disease has relapsed. Such advantages could be given to the tumor cells by the ability of TNF α to promote EMT, and by similar (although to a lower extent) or other tumor-promoting activities of IL-1 β (e.g. joint activities with other pro-malignancy elements). Mainly, the findings on TNF α contribute to the overall view of the roles played by this cytokine in breast cancer. While TNF α was found to have multiple tumor-promoting roles in this disease [25-59,84], there are malignant diseases in which it was described as an anti-malignancy factor [25,60,61]. Our present results provide further evidence to the major importance of TNF α

in supporting breast malignancy, and strengthen the need to consider this cytokine as a therapeutic target in this disease.

Based on our findings, the possibility exists that different inflammatory mediators act in complementary manners at the tumor microenvironment to support processes of tumor growth and progression, with potentially major roles for TNF α . Such a possibility emphasizes the need to perform extensive in vitro and in vivo studies, in which the combined roles of such factors will be investigated, to be then followed by determining the effects of joint inhibitory measures on disease course. Indeed, in our analyses we are now taking the first steps in this direction. It is possible that such an approach may hamper different pro-tumorigenic mechanisms and signaling pathways simultaneously, leading to more inhibitory effects on breast tumor growth and metastasis.

Additional material

Additional file 1: TNF α and IL-1 β up-regulate the release of CCL2 by human breast tumor cells. T47D (A) and MCF-7 (B) human breast tumor cells were stimulated for 24-48 hr with human TNF α or IL-1 β . CCL2 levels were determined in the cell supernatants by ELISA, at the linear range of absorbance. The results are representatives of $n \geq 3$. * $p < 0.05$, ** $p < 0.01$ in comparison to unstimulated cells.

Additional file 2: TNF α and IL-1 β up-regulate the release of CCL5 by human breast tumor cells. T47D (A) and MCF-7 (B) human breast tumor cells were stimulated for 24-48 hr with human TNF α or IL-1 β . CCL5 levels were determined in the cell supernatants by ELISA, at the linear range of absorbance. The results are representatives of $n \geq 3$. ** $p < 0.01$, *** $p < 0.001$ in comparison to unstimulated cells.

Abbreviations

DCIS: Ductal Carcinoma *In Situ*; EMT: Epithelial-to-Mesenchymal Transition; ER: Estrogen Receptor α ; IDC: Invasive Ductal carcinoma; IHC: Immunohistochemistry. IL-1 β : Interleukin 1 β ; PR: Progesterone Receptor; TNF α : Tumor Necrosis Factor α .

Acknowledgements

(1) The study was supported by The Israel Science Foundation and by Federico Foundation.
(2) In memoriam of Dr. Yaal-Hahoshen, who passed away on January 2009. Dr. Yaal-Hahoshen was involved in the initial stages of study design, before its modification to its present final format.

Author details

¹Department of Cell Research and Immunology, George S. Wise Faculty of Life Sciences, Tel Aviv University, Israel. ²Department of Surgery A, Meir Medical Center and the Sackler School of Medicine, Tel Aviv University, Israel. ³Department of Oncology, Tel Aviv Sourasky Medical Center, and the Sackler School of Medicine, Tel Aviv University, Israel. ⁴Department of Pathology, Tel Aviv Sourasky Medical Center, and the Sackler School of Medicine, Tel Aviv University, Israel. ⁵Statistical Service, Tel Aviv Sourasky Medical Center, and the Sackler School of Medicine, Tel Aviv University, Israel. ⁶Current affiliation: Department of Surgery B, Sheba Medical Center and the Sackler School of Medicine, Tel Aviv University, Israel.

Authors' contributions

GS was the major contributor to the study of normal individuals and breast cancer patients. She performed a large amount of the IHC stainings and was in charge of organization of the data, and also took charge of the migration

and invasion assays. MOS was the major contributor to the EMT part of the project, and also performed some of the IHC stainings. IH coordinated the study in the Meir Medical Center, and performed some of the IHC stainings. NYH was involved in the initial stages of study design, and coordinated the study in Sourasky Medical Center. LLT was the pathologist who analyzed the biopsy sections. ES performed the statistical analyses. TLR has contributed to the migration and invasion assays. PW performed analyses related to EMT in the tumor cells. TM participated in EMT analyses. MG participated in the design of the study in Meir Medical Center. ABB was the principal investigator responsible for the whole study, including all its parts. All authors have approved the submission of the manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 22 October 2010 Accepted: 12 April 2011

Published: 12 April 2011

References

- Lewis CE, Hughes R: Inflammation and breast cancer. Microenvironmental factors regulating macrophage function in breast tumours: hypoxia and angiopoietin-2. *Breast Cancer Res* 2007, **9**(3):209.
- Lin EY, Pollard JW: Tumor-associated macrophages press the angiogenic switch in breast cancer. *Cancer Res* 2007, **67**(11):5064-5066.
- Soria G, Ben-Baruch A: The inflammatory chemokines CCL2 and CCL5 in breast cancer. *Cancer Lett* 2008, **267**(2):271-285.
- Goldberg JE, Schwertfeger KL: Proinflammatory cytokines in breast cancer: mechanisms of action and potential targets for therapeutics. *Curr Drug Targets* 2010, **11**(9):1133-1146.
- Ben-Baruch A: The multifaceted roles of chemokines in malignancy. *Cancer Metastasis Rev* 2006, **25**(3):357-371.
- Conti I, Rollins BJ: CCL2 (monocyte chemoattractant protein-1) and cancer. *Semin Cancer Biol* 2004, **14**(3):149-154.
- Soria G, Ben-Baruch A: The CCL5/CCR5 axis in cancer. Springer Publishers;
- Pinilla S, Alt E, Abdul Khalek FJ, Jotzu C, Muehlberg F, Beckmann C, Song YH: Tissue resident stem cells produce CCL5 under the influence of cancer cells and thereby promote breast cancer cell invasion. *Cancer Lett* 2009, **284**(1):80-85.
- Zhang Y, Yao F, Yao X, Yi C, Tan C, Wei L, Sun S: Role of CCL5 in invasion, proliferation and proportion of CD44+/CD24- phenotype of MCF-7 cells and correlation of CCL5 and CCR5 expression with breast cancer progression. *Oncol Rep* 2009, **21**(4):1113-1121.
- Lu X, Kang Y: Chemokine (C-C motif) ligand 2 engages CCR2+ stromal cells of monocytic origin to promote breast cancer metastasis to lung and bone. *J Biol Chem* 2009, **284**(42):29087-29096.
- Fujimoto H, Sangai T, Ishii G, Ikehara A, Nagashima T, Miyazaki M, Ochiai A: Stromal MCP-1 in mammary tumors induces tumor-associated macrophage infiltration and contributes to tumor progression. *Int J Cancer* 2009, **125**(6):1276-1284.
- Dwyer RM, Potter-Beirne SM, Harrington KA, Lowery AJ, Hennessy E, Murphy JM, Barry FP, O'Brien T, Kerin MJ: Monocyte chemotactic protein-1 secreted by primary breast tumors stimulates migration of mesenchymal stem cells. *Clin Cancer Res* 2007, **13**(17):5020-5027.
- Molloy AP, Martin FT, Dwyer RM, Griffin TP, Murphy M, Barry FP, O'Brien T, Kerin MJ: Mesenchymal stem cell secretion of chemokines during differentiation into osteoblasts, and their potential role in mediating interactions with breast cancer cells. *Int J Cancer* 2009, **124**(2):326-332.
- Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, Richardson AL, Polyak K, Tubo R, Weinberg RA: Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature* 2007, **449**(7162):557-563.
- Yaal-Hahoshen N, Shina S, Leider-Trejo L, Barnea I, Shabtai EL, Azenshtein E, Greenberg I, Keydar I, Ben-Baruch A: The chemokine CCL5 as a potential prognostic factor predicting disease progression in stage II breast cancer patients. *Clin Cancer Res* 2006, **12**(15):4474-4480.
- Soria G, Ben-Baruch A: Concomitant expression of the chemokines RANTES and MCP-1 in human breast cancer: A basis for tumor-promoting interactions. *Cytokine* 2008, **44**:191-200.
- Valkovic T, Lucin K, Krstulja M, Dobi-Babic R, Jonjic N: Expression of monocyte chemotactic protein-1 in human invasive ductal breast cancer. *Pathol Res Pract* 1998, **194**(5):335-340.

18. Saji H, Koike M, Yamori T, Saji S, Seiki M, Matsushima K, Toi M: **Significant correlation of monocyte chemoattractant protein-1 expression with neovascularization and progression of breast carcinoma.** *Cancer* 2001, **92**(5):1085-1091.
19. Goede V, Brogelli L, Ziche M, Augustin HG: **Induction of inflammatory angiogenesis by monocyte chemoattractant protein-1.** *Int J Cancer* 1999, **82**(5):765-770.
20. Luboshits G, Shina S, Kaplan O, Engelberg S, Nass D, Lifshitz-Mercer B, Chaitchik S, Keydar I, Ben-Baruch A: **Elevated expression of the CC chemokine regulated on activation, normal T cell expressed and secreted (RANTES) in advanced breast carcinoma.** *Cancer Res* 1999, **59**(18):4681-4687.
21. Salcedo R, Ponce ML, Young HA, Wasserman K, Ward JM, Kleinman HK, Oppenheim JJ, Murphy WJ: **Human endothelial cells express CCR2 and respond to MCP-1: direct role of MCP-1 in angiogenesis and tumor progression.** *Blood* 2000, **96**(1):34-40.
22. Robinson SC, Scott KA, Wilson JL, Thompson RG, Proudfoot AE, Balkwill FR: **A chemokine receptor antagonist inhibits experimental breast tumor growth.** *Cancer Res* 2003, **63**(23):8360-8365.
23. Stormes KA, Lemken CA, Lepre JV, Marinucci MN, Kurt RA: **Inhibition of metastasis by inhibition of tumor-derived CCL5.** *Breast Cancer Res Treat* 2005, **89**(2):209-212.
24. Adler EP, Lemken CA, Katchen NS, Kurt RA: **A dual role for tumor-derived chemokine RANTES (CCL5).** *Immunol Lett* 2003, **90**(2-3):187-194.
25. Balkwill F: **TNF-alpha in promotion and progression of cancer.** *Cancer Metastasis Rev* 2006, **25**(3):409-416.
26. Apte RN, Krelin Y, Song X, Dotan S, Recih E, Elkabets M, Carmi Y, Dvorkin T, White RM, Gayvoronsky L, et al: **Effects of micro-environment- and malignant cell-derived interleukin-1 in carcinogenesis, tumour invasiveness and tumour-host interactions.** *Eur J Cancer* 2006, **42**(6):751-759.
27. Nicolini A, Carpi A, Rossi G: **Cytokines in breast cancer.** *Cytokine Growth Factor Rev* 2006, **17**(5):325-337.
28. Jin L, Yuan RQ, Fuchs A, Yao Y, Joseph A, Schwall R, Schnitt SJ, Guida A, Hastings HM, Andres J, et al: **Expression of interleukin-1beta in human breast carcinoma.** *Cancer* 1997, **80**(3):421-434.
29. Lewis AM, Varghese S, Xu H, Alexander HR: **Interleukin-1 and cancer progression: the emerging role of interleukin-1 receptor antagonist as a novel therapeutic agent in cancer treatment.** *J Transl Med* 2006, **4**:48.
30. Miller LJ, Kurtzman SH, Anderson K, Wang Y, Stankus M, Renna M, Lindquist R, Barrows G, Kreutzer DL: **Interleukin-1 family expression in human breast cancer: interleukin-1 receptor antagonist.** *Cancer Invest* 2000, **18**(4):293-302.
31. Pantschenko AG, Pushkar I, Anderson KH, Wang Y, Miller LJ, Kurtzman SH, Barrows G, Kreutzer DL: **The interleukin-1 family of cytokines and receptors in human breast cancer: implications for tumor progression.** *Int J Oncol* 2003, **23**(2):269-284.
32. Kurtzman SH, Anderson KH, Wang Y, Miller LJ, Renna M, Stankus M, Lindquist RR, Barrows G, Kreutzer DL: **Cytokines in human breast cancer: IL-1alpha and IL-1beta expression.** *Oncol Rep* 1999, **6**(1):65-70.
33. Pusztai L, Clover LM, Cooper K, Starkey PM, Lewis CE, McGee JO: **Expression of tumour necrosis factor alpha and its receptors in carcinoma of the breast.** *Br J Cancer* 1994, **70**(2):289-292.
34. Miles DW, Happerfield LC, Naylor MS, Bobrow LG, Rubens RD, Balkwill FR: **Expression of tumour necrosis factor (TNF alpha) and its receptors in benign and malignant breast tissue.** *Int J Cancer* 1994, **56**(6):777-782.
35. Basolo F, Conaldi PG, Fiore L, Calvo S, Toniolo A: **Normal breast epithelial cells produce interleukins 6 and 8 together with tumor-necrosis factor: defective IL6 expression in mammary carcinoma.** *Int J Cancer* 1993, **55**(6):926-930.
36. Leek RD, Landers R, Fox SB, Ng F, Harris AL, Lewis CE: **Association of tumour necrosis factor alpha and its receptors with thymidine phosphorylase expression in invasive breast carcinoma.** *Br J Cancer* 1998, **77**(12):2246-2251.
37. Garcia-Tunon I, Ricote M, Ruiz A, Fraile B, Paniagua R, Royuela M: **Role of tumor necrosis factor-alpha and its receptors in human benign breast lesions and tumors (in situ and infiltrative).** *Cancer Sci* 2006, **97**(10):1044-1049.
38. Sheen-Chen SM, Chen WJ, Eng HL, Chou FF: **Serum concentration of tumor necrosis factor in patients with breast cancer.** *Breast Cancer Res Treat* 1997, **43**(3):211-215.
39. Cui LF, Guo XJ, Wei J, Liu FF, Gu F, Fan Y, Lang RG, Fu L: **[Significance of interleukin-1beta expression and microvascular density in invasive micropapillary carcinoma of breast].** *Zhonghua Bing Li Xue Za Zhi* 2008, **37**(9):599-603.
40. Snoussi K, Strosberg AD, Bouaouina N, Ben Ahmed S, Chouchane L: **Genetic variation in pro-inflammatory cytokines (interleukin-1beta, interleukin-1alpha and interleukin-6) associated with the aggressive forms, survival, and relapse prediction of breast carcinoma.** *Eur Cytokine Netw* 2005, **16**(4):253-260.
41. Chavey C, Bibeau F, Gourgou-Bourgade S, Burlinchon S, Boissiere F, Laune D, Roques S, Lazennec G: **Oestrogen receptor negative breast cancers exhibit high cytokine content.** *Breast Cancer Res* 2007, **9**(1):R15.
42. Szlosarek P, Charles KA, Balkwill FR: **Tumour necrosis factor-alpha as a tumour promoter.** *Eur J Cancer* 2006, **42**(6):745-750.
43. Mestiri S, Bouaouina N, Ahmed SB, Khedhaier A, Jrad BB, Remadi S, Chouchane L: **Genetic variation in the tumor necrosis factor-alpha promoter region and in the stress protein hsp70-2: susceptibility and prognostic implications in breast carcinoma.** *Cancer* 2001, **91**(4):672-678.
44. Mestiri S, Bouaouina N, Ben Ahmed S, Chouchane L: **A functional polymorphism of the tumor necrosis factor receptor-II gene associated with the survival and relapse prediction of breast carcinoma.** *Cytokine* 2005, **30**(4):182-187.
45. Bozcuk H, Uslu G, Samur M, Yildiz M, Ozben T, Ozdogan M, Artac M, Altunbas H, Akan I, Savas B: **Tumour necrosis factor-alpha, interleukin-6, and fasting serum insulin correlate with clinical outcome in metastatic breast cancer patients treated with chemotherapy.** *Cytokine* 2004, **27**(2-3):58-65.
46. Rubio MF, Werbajh S, Cafferata EG, Quaglino A, Colo GP, Nojek IM, Kordon EC, Nahmod VE, Costas MA: **TNF-alpha enhances estrogen-induced cell proliferation of estrogen-dependent breast tumor cells through a complex containing nuclear factor-kappa B.** *Oncogene* 2006, **25**(9):1367-1377.
47. Seeger H, Wallwiener D, Mueck AO: **Different effects of estradiol and various antiestrogens on TNF-alpha-induced changes of biochemical markers for growth and invasion of human breast cancer cells.** *Life Sci* 2006, **78**(13):1464-1468.
48. Sirotkovic-Skerlev M, Cacev T, Krizanac S, Kulic A, Pavelic K, Kapitanovic S: **TNF alpha promoter polymorphisms analysis in benign and malignant breast lesions.** *Exp Mol Pathol* 2007, **83**(1):54-58.
49. Rivas MA, Carnevale RP, Proietti CJ, Rosembli C, Beguelin W, Salatino M, Charreau EH, Frahm I, Sapia S, Brouckaert P, et al: **TNF alpha acting on TNFR1 promotes breast cancer growth via p42/P44 MAPK, JNK, Akt and NF-kappa B-dependent pathways.** *Exp Cell Res* 2008, **314**(3):509-529.
50. Hagemann T, Robinson SC, Schulz M, Trumper L, Balkwill FR, Binder C: **Enhanced invasiveness of breast cancer cell lines upon co-cultivation with macrophages is due to TNF-alpha dependent up-regulation of matrix metalloproteases.** *Carcinogenesis* 2004, **25**(8):1543-1549.
51. Carpenter PM, Gatanaga T, Nguyen HP, Hiserodt JC: **Lymphocyte and monocyte-induced motility of MCF-7 cells by tumor necrosis factor-alpha.** *Int J Cancer* 1997, **71**(1):64-70.
52. Varela LM, Stangle-Castor NC, Shoemaker SF, Shea-Eaton WK, Ip MM: **TNFalpha induces NFkappaB/p50 in association with the growth and morphogenesis of normal and transformed rat mammary epithelial cells.** *J Cell Physiol* 2001, **188**(1):120-131.
53. Warren MA, Shoemaker SF, Shealy DJ, Bshar W, Ip MM: **Tumor necrosis factor deficiency inhibits mammary tumorigenesis and a tumor necrosis factor neutralizing antibody decreases mammary tumor growth in neu/erbB2 transgenic mice.** *Mol Cancer Ther* 2009, **8**(9):2655-2663.
54. Pantschenko AG, Pushkar I, Miller LJ, Wang YP, Anderson K, Peled Z, Kurtzman SH, Kreutzer DL: **In vitro demonstration of breast cancer tumor cell sub-populations based on interleukin-1/tumor necrosis factor induction of interleukin-8 expression.** *Oncol Rep* 2003, **10**(4):1011-1017.
55. De Larco JE, Wuertz BR, Rosner KA, Erickson SA, Gamache DE, Manivel JC, Furcht LT: **A potential role for interleukin-8 in the metastatic phenotype of breast carcinoma cells.** *Am J Pathol* 2001, **158**(2):639-646.
56. Nozaki S, Sledge GW Jr, Nakshatri H: **Cancer cell-derived interleukin 1alpha contributes to autocrine and paracrine induction of pro-metastatic genes in breast cancer.** *Biochem Biophys Res Commun* 2000, **275**(1):60-62.
57. Palmieri C, Roberts-Clark D, Assadi-Sabet A, Coope RC, O'Hare M, Suinters A, Hanby A, Slade MJ, Gomm JJ, Lam EW, et al: **Fibroblast growth factor 7,**

- secreted by breast fibroblasts, is an interleukin-1beta-induced paracrine growth factor for human breast cells. *J Endocrinol* 2003, **177**(1):65-81.
58. Verhasselt B, Van Damme J, van Larebeke N, Put W, Bracke M, De Potter C, Mareel M: Interleukin-1 is a motility factor for human breast carcinoma cells in vitro: additive effect with interleukin-6. *Eur J Cell Biol* 1992, **59**(2):449-457.
59. Reed JR, Leon RP, Hall MK, Schwertfeger KL: Interleukin-1beta and fibroblast growth factor receptor 1 cooperate to induce cyclooxygenase-2 during early mammary tumorigenesis. *Breast Cancer Res* 2009, **11**(2):R21.
60. Balkwill F: Tumour necrosis factor and cancer. *Nat Rev Cancer* 2009, **9**(5):361-371.
61. Bertazza L, Mocellin S: Tumor necrosis factor (TNF) biology and cell death. *Front Biosci* 2008, **13**:2736-2743.
62. Page DL: Breast lesions, pathology and cancer risk. *Breast J* 2004, **10**(Suppl 1):S3-4.
63. Cuzick J: Treatment of DCIS—results from clinical trials. *Surg Oncol* 2003, **12**(4):213-219.
64. Leonard GD, Swain SM: Ductal carcinoma in situ, complexities and challenges. *J Natl Cancer Inst* 2004, **96**(12):906-920.
65. Neumark E, Cohn MA, Lukanidin E, Witz IP, Ben-Baruch A: Possible co-regulation of genes associated with enhanced progression of mammary adenocarcinomas. *Immunol Lett* 2002, **82**(1-2):111-121.
66. Neumark E, Sagi-Assif O, Shalmon B, Ben-Baruch A, Witz IP: Progression of mouse mammary tumors: MCP-1-TNFalpha cross-regulatory pathway and clonal expression of promalignancy and antimalignancy factors. *Int J Cancer* 2003, **106**(6):879-886.
67. Azenshtein E, Luboshits G, Shina S, Neumark E, Shahbazian D, Weil M, Wigler N, Keydar I, Ben-Baruch A: The CC chemokine RANTES in breast carcinoma progression: regulation of expression and potential mechanisms of promalignant activity. *Cancer Res* 2002, **62**(4):1093-1102.
68. Ali S, Kaur J, Patel KD: Intercellular cell adhesion molecule-1, vascular cell adhesion molecule-1, and regulated on activation normal T cell expressed and secreted are expressed by human breast carcinoma cells and support eosinophil adhesion and activation. *Am J Pathol* 2000, **157**(1):313-321.
69. Seeger H, Wallwiener D, Mueck AO: Effects of estradiol and progestogens on tumor-necrosis factor-alpha-induced changes of biochemical markers for breast cancer growth and metastasis. *Gynecol Endocrinol* 2008, **24**(10):576-579.
70. Kelly RW, Carr GG, Riley SC: The inhibition of synthesis of a beta-chemokine, monocyte chemoattractant protein-1 (MCP-1) by progesterone. *Biochem Biophys Res Commun* 1997, **239**(2):557-561.
71. Polyak K, Weinberg RA: Transitions between epithelial and mesenchymal states: acquisition of malignant and stem cell traits. *Nat Rev Cancer* 2009, **9**(4):265-273.
72. Hollier BG, Evans K, Mani SA: The epithelial-to-mesenchymal transition and cancer stem cells: a coalition against cancer therapies. *J Mammary Gland Biol Neoplasia* 2009, **14**(1):29-43.
73. Vincent-Salomon A, Thiery JP: Host microenvironment in breast cancer development: epithelial-mesenchymal transition in breast cancer development. *Breast Cancer Res* 2003, **5**(2):101-106.
74. Jechlinger M, Grunert S, Tamir IH, Janda E, Ludemann S, Waerner T, Seither P, Weith A, Beug H, Kraut N: Expression profiling of epithelial plasticity in tumor progression. *Oncogene* 2003, **22**(46):7155-7169.
75. Prindull G: Hypothesis: cell plasticity, linking embryonal stem cells to adult stem cell reservoirs and metastatic cancer cells? *Exp Hematol* 2005, **33**(7):738-746.
76. Blick T, Widodo E, Hugo H, Waltham M, Lenburg ME, Neve RM, Thompson EW: Epithelial mesenchymal transition traits in human breast cancer cell lines. *Clin Exp Metastasis* 2008, **25**(6):629-642.
77. Hugo H, Ackland ML, Blick T, Lawrence MG, Clements JA, Williams ED, Thompson EW: Epithelial-mesenchymal and mesenchymal-epithelial transitions in carcinoma progression. *J Cell Physiol* 2007, **213**(2):374-383.
78. Freund A, Chauveau C, Brouillet JP, Lucas A, Lacroix M, Licznar A, Vignon F, Lazennec G: IL-8 expression and its possible relationship with estrogen-receptor-negative status of breast cancer cells. *Oncogene* 2003, **22**(2):256-265.
79. Bagatell R, Khan O, Paine-Murrieta G, Taylor CW, Akinaga S, Whitesell L: Destabilization of steroid receptors by heat shock protein 90-binding drugs: a ligand-independent approach to hormonal therapy of breast cancer. *Clin Cancer Res* 2001, **7**(7):2076-2084.
80. Walsh MD, Luckie SM, Cummings MC, Antalis TM, McGuckin MA: Heterogeneity of MUC1 expression by human breast carcinoma cell lines in vivo and in vitro. *Breast Cancer Res Treat* 1999, **58**(3):255-266.
81. Bussolati B, Biancone L, Cassoni P, Russo S, Rola-Pleszczynski M, Montrucchio G, Camussi G: PAF produced by human breast cancer cells promotes migration and proliferation of tumor cells and neo-angiogenesis. *Am J Pathol* 2000, **157**(5):1713-1725.
82. Line BR, Feustel PJ, Festin SM, Andersen TT, Dansereau RN, Lukasiewicz RL, Zhu S, Bennett JA: Scintigraphic detection of breast cancer xenografts with Tc-99m natural and recombinant human alpha-fetoprotein. *Cancer Biother Radiopharm* 1999, **14**(6):485-494.
83. Fan F, Stoeltzing O, Liu W, McCarty MF, Jung YD, Reinmuth N, Ellis LM: Interleukin-1beta regulates angiopoietin-1 expression in human endothelial cells. *Cancer Res* 2004, **64**(9):3186-3190.
84. Apte RN, Dotan S, Elkabets M, White MR, Reich E, Carmi Y, Song X, Dvozkin T, Krelin Y, Voronov E: The involvement of IL-1 in tumorigenesis, tumor invasiveness, metastasis and tumor-host interactions. *Cancer Metastasis Rev* 2006, **25**(3):387-408.

Pre-publication history

The pre-publication history for this paper can be accessed here:
<http://www.biomedcentral.com/1471-2407/11/130/prepub>

doi:10.1186/1471-2407-11-130

Cite this article as: Soria et al.: Inflammatory mediators in breast cancer: Coordinated expression of TNF α & IL-1 β with CCL2 & CCL5 and effects on epithelial-to-mesenchymal transition. *BMC Cancer* 2011 **11**:130.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

