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Inhibition of Granulopoiesis in Vivo and in Vitro by β -Lactam Antibiotics

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β -Lactam antibiotics can induce severe neutropenia by a hitherto unknown mechanism. Fifty cases of β -lactam antibiotic-induced neutropenia ($<1,000$ neutrophils/ mm^3) from 17 hospitals were analyzed and compared with 140 literature cases. The incidence of neutropenia was 5%–15% in patients treated for ≥ 10 days with large doses of any β -lactam antibiotic but $<0.1\%$ with shorter duration of therapy. In $>95\%$ of cases recovery occurred between one to seven days after withdrawal of β -lactam antibiotics. Bone marrow aspirates were characterized by a lack of well-differentiated myeloid elements in the presence of numerous immature granulocyte precursors. Nine penicillins and eight cephalosporins inhibited in vitro granulopoiesis in a dose-dependent manner. There was a good correlation between the inhibitory capacity of β -lactam antibiotics in vitro and the doses inducing neutropenia in vivo. These observations may be relevant for therapy in the granulocytopenic patient.

Penicillins and cephalosporins (β -lactam antibiotics) are remarkably nontoxic within a broad dose range. Nevertheless, they induce a variety of adverse reactions, most of which are believed to be of an allergic nature [1]. Severe neutropenia is a well-known complication of high-dose therapy with β -lactam antibiotics [2]. There are still many unanswered clinical and pathogenetic questions. It is not known whether β -lactam antibiotic-induced neutropenia is a result of direct toxicity to the bone marrow or of immune-mediated bone marrow injury. In an attempt to increase our understanding of β -lactam antibiotic-induced neutropenia, we analyzed 50 of our patients with this problem, found an additional 140 literature cases, and assessed the influence of 17 selected β -lactam antibiotics on in vitro granulopoiesis.

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Materials and Methods

Selection of cases. Neutropenia was defined as a neutrophil count of $\leq 1,000/\text{mm}^3$. Criteria for assessing the causal relation between neutropenia and β -lactam antibiotics were adapted from those of Karch and Lasagna [3]. Neutropenia was thus induced by β -lactam antibiotics only if it followed a reasonable time sequence from administration of the drug, disappeared on stopping administration of the drug, and was not explainable by the patient's clinical condition.

Cases were excluded in which other drugs had to be considered as a possible cause or in which the cause-and-effect relation remained otherwise questionable.

Cases of β -lactam antibiotic-induced neutropenia came to our attention in one of the three following ways: (1) they were encountered during a multicenter study concerning the side effects of penicillin G [3] and other β -lactam antibiotics (authors' unpublished observations); (2) they occurred from September 1980 to December 1983 in the Medical Clinic of the University Hospital, Zurich; or (3) specimens from such cases were sent to us for evaluation of suspected β -lactam antibiotic-induced reactions.

The literature cases were obtained through a computer search (MEDLINE; National Library of Medicine, Bethesda, Md) from 1945 to 1983.

Analysis of clinical data. The total dose of β -lactam antibiotic given within the course of therapy before neutropenia developed was determined for each patient, and the mean daily dose (MDD) was

then calculated. In children the doses per kilogram of body weight were translated to the same relative doses given to a patient weighing 65 kg. Therapy breaks of <48 hr were ignored. If more than one β -lactam antibiotic was involved, the drug given while neutropenia developed was considered as the main cause. If two β -lactam antibiotics were given simultaneously, the one given in the higher total dose was considered as the main cause.

Concomitant symptoms of neutropenia were defined as follows. Eosinophilia was $\geq 10\%$ of the white blood cell count or an absolute count of $\geq 750/\text{mm}^3$. Thrombocytopenia was $\leq 100,000$ platelets/ mm^3 . Reticulocytopenia was $\leq 0.25\%$ of the red blood cell count. Fever was a temperature ≥ 38.0 C after at least three afebrile days (≤ 37.2 C) with no other cause of fever evident.

Bone marrow cultures. Cancellous bone samples from hematologically normal donors undergoing orthopedic surgery were collected in Hanks' solution.

A modification of the method of Pike and Robinson [4] for culture of myeloid progenitor cells was used. Dextran-sedimented bone marrow cells were plated at a concentration of 10^5 mononuclear cells/ml of supplemented minimal essential medium containing 0.3% agar and 20% pretested fetal calf serum. Human placenta-conditioned medium [5] served as the source of colony-stimulating factor. Cultures were set up in triplicate and incubated at 37 C in a humidified atmosphere plus 5% CO_2 . Af-

ter eight days aggregates of ≥ 40 cells were scored as colonies and those of three to 39 cells as clusters. Because cluster formation always paralleled colony formation, only the latter was considered for final evaluation.

β -Lactam antibiotics were dissolved in 0.9% NaCl at 100 mg/ml and further diluted with supplemented medium. In some experiments β -lactam antibiotics were added after the culture process was started by overlaying the drug in 0.1 ml of complete medium on top of the agar culture to the final concentration required.

Results

In vivo data. Table 1 summarizes data in 50 cases of β -lactam antibiotic-induced neutropenia from a total of 17 hospitals.

Ninety-six percent of these cases were detected only after therapy with β -lactam antibiotics of ≥ 10 days. In cases in which neutrophil counts were done daily, a decrease in count was seen over several days until total agranulocytosis was established or therapy with β -lactam antibiotics was discontinued.

In contrast to the duration of therapy, MDDs, and thus total doses given before neutropenia developed, varied markedly with different β -lactam antibiotics. In the cases of penicillin G, floxacillin, and ceftriaxone, with six or more cases each, these differences were significant (table 1).

Table 1. Data from 50 cases of β -lactam antibiotic-induced neutropenia from 17 hospitals.

Substance	No. of cases	Mean (range) duration of therapy (days)	Mean \pm SD dose (g)		No. of cases with		
			Total	Daily	Rash	Eosino-	
						philia	Fever
Penicillin G	26	23 (13-34)	268 \pm 97*	11.6 \pm 3.5 [†]	11	10	11
Floxacin	7	27 (21-40)	197 \pm 67* [‡]	7.2 \pm 1.1 ^{†§}	4	2	3
Ceftriaxone	6	21 (8-25)	51 \pm 29 [‡]	2.3 \pm 1.0 [§]	0	3	2
Amoxicillin	2	13 (6-20)	216 \pm 275	12.0 \pm 12.0	0	0	0
Piperacillin	2	22 (17-27)	400 \pm 232	17.4 \pm 4.9	0	0	0
Cefotaxime	2	29 (27-31)	208 \pm 101	7.0 \pm 2.8	0	0	1
Apalcillin	1	16	162	9.0	0	1	0
Ticarcillin	1	15	516	34.4	0	1	0
Mezlocillin	1	36	594	16.5	1	1	1
Cephalothin	1	18	139	7.7	0	0	0
Cephacetrile	1	36	432	12.0	1	0	1
Total	50	23.2 (6-40)	243 \pm 139	10.3 \pm 5.9	17	18	19

* $P < .05$ by t test.

[†] $P < .01$ by t test.

[‡] $P < .01$ by t test.

[§] $P < .01$ by t test.

On discontinuation of therapy with β -lactam antibiotics, complete recovery took place within one to seven days.

Simultaneous eosinophilia, rash, and/or fever were observed in 35 (70%) of the cases (table 1). In 19 (38%) cases one symptom was seen, in 14 (28%) cases two symptoms were present, and in 2 (4%) cases all three occurred together. There did not appear to be any predictable combination of these symptoms, which disappeared within one to four days after cessation of therapy with β -lactam antibiotics.

Thrombocytopenia was noted in nine of 36 patients for whom platelet counts were done and reticulocytopenia in two of 19 cases.

Bone marrow findings. In 24 cases a bone marrow aspirate was available. Numerical values of differential counts are not presented because bone marrow aspiration was done at different intervals with regard to the nadir of neutropenia.

Lack of well-differentiated myeloid elements in the presence of still numerous granulocyte precursors was a typical finding. The mitotic pool of granulopoiesis was found to be normocellular or even hypercellular; however, occasionally it was hypocellular. Severe hypocellularity has only been found in cases of total agranulocytosis. Such a "maturation arrest" in aspirates taken immediately at the termination of therapy was usually at the promyelocyte stage, whereas ≥ 24 hr later it was at the myelocyte or metamyelocyte stage.

In four cases bone marrow aspirates were performed at the beginning of the decrease in number of peripheral granulocytes. In these cases myeloblasts

and promyelocytes were still numerous; myelocytes were greatly reduced in number or lacking, whereas the numbers of metamyelocytes, band forms, and segmented granulocytes were still normal.

In vitro data. Addition of various β -lactam antibiotics to normal bone marrow cultures resulted in a dose-dependent inhibition of myeloid colony formation, as illustrated for penicillin G, cloxacillin, and ceftizoxime in figure 1. There were impressive differences in absolute potency of the β -lactam antibiotics studied. However, the shape of the dose-effect curves was always similar. Concentration of β -lactam antibiotics leading to 50% inhibition of colony formation (IC_{50}) were interpolated from the dose-effect curves and served as a simple measure of in vitro toxicity (table 2). Cephalosporins were from three to 25 times more effective than semisynthetic penicillins. The sensitivity of cells from different bone marrow donors to a particular substance varied greatly. In some instances, especially in the low concentration range, deviations from median values exceeding 50% were seen. However, the relation of inhibition of a given bone marrow sample by different β -lactam antibiotics remained constant.

Overnight exposure (18 hr) of bone marrow cells to β -lactam antibiotics in concentrations otherwise resulting in 90%–100% inhibition, followed by washing for removal of drug, did not affect colony formation. The trypan blue-exclusion test showed full viability of cells during preincubation as compared with controls (viability, >95%).

However, when β -lactam antibiotics were added after the culture process had begun, the inhibitory effect was progressively diminished. In five culture

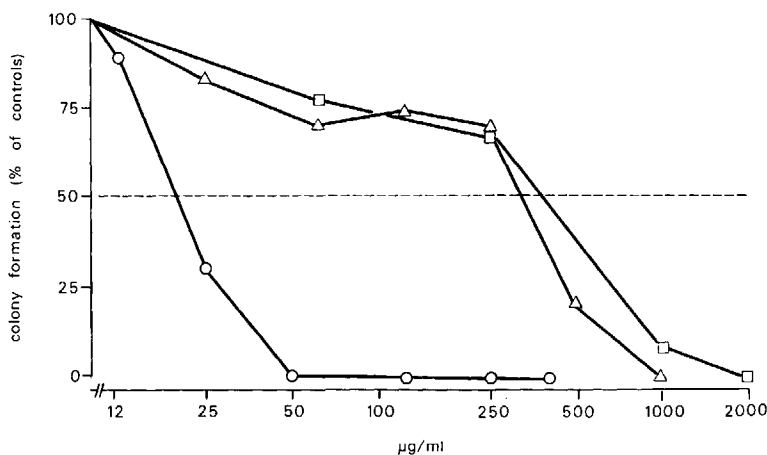


Figure 1. Inhibition of colony formation by ceftizoxime (O), cloxacillin (Δ), and penicillin G (□). Data are median values from differing numbers of cultures (table 2).

Table 2. IC_{50} values of penicillin G, eight semisynthetic penicillins, and eight cephalosporins in vitro.

Substance	Medium (range) IC_{50} (μ g/ml)	No. of cultures
Penicillin		
Mezlocillin	550 (280->1,000)	4
Ticarcillin	520 (300->1,000)	9
Piperacillin	470 (153->1,000)	5
Methicillin	470 (155-670)	6
Penicillin G	380 (45-560)	15
Oxacillin	370 (62.5-560)	4
Cloxacillin	340 (251-380)	4
Azlocillin	320 (12-1,000)	6
Floxacillin	105 (<62.5-230)	6
Cephalosporin		
Moxalactam	35 (12-62)	4
Cefoxitin	32 (16-36)	3
Ceftriaxone	30 (2-43)	5
Ceftizoxime	20 (10-33)	4
Cephalothin	16 (16-17)	3
Cefazolin	15 (7-18)	3
Ceftazidime	12 (1-16)	3
Cefotaxime	7 (1-8)	4

experiments with ceftazidime we found inhibition to be reduced by $\sim 50\%$ when the drug was added after 24 hr and by $\sim 75\%$ after 48hr. Addition after ≥ 72 hr no longer induced significant inhibition.

Discussion

In vivo data. In the literature we found an additional 140 cases fulfilling the above criteria for β -lactam antibiotic-induced neutropenia (table 3). The literature data largely confirmed the observations with our 50 subjects and allowed estimation of the incidence of β -lactam antibiotic-induced neutropenia and the risk of a relapse when the patient is rechallenged.

Relation of neutropenia to duration and dosage of therapy with β -lactam antibiotics. In 91% of the literature cases neutropenia did not occur before day 10 after therapy was started. Indeed, duration of therapy with β -lactam antibiotics of at least 10 days appears to be a threshold for inducing neutropenia. In

Table 3. Data from 140 literature cases of β -lactam antibiotic-induced neutropenia.

Substance	No. of cases	Mean (range) duration of therapy (days)	Mean \pm SD dose (g)		Reference
			Total	Daily	
Oxacillin	22	24 (17-75)	249 \pm 114	11.1 \pm 4.8	6-16
Penicillin G	20	24 (10-36)	335 \pm 301	14.1 \pm 12	16-31
Cephalothin	18	20 (2-34)	141 \pm 83	7.4 \pm 2.9	11, 32-38
Nafcillin	18	18 (2-38)	169 \pm 68	10.5 \pm 2.8	39-48
Methicillin	9	17 (4-21)	171 \pm 102	9.5 \pm 3.4	49-54
Cloxacillin	8 (+ 1)*	25 (17-40)	216 \pm 82	9.3 \pm 3.5	24, 55-57
Ampicillin	8	20 (9-32)	198 \pm 125	10.9 \pm 7.7	16, 23, 58-62
Carbenicillin	6	20 (15-24)	536 \pm 213	25.8 \pm 9.3	23, 24, 63
Cefotaxime	5	22 (9-37)	171 \pm 162	6.5 \pm 3.7	16, 64, 65
Piperacillin	4	24 (22-26)	373 \pm 142	15.4 \pm 5.9	46, 66
Ticarcillin	4	15 (9-29)	246 \pm 111	18.5 \pm 5.5	67-69
Ceftazidime	3	14 (7-25)	66 \pm 73	4.1 \pm 2.3	16, 70
Amoxicillin	2	21 (20-22)	74 \pm 2	3.5 \pm 0.3	16
Cephaloridine	2	16 (5-27)	115 \pm 70	9.6 \pm 4.9	71, 72
Mezlocillin	2	17 (12-22)	134 \pm 65	9.5 \pm 7.8	73, 74
Ceftriaxone	2	6 (5-7)	30 \pm 7	4.9	75
Azlocillin	1	19	223	11.7	76
Phenethicillin	1	23	138	5.0	29
Cefamandole	1	24	254	10.6	11
Cefuroxime	1	28	168	6.0	30
Cefoperazone	1	18	108	6.0	77
Cephapirin	1	6	24	4.0	78
Total	140	20.6 (2-75)	224 \pm 178	10.95 \pm 7.5	

* The additional case was seen after 263 days of peroral therapy (1.5 g per day).

Table 4. Incidence of neutropenia in four different series of patients treated with methicillin.

Reference	Mean duration of therapy (days)	Mean total dose (g)	Incidence (%)
Nahata et al. [44]	6.5	45	0
Kitzing et al. [51]	10	130	0
	21	273	12
Kancir et al. [85]	27	>325	12.2

patients treated with large doses of β -lactam antibiotics for ≥ 10 days, neutropenia developed with an incidence of from 5% to >15% [3, 44, 48, 51, 54, 64, 66, 75, 79–84]. With shorter therapy courses, however, the incidence can be estimated to be as low as 0.1%.

This point can be illustrated with three articles that studied a total of 144 patients receiving methicillin (table 4). Yow et al. [54], in a 10 year assessment of methicillin-associated side effects, found leukopenia to occur in 16 of 102 children who received 200 mg of methicillin/kg per day for ≥ 10 days. In four instances the absolute neutrophil count dropped to $< 500/\text{mm}^3$. In contrast, nearly 3,000 patients who received the drug for < 10 days did not develop leukopenia.

Besides intrinsic myelosuppressive activity, pharmacokinetic properties might be important for the overall myelotoxicity of a given β -lactam antibiotic. Because ceftriaxone has a very long serum half-life, neutropenia already occurred in a dose range lower than with any other β -lactam antibiotic. However, this observation need not imply an increased risk because this substance reaches sufficient antibacterial activity with lower daily doses as well. Pharmacokinetic factors might also account for the fact that in the 15 literature cases in which treatment was oral [10, 16, 29, 56, 57, 61] or im [29, 33, 52, 53, 72, 78] total doses of β -lactam antibiotics given were lower than when treatment was iv, although duration of such treatment was the same.

Complete recovery within one to seven days after discontinuation of therapy with β -lactam antibiotics was observed in 94% of the well-documented literature cases. Duration of recovery of four or more days was only seen in cases of total agranulocytosis. Permanent damage of granulopoiesis was never seen. In one case a decreased neutrophil count remained constant [41], and in three cases the count even returned to normal levels during continuous therapy [32, 39, 42].

Exposure to the same or a different β -lactam antibiotic induced a relapse in only 12 (31%) of the 39 patients rechallenged (including our cases) [6, 7, 11, 13, 26, 27, 29, 30, 32, 36, 40–42, 46, 49, 52, 59, 63, 67, 69, 85]. The induction of a relapse depended clearly on the duration and dosage of therapy with β -lactam antibiotic, as did the first induction of neutropenia. This dependence was true irrespective of the time between complete recovery from neutropenia and rechallenge. There have been relapses reported months and even years after the first episode of neutropenia [27, 29].

Relation of neutropenia to other symptoms. Symptoms accompanying neutropenia in the literature cases were similar to those seen in our cases (table 5).

Eosinophilia, rash, and fever combined were reported in 10 cases, two of the symptoms appeared in 17 cases, and an isolated symptom was seen in 37 cases. The three symptoms appear to represent a syndrome that is pathogenetically related to neutropenia.

Thrombocytopenia and reticulocytopenia occurring along with neutropenia suggest that β -lactam antibiotics can inhibit all three hematopoietic cell lines. However, when bone marrow aspirates were analyzed no clear substrate for this inhibition could be identified.

In vitro data. All of the β -lactam antibiotics studied inhibited in vitro granulopoiesis dose dependently. However, progenitor cells were not directly killed by β -lactam antibiotics. If the bone marrow cells were incubated overnight with β -lactam antibiotics followed by washing and plating, no inhibition of colony formation was observed, and the developmental potential of the cells appeared to be maintained. However, β -lactam antibiotics were only fully effective during the first 24 hr of the culture pro-

Table 5. Concomitant symptoms in 140 literature cases of β -lactam antibiotic-induced neutropenia.

Symptom	No. positive/no. of patients studied (%)
Eosinophilia*	25/79 (32)
Rash	26/64 (41)
Fever*	52/76 (68)
Thrombocytopenia	9/69 (13)
Reticulocytopenia	12/27 (44)

* If exact values were lacking, these symptoms were considered present when they were at least stated qualitatively.

cess. An inhibitory effect of β -lactam antibiotics on *in vitro* granulopoiesis is thus seen only when these drugs are present during the early proliferative phase. Furthermore, we have noted that addition of acute-phase serum and/or lymphocytes from two patients with penicillin G-induced neutropenia to autologous bone marrow cultures did not alter the effects of β -lactam antibiotics [86]. These *in vitro* findings taken together are consistent only with a nonimmunologic pathogenesis of β -lactam antibiotic-induced neutropenia, assuming that similar mechanisms are operative *in vivo*.

Several articles have reported drug-dependent antibodies to neutrophils, which could explain β -lactam antibiotic-induced neutropenia by a mechanism similar to that seen in penicillin-induced immunohemolytic anemia [16, 25, 87, 88]. With a solid-phase RIA we found circulating IgG antibodies to penicillins in a series of patients after treatment with large doses of penicillin G [2, 89] as well as ampicillin, floxacillin, or piperacillin [90], whether the subjects had neutropenia or not. Circulating antibodies to penicilloylated cells may only reflect an increase in level of circulating antibodies to drug, which appears to be common after very high-dose therapy with β -lactam antibiotics.

Inhibitory capacity (IC_{50}) of different β -lactam antibiotics in bone marrow cultures correlates well with the MDDs inducing neutropenia in patients (figure 2). Bone marrow cultures seem to allow for an approximate estimation of the myelotoxicity that would be expected *in vivo*. However, there are important discrepancies between semisolid agar cultures and the situation *in vivo*, mainly concerning the kinetics and protein binding of β -lactam antibiotics. On the basis of preliminary experience, we believe that with a liquid culture system allowing for multiple addition and removal of the drug during the culture process, an improved test model could be built up.

The inhibitory effect of β -lactam antibiotics on cell proliferation appears not to be restricted to granulopoiesis. Some cephalosporins were found to dose dependently inhibit lymphocyte proliferation after stimulation with mitogens [91, 92]. Preincubation with moxalactam followed by washing did not affect viability and subsequent proliferation of cells [92]. Similar effects were shown with penicillin G in cultures of isolated rat hepatocytes [93, 94]. It remains unclear how this inhibition of cell proliferation is achieved. Because the effect can be abolished by thorough washing after preincubation, stable

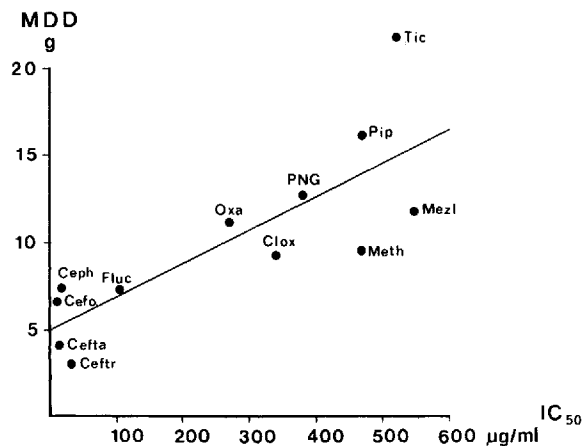


Figure 2. Correlation ($r = .804$; $P < .01$) of MDDs inducing neutropenia *in vivo* with IC_{50} values *in vitro*: PNG = penicillin G; Cefo = cefotaxime; Cefta = ceftazidime; Ceftr = ceftriaxone; Ceph = cephalothin; Clox = cloxacillin; Fluc = flucloxacillin (Floxacillin); Meth = methicillin; Mezl = mezlocillin; Oxa = oxacillin; Pip = piperacillin; and Tic = ticarcillin.

covalent binding of carbonyl groups to amino groups of membrane proteins or simple uptake and binding to intracellular structures are unlikely to be decisive pathogenic steps.

Although the *in vitro* observations as a whole cannot yet be interpreted comprehensively, they match well the typical findings in the bone marrow aspirate of patients with β -lactam antibiotic-induced neutropenia. As described above, immature myeloid precursor cells are usually numerous, whereas more mature forms are lacking. Furthermore, bone marrow aspirates taken early during development of neutropenia suggest that postmitotic maturation is not affected. β -lactam antibiotics appear not to affect the granulocytic stem cell itself but might exert their inhibitory effects on other precursor cells within the mitotic pool. Consequently, "proliferation arrest" would describe what seems to happen better than the generally used phrase "maturation arrest." This particular proliferation disturbance may explain the rapid recovery from neutropenia generally seen after withdrawal of therapy with β -lactam antibiotics.

Clinical consequences. In only eight (4.2%) of the 190 cases analyzed did a secondary infection ascribable to neutropenia develop, and it was never fatal. There might be some bias in that cases without documented recovery on discontinuation of therapy with β -lactam antibiotics were not considered. There are literature cases otherwise fulfilling our

criteria in which death occurred from various causes before the peripheral granulocyte count rose again [73]. Nevertheless, the risk for important sequelae of β -lactam antibiotic-induced neutropenia appears to be low.

The more important question is what consequences do the myelosuppressive effects of high-dose therapy with β -lactam antibiotics have in the treatment of the granulocytopenic patient, especially after cytostatic treatment? In such cases long-term therapy with cumulative doses, often of >200 g, of one or more β -lactam antibiotics are given. In hematologically normal individuals this treatment would lead to neutropenia in 5% to 15% of those treated. Currently several studies are being conducted to assess whether double β -lactam antibiotic regimens can reduce toxicity of β -lactam-aminoglycoside combinations without giving up double antibiotic coverage. Indeed, in a recent trial a double β -lactam antibiotic regimen resulted in lower nephrotoxicity but also in a significantly longer duration of granulocytopenia [95].

In an ongoing retrospective study we have analyzed 227 granulocytopenic episodes (granulocyte count, <500/mm³) in patients with acute myelogenous leukemia. We found that beginning therapy with β -lactam antibiotics simultaneously with or very early after cytostatic treatment is associated significantly ($P < .001$) with a longer duration of granulocytopenia. Consequently, in patients with granulocytopenia for >20 days, the mean total dose of β -lactam antibiotics and the mean dosage of β -lactam antibiotic per granulocytopenic day were disproportionately higher as compared with the remaining patients treated with β -lactam antibiotics (authors' unpublished observations).

Duration and severity of granulocytopenia remain the most important prognostic factors for the patient with neutropenia after cytostatic therapy for malignant disease. It is very important to clarify whether β -lactam antibiotics can delay regeneration of myelopoiesis after cytostatic therapy and how such an effect could be recognized.

References

1. Erffmeyer JE. Adverse reactions to penicillin. *Ann Allergy* 1981;47:288-300
2. Nefel KA, Wälti M, Spengler H, von Felten A, Weitzman SA, Bürgi H, de Weck AL. Neutropenia after penicillins: toxic or immune-mediated? *Klin Wochenschr* 1981;59:877-88
3. Karch FE, Lasagna L. Adverse drug reactions — a critical review. *JAMA* 1975;234:1236-41
4. Pike BL, Robinson WA. Human bone marrow colony growth in agar-gel. *J Cell Physiol* 1970;76:77-84
5. Schlunk T, Schleyer M. The influence of culture conditions on the production of colony-stimulating activity by human placenta. *Exp Hematol* 1980;8:179-84
6. Ahern MJ, Hicks JE, Andriole VT. Neutropenia during high dose intravenous oxacillin therapy. *Yale J Biol Med* 1976;49:351-60
7. Brook I. Leukopenia and granulocytopenia after oxacillin therapy. *South Med J* 1977;70:565-6
8. Chu J-Y, O'Connor DM, Schmidt RR. The mechanism of oxacillin-induced neutropenia [letter]. *J Pediatr* 1977;90:668-9
9. Fallon JA, Tall AR, Janis MG, Brauer MJ. Oxacillin-induced granulocytopenia. *Acta Haematol (Basel)* 1978;59:163-70
10. Freedman MA. Oxacillin — apparent hematologic and hepatic toxicity. *Rocky Mountain Medical Journal* 1965;62(1):34-6
11. Homayouni H, Gross PA, Setia U, Lynch TJ. Leukopenia due to penicillin and cephalosporin homologues. *Arch Intern Med* 1979;139:827-8
12. Kahn JB. Oxacillin-induced agranulocytosis [letter]. *JAMA* 1978;240:2632
13. Leventhal JM, Silken AB. Oxacillin-induced neutropenia in children. *J Pediatr* 1976;89:769-71
14. Ortvals DW, Marr JJ. Profound neutropenia caused by oxacillin. *South Med J* 1977;70:1245-7
15. Passoff TL, Sherry HS. Oxacillin induced neutropenia. A case report. *Clin Orthop* 1978;135:69-70
16. Rouviex B, Lassoued K, Vittecoq D, Regnier B. Neutropenia due to β lactamase antibodies. *Br Med J* 1983;287:1832-4
17. Bönner G, Helber A, Saborowski F. Granulozytopenie unter hochdosierter, intravenöser Penicillintherapie. *Therapiewoche* 1979;29:3925-8
18. Colvin B, Rogers M, Layton C. Benzylpenicillin-induced leukopenia. Complication of treatment of bacterial endocarditis. *Br Heart J* 1974;36:216-9
19. Corbett GM, Perry DJ, Shaw TRD. Penicillin-induced leukopenia [letter]. *N Engl J Med* 1982;307:1642-3
20. Forshaw J. Penicillin-induced granulopenia [letter]. *Br Med J* 1968;3:184
21. Joorabchi B, Kohout E. Apparent penicillin-induced arrest of mature bone marrow elements. *Br Med J* 1973;2:26-7
22. Korting HC, Pfeiffer B, Schranz W. Penicillin-G-induzierte Leukopenia bei Endocarditis lenta. *Z Kardiol* 1979;68:177-9
23. Kumar K, Kumar A. Reversible neutropenia associated with ampicillin therapy in pediatric patients. *Drug Intell Clin Pharm* 1981;15:802-6
24. Miro JM, Gatell JM, Moreno A, Mensa J, Roca J, Garcia San Miguel J. More on penicillin-induced neutropenia [letter]. *N Engl J Med* 1983;308:901-2
25. Murphy MF, Riordan T, Minchinton RM, Chapman JF, Amess JAL, Shaw EJ, Waters AH. Demonstration of an immune-mediated mechanism of penicillin-induced neutropenia and thrombocytopenia. *Br J Haematol* 1983;55:155-60
26. Paradisi F, Cioffi R, Ceccarelli G. Granulocitopenia e piastrinopenia probabilmente dovute alla benzil-penicillina ed alla cefalotina. *Giornale di Malattie Infettive e Parassitarie* 1977;29:251-6

27. Petz LD, Fudenberg HH. Coombs-positive hemolytic anemia caused by penicillin administration. *N Engl J Med* 1966;**274**:171-8
28. Postelnick M, Gaskins JD. Penicillin G-induced granulocytopenia. *Drug Intell Clin Pharm* 1981;**15**:289
29. Rossiter MA, Gray IR, Shinton NK. Penicillin-induced haemolytic anaemia [letter]. *Br Med J* 1968;**3**:616-7
30. Timmis AD, Crofts MA, Metcalfe J, Monaghan MJ, Sharp J, Jackson G. Gonococcal endocarditis with penicillin-induced bone marrow hypoplasia. Role of endocardiography. *JAMA* 1981;**246**:672-3
31. Yap PK. Severe leucopenia and fever due to benzylpenicillin – a rare complication of treatment with a common antibiotic. *Singapore Med J* 1981;**22**:99-101
32. Allo M, Silva J Jr. Antibiotic agranulocytosis: association with cephalothin and carbenicillin. *South Med J* 1977;**70**:1017-9
33. Davis A, Seligman SJ, Hewitt WL, Finegold SM. Neutropenic reaction to cephalothin therapy. In: Sylvester JC, ed. *Antimicrobial Agents and Chemotherapy – 1963*. Washington, DC: American Society for Microbiology, 1963:272-7
34. DiCato M-A, Ellman L. Cephalothin-induced granulocytopenia [letter]. *Ann Intern Med* 1975;**83**:671-2
35. Galanaud P, Second P, Subtil E, Derenne J-P. A propos de 4 observations d'agranulocytose au cours de traitements par la céphalotine. *Ann Med Interne (Paris)* 1976;**127**:579-83
36. Holt S, Khan MM, Orlans DA, Epstein EJ. Cephalothin induced neutropenia during the treatment of bacterial endocarditis. *Scott Med J* 1978;**23**:135-9
37. LePorrier M, Marquet M, Prier A, Binet J-L. Agranulocytose aiguë réversible lors d'un traitement par la cefalexine. *Ann Med Interne (Paris)* 1976;**127**:461-6
38. Tartas NE, Bullorsky EO, Hevia JEJ, Avalos JCS. Pancytopenia induced by cephalothin. *JAMA* 1981;**245**:1148-9
39. Carpenter J. Neutropenia induced by semisynthetic penicillin. *South Med J* 1980;**73**:745-8
40. Couchonnal GJ, Hinthorn DR, Hodges GR, Liu C. Nafcillin-associated granulopenia. *South Med J* 1978;**71**:1356-8
41. Dutro MP, Piccoro JJ Jr, Wilson HD. Nafcillin-induced neutropenia in two children. *Am J Hosp Pharm* 1981;**38**:889-92
42. Greene GR, Cohen E. Nafcillin-induced neutropenia in children. *Pediatrics* 1978;**61**:94-7
43. Markowitz SM, Rothkopf M, Holden FD, Stith DM, Duma RJ. Nafcillin-induced agranulocytosis. *JAMA* 1975;**232**:1150-2
44. Nahata MC, DeBolt SL, Powell DA. Adverse effects of methicillin, nafcillin and oxacillin in pediatric patients. *Dev Pharmacol Ther* 1982;**4**:117-23
45. Sandberg M, Tuazon CU, Sheagren JN. Neutropenia probably resulting from nafcillin. *JAMA* 1975;**232**:1152-4
46. Wilson C, Greenwood G, Remington JS, Vosti KL. Neutropenia after consecutive treatment courses with nafcillin and piperacillin [letter]. *Lancet* 1979;**1**:1150
47. Wolf DJ, Resnick GD. Nafcillin-induced neutropenia. *NY State J Med* 1978;**78**:256-9
48. Zakhireh B, Root RK. Unusually high occurrence of drug reactions with nafcillin. *Yale J Biol Med* 1978;**51**:449-55
49. Bullock WE Jr. Evaluation of 2,6-dimethoxyphenyl penicillin therapy in staphylococcal sepsis. In: Finland, M, Savage GM, eds. *Antimicrobial Agents and Chemotherapy – 1961*. Washington, DC: American Society for Microbiology, 1961:770-8
50. Godin M, Deshayes P, Ducastelle T, Delpech A, Leloët X, Fillastre JP. Agranulocytosis, haemorrhagic cystitis and acute interstitial nephritis during methicillin therapy [letter]. *J Antimicrob Chemother* 1980;**6**:296-7
51. Kitzing W, Nelson JD, Mohs E. Comparative toxicities of methicillin and nafcillin. *Am J Dis Child* 1981;**135**:52-5
52. Levitt BH, Gottlieb AJ, Roenberg IR, Klein JJ. Bone marrow depression due to methicillin, a semisynthetic penicillin. *Clin Pharmacol Ther* 1964;**5**:301-6
53. McElfresh AE, Huang NN. Bone-marrow depression resulting from the administration of methicillin. With a comment on value of serum iron determination. *N Engl J Med* 1962;**266**:246-7
54. Yow MD, Taber LH, Barrett FF, Mintz AA, Blankinship GR, Clark GE, Clark DJ. A ten-year assessment of methicillin-associated side effects. *Pediatrics* 1976;**58**:329-34
55. Krafft T, Pugin P, Miescher PA. Agranulocytose et cloxaciline intraveineuse. *Schweiz Med Wochenschr* 1978;**108**:1821-3
56. Shah I, Kumar KS, Lerner AM. Agranulocytosis associated with chronic oral administration of cloxacillin for suppression of staphylococcal osteomyelitis. *Am J Hematol* 1982;**12**:203-6
57. Westerman EL, Bradshaw MW, Williams TW Jr. Agranulocytosis during therapy with orally administered cloxacillin. *Am J Clin Pathol* 1978;**69**:559-60
58. Fournier-Carrie M, Vigne G, Duruy H. Granulopénie après traitement par ampicilline. *Nouvelle Presse Médicale* 1983;**12**:1606-7
59. Graf M, Tarlov A. Agranulocytosis with monohistiocytosis associated with ampicillin therapy. *Ann Intern Med* 1968;**69**:91-5
60. Korting HC, Schranz W. Knochenmarksdepression mit peripherer Panzytopenie unter der Gabe einer Ampicillin/Oxacillin-Kombination. *Z Kardiol* 1979;**68**:772-5
61. Sidi Y, Livni E, Solomon F, Pinkhas J. Ampicillin-induced neutropenia. *Haematologica (Pavia)* 1981;**66**:216-9
62. Stille W. Reversible allergische Agranulozytose unter der Therapie mit Ampicillin. *Med Welt* 1969;**36**:1981-4
63. Reyes MP, Palutke M, Lerner AM. Granulocytopenia associated with carbenicillin: five episodes in two patients. *Am J Med* 1973;**54**:413-8
64. Mader JT, LeFrock JL, Hyams KC, Molavi A, Reinartz JA. Cefotaxime therapy for patients with osteomyelitis and septic arthritis. *Rev Infect Dis* 1982;**4**(Suppl):S472-80
65. Ohsawa T, Furukawa F. Neutropenia associated with cefotaxime. *Drug Intell Clin Pharm* 1983;**17**:739-41
66. Bouza E, Buzón L, Martínez-Beltran J, Rodríguez M, Moneo I, Burgaleta C. Complications of the treatment of bacterial infections with piperacillin. In: Nelson JD, Grassi C, eds. *Current chemotherapy and infectious diseases*. Washington, DC: American Society for Microbiology, 1980:300-3
67. Gastineau D, Spector R, Phillips D. Severe neutropenia associated with ticarcillin therapy [letter]. *Ann Intern Med* 1981;**94**:711-2
68. Irvine AE, Morris TCM, Kelly GJ, McCracken N. Ticarcillin-induced neutropenia corroborated by an in vitro CFU-C toxicity. *Acta Haematol (Basel)* 1983;**70**:364-8
69. Ohning BL, Reed MD, Doershuk CF, Blumer JL. Ticarcillin-

- associated granulocytopenia. *Am J Dis Child* 1982;136:645-6
70. Eron LJ, Park CH, Hixon DL, Goldenberg RI, Poretz DM. Ceftazidime in patients with pseudomonas infections. *J Antimicrob Chemother* 1983;12(Suppl A):161-9
 71. Stewart GT, Holt RJ. Laboratory and clinical results with cephaloridine. *Lancet* 1964;2:1305-9
 72. Stille W, Becker HJ. Allergische Neutropenie unter der Behandlung mit Cephaloridin. *Int J Clin Pharmacol Ther Toxicol. Sonderheft Cephalosporine* 1968:92-4
 73. Schmid L, Heit W, Flury R. Agranulocytosis associated with semisynthetic penicillins and cephalosporines: review of 7 cases. *Blut* 1984;48:11-8
 74. Tsujimoto T, Yamaguchi S, Maruyama H. Clinical evaluation of mezlocillin in infectious diseases. *Chemotherapy (Tokyo)* 1979;27(Suppl 1):184-90
 75. Aronoff SC, Murdell D, O'Brien CA, Klinger JD, Reed MD, Blumer JL. Efficacy and safety of ceftriaxone in serious pediatric infections. *Antimicrob Agents Chemother* 1983;24:663-6
 76. Hofmann G, Saule H. Neutropenia als Nebenwirkung von Azlocillin bei einem Kind. *Klin Pädiatr* 1978;19:418-20
 77. Warren JW, Miller EH Jr, Fitzpatrick B, DeFranco DE, Caplan ES, Tenney JH, Anthony WC. A randomized, controlled trial of cefoperazone vs. cefamandole-tobramycin in the treatment of putative, severe infections with gram-negative bacilli. *Rev Infect Dis* 1983;5(Suppl):S173-80
 78. Levison ME, Bran JL, Jepson JH, Kaye D. Neutropenia associated with cephapirin therapy. *Antimicrob Agents Chemother* 1972;1:174-6
 79. Korzeniowski O, Sande MA, the National Collaborative Endocarditis Study Group. Combination antimicrobial therapy for *Staphylococcus aureus* endocarditis in patients addicted to parenteral drugs and in nonaddicts. A prospective study. *Ann Intern Med* 1982;97:496-503
 80. Marier RL, Sanders CV, Faro S, Janney A, Williams WW, Derks F, Aldridge KE. Piperacillin v carbenicillin in the therapy for serious infections. *Arch Intern Med* 1982;142:2000-5
 81. St John MA, Prober CG. Side effects of cloxacillin in infants and children. *Can Med Assoc J* 1981;125:458-60
 82. Winston DJ, Murphy W, Young LS, Hewitt WL. Piperacillin therapy for serious bacterial infections. *Am J Med* 1980;69:255-61
 83. Eron LJ, Goldenberg RI, Poretz DM, Park CH. Piperacillin therapy for pseudomonas infections. *South Med J* 1983;76:859-62
 84. Neftel KA, Wälti M, Schulthess HK, Gubler J. Adverse reactions following intravenous penicillin-G relate to degradation of the drug in vitro. *Klin Wochenschr* 1984;62:25-9
 85. Kancir LM, Tuazon CU, Cardella TA, Sheagren JN. Adverse reactions to methicillin and nafcillin during treatment of serious *Staphylococcus aureus* infections. *Arch Intern Med* 1978;138:909-11
 86. Müller MR, Neftel KA, Wälti M, Hauser SP, Erni J. Inhibition of in vitro granulopoiesis by betalactam antibiotics [abstract]. *Exp Hematol* 1983;11(Suppl 14):181
 87. Weitzman SA, Stosel TP, Desmond M. Drug-induced immunological neutropenia. *Lancet* 1978;1:1068-72
 88. Levine AS, Weiner RS, Fudenberg HH, Spath P, Petz L. Granulocytopenia caused by anticephalothin antibodies [abstract]. *Clinical Research* 1971;19:424A
 89. Neftel KA, Wälti M, Spengler H, de Weck AL. Effect of storage of penicillin-G solutions on sensitisation to penicillin-G after intravenous administration. *Lancet* 1982;1:986-8
 90. Neftel KA, Schulthess HK, Wälti M, Jäger A, Hirschi T, Gatell IM. Spättyreaktionen auf Penicilline sind dosisabhängig und korrelieren nicht mit Medikamenten-spezifischen Serumantikörpern [abstract]. *Schweiz Med Wochenschr* 1984;114:1778
 91. Chaperon EA, Sanders WE Jr. Suppression of lymphocyte responses by cephalosporins. *Infect Immun* 1978;19:378-84
 92. Manzella JP, Clark JK. Effects of moxalactam and cefuroxime on mitogen-stimulated human mononuclear leukocytes. *Antimicrob Agents Chemother* 1983;23:360-3
 93. Vonen B, Mørland J. Penicillin toxicity in isolated rat hepatocytes revealed by decreased incorporation of valine into proteins. *Acta Pharmacol Toxicol (Copenh)* 1982;51:81-6
 94. Schwarze PE, Seglen PO. Effects of antibiotics on protein synthesis and degradation in primary cultures of rat hepatocytes. *In Vitro* 1981;17:71-6
 95. Young LS. Antimicrobial chemotherapy of tumor patients in the United States. *Fortschritte der Antimikrobielle und Antineoplastischen Chemotherapie* 1984;3-6:953-8