

Intercontinental spread of OXA-48 beta-lactamase-producing *Enterobacteriaceae* over a 11-year period, 2001 to 2011

A Potron¹, L Poirel^{1,2}, E Rondinaud¹, P Nordmann (patrice.nordmann@unifr.ch)^{1,2}

1. INSERM U914, Emerging Resistance to Antibiotics, Faculté de Médecine et Université Paris-Sud, K. Bicêtre, France
2. Medical and Molecular Microbiology Unit, Department of Medicine, Faculty of Science, University of Fribourg, Fribourg, Switzerland

Citation style for this article:

Potron A, Poirel L, Rondinaud E, Nordmann P. Intercontinental spread of OXA-48 beta-lactamase-producing *Enterobacteriaceae* over a 11-year period, 2001 to 2011. *Euro Surveill.* 2013;18(31):pii=20549. Available online: <http://www.eurosurveillance.org/ViewArticle.aspx?ArticleId=20549>

Article submitted on 14 November 2012 / published on 01 August 2013

OXA-48 beta-lactamase producers are emerging as an important threat mostly in the Mediterranean area. We report here the molecular epidemiology of a collection of OXA-48 beta-lactamase-positive enterobacterial isolates (n=107) recovered from European and north-African countries between January 2001 and December 2011. This collection included 67 *Klebsiella pneumoniae*, 24 *Escherichia coli* and 10 *Enterobacter cloacae*. Using the EUCAST breakpoints, ninety-eight isolates (91.6%) were of intermediate susceptibility or resistant to ertapenem, whereas 66% remained susceptible to imipenem. Seventy-five per cent of the isolates co-produced an extended-spectrum beta-lactamase, most frequently CTX-M-15 (77.5%). Susceptibility testing to non-beta-lactam antibiotics showed that colistin, tigecycline, amikacin, and fosfomycin remain active against most of the isolates. Multilocus sequence typing indicated that the most common sequence types (ST) were ST101 and ST38 for *K. pneumoniae* and *E. coli*, respectively. The *bla*_{OXA-48} gene was located on a 62 kb IncL/M plasmid in 92.5% of the isolates, indicating that a single plasmid was mainly responsible for the spread of that gene. In addition, this study identified multiple cases of importation of OXA-48 beta-lactamase producers at least in Europe, and spread of OXA-48 beta-lactamase producers giving rise to an endemic situation, at least in France.

Introduction

Currently, an emergence of carbapenem resistance in *Enterobacteriaceae* is reported, mostly related to the spread of carbapenemases [1]. Those carbapenem-hydrolysing beta-lactamases belong to the Ambler class A (e.g. KPC), class B (e.g. IMP, VIM and NDM) [1], and class D (e.g. OXA-48 and its variants possessing weaker but significant carbapenemase activity) [2]. OXA-48 had first been identified from a clinical *Klebsiella pneumoniae* isolate recovered in Istanbul, Turkey, in 2001 [3]. The corresponding gene, namely *bla*_{OXA-48}, was then also identified in *Escherichia coli* and *Citrobacter freundii*, still in Turkey [4]. For

several years, OXA-48 was identified only in Turkey, and almost all OXA-48 beta-lactamase producers were reported from patients hospitalised in Turkey or with a link to that country [4,5]. Since 2008, this gene has been identified in many other countries, most often in *K. pneumoniae* isolates [2,5-10]. OXA-48 is now identified in the Middle East and in North African countries, and those countries are considered as reservoirs of OXA-48 beta-lactamase producers [2]. In addition to sporadic cases, an increasing number of outbreaks due to OXA-48-producing *K. pneumoniae* are currently observed, not only in Turkey but also in Belgium, France, Greece, the Netherlands and Spain [2,11-13]. *K. pneumoniae* strains belonging to specific sequence types (ST), such as ST395 and ST101, have been involved in those outbreaks [12,14].

In order to gain further understanding of that phenomenon, our study aimed at comparing the genetic features of OXA-48 beta-lactamase-producing strains recovered from various countries by analysing an existing collection of 107 *bla*_{OXA-48}-positive enterobacterial isolates. The genetic context and the location of the *bla*_{OXA-48} gene were investigated, as well as resistance to broad-spectrum beta-lactams and non-beta-lactam antibiotics.

Methods

Bacterial isolates

A total of 107 OXA-48 beta-lactamase-producing enterobacterial isolates were investigated retrospectively. *Enterobacteriaceae* producing OXA-48-like beta-lactamases were not included in this study. All isolates had been recovered from clinical specimens except a single isolate (one *Serratia marcescens* strain from an environmental water sample in Morocco), and had been received between January 2001 and December 2011 in our National Reference Laboratory which is also used as an International Reference Laboratory by many colleagues worldwide who send us their isolates for

further characterisation. Of identical strains in an outbreak, only one was included in this work. The distribution of clinical samples was as follows: rectal swabs (n=33), urine samples (n=24), blood samples (n=12), wound samples (n=7), respiratory specimens (n=4), catheters (n=4), bone specimens (n=2), peritoneal fluids (n=2), and placenta specimen (n=1). One sample per patient was included. Detailed information could not be obtained for 18 clinical samples. The isolates were identified to species level using the API 20E system (bioMérieux, La Balme-les-Grottes, France).

Susceptibility testing

Routine antibiograms were determined by disk diffusion method on Mueller-Hinton (MH) agar (Bio-Rad, Marnes-la-Coquette, France) and interpreted using the guidelines of the Clinical and Laboratory Standards Institute (CLSI) (updated 2012) and of the European Committee on Antimicrobial Susceptibility Testing (EUCAST) for tigecycline and colistin [15,16]. In addition, MICs were determined for imipenem, meropenem, ertapenem, cefotaxime, and ceftazidime using E-test (bioMérieux, La Balme-les-Grottes, France). The production of extended-spectrum beta-lactamases (ESBL) was evidenced by a double-disk synergy test performed with cefepime, ceftazidime, and ticarcillin/clavulanic acid disks [17] and more recently by using the rapid ESBL NDP test [18].

PCR and sequencing of beta-lactamase-encoding genes

Whole-cell DNA was extracted using the QiaAmp DNA minikit and following the manufacturer's recommendations (Qiagen, Courtaboeuf, France). All isolates were screened by PCR for the Ambler class A and B carbapenemase-encoding genes *bla*_{KPC}, *bla*_{IMP}, *bla*_{VIM}, *bla*_{NDM} [19-20]. For each isolate, the *bla*_{OXA-48} gene was amplified using primers preOXA-48A and preOXA-48B, and subsequently sequenced [21]. Detection of other beta-lactamase genes such as *bla*_{TEM}, *bla*_{SHV}, *bla*_{CTX-M}, *bla*_{AmpC}-like, and *bla*_{OXA-1} was performed with internal primers, as described previously [19,22]. PCR products were analysed on agarose gel and sequenced by using the amplification primers with an automated sequencer (ABI PRISM 3100; Applied Biosystems). The nucleotide and deduced protein sequences were analysed using software from the National Center for Biotechnology Information (www.ncbi.nlm.nih.gov).

Strain typing

Multilocus sequence typing (MLST) with seven housekeeping genes (*rpoB*, *gapA*, *mdh*, *pgi*, *phoE*, *infB* and *tonB*) was performed for *K. pneumoniae* isolates according to Diancourt et al. [23]. Allele sequences and STs were verified at <http://pubmlst.org/Kpneumoniae>. Fragments of seven housekeeping genes (*adk*, *fumC*, *gyrB*, *icd*, *mdh*, *purA* and *recA*) were amplified and sequenced for *E. coli* isolates as described on the following website <http://mlst.ucc.ie/mlst/dbs/Ecoli>. A different allele number was given to each distinct sequence within a locus, and a distinct ST number

was attributed to each distinct combination of alleles. *E. coli* isolates were assigned to the major *E. coli* phylogenetic groups (A, B1, B2, and D) by multiplex PCR, as described [24]. The genetic relationship between the *Enterobacter cloacae* isolates was studied using Diversilab, a semi-automated typing system based on repetitive sequence-based PCR (rep-PCR) following the manufacturer's instructions (bioMérieux).

Plasmid DNA analysis, transformation and mating-out assays

Plasmid DNA was extracted from the isolates using the Kieser technique [25]. *E. coli* NCTC50192, harbouring four plasmids of 154, 66, 48 and 7 kb, was used as plasmid size marker. Plasmid DNAs were analysed by agarose gel electrophoresis. Direct transfer of the carbapenem resistance markers was attempted by liquid mating-out assays at 37°C, using *E. coli* J53 as recipient, or by electrotransformation of plasmid DNA, using *E. coli* TOP10 as recipient as reported [3,4]. Selection was performed on agar plates supplemented with ertapenem (0.5 µg/ml) and azide (100 µg/ml) for mating-out assays. In order to search for a possible chromosomal location of the *bla*_{OXA-48} gene in *E. coli* isolates 19 to 24, restriction with endonuclease I-CeuI followed by pulsed-field gel electrophoresis (PFGE) analysis was performed as described [26].

Replicon and transposon typing

PCR-based replicon typing (PBRT) of the main plasmid incompatibility groups reported in *Enterobacteriaceae* was performed as described [27] and using the specific primers designed from plasmid pOXA-48a [28]. Genetic structures surrounding the *bla*_{OXA-48} gene were determined according to the Tn1999-like PCR-mapping scheme as described [29].

Results

Bacterial isolates

A total of 107 isolates were studied, including *K. pneumoniae* (n=67), *E. coli* (n=24), and *E. cloacae* (n=10) (Table 1). Other enterobacterial species were identified: *Citrobacter koseri* (n=2), *C. freundii* (n=1), *Klebsiella oxytoca* (n=1), *Providencia rettgeri* (n=1), and *S. marcescens* (n=1). They had been isolated in France (n=61), Morocco (n=22), Turkey (n=11), Egypt (n=3), Lebanon (n=2), Tunisia (n=2), Switzerland (n=2), South Africa (n=2), Belgium (n=1), and the Netherlands (n=1), respectively (Table 1). Among the 61 strains collected in France, 30 had a history of international travel to the following countries: Morocco (n=14), Tunisia (n=2), Libya (n=5), Algeria (n=4), Egypt (n=3), Senegal (n=1), and Kuwait (n=1). In 12 cases, no travel history from a foreign country was identified. For the remaining 19 cases, no precise travel information could be obtained (Table 1).

TABLE 1A

Genetic features associated with OXA-48 beta-lactamase producers, 2001–11 (n=107)

Species	Country of isolation	Travel history	beta-lactams MICs (µg/mL)				Sequence type	Genetic location of <i>bla</i> _{OXA-48}	Incompatibility group of <i>bla</i> _{OXA-48} plasmid	Non-beta-lactam-associated resistances	Associated beta-lactam resistance determinants ^a	Phylogenetic group	Transposon bearing <i>bla</i> _{OXA-48}
			ERT	IMP	MER	CAZ							
<i>Enterobacter cloacae</i>	France	Morocco	8	1	1	64	>256	ND	Q, Gm, Tm, TET, Cm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2	
<i>E. cloacae</i>	France	?	>32	1	1.5	32	>256	ND	Q, Tm, Ak, TET, TGC, Cm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2	
<i>E. cloacae</i>	France	None	4	0.5	0.5	>256	>256	ND	Q, Ami, TET, TGC, Cm, Fos, SXT, FT	CTX-M-15, TEM-1, OXA-1, DHA-7, SHV-12	ND	Tn1999.2	
<i>E. cloacae</i>	France	Algeria	3	0.75	0.38	48	>256	ND	FOS	CTX-M-15	ND	Tn1999.4	
<i>E. cloacae</i>	France	Morocco	16	1	0.75	32	>256	ND	Q, Gm, Tm, Cm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2	
<i>E. cloacae</i>	France	Morocco	1.5	0.5	0.25	16	>256	ND	Q, Gm, Tm, TET, Cm, SXT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.1	
<i>E. cloacae</i>	Morocco	None	1	0.5	0.38	0.5	8	ND	Ami, TET, SXT, FT	CTX-M-9, TEM-1	ND	Tn1999.2	
<i>E. cloacae</i>	Morocco	None	1	0.5	0.38	48	16	ND	OFX, Ami, TET, SXT, FT	CTX-M-9, TEM-1, SHV-12	ND	Tn1999.2	
<i>E. cloacae</i>	Morocco	None	0.75	0.38	0.19	8	128	ND	Gm, Tm, TET, Cm, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.2	
<i>E. cloacae</i>	Morocco	None	6	1	1	48	16	ND	OFX, Gm, Tm, Cm, SXT, FT	TEM-1, DHA-7, SHV-12	ND	Tn1999.2	
<i>C. koseri</i>	France	?	2	0.38	0.38	0.38	2	ND	None	None	ND	Tn1999.2	
<i>C. koseri</i>	France	?	2	0.75	0.38	2	2	ND	None	None	ND	Tn1999.2	
<i>Klebsiella oxytoca</i>	Morocco	None	4	1	0.5	256	256	ND	Q, Gm, Tm, TET, Cm, SXT	CTX-M-15, TEM-1	ND	Tn1999.2	
<i>Citrobacter freundii</i>	France	None	4	0.75	0.38	64	12	ND	Q, Gm, Tm, TET, TGC, Cm, SXT	SHV-12, TEM-1	ND	Tn1999.2	
<i>Providencia rettgeri</i>	Turkey	None	>32	>32	>32	32	16	ND	Q, Gm, Tm, TET, Cm, SXT, FT	TEM-101	ND	Tn1999.1	
<i>Serratia marcescens</i>	Morocco	None	>32	8	4	1	3	ND	Q, TET, Cm, SXT, FT	OXA-1	ND	Tn1999.1	
<i>E. coli</i>	France	Algeria	0.5	0.38	0.12	16	>256	10	Q, SXT	CTX-M-15, TEM-1	A	Tn1999.4	

Ak: amikacin; Ami: aminoglycosides; Cm: chloramphenicol; Cs: colistin; Δ Tn1999: truncated transposon Tn1999; FOS: fosfomicin; FT: nitrofurantoin; Gm: gentamicin; MIC: minimum inhibitory concentration; ND: not determinable; OFX: ofloxacin; Q: fluoroquinolones; SXT: sulfamethoxazole-trimethoprim; TET: tetracycline; TGC: tigecycline; Tm: tobramycin.

^a Resistance markers being co-harboured by the *bla*_{OXA-48}-carrying plasmid are underlined.

TABLE 1B

Genetic features associated with OXA-48 beta-lactamase producers, 2001–11 (n=107)

Species	Country of isolation	Travel history	beta-lactams MICs (µg/mL)				Sequence type	Genetic location of <i>bla</i> _{OXA-48}	Incompatibility group of <i>bla</i> _{OXA-48} -positive plasmid	Non-beta-lactam-associated resistances	Associated beta-lactam resistance determinants ^a	Phylogenetic group	Transposon bearing <i>bla</i> _{OXA-48}
			ERT	IMP	MER	CAZ							
<i>E. coli</i>	France	Libya	1	0.5	0.19	>256	32	Plasmidic	Incl/M	Ak, Tm, TET, Cm, SXT	CMY-2, VEB-8, TEM-1	A	Tn1999.2
<i>E. coli</i>	France	Egypt	3	0.5	0.19	0.5	24	Chromosomal	ND	OFX, Gm, Tm, SXT	CTX-M-24, TEM-1	D	ΔTn1999.2
<i>E. coli</i>	France	Turkey	3	0.75	0.38	0.75	24	Chromosomal	ND	OFX, Gm, Tm, SXT	CTX-M-24, TEM-1	D	ΔTn1999.2
<i>E. coli</i>	France	Egypt	0.5	0.25	0.19	0.5	24	Chromosomal	ND	OFX, Gm, Tm, SXT	CTX-M-24, TEM-1	D	ΔTn1999.2
<i>E. coli</i>	France	None	1	0.38	0.25	0.5	24	Chromosomal	ND	OFX, Gm, Tm, SXT	CTX-M-24, TEM-1	D	ΔTn1999.2
<i>E. coli</i>	Switzerland	?	8	0.75	1	1.5	48	Chromosomal	ND	OFX, Gm, Tm, SXT	CTX-M-24, TEM-1	D	ΔTn1999.2
<i>E. coli</i>	Egypt	None	2	0.5	0.25	0.5	24	Chromosomal	ND	OFX, Gm, Tm, SXT	CTX-M-24, TEM-1	D	ΔTn1999.2
<i>E. coli</i>	Lebanon	None	2	0.75	0.25	1.5	48	Plasmidic	Incl/M	SXT	CTX-M-14, TEM-1	D	Tn1999.2
<i>E. coli</i>	France	?	0.5	0.38	0.12	0.12	0.09	Plasmidic	Incl/M	Q, TET, SXT	TEM-1	A	Tn1999.2
<i>E. coli</i>	France	Egypt	0.5	0.5	0.19	8	>256	Plasmidic	Incl/M	TET, SXT	CTX-M-15, TEM-1	D	Tn1999.2
<i>E. coli</i>	France	None	0.75	0.5	0.25	0.12	0.38	Plasmidic	Incl/M	None	TEM-1	B2	Tn1999.2
<i>E. coli</i>	Egypt	None	>32	32	>32	>256	256	Plasmidic	Incl/M	Q, Ami, TET, Cm, SXT	TEM-1, VIM-1, CMY-4	B1	Tn1999.1
<i>E. coli</i>	France	?	3	1.5	0.5	0.19	1	Plasmidic	Incl/M	TET, SXT	TEM-1	D	Tn1999.2
<i>E. coli</i>	France	Morocco	2	0.38	0.25	0.5	0.5	Plasmidic	Incl/M	Q, TET	None	A	Tn1999.2
<i>E. coli</i>	France	?	0.75	0.38	0.19	16	128	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT	CTX-M-15, OXA-1	A	Tn1999.2
<i>E. coli</i>	France	?	0.75	0.38	0.19	8	64	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT	CTX-M-15, OXA-1	A	Tn1999.2
<i>E. coli</i>	Turkey	None	24	1.5	12	16	192	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT	CTX-M-15, TEM-1, OXA-1	D	Tn1999.1
<i>E. coli</i>	Turkey	None	>32	1.5	12	16	256	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT	CTX-M-15, TEM-1, OXA-1	D	Tn1999.1
<i>E. coli</i>	France	Morocco	1.5	0.38	0.25	24	>256	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, SXT	CTX-M-15, TEM-1, OXA-1	A	Tn1999.1
<i>E. coli</i>	France	?	1.5	0.5	0.19	24	256	Plasmidic	Inc F	None	CTX-M-15, TEM-1	D	ΔTn1999.1
<i>E. coli</i>	France	?	1	0.5	0.19	0.12	0.75	Plasmidic	Incl/M	None	None	B2	Tn1999.2
<i>E. coli</i>	France	None	1	0.5	0.19	1.5	24	Plasmidic	Incl/M	TET	CTX-M-15	D	Tn1999.2

Ak: amikacin; Ami: aminoglycosides; Cm: chloramphenicol; Cs: colistin; ΔTn1999: truncated transposon Tn1999; FOS: fosfomicin; FT: nitrofurantoin; Gm gentamicin; MIC: minimum inhibitory concentration; ND: not determinable; OFX: ofloxacin; Q: fluoroquinolones; SXT: sulfamethoxazole-trimethoprim; TET: tetracycline; TGC: tigecycline; Tm: tobramycin.

^a Resistance markers being co-harboured by the *bla*_{OXA-48}-carrying plasmid are underlined.

TABLE 1C

Genetic features associated with OXA-48 beta-lactamase producers, 2001–11 (n=107)

Species	Country of isolation	Travel history	beta-lactams MICs (µg/mL)				Sequence type	Genetic location of <i>bla</i> _{OXA-48}	Incompatibility group of <i>bla</i> _{OXA-48} plasmid	Non-beta-lactam-associated resistances	Associated beta-lactam resistance determinants ^a	Phylogenetic group	Transposon bearing <i>bla</i> _{OXA-48}	
			ERT	IMP	MER	CAZ								CTX
<i>E. coli</i>	France	Morocco	0.75	0.5	0.19	1	16	2969	Plasmidic	Incl/M	TET	CTX-M-15	D	Tn1999.2
<i>K. pneumoniae</i>	Morocco	None	6	0.75	0.75	192	256	11	Plasmidic	Incl/M	Q, Ami, TET, Cm, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Turkey	None	≥32	≥32	≥32	≥256	64	14	Plasmidic	Incl/M	Q, Ami, Cm, SXT	SHV-2a, TEM-1, OXA-47	ND	Tn1999.1
<i>K. pneumoniae</i>	Turkey	None	≥32	2	4	256	48	14	Plasmidic	Incl/M	Q, Ak, Tm, FT	SHV-12, TEM-1, OXA-1	ND	Tn1999.1
<i>K. pneumoniae</i>	Turkey	None	≥32	≥32	≥32	≥256	48	14	Plasmidic	Incl/M	Q, Ami, Cm, FOS	OXA-1, TEM-1, SHV-12	ND	Tn1999.1
<i>K. pneumoniae</i>	Egypt	None	1.5	2	0.75	0.19	1.5	14	Plasmidic	Incl/M	FT	OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	?	≥32	8	6	1	3	15	Plasmidic	Incl/M	Q, Tm, TET, TGC, Cm, FOS, SXT, FT	TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	Morocco	2	0.5	0.5	192	≥256	15	Plasmidic	Incl/M	Q, Gm, Tm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	Morocco	2	1	0.5	48	128	15	Plasmidic	Incl/M	Q, Gm, Tm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Morocco	None	12	1	2	8	6	15	Plasmidic	Incl/M	Q, Ami, TET, TGC, Cm, FOS, SXT, FT	DHA-1, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Morocco	None	≥32	≥32	≥32	256	≥256	15	Plasmidic	Incl/M	Q, Gm, Tm, TET, FOS, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.1
<i>K. pneumoniae</i>	Morocco	None	12	1	2	8	6	15	Plasmidic	Incl/M	Q, Ami, TET, TGC, Cm, FOS, SXT, FT	DHA-1, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Morocco	None	2	0.38	0.5	24	≥256	15	Plasmidic	Incl/M	Q, Gm, Tm, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Turkey	None	2	0.5	0.5	64	≥256	16	Plasmidic	Incl/M	Q, Tm, Ak, TET, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Morocco	None	0.38	0.38	0.5	12	96	25	Plasmidic	Incl/M	Q, Gm, Tm, TET, TGC, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Morocco	None	0.38	0.38	0.5	12	96	25	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.1
<i>K. pneumoniae</i>	France	Koweit	≥32	32	24	1.5	2	29	Plasmidic	Incl/M	OFX, TET, TGC, Cm, FOS, SXT, FT	None	ND	Tn1999.2

Ak: amikacin; Ami: aminoglycosides; Cm: chloramphenicol; Cs: colistin; ΔTn1999: truncated transposon Tn1999; FOS: fosfomicin; FT: nitrofurantoin; Gm: gentamicin; MIC: minimum inhibitory concentration; ND: not determinable; OFX: ofloxacin; Q: fluoroquinolones; SXT: sulfamethoxazole-trimethoprim; TET: tetracycline; TGC: tigecycline; Tm: tobramycin.

^a Resistance markers being co-harboured by the *bla*_{OXA-48}-carrying plasmid are underlined.

TABLE 1D

Genetic features associated with OXA-48 beta-lactamase producers, 2001–11 (n=107)

Species	Country of isolation	Travel history	beta-lactams MICs (µg/mL)				Sequence type	Genetic location of bla _{OXA-48}	Incompatibility group of bla _{OXA-48} -positive plasmid	Non-beta-lactam-associated resistances	Associated beta-lactam resistance determinants ^a	Phylogenetic group	Transposon bearing bla _{OXA-48}
			ERT	IMP	MER	CAZ							
<i>K. pneumoniae</i>	France	Morocco	0.5	0.38	0.25	0.19	0.38	Plasmidic	Incl/M	None	None	ND	Tn1999.2
<i>K. pneumoniae</i>	France	None	0.75	0.5	0.25	0.09	0.38	Plasmidic	Incl/M	TET, FT	None	ND	Tn1999.2
<i>K. pneumoniae</i>	France	?	1	0.5	0.25	0.12	0.5	Plasmidic	Incl/M	None	TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	?	1	0.5	0.25	0.12	0.25	Plasmidic	Incl/M	FOS	TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	?	0.75	0.5	0.25	0.19	0.25	Plasmidic	Incl/M	None	TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	None	4	4	1	0.5	0.5	Plasmidic	Incl/M	TET, FOS	None	ND	Tn1999.2
<i>K. pneumoniae</i>	Tunisia	None	≥32	1.5	12	48	≥256	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	?	≥32	24	16	32	≥256	Plasmidic	Incl/M	Q, Gm, Tm, TET, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Tunisia	None	≥32	2	8	≥256	≥256	Plasmidic	Incl/M	Ami, TET, Cm, FOS, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	None	3	0.5	0.38	48	96	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Switzerland	?	6	0.5	0.75	48	≥256	Plasmidic	Incl/M	Q, Tm, Ak, TET, SXT, FT, TGC	CTX-M-15, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	Morocco	≥32	3	16	192	≥256	Plasmidic	Incl/M	Q, Gm, Tm, TET, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	None	4	0.5	0.5	48	128	Plasmidic	Incl/M	Q, Tm, SXT, FT	CTX-M-15, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	Libya	≥32	3	8	≥256	≥256	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	France	Libya	≥32	≥32	≥32	≥256	≥256	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	South Africa	?	≥32	3	8	≥256	≥256	Plasmidic	Incl/M	Q, Gm, Tm, TET, TGC, Cm, SXT, FT	CTX-M-15, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	South Africa	?	≥32	3	8	≥256	≥256	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cs, SXT, FT	CTX-M-15, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Morocco	None	≥32	≥32	16	192	≥256	Plasmidic	Incl/M	Q, Gm, Tm, TET, FOS, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i>	Morocco	None	≥32	≥32	≥32	256	≥256	Plasmidic	Incl/M	Q, Tm, Ak, TET, FOS, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.2

Ak: amikacin; Ami: aminoglycosides; Cm: chloramphenicol; Cs: colistin; ΔTn1999: truncated transposon Tn1999; FOS: fosfomicin; FT: nitrofurantoin; Gm: gentamicin; MIC: minimum inhibitory concentration; ND: not determinable; OFX: ofloxacin; Q: fluoroquinolones; SXT: sulfamethoxazole-trimethoprim; TET: tetracycline; TGC: tigecycline; Tm: tobramycin.

^a Resistance markers being co-harboured by the bla_{OXA-48}-carrying plasmid are underlined.

TABLE 1E

Genetic features associated with OXA-48 beta-lactamase producers, 2001–11 (n=107)

Species	Country of isolation	Travel history	beta-lactams MICs (µg/mL)				Sequence type	Genetic location of <i>bla</i> _{OXA-48}	Incompatibility group of <i>bla</i> _{OXA-48} positive plasmid	Non-beta-lactam-associated resistances	Associated beta-lactam resistance determinants ^a	Phylogenetic group	Transposon bearing <i>bla</i> _{OXA-48}	
			ERT	IMP	MER	CAZ								CTX
<i>K. pneumoniae</i> 78	Morocco	None	2	0.38	0.5	192	256	101	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 79	Morocco	None	2	0.38	0.5	>256	>256	101	Plasmidic	Incl/M	Q, Tm, Ak, TET, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 80	Belgium	None	4	1	1	1	0.5	147	Plasmidic	Incl/M	Q, TET, TgC, Cm, SXT	None	ND	Tn1999.2
<i>K. pneumoniae</i> 81	Turkey	None	>2	32	8	96	>256	147	Plasmidic	Incl/M	Q, Tm, Ak, TET, TgC, Cm, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 82	France	Tunisia	3	0.38	0.38	64	>256	147	Plasmidic	Incl/M	Q, Gm, Tm, TET, TgC, Cm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 83	France	?	2	0.38	0.38	96	192	147	Plasmidic	Incl/M	Q, Tm, Ak, FT	CTX-M-15, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i> 84	France	Libya	3	1.5	3	>256	>256	147	Plasmidic	Incl/M	Q, Gm, Tm, Ak, TET, Cm, SXT, FT	CTX-M-15, TEM-1, OXA-1, CMY-2	ND	Tn1999.2
<i>K. pneumoniae</i> 85	France	Libya	>2	3	8	>256	>256	147	Plasmidic	Incl/M	Q, Ami, TET, Cm, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1, CMY-2	ND	Tn1999.2
<i>K. pneumoniae</i> 86	France	?	1	0.25	0.25	48	>256	307	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 87	France	Algeria	0.5	0.38	0.19	32	192	336	Plasmidic	Incl/M	Q, Tm, Ak, SXT	CTX-M-15, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 88	Morocco	None	1	0.5	0.5	8	48	392	Plasmidic	Incl/M	Q, Gm, Tm, TET, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i> 89	Morocco	None	12	6	3	24	>256	392	Plasmidic	Incl/M	Q, Gm, Tm TET, Cm, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 90	France	None	3	0.75	0.75	96	>256	395	plasmidic	Incl/M	Q, Tm, Gm, TET, FOS, SXT, FT, Cm	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 91	France	None	2	0.5	0.38	1	1	395	Plasmidic	Incl/M	Q, Tm, Ak, TET, TgC, Cm, FOS, SXT, FT	OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 92	Netherlands	None	2	0.5	0.38	32	192	395	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 93	France	Morocco	2	0.38	0.38	256	256	395	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2

Ak: amikacin; Ami: aminoglycosides; Cm: chloramphenicol; Cs: colistin; ΔTn1999: truncated transposon Tn1999; FOS: fosfomicin; FT: nitrofurantoin; Gm: gentamicin; MIC: minimum inhibitory concentration; ND: not determinable; OFX: ofloxacin; Q: fluoroquinolones; SXT: sulfamethoxazole-trimethoprim; TET: tetracycline; TgC: tigecycline; Tm: tobramycin.

^a Resistance markers being co-harboured by the *bla*_{OXA-48}-carrying plasmid are underlined.

TABLE 1F

Genetic features associated with OXA-48 beta-lactamase producers, 2001–11 (n=107)

Species	Country of isolation	Travel history	beta-lactams MICs (µg/mL)					Sequence type	Genetic location of bla _{OXA-48}	Incompatibility group of bla _{OXA-48} -positive plasmid	Non-beta-lactam-associated resistances	Associated beta-lactam resistance determinants ^a	Phylogenetic group	Transposon bearing bla _{OXA-48}
			ERT	IMP	MER	CAZ	CTX							
<i>K. pneumoniae</i> 94	France	Morocco	≥32	24	32	128	≥256	395	Plasmidic	Incl/M	Q, Gm, Tm, TET, TGC, Cm, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 95	Morocco	None	3	0.5	0.5	≥256	≥256	395	Plasmidic	Incl/M	Q, Gm, Tm, TET, TGC, Cm, FOS, SXT, FT	CTX-M-15, TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i> 96	Morocco	None	3	0.5	0.5	≥256	≥256	395	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 97	France	Morocco	1.5	3	0.25	32	96	405	Plasmidic	Incl/M	Q, Gm, Tm, TET, FOS, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.1
<i>K. pneumoniae</i> 98	Lebanon	None	≥32	≥32	≥32	0.38	3	496	Plasmidic	Incl/M	FOS, SXT, FT	None	ND	Tn1999.2
<i>K. pneumoniae</i> 99	France	Algeria	1	0.38	0.19	0.25	0.25	530	Plasmidic	Incl/M	FT	TEM-1	ND	Tn1999.2
<i>K. pneumoniae</i> 100	France	?	4	0.75	0.5	0.5	0.75	685	Plasmidic	Incl/M	FT	None	ND	Tn1999.2
<i>K. pneumoniae</i> 101	Turkey	?	0.38	0.38	0.12	0.09	0.12	981	Plasmidic	Incl/M	OFX	None	ND	Tn1999.2
<i>K. pneumoniae</i> 102	Turkey	None	≥32	≥32	32	32	≥256	982	Plasmidic	Incl/M	OFX, Tm, TET, TGC, FOS, SXT, FT	CTX-M-15, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 103	Turkey	None	≥32	2	6	≥256	48	984	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, SXT, FT	SHV-12, TEM-1	ND	Tn1999.1
<i>K. pneumoniae</i> 104	France	Senegal	1	0.5	0.25	12	48	985	Plasmidic	Incl/M	OFX, Tm, Gm, TET, TGC, Cm, FOS, SXT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 105	France	Morocco	3	0.5	0.38	96	256	986	Plasmidic	Incl/M	Q, Gm, Tm, TET, Cm, SXT, FT	CTX-M-15, TEM-1, OXA-1	ND	Tn1999.2
<i>K. pneumoniae</i> 106	France	?	3	0.5	0.38	1.5	0.75	987	Plasmidic	Incl/M	OFX, TET, TGC, Cm, FT	None	ND	Tn1999.2
<i>K. pneumoniae</i> 107	Morocco	None	1	0.38	0.5	12	3	1027	Plasmidic	Incl/M	Q, Ami, TET, Cm, FOS, SXT, FT	TEM-1, SHV-27, DHA-1	ND	Tn1999.2

Ak: amikacin; Ami: aminoglycosides; Cm: chloramphenicol; Cs: colistin; ΔTn1999: truncated transposon Tn1999; FOS: fosfomicin; FT: nitrofurantoin; Gm: gentamicin; MIC: minimum inhibitory concentration; ND: not determinable; OFX: ofloxacin; Q: fluoroquinolones; SXT: sulfamethoxazole-trimethoprim; TET: tetracycline; TGC: tigecycline; Tm: tobramycin.

^a Resistance markers being co-harboured by the bla_{OXA-48}-carrying plasmid are underlined.

TABLE 2

Susceptibility to carbapenems of *Klebsiella pneumoniae*, *Escherichia coli* and *Enterobacter cloacae* isolates, 2001–11 (n=107)

Species	Antimicrobial drug	Susceptibility Number (%) of isolates		
		Susceptible	Intermediate	Resistant
<i>K. pneumoniae</i> (n=67)	Imipenem	40 (60)	13 (19)	14 (21)
	Ertapenem	5 (7)	9 (13)	53 (79)
	Meropenem	40 (60)	6 (9)	21 (31)
<i>E. coli</i> (n=24)	Imipenem	20 (84)	3 (12)	1 (4)
	Ertapenem	4 (17)	8 (33)	12 (50)
	Meropenem	21 (88)	0 (0)	3 (12)
<i>E. cloacae</i> (n=10)	Imipenem	10 (100)	0 (0)	0 (0)
	Ertapenem	0 (0)	3 (30)	7 (70)
	Meropenem	9 (90)	1 (10)	0 (0)

The percentages are rounded so as to add up to 100%.

The results in this Table are from E-tests.

Susceptibility to carbapenems and broad-spectrum cephalosporins

Results of susceptibility testing are shown in Tables 2 and 3. According to the CLSI guidelines, 40 (60%) *K. pneumoniae* isolates, 20 (83%) *E. coli* isolates, and 10 (100%) *E. cloacae* isolates were susceptible to imipenem (Table 2). In addition, 40 (60%) *K. pneumoniae*, 21 (88%) *E. coli*, and 9 (90%) *E. cloacae* isolates were susceptible to meropenem. By contrast, 62 (92%) *K. pneumoniae*, 20 (83%) *E. coli*, and 10 (100%) *E. cloacae* isolates were found of intermediate susceptibility or resistant to ertapenem (Table 2). Regarding the broad-spectrum cephalosporins, 73 (68%) and 90 (84%) isolates were resistant or of intermediate susceptibility to ceftazidime and cefotaxime, respectively (Table 1 and 3).

Beta-lactamase genes

Among the 107 OXA-48-producing isolates, 80 (75%) co-produced an ESBL. A *bla*_{CTX-M}-like gene was detected in 71 (66%) of the isolates (89% of the ESBL-producing isolates). Among the different CTX-M variants identified, those belonging to the CTX-M-1 and CTX-M-9 groups accounted for 87.5% (n=62) and 12.5% (n=9), respectively. CTX-M-15 was the only representative of the CTX-M-1 group. In the CTX-M-9 group, the *bla*_{CTX-M-14} was identified in a single *E. coli* (Table 1). Two *E. cloacae* isolates harboured a *bla*_{CTX-M-9} gene and 6 *E. coli* isolates harboured a *bla*_{CTX-M-24} gene. The other ESBL determinants were SHV-2a (one *K. pneumoniae*), SHV-12 (three *K. pneumoniae*, two *E. cloacae*, and one *C. freundii*), SHV-27 (one *K. pneumoniae*) and TEM-101 (*P. rettgeri* isolate no. 15). Among the SHV-12-producing isolates, one *E. cloacae* co-produced CTX-M-9 (Table 1). In addition, a novel VEB variant, namely VEB-8, was identified in a single *E. coli* isolate from Libya that

co-produced CMY-2 (*E. coli* 18). VEB-8 differed from VEB-5 by a single amino acid substitution (GenBank accession number JX679208) [30,31]. It is interesting to note that ESBLs were not related to date or geographic area of isolation.

Furthermore, nine isolates (8.5%) co-produced a plasmid-mediated AmpC-type beta-lactamase. Four isolates (3.8%) produced a CMY-type beta-lactamase, namely CMY-4 in a single *E. coli* isolate from Egypt and CMY-2 in three isolates (a single *E. coli* and two *K. pneumoniae* isolates from Libya). Five isolates (5%) produced a DHA-like AmpC, namely DHA-1 in three *K. pneumoniae* isolates and DHA-7 in two *E. cloacae* isolates. All the DHA-producing isolates originated from Morocco. A single isolate (*E. coli* 29 from Egypt) co-produced OXA-48 and another carbapenemase, namely VIM-1, in addition to CMY-4. The non-ESBL beta-lactamases TEM-1 and OXA-1 were detected in 79 (74%) and 47 (44%) isolates, respectively.

Susceptibility to non-beta-lactam antibiotics

Results of susceptibility testing for non-beta-lactam antibiotics are shown in Table 3. Four antibiotics were active against the majority of the isolates; 104 (99%) of the 107 isolates were susceptible to colistin, 90 (84.1%) to tigecycline, 83 (77.6%) to amikacin, and 77 (72%) to fosfomycin. Conversely, 84 (78.5%) of the 107 isolates were resistant to sulfamethoxazole-trimethoprim, 72 (67.3%) to tetracycline, 64 (59.8%) to ciprofloxacin, and 61 (57%) to gentamicin. Resistant isolates that produced an ESBL were mostly resistant also to non-beta-lactam antibiotics (Table 3).

TABLE 3

Susceptibility of the study isolates determined by disk diffusion method, 2001–11 (n=107)

Antimicrobial drug	Susceptibility Number (%) of isolates								
	Susceptible			Intermediate			Resistant		
	Total	ESBL	Non-ESBL	Total	ESBL	Non-ESBL	Total	ESBL	Non-ESBL
Ceftazidime	34 (31.8)	10 (9.4)	24 (22.4)	9 (8.4)	7 (6.5)	2 (1.9)	64 (59.8)	63 (58.9)	1 (0.9)
Cefotaxime	17 (15.9)	0 (0)	17 (15.9)	8 (7.5)	1 (1.0)	7 (6.5)	82 (76.6)	79 (73.8)	3 (2.8)
Tetracycline	34 (31.8)	21 (19.6)	13 (12.2)	1 (0.9)	1 (0.9)	0 (0)	72 (67.3)	58 (54.2)	14 (13.1)
Tigecycline	90 (84.1)	69 (64.5)	21 (19.6)	5 (4.7)	4 (3.7)	1 (1.0)	12 (11.2)	7 (6.5)	5 (4.7)
Fosfomycin	77 (72.0)	58 (54.2)	19 (17.8)	2 (1.8)	2 (1.8)	0 (0)	28 (26.2)	20 (18.7)	8 (7.5)
Sulfamethoxazol/ trimethoprim	23 (21.5)	7 (6.5)	16 (15.0)	0 (0)	0 (0)	0 (0)	84 (78.5)	73 (68.2)	11 (10.3)
Colistin	104 ^a (99.0)	78 (74.3)	26 (24.7)	0 (0)	0 (0)	0 (0)	1 (1.0)	1 (1.0)	0 (0)
Ciprofloxacin	39 (36.4)	21 (19.6)	18 (16.8)	4 (3.8)	4 (3.8)	0 (0)	64 (59.8)	55 (51.4)	9 (8.4)
Amikacin	83 (77.6)	60 (56.1)	23 (21.5)	15 (14.0)	14 (13.1)	1 (0.9)	9 (8.4)	6 (5.6)	3 (2.8)
Gentamicin	43 (40.2)	19 (17.8)	24 (22.4)	3 (2.8)	3 (2.8)	0 (0)	61 (57.0)	58 (54.2)	3 (2.8)

ESBL: extended-spectrum beta-lactamases.

^a The *Providencia rettgeri* and the *Serratia marcescens* isolates were excluded because of their natural resistance to colistin.

The percentages are rounded so as to add up to 100%.

Phylogenetic groups of the *Escherichia coli* isolates

More than half of *E. coli* isolates belonged to the phylogenetic group D (14 of the 24 *E. coli* isolates), seven *E. coli* isolates belonged to the phylogenetic group A, two belonged to the phylogenetic group B2, and one isolate belonged to the phylogenetic group B1 (Table 1).

Mulilocus sequence typing

The distribution of the sequence types among the *K. pneumoniae* and *E. coli* isolates is shown in Figures 1 and 2, respectively. ST101 was the most commonly observed ST for the *K. pneumoniae* isolates, accounting for 17 out of 67 isolates (25.4%), followed by ST395 and ST15 (7 isolates, 10.5%) (Figure 1). Six isolates (9%) belonged to ST147 (9%) and the other isolates to diverse STs, namely ST14 (n=4), ST45 (n=4), ST25 (n=2), ST392 (n=2), and one to other STs (Figure 1). Among the 24 OXA-48-positive *E. coli* isolates, seven belonged to ST38 (29.2%). The remaining 17 isolates belonged to STs 10, 617, 648 and 2969 (two isolates each) and to STs 46, 69, 95, 101, 362, 410, 746, 963 and 1092 (one isolate each) (Figure 2).

Since no MLST system has been developed for typing the *E. cloacae* species, these isolates were genotyped using the DiversiLab method. *E. cloacae* 7 and 9 recovered from Morocco were closely related, and *E. cloacae* 1, 2 and 5 (also from Morocco) belonged to the same cluster. The other *E. cloacae* isolates were distinct (data not shown).

Genetic location the *bla*_{OXA-48} gene

Using the specific primers designed from the reference plasmid pOXA-48a of *K. pneumoniae* 11978 [27] to amplify its replicase gene, 99 of the 107 isolates (92.5%) carried an IncL/M-pOXA-48a-like backbone. For the eight other isolates (*P. rettgeri* isolate no. 15, *E. coli* isolates no. 19 to 24, and *E. coli* isolate no. 37), mating-out assays were performed and transconjugants harbouring the *bla*_{OXA-48} gene were obtained for *P. rettgeri* isolate no. 15 and *E. coli* isolate no. 37. Plasmid DNA analysis of the two *E. coli* transconjugants revealed a single plasmid. Those two *bla*_{OXA-48}-positive plasmids corresponded to a ca. 150 kb IncA/C-type plasmid identified from *P. rettgeri* isolate no. 15 from Turkey and a ca. 160-kb IncF-type plasmid from an *E. coli* isolate from France. Despite repeated attempts, transconjugants or transformants were not obtained for six of the seven *E. coli* isolates belonging to ST38. Interestingly, I-Ceul analysis confirmed the chromosomal location of the *bla*_{OXA-48} gene in those six isolates (data not shown). Furthermore, one out of the seven *bla*_{OXA-48}-positive ST38 *E. coli* harboured the epidemic OXA-48 IncL/M-type plasmid.

Genetic environment of the *bla*_{OXA-48} gene

The *bla*_{OXA-48} gene was flanked by two copies of IS1999. In 21 isolates (19.6%), the upstream copy remained intact. This structure corresponded to transposon Tn1999, whereas 84 isolates (78.5%) had a Tn1999.2 transposon structure in which the IS1999 is disrupted by insertion of an IS1R element [4]. In two isolates

FIGURE 1

Sequence types represented among OXA-48-producing *Klebsiella pneumoniae* isolates, 2001–11 (n=67)

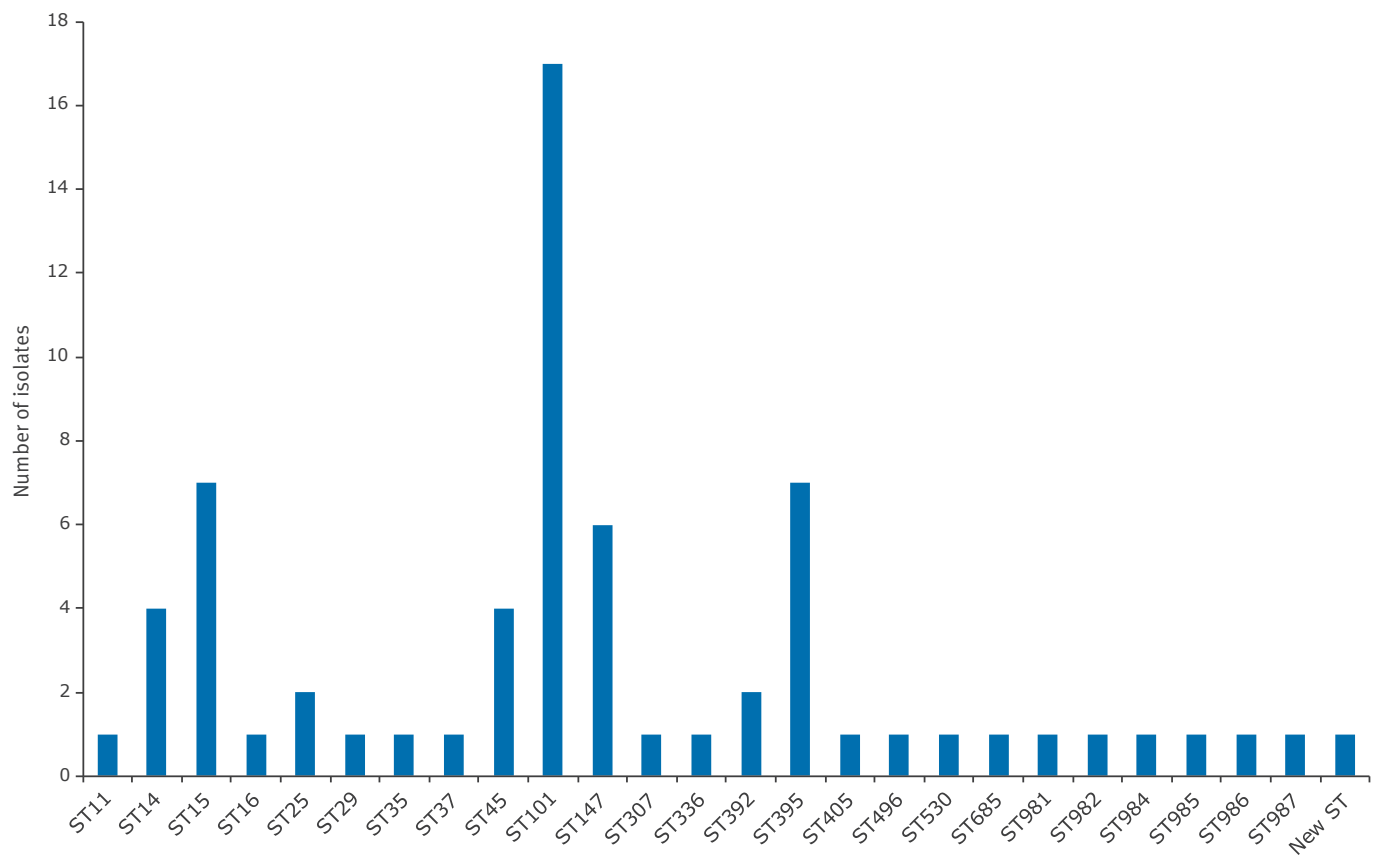
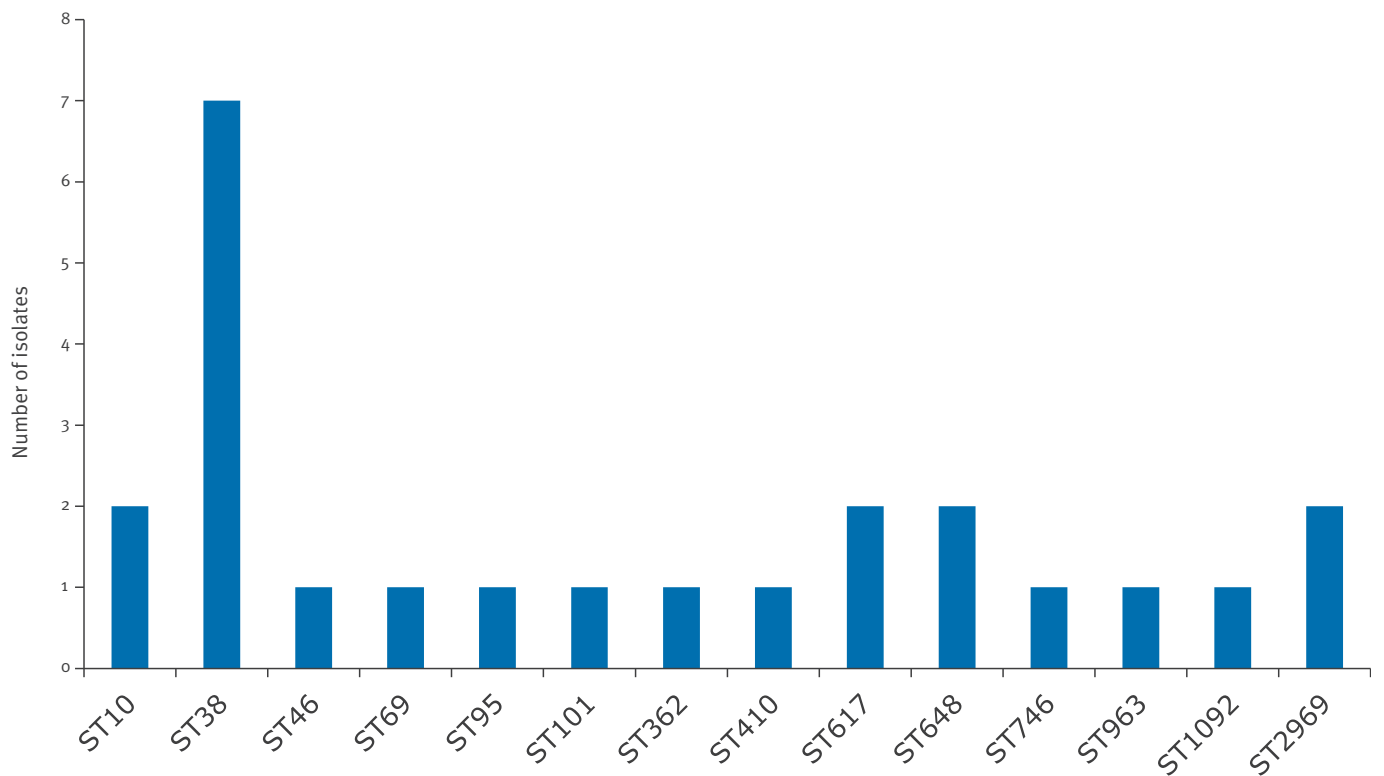


FIGURE 2

Sequence types represented among OXA-48-producing *Escherichia coli* isolates, 2001–11 (n=24)



(*E. cloacae* isolate no. 4 and *E. coli* isolate no. 17, recovered from the same patient), a new Tn₁₉₉₉ derivative was identified. This new transposon Tn_{1999.4} was composed of Tn_{1999.2} disrupted by another transposon, Tn₂₀₁₅ which, in turn, was composed of ISEcp1, *bla*_{CTX-M-15} and a truncated Tn2 transposase [32].

Discussion

We have analysed here many different features of 107 known OXA-48-positive enterobacterial isolates which are widely distributed at least in several European and North African countries, and also in Turkey. Noticeably, 25% of the OXA-48 beta-lactamase producers remained susceptible to broad-spectrum cephalosporins, which therefore present possible therapeutic options. At least positive therapeutic outcomes have been obtained using an animal model of infection and broad-spectrum cephalosporins [33]. Those ESBL-negative isolates were most often susceptible to the other classes of antibiotics, which is in line with the fact that the epidemic plasmid encoding the *bla*_{OXA-48} gene does not carry additional resistance determinants [28].

However, 75% of the OXA-48-positive isolates in our study harboured an additional ESBL-encoding gene that confers resistance to broad-spectrum cephalosporins. We have recently reported the genetic association of the *bla*_{CTX-M-15} and *bla*_{OXA-48} genes on the same transposon, indicating that this combination of multidrug-resistance genes may spread further in the future [32]. In addition, most of those ESBL-producing isolates were resistant to non-beta-lactam antibiotics, due to other resistance mechanisms. It is worth mentioning that 70 isolates (65%) were susceptible to imipenem and meropenem according to CLSI guidelines, further complicating the detection of OXA-48-producing isolates in laboratories. Conversely, most isolates showed intermediate susceptibility or resistance to ertapenem. Ertapenem may thus be the most appropriate carbapenem molecule for detecting OXA-48 producers. Therefore, a selective medium containing ertapenem has recently been developed for the detection of all types of carbapenemase producers including the OXA-48 beta-lactamase producers [34]. Taking into account the fact that 75% of the OXA-48 isolates were ESBL producers and the level of resistance to non-beta-lactam molecules, treatment options for infections caused by OXA-48 beta-lactamase producers may be limited. The efficacy of carbapenems in treating infections due to OXA-48 beta-lactamase producers with susceptibility or low-level resistance to several carbapenems remains debatable, because carbapenems have been shown to be an inefficient therapy for treating mice with induced peritonitis caused by an OXA-48-producing *K. pneumoniae* [33]. Also, imipenem-containing therapy failed to treat several OXA-48 infections in humans [4,11]. A single report described imipenem as efficient treatment against bacteraemia due to an OXA-48 *K. pneumoniae* isolate [35]. Controlled trials are needed to evaluate

the real clinical efficacy of carbapenems in treating infections due to OXA-48 beta-lactamase producers.

The clonal distribution of OXA-48 beta-lactamase-positive isolates is interesting because a quarter of the *K. pneumoniae* isolates belonged to ST101. OXA-48-positive *K. pneumoniae* isolates belonging to ST101 have recently been implicated in an outbreak in Spain, and have also been detected in Tunisia [11,12]. We report here that the ST101 isolates were recovered from Tunisia, Morocco, and from South Africa and France from patients who did not travel abroad, suggesting that this ST has now widely spread in European countries and in Africa. Seven *K. pneumoniae* isolates belonged to ST395, a ST implicated in clonal outbreaks in Europe [11,14]. Interestingly, we detected seven ST15 among *K. pneumoniae* isolates recovered from patients who had a link with Morocco. That sequence type corresponds to an internationally occurring clone and has been associated with different ESBL genes, but also with the metallo-beta-lactamase genes coding for NDM and VIM [36,37]. The occurrence of OXA-48 beta-lactamase in a ST15 *K. pneumoniae* isolate had been reported only once, in 2012, in an isolate from Finland [38]. Those data are likely to indicate that a novel OXA-48 *K. pneumoniae* clone belonging to ST15 may emerge in Morocco. *K. pneumoniae* isolates belonging either to ST392 or ST147 (differing at a single locus) were identified in a total of eight isolates, with the two ST392 collected in Morocco and the six ST147 collected in Belgium, Turkey and France, and also from patients originating from Tunisia or Libya. This result highlights the dissemination of another OXA-48-producing clone, mainly in the Mediterranean area. The other *K. pneumoniae* isolates belonged to diverse ST, supporting the hypothesis of the widespread dissemination of a single *bla*_{OXA-48}-positive IncL/M plasmid among various genetic backgrounds. Overall, there is no association between ST type and ESBL type among OXA-48 producers.

Among the 24 *E. coli* isolates, seven were of ST38, showing that this clone is widely disseminated, as previously suggested [7,39]. Interestingly, the *bla*_{OXA-48} gene was chromosomally located in six of those isolates, as was speculated for the ST38 *E. coli* isolates recovered in the United Kingdom [7]. Such chromosomal location of the *bla*_{OXA-48} gene in *E. coli* may be associated to a lower level of resistance (a single gene copy). The other 17 *E. coli* isolates were genetically distinct. Furthermore, it is interesting to note that 16 of the 24 OXA-48-positive *E. coli* belonged to phylogenetic group D or B2, which mainly include virulent strains. The *E. cloacae* isolates were overall clonally diverse.

As suggested previously, the *bla*_{OXA-48} gene was located on a 62 kb IncL/M plasmid in most of our isolates (n=99, 92.5%), indicating that current spread of OXA-48 beta-lactamase producers is mainly related to the diffusion of this plasmid. The dissemination of the *bla*_{OXA-48} gene is also associated with the spread of different clones.

Interestingly, 20% of OXA-48-producing isolates collected in France were considered to be autochthonous, indicating that the *bla*_{OXA-48} gene has already spread in the community in France. This latter result indicates ongoing diffusion of OXA-48-type genes in Europe.

Acknowledgements

This work was partially funded by a grant from the INSERM (U914) and the Université Paris XI, France. We thank platform Genotyping of Pathogens and Public Health (Institut Pasteur, Paris, France) for coding MLST alleles and profiles and making them available at www.pasteur.fr/mlst.

References

- Nordmann P, Naas T, Poirel L. Global spread of carbapenemase-producing *Enterobacteriaceae*. *Emerg Infect Dis*. 2011;17(10):1791-8. <http://dx.doi.org/10.3201/eid1710.110655> PMID:22000347 PMCID:PMC3310682
- Poirel L, Potron A, Nordmann P. OXA-48-like carbapenemase: the phantom menace. *J Antimicrob Chemother*. 2012;67(7):1597-606. <http://dx.doi.org/10.1093/jac/dks121> PMID:22499996
- Poirel L, Héritier C, Tolün V, Nordmann P. Emergence of oxacillinase-mediated resistance to imipenem in *Klebsiella pneumoniae*. *Antimicrob Agents Chemother*. 2004;48(1):15-22. <http://dx.doi.org/10.1128/AAC.48.1.15-22.2004> PMID:14693513 PMCID:PMC310167
- Carrère A, Poirel L, Yilmaz M, Akan OA, Feriha C, Cuzon G, et al. Spread of OXA-48-encoding plasmid in Turkey and beyond. *Antimicrob Agents Chemother*. 2010;54(3):1369-73. <http://dx.doi.org/10.1128/AAC.01312-09> PMID:20086157 PMCID:PMC2825965
- Carrère A, Poirel L, Eraksoy H, Gagatay AA, Badur S, Nordmann P. Spread of OXA-48-positive carbapenem-resistant *Klebsiella pneumoniae* isolates in Istanbul, Turkey. *Antimicrob Agents Chemother*. 2008;52(8):2950-4. <http://dx.doi.org/10.1128/AAC.01672-07> PMID:18519712 PMCID:PMC2493117
- Adler A, Shklyar M, Schwaber MJ, Navon-Venezia S, Dhaher Y, Edgar R, et al. Introduction of OXA-48-producing *Enterobacteriaceae* to Israeli hospitals by medical tourism. *J Antimicrob Chemother*. 2011;66(12):2763-6. <http://dx.doi.org/10.1093/jac/dkr382> PMID:22191089
- Dimou V, Dhanji H, Pike R, Livermore DM, Woodford N. Characterization of *Enterobacteriaceae* producing OXA-48-like carbapenemases in the UK. *J Antimicrob Chemother*. 2012;67(7):1660-5. <http://dx.doi.org/10.1093/jac/dks124> PMID:22532467
- Glupczynski Y, Huang T, Bouchahrouf W, Rezende de Castro R, Bauraing C, Gérard M, et al. Rapid emergence and spread of OXA-48-producing carbapenem-resistant *Enterobacteriaceae* isolates in Belgian isolates. *Int J Antimicrob Agents*. 2012;39(2):168-72. <http://dx.doi.org/10.1016/j.ijantimicag.2011.10.005> PMID:22115539
- Pfeifer Y, Schlatterer K, Engelmann E, Schiller RA, Frangenberg HR, Stiewe D, et al. Emergence of OXA-48-type carbapenemase-producing *Enterobacteriaceae* in German hospitals. *Antimicrob Agents Chemother*. 2012;56(4):2125-8. <http://dx.doi.org/10.1128/AAC.05315-11> PMID:22290940 PMCID:PMC3318349
- Poirel L, Carbonnelle E, Bernabeu S, Gutmann L, Rotimi V, Nordmann P. Importation of OXA-48-producing *Klebsiella pneumoniae* from Kuwait. *J Antimicrob Chemother*. 2012;67(8):2051-2. <http://dx.doi.org/10.1093/jac/dks167> PMID:22577102
- Cuzon G, Ouanich J, Gondret R, Naas T, Nordmann P. Outbreak of OXA-48-positive carbapenem-resistant *Klebsiella pneumoniae* isolates in France. *Antimicrob Agents Chemother*. 2011;55(5):2420-3. <http://dx.doi.org/10.1128/AAC.01452-10> PMID:21343451 PMCID:PMC3088266
- Pitart C, Solé M, Roca I, Fábrega A, Vila J, Marco F. First outbreak of a plasmid-mediated carbapenem-hydrolyzing OXA-48 β -lactamase in *Klebsiella pneumoniae* in Spain. *Antimicrob Agents Chemother*. 2011;55(9):4398-401. <http://dx.doi.org/10.1128/AAC.00329-11> PMID:21746954 PMCID:PMC3165339
- Voulgari E, Zarkotou O, Ranellou K, Karageorgopoulos DE, Vrioni G, Mamali V, et al. Outbreak of OXA-48 carbapenemase-producing *Klebsiella pneumoniae* in Greece involving an ST11 clone. *J Antimicrob Chemother*. 2013;68(1):84-8. <http://dx.doi.org/10.1093/jac/dks356> PMID:22945916
- Potron A, Kalpoe J, Poirel L, Nordmann P. European dissemination of a single OXA-48-producing *Klebsiella pneumoniae* clone. *Clin Microb Infect*. 2011;17(12):E24-6. <http://dx.doi.org/10.1111/j.1469-0691.2011.03669.x> PMID:21973185
- Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing; twenty-second informational supplement. CLSI document M100-S22. Wayne, PA: CLSI, 2012. Available from: <http://antimicrobianos.com.ar/ATB/wp-content/uploads/2012/11/M100S22E.pdf>
- European Committee on Antimicrobial Susceptibility Testing (EUCAST). Breakpoint tables for interpretation of MICs and zone diameters, Version 2.0. EUCAST; 2012. Available from: http://www.eucast.org/fileadmin/src/media/PDFs/EUCAST_files/Disk_test_documents/EUCAST_breakpoints_v_2.0_120101.pdf
- Drieux L, Brossier F, Sougakoff W, Jarlier V. Phenotypic detection of extended-spectrum β -lactamase production in *Enterobacteriaceae*: review and bench guide. *Clin Microbiol Infect*. 2008;14(1):90-103. <http://dx.doi.org/10.1111/j.1469-0691.2007.01846.x> PMID:18154532
- Nordmann P, Dortet L, Poirel L. Rapid detection of extended-spectrum- β -lactamase-producing *Enterobacteriaceae*. *J Clin Microbiol*. 2012;50(9):3016-22. <http://dx.doi.org/10.1128/JCM.00859-12> PMID:22760052 PMCID:PMC3421789
- Poirel L, Dortet L, Bernabeu S, Nordmann P. Genetic features of bla_{NDM-1}-positive *Enterobacteriaceae*. *Antimicrob Agents Chemother*. 2011;55(11):5403-7. <http://dx.doi.org/10.1128/AAC.00585-11> PMID:21859933 PMCID:PMC3195013
- Poirel L, Walsh TR, Cuvillier V, Nordmann P. Multiplex PCR for acquired carbapenemase genes. *Diagn Microbiol Infect Dis*. 2011;70(1):119-23. <http://dx.doi.org/10.1016/j.diagmicrobio.2010.12.002> PMID:21398074
- Potron A, Nordmann P, Lafeuille E, Al Maskari Z, Al Rashdi F, Poirel L. Characterization of OXA-181, a carbapenem-hydrolyzing class D β -lactamase from *Klebsiella pneumoniae*. *Antimicrob Agents Chemother*. 2011;55(10):4896-9. <http://dx.doi.org/10.1128/AAC.00481-11> PMID:21768505 PMCID:PMC3186949
- Poirel L, Guibert M, Girlich D, Naas T, Nordmann P. Cloning, sequence analyses, expression, and distribution of amp^C-amp^R from *Morganella morganii* clinical isolates. *Antimicrob Agents Chemother*. 1999;43(4):769-76. PMID:10103179 PMCID:PMC89205
- Diancourt L, Passet V, Verhoef J, Grimont PA, Brisse S. Multilocus sequence typing of *Klebsiella pneumoniae* nosocomial isolates. *J Clin Microbiol*. 2005;43(8):4178-82. <http://dx.doi.org/10.1128/JCM.43.8.4178-4182.2005> PMID:16081970 PMCID:PMC1233940
- Clermont O, Bonacorsi S, Bingen E. Rapid and simple determination of the *Escherichia coli* phylogenetic group. *Appl Environ Microbiol*. 2000;66(10):4555-8. <http://dx.doi.org/10.1128/AEM.66.10.4555-4558.2000> PMID:11010916 PMCID:PMC92342
- Kieser T. Factors affecting the isolation of CCC DNA from *Streptomyces lividans* and *Escherichia coli*. *Plasmid*. 1984;12(1):19-36. [http://dx.doi.org/10.1016/0147-619X\(84\)90063-5](http://dx.doi.org/10.1016/0147-619X(84)90063-5)
- Liu SL, Hessel A, Sanderson KE. Genomic mapping with I-Ceul, an intron-encoded endonuclease specific for genes for ribosomal RNA, in *Salmonella* spp., *Escherichia coli*, and other bacteria. *Proc Natl Acad Sci USA*. 1993;90(14):6874-8. <http://dx.doi.org/10.1073/pnas.90.14.6874> PMID:8347173 PMCID:PMC47035
- Carattoli A, Bertini A, Villa L, Falbo V, Hopkins KL, Threlfall EJ. Identification of plasmids by PCR-based replicon typing. *J Microbiol Methods*. 2005;63(3):219-28.
- Poirel L, Bonnin RA, Nordmann P. Genetic features of the widespread plasmid coding for the carbapenemase OXA-48. *Antimicrob Agents Chemother*. 2012;56(1):559-62. <http://dx.doi.org/10.1128/AAC.05289-11> PMID:22083465 PMCID:PMC3256075
- Aubert D, Naas T, Héritier C, Poirel L, Nordmann P. Functional characterization of IS₁₉₉₉, an IS₄ family element involved in mobilization and expression of β -lactam resistance genes. *J Bacteriol*. 2006;188(18):6506-14. <http://dx.doi.org/10.1128/JB.00375-06> PMID:16952941 PMCID:PMC1595497
- Jacoby G. β -Lactamase Classification and Amino Acid Sequences for TEM, SHV and OXA Extended-Spectrum

- and Inhibitor Resistant Enzymes. Burlington: Lahey Clinic. [Accessed Jul 2013]. Available from: www.lahey.org/studies/
31. Hidalgo L, Hopkins KL, Wareham DW, Gutierrez B, Gonzalez-Zorn B. Association of extended-spectrum β -lactamase VEB-5 and 16S rRNA methyltransferase ArmA in *Salmonella enterica* from the United Kingdom. *Antimicrob Agents Chemother.* 2012;56(9):4985-7. <http://dx.doi.org/10.1128/AAC.00381-12> PMID:22710120 PMCID:PMC3421862
 32. Potron A, Nordmann P, Rondinaud E, Jaureguy F, Poirel L. A mosaic transposon encoding OXA-48 and CTX-M-15; towards the panresistance. *J Antimicrob Chemother.* 2013;68(2):476-7. <http://dx.doi.org/10.1093/jac/dks397> PMID:23027715
 33. Mimos O, Grégoire N, Poirel L, Marliat M, Couet W, Nordmann P. Broad-spectrum β -lactam antibiotics for treating experimental peritonitis in mice due to *Klebsiella pneumoniae* producing the carbapenemase OXA-48. *Antimicrob Agents Chemother.* 2012;56(5):2759-60. <http://dx.doi.org/10.1128/AAC.06069-11> PMID:22330912 PMCID:PMC3346608
 34. Nordmann P, Girlich D, Poirel L. Detection of carbapenemase producers in Enterobacteriaceae by use of a novel screening medium. *J Clin Microb.* 2012;50(8):2761-6. <http://dx.doi.org/10.1128/JCM.06477-11> PMID:22357501 PMCID:PMC3421537
 35. Maherault AC, Nordmann P, Therby A, Pangon B. Efficacy of imipenem for the treatment of bacteremia due to an OXA-48-producing *Klebsiella pneumoniae* isolate. *Clin Infect Dis.* 2011;54(4):577-8. <http://dx.doi.org/10.1093/cid/cir887> PMID:22157173
 36. Poirel L, Benouda A, Hays C, Nordmann P. Emergence of NDM-1-producing *Klebsiella pneumoniae* in Morocco. *J Antimicrob Chemother.* 2011;66(12):2781-3. <http://dx.doi.org/10.1093/jac/dkr384> PMID:21930570
 37. Sanchez-Romero I, Asensio A, Oteo J, Munoz-Algarra M, Isidoro B, Vindel A, et al. Nosocomial outbreak of VIM-1-producing *Klebsiella pneumoniae* isolates of multilocus sequence type 15: molecular basis, clinical risk factors, and outcome. *Antimicrob Agents Chemother.* 2012;56(1):420-7. <http://dx.doi.org/10.1128/AAC.05036-11> PMID:22005997 PMCID:PMC3256054
 38. Osterblad M, Kirveskari J, Hakanen AJ, Tissari P, Vaara M, Jalava J. Carbapenemase-producing Enterobacteriaceae in Finland: the first years (2008-11). *J Antimicrob Chemother.* 2012;67(12):2860-4. <http://dx.doi.org/10.1093/jac/dks299> PMID:22855858
 39. Poirel L, Bernabeu S, Fortineau N, Podglajen I, Lawrence C, Nordmann P. Emergence of OXA-48-producing *Escherichia coli* clone ST38 in France. *Antimicrob Agents Chemother.* 2011;55(10):4937-8. <http://dx.doi.org/10.1128/AAC.00413-11> PMID:21768512 PMCID:PMC3186974