

Open access • Journal Article • DOI:10.1007/S00415-017-8474-3

Intermediate Charcot–Marie–Tooth disease: an electrophysiological reappraisal and systematic review — Source link

José Berciano, Antonio G. García, Elena Gallardo, K. Peeters ...+6 more authors Institutions: University of Cantabria, University of Antwerp Published on: 31 Mar 2017 - Journal of Neurology (Springer Berlin Heidelberg)

Related papers:

- Clinical implications of genetic advances in Charcot–Marie–Tooth disease
- · Charcot-Marie-Tooth Disease Subtypes and Genetic Testing Strategies
- Epidemiologic Study of Charcot-Marie-Tooth Disease: A Systematic Review
- Charcot–Marie–Tooth disease: frequency of genetic subtypes and guidelines for genetic testing
- CMT subtypes and disease burden in patients enrolled in the Inherited Neuropathies Consortium natural history study: a cross-sectional analysis





This item is the archived peer-reviewed author-version of:

Intermediate Charcot-Marie-Tooth disease : an electrophysiological reappraisal and systematic review

Reference:

Berciano José, García Antonio, Gallardo Elena, Peeters Kristien, Pelayo-Negro Ana L., Alvarez-Paradelo Silvia, Gazulla José, Martínez-Tames Miriam, Infante Jon, Jordanova Albena.- Intermediate Charcot-Marie-Tooth disease : an electrophysiological reappraisal and systematic review Journal of neurology - ISSN 0340-5354 - Heidelberg, Springer heidelberg, 264:8(2017), p. 1655-1677 Full text (Publisher's DOI): https://doi.org/10.1007/S00415-017-8474-3 To cite this reference: http://hdl.handle.net/10067/1421910151162165141

uantwerpen.be

Institutional repository IRUA

-1-

Intermediate Charcot-Marie-Tooth disease: an electrophysiological reappraisal and systematic review

José Berciano^{1*}, Antonio García^{2*}, Elena Gallardo³, Kristien Peeters⁴, Ana L. Pelayo-Negro¹, Silvia Álvarez-Paradelo², José Gazulla⁵, Miriam Martínez-Tames⁶, Jon Infante¹ and Albena Jordanova⁴

¹Service of Neurology, Hospital Universitario Marqués de Valdecilla, Instituto de Investigación Marqués de Valdecilla (IDIVAL), Universidad de Cantabria (UC), and Centro de Investigación Biomédica en Red de Enfermedades Neurodegenerativas (CIBERNED), Santander, Spain

²Service of Clinical Neurophysiology, Hospital Universitario Marqués de Valdecilla, Instituto de Investigación Marqués de Valdecilla (IDIVAL), Universidad de Cantabria (UC), and Centro de Investigación Biomédica en Red de Enfermedades Neurodegenerativas (CIBERNED), Santander, Spain

³Service of Radiology, Hospital Universitario Marqués de Valdecilla, Instituto de Investigación Marqués de Valdecilla (IDIVAL), Universidad de Cantabria (UC), and Centro de Investigación Biomédica en Red de Enfermedades Neurodegenerativas (CIBERNED), Santander, Spain

⁴VIB Center for Molecular Neurology, University of Antwerp, Antwerp, Belgium

⁵Service of Neurology, Hospital Universitario Miguel Servet, Zaragoza, Spain

⁶Medical Student, Universidad de Cantabria (UC), Spain

*These authors contributed equally to this work

For: Journal of Neurology (Systematic review)

Article content: Text, 7,900 words; Abstract, 301 words; 156 References; 5 Figures; and 2 Tables

Correspondence to: Dr José Berciano, Professor emeritus, Department of Medicine and Psychiatry, "Edificio Escuela Universitaria de Enfermería (cuarta planta)", Avda. de Valdecilla s/n, University of Cantabria, 39008 Santander, Spain; E-mail: jaberciano@humv.es

Abstract

Charcot-Marie-Tooth disease (CMT) is the most frequent form of inherited neuropathy with great variety of phenotypes, inheritance patterns and causative genes. According to median motor nerve conduction velocity (MNCV), CMT is divided into demyelinating (CMT1) with MNCV below 38 m/s, axonal (CMT2) with MNCV above 38 m/s, and intermediate CMT with MNCV between 25 and 45 m/s. In each category, transmission may be autosomal dominant, autosomal recessive or X-linked. The nosology of intermediate CMT is controversial because of concerns about electrophysiological delimitation. A systematic computer-based literature search was conducted on PubMed, using the following MeSH: i/ intermediate Charcot-Marie-Tooth; ii/ X-linked intermediate Charcot-Marie-Tooth; and iii/ X-linked Charcot-Marie-Tooth and electrophysiology. We retrieved 225 articles reporting X-linked CMT or intermediate CMT with electrophysiological information. After eligibility, 156 papers were used for this review. In assessing median MNCV, compound muscle action potential (CMAP) amplitudes were taken into account. In cases with attenuated CMAP and wherever possible, proximal median MNCV was used for accurate definition of conduction slowing in the intermediate range. In the vast majority of males with X-linked CMT associated to GJB1 mutation (CMTX1), median MNCV was intermediate. CMT associated with DRP2 mutation is another well-documented X-linked intermediate disorder. Autosomal dominant intermediate CMT (DI-CMT) encompasses 11 different types, six of them with assigned phenotype MIM number, the remaining five being unnumbered. Based on available electrophysiological information, we wonder if DI-CMTA should be reclassified within CMT2. Autosomal recessive intermediate CMT (RI-CMT) covers four numbered MIM phenotypes though, in accordance with reported electrophysiology, two of them (RI-CMTB and RI-CMTD) should probably be

reclassified within AR-CMT2. We conclude that intermediate CMT is a complex inherited syndrome, whose characterization requires a specific electrophysiological protocol comprising evaluation of upper-limb proximal nerve trunks when distal CMAP amplitudes are reduced, and that an updated version of MIM phenotype numbering is needed.

Keywords: Autosomal dominant inheritance. Autosomal recessive inheritance. Axonal degeneration. Charcot-Marie-Tooth disease. Compound motor action potential. Demyelination. Electrophysiology. Intermediate Charcot-Marie-Tooth disease. Motor conduction velocity. Nerve biopsy. OMIM. PRISMA statement. Proximal motor nerve conduction velocity. Systematic review. X-linked inheritance.

Introduction

Charcot-Marie-Tooth disease (CMT) is the most frequent form of sensorimotor inherited neuropathy with a prevalence ratio of 28 cases/100,000 inhabitants [1]. CMT was initially classified according to the mode of transmission (autosomal dominant, autosomal recessive or X-linked), and electrophysiological or nerve biopsy features [2]. Characteristically, median motor nerve conduction velocity (MNCV) is below 38 m/s for the demyelinating form (CMT1) and above 38 m/s for the axonal form (CMT2) [3]. For want of a better name, Davis and colleagues introduced the term intermediate for designating dominant CMT (DI-CMT) patients with clinico-electrophysiological and pathological features not fitting into either CMT1 or CMT2 [4]. Their intermediate series comprised 49 affected individuals (31 male and 18 females) coming from seven kinships. It is worth noting that in four of their kinships "males tended to be more affected than females. The degree of sparing of females was illustrated by the presence of asymptomatic affected women in the seventh decade in kinship 22". Although these features pointed to an X-linked disorder, the authors did not consider this possibility in their paper. In short, DI-CMT was originally characterized by the following features: i/ absence of clinically observed nerve hypertrophy; ii/ median MNCV between 25 and 45 m/s (according to their table 1a, mean and standard deviation of 34.6 ± 6.3); iii/ prolonged distal motor latencies (DML) (mean, 5.0 ± 1.3 ms); iv/ preserved mean compound muscle action potential (CMAP) amplitude (mean, 4.6 mV; neither range nor standard deviation is given); and iv/ nerve biopsy showing axonal changes, clusters of regenerating myelinated fibres, loss of larger fibres noted from unimodal diameter histograms, and onion bulbs with fewer lamellae than in CMT1 [5].

The distinction of an intermediate CMT subtype was initially a controversial issue since Buchthal and Behse, in their seminal CMT study comprising 77 patients, did

-4-

not find any evidence of an intermediate type [6]. In fact, they reported that motor and sensory conduction velocities confirmed the existence of the two main types of CMT: hypertrophic with conduction velocity along the sural nerve (also superficial peroneal and median nerves) diminished to less than 60% of normal, and neuronal with conduction velocity preserved or slowed but never to less than 60% of normal.

In a series of 131 CMT patients, Bouché and colleagues found two main groups: 55 patients whose median MNCVs were below 30 m/s (mean, 19.47±5.46), and 64 whose corresponding MNCVs were above 40 m/s (mean, 50.72 ± 5.80) [7]. It is worth noting that CMAP amplitude values are not given. There remained a third group, comprising 12 patients (6 dominant and 6 sporadic), that could not be classified since median MNCVs ranged between 30 and 40 m/s. Based upon these electrophysiological features and nerve biopsy findings [8], the authors suggested that this group might be included within intermediate CMT. Since then, the electrophysiological notion of intermediate CMT has widely been applied for patients showing either median MNCV between 25 and 45 m/s or between 30 and 40 m/s. More recently and starting from a comprehensive investigation of genetic testing strategies in CMT, Saportaand colleagues restricted the intermediate range of upper-limb MNCV from >35 to \leq 45 m/s [9]. Furthermore, these authors established for ulnar nerve that if the CMAP is >0.5mV, a conduction velocity of <38 m/s is considered demyelinating, and 38-45 m/s is intermediate [10]. As stated by Nicholson and Myers selection of nerve conduction values in different affected individuals in a family is challenging, because slow nerve conduction values can be found in axonal neuropathies as a result of the loss of large rapidly conduction fibres [11].

It is timely to remember the two basic patterns of motor conduction electrophysiological alteration occurring in axonal loss lesions [12]: i/ at one extreme, there may be severe axonal loss with sparing of only a few of the fastest fibres remaining, which leads to CMAP amplitude reduction with relative preservation of MNCV and DML; and ii/ at the other extreme, as usually occurs in axonal CMT, if all axons are lost, except for a few of the normal slowly conducting fibres, the amplitude also falls dramatically, but conduction velocity can only drop as low as 35 m/s (=75%) of the lower limit of normal [LLN]), and DML also prolong to 130% of the upper limit of normal. In axonal polyneuropathies, CMAP amplitude reduction is linearly correlated with decrease in median and peroneal MNCV, though in the long-term such nerve conduction alterations may be aggravated by the appearance of slow-regenerating fibres, and axonal atrophy with or without secondary de- or re-myelination [13]. However, the pattern and degree of axonal loss cannot be entirely determined from routine electrodiagnostic or strength testing due to collateral reinnervation, this calling for special techniques such as motor unit number estimation [14]. The common pathological hallmark of axonal CMT is length-dependent loss of large myelinated fibres with variable signs of regeneration and axonal atrophy[15-17] Applying a conversion factor of 6 [18] to sural nerve ultrathin images from a severe axonal CMT associated to homozygous E210X in the NEFL gene, we have estimated that conduction velocity of this nerve would be 18 m/s, namely, comparable to those found in median nerve, 22 m/s (CMAP, 1.2 mV), and ulnar nerve, 17 m/s (CMAP, 0.1 mV) [19, 20]. This conduction estimation could be even lower using a conversion factor of 4.3 reported in normal sural nerve [21].

At this point, we would like to emphasize that in any axonal CMT with severe amplitude reduction of distal CMAPs in upper-limb nerves, the corresponding MNCVs may fall down to the intermediate range or even below 25 m/s. In the seminal CMT2 series by Bienfaitand colleagues, comprising 61 patients from 18 kinships, median MNCV was less than 38 m/s in nine patients from six families, all of them exhibiting severely reduced distal CMAPs (<1 mV) [22, 23]. Wisely the authors wrote that "nerve conduction velocities may be reduced because of loss of large-diameter fibres or secondary demyelination or both... Some authors adhere strictly to intermediate MCVs (between 20 and 45 m/s) in all family members, but others include families with wide ranges of MCV, making classification difficult... It remains a semantic discussion whether families with a wide range of MCVs should be included in a separate DI-CMT group or in CMT2". We addressed this question in six patients from two DI-CMT pedigrees associated with previously reported *NEFL* mutations [20, 24]. For the segment elbow-wrist, median MNCVs were systematically in the intermediate or even in the demyelinating range, both in patients with preserved CMAPs or attenuated distal CMAPs. Figure 1 illustrates that in case of severe distal CMAP attenuation with median MNCV in the demyelinating range, recording of proximal nerve segments (here axilla-elbow) might be a useful method for accurate detection of the intermediate conduction pattern.

CMT encompasses clinically and genetically heterogeneous syndromes, pathogenic mutations having been reported in over 80 genes with the incorporation of next-generation sequencing [25, 26]. Intermediate CMT has been associated with Xlinked gene mutations (mainly *GBJ1* mutations), and gene mutations with either autosomal dominant (DI-CMT) or autosomal recessive (RI-CMT) transmission [11, 26-28]. DI-CMT comprises six loci with five cloned genes (*DNM2*, *YARS*, *MPZ*, *INF2*, and *GNB4*), whereas mutations in four genes (*GDAP1*, *KARS*, *PLEKHG5* and *COX6A1*) have been associated with RI-CMT. Frequencies of intermediate CMT have ranged between 2.9% and 15.6% (median, 6.8%) in comparison with the total CMT rate, its prevalence in the Republic of Ireland being 0.61 cases/100,000 (95% IC, 0.35-0.87) [9, 29-34].

Given that the notion on intermediate CMT is to some degree controversial, an up-to-date overview of syndromes included under such designation is currently needed. In light to this, we aimed to systematically review the nosology of syndromes enclosed under the umbrella of intermediate CMT, taking into account reappraised motor nerve conduction parameters in upper-limb nerves.

Methods

Literature search strategy

A systematic computer-based literature search was conducted in October 27, 2016 on PubMed database. We used three Medical Subject Headings (MeSH) terms in title: i/ intermediate Charcot-Marie-Tooth; ii/ X-linked intermediate Charcot-Marie-Tooth; and iii/ X-linked Charcot-Marie-Tooth and electrophysiology. We selected papers published in English, French or Spanish languages. We also screened the database Online Mendelian Inheritance in Man (OMIM; <u>https://www.omim.org/</u>), Inherited Peripheral Neuropathies Mutation Database (<u>http://www.molgen.ua.ac.be/CMTMutations/</u>), and Neuromuscular Home Page (<u>http://neuromuscular.wustl.edu/time/hmsn.html#</u>). A manual search of relevant review papers was also performed. Our search methodology was done following the standard guidelines for systematic literature reviews outlined in the PRISMA statement [35].

Quality assessment

Given that intermediate CMT is an electrophysiological notion, we considered that reassessment of electrophysiological parameters in each publication is a crucial issue. Following the original description of intermediate CMT [4], we focused on median nerve motor conduction parameters, and alternatively on those of ulnar nerve, accepting that intermediate median MNCV fits in between 25 and 45 m/s with relative preservation of distal CMAP. For comparison purposes, control MNCV values were those used in our laboratory, reported elsewhere [20, 24, 36]. Starting from nerve biopsv features, it is a well-established histological fact that both in CMT2 and in intermediate CMT loss of myelinated fibres mainly involves the largest peak ($\geq 7\mu m$) [5, 6, 8, 16, 17]. With a cutoff of 7 μ m, in normal nerves 32% to 45% of myelinated fibres fall above, and 55% to 68% fall below this value [37]. If we assume a selective loss of large myelinated fibres, distal median nerve CMAP amplitude could fall into values between 2.2 and 2.7 mV (LLN, \geq 4). Accepting this corollary and applying conversion factors of 4.3 or 6 (see above), such large myelinated fibre loss could account for a decline of median MNCV between 26 and 36 m/s. Under such circumstances, when confronted with nerve conduction values below 38 m/s, characteristic of CMT1 [3], we should bear in mind that this could merely be accounted for by axonal loss. Therefore, in assessing median/ulnar MNCV in the forearm segment we invariably analyzed distal CMAP amplitude; in cases with CMAP reduction by half of the normal LLN or more, we also analyzed if proximal conduction values, axilla-elbow segment, were available (see figure 1). In short, in our view an accurate definition of intermediate CMT requires availability of nerve segments with normal or relatively preserved CMAP amplitudes.

The results of nerve biopsy in intermediate CMT, particularly in CMTX1 males, have been a matter of some controversy, given that they were initially interpreted as primary axonal pathology [38], demyelinating [39] or mixed pathology [40]. To clarify the issue we have paid particular attention to papers with detailed nerve histological

-9-

descriptions including fibre teasing or longitudinal thin sections, and fibre morphometry [2, 41].

Results

We retrieved 225 articles reporting X-linked CMT or intermediate CMT, although those containing electrophysiological information were selected for this review (see below). We will present our review results according to appearance order in table 1.

X-linked Charcot-Marie-Tooth disease (CMTX)

CMTX accounts for around 10% of all CMT forms, five phenotypes having been recognized [26]. By far, CMTX1 (MIM#302800) is the most frequent phenotype, which is associated with a plethora of point *GJB1* mutations, including a deletion of the entire coding sequence of the gene [42]. It has been established that most *GJB1* mutations cause a simple loss of function and that no particular mutation appeared more severe than deletion of the entire gene [43].

The CMTX1 phenotype is characterized by progressive muscle atrophy and weakness, areflexia and variable sensory abnormalities; central nervous system (CNS) involvement may also occur. Males have moderate to severe symptoms, whereas heterozygous females are usually less affected [42, 44, 45]. Although the CMTX1 electrophysiological pattern in males is currently accepted as a prototypic example of intermediate disorder, the issue is not without controversy. With heterogeneity between affected males and females, the results of nerve conduction studies differed in the three initial studies (table 2) [38, 39, 46, 47]. Phillips and colleagues found that ulnar MNCVs ranged between 33 and 42 m/s (mean, 38) in males, and between 44 and 56 m/s (mean, 49) in females; MNCV slowing was most prominent in individuals who had CMAP amplitude reduction [46]. They considered that the observed phenotype was similar to that of intermediate CMT and, despite not having performed nerve biopsy, wisely proposed that electrophysiology indicates a mixed axonal and demyelinating disorder. In the pedigree reported by Rozear and colleagues median MNCV ranged between 28 and 37 m/s (mean, 34) in males, and between 38 and 53 m/s (mean, 50) in affected females [39]. Neither CMAP values nor DML values are given. Sural nerve biopsy in a male patient showed axonal loss, variation of myelin thickness, onion bulbs, and, on fibre teasing, shortened internodal lengths and variation in myelin thickness. These electrophysiological and pathological data in male patients were interpreted as characteristic of a demyelinating hypertrophic neuropathy. In a large Canadian kindred comprising 13 male patients and 19 affected females examined, Hahn and colleagues recorded peroneal MNCVs distally to extensor digitorum brevis muscle and proximally to anterolateral leg muscle compartment [38]. Where CMAP amplitudes were preserved, MNCVs were either normal or slightly reduced (30 to 39 m/s). Nerve biopsies in two cases showed loss of myelinated and unmyelinated fibres, regenerative sprouting and secondary demyelination. The authors conclude that this CMTX variant is the result of primary axonal degeneration.

Given the complex nosology of CMTX1, it is timely to make a brief comment on animal models of the disease. Initial characterization of *GJB1*-null mice showed that demyelination preceded axonal loss [48]. Indeed, connexin32-null mice developed, by three months of age, demyelinating peripheral neuropathy with widened incisures and accumulations of adaxonal cytoplasm ("periaxonal collars"), though main electrophysiological findings were significant decrease of M responses and slight conduction slowing indicative of mixed axonal and demyelinating pathology [49, 50]. Re-investigation of the issue has demonstrated that axonal abnormalities, including impaired cytoskeletal organization and defects in axonal transport, preceded demyelination in this mouse model [51]. Prominent periaxonal collars have also been reported in nerve biopsies of CMTX1 patients [40, 52-55], but not in other inherited neuropathies [50]. The disorganization of the inner Schwann cell compartment might reflect poor communication between outer and inner cytoplasmic aspects of myelinating Schwann cells, resulting from altered gap junctions, which eventually leads to cytoplasmic abnormalities [49, 50]. Alternatively and accepting the relevant role of early axonal dysfunction, such periaxonal collars could represent a non-specific mechanism by which the Schwann cells clear debris and help maintaining the integrity of the axon under normal and pathologic conditions [56, 57]. Be that as it may, the corollary of these experimental studies is that both primary axonal dysfunction evolving into demyelination or primary demyelination evolving into axonal degeneration may occur in CMTX1.

Table 2 summarizes 40 papers describing motor conduction parameters in median nerve (alternatively ulnar nerve or peroneal nerve) reported between 1985 and 2016 [38-40, 43, 45, 46, 52, 53, 55, 58-88]. In the five largest CMTX1 series, mean median MNCV in males ranged from 33.2 to 36.0 m/s, mean median CMAP amplitudes varying between 2.0 and 3.7 mV [40, 45, 63, 69, 70]. Furthermore, the distribution of median MNCVs was unimodal peaking at 33 m/s [70]. In one series, 90% of CMTX1 males showed intermediate median MNCV (30-40 m/s), just one out of 21 cases exhibiting normal MNCV [63]; in another series, however, male median MNCV was normal in 20% (\geq 49 m/s), intermediate (33-41 m/s) in 40%, and slow (<32 m/s) in the remaining 20% [45]. In case report or family report papers, median MNCVs and CMAP amplitudes in males were almost always abnormal (\leq 48 m/s and \leq 3.9 mV); exceptions are as follows (see Table 2): i/ two affected boys, aged 3 and 6 years, were normal clinically and electrophysiologically thus indicating that CMTX1 phenotype is age dependent [38, 43, 63]; ii/ occasionally, male patients had normal MNCV with reduced CMAP amplitude, a feature giving support to the notion of a primary axonal involvement [38, 63]; and iii/ conversely, there were a few patients with intermediate median or ulnar MNCV and preserved CMAP amplitude, a binomial pointing to a demyelinating disorder [55, 67, 71, 73, 74]. A significant positive correlation was found between median, ulnar and peroneal MNCV slowing and CMAP amplitude reduction [63]. Where reported, DML and F-wave latencies were usually and harmonically delayed with the degree of MNCV slowing (see table 2). In no case exhibiting median or ulnar MNCV slowing with CMAP attenuation was proximal (axilla-elbow segment) MNCV evaluated.

Nerve biopsy in CMTX1 male patients has been reported in 17 papers (table 2) [38, 40, 47, 52, 53, 55, 63-65, 68, 70, 71, 74, 75, 80, 83, 85]. Leaving the initial controversy over pure demyelination versus pure axonal pathology into aside (see above), the common denominator of histological studies is a variable combination of demyelination and axonal degeneration, which can be summarized as follows: i/ on semithin sections, reduction of larger myelinated fibres and clusters of regeneration, with a shift of the fibre histogram to the smaller fibre range; ii/ variable presence of onion bulb formation; iii/ on fibre teasing and longitudinal semithin sections, paranodal demyelination, widening of the nodal gap, and to a lesser degree internodal de/remyelination; and iv/ on ultrastructural study, periaxonal collars (see above), thinly myelinated fibres, clusters of regeneration, and onion bulbs. The observed mixed pathology indicate that mutation and loss of function of connexin 32 gap junctions lead to profound alterations in the Schwann cell-axon unit with deleterious effects on both Schwann cells and myelin, as well as on the functions and integrity of axons [40]. In

short, these descriptions corroborate the former pathological hallmark reported in intermediate CMT [5, 8].

CMT associated with mutation in X-linked DRP2

In a unique pedigree, Brennan and colleagues described a 60-year-old man presenting with a 10-year history of length-dependent sensorimotor neuropathy [89]. A maternal grand-father had been diagnosed of peripheral neuropathy; his mother and two maternal aunts presented hammertoes. On three serial electrophysiological studies performed over a 4-year period, ulnar MNCVs ranged between 43 and 45 m/s, CMAP amplitudes being normal (13.4-8.2 mV); worthy of note is the fact that motor conduction study included two recordings from axilla that showed velocities of 36 and 35 m/s with preserved CMAP amplitudes. DML were normal or minimally prolonged, whereas Flatencies were delayed (37.6 ms in median nerve and 37.4 ms in ulnar nerve; normal, \leq 31 and \leq 32 ms, respectively). Whole exome sequencing revealed a stop codon mutation in DRP2, c805C>T (Q269*). Skin biopsies revealed a lack of DRP2 in myelinated nerves that exhibited no distinct Cajal bands and periaxonal collars (see above). DRP2 interacts with periaxin and dystroglican to form the periaxin-DRP2dystroglican complex, which plays a role in the maintenance of the Cajal bands on myelinating Schwann cells. In short, this case is an emblematic example of intermediate CMT with motor conduction slowing in the range formerly reported by Davis and colleagues [4], here being more marked in proximal nerve segments.

DI-CMTA (MIM#606483)

Rossi and colleagues reported a 5-generation CMT pedigree with male-to-male transmission and 21 affected members, 15 of them having been studied clinically and

electrophysiologically [90]. Sensorimotor semeiology appeared in the second decade and stabilized by the fourth decade. Median MNCV ranged between 25 and 45 m/s, but data on CMAP amplitudes are not given. Ulnar MNCV varied from 27 to 61 m/s (mean, 37.8), but again CMAP values are lacking. Nerve biopsy studies showed signs of axonal degeneration and demyelination [91]. The authors concluded that their electrophysiological and histological data support the proposal for an intermediate form of CMT. After discarding mutations of *PMP22*, *MPZ* and *NEFL*, linkage analysis demonstrated linkage to chromosome 10q24.1-q25.1 [92, 93].

DI-CMTB (MIM#606482)

By linkage analysis in two large Australian and US intermediate CMT pedigrees, a second locus was mapped to chromosome 19p [94-96]. Concise electrophysiological descriptions indicate that conduction velocities ranged from 24 to 54 m/s in the Australian pedigree, and that median nerve conduction velocities in two patients from the DUK118 pedigree were 36 and 46 m/s. Sural-nerve biopsy showed similar findings to those reported in DI-CMTA. In these two pedigrees and in one additional originating from Belgium, Züchner and colleagues identified unique mutations in the pleckstrin homology domain of dynamin 2 (*DNM2*) [97]. DNM2 belongs to the family of large GTPases and is part of the cellular fusion-fission apparatus.

Claeys and colleagues described detailed clinical, haematological, electrophysiological and sural nerve biopsy findings in 34 patients belonging to six independent families in whom a *DNM2* mutation had been identified [98], including those reported by Züchner and colleagues [97] and by us [99]. Patients presented with a classical CMT phenotype, which was mild to moderately severe. The disease could cosegregate with neutropenia or cataracts. The mean age of onset was 16 years, varying between 2 and 50 years. In 27 out the 34 patients electrophysiologically evaluated, median MNCVs ranged from 26 m/s to normal values indicating that the phenotypic spectrum of DNM2 mutations encompasses both intermediate and axonal patterns. Distinction of such patterns is not always an easy task. This question is illustrated in figures 2 and 3 corresponding to a pedigree associated with DNM2 G358R mutation [99]. Briefly, all three patients (proband and two affected daughters) showed marked lower-leg muscle atrophy and inexcitability of lower-limb nerves, a fact indicative of severe axonal degeneration. Starting from median MNCV in the proband, who had marked hand atrophy and even after examining the axilla-elbow segment, a diagnosis of DI-CMT could have been entertained, though widespread median and ulnar nerve CMAP attenuation pointed to an axonal disorder. It is worth remembering that a single conduction velocity value is not definitive of intermediate CMT, a term that should only be applied to the form and not the nerve conduction value [11]. This notion is applicable to the current family as affected proband's daughters had preserved median motor conduction parameters, indicative that in both we are confronted with a length- and agedependent axonal disorder of lower-limb nerves, not yet involving upper-limb nerves. Therefore, in addition to DI-CMT, DNM2 mutations may be associated with axonal CMT [100, 101], currently classified as CMT2M [26]. DI-CMTB and CMT2M are included under the same "Phenotype MIM number", 606482; in our view, this calls for

DI-CMTC (MIM#608323)

future revision.

Jordanova and colleagues reported two large unrelated families with DI-CMT linked to a novel locus on chromosome 1p35 [102]. Three years later, they identified two heterogeneous missense mutations (G41R and E196K) and one *de novo* deletion (15156delVKQV) in tyrosil-tRNA synthetase (YARS) in three unrelated families with DI-CMTC [103]. Aminoacyl-tRNA synthetases (ARSs) are ubiquitously expressed and essential enzymes responsible for charging tRNA with cognate amino-acids. Mutations in 28 of the 37 nuclear-encoded human ARS genes have been linked to a variety of recessive or dominant tissue-specific disorders [104]. YARS (also known as TyrRS) mutations causing DI-CMTC alter the normal function and distribution of YARS in neuronal endings, which in turn could affect synaptic plasticity resulting in axonal degeneration [103]. *Drosophila* model of the three DI-CMTC associated *YARS* mutations, which recapitulates several hallmarks of the human disease, suggests that the resulting phenotype is most likely due to gain-of-function alteration of the mutant TyRS or to an interference of unknown function of wild type protein [105].

Thomas and colleagues have reported detailed clinical, electrophysiological and morphological features in the original Bulgarian and US families [106], upon whom the disease locus was mapped [102]. Twenty-one affected individuals from the US family and 27 from the Bulgarian family were evaluated. Essentially, the clinical picture is a sensorimotor neuropathy, predominating in lower limbs with pes cavus, hammertoes and atrophy of extensor digitorum brevis. Mean ages of onset were 9 years in US patients and 20 years in Bulgarian patients. Median MNCV ranged from 29.5 to 45.6 m/s (mean, 34.1) in US family, and from 24.7 to 57.8 m/s (mean, 41.5) in Bulgarian family, CMAP amplitudes varying from 3.0 to 13.6 mV (mean, 7.7) and from 1.1 to 8.1 mV (mean, 5.1), respectively. It is worth noting that median CMAP amplitudes were abnormal 7 out of 41 studies; median DML were prolonged in ~50% of studies. Median MNCV against age suggested progressive slowing with stabilization in the fourth decade. Since CMAP amplitudes remained stable, axon loss probably plays a lesser role in the declining of nerve conduction velocities. Sural nerve biopsies of five patients revealed age-dependent axonal degeneration, reduced number of large myelinated fibres, some remyelinated axons, clusters of regeneration, and no onion bulbs. In short, the reported DI-CMTC phenotype is in line with the first description of intermediate CMT [4, 5].

DI-CMTD (MIM#607791)

Under the rubric of intermediate hereditary motor and sensory neuropathy, Mastaglia and colleagues reported a 4-generation CMT pedigree associated with *MPZ* D6Y mutation [107]. Electrophysiology in eight patients showed median MNCV ranging between 24 and 41 m/s (mean, 35), and ulnar MNCV ranging between 33 and 48 m/s (mean, 42.3). No data on CMAP are given, though sensory nerve potentials at the wrist were markedly delayed and attenuated in all affected members. Sural nerve biopsy in two cases showed advanced fibre loss, some thinly myelinated internodes, and absence of onion bulbs. Without knowing CMAP amplitude values, it does not seem possible to us to discard that the observed motor conduction slowing could merely be due to axonal loss (see above). In any case, three definite DI-CMT pedigrees associated with *MPZ* mutation have been reported [108-110], briefly analyzed below.

In a late-onset CMT family associated with *MPZ* K236del mutation, comprising two symptomatic and two sub-clinical heterozygous mutation carriers, median MNCV ranged from 29 to 56 m/s with normal CMAP amplitudes, and prolonged DML in two cases [108].

In a 4-generation family with a classic CMT phenotype harbouring *MPZ* K214M mutation, median MNCV in seven patients ranged from 34.2 to 40.1 m/s, CMAP amplitudes varying between 2.1 and 9.6 mV (\geq 4 in four patients) [109]. Intriguingly, examination of the axilla-elbow segment, performed in five patients, also showed motor

conduction slowing in the intermediate range. Median nerve DML ranged from 4.4 and 7.1 ms; F-latencies were prolonged in all three evaluated cases (37.5, 38 and 38.4 ms). As stated by the authors, these features cannot be accounted for by axonal degeneration and therefore unequivocally indicate that nerve conduction parameters are in the intermediate range.

A pedigree with *MPZ* W101X mutation, comprising 9 patients across 4 generations, showed a unique phenotype consisting of debilitating pain [110]. In the four patients electrophysiologically evaluated, median MNCV ranged from 38 to 43 m/s, CMAP amplitudes being always normal. DMLs were prolonged (4.5 to 4.9 ms) in three patients, whereas prolongation of F-latencies (32.1 and 35.6 ms) was noted in two out the three patients tested.

DI-CMTE (MIM#610982)

Inverted forming-2 (*INF2*) mutations are associated with 75% of patients suffering from DI-CMT and focal segmental glomerulosclerosis (FSGS) [111]. INF2 belongs to the group of proteins that regulates nucleation and elongation of actin filaments; when mutated, INF2 dysregulates actin-dependent processes, ultimately interfering with myelination and mitochondrial dynamics [112]. So far, 21 proband cases of CMT-FSGS have been reported associated with 15 different heterozygous point *INF2* mutations in the DID domain, which may appear either in dominant kinships or in sporadic cases (*de novo* mutation) [111-116]. Clinical phenotypes were similar to those seen in other CMT syndromes, onset usually occurring in the first or second decade, before the end-stage renal disease. There may be manifestations of CNS involvement. Median MNCVs, recorded in 18 patients, were as follows: i/ in nine patients ranged from 13 to 42 m/s (mean, 32), CMAP amplitudes not being reported [111, 112]; and ii/ as is characteristic

of intermediate CMT (see above), in the remaining nine patients ranged from 27 to 45 m/s (mean, 39), CMAP amplitudes being systematically normal [114-116]. Nerve biopsy studies have revealed a uniform pattern consisting of chronic demyelination associated with progressive axonal loss, accumulation of β -actin in the cytoplasm of Schwann cells, and cytoplasmic supernumerary elongated extensions in unmyelinating Schwann cells. These morphological features suggest that DI-CMTE is the first peripheral nerve disorder associated with Schwann cell actinopathy [113].

DI-CMTF (MIM#615185)

Lee and colleagues reported a dominant CMT pedigree with 11 affected members, six of them being symptomatic with onset between 10 and 45 years [117]. Clinical severity varied from no symptoms to being wheelchair-bound. Median nerve electrophysiology showed the characteristic intermediate CMT: MNCVs ranged from 16.5 to 45.7 m/s (mean, 32), and CMAP amplitudes from 3.2 to 9.6 mV (mean, 6.3). Sural biopsy in two patients revealed reduced myelinated fibres, clusters of regeneration and onion bulbs. The disease was mapped to 3q28-q29. Subsequent exome sequencing revealed a heterozygous mutation (G53D) in *GNB4*, encoding guanine-nucleotide-binding protein subunit beta-4 (G β_4), to cosegregate with the CMT phenotype in this family [118]. Therefore, the study identified *GNB4* mutation as a novel cause of CMT highlighting the importance of G β_4 -related G-protein-coupled-receptor signalling in peripheral-nerve function in humans. Another pathogenic heterozygous *GNB4* mutation (K57E) has been reported in a severe CMT1 patient [119].

DI-CMT and NEFL mutation

Neurofilament triplet proteins (heavy, medium and light) are the major intermediate filaments present in adult neurons and their expression is restricted to neuronal cell types of CNS and peripheral nervous system (PNS). Their main roles are to increase the axonal calibre of myelinated axons and consequently their conduction velocity, and contribute to the dynamic properties of the axonal cytoskeleton during neuronal differentiation, axon outgrowth, regeneration and guidance [120, 121]. CMT has mainly been associated with NEFL dominant missense mutations causing disruption of neurofilament assembly and aggregate formation with interference of axonal transport [122]. Resulting phenotypes have being classified as CMT1F (MIM#607734) and CMT2E (MIM#607684) [26]. There have been several descriptions of NEFL mutations associated with an intermediate electrophysiological pattern systematically accompanied by variably reduced CMAP amplitudes, making it difficult to establish whether nerve conduction slowing is attributable to reduction of axonal diameters [122-125]. We have reported intermediate electrophysiological patterns in two dominant CMT pedigrees associated with either NEFL E396K mutation [20] or NEFL N98S mutation [24]. Our first pedigree comprised four patients over two generations, aged between 35 and 59 years. The clinical picture was characterized by pes cavus, sensorimotor neuropathy and spastic gait. Both older patients showed ascending leg weakness to involve pelvic musculature. Figure 1 illustrates median MNCV study in the oldest patient; the remaining three showed intermediate median MNCVs with normal or minimally attenuated CMAPs. Median F waves and DML were moderately delayed. Our second pedigree comprised two patients, the proband and her son, aged 38 and 5 years. The proband showed delayed motor milestones that, as of the second decade, evolved into severe phenotype consisting of sensorimotor neuropathy, pes cavus,

clawing hands, gait and kinetic cerebellar ataxia, nystagmus and dysarthria, she becoming wheelchair bound. Her median MNCV study is illustrated in figure 4. As a whole, electrophysiological recordings indicate that accurate detection of intermediate motor conduction slowing may require exploration of proximal nerve segments in upper-limb nerves. Intriguingly, another *NEFL* N98S mutation with intermediate phenotype has recently been reported [126]. DI-CMT associated with *NEFL* mutations is a distinct and complex syndrome implying a simultaneous involvement of the PNS and CNS, which also calls for specific OMIM numbering.

DI-CMT associated with mutation in other genes

Mutations in the *mitofusin 2* (*MFN2*) are the most frequent cause of CMT2A (MIM# 609260) [127, 128]. In a Norwegian genetic survey of 232 consecutive unselected CMT pedigrees, *MFN2* point mutations were identified in 8 (3.4%) of them [129]. Pedigree 8 in this study, with two male patients, aged 23 and 50 years, harbouring heterozygous *MFN2* A716T mutation, showed median/ulnar MNCVs that ranged from 37/39 to 44/43 m/s with normal or slightly reduced CMAP amplitudes. This is probably the only autosomal dominant *MFN2* pedigree so far reported fulfilling electrophysiological criteria of intermediate CMT.

Point *AARS* mutations are associated with CMT2N (MIM#613287) [130]. In a cohort of six families with dominant *AARS*-related neuropathies, median MNCVs, obtained in eight patients, ranged from 26 to 47.6 m/s (mean, 36), CMAP amplitudes varying between 2 and 18 mV (mean, 7.2) [131]. Therefore, the electrophysiological pattern in these families fits well into that of intermediate CMT.

Spaans and colleagues reported a unique dominant phenotype combining myotonic dystrophy type 1, CMT, encephalopathic attacks, and sensoneurial hearing loss [131-134]. Median MNCV in 14 patients ranged from 23 to 48 m/s (mean, 37); although individual CMAP values are not indicated, it is specifically stated that in four patients the negative peak of the relevant CMAP was lower than 0.5 mV, which means that the corresponding MNCV value does not reliably represent the degree of demyelination [131]. Thus, it seems that intermediate CMT is an integral part of this exceptional pedigree.

Dynactin 2 (DCTN2) is a subunit of dynactin, a multiprotein complex associated with dynein. Mice over-expressing dynamitin, the p50 subunit of DCNT2, demonstrate a late-onset progressive motor unit degenerative syndrome [135]. Braathen and colleagues have identified a unique *DCNT2* H113Y DI-CMT pedigree with late onset phenotype [136]. In four electrophysiologically evaluated patients, median MNCV ranged from 35.5 to 50 m/s (mean, 46), and CMAP amplitudes from 1.5 to 6.8 mV (mean, 4). So far, no other intermediate CMT syndromes associated with *DCNT2* mutation have been reported.

Finally, in axonal dominant CMT pedigrees associated with *EGR2* R409Q mutation [137] or *HSP27* R17W mutation [138], there were isolated patients exhibiting intermediate median MNCV with normal or minimally attenuated CMAP amplitudes. We entirely agree in classifying both pedigrees within CMT2 given that axonal conduction pattern occurred in the majority of patients.

RI-CMTA (MIM#608340)

Mutations in ganglioside-induced differentiation associated protein (GDAP1), a new subfamily of the glutation-S-transferases, were initially described in autosomal recessive families showing a severe CMT phenotype sometimes with vocal cord paralysis, which are currently classified as CMT4A (MIM#214400) or AR-CMT2

(MIM#607706) [139, 140]. Heterozygous *GDAP1* mutations cause CMT2K, usually characterized by later onset, milder clinical picture, and axonal electrophysiology (MIM#607831) [141-144].

Baxter and colleagues found that motor conduction velocities ranged between 27 and 35 m/s; regrettably, reference to CMAP amplitudes is lacking [139]. Looking at their "Web Figure A", the outstanding histological feature in nerve biopsy is a decrease of large myelinated fibres without clear evidence to frequent onion bulbs, as mentioned in the paper. Although conduction values are in the intermediate CMT range, without knowing CMAP values this diagnostic label cannot be established with certainty.

In the series by Cuesta and colleagues [140], detailed electrophysiological and nerve biopsy studies showed the characteristic findings of a severe axonal CMT [145, 146]. Intriguingly, where peripheral nerves were inexcitable on distal stimulation, the motor latency of the axillary nerve was not delayed, as would have been expected in any intermediate CMT, but normal.

In another series of seven AR-CMT families associated with *GDAP1* mutations, the range of MNCV was variable: some patients having normal or near normal MNCV, whereas in other showing severe slowed MNCV [147]. The peripheral nerve biopsy findings were equally variable and showed features of demyelination or axonal degeneration.

The eponym RI-CMTA is used in OMIM in relation to the Polish pedigree reported by Kabzińska and colleagues, comprising two severely affected sisters, aged 29 and 21 years, harbouring homozygous *GDAP1* G327D mutation [148]. In elder patient ulnar MNCV was 32.7 m/s with CMAP of 0.2 mV and DML of 6.0 ms. In her sister ulnar and median nerves were inexcitable, whereas the axillary nerve exhibited attenuated CMAP (0.28 mV) with normal latency. Sural nerve biopsy showed marked loss of large myelinated fibres, clusters of regeneration, and a few onion bulbs. We concur with the authors that the current pedigree is a prototypic example of axonal CMT with autosomal recessive transmission.

Senderek and colleagues introduced the term "intermediate" in the title of a paper describing two Turkish families with mutations in *GDAP1* gene in autosomal recessive CMT neuropathy [149]. In one patient of each family, aged 7 and 6 years, median MNCV were 31 and 26.4 m/s with CMAP amplitudes of 0.5 and 0.3 mV, respectively. Sural nerve biopsies showed similar findings consisting of marked loss of large myelinated axons, active axonal degeneration, numerous clusters of regeneration, and occasional onion bulbs. As previously discussed, both electrophysiological and histological features may be explained by severe, primary axonal pathology.

Under the rubric of recessive intermediate CMT, Chung and colleagues reported two unrelated patients harbouring homozygous H256R or compound heterozygous P111H and V219G in *GDAP1* [150]. Patients were aged 5 (patient 1) and 8 (patient 2) years, and showed a severe CMT phenotype. In patient 1, at ages 4 and 5, median MNCVs were 38.7 and 37.5 m/s with CMAP amplitudes of 0.1 and 0.3 ms, respectively. In patient 2, at ages 7, 7.5 and 8 years, MNCVs were 44.8, 43.8 and 44.1 m/s with CMAP amplitudes of 4.5, 2.7 and 3.9 mV, respectively. Sural nerve biopsies showed decreased number of large myelinated fibres with unimodal histogram, clusters of regeneration, and onion- or pseudo-onion bulb formations. Starting from electrophysiological parameters and considering normal median MNCV and CMAP values at ages 1-6 and 6-12 years [35], only patient 2 could be included within the RI-CMTA. To further explore the role of ARSs in CMT disease (see above), McLaughlin and colleagues carried out a large-scale mutation screening of the 37 human ARS genes in a cohort of 355 patients with a phenotype consistent with CMT [151]. They identified two variants (L133H and Y173SfsX7) in the lysyl-tRNA synthesase (*KARS*) gene in a CMT patient showing developmental delay, self-abuse behaviour, dysmorphic features, and vestibular Schwannoma. Median/ulnar MNCVs were 39.5/30.6 m/s with CMAP amplitude of 0.5 mV. Although these MNCV values are in the intermediate range, severe CMAP attenuation, indicative of axonal loss, may account for the observed conduction slowing, and therefore the case should probably be reclassified as AR-CMT2.

RI-CMTC (MIM#615376)

Mutation in *Pleckstrin homology domain-containing family G member 5 (PLEKHG5)* gene has been reported in severe childhood onset lower motor neuron disease, and in three well-defined RI-CMTC pedigrees [152-154]. PH domain of PLEKHG5 contributes to the allosteric regulation of the RhoGEF domain, activating GTPases by stimulating the exchange of GDP to GTP, thereby initiating various signalling mechanisms that regulate neuronal shape and plasticity, dendrite growth, synapse formation, and neuronal survival.

Azzedine and colleagues reported two consanguineous pedigrees originated from Portugal and Morocco harbouring two different homozygous truncating mutations in the *PLEKHG5* gene [153]. The Portuguese family comprised two affected siblings, aged 43 and 53 years, and four unaffected siblings; in the Moroccan family four out of eight siblings were affected, aged between 42 and 51 years. With onset from the first to the fifth decade of life, the clinical picture consisted of distal lower- and upper-limb amyotrophy, areflexia, distal sensory loss, and foot deformities. There was no CNS semeiology. Median MNCV in four cases ranged from 35 to 39 m/s, CMAP amplitudes being normal in three patients or minimally reduced (3.7 mV) in the remaining one. Sural nerve biopsy showed reduced fibre density to about 70% of normal, with predominant loss of larger fibres and remyelinated fibres.

In a 19-year-old Korean patient, Kim and colleagues identified a novel compound heterozygous (T663M and G820R) mutations in the *PLEKHG5* gene [154]. The patient showed early onset CMT with moderate CMT neuropathy score (15 for a maximum of 36). At ages 14, 15 16 and 19 years, median MNCV were 25.3, 24.7, 25.6 and 29.2 m/s, CMAP amplitudes being always normal (between 5.5 and 9.5 mV). Median DMLs were prolonged (between 5.4 and 7 ms). Histological findings in sural nerve included complete loss of large- and medium-size myelinated fibres to 297/mm², rare regeneration clusters, and abnormal PLEKHG5 reaction pattern on immuno-histochemical study.

RI-CMTD (MIM#616039)

To identify the genetic background of Japanese CMT, Tamiya and colleagues analyzed the disease-causing mutation in 350 affected individuals, pathogenic mutations being found in 50% of demyelinating CMT and 20% of axonal CMT [155]. Using whole-genome sequencing, the authors found *COX6A1* mutation (c.247-10_247-6del CACTC) in two families with affected members from consanguineous marriages at different sites in Japan. The affected member in family 2, aged 39 years, had minimally reduced or normal median MNCVs (right median nerve, 45.3 m/s; left median nerve, 49.6 m/s) with preserved CMAP amplitudes, and inexcitable tibial nerve. We entirely agree with

the authors that, as a whole, such electrophysiology is indicative of axonal CMT almost selectively involving lower-limb nerves. The affected siblings of family 1 (V-1 and V-2), aged 30-39 years, showed similar clinical findings consisting of early and severe CMT phenotype. Electrophysiological and pathological data are tabulated in their tables S1 and S2. At 8 years of age, patient V-1 showed median/ulnar MNCV of 43.4/45.0 m/s with undetectable sensory nerve action potentials. Sural nerve biopsy showed decrease of myelinated fibres and onion bulbs. At ages 30-39 electrophysiological features were as follows: i/ patient V-1, median/ulnar MNCVs of 35.7/40.7 m/s with CMAP amplitudes of 0.7/1.3 mV, respectively; and ii/ patient V-2, median/ulnar MNCVs of 34.8/37.3 m/s with CMAP amplitudes of 0.08/0.05 mV, respectively. In our view, these electrophysiological data are indicative of a severe axonal neuropathy.

In a recent report, Laššuthová and colleagues confirmed that *COX6A1* homozygous mutation (according to the authors, the correct nomenclature for the deletion should be c.247-7_247-3del) may cause severe, early-onset axonal CMT [156]. In their 37-year-old patient, median/ulnar MNCV were 44 and 46 m/s, CMAP amplitudes being 1.2/1.1 mV, respectively

As said before, exploration of proximal nerve segments of median or ulnar nerves would have been very useful to demonstrate a possible pattern of intermediate motor nerve conduction (see figures 1 and 4). In any case, we are persuaded that up to now the reported COX6A1-related disorders should be re-classified under the rubric of AR-CMT2, thus RI-CMTD becoming vacant.

Discussion

To the best of our knowledge, this is the first systematic review of intermediate CMT. The notion of intermediate CMT initially emerged as a particular pattern of median MNCV slowing, different from that of CMT1 (usually <25 m/s) and CMT2 (usually >45 m/s) [4]. So, MNCV limits were firstly established between 25 and 45 m/s, soon after being narrowed down to 30-40 m/s [7]. Although these electrophysiological limits have widely been accepted in the literature, one important drawback is that the contribution of CMAP amplitude reduction, reflecting axonal loss, to conduction slowing was not specifically taken into account. It is important to note that nerve biopsies in original intermediate CMT series showed similar pathological features consisting of loss of large myelinated fibres, clusters of regeneration, and a few onion bulbs [5, 8], that is, a combination of axonal and demyelinating changes.

As reported for sural nerve in patients with either CMT or acquired neuropathy, the velocity is predictable from the largest fibres present in the biopsy [11, 21]. Overall, nerve biopsy and autopsy studies in CMT2 have demonstrated length-dependent loss of large myelinated fibres ($\geq 7\mu$ m) and clusters of regeneration [15-17]. In regards to median MNCV at the elbow-wrist segment and applying sural nerve conversion factor of 3.85 reported in axonal neuropathy [21], such pathologic framework could imply a secondary slowing down of motor conduction into the intermediate range or even into the demyelinating range. Under such circumstances is not an easy task to determine whether median MNCV slowing in intermediate CMT is accounted for by loss of rapidly conducting fibres or by demyelination [11]. Based on our experience with intermediate CMT [20, 24] and previous electrophysiological and pathological considerations (see above), figure 5 provides a proposed algorithm of the electrophysiological approach when investigating a patient with presumptive intermediate CMT.

Having in mind all these aspects, we proceeded to review PubMed descriptions published under the rubric of intermediate CMT or X-linked CMT. We adhered to the classification proposed in updated review CMT papers [26, 27] (see table 1).

CMTX1 in males is the most frequent cause of intermediate CMT. We found 40 papers describing median MNCV study (alternatively ulnar or peroneal nerves) (see table 2). The great majority of male CMTX1 patients showed median MNCV in the intermediate range, though distal CMAP amplitudes were often attenuated, making it difficult to establish the contribution of axonal loss to conduction slowing.

DRP2 mutation causes an exceptional, well-defined form of X-linked intermediate CMT [89], whose phenotype MIM number has not yet been assigned.

DI-CMT encompasses six numbered MIM phenotypes (see table 1). DI-CMTA is assigned to the pedigree reported by Rossi and colleagues [90]. Its locus mapped to chromosome 10q [92, 93], but the responsible gene has not yet been cloned. Certainly, in this pedigree ulnar MNCVs were within the intermediate range (see above), but considering that CMAP amplitudes are lacking, the question arises as to whether this family could merely represent an example of CMT2 with slowed motor conduction due to potentially severe loss of large axons [22, 23]. DI-CMTB is associated with *DNM2* gene mutations covering a wide phenotypic expression, which includes axonal and intermediate CMT phenotypes sometimes associated with neutropenia or cataracts [98]. Our nerve conduction study in a *DNM2* G359R CMT2 pedigree illustrates that deciphering of the electrophysiological hallmark may require not only investigation of the proband, but also of other less affected secondary cases (see figures 2, 3 and 5) [99], thus emphasizing the notion that the term intermediate should only be applied to the

-30-

form and not to nerve conduction values [11]. *YARS* mutations are associated with DI-CMTC, a well-defined electrophysiological intermediate pattern in the reported pedigrees [102, 103, 106]. DI-CMTD was initially reported in a family harbouring *MPZ* D6Y mutation, whose eight patients showed median MNCV in the intermediate range, CMAP amplitudes being lacking [107]. Afterwards, there have been three family reports conforming to the characteristic hallmark of intermediate CMT [108-110]. The relevant diagnostic role of investigating proximal median MNCV for accurate detection of intermediate conduction slowing is demonstrated in one of these pedigrees [110]. DI-CMTE is due to *INF2* mutations, manifesting with intermediate CMT and FSGS [111]. DI-CMTF caused by *GNB4* mutations is another well-defined intermediate phenotype [117, 118], though this gene mutation may also cause CMT1 with severe nerve

conduction slowing [119].

Additional DI-CMT forms, not yet numbered in OMIM, are as follows: i/ that associated with *NEFL* mutations, usually showing CNS involvement [20, 24]; ii/ that associated with *MFN2* mutations, just one kinship having been reported [129]; iii/ that associated with *AARS* mutations [131]; iv/ the exceptional phenotype combining myotonic dystrophy type 1 and CMT [132-134]; and v/ that associated with *DCNT2* mutation [136].

Currently, RI-CMT comprises four phenotypes (see table 1). RI-CMTA is associated with *GDAP1* mutation. Most *GDAP1* mutations cause AR-CMT2 and less frequently CMT2K [140, 141, 143-146]. Median MNCVs in the intermediate range have been reported in several pedigrees [136, 145, 146], but their correct electrophysiological classification is not possible given that CMAP amplitudes were severely reduced or not specified. In fact, just one the patients reported by Chung and colleagues could be included within RI-CMTA [150]. Individualization of RI-CMTB has been based upon a single pedigree associated with *KARS* mutation [151]. Our review indicates that electrophysiology is compatible with a severe axonal neuropathy, so this pedigree should be reclassified within AR-CMT2 phenotypes, RI-CMTB becoming a vacant rubric. RI-CMTC is a well-documented intermediate CMT phenotype caused by *PELKHG5* mutation [153, 154]. RI-CMTD was individualized in two Japanese pedigrees with *COX6A1* mutation, showing either median MNCV over 45 m/s, or median MNCV in the intermediate range with severe CMAP attenuation [155]. Therefore, we propose that this disorder should be reclassified within AR-CMT2, RI-CMTC being another vacant rubric.

We conclude that intermediate CMT is a complex inherited syndrome, whose characterization requires a specific electrophysiological protocol comprising evaluation of upper-limb proximal nerve trunks when distal CMAP amplitudes are reduced or even complementary evaluation of other affected kinship members. An updated version of MIM phenotype numbering is needed.

Acknowledgements. The authors are grateful to Mr Mario Corral (Director of "Marquesa de Pelayo" Library, Santander, Spain) for his invaluable help in literature search. The authors also thank Drs Mario Reviriego and Antonio Murciano (Smart Intelligence Services, SL, Madrid, Spain) for technical support, Mrs Marta de la Fuente for secretarial work, and Miss Mar Ruiz for her help in contacting our patients.

Compliance with ethical standards.

Conflicts of interests. None.

REFERENCES

1. Combarros O, Calleja J, Polo JM, Berciano J (1987) Prevalence of hereditary motor and sensory neuropathy in Cantabria. Acta Neurol Scand 75: 9-12. 2. Dyck PJ (2005) Inherited neuronal degeneration and atrophy affecting peripheral motor, sensory, and autonomic neurons. In: Dyck PJ, Thomas PK, Lambert EH and Bunge R (eds): Peripheral neuropathy, 2nd ed. Philadelphia, WB Saunders, p 1600-1655. 3. Harding AE, Thomas PK (1980) The clinical features of hereditary motor and sensory neuropathy types I and II. Brain 103: 259-280. 4. Davis CJ, Bradley WG, Madrid R (1978) The peroneal muscular atrophy syndrome: clinical, genetic, electrophysiological and nerve biopsy studies. I. Clinical, genetic and electrophysiological findings and classification. J Genet Hum 26: 311-49. 5. Madrid R, Bradley WG, Davis CJ (1977) The peroneal muscular atrophy syndrome. Clinical, genetic, electrophysiological and nerve biopsy studies. Observations on pathological changes in sural nerve biopsies. J Neurol Sci 32: 91-122. Buchthal F, Behse F (1977) Peroneal muscular atrophy (PMA) and related disorders. I. Clinical manifestations as related to biopsy findings, nerve conduction and electromyography. Brain 100: 41-66. 7. Bouché P, Gherardi R, Cathala HP, Lhermitte F, Castaigne P (1983) Peroneal muscular atrophy. Part 1. Clinical and electrophysiological study. J Neurol Sci 61: 389-399. 8. Gherardi R, Bouché P, Escourolle R, Hauw JJ (1983) Peroneal muscular atrophy. Part 2. Nerve biopsy studies. J Neurol Sci 61: 401-416. 9. Saporta AS, Sottile SL, Miller LJ, Feely SM, Siskind CE, Shy ME (2011) Charcot-Marie-Tooth disease subtypes and genetic testing strategies. Ann Neurol 69: 22-33. 10. Klein CJ, Duan X, Shy ME (2013) Inherited neuropathies: clinical overview and update. Muscle Nerve 48: 604-622. 11. Nicholson G, Myers S (2006) Intermediate forms of Charcot-Marie-Tooth neuropathy: a review. Neuromolecular Med 8: 123-130. 12. Preston DC, Shapiro BE (2005) Electromyography and neuromuscular disorders. Clinicalelectrophysiologic correlations. 2nd ed. Elsevier/Butterworth Heinemann, Philadelphia. 13. Tankisi H, Pugdahl K, Johnsen B, Fuglsang-Frederiksen A (2007) Correlations of nerve conduction measures in axonal and demyelinating polyneuropathies. Clin Neurophysiol 118: 2383-2392. 14. Lawson VH, Gordon Smith A, Bromberg MB (2003) Assessment of axonal loss in Charcot-Marie-Tooth neuropathies. Exp Neurol 184: 753-757. 15. Behse F, Buchthal F (1977) Peroneal muscular atrophy (PMA) and related disorders. II. Histological findings in sural nerves. Brain 100: 67-85. 16. Berciano J, Combarros O, Figols J, Calleja J, Cabello A, Silos I, Coria F (1986) Hereditary motor and sensory neuropathy type II. Clinicopathological study of a family. Brain 109: 897-914.

	17. Hahn AF (1993) Hereditary motor and sensory neuropathy: HMSN type II (neuronal type) and
1	X-linked HMSN Brain Pathol 3: 147-155
2	18 Waxman SG (1080) Determinants of conduction valuative in muclimated nerve fibers. Musela
3 4	Nome 2: 141, 150
5	10 Nerve 5. 141-150.
6	19. Yum Sw, Zhang J, Mo K, Li J, Scherer SS (2009) A novel recessive NeTI mutation causes a
8	severe, early-onset axonal neuropathy. Ann Neurol 66: 759-770.
9	20. Berciano J, Peeters K, García A, López-Alburquerque T, Gallardo E, Hernández-Fabián A,
10	Pelayo-Negro AL, De Vriendt E, Infante J, Jordanova A (2016) NEFL N98S mutation: another
12	cause of dominant intermediate Charcot-Marie-Tooth disease with heterogeneous early-onset
13	phenotype. J Neurol 263: 361-369.
15	21. Oh SJ, Hemmi S, Kurokawa K, Hatanaka Y (2010) Intraoperative on-nerve nerve conduction
16	study and conversion factor in the sural nerve. Muscle Nerve 42: 373-378.
18	22. Bienfait HM, Verhamme C, van Schaik IN, Koelman JH, de Visser BW, de Haan RJ, Baas F,
19	van Engelen BG, de Visser M (2006). Comparison of CMT1A and CMT2: similarities and
20 21	differences. J Neurol 253: 1572-1580.
22	23. Bienfait HM, Baas F, Koelman JH, de Haan RJ, van Engelen BG, Gabreëls-Festen AA,
23	Ongerboer de Visser BW Meggouh F Weterman MA De Jonghe P Timmerman V de Visser
25	M (2007) Phenotype of Charcot-Marie-Tooth disease Type 2 Neurology 68: 1658-1667
26 27	24. Parajano I. Garaja A. Paatara K. Gallarda E. Da Vriandt E. Dalava Nagra A. Infanta I.
28	24. Bereland J, Gareta A, Feeters K, Ganardo E, De Vitendi E, Felayo-Negro AL, infante J,
29	Jordanova A (2015) NEFL E396K mutation is associated with a novel dominant intermediate
30 31	Charcot-Marie-Tooth disease phenotype. J Neurol 262: 1289-1300.
32	25. Rossor AM, Polke JM, Houlden H, Reilly MM (2013) Clinical implications of genetic advances
33 34	in Charcot-Marie-Tooth disease. Nat Rev Neurol 9: 562-71.
35	26. Rossor AM, Tomaselli PJ, Reilly MM (2016) Recent advances in the genetic neuropathies. Curr
36	Opin Neurol 29: 537-48.
38	27. Liu L, Zhang R (2014) Intermediate Charcot-Marie-Tooth disease. Neurosci Bull 30: 999-1009.
39	28. Baets J, De Jonghe P, Timmerman V (2014). Recent advances in Charcot-Marie-Tooth disease.
40 41	Curr Opin Neurol 27: 532-540.
42	29. Nicolaou P. Zamba-Papanicolaou E. Koutsou P. Kleopa KA. Georghiou A. Hadiigeorgiou G.
43	Panadimitriou A Kyriakides T Christodoulou K (2010) Charcot-Marie-Tooth disease in
45	Currence anidamialogical alinical and constitution Alexandraticities Neuroanidamialogy 25: 171-177
46	Cyprus. epidemiological, chinical and genetic characteristics. Neuroepidemiology 55. 171-177.
47	30. Murphy SM, Laura M, Fawcett K, Pandraud A, Liu YT, Davidson GL, Rossor AM, Poike JM,
49	Castleman V, Manji H, Lunn MP, Bull K, Ramdharry G, Davis M, Blake JC, Houlden H, Reilly
50 51	MM (2012) Charcot-Marie-Tooth disease: frequency of genetic subtypes and guidelines for
52	genetic testing. J Neurol Neurosurg Psychiatry 83: 706-710.
53	31. Braathen GJ (2012) Genetic epidemiology of Charcot-Marie-Tooth disease. Acta Neurol Scand
54 55	Suppl (193): iv-22.
56	
57 58	
59	
60	
©⊥ 62	
63	
64 65	

32.	. Østern R, Fagerheim T, Hjellnes H, Nygård B, Mellgren SI, Nilssen Ø (2013) Diagnostic
	laboratory testing for Charcot Marie Tooth disease (CMT): the spectrum of gene defects in
	Norwegian patients with CMT and its implications for future genetic test strategies. BMC Med
	Genet 14: 94.
33.	. Laššuthová P, Šafka Brožková D, Krůtová M, Neupauerová J, Haberlová J, Mazanec R, Dřímal
	P, Seeman P (2016) Improving diagnosis of inherited peripheral neuropathies through gene panel
	analysis. Orphanet J Rare Dis 11: 118.
34.	. Lefter S, Hardiman O, Ryan AM (2017) A population-based epidemiologic study of adult
	neuromuscular disease in the Republic of Ireland. Neurology 88: 304-313.
35.	Moher D, Liberati A, Tetzlaff J, Altman DG; PRISMA Group (2009) Preferred reporting items
	for systematic reviews and meta-analyses: the PRISMA statement. J Clin Epidemiol 62: 1006-
	1012.
36.	. García A, Calleja J, Antolín FM, Berciano J (2000) Peripheral motor and sensory nerve
	conduction studies in normal infants and children. Clin Neurophysiol 111: 513-520.
37.	. Midroni G, Bilbao JM (1995) Biopsy diagnosis of peripheral neuropathy. Boston Butterworth-
	Heineman, Boston, p 35-43.
38.	. Hahn AF, Brown WF, Koopman WJ, Feasby TE (1990) X-linked dominant hereditary motor and
	sensory neuropathy. Brain 113: 1511-1525.
39.	. Rozear MP, Pericak-Vance MA, Fischbeck K, Stajich JM, Gaskell PC Jr, Krendel DA, Graham
	DG, Dawson DV, Roses AD (1987). Hereditary motor and sensory neuropathy, X-linked: a half
	century follow-up. Neurology 37: 1460-1465.
40.	. Hahn AF, Bolton CF, White CM, Brown WF, Tuuha SE, Tan CC, Ainsworth PJ (1999)
	Genotype/phenotype correlations in X-linked dominant Charcot-Marie-Tooth disease. Ann N Y
	Acad Sci 883: 366-382.
41.	. Sommer CL, Brandner S, Dyck PJ, Harati Y, LaCroix C, Lammens M, Magy L, Mellgren SI,
	Morbin M, Navarro C, Powell HC, Schenone AE, Tan E, Urtizberea A, Weis J; Peripheral Nerve
	Society (2010) Peripheral Nerve Society Guideline on processing and evaluation of nerve
	biopsies. J Peripher Nerv Syst 15: 164-175.
42.	. Scherer SS, Kleopa KA (2012) X-linked Charcot-Marie-Tooth disease. J Peripher Nerv Syst 17
	Suppl 3: 9-13.
43.	. Shy ME, Siskind C, Swan ER, Krajewski KM, Doherty T, Fuerst DR, Ainsworth PJ, Lewis RA,
	Scherer SS, Hahn AF (2007) CMT1X phenotypes represent loss of GJB1 gene function.
	Neurology 68: 849-855.
44.	. Siskind CE, Murphy SM, Ovens R, Polke J, Reilly MM, Shy ME (2011) Phenotype expression
	in women with CMT1X. J Peripher Nerv Syst 16: 102-107.
45.	. Jerath NU, Gutmann L, Reddy CG, Shy ME (2016) Charcot-Marie-Tooth disease type 1X in
	women: Electrodiagnostic findings. Muscle Nerve 54: 728-732.
46.	Phillips LH, Kelly TE, Schnatterly P, Parker D (1985) Hereditary motor-sensory neuropathy
	(HMSN): possible X-linked dominant inheritance. Neurology 35: 498-502.

	47. Fischbeck KH, ar-Rushdi N, Pericak-Vance M, Rozear M, Roses AD, Fryns JP (1986) X-linked
1	neuropathy: gene localization with DNA probes. Ann Neurol 20: 527-532.
2	48 Kleona KA Abrams CK Scherer SS (2012) How do mutations in GIB1 cause X-linked
3 4	Charact Maria Tasth Jianas 2 Davis Des 1497, 100 205
5	Charcot-Marie-Tooth disease? Brain Kes 1487: 198-205.
6	49. Anzini P, Neuberg DH, Schachner M, Nelles E, Willecke K, Zielasek J, Toyka KV, Suter U,
/	Martini R (1997) Structural abnormalities and deficient maintenance of peripheral nerve myelin
9	in mice lacking the gap junction protein connexin 32. J Neurosci 17: 4545-51.
10	50. Scherer SS, Xu YT, Nelles E, Fischbeck K, Willecke K, Bone LJ (1998) Connexin32-null mice
12	develop demyelinating peripheral neuropathy. Glia 24: 8-20.
13	51. Vavlitou N. Sargiannidou I. Markoullis K. Kvriacou K. Scherer SS. Kleopa KA (2010) Axonal
14	nathology precedes demyelination in a mouse model of X-linked demyelinating/type I Charcot-
16	Maria Taath reuronathu. I Neuronathal Eur Neurol 60: 045-058
17	Marie 100th neuropathy. J Neuropathol Exp Neurol 69. 943-938.
18 19	52. Senderek J, Hermanns B, Bergmann C, Boroojerdi B, Bajbouj M, Hungs M, Ramaekers VT,
20	Quasthoff S, Karch D, Schröder JM (1999) X-linked dominant Charcot-Marie-Tooth neuropathy.
21	clinical, electrophysiological, and morphological phenotype in four families with different
22	connexin32 mutations. J Neurol Sci 167: 90-101.
24	53. Hahn AF, Ainsworth PJ, Naus CC, Mao J, Bolton CF (2000) Clinical and pathological
25	observations in men lacking the gap junction protein connexin 32. Muscle Nerve Suppl 9: S39-
20 27	48
28	54 Hahn AF Ainsworth PI Bolton CF Bilbao IM Vallat IM (2001) Pathological findings in the v
29 30	1. Hann AI, Answord II, Doron CI, Dhoao Jwi, Vanat Jwi (2001) Famological midings in the A
31	linked form of Charcot-Marie-Tooth disease: a morphometric and ultrastructural analysis. Acta
32	Neuropathol 101: 129-139.
33 34	55. Kleopa KA, Zamba-Papanicolaou E, Alevra X, Nicolaou P, Georgiou DM, Hadjisavvas A,
35	Kyriakides T, Christodoulou K (2006) Phenotypic and cellular expression of two novel
36	connexin32 mutations causing CMT1X. Neurology 66: 396-402.
37	56. Spencer PS, Thomas PK (1974) Ultrastructural studies of the dying-back process. II. The
39	sequestration and removal by Schwann cells and oligodendrocytes of organelles from normal
40	and diseases axons. I Neurocytol 3: 763-383
41 42	57 Parajana I. Caraja A. Figala I. Muñoz P. Parajana MT. Lafarga M. (2000) Paringurium
43	57. Berelano J, Garela A, Figois J, Munoz K, Berelano MT, Latarga M (2000) Fernicultum
44	contributes to axonal damage in acute inflammatory demyelinating polyneuropathy. Neurology.
46	55: 552-559.
47	58. Mostacciuolo ML, Müller E, Fardin P, Micaglio GF, Bardoni B, Guioli S, Camerino G, Danieli
48 49	GA (1991) X-linked Charcot-Marie-Tooth disease. A linkage study in a large family by using 12
50	probes of the pericentromeric region. Hum Genet 87: 23-27.
51	59. Ionasescu VV, Trofatter J, Haines JL, Summers AM, Ionasescu R, Searby C (1992) X-linked
52 53	recessive Charcot-Marie-Tooth neuropathy: clinical and genetic study. Muscle Nerve 15: 368-
54	373
55 56	60 Nicholson G. Nach I (1002) Intermediate narya conduction valuation define V linked Charact
57	00. Metholson G, Nash J (1995) Intermediate herve conduction velocities denne X-miked Charcot-
58	Marie-1 ooth neuropathy families. Neurology 43: 2558-2564.
59 60	
61	
62	
63 64	
65	

61.	Oterino A, Montón FI, Cabrera VM, Pinto F, Gonzalez A, Lavilla NR (1996) Arginine-164-
	tryptophan substitution in connexin32 associated with X linked dominant Charcot-Marie-Tooth
	disease. J Med Genet 33: 413-415.
62.	Ionasescu V, Ionasescu R, Searby C (1996) Correlation between connexin 32 gene mutations
	and clinical phenotype in X-linked dominant Charcot-Marie-Tooth neuropathy. Am J Med Genet
	63: 486-491.
63.	Birouk N, LeGuern E, Maisonobe T, Rouger H, Gouider R, Tardieu S, Gugenheim M, Routon
	MC, Léger JM, Agid Y, Brice A, Bouche P (1998) X-linked Charcot-Marie-Tooth disease with
	connexin 32 mutations: clinical and electrophysiologic study. Neurology 50: 1074-1082.
64.	Ainsworth PJ, Bolton CF, Murphy BC, Stuart JA, Hahn AF (1998) Genotype/phenotype
	correlation in affected individuals of a family with a deletion of the entire coding sequence of the
	connexin 32 gene. Hum Genet 103: 242-244.
65.	Sander S, Nicholson GA, Ouvrier RA, McLeod JG, Pollard JD (1998). Charcot-Marie-Tooth
	disease: histopathological features of the peripheral myelin protein (PMP22) duplication
	(CMT1A) and connexin32 mutations (CMTX1). Muscle Nerve 21: 217-225.
66.	Gutierrez A, England JD, Sumner AJ, Ferer S, Warner LE, Lupski JR, Garcia CA (2000)
	Unusual electrophysiological findings in X-linked dominant Charcot-Marie-Tooth disease.
	Muscle Nerve 23: 182-188.
67.	Dupré N, Cossette L, Hand CK, Bouchard JP, Rouleau GA, Puymirat J (2001) A founder
	mutation in French-Canadian families with X-linked hereditary neuropathy. Can J Neurol Sci 28:
	51-55.
68.	Nakagawa M, Takashima H, Umehara F, Arimura K, Miyashita F, Takenouchi N, Matsuyama
	W, Osame M (2001) Clinical phenotype in X-linked Charcot-Marie-Tooth disease with an entire
	deletion of the connexin 32 coding sequence. J Neurol Sci 185: 31-37.
69.	Dubourg O, Tardieu S, Birouk N, Gouider R, Léger JM, Maisonobe T, Brice A, Bouche P,
	LeGuern E (2001) Clinical, electrophysiological and molecular genetic characteristics of 93
	patients with X-linked Charcot-Marie-Tooth disease. Brain 124: 1958-1967.
70.	Hattori N, Yamamoto M, Yoshihara T, Koike H, Nakagawa M, Yoshikawa H, Ohnishi A,
	Hayasaka K, Onodera O, Baba M, Yasuda H, Saito T, Nakashima K, Kira J, Kaji R, Oka N,
	Sobue G; Study Group for Hereditary Neuropathy in Japan (2003) Demyelinating and axonal
	features of Charcot-Marie-Tooth disease with mutations of myelin-related proteins (PMP22,
	MPZ and Cx32): a clinicopathological study of 205 Japanese patients. Brain 126: 134-1351.
71.	Capasso M, Di Muzio A, Ferrarini M, De Angelis MV, Caporale CM, Lupo S, Cavallaro T,
	Fabrizi GM, Uncini A (2004) Inter-nerves and intra-nerve conduction heterogeneity in CMTX
	with Arg(15)Gln mutation. Clin Neurophysiol 115: 64-70.
72.	Houlden H, Girard M, Cockerell C, Ingram D, Wood NW, Goossens M, Walker RW, Reilly
	MM (2004) Connexin 32 promoter P2 mutations: a mechanism of peripheral nerve dysfunction.
	Ann Neurol 56: 730-734.
73.	Ryan MM, Jones HR Jr (2005) CMTX mimicking childhood chronic inflammatory
	demyelinating neuropathy with tremor. Muscle Nerve 31: 528-530.

	74. Vondracek P, Seeman P, Hermanova M, Fajkusova L (2005). X-linked Charcot-Marie-Tooth
1	disease: phenotypic expression of a novel mutation Ile127Ser in the GJB1 (connexin 32) gene.
2	Muscle Nerve $31 \cdot 252 - 255$
3	
5	75. Liang GS, de Miguel M, Gomez-Hernandez JM, Glass JD, Scherer SS, Mintz M, Barrio LC,
6	Fischbeck KH (2005) Severe neuropathy with leaky connexin32 hemichannels. Ann Neurol 57:
7	749-754.
8	76 Zhang RX Luo W Zi XH Xia K Cai F Xiao IF Zhao GH Zhang FF Shen L Jiang H Tang
10	DS (2005) Mutation concerning of Cu22 in Han Chinasa national with Charact Maria Taoth
11	BS (2005) Mutation screening of Cx52 in Han Chinese patients with Charcot-Iviane-100th
12	disease. Beijing Da Xue Xue Bao 37: 68-71.
13	77. Huttner IG, Kennerson ML, Reddel SW, Radovanovic D, Nicholson GA (2006) Proof of genetic
15	heterogeneity in X-linked Charcot-Marie-Tooth disease. Neurology 67: 2016-2021.
16	78. Beauvais K, Furby A, Latour P (2006) Clinical, electrophysiological and molecular genetic
17	studies in a family with X linked dominant Charact Maria Tooth neuronathy presenting a neval
19	studies in a family with x-miked dominant Charcot-Marie-Tooth neuropathy presenting a nover
20	mutation in GJB1 Promoter and a rare polymorphism in LITAF/SIMPLE. Neuromuscul Disord
21	16: 14-18.
22	79. Vazza G, Merlini L, Bertolin C, Zortea M, Mostacciuolo ML (2006) A novel 9-bp insertion in
24	the GJB1 gene causing a mild form of X-linked CMT with late onset. Neuromuscul Disord 16:
25	878-881
26	
28	80. Kim HS, Chung KW, Kang SH, Choi SK, Cho SY, Koo H, Kim SB, Choi BO (2010) Myotonic
29	dystrophy type I combined with X-linked dominant Charcot-Marie-Tooth neuropathy.
30	Neurogenetics 11: 425-433.
31 32	81. Vrancken AF, Spliet WG, van Ruissen F (2010) X-linked Charcot-Marie-Tooth disease with
33	novel c 47A>T GJB1 gene mutation J Perinher Nerv Syst 15: 156-157
34	22. Sakagushi II. Vamashita S. Miyra A. Hirahara T. Kimura E. Maada V. Tarasaki T. Hirana T.
35	62. Sakaguchi H, Famashita S, Mitura A, Hiranara T, Kiniura E, Maeua T, Terasaki T, Hirano T,
37	Uchino M (2011) A novel GJB1 frameshift mutation produces a transient CNS symptom of X-
38	linked Charcot-Marie-Tooth disease. J Neurol 258: 284-290.
39	83. Yiu EM, Geevasinga N, Nicholson GA, Fagan ER, Ryan MM, Ouvrier RA (2011) A
40	retrospective review of X-linked Charcot-Marie-Tooth disease in childhood. Neurology 76: 461-
42	466
43	04 Marchar CM Dellas I Marii II Dialas I Deiniana I. Conservato M. Harddan II Dura dara C. Deilla
44	84. Murphy SM, Poike J, Manji H, Blake J, Keiniger L, Sweeney M, Houlden H, Brandner S, Keiliy
46	MM (2011) A novel mutation in the nerve-specific 5'UTR of the GJB1 gene causes X-linked
47	Charcot-Marie-Tooth disease. J Peripher Nerv Syst 16: 65-70.
48	85. Chen SD, Li ZX, Guan YT, Zhou XJ, Jiang JM, Hao Y (2011) A novel mutation of gap junction
49 50	protein ß 1 gene in X-linked Charcot-Marie-Tooth disease. Muscle Nerve 43: 887-892.
51	86. Borgulová I. Mazanec P. Sakmarvová I. Havlová M. Safka Brožková D. Seeman P. (2013)
52	biguiova i, mazance K, sakinaryova i, naviova ivi, saika Biozkova D, seeman i (2015)
53 54	Mosaicism for GJB1 mutation causes milder Charcot-Marie-Tooth X1 phenotype in a
55	heterozygous man than in a manifesting heterozygous woman. Neurogenetics 14: 189-195.
56	87. Zhao Y, Xie Y, Zhu X, Wang H, Li Y, Li J (2014) Transient, recurrent, white matter lesions in
57	x-linked Charcot-Marie-tooth disease with novel mutation of gap junction protein beta 1 gene in
59	China: a case report. BMC Neurol 14: 156.
60	
61	
o∠ 63	
-	

88.	Liu L, Li X, Hu Z, Zi X, Zhao X, Xie Y, Huang S, Xia K, Tang B, Zhang R (2016). Phenotypes
	and cellular effects of GJB1 mutations causing CMT1X in a cohort of 226 Chinese CMT
	families. Clin Genet Nov 2. doi: 10.1111/cge.12913. [Epub ahead of print].
89.	Brennan KM, Bai Y, Pisciotta C, Wang S, Feely SM, Hoegger M, Gutmann L, Moore SA,
	Gonzalez M, Sherman DL, Brophy PJ, Züchner S, Shy ME (2015) Absence of Dystrophin
	Related Protein-2 disrupts Cajal bands in a patient with Charcot-Marie-Tooth disease.
	Neuromuscul Disord 25: 786-793.
90.	Rossi A, Paradiso C, Cioni R, Rizzuto N, Guazzi G (1985) Charcot-Marie-Tooth disease: study
	of a large kinship with an intermediate form. J Neurol 232: 91-98.
91.	Malandrini A, Ceuterick C, Villanova M, Gambelli S, Berti G, Rossi A, Guazzi GC (2001)
	Ultrastructural findings in the peripheral nerve in a family with the intermediate form of
	Charcot-Marie-Tooth disease. J Submicrosc Cytol Pathol 33: 59-63.
92.	Villanova M, Timmerman V, De Jonghe P, Malandrini A, Rizzuto N, Van Broeckhoven C,
	Guazzi G, Rossi A (1998) Charcot-Marie-Tooth disease: an intermediate form. Neuromuscul
	Disord 8: 392-393.
93.	Verhoeven K, Villanova M, Rossi A, Malandrini A, De Jonghe P, Timmerman V (2001)
	Localization of the gene for the intermediate form of Charcot-Marie-Tooth to chromosome
	10q24.1-q25.1.Am J Hum Genet 69: 889-894.
94.	Kennerson ML, Zhu D, Gardner RJ, Storey E, Merory J, Robertson SP, Nicholson GA (2001)
	Dominant intermediate Charcot-Marie-Tooth neuropathy maps to chromosome 19p12-p13.2.
	Am J Hum Genet 69: 883-888.
95.	Zhu D, Kennerson M, Merory J, Chrast R, Verheijen M, Lemke G, Nicholson G (2003) Refined
	localization of dominant intermediate Charcot-Marie-Tooth neuropathy and exclusion of seven
	known candidate genes in the region. Neurogenetics 4: 179-183.
96.	Speer MC, Graham FL, Bonner E, Collier K, Stajich JM, Gaskell PC, Pericak-Vance MA, Vance
	JM (2002) Reduction in the minimum candidate interval in the dominant-intermediate form of
	Charcot-Marie-Tooth neuropathy to D19S586 to D19S432. Neurogenetics 4:83-85.
97.	Züchner S, Noureddine M, Kennerson M, Verhoeven K, Claeys K, De Jonghe P, Merory J,
	Oliveira SA, Speer MC, Stenger JE, Walizada G, Zhu D, Pericak-Vance MA, Nicholson G,
	Timmerman V, Vance JM (2005) Mutations in the pleckstrin homology domain of dynamin 2
	cause dominant intermediate Charcot-Marie-Tooth disease. Nat Genet 37: 289-294.
98.	Claeys KG, Züchner S, Kennerson M, Berciano J, Garcia A, Verhoeven K, Storey E, Merory JR,
	Bienfait HM, Lammens M, Nelis E, Baets J, De Vriendt E, Berneman ZN, De Veuster I, Vance
	JM, Nicholson G, Timmerman V, De Jonghe P (2009) Phenotypic spectrum of dynamin 2
	mutations in Charcot-Marie-Tooth neuropathy. Brain 132: 1741-1752.
99.	Gallardo E, Claeys KG, Nelis E, García A, Canga A, Combarros O, Timmerman V, De Jonghe
	P, Berciano J (2008) Magnetic resonance imaging findings of leg musculature in Charcot-Marie-
	Tooth disease type 2 due to dynamin 2 mutation. J Neurol 255: 986-992.

	100.Fabrizi GM, Ferrarini M, Cavallaro T, Cabrini I, Cerini R, Bertolasi L, Rizzuto N (2007) Two
1	novel mutations in dynamin-2 cause axonal Charcot-Marie-Tooth disease. Neurology 69: 291-
2	295
3 4	101 Ditaur M. Stailauis T. Drudhan D. Mauraca C.A. Lataur D. Varmarah D. Guishanau D. (2009) A
5	101.Bhoun M, Stojković I, Fludnon B, Maurage CA, Latour P, Vermersch P, Guicheney P (2008) A
6	novel mutation in the dynamin 2 gene in a Charcot-Marie-Tooth type 2 patient: clinical and
/	pathological findings. Neuromuscul Disord 18: 334-338.
9	102. Jordanova A, Thomas FP, Guergueltcheva V, Tournev I, Gondim FA, Ishpekova B, De Vriendt
10	E, Jacobs A, Litvinenko I, Ivanova N, Buzhov B, De Jonghe P, Kremensky I, Timmerman V
12	(2003) Dominant intermediate Charcot-Marie-Tooth type C maps to chromosome 1p34-p35. Am
13	I Hum Genet 73: 1423-1430
14	103 Jordanova A. Jrohi I. Thomas FP. Van Dijck P. Maarschaart K. Dawil M. Diarick I. Jacobs A.
16	D M : KE C . KI W D CN T . L C . K DANK M, DIELK I, JACOB A,
17	De Vriendt E, Guergueltcheva V, Rao CV, Tournev I, Gondim FA, D'Hooghe M, Van Gerwen
18	V, Callaerts P, Van Den Bosch L, Timmermans JP, Robberecht W, Gettemans J, Thevelein JM,
20	De Jonghe P, Kremensky I, Timmerman V (2006). Disrupted function and axonal distribution of
21	mutant tyrosyl-tRNA synthetase in dominant intermediate Charcot-Marie-Tooth neuropathy. Nat
22	Genet 38: 197-202.
24	104. Oprescu SN, Griffin LB, Beg AA, Antonellis A (2017) Predicting the pathogenicity of
25	aminoacyl-tRNA synthetase mutations. Methods 113: 139-151.
26 27	105 Storkebaum F. Leitão-Goncalves R. Godenschwege T. Nangle I. Meija M. Bosmans I. Ooms T.
28	Looks A. Von Dijak D. Vong VI. Schimmel D. Narge V. Timmermen V. Cellearte D. Jordenova
29	Jacobs A, Van Dijck P, Yang XL, Schimmel P, Norga K, Timmerman V, Canaens P, Jordanova
30 31	A (2009) Dominant mutations in the tyrosyl-tRNA synthetase gene recapitulate in Drosophila
32	features of human Charcot-Marie-Tooth neuropathy. Proc Natl Acad Sci U S A 106: 11782-
33	11787.
34 35	106. Thomas FP, Guergueltcheva V, Gondim FA, Tournev I, Rao CV, Ishpekova B, Kinsella LJ, Pan
36	Y, Geller TJ, Litvinenko I, De Jonghe P, Scherer SS, Jordanova A (2016) Clinical,
37	neurophysiological and morphological study of dominant intermediate Charcot-Marie-Tooth
39	type C neuropathy I Neurol 263: 467-476
40	107 Mastaclia El Naviale KI Stall D. Deilling DA. Edmandaton IE. Danage SM. Wilton SD.
41 42	107. Mastagna FL, Nowak KJ, Stein R, Phillips BA, Edmondston JE, Dorosz SM, witten SD,
43	Hallmayer J, Kakulas BA, Laing NG (1999) Novel mutation in the myelin protein zero gene in a
44	family with intermediate hereditary motor and sensory neuropathy. J Neurol Neurosurg
45 46	Psychiatry 67: 174-179.
47	108. Sowden JE, Logigian EL, Malik K, Herrmann DN (2005) Genotype-phenotype correlation in a
48	family with late onset CMT and an MPZ lys236del mutation. J Neurol Neurosurg Psychiatry; 76:
49 50	442-444.
51	109 Banchs I. Casasnovas C. Montero J. Volpini V. Martínez-Matos JA (2010) Charcot-Marie-Tooth
52 53	disease with intermediate conduction velocities caused by a novel mutation in the MPZ gene
54	Mugala Namua 42, 194, 199
55	Muscle Nerve 42. 184-188.
56 57	110. Ramirez JD, Barnes PR, Mills KR, Bennett DL (2012) Intermediate Charcot-Marie-Tooth
58	disease due to a novel Trp101Stop myelin protein zero mutation associated with debilitating
59	neuropathic pain. Pain 153: 1763-1768.
6U 61	
62	
63 64	
65	

	111.Boyer O, Nevo F, Plaisier E, Funalot B, Gribouval O, Benoit G, Huynh Cong E, Arrondel C,
1	Tête MJ, Montjean R, Richard L, Karras A, Pouteil-Noble C, Balafrej L, Bonnardeaux A,
2	Canaud G. Charasse C. Dantal J. Deschenes G. Deteix P. Dubourg O. Petiot P. Pouthier D.
4	Laguarn E. Guiachan Mantal A. Brautin I. Gublar MC. Sauniar S. Banaa B. Vallet IM. Alansa
5	Leguern E, Guidenon-Mainer A, Broutin I, Gubier MC, Saumer S, Konco F, Vanat JM, Alonso
6	MA, Antignac C, Mollet G (2011) INF2 mutations in Charcot-Marie-Tooth disease with
/	glomerulopathy. N Engl J Med 365: 2377-2388.
9	112. Mademan I, Deconinck T, Dinopoulos A, Voit T, Schara U, Devriendt K, Meijers B, Lerut E, De
10	Jonghe P, Baets J (2013) De novo INF2 mutations expand the genetic spectrum of hereditary
11 12	neuropathy with glomerulopathy. Neurology 81: 1953-1958.
13	113 Mathis S. Funalot B. Boyer O. Lacroix C. Marcorelles P. Magy I. Richard I. Antignac C. Vallat
14	IN (2014) Nouronothalagia characterization of INE2 related Charact Maria Tagth diagona
15 16	JM (2014) Neuropathologic characterization of INF2-related Charcot-Marie-Tooth disease.
17	evidence for a Schwann cell actinopathy. J Neuropathol Exp Neurol 73: 223-233.
18	114.Park HJ, Kim HJ, Hong YB, Nam SH, Chung KW, Choi BO (2014) A novel INF2 mutation in a
20	Korean family with autosomal dominant intermediate Charcot-Marie-Tooth disease and focal
21	segmental glomerulosclerosis. J Peripher Nerv Syst 19: 175-179.
22	115.Roos A, Weis J, Korinthenberg R, Fehrenbach H, Häusler M, Züchner S, Mache C, Hubmann H,
23	Auer-Grumbach M. Senderek J (2015) Inverted formin 2-related Charcot-Marie-Tooth disease:
25	extension of the mutational spectrum and pathological findings in Schwann cells and avons. I
26	Description Norma System 20, 52, 50
28	Peripher Nerv Syst 20: 52-59.
29	116.Jin S, Wang W, Wang R, Lv H, Zhang W, Wang Z, Jiao J, Yuan Y (2015) INF2 mutations
30 31	associated with dominant inherited intermediate Charcot-Marie-Tooth neuropathy with focal
32	segmental glomerulosclerosis in two Chinese patients. Clin Neuropathol 34: 275-281.
33	117.Lee YC, Lee TC, Lin KP, Lin MW, Chang MH, Soong BW(2010) Clinical characterization and
34	genetic analysis of a possible novel type of dominant Charcot-Marie-Tooth disease.
36	Neuromuscul Disord 20: 534-539
37	118 Soong RW Huang VH Tsai PC Huang CC Pan HC Lu VC Chien HL Liu TT Chang MH
38 39	Lin KD, Tra DL, Kas LG, Las XC (2012) Essent as many identifies CNDA matations as a
40	Lin KP, Tu PH, Kao LS, Lee YC (2013) Exome sequencing identifies GNB4 mutations as a
41	cause of dominant intermediate Charcot-Marie-Tooth disease. Am J Hum Genet 92: 422-430.
42	119. Laššuthová P, Šafka Brožková D, Neupauerová J, Krůtová M, Mazanec R, Seeman P (2017)
44	Confirmation of the GNB4 gene as causal for Charcot-Marie-Tooth disease by a novel de novo
45	mutation in a Czech patient. Neuromuscul Disord 27: 57-60.
47	120.Perrot R, Berges R, Bocquet A, Eyer J (2008) Review of the multiple aspects of neurofilament
48	functions, and their possible contribution to neurodegeneration. Mol Neurobiol 38: 27-65.
49 50	121 Liem RK Messing A (2009) Dysfunctions of neuronal and glial intermediate filaments in
51	diagona L Clin Invest 110: 1914 1924
52	disease. J Chill Hivest 117. 1814-1824.
53 54	122.Perez-Olle R, Leung CL, Liem RK (2002) Effects of Charcot-Marie- I ooth-linked mutations of
55	the neurofilament light subunit on intermediate filament formation. J Cell Sci 115: 4937-4946.
56	123.Züchner S, Vorgerd M, Sindern E, Schröder JM (2004) The novel neurofilament light (NEFL)
57	mutation Glu397Lys is associated with a clinically and morphologically heterogeneous type of
59	Charcot-Marie-Tooth neuropathy. Neuromuscul Disord 14: 147-157.
60 61	
o⊥ 62	
63	
64 65	
CO	

	124. Miltenberger-Miltenyi G, Janecke AR, Wanschitz JV, Timmerman V, Windpassinger C, Auer-
1	Grumbach M, Löscher WN (2007) Clinical and electrophysiological features in Charcot-Marie-
2	Tooth disease with mutations in the NEFL gene. Arch Neurol 64: 966-970.
3	125 Lin KD Soong DW Vang CC, Huang LW, Chang MH, Loo HL Antonollis A, Loo VC (2011)
5	125.Lin KF, Soong BW, Tang CC, Huang LW, Chang WH, Lee HI, Antonenis A, Lee TC (2011)
6	The mutational spectrum in a cohort of Charcot-Marie-Tooth disease type 2 among the Han
/	Chinese in Taiwan. PLoS One 6: e29393.
9	126. Yang Y, Gu LQ, Burnette WB, Li J (2016) N98S mutation in NEFL gene is dominantly
10	inherited with a phenotype of polyneuropathy and cerebellar atrophy. J Neurol Sci 365: 46-47.
11	127 Lawson VH, Graham BV, Flanigan KM (2005) Clinical and electrophysiologic features of
13	CMT2A with mutations in the mitafusin 2 game. Neurology 65: 197-204
14	120 Markannan K. Charre K.C. Züchner C. Caleridar D. Weis I. Contarials C. Landenson A. Malia F.
15 16	128. vernoeven K, Claeys KG, Zuchner S, Schröder JM, weis J, Ceuterick C, Jordanova A, Nells E,
17	De Vriendt E, Van Hul M, Seeman P, Mazanec R, Saifi GM, Szigeti K, Mancias P, Butler IJ,
18	Kochanski A, Ryniewicz B, De Bleecker J, Van den Bergh P, Verellen C, Van Coster R,
19 20	Goemans N, Auer-Grumbach M, Robberecht W, Milic Rasic V, Nevo Y, Tournev I,
21	Guergueltcheva V, Roelens F, Vieregge P, Vinci P, Moreno MT, Christen HJ, Shy ME, Lupski
22	JR. Vance JM. De Jonghe P. Timmerman V (2006) MFN2 mutation distribution and
23	genotype/nhenotype correlation in Charcot-Marie-Tooth type 2 Brain 129: 2093-2102
25	120 Desether CL Send IC Lebets A. Henry H. Deserli MD (2010) MEN2 a sist mutation accurate
26	129. Braatnen GJ, Sand JC, Lobato A, Høyer H, Russell MB (2010) MFN2 point mutations occur in
27	3.4% of Charcot-Marie-Tooth families. An investigation of 232 Norwegian CMT families. BMC
29	Med Genet 11: 48.
30	130. Latour P, Thauvin-Robinet C, Baudelet-Méry C, Soichot P, Cusin V, Faivre L, Locatelli MC,
31 32	Mayençon M, Sarcey A, Broussolle E, Camu W, David A, Rousson R (2010) A major
33	determinant for binding and aminoacylation of tRNA(Ala) in cytoplasmic Alanyl-tRNA
34	synthetase is mutated in dominant avonal Charcot-Marie-Tooth disease. Am I Hum Genet 86:
35 36	
37	
38	131.Bansagi B, Antoniadi T, Burton-Jones S, Murphy SM, McHugh J, Alexander M, Wells R,
39 40	Davies J, Hilton-Jones D, Lochmüller H, Chinnery P, Horvath R (2015) Genotype/phenotype
41	correlations in AARS-related neuropathy in a cohort of patients from the United Kingdom and
42	Ireland. J Neurol 262: 1899-1908.
43 44	132. Spaans F, Jennekens FG, Mirandolle JF, Bijlsma JB, de Gast GC (1986) Myotonic dystrophy
45	associated with hereditary motor and sensory neuropathy. Brain 109: 1149-1168
46	133 Spears E Eaber CG Smeets HI Hofman DA Braida C Monekton DG de Die Smulders CE
47 48	
49	(2009) Encephalopathic attacks in a family co-segregating myotonic dystrophy type 1, an
50	intermediate Charcot-Marie-Tooth neuropathy and early hearing loss. J Neurol Neurosurg
51 52	Psychiatry 80: 1029-1035.
53	134.Brunner HG, Spaans F, Smeets HJ, Coerwinkel-Driessen M, Hulsebos T, Wieringa B, Ropers
54 55	HH (1991) Genetic linkage with chromosome 19 but not chromosome 17 in a family with
56	myotonic dystrophy associated with hereditary motor and sensory neuronathy. Neurology 41
57	80-84
58	00-0 - .
59 60	
61	
62 62	
ьз 64	
65	

	135.LaMonte BH, Wallace KE, Holloway BA, Shelly SS, Ascaño J, Tokito M, Van Winkle T,
1	Howland DS, Holzbaur EL (2002) Disruption of dynein/dynactin inhibits axonal transport in
2	motor neurons causing late-onset progressive degeneration Neuron 34: 715-727
3	12(Denother CL Harver H. Denle CL. Tenders K. Chielbard CE. Denoull MD (2016) Mariante in the
5	136. Braatnen GJ, Høyer H, Busk ØL, Tveten K, Skjelbred CF, Russell MB (2016) variants in the
6	genes DCTN2, DNAH10, LRIG3, and MYO1A are associated with intermediate Charcot-Marie-
7	Tooth disease in a Norwegian family. Acta Neurol Scand 134: 67-75.
o 9	137. Sevilla T, Sivera R, Martínez-Rubio D, Lupo V, Chumillas MJ, Calpena E, Dopazo J, Vílchez
10	II Palau F Espinós C (2015) The EGR2 gene is involved in axonal Charcot-Marie-Tooth
11	disance Fur I Neural 22: 1549 1555
13	disease. Eur J Neuror 22, 1546-1555.
14	138.Solla P, Vannelli A, Bolino A, Marrosu G, Coviello S, Murru MR, Tranquilli S, Corongiu D,
15	Benedetti S, Marrosu MG (2010) Heat shock protein 27 R127W mutation: evidence of a
16	continuum between axonal Charcot-Marie-Tooth and distal hereditary motor neuropathy. J
18	Neurol Neurosurg Psychiatry 81: 958-962.
19	139 Baxter RV Ben Othmane K. Rochelle IM. Staijch IF. Hulette C. Dew-Knight S. Hentati F. Ben
20	Hamida M. Dal S. Stanzan IE. Cilbart ID. Dariaak Varias MA Varias IM (2002) Canaliasida
22	Hamida M, Bei S, Stenger JE, Gilbert JK, Pericak-Vance MA, Vance JM (2002) Ganglioside-
23	induced differentiation-associated protein-1 is mutant in Charcot-Marie-Tooth disease type
24	4A/8q21. Nat Genet 30: 21-22.
25 26	140. Cuesta A, Pedrola L, Sevilla T, García-Planells J, Chumillas MJ, Mayordomo F, LeGuern E,
27	Marín I. Vílchez JJ. Palau F (2002) The gene encoding ganglioside-induced differentiation-
28	associated protein 1 is mutated in avonal Charcot-Marie-Tooth type 44 disease. Nat Genet 30
29	
31	22-25.
32	141. Claramunt R, Pedrola L, Sevilla T, López de Munain A, Berciano J, Cuesta A, Sánchez-Navarro
33	B, Millán JM, Saifi GM, Lupski JR, Vílchez JJ, Espinós C, Palau F (2005) Genetics of Charcot-
34 35	Marie-Tooth disease type 4A: mutations, inheritance, phenotypic variability, and founder effect.
36	J Med Genet 42: 358-365
37	1/2 Crimella C. Tonelli A. Airoldi G. Baschirotto C. D'Angelo MG. Bonato S. Losito I. Trabacca A
38 39	Prezidente en la contra
40	Bresolin N, Bassi MT (2010) The GST domain of GDAP1 is a frequent target of mutations in
41	the dominant form of axonal Charcot Marie Tooth type 2K. J Med Genet 47: 712-716.
42	143. Sivera R, Espinós C, Vílchez JJ, Mas F, Martínez-Rubio D, Chumillas MJ, Mayordomo F,
44	Muelas N, Bataller L, Palau F, Sevilla T (2010) Phenotypical features of the p.R120W mutation
45	in the GDAP1 gene causing autosomal dominant Charcot-Marie-Tooth disease. J Peripher Nerv
46	Synt 15: 224 244
47	
49	144.Zimon M, Baets J, Fabrizi GM, Jaakkola E, Kabzińska D, Pilch J, Schindler AB, Cornblath DR,
50	Fischbeck KH, Auer-Grumbach M, Guelly C, Huber N, De Vriendt E, Timmerman V, Suter U,
51 52	Hausmanowa-Petrusewicz I, Niemann A, Kochański A, De Jonghe P, Jordanova A (2011)
53	Dominant GDAP1 mutations cause predominantly mild CMT phenotypes. Neurology 77: 540-
54	548
55 56	145 Soville T. Cueste A. Chumilles MI. Mayordome E. Dedrole I. Deley F. Mileher, H. (2002)
57	145. Sevina I, Cuesta A, Chummas IVIJ, Mayordomo F, Pedrola L, Palau F, Vilchez JJ (2003).
58	Clinical, electrophysiological and morphological findings of Charcot-Marie-Tooth neuropathy
59	with vocal cord palsy and mutations in the GDAP1 gene. Brain 126: 2023-2033.
6U	
62	
63	
6/	

	146.Sevilla T, Jaijo T, Nauffal D, Collado D, Chumillas MJ, Vilchez JJ, Muelas N, Bataller L,
1	Domenech R, Espinós C, Palau F (2008) Vocal cord paresis and diaphragmatic dysfunction are
2	severe and frequent symptoms of GDAP1 associated neuropathy. Brain 131: 3051 3061
3	severe and nequent symptoms of ODAT 1-associated neuropathy. Brain 151, 5051-5001.
5	147.Nelis E, Erdem S, Van Den Bergh PY, Belpaire-Dethiou MC, Ceuterick C, Van Gerwen V,
6	Cuesta A, Pedrola L, Palau F, Gabreëls-Festen AA, Verellen C, Tan E, Demirci M, Van
7	Broeckhoven C, De Jonghe P, Topaloglu H, Timmerman V (2002) Mutations in GDAP1:
8	autocomal recognize CMT with demvelingtion and evenengthy. Neurology 50: 1865-1872
9	autosoniai recessive CMT with demyennation and axonopathy. Neurology 59, 1805-1872.
10	148.Kabzińska D, Niemann A, Drac H, Huber N, Potulska-Chromik A, Hausmanowa-Petrusewicz I,
12	Suter U, Kochański A (2011) A new missense GDAP1 mutation disturbing targeting to the
13	mitochondrial membrane causes a severe form of AR-CMT2C disease. Neurogenetics 12: 145-
14	153
16	
17	149. Senderek J, Bergmann C, Ramaekers VT, Nelis E, Bernert G, Makowski A, Züchner S, De
18	Jonghe P, Rudnik-Schöneborn S, Zerres K, Schröder JM (2003) Mutations in the ganglioside-
19	induced differentiation-associated protein-1 (GDAP1) gene in intermediate type autosomal
20 21	recessive Charcot-Marie-Tooth neuropathy, Brain 126: 642-649
22	150 CL KW H VOL HLL HK K H V H K OD D L CLK' HDL CL'DO
23	150. Chung KW, Hyun YS, Lee HJ, Jung HK, Koo H, Yoo JH, Kim SB, Park CI, Kim HN, Choi BO
24	(2011) Two recessive intermediate Charcot-Marie-Tooth patients with GDAP1 mutations. J
25 26	Peripher Nerv Syst 16: 143-146.
27	151.McLaughlin HM, Sakaguchi R, Liu C, Igarashi T, Pehlivan D, Chu K, Iver R, Cruz P, Cherukuri
28	DE Hanson NE Mullikin IC: NISC Comparative Sequencing Program Dissocker I.C. Wilson
29	FF, Hansen NF, Munikin JC, NISC Comparative Sequencing Flogram, Diesecker EG, wilson
30 31	TE, Ionasescu V, Nicholson G, Searby C, Talbot K, Vance JM, Züchner S, Szigeti K, Lupski JR,
32	Hou YM, Green ED, Antonellis A (2010) Compound heterozygosity for loss-of-function lysyl-
33	tRNA synthetase mutations in a patient with peripheral neuropathy. Am J Hum Genet 87: 560-
34	566
36	152 Maystadt I. Pozsähazy P. Parkats M. Duguo S. Vannuffal P. Pamaala S. Lambart P. Najimi M
37	152. Maystaut 1, Rezsonazy R, Baikats M, Duque S, Valinutier F, Remacie S, Lamoert B, Najimi M,
38	Sokal E, Munnich A, Viollet L, Verellen-Dumoulin C (2007) The nuclear factor kappaB-
39	activator gene PLEKHG5 is mutated in a form of autosomal recessive lower motor neuron
40	disease with childhood onset. Am J Hum Genet 81: 67-76.
42	153 Azzedine H. Zavadakova P. Planté-Bordeneuve V. Vaz Pato M. Pinto N. Bartesaghi I., Zenker J.
43	Doirot O. Dornard Mariagal N. Arnoud Couttonaira E. Cartoni D. Title A. Vanturini C. Médard
44	Ponot O, Bernard-Marissar N, Arnaud Gouttenone E, Cartoni K, Thie A, Venturni G, Medard
46	JJ, Makowski E, Schöls L, Claeys KG, Stendel C, Roos A, Weis J, Dubourg O, Leal Loureiro J,
47	Stevanin G, Said G, Amato A, Baraban J, LeGuern E, Senderek J, Rivolta C, Chrast R (2013)
48	PLEKHG5 deficiency leads to an intermediate form of autosomal-recessive Charcot-Marie-
49 50	Tooth disease Hum Mol Genet 22: 4224-4232
51	154 Kim HI Hans VD Barla M. Chai VD Kim VI Vara DD Kaa H. Vara HI Kim OD Barla M
52	154.Kim HJ, Hong YB, Park JM, Chol YK, Kim YJ, Yoon BK, Koo H, Yoo JH, Kim SB, Park M,
53	Chung KW, Choi BO (2013) Mutations in the PLEKHG5 gene is relevant with autosomal
54 55	recessive intermediate Charcot-Marie-Tooth disease. Orphanet J Rare Dis 8:104.
56	
57	
58	
59 60	
61	
62	
63 64	
04 65	

155. Tamiya G, Makino S, Hayashi M, Abe A, Numakura C, Ueki M, Tanaka A, Ito C, Toshimori K, Ogawa N, Terashima T, Maegawa H, Yanagisawa D, Tooyama I, Tada M, Onodera O, Hayasaka K (2014) A mutation of COX6A1 causes a recessive axonal or mixed form of Charcot-Marie- Tooth disease. Am L Hum Genet 95: 294-300
 156.Laššuthová P, Beharka R, Krůtová M, Neupauerová J, Seeman P (2016) COX6A1 mutation causes axonal hereditary motor and sensory neuropathy - the confirmation of the primary report. Clin Genet 89: 512-514.

FIGURE LEGENDS

Figure 1. Median MNCV study in a 59-year-old patient belonging to a DI-CMT family associated with *NEFL* E396K mutation [24]. (**A**) The recorded MCV value of 36 m/s (elbow-wrist segment), within the intermediate range, could be attributed to severe CMAP amplitude reduction (to 15% of the LLN, \geq 4 mV), namely, to loss of large myelinated fibres. This question is clarified here analysing the segment axilla-elbow, with stimulation at axilla and elbow, and recording from flexor digitorum sublimis (FDS) (**B**). Note that despite CMAP amplitudes being preserved, the recorded MCV, 44 m/s (normal, \geq 54), is in the intermediate range.

Figure 2. Clinical pictures of the proband (**A-C**), aged 55 years, and her younger affected daughter (**D**, **E**), aged 23 years, suffering from axonal CMT2 associated with *DNM2* Gly358Arg mutation [99]. These patients are identified as CMT-103/I.2 and CMT-103/II.3 in the series by Claeys and colleagues [98]. In the proband note marked lower-leg amyotrophy (**A**) and hand wasting involving first dorsal interossei (**B**) and thenar eminences (**C**). Her daughter shows preserved hand musculature (**D**), but evident lower-leg amyotrophy. MNCV study revealed absence of responses in tibial and peroneal nerves, median MNCV study being illustrated in figure 3.

Figure 3. Median MNCV study in three patients from a pedigree with CMT2 associated with *DNM2* Gly358Arg mutation (see figure 2). (A) In the 55-year-old proband patient showing marked hand amyotrophy, CMAPs are severely attenuated in APB muscles, elbow-wrist conduction velocity being 33 m/s. As argued in figure 1, one may wonder whether so marked motor conduction slowing could be accounted for by axonal loss or

demyelination. To clarify the issue we recorded median MNCV in the segment axillaelbow with recording in FDS. The recorded CMAPs are now of greater amplitude but still clearly attenuated, so the recorded MNCV, 44 m/s, might be interpreted either intermediate or secondary to axonal loss. Electrophysiological evaluation of her two affected daughters, not showing hand amyotrophy, is crucial in this regard. Indeed, median MNCV study in elder affected daughter (**B**), aged 32 years, and younger affected daughter (**C**; see figure 2D, E) shows almost normal conduction parameters, the usual electrophysiologic hallmark of uninvolved nerves in axonal CMT.

Figure 4. Median MNCV study in a 31-year-old patient belonging to a DI-CMT family associated to *NEFL* N98S mutation [20]. (**A**) The recorded MNCV value of 18.3 m/s (elbow-wrist segment; normal, \geq 49), within the demyelinating range, could be attributed greatly to severe CMAP amplitude reduction in abductor pollicis brevis (APB) (to 5% of the LLN), namely, to presumptive complete loss of large myelinated fibres. (**B**) This question is again clarified analyzing the segment axilla-elbow, with stimulation at axilla and elbow, and recording in flexor digitorum sublimis (FDS). Note that despite CMAP amplitudes being relatively preserved, recorded MNCV, 47.5 m/s (normal, \geq 55), is in the intermediate range. As argued in the text, proximal MNCV study might be essential for accurately defining the CMT pattern of conduction slowing.

Figure 5. An algorithm for electrophysiological approach in CMT focused on the intermediate form of the disease. For further details, see also figures 1, 3 and 4. For abbreviations see text.











Figure



Figure

Table			

Table 1. Classification of in	termediate CMT		
Transmission and types	Gene	Loci	OMIM number
<u>X-linked</u>			
CMTX1	GJB1	Xq13.1	302800
DRP2-disorder	DRP2	Xq22.1	Not yet assigned
Autosomal dominant			
DI-CMTA*	Not cloned	10q24.1	Not cloned
DI-CMTB	DNM2	19p13.2	606482
DI-CMTC	YARS	1p35.1	608323
DI-CMTD	MPZ	1q23.3	607791
DI-CMTE	INF2	14q32.33	614455
DI-CMTF	GNB4	3q26.33	615185
Other dominant types			
DI-CMT/NEFL	NEFL	8p21.2	Not yet assigned
DI-CMT/MNF2	MNF2	1p36.22	Not yet assigned
DI-CMT/AARS	AARS	16q22.1	Not yet assigned
DI-CMT/MD1	Not cloned	19q13.32	Not yet assigned
DI-CMT/DCTN2	DCTN2	12q13.3	Not yet assigned
Autosomal recessive			
RI-CMTA	GDAP1	8q21.11	608340
RI-CMTB*	KARS	16q23.1	613641
RI-CMTC	PLEKHG5	1p36.31	615376
RI-CMTD*	COX6M	12q24.31	616039

(*) Phenotypes that could be classified within CMT2 or AR-CMT2

Table 2. Median ner	rve motor conduction par	rameters in CMTX1 m	ales		
Reference	No. patients electrophysiologically evaluated	MNCV m/s Range (Mean ± SD)	CMAP mV Range (Mean ± SD)	DML ms Range (Mean ± SD)	Comments
Phillips <i>et al</i> ⁴⁴	9	NR	NR	NR	Ulnar MNCV ranged from 33.1 to 44.7 (mean 38). Slowing was in parallel with loss of CMAP amplitude. Ulnar MNCV in 6 obligate heterozygous women ranged from 43.7 to 56.4 (mean 49.4)
Rozear <i>et al</i> ³⁷	4	28 - 37 (34.5)	NR	NR	Median MNCV in 6 obligate heterozygous women ranged from 38 to 53 (mean 50)
Hahn <i>et al</i> ³⁸	See comments	NR	NR	NR	Peroneal MNCV showed abnormalities in 28/31 cases (males and females). Modest reduction in MNCV paralleled changes in CMAP amplitudes for EDB and anterolateral leg muscles. Electrophysiology was normal in one affected male at age 3, who later on developed signs of the disease
Mostacciuolo et al ⁵⁸	41	NR	NR	NR	Mean ulnar MNCVs were 26 ± 5.9 in males and 38 m/s in females
Ionasescu <i>et al⁵⁹</i>	6	NR	NR	NR	Electrophysiologic findings were consistent with both demyelination (slowing) and axonal denervation in all male patients
Nicholson and Nash ⁶⁰	15	(30.8 ± 5.9)	NR	NR	In comparison with CMT1A, mean median MNCVs were faster with an average of 10 m/s for males and 20 m/s for females
Oterino <i>et al</i> ⁶¹	7	0 - 40 (24.2)	NR	NR	Only 2 out of 7 male patients remained asymptomatic but areflexic
Ionasescu <i>et al</i> ⁶²	6	NR	NR	NR	Ibid Ionasescu (1992). Prolonged DMLs (6-7 ms) were present. The CMT obligate female carriers showed normal electrophysiology
Birouk <i>et al</i> ⁶³	21	31 - 60 (36.2 ± 6.7)	0.3 - 6.3 (2.6 ± 1.8)	2.3 - 7 4.7 ± 1.1)	In 90% of CMTX1 males, median MNCV was intermediate ranging from 30 to 40 m/s. Only one male had normal MNCV. There was significant correlation between the decrease in CMAP amplitude and MNCV in the median, ulnar and peroneal nerves. In 27 obligate female carriers, median MNCVs ranged between 31 and 55 m/s (44 ± 7.3). Two children with the mutation, a 6-year-old boy and a 8-year-old girl, were normal clinically and electrophysiologically
Ainsworth et al ⁶⁴	1	NR	NR	NR	Ulnar MNCV was 34.8 m/s; CMAP was 3.83 mV
Sander <i>et al</i> ⁶⁵	4	25 – 40 (32)	NR	NR	Median MNCV in a female patient was 30 m/s
Senderek <i>et al</i> ⁵²	9	27.7 – 33.3 (31)	Reduced	NR	Median MNCV in 9 female patients ranged from 35 to 49 m/s (mean, 40)

Table 2. Median ne	erve motor conduction par	ameters in CMTX1 ma	ales (cont. 1)		
Reference	No. patients electrophysiologically evaluated	MNCV m/s Range (Mean ± SD)	CMAP mV Range (Mean ± SD)	DML ms Range (Mean ± SD)	Comments
Hahn <i>et at</i> ⁴⁰	53	23 - 50 (34.5 ± 6.1)	0-12 (3.7 ± 3.7)	3 - 12 (5.4 ± 1.8)	Ten (34%) male patients showed CMAP amplitude ≤ 0.5 mV. F-wave latencies ranged between 38 and 53 ms (mean, 47). In 63 affected females, median MNCV ranged between 33 and 60 m/s (mean 45.8 \pm 7.3); CMAP amplitudes ranged from 2 to 17 mV (7.8 \pm 3.4); DML ranged from 3 to 5 ms (4.1 \pm 0.6); and F-waves ranged from 30 to 36 ms (mean, 33)
Hahn <i>et al</i> ⁵³	_	27.9	4.9	4.8	This pedigree comprises two male patients. Ulnar MNCVs were 32.3 and 38.0 m/s; CMAPs were 3.8 and 6.6 mV; and DML were 4.5 and 5.0 ms. F-wave in one case was 42.8 ms
Gutiérrez <i>et al</i> ⁶⁶	6	25 - 54	0.045 - 2.2	3.8 - 6	F-wave latency recorded in one case was 32.1 ms. Terminal latency index was preserved. Ulnar MNCV ranged between 27 and 39 m/s
Dupré <i>et al</i> ⁶⁷	Q	22 - 33 (29.0 ± 4.8)	0.3 - 6 (2.0 ± 2.2)	6.6 - 7.7 (7.0 ± 0.9)	One male patient showed no response. In six female patients median MNCV ranged from 41 to 57 m/s (49 ± 6). Three patients with reduced median MNCV had normal CMAP amplitudes
Nakagawa <i>et al</i> ⁶⁸	0	26.5;30.1	0.1;3.6	5.7;5.5	F-wave latency, recorded in one case, was 48 ms. Ulhar MNCVs were 33.3 / 33.2; CMAP amplitudes were 4.0 / 3.4; DMLs were 4.5 / 5.0; and F-wave latencies were 48.2 / 47.8
Dubourg <i>et al</i> ⁶⁹	41	20 - 60 (34.5 ± 6.8)	0 - 8.6 (2.2 ± 2.1)	2.3 – 8.8 (4.9 ± 1.2)	In five male patients there was no response. In this comprehensive study the authors analyzed ulnar and peroneal MNCV, and included 52 female patients. In males mean MNCV in all three nerves ranged between 30 to 40 m/s, but were slightly below normal values in females. MNCV below 30 m/s were found in 18% of men but only in 5% of women
Hattori <i>et al</i> ⁷⁰	42	22.8 - 46.6 (33.2 ± 5.7)	0 - 8.1 (2.0 ± 1.8)	NR (5.3 ± 1.7)	Median MNCVs were discordant in terms of the 38 m/s cut-off value among siblings in the six families examined. CMAP amplitude significantly decreased with age and duration of illness
Capasso <i>et al</i> ⁷¹	0	23; 31	0.8;1.4	5.7; 6.2	The authors also studied ulnar MNCVs that were 36 m/s and 30 m/s; CMAP amplitudes were 5.3 and 5.2 mV; and DML were 3.0 and 3.7 ms. They considered that ulnar MNCV slowing with normal or relatively preserved CMAP amplitudes point to a demyelinating disorder
Houlden <i>et al</i> ⁷²	See comments	See comments	See comments	See comments	In male patients (5 according to their figure 1), ulnar MNCV ranged from 28 to 36 m/s (mean 32 ± 4.0), whereas in affected females (11) ranged from 34 to 55 m/s (46 ± 2.9). No data on CMAP amplitudes or DML are given

Table 2. Median ne	rve motor conduction par	ameters in CMTX1 n	nales (cont. 2)		
Reference	No. patients electrophysiologically	MNCV m/s Range	CMAP mV Range	DML ms Range	Comments
	evaluated	$(Mean \pm SD)$	$(Mean \pm SD)$	$(Mean \pm SD)$	
Ryan and Jones ⁷³		38	3.87	5.6	Tabulated data are those of right median nerve. Left median MNCV was 41 m/s; CMAP was 7.55 mV, and DML was 3.9 ms, In this 13 year-old patient, there was response to immunoglobulin therapy
Vondracek et al ⁷⁴	Π	33	5.9	4.8	Median MNCVs in the intermediate range were also found in two affected women
Liang <i>et al</i> ⁷⁵	0	0	0	0	The authors describe a 13-year-old girl with severe Dejerine-Sottas phenotype and $MNCV$ of 18-20 m/s
Zhang <i>et al</i> ⁷⁶	20	10.7 - 45	NR	NR	In 18 affected females median MNCV ranged between 23 and 61 m/s
Huttner <i>et al</i> ⁷⁷	11	25 - 57 (42.1 ± 9.4)	0.5 - 11 (4.1 ± 3.1)	3.0 - 6.6 (4.9 ± 9.4)	There were no responses in one patient
Kleopa <i>et al⁵⁵</i>	e	23; 38	0.19;5.7	7.2 ,4.2	In one male patient median nerve showed no responses. Ulnar MNCV in all three patients were 32, 38, 38 m/s with CMAP of 0.87, 2.31 and 6.5 mV, respectively
Beauvais <i>et al</i> ⁷⁸	1	34	5.3	NR	One affected woman showed median MNCV of 34 m/s and CMAP of 5.3 mV
Vazza et al ⁷⁹	7	36;31	0.3;1.3	NR ; 5.9	One affected woman showed median MNCV of 38 m/s and CMAP of 8 mV
Shy <i>et al</i> ⁴³	73	See comments	See comments	See comments	This comprehensive study demonstrates important issues: i/ in all patients disability increased with age and loss of motor units; ii/ ulnar MNCVs (wrist-elbow) were predominantly in the intermediate range (30-50 m/s); iii/ occasional patients had forearm ulnar MNCV < 30 m/s, which was always associated with markedly reduced CMAP amplitudes; and iv/ most <i>GJB1</i> mutations cause neuropathty by a loss of normal connexin 32 function
Kim <i>et al</i> ⁸⁰	1	See comments	See comments	See comments	Ulnar MNCV was 34.8 m/s; CMAP was 4.1 mV; and DML was 3.9 ms
Vrancken <i>et al</i> ⁸¹	0	33;33	4.8;7.9	4.7;4.5	Ulnar MNCV showed similar parameters. In an affected woman median MNCV was 53 m/s; CMAP amplitude was 7.3 mV; and DML was 4.0 ms
Sakaguchi <i>et al</i> ⁸²	1	36.9	0.59	5.8	Ulnar nerve MNCV was 37 m/s with CMAP of 2.1 mV, and DML of 3.8 m/s. There was response to immunoglobulin therapy

Table 2. Median ne	rve motor conduction par	ameters in CMTX1 m	ales (cont. 3)		
Reference	No. patients electrophysiologically evaluated	MNCV m/s Range (Mean ± SD)	CMAP mV Range (Mean ± SD)	DML ms Range (Mean ± SD)	Comments
Yiu <i>et al</i> ⁸³	3 See comment	30;40	NR	4.5	Electrophysiology is complemented by ulnar, peroneal and tibial MNCV. Intermediate MNCV, particularly in combination with axonal features, should raise the possibility of CMTX
Murphy <i>et al</i> ⁸⁴	ς	32 – 42 (34.7)	0.5 - 0.9	NR	Median MNCV in three females ranged from 32 to 53.7 m/s (mean 44.9). Ulnar nerve was also evaluated
Chen <i>et al</i> ⁸⁵	7	32.8 ,28.5	3.5;0.2	4.7;3.5	F-wave latency in one patient was 43.8 ms. Ulnar nerve was also evaluated. In a female patient, median MNCV was 43.9 m/s with CMAP of 0.9 mV
Borgulová <i>et al</i> ⁸⁶	_	43.9	2.2	4.85	In another male patient peroneal MNCV was 39.8 m/s with CMAP of 3.3 mV. In three female patients median MNCV ranged between 40 and 49 m/s, CMAPs being normal or slightly reduced
Zhao <i>et al</i> ⁸⁷	1	33	7.7	2.7	In this 15-year-old patient contralateral median MNCV was 40 m/s with CMAP of 1.7 mV
Jerath <i>et al</i> ⁴⁵	20	21 - 44 (35 ± 1.5)	3 ± 0.6	5 ± 0.3	Median MNCV were normal in 4 (20%) male patients, intermediate (33-41 m/s) in 8 (40%), and slow (< 32 m/s) in 8 (40%)
Liu et al ⁸⁸	S	29.1 – 37.2	0.1 - 17.2	NR	Median MNCV were from 29.1 to 40 m/s in 5 out of the 31 proband male patients. CMAP amplitudes ranging from 0.1 to 17.2 mV. This comprehensive study comprises 226 CMT pedigrees, 31 (13.7%) being categorized as CMTX1, representing 65% of all intermediate CMT

NR= not recorded; SD= standard deviation; for the remaining abbreviations see text.