

University of Pennsylvania ScholarlyCommons

Departmental Papers (EES)

Department of Earth and Environmental Science

July 2005

Intertidal Mangrove Foraminifera From The Central Great Barrier Reef Shelf, Australia: Implications for Sea-Level Reconstruction

Sarah A. Woodroffe University of Durham

Benjamin P. Horton University of Pennsylvania, bphorton@sas.upenn.edu

Piers Larcombe Centre for Environment, Fisheries and Aquaculture Science

John E. Whittaker Natural History Museum

Follow this and additional works at: https://repository.upenn.edu/ees_papers

Recommended Citation

Woodroffe, S. A., Horton, B. P., Larcombe, P., & Whittaker, J. E. (2005). Intertidal Mangrove Foraminifera From The Central Great Barrier Reef Shelf, Australia: Implications for Sea-Level Reconstruction. Retrieved from https://repository.upenn.edu/ees_papers/47

Published in Journal of Foraminiferal Research, Volume 35, Number 3, July 2005, pages 259–270.

This paper is posted at ScholarlyCommons. https://repository.upenn.edu/ees_papers/47 For more information, please contact repository@pobox.upenn.edu.

Intertidal Mangrove Foraminifera From The Central Great Barrier Reef Shelf, Australia: Implications for Sea-Level Reconstruction

Abstract

Contemporary foraminiferal samples and environmental information were collected from three fringing mangrove environments (Sandfly Creek Transect 1 and 2, and Cocoa Creek) in Cleveland Bay, and an estuarine mangrove environment (Saunders Creek) in Halifax Bay, on the central Great Barrier Reef (GBR) coastline, Australia, to elucidate the relationship of the foraminiferal assemblages with the environment. The data support the vertical zonation concept, which suggests that the distribution of foraminifera in the intertidal zone is usually a direct function of elevation, with the duration and frequency of subaerial exposure as the most important factor. An agglutinated foraminiferal assemblage dominated by *Miliammina fusca, Trochammina inflata, Ammotium directum* and *Haplophragmoides* sp. exists at the landward edge of the field sites, in a zone between just above Mean Low Water of Neap Tides to Highest Astronomical Tide level (a vertical range of 1.8 m). In addition, a foraminiferal assemblage dominated by *Ammonia aoteana* is found at all sites, existing between just below Mean Low Water of Neap Tides and Mean High Water of Neap Tides (a vertical range of 0.8 m). These assemblages may be used to reconstruct sea level from fossil cores from the area.

Comments

Published in Journal of Foraminiferal Research, Volume 35, Number 3, July 2005, pages 259–270.

INTERTIDAL MANGROVE FORAMINIFERA FROM THE CENTRAL GREAT BARRIER REEF SHELF, AUSTRALIA: IMPLICATIONS FOR SEA-LEVEL RECONSTRUCTION

SARAH A. WOODROFFE¹, BEN P. HORTON², PIERS LARCOMBE³ AND JOHN E. WHITTAKER⁴

ABSTRACT

Contemporary foraminiferal samples and environmental information were collected from three fringing mangrove environments (Sandfly Creek Transect 1 and 2, and Cocoa Creek) in Cleveland Bay, and an estuarine mangrove environment (Saunders Creek) in Halifax Bay, on the central Great Barrier Reef (GBR) coastline, Australia, to elucidate the relationship of the foraminiferal assemblages with the environment. The data support the vertical zonation concept, which suggests that the distribution of foraminifera in the intertidal zone is usually a direct function of elevation, with the duration and frequency of subaerial exposure as the most important factor. An agglutinated foraminiferal assemblage dominated by Miliammina fusca, Trochammina inflata, Ammotium directum and Haplophragmoides sp. exists at the landward edge of the field sites, in a zone between just above Mean Low Water of Neap Tides to Highest Astronomical Tide level (a vertical range of 1.8 m). In addition, a foraminiferal assemblage dominated by Ammonia aoteana is found at all sites, existing between just below Mean Low Water of Neap Tides and Mean High Water of Neap Tides (a vertical range of 0.8 m). These assemblages may be used to reconstruct sea level from fossil cores from the area.

INTRODUCTION

The study of Holocene relative sea-level (RSL) changes in the Great Barrier Reef (GBR) region of Australia is particularly important because of its tectonic stability and its great distance from the centers of former ice caps. Observations of sea-level change in far field locations such as Australia provide the most direct estimate of the volume of grounded ice existing at various time periods between the Last Glacial Maximum and present (Milne and others, 2002). In addition, evidence of a mid-Holocene high stand of approximately +3 m along parts of the east coast of Australia and the nature of subsequent RSL fall is of interest to geophysical modelers because it yields information on the contribution of hydro-isostasy, equatorial ocean siphoning and crustal levering to late Holocene relative sea-level change in tropical areas. This increases our understanding of solid earth geophysics (e.g., Mitrovica and Peltier, 1991; Nakada and Lambeck, 1989; Yokoyama and others, 2001; Lambeck and others, 2002).

One approach to the study of RSL change is to use microfossil sea-level indicators such as foraminifera, diatoms and pollen, which are contained in both contemporary and fossil sedimentary deposits. Sea-level indicators developed from observed changes in microfossil assemblages have been used for the past 30 years or so to provide reconstructions of Holocene RSL change for many areas, including Britain, Europe and North America, and have been the primary source of data for developing and testing models of RSL change (e.g., Shennan and others, 2002; Peltier, 2002; Shennan and Horton, 2002). Marsh foraminifera, in particular, are highly accurate sea-level indicators because they have narrow ecological tolerances and narrow vertical zones in the intertidal zone (Scott and Medioli, 1978, 1980a). However, all microfossil data and their associated RSL reconstructions are subject to errors, which limit the precision of determining both age and elevation, and as smaller magnitude sea-level changes are analyzed, the magnitude of errors becomes increasingly important. To address this limitation we must use the most precise indicators available, and use statistically robust quantitative techniques in reconstructions.

To this end, a new generation of microfossil-based quantitative paleoenvironmental reconstructions has been developed (e.g., Horton, 1999; Zong and Horton, 1999; Edwards and Horton, 2000; Gehrels and others, 2001; Horton and others, 2003), allowing a relatively precise reconstruction of former sea levels, using a statistically-based relationship between contemporary foraminiferal assemblages, their relationships to sea level and their fossil counterparts. This developing research field is generating high precision in RSL reconstructions (Horton and others, 2000; Edwards and Horton, 2000; Horton and Edwards, 2005; Sawai and others, 2004).

In contrast to mid-latitude, temperate environments, there have been relatively few studies of intertidal foraminifera and their relationship to RSL in tropical coastal environments. The studies include Michie (northern Australia, 1987), Scott and others (Brazil, 1990), Barbosa and Suguio (Brazil, 1999), Hayward and others (New Zealand, 1999b, 2004b), Debenay and others (west coast of Africa, New Caledonia and northern Australia, 2000). Haslett (northern Australia, 2001), Debenay and others (French Guiana, 2002), Javaux and Javaux and Scott (Bermuda 1999, 2003), Horton and others (Great Barrier Reef coastline, Australia, 2003), and Horton and others (Indonesia, 2005), Furthermore, most studies concentrate on individual sites and do not investigate the potential for diversity in foraminiferal faunas over local or regional scales. The tropical environment has many different challenges for foraminiferal assemblages, including the widespread presence of bioturbators

¹ Department of Geography, University of Durham, South Road, Durham, DH1 3LE, UK. E-mail: S.A.Woodroffe@durham.ac.uk

² Department of Earth and Environmental Science, University of Pennsylvania, Philadelphia, PA, 19104-6316, USA.

³ Centre for Environment, Fisheries and Aquaculture Science, Pakefield Road, Lowestoft, Suffolk, NR33 0HT, UK.

⁴ Micropalaeontology Research, Department of Palaeontology, The Natural History Museum, London SW7 5BD, UK.

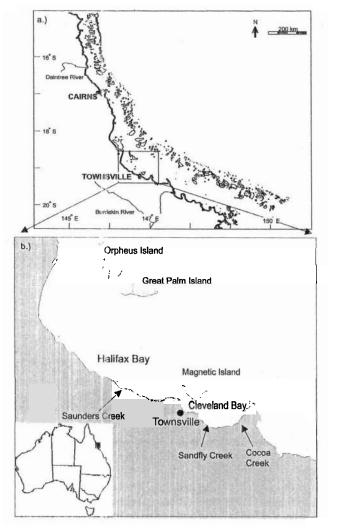


FIGURE 1. Location map of the central Great Barrier Reef showing sites studied. (a) The Great Barrier Reef coastline from Cape Tribulation in the north to the Whitsunday Islands in the south. (b) The central Great Barrier Reef province around Townsville showing the location of the field sites Saunders Creek, Sandfly Creek and Cocoa Creek.

(e.g., fiddler crabs) which disturb the surface sediment, creating potentially increased sources of error when trying to reconstruct former sea levels and post-depositional changes in foraminiferal assemblages due to disaggregation and dissolution. This paper documents the characteristics of modern intertidal environments from four different locations in the central GBR province, and compares and contrasts their foraminiferal faunas, identifying implications for sea-level reconstructions.

STUDY AREAS

The shoreline of the central GBR is characterized by a series of north-facing coastal embayments (e.g., Halifax Bay, Cleveland Bay, Bowling Green Bay), which are protected from the dominant southeast trade wind by granite headlands or sand spits, but are open to northerly and northeasterly weather and the impact of occasional tropical cyclones (Belperio, 1983; Carter and others, 1993). Cleveland Bay lies immediately offshore from Townsville, Queensland, and Halifax Bay lies north of it, approximately 600 km north of the southernmost limit of the GBR (Fig. 1). The bays lie at approximately 19° S 146°--147° 30' E. Cleveland Bay is approximately 20 km square and is landlocked around its southern and eastern margins by the mainland. Within the bay lies Magnetic Island, which is granitic and \sim 12 km in diameter, and rises to an altitude of 495 m. This island shields the northerly part of Cleveland Bay. The southern part of the bay is shielded by the granite headland of Cape Cleveland, which rises to an elevation of 557 m. Halifax Bay is more open, but the granite promontory of Cape Pallarenda also shields the southern half. Cleveland and Halifax Bays are relatively shallow, both reaching a maximum water depth of 15 m at their seaward edges (Carter and others, 1993). Complex water motions occur in Cleveland Bay, including the effects of refracted SE-generated swell waves, and the tidal range for both bays is generally mesotidal (~2.3 m; Carter and others, 1993; Larcombe and others, 1995). Australian Height Datum (AHD) is the local height datum used in this study. Mean Sea Level is 0.1 m above 0 m AHD. Lowest Astronomical Tide (LAT) is -1.86 m AHD, Mean High Water of Spring Tides (MHWST) is +1.21 m AHD, and Highest Astronomical Tide (HAT) is +2.15 m AHD.

COCOA CREEK, CLEVELAND BAY

Cocoa Creek is the southernmost of four tidal creeks entering Cleveland Bay. The main channel meanders for 9.5 km through an extensive chenier plain close to the granite escarpment of Cape Cleveland, and extends 600 m seawards of the last chenier ridge through an extensive, well-developed mangrove fringe that is in places up to 400 m wide. The mangroves are mature and florally diverse, with trees up to 8 m in height. The mean spring tide range is 2.3 m. The transect at Cocoa Creek was taken perpendicular to the shoreline, in a series of sub-transects which cover the range from -4.5 m AHD (-2.72 m LAT) to 1.16 m AHD (0.05 m below MHWST, Table 1). We collected data from 35 sample stations. We took samples at 10-cm elevation intervals where possible (apart from below LAT, where this was

TABLE 1. Environmental and elevation information for the 4 contemporary mangrove transects in Cleveland Bay and Halifax Bay.

	Cocoa Creek	Sandfly Creek Transect 1	Sandfly Creek Transect 2	Saunders Creek
PH range	5.5-8.28	4.14-7.32	7.16-8.2	5.6-7.87
Salinity range (ppt)	5.9-19.5	7.5–13	8-15	5.3-28
% LOI range	2.2-19.5	0.5-9.5	0.4-2.6	0.5-7.0
% Sand range	0-0	0-86.2	0-75.3	0-97.6
Vegetation cover (%) range	0-90	0-90	0-75	0-90
Elevation range (m) AHD	-4.58 to 1.16	-0.346 to $+0.894$	-0.856 to $+0.304$	-0.466 to +1.694

not possible). The shallow subtidal and lower intertidal environment (-4.5 m AHD to -0.05 m AHD) is dominated by a silty substrate (~70% silt), with a low organic content (<5%), low salinity and neutral pH. The fringing *Rhizophora stylosa*-dominated floral zone starts at 0 m AHD, marked by a distinct change in grain size distributions from ~70% silt to ~90% clay, an increase in organic content to ~10%, increasing salinity (10–15‰) and a drop in pH. The *Rhizophora stylosa* mangroves are mature and vegetation cover is dense. At 0.78 m AHD there is a transition to a *Ceriops* sp. floral zone, which is accompanied by an increase in organic content to ~19%, a further increase in salinity to ~20‰ and a further drop in pH. The transect stops at 1.16 m AHD against an unvegetated, sand-rich, 3-m-high chenier ridge.

SANDFLY CREEK, CLEVELAND BAY

Sandfly Creek is the northernmost and smallest of the four tidal estuaries in Cleveland Bay (Fig. 1). The main channel of Sandfly Creek meanders through a series of chenier ridges, and extends 800 m seaward of the final ridge through fairly mature mangroves and shallow tidal flats. The fringing mangroves extend along the coast on either side of the mouth of the creek for some distance, with large stands of *Rhizophora stylosa* and *Avicennia marina* at the mouth of the creek (Bunt and Bunt, 1999). The mean spring tidal range is 2.3 m.

Sandfly Creek Transect 1

This transect was taken diagonally from the creek mouth, perpendicular to the shoreline, and has 14 sample stations, which we placed strategically where there are changes in topography or vegetation. The transect covers a range from just below MLWST to MHWST (Table 1). The sedimentary environment consists of a silt-rich unvegetated tidal flat below approximately -0.3 m AHD, moving into a sparsely vegetated fringing Avicennia marina-dominated floral zone at approximately 0 m AHD, with an increasingly sandy substrate. Salinity is low in these two zones, whereas pH is neutral. At 0.15 m AHD, there is a transition to a densely vegetated Rhizophora stylosa-dominated floral zone, with between 80-90% vegetation cover. The substrate remains predominantly sandy. Salinity increases while pH decreases through this zone. At approximately 0.6 m AHD, a transition occurs to a second, more dense Avicennia marina-dominated floral zone. This floral zone is less dense than the Rhizophora stylosa-dominated zone it replaces. The substrate becomes more clay-rich, salinity continues to increase (~13‰) and pH rises slightly. This zone is interrupted at 0.9 m AHD by an unvegetated, sand-rich chenier ridge, approximately 5 m wide and 2 m high. The mangrove continues behind this chenier ridge, consisting predominantly of Aegiceras corniculatum mangrove species, which are infrequently inundated by saline waters from the estuarine channel.

Sandfly Creek Transect 2

This transect was taken perpendicular to the shoreline through a series of floral mangrove zones, approximately

100 m west of the mouth of Sandfly Creek, and has 20 sample stations, which we placed strategically where there were marked changes in topography or vegetation. The transect covers a range from just below MLWST to MHWST (Table 1). There is an unvegetated, silty tidal flat at approximately -0.8 m AHD, grading into a sparsely vegetated fringing Avicennia marina-dominated floral zone at -0.6 m AHD. A few small, sandy tidal channels occur in this zone. Salinity is low in these zones (~8%) and pH is relatively high. At -0.3 m AHD, there is a transition to a poorly developed Rhizophora stylosa-dominated floral zone. The Rhizophora stylosa mangroves are juvenile and do not have well-developed prop roots. Salinity increases through this zone (\sim 13‰) and pH drops to near neutral. At 0 m AHD, there is a transition to an open Avicennia marinu-dominated floral zone. Salinity increases again through this zone (~15‰), and pH remains fairly neutral. This zone is interrupted at 0.25 m AHD by an unvegetated, sand-rich chenier ridge, which is also present at Transect 1.

SAUNDERS CREEK, HALIFAX BAY

Approximately 50 km north of Townsville, in southern Halifax Bay, is Saunders Creek, which meanders through dense, estuarine mangroves towards the final shoreline, formed by a beach ridge. The transect at Saunders Creek was taken perpendicular to the estuarine channel, with a total of 10 sample stations, which we placed strategically where there were marked changes in topography or vegetation. The transect covers a range from just above MLWST to just below Highest Astronomical Tide (HAT; Table 1). The unvegetated channel is at approximately -0.5 m AHD, and has a very high sand percentage (95%), very low salinity (<5%) and relatively low pH. At -0.4 m AHD, there is a transition to a densely vegetated Rhizophora stylosadominated floral zone, with increasing salinity values $(\sim 10\%)$, a silt-rich substrate (44% silt) and increasing pH levels. Vegetation cover is approximately 95% in this zone. At 1.4 m AHD, there is a transition to a narrow Avicennia marina-dominated floral zone, which has high salinity (~18‰) and high pH. At 1.55 m AHD, there is a transition to an open, Aegiceras corniculatum-dominated floral zone. Salinity continues to increase to approximately 20%, whereas pH remains high. The substrate is silt-rich, and the density of vegetation cover decreases to approximately 60%. At 1.6 m AHD, there is a transition from an Aegiceras corniculatum-dominated floral zone to a saltpan. Salinity increases to approximately 28‰, and vegetation cover decreases to approximately 10% and is made up of small stands of Salicornia australis and other salt-tolerant marsh species.

MATERIALS AND METHODS

We collected foraminifer samples (10 cm² surface sample by 1 cm thick) and environmental samples (30 cm² surface sample by 1 cm thick) from transects which crossed the whole of the mangrove zonation and intertidal zone where possible (following Horton, 1999). All readings were taken at low tide during a neap tidal cycle in the southern hemisphere winter. The environmental samples were analyzed for grain size, loss on ignition, pH and salinity. Salinity and pH analyses were performed by adding 25 ml of distilled water to 5 g of sediment and measuring levels in the water. Surface samples were generally concentrated within subenvironments near to mean sea level, particularly within the mangrove swamp, intertidal mudflat and shallow subtidal mudflat. All stations were leveled using a level and staff. An altitude for each station in relation to Australian Height Datum was achieved by either leveling the altitude of the swash mark from the previous high tide, or leveling to the sea and using a 'timed still' water reading to relate the altitudes to the tidal curve for Townsville (following Horton and others, 2003).

FORAMINIFERAL ANALYSIS

Sample preparation followed that of Scott and others (2001). Each sample was placed in buffered ethanol with the protein stain rose Bengal to identify organisms living at the time of collection (after Murray, 1991). Samples were subdivided into eight aliquots using a wet-splitter (Scott and others, 2001), and 200 tests were counted where possible. Taxonomy (Pl. 1) follows Albani (1968), Haig (1988), Bronniman and Whittaker (1993), Wynn-Jones (1994), Yassini and Jones (1995), Hayward and others (1999a), Revets (2000) and Horton and others (2003). Recent advances in molecular and morphometric analysis (Hayward and others, 2004a) have allowed the distinction of different morphological types of Ammonia, commonly described in the literature as Ammonia beccarii. In light of this work we have illustrated most morphological types described in this paper as Ammonia aoteana (Pl. 1).

The foraminiferal data are expressed as a percentage of dead assemblages only (following Horton, 1999). Horton (1999) found that the dead assemblage most closely resembles fossilized assemblages found in cores. However, the issue of using live, dead or total assemblages remains a matter of contention (eg., Scott and Medioli, 1980b; Murray, 2000). The dead assemblage differs from the live assemblage through life processes and postmortem changes (Murray, 1991). It has been argued by Murray (1991, 2000) and others (Horton and Edwards, 2004 in press; Horton and others, 2005) that the live component is variable and is not transferred into subsurface environments; therefore, its inclusion would degrade the utility of the dataset. By using the dead assemblage, we are able to compare our data with that of Horton and others (2003) from the same coastline. Dead individuals contribute at least 86% of the total number of tests counted at each site.

The foraminiferal preservation was generally very good, although samples in Sandfly Creek Transect 1 often yielded less than 40 specimens. For samples from the other three transects, a total of at least 200 was reached (following Patterson and Fishbein, 1989). We used unconstrained incremental sum-of-squares cluster analysis to detect, describe and classify patterns within the foraminiferal data from Cocoa Creek, Sandfly Creek Transect 2 and Saunders Creek. We used unconstrained cluster analysis based on unweighted Euclidean distance with foraminifera as unstandardized percentage abundances to classify contemporary samples into more-or-less homogeneous groups (clusters). Detrended Correspondence Analysis (DCA) was used to represent samples as points in multidimensional space. Only samples with counts greater than 160 individuals and species that reach 5% of the total sum were included. Cluster analysis is effective in classifying the samples according to their foraminiferal assemblage, but Detrended Correspondence Analysis gives further information about the pattern of variation within and between groups, which is important because the precise boundaries between clusters can be arbitrary. The elevation of each station within the reliable clusters was analyzed to determine a vertical zonation of each intertidal environment. Cluster analysis and DCA are illustrated for Cocoa Creek in Figure 2. Other sites are not shown, but this information is available from the authors. A repository with all foraminiferal data from this study is located at http:// www.CushmanFoundation.org, with reference number JFR DR200508.

RESULTS

A total of 72 dead foraminiferal species were found in samples taken from the intertidal and shallow subtidal zones. The composition of the foraminiferal assemblages and their vertical zonation are given below.

COCOA CREEK

There were abundant foraminifera within this transect, with counts in excess of 200 possible at all sample stations (Figs. 2, 3). The dominant species in the subtidal and intertidal mudflat were Pararotalia venusta, Parrellina hispidula, Ammonia aoteana and A. tepida. The fauna in the mudflat zone was highly diverse, with up to 30 different species recorded at each sample station. The Rhizophora stylosa zone marks a transition from a fully calcareous assemblage to a mixed calcareous and agglutinated assemblage, dominated by A. aoteana, Rosalina sp., Miliammina fusca and Paratrochammina stoeni. The uppermost part of the transect, in the Ceriops sp. zone close to MHWST, has a fully agglutinated assemblage dominated by M. fusca, Trochammina inflata and Haplophragmoides sp. Cluster analysis of foraminiferal death assemblages at each sample station detects three zones (Fig. 2):

- Zone CC I is an agglutinated foraminiferal zone with a low species diversity (approximately 12 species per sample). The dominant species are *Miliammina fusca*, *Trochammina inflata*, *Ammotium directum* and *Haplophragmoides* sp. The elevation range of this zone is 0.32–1.16 m AHD (vertical range of 0.84 m).
- Zone CC IIa is a mixed calcareous and agglutinated foraminiferal zone with a reasonably diverse fauna (approximately 18 species found at each sample station). The dominant species are A. aoteana, Rosalina sp., Paratrochammina stoeni and M. fusca. The elevation range of this zone is 0.04–0.30 m AHD (vertical range of 0.26 m).
- Zone CC IIb is a fully calcareous foraminiferal zone with a highly diverse fauna (approximately 30 species found at each sample station). The dominant species are *Pararotalia venusta*, *Parrellina hispidula*, *A. aoteana* and *A. tepida. Pararotalia venusta* alone accounts for between 20-54 % of the count at each sample station. The elevation range of this zone is -4.58--0.05 m AHD (vertical range of 4.53 m).

SANDFLY CREEK TRANSECT 1

The foraminiferal preservation in the surface sediments ranged between several hundred and 0 tests per 10 cm³ (Fig. 4). The tidal mudflat had abundant and diverse calcareous species, and counts in excess of 200 were possible. The calcareous species Miliolinella lakemarquiensis, Ammonia tepida and A. aoteana were dominant. In the densely vegetated, fringing Rhizophora stylosa floral zone and the more sparsely vegetated Avicennia marina floral zone, the foraminiferal counts were extremely low (as few as 8 specimens per 10cc sample), consisting of a mixture of calcareous and agglutinated species, mostly Trochammina inflata and Cribroelphidium poeynum. Only one assemblage zone was detected and classified at this site, being a calcareous zone dominated by Ammonia aoteana (T1 II). The elevation range of this zone is -0.36-0.36 m. a vertical range of 0.61 m.

SANDFLY CREEK TRANSECT TWO

Foraminiferal preservation in this transect was good, and counts in excess of 200 were possible (Fig. 5). The dominant species in the tidal mudflat were Miliolinella lakemarquiensis, Ammonia tepida and A. aoteana, together accounting for between 40 and 50% of the total foraminiferal count. Generally, the assemblage in the mudflat zone was highly diverse, with up to 30 species recorded at each sample station. The foraminiferal fauna was uniform through the fringing Avicennia marina- and Rhizophora stylosadominated floral zones, but showed a slight increase in Ammonia tepida at the expense of A. aoteana through the Rhizophora stylosa zone. The second Avicennia marina-dominated floral zone had fluctuating values of Ammonia aoteana and A. tepida, but retained species diversity. The uppermost transect station, situated at the base of the chenier ridge, had a unique fauna consisting primarily of Quinqueloculina suborbicularis and A. aoteana (together totaling 77% of the count at this sample station). Cluster analysis of foraminiferal death assemblages at each sample station detects two zones:

- Zone T2 IIa is dominated by calcareous species, with moderately low species diversity (approximately 15 species found at each sample station). The dominant species are *Ammonia aoteana* and *Quinqueloculina suborbicularis*, with a maximum relative abundance of 45%. The elevation range of this zone is 0.1–0.3 m AHD (vertical range of 0.2 m).
- Zone T2 IIb is a calcareous foraminifera-dominated zone, with a highly diverse fauna (approximately 25 species found at each sample station). Ammonia tepida and Miliolinella lakemarquiensis are the dominant species, with a maximum relative abundance of 56 %. Other notable species include A. aoteana, Triloculina oblonga and Wiesnerella auriculata. The elevation range of this zone is -0.9-0.2 m AHD (vertical range of 1.1 m).

SAUNDERS CREEK

In the tidal creek, the assemblage is dominated by Ammonia aoteana (up to 80% of the total count), with low but persistent occurrences of agglutinated species Miliammina fusca and Trochammina inflata (Fig. 6). Through the fringing Rhizophora stylosa floral zone, Ammonia aoteana decreases and agglutinated species increase rapidly. Miliammina fusca peaks through this floral zone. The ratio of agglutinated to calcareous species is 60:40 through the Rhizophora zone. Moving into the Avicennia marina-dominated floral zone, Trochammina inflata gains in relative importance, and in the Aegiceras corniculatum floral zone, Trochammina inflata is the dominant species (60% of the counts). In the upper intertidal saltpan, both Trochammina inflata and Miliammina fusca dominate. Multivariate analysis of foraminiferal death assemblages from Saunders Creek delineates two zones:

- Zone SC I has both calcareous and agglutinated species and consists of eight samples. Species diversity is fairly low (approximately 15 species per sample). The dominant species are *Miliammina fusca*, *Trochammina inflata* and *Ammonia aoteana*, together accounting for a maximum of 84% relative abundance. The elevation range of this zone is -0.1-1.7 m AHD.
- Zone SC IIa is dominated by calcareous species, and consists of two samples from the lower end of the transect, in the tidal creek. Species diversity is low (approximately 10 species per sample), and the dominant species is *Ammonia aoteana*, with a maximum relative abundance of 68%. The elevation range of this zone is -0.5-0.1 m AHD (vertical range of 0.4 m).

DISCUSSION

The four transects from Cleveland Bay and Halifax Bay each show different foraminiferal assemblages across a tropical mangrove environment (Fig. 7). Sandfly Creek Transect 1 had very high and very low counts, in excess of 200 tests per 10 cm³ in the mudflat environment, but in the fringing Rhizophora stylosa and sparsely vegetated Avicennia marina zones, counts were as low as 8 tests per 10 cm³. All of the other sites had abundant foraminifera within them. There are many controlling factors on foraminiferal abundance and type, including nutrition, dissolved oxygen conditions, pH, salinity, substrate and temperature (e.g., Murray, 1968; Alve and Nagy, 1986; Boltovskoy and others, 1991; de Rijk, 1995; Barbosa and Suguio, 1999; Debenay and others, 2002; Horton and others, 2003). Barbosa and Suguio (1999) and Horton and others (2003) show that the foraminiferal assemblages in the coastal mangroves of Brazil and the GBR coastline, respectively, are controlled by the elevation with respect to the tidal frame. However, ground temperature, sparse vegetation cover and low organic content are particularly important considerations in tropical locations, where ponded water in the intertidal zone can reach temperatures of over 40°C between tidal inundations and there is less organic food available (Murray, 1968; Dublin-Green, 1992; de Rijk, 1995).

Sandfly Creek Transect 1 was relatively acidic. The pH variations within intertidal environments are generally greater than in any other marine environment (Phleger and Bradshaw, 1966), and pH has an effect on both the protoplasmic cell and the calcareous test of foraminifera. Experiments using live *Ammonia* in normal salinity waters of different pH show that decalcification begins at values below 7.5 (Le

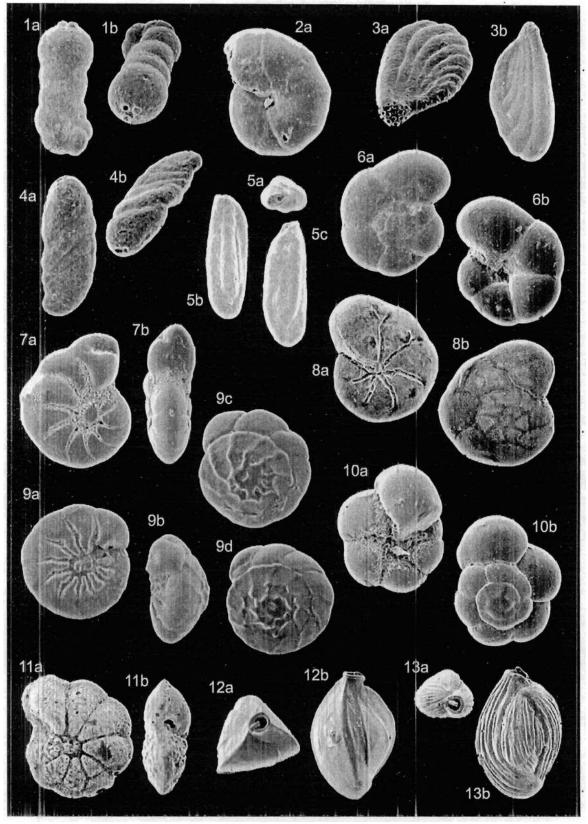


PLATE 1

Fig. 1a Acupeina triperforata (Millett) side view, ×120. Fig. 1b Acupeina triperforata (Millett) oblique apertural view, ×120. Fig. 2a Haplophragmoides sp. (D'Orbigny) side view, ×120 Fig. 3a Ammoastuta salsa (Cushman & Bronniman) oblique apertural view, ×150. Fig. 3b Ammoastuta salsa (Cushman & Bronniman) side view, ×150. Fig. 4a Ammotium directum (Cushman & Bronniman) side view, ×150. Fig. 4b Ammotium directum (Cushman & Bronniman) oblique apertural view, ×160. Fig. 5a Miliammina fusca (Brady) apertural view, ×176. Fig. 5b Miliammina fusca (Brady) side view, ×136. Fig. 5c Miliammina fusca (Brady) side view, ×136. Fig. 6a Trochammina inflata (Montagu) spiral

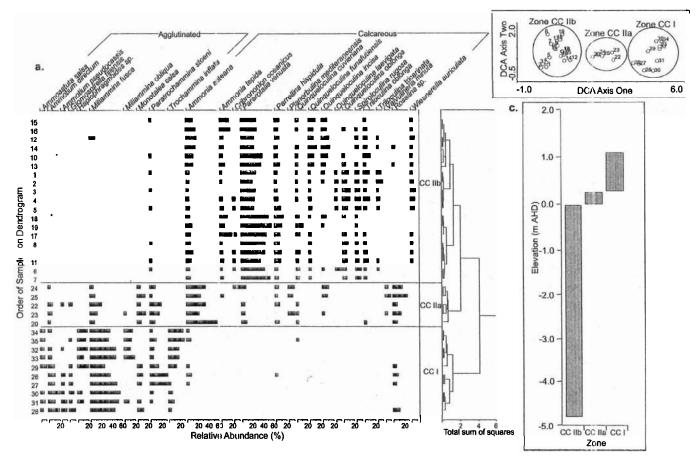


FIGURE 2. (a) Unconstrained incremental sum-of-squares cluster analysis based on unweighted Euclidean distance of foraminiferal death assemblages from Cocoa Creek. Only samples with counts > 160 individuals and species that reach 5% of the total sum are included. (b) DCA biplot showing the 3 main foraminiferal assemblages. (c) Box plots showing maximum and minimum elevations (m, Australian Height Datum) for the faunal zones of Cocoa Creek.

Cadre and others, 2003), and Alve and Nagy (1986) report dissolution of tests under a pH range of 6.5–7.2. Low pH is unfavorable for calcareous foraminifera, and individuals must spend considerable energy recalcifying their tests.

The low intertidal mudflat at Sandfly Creek Transect 1 had an average pH of 7, whereas the pH in the vegetated *Rhizophora stylosa* and *Avicennia marina* zones ranges from 6.2–6.6, within and below the reported pH range which may cause dissolution of tests. This may explain the total absence of calcareous foraminifera in Sandfly Creek Transect 1 through the vegetated zones, which are below the elevation range where agglutinated foraminifera are found. Levels of pH through the vegetated zones at Sandfly Creek Transect 2 remain above 7.5, so total dissolution of calcareous tests does not occur. Debenay and others (2002) argue that decomposition of leaf litter in mangrove zones by bacterial activity lowers pH and contributes to the disappearance of calcareous foraminifera. These local-scale variations in pH levels impact greatly on foraminiferal presence and abundance.

Multivariate analysis of samples from Sandfly Creek Transect 2, Cocoa Creek and Saunders Creek shows a series of foraminiferal assemblages which cluster in relation to elevation. However, none of the transects cover the whole intertidal zone from LAT to HAT because of morphological constraints such as chenier ridges (Fig. 7). This is reflected in the foraminiferal assemblage zones found at each location.

Agglutinated species Miliammina fusca, Trochammina inflata, Ammotium directum and Haplophragmoides sp. dominate zones CC I and SC I, along with persistent occurrences of Ammonia aoteana at Saunders Creek only.

←

view, ×176. Fig. 6b Trochammina inflata (Montagu) edge view, ×176. Fig. 7a Haynesina depressula (Walker & Jacob) spiral view, ×150. Fig. 7b Haynesina depressula (Walker & Jacob) edge view, ×150. Fig. 8a Rosalina sp. side view, ×200. Fig. 8b Rosalina sp. side view, ×200. Fig. 9a Ammonia aoteana (Finlay) umbilical view, ×220. Fig. 9b Ammonia aoteana (Finlay) edge view, ×250. Fig. 9c Ammonia aoteana (Finlay) spiral view, ×220. Fig. 9d Ammonia aoteana (Finlay) spiral view, ×250. Fig. 10a Ammonia tepida (Cushman) apertural view, ×200. Fig. 11b Pararotalia venusta (Brady) side view, ×200. Fig. 11b Pararotalia venusta (Brady) apertural view, ×200. Fig. 12a Triloculina tricarinata (d'Orbigny) apertural view, ×200. Fig. 13b Quinqueloculina poeyana (d'Orbigny) side view, ×200.

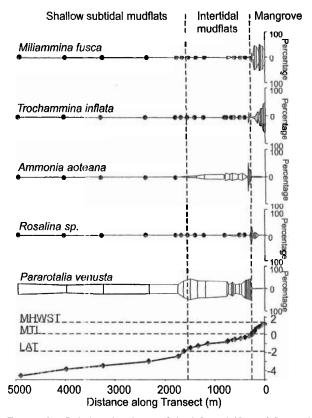


FIGURE 3. Relative abundance of dead foraminifera of five main foraminiferal species and populations from Cocoa Creek. The elevation (m, Australian Height Datum), tidal heights, floral zonation and sampling stations are indicated.

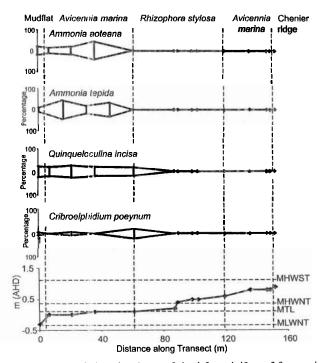


FIGURE 4. Relative abundance of dead foraminifera of four main foraminiferal species and populations from Sandfly Creek Transect 1. The elevation (m, Australian Height Datum), tidal heights, floral zonation and sampling stations are indicated.

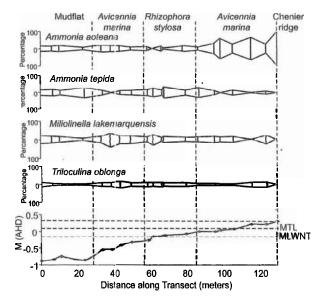


FIGURE 5. Relative abundance of dead foraminifera of four main foraminiferal species and populations from Sandfly Creek Transect 2. The elevation (m, Australian Height Datum), tidal heights, floral zonation and sampling stations are indicated.

These zones are found towards the landward edge of the mangrove study sites at Cocoa Creek and Saunders Creek, in the elevation range -0.1-1.7 m AHD (range 1.8 m, just above MLWNT to close to HAT). Similar faunal assemblages have been found in other tropical locations. Horton and others (2003) identify two faunal zones dominated by agglutinated foraminifera at the landward edge of a separate transect at Cocoa Creek, Cleveland Bay, Australia. In Indonesia, Horton and others (2005) also identify an agglutinated upper mangrove assemblage with *Trochammina inflata* and *Miliammina fusca* on islands off southeastern Sula-

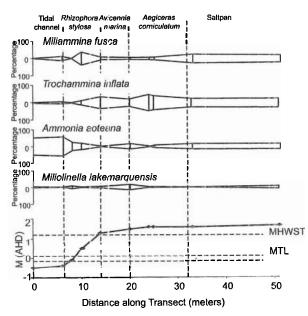


FIGURE 6. Relative abundance of dead foraminifera of four main foraminiferal species and populations from Saunders Creek. The elevation (m, Australian Height Datum), tidal heights, floral zonation and sampling stations are indicated.

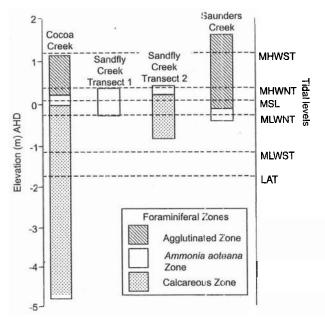


FIGURE 7. Summary of foraminiferal death assemblages at each field location, with tidal levels for Halifax and Cleveland bays.

wesi, and furthermore, Debenay and others (2000) have identified agglutinated foraminifera (*Jadammina macrescens* and *Trochammina inflata*) in upper marshes in New Caledonia and Cairns, northeastern Queensland, Australia.

Zones CC IIa, T1 II, T2 IIa and SC IIa at Cocoa, Sandfly and Saunders creeks have assemblages with low species diversity and dominated by *Ammonia aoteana*, accounting for 20–68% of the total count at each sample station in each zone. The elevation range is -0.5-0.3 m AHD (range 0.8 m, just below MLWNT to just below MHWNT). Other studies from tropical and subtropical locations have shown an *Ammonia*-dominated assemblage in the mid-intertidal zone (e.g., Haslett, 2001, in the upper part of the tidal flat at the Barron River estuary, Cairns, northeastern Queensland, Australia; Hayward and others, 1999, in tidal flats and mangrove forests in New Zealand; and Horton and others, 2005, in Indonesia). It is also found in a lower estuarine environment in Brazil (Barbosa and Suguio, 1999).

Faunal zones CC IIb and T2 IIb are found at the seaward edge of transects at Cocoa and Sandíly Creeks. They have diverse calcareous assemblages dominated by Pararotalia venusta, Ammonia tepida and Parrellina hispidula, and an elevation range of -4.58-0.20 m AHD (range 4.78 m, -2.72 m LAT to just above MTL). Haslett (2001) found a diverse foraminiferal assemblage, dominated by A. beccarii but with many other shallow marine benthic and planktonic species, in the lower intertidal zone of the Barron River estuary, and Horton and others (2003) observe a similar calcareous dominated assemblage at Cocoa Creek in the intertidal mudflats. This study is one of the first in tropical Australia to extend the sampling of intertidal mudflats to below the limit of LAT, into the shallow subtidal area. It shows the extension of this diverse calcareous zone beneath the intertidal zone and implies that for this low intertidal/ shallow subtidal assemblage, the duration and frequency of tidal inundation may not be the most important limiting factor.

No clear link between foraminiferal assemblages and floral zones can be established from our new field sites. The Ammonia aoteana-dominated zone is correlated with an upper mangrove, Avicennia marina-dominated floral assemblage at Sandfly Creek, a fringing Rhizophora stylosa zone at Cocoa Creek, and with a vegetation-free creek bed at Saunders Creek. Sandfly Creek Transect 2 shows a transition from mudflat to Rhizophora stylosa-dominated floral assemblage at a lower elevation than at the other two sites. This may explain why the Ammonia aoteana zone at Sandfly Creek is higher in the mangrove sequence than at the other two sites, despite being at a similar elevation. Localized differences in mangrove zonation may be determined on a small scale by factors such as sediment transport and nutrient availability, as well as by tidal inundation frequency (Bunt and Bunt, 1999). Floral zones appear to respond to the local environment, while foraminiferal zonation occurs at similar elevations at different types of sites.

IMPLICATIONS FOR SEA-LEVEL STUDIES

Past foraminiferal studies in temperate environments (e.g., Scott and Medioli, 1978; Jennings and Nelson, 1992; Horton, 1999; Gehrels and others, 2001) have indicated that a vertical zonation of foraminifera occurs in the intertidal zone, where the distribution of foraminifera is a direct function of elevation, with the duration and frequency of intertidal exposure as the most important environmental factors. Scott and others (2001) state that a vertical zonation of marsh foraminiferal assemblages exists on a worldwide scale, and suggest that the same 8-10 species of marsh foraminifera are ubiquitous worldwide in the upper part of the intertidal zone. Information on vertical zonation trends in tropical environments are more limited, and the evidence which exists points to a range of environmental factors being important. In New Zealand, salinity and elevation are shown as jointly the most important factors governing foraminiferal distribution (Hayward, 1999a, 1999b; 2004b), and Debenay and others (2000) show that in mangroves a range of environmental factors are important, including salinity and the presence or absence of vegetation.

An important issue when considering the applicability of foraminiferal zones in sea-level reconstructions is the preservation of foraminifera in fossil deposits. Agglutinated foraminifera within high-marsh environments have been used to precisely reconstruct former sea levels in temperate salt marshes (e.g., Gehrels, 2000; Gehrels and others, 2002; Horton and others, 2005). Many fossil cores have been collected from tropical mangrove and estuarine environments on the Great Barrier Reef coastline. These deposits have many calcareous benthic foraminifera preserved within silty clay horizons, but no foraminifera are preserved within organic-rich horizons (e.g., Fig. 8). The organic horizons have grain size characteristics and organic content similar to those of modern mangrove environments, and preserve abundant mangrove pollen. Taphonomic loss of foraminiferal tests due to post-mortem disaggregation is an important obstacle in the use of mangrove (mainly agglutinated) foraminifera in paleoenvironmental reconstructions in this location. The low pH environment of mangroves is not favorable for the preservation of calcareous foraminifera; however, agglutinated

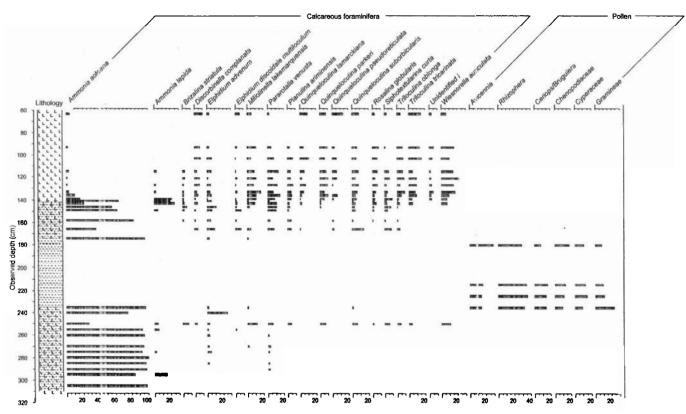


FIGURE 8. Foraminifera and pollen preserved in a core (KG931 VC1) collected from a shallow subtidal environment in Cleveland Bay, north Queensland. No foraminifera are preserved in the organic section of the core between 180–235cm; however, mangrove pollen are well preserved through this section. Lithology modified from Troels-Smith. Symbols: LLL—silty clay, horizontal dashes---organic material.

foraminifera are also not readily preserved in mangrove horizons. Studies show that thin-shelled tests such as those of *Miliammina obliqua* rapidly disaggregate on burial (Hayward and others. 2004) and even handling of materials in the field and laboratory may affect test preservation (Wang and Chappell, 2001). We do not find even the most robust agglutinated species (e.g., *Trochammina inflata*) preserved in cores. Because of the absence of upper intertidal agglutinated foraminifera in fossil deposits from the Great Barrier Reef coastline, it is important for us to understand contemporary vertical zonation trends within mid- to low-intertidal calcareous faunas which live in muddy environments. These may be used, albeit with potentially decreased precision, to allow quantitative sea-level reconstructions using foraminiferal assemblages from this area.

An assemblage made up solely of calcareous species, but which exists only between MLWNT and MHWNT (range of ~ 0.7 m) and is dominated by *Ammonia aoteana* may be a useful sea-level indicator in this environment because of the problems associated with removal of agglutinated foraminifera from fossil sediments. This assemblage often exists below the limit of fringing mangroves, and therefore is unlikely to be affected by large pH excursions experienced in vegetated zones. However, in some locations where mangroves extend lower into the intertidal zone (e.g., Sandfly Creek Transect 1), low pH within the mangrove will likely cause the dissolution of calcareous tests such as those of *A. aoteana*.

Where agglutinated foraminifera are preserved in fossil deposits, a modern agglutinated assemblage which exists be-

tween MLWNT and HAT (range 1.8 m), consisting primarily of *Miliammina fusca*, *Trochammina inflata*, *Ammotium directum* and *Haplophragmoides* sp. may be the most useful sea-level indicator, depending on the degree of preservation of calcareous foraminifera in the fossil cores. The usefulness of different modern foraminiferal assemblages in reconstructing sea levels should, therefore, be assessed on a core by core basis.

In the tropical mangrove environments studied, local scale factors affect foraminiferal distributions. Limitations to using foraminifera as precise indicators in sea-level reconstructions include low pH excursions causing dissolution of calcareous tests and the potential problem of taphonomic loss of agglutinated foraminifera in fossil deposits.

CONCLUSIONS

Statistical analysis of the foraminiferal death assemblages from all sites supports numerous studies from temperate and tropical regions that indicate a vertical zonation of foraminifera exists within the intertidal zone. An upper mangrove foraminiferal zone dominated by agglutinated species, including *Trochammina inflata, Paratrochammina stoeni* and *Miliammina fusca*, exists in the higher tidal elevation range (-0.1-1.7 m AHD) at Cocoa Creek and Saunders Creek. A mid-intertidal foraminiferal zone dominated by *Ammonia aoteana* (-0.5-0.3 m AHD) exists at all sites. A low intertidal foraminiferal zone, dominated by *A. aoteana, A. tepida* and numerous other calcareous species (-4.58-0.2 mAHD), exists at Cocoa Creek and Sandfly Creek. The only for a miniferal zone present at all locations is an A. aoteanadominated for a miniferal assemblage between -0.5-+0.3 m AHD (just above MLWNT to MHWNT).

ACKNOWLEDGMENTS

The authors thank Scott Smithers for his assistance in the field, and James Cook University, Townsville, Australia, which hosted Woodroffe whilst the fieldwork was undertaken. We also thank Dr. R. Watkins for kindly supplying pollen data from a studied core. We acknowledge and greatly appreciate funding by the Ouaternary Research Association. This research was carried out while in receipt of a Natural Environment Research Council award (NER/S/C/2002/ 10581). Special acknowledgments are given to Dave Scott, Bruce Hayward and an anonymous reviewer for their valuable comments on the original version of this paper. The authors thank the cartography department at the Department of Geography, University of Durham, for producing the figures, and to all members of the Environmental Research Centre, University of Durham, for their help and advice. This paper is a contribution to IGCP project 495.

REFERENCES

- ALBANI, A. D., 1968, Recent foraminifera from Port Hacking, New South Wales: Contributions from the Cushman Foundation for Foraminiferal Research, v. 19, p. 85–119.
- ALVE, E., and NAGY, J., 1986, Estuarine foraminiferal distribution in Sandebukta, a branch of the Oslo fjord: Journal of Foraminiferal Research, v. 16, p. 261–284.
- BARBOSA, C. F. and SUGUIO, K., 1999, Biosedimentary facies of a subtropical microtidal estuary—an example from southern Brazil: Journal of Sedimentary Research, v. 69, p. 576–587.
- BEAMAN, R., LARCOMBE, P., and CARTER, K. M., 1994, New evidence for the holocene sea-level high from the inner shelf, central Great-Barrier-Reef, Australia: Journal of Sedimentary Research, Section A—Sedimentary Petrology and Processes, v. 64, p. 881–885.
- BELPERIO, A. P., 1983, Terrigenous sedimentation in the central Great Barrier Reef lagoon: a model from the Burdekin region: Bureau of Mineral Resources Journal of Australian Geology and Geophysics v. 8, p. 179–190.
- BOLTOVSKOY, E., SCOTF, D. B.,, and MEDIOLI, F. S., 1991, Morphological variations of benthic foraminiferal tests in response to changes in ecological parameters: Journal of Paleontology, v. 65, p. 175-185.
- ,and WRIGHT, R., 1976, Recent Foraminifera: Junk, The Hague, 515 pp.
- BRONNIMAN, P., and WHITTAKER, J. E., 1993, Taxonomic revision of some recent agglutinated foraminifera from the Malay archipelago in the Millett Collection, Natural History Museum, London: Bulletin of the Natural History Museum, London (Zoology), v. 59, p. 107-124.
- BUNT, J. S., and BUNT, E. D., 1999, Complexity and variety of zonal pattern in the mangroves of the Hinchinbrook area, Northeastern Australia: Mangroves and Saltmarshes, v. 3, p. 165–176.
- CARTER, R. M., JOHNSON, D. P., and COOPER, K. G., 1993, Episodic post glacial sea-level rise and the sedimentary evolution of a tropical continental embayment (Cleveland Bay, Great Barrier Reef Shelf, Australia): Australian Journal of Earth Sciences, v. 40, p. 229-255.
- DEBENAY, J. -P., GUILLOU, J., REDOIS, F., and GESLIN, E., 2000, Distribution trends of foraminiferal assemblages in paralic environments, *in* Martin, R. E. (ed.), Environmental micropaleontology, Topics in Geobiology, vol. 15: Kluwer Academic Publishers, New York, p. 39–67.
 - —, GUIRAL, D., and PARRA, M., 2002. Ecological factors acting on the microfauna in mangrove swamps. The case of foraminiferal assemblages in French Guiana: Estuarine, Coastal and Shelf Science, v. 55, p. 509–533.

- DUBLIN-Green, C. O., 1992, Benthic foraminiferal ecology and sediment distribution in the Bonny River, Niger Delta: Unpublished Ph.D. Dissertation, University of London, United Kingdom.
- EDWARDS, R. J., and HORTON, B. P., 2000, High resolution records of relative sea-level change from UK saltmarsh foraminifera: Marine Geology, v. 169, p. 41–56.
- GEHRELS, W.R., 2000, Using foraminiferal transfer functions to produce high-resolution sea-level records from salt-marsh deposits, Maine, USA: The Holocene, v. 10, p. 367–376.
- ——, ROE, H. M., and CHARMAN, D. J., 2001, Foraminifera, testate amoebae and diatoms as sea-level indicators in UK saltmarshes: A quantitative multiproxy approach: Journal of Quaternary Science, v. 16, p. 201–220.
- ——, BELKNAP, D. F., BLACK, S., and NEWNHAM, R. M., 2002, Rapid sea-level rise in the Gulf of Maine, USA, since AD 1800: The Holocene, v. 12, p. 383–389.
- HASLETT, S. K., 2001, The Palaeoenvironmental implications of the distribution of intertidal foraminifera in a tropical Australian estuary: a reconnaissance study: Australian Geographical Studies, v. 39, p. 67–74.
- HAIG, D. W., 1988, Miliolid foraminifera from inner neritic sand and mud facies of the Papuan Lagoon, New Guinea: Journal of Foraminiferal Research, v. 18, p. 203–236.
- HAYWARD, B.W., GRENFELL, H. R., REID, C. M., and HAYWARD, K. A., 1999a, Recent New Zealand shallow-water benthic foraminifera: taxonomy, ecological distribution, biogeography, and use in palaeoenvironmental assessment: Institute of Geological and Nuclear Science Ltd., Lower Hutt, New Zealand, 258 p.
- —, GRENFELL, H. R., and SCOTT, D. B., 1999b, Tidal range of marsh foraminifera for determining former sea-level heights in New Zealand: New Zealand Journal of Geology and Geophysics, v. 42, p. 395-413.
- ———, HOLZMANN, M., GRENFELL, H. R., PAWOWSKI, J., and TRIGGS, C. M., 2004a, Morphological distinction of molecular types in *Ammonia*—towards a taxonomic revision of the world's most commonly misidentified foraminifera: Marine Micropalaeontology, v. 50, p. 237–271.
- SCOTT, G. H., GRENFELL, H. R., CARTER, R., and LIPPS, J. H., 2004b, Techniques for estimation of tidal elevation and confinement (~salinity) histories of sheltered harbours and estuaries using benthic foraminifera: examples from New Zealand: The Holocene, v. 14, p. 218–232.
- HORTON, B. P., 1999, The distribution of contemporary intertidal foraminifera at Cowpen Marsh, Tees Estuary, UK: Implications for studies of Holocene sea-level changes: Palaeogeography, Palaeoclimatology, Palaeoecology, v. 149, p.127-149.
- ——, EDWARDS, R. J., and LLOYD, J. M., 2000, Implications of a microfossil-based transfer function in Holocene sea-level studies, in Shennan, I., and Andrews, J. (eds.), Holocene Land-Ocean Interaction and Environmental Change around the North Sea: Geological Society, London, Special Publication v. 166, p. 41–54.
- —, LARCOMBE, P., WOODROFFE, S. A., WHITTAKER, J. E., WRIGHT, M. R., and WYNN, C., 2003, Contemporary foraminiferal distributions of the GBR coastline, Australia: implications for sea-level reconstructions: Marine Geology, v. 198, p. 225–243.
- —, and EDWARDS, R. J., 2005, The application of local and regional transfer functions to the reconstruction of Holocene sea levels, north Norfolk, England: The Holocene, v. 15, p. 143–155.
- , WHITTAKER, J. E., THOMSON, K. H., HARDBATTLE, M. I. J., WOODROFFE, S. A., and WRIGHT, M. R., 2005, The development of a modern foraminiferal data set for sea-level reconstructions, Wakatobi Marine National Park, Southeast Sulawesi, Indonesia: Journal of Foraminiferal Research, vol. 35, p. 1–14.
- JAVAUX, E. J., 1999, Benthic Foraminifera from the Modern Sediments of Bermuda: Implications for Holocene Sea-Level Studies: Dalhousie University, Nova Scotia, Canada, 625 p.
- —, and SCOTT, D. B., 2003, Illustration of modern benthic foraminifera from Bermuda and remarks on distribution in other subtropical/tropical areas: Palaeontologica Electronica, v. 6, 29 pp. 2.1MB; http://palaeo-electronica.org/paleo/2003_1/benthic/issue1_ 03.htm
- JENNINGS, A. E., and NELSON, A. R., 1992, Foraminiferal assemblage zones in Oregon tidal marshes—relation to marsh floral zones and sea-level: Journal of Foraminiferal Research, v. 22, p. 13–29.

- LARCOMBE, P., RIDD, P. V., PRYTZ, A., and WILSON, B., 1995, Factors controlling suspended sediment on inner-shelf coral reefs, Townsville, Australia: Coral Reefs, v. 14, 163–171.
- LAMBECK, K., YOKOYAMA, Y., and PURCELL, T., 2002, Into and out of the Last Glacial Maximum: Sea-level change during oxygen isotope stages 3 and 2: Quaternary Science Reviews, v. 21, p. 343-360.
- LE CADRE, V., DEBENAY, J. -P., and LESOURD, M., 2003, Low pH effects on *Amunonia beccarii* test deformation: implications for using test deformations as a pollution indicator: Journal of Foraminiferal Research, v. 33, p. 1–9.
- MICHIE, M. G., 1987, Distribution of foraminifera in a macrotidal tropical estuary: Port Darwin, Northern Territory of Australia: Australian Journal of Marine and Freshwater Research, v. 38, p. 249– 259.
- MILNE, G. A., MITROVICA, J. X., and SCHRAG, D. P., 2002, Estimating past continental ice volume from sea-level data: Quaternary Science Reviews, v. 21, p. 361–376.
- MITROVICA, J. X., and PELTIER, W. R., 1991, On postglacial geoid subsidence over the equatorial oceans: Journal of Geophysical Research, v. 96, p. 20,053–20,071.
- MURRAY, J. W., 1968, The living Foraminiferida of Christchurch Harbour, England: Micropalaeontology, v. 14, p. 83–96.
- 2000, JFR comment: the enigma of the continued use of total assemblages in ecological studies of benthic foraminifera: Journal of Foraminiferal Research, v. 30, p. 244–245.
- NAKADA, M., and LAMBECK, K., 1989, Late Pleistocene and Holocene sea-level change in the Australian region and mantle rheology: Geophysical Journal, v. 96, p. 497–517.
- PATTERSON, R. T., and FISHBEIN, E., 1989, Re-examination of the statistical methods used to determine the number of point counts needed for micropaleontological quantitative research: Journal of Paleontology, v. 63, p. 245–248.
- PELTIER, W. R., 2002, On eustatic sea level history: Last Glacial Maximum to Holocene: Quaternary Science Reviews, v. 21, p. 377– 396.
- PHELGER, F. B., and BRADSHAW, J. S., 1966, Sedimentary environments in a marine marsh: Science, v.154, p. 1551–1553.
- REVERS, S. A., 2000, Foraminifera of Leschenault Inlet: Journal of the Royal Society of Western Australia, v. 83, p. 365-375.
- DE RIJK, S., 1995, Agglutinated foraminifera as indicators of salt marsh development in relation to late Holocene sea-level rise: Unpublished Ph.D. Dissertation, Free University, Amsterdam.
- SAWAI, Y., HORTON, B. P., and NAGUMO, T., 2004, The development of a diatom-based transfer function along the Pacific coast of eastern Hokkaido, northern Japan—an aid in paleoseismic studies of

the Kuril subduction zone: Quaternary Science Reviews, v. 23, p. 2467–2483.

- SCOTT, D. B., and MEDIOLI, F. S., 1978, Vertical zonations of marsh foraminifera as accurate indicators of former sea-levels: Nature v. 272, p. 528–531.
- ——, and MEDIOLI, F. S., 1980a, Quantitative studies of marsh foraminiferal distributions in Nova Scotia: implications for sea level studies: Cushman Foundation for Foraminiferal Research, Special Publication, no. 17, 58 p.
- ------, and MEDIOLI, F. S., 1980b, Living vs. total foraminiferal populations: their relative usefulness in paleoecology: Journal of Paleontology v. 54, p. 814–831.
- —, MEDIOLI, F. S., and SCHAFER, C. T., 2001, Monitoring in coastal environments using foraminifera and thecamoebian indicators: Cambridge University Press, 177 p.
- SCHNACK, E. S., FERRERO, L., ESPINOSA, M., and BARBOSA, C. F. 1990, Recent marsh foraminifera from the east coast of South America: comparison to the northern hemisphere, *in* Hemleben, C., Kaminski, M. A., Kuhnt, W., and Scott, D. B. (eds.), Paleoecology, Biostratigraphy, Paleoceanography and Taxonomy of Agglutinated Foraminifera, NATO ASI Series C, 327, Math and Physical Sciences, p. 717–738.
- SHENNAN, I., and HORTON, B. P., 2002, Holocene land and sea-level changes in Great Britain: Journal of Quaternary Science, v. 17, p. 511–526.
- —, HORTON, B., PELTIER, W.R., and DRUMMOND, R., 2002, Global to local scale parameters determining relative sea-level changes and the post-glacial isostatic adjustment of Great Britain: Quaternary Science Reviews, v. 21, p. 397–408.
- WANG, P., and CHAPPELL, J., 2001, Foraminifera as Holocene environmental indicators in the South Alligator River, Northern Australia: Quaternary International, v. 83–85, p. 47–62.
- WYNN-JONES, R., 1994, The Challenger Foraminifera: Oxford Science Publications, Oxford, 149 p.
- YASSINI, I., and JONES, B. G., 1995, Foraminiferida and Ostracoda from Estuarine and Shelf Environments on the Southeastern Coast of Australia: University of Wollongong Press, Wollongong, Australia, 484 p.
- YOKOYAMA, Y., DE DECKKER, P., LAMBECK, K., JOHNSTON, P., and FIFIELD, L. K., 2001, Sea-level at the Last Glacial Maximum: evidence from northwestern Australia to constrain ice volumes for oxygen isotope stage 2: Palaeogeography, Palaeoclimatology, Palaeoecology, v. 165. p. 281–297.
 ZONG, Y., and HORTON, B. P., 1999, Diatom-based tidal-level transfer
- ZONG, Y., and HORTON, B. P., 1999, Diatom-based tidal-level transfer functions as an aid in reconstructing Quaternary history of sealevel movements in the UK: Journal of Quaternary Science, v. 14, p. 153–167.

Received 12 August 2003 Accepted 1 December 2004