

1 Intra-annual dynamics of non-structural carbohydrates in the  
2 cambium of mature conifer trees reflects radial growth demands

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10 Running head: NSC IN THE CAMBIUM OF MATURE CONIFERS

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## 21 **Summary**

22 The presence of soluble carbohydrates in the cambial zone, either from  
23 sugars recently produced during photosynthesis or starch remobilized from  
24 storage organs, is necessary for radial tree growth. However, considerable  
25 uncertainties on carbohydrate dynamics and the consequences on tree  
26 productivity exist. This study aims to better understand the variation of the  
27 different carbon pools at intra-annual resolution by quantifying how cambial  
28 zone sugar and starch concentrations fluctuate over the season and in  
29 relation to cambial phenology. A comparison between two physiologically  
30 different species growing at the same site, i.e., the evergreen *Picea abies* and  
31 the deciduous *Larix decidua* and between *Larix decidua* from two contrasting  
32 elevations is presented to identify mechanisms of growth limitation.

33 Results indicate that the annual cycle of sugar concentration within the  
34 cambial zone is coupled to the process of wood formation. The highest sugar  
35 concentration is observed when the number of cells in secondary wall  
36 formation and lignification stages is at a maximum, subsequent to most radial  
37 growth. Starch disappears in winter while other freeze-resistant non-structural  
38 carbohydrates (NSC) increase. Slight differences in NSC concentration  
39 between species are consistent with the differing climate sensitivity of the  
40 evergreen and deciduous species investigated. The general absence of  
41 differences between elevations suggest that the cambial activity of trees  
42 growing at the treeline was not limited by the availability of carbohydrates at  
43 the cambial zone but instead by environmental controls on growing season  
44 duration.

45 Keywords: NSC, cambium, *Larix decidua*, *Picea abies*, intra-annual analysis,  
46 tree line, phenology

47

## 48 **Introduction**

49 Tree growth and survival depends not only on their capacity to produce and  
50 use carbohydrates, but also on efficient carbohydrate storage and  
51 remobilization. Carbohydrates are supplied by fixing atmospheric carbon (C)  
52 during photosynthesis and are vital in almost all plant physiological processes  
53 including the maintenance of existing tissue, the formation and enlargement of  
54 organs, and all associated metabolic processes. Investigating ecosystem  
55 productivity under changing climatic conditions requires understanding how  
56 various physiological and environmental factors constrain plant growth. In this  
57 regard, studies of non-structural carbohydrates (NSC; defined here as soluble  
58 sugars + starch) have been widely used to assess the source-sink balance of  
59 trees ([Barbaroux and Bréda 2002](#); [Damesin and Lelarge 2003](#); [Fischer and  
60 Holl 1992](#); [Gruber et al. 2012](#); [Michelot et al. 2012](#); [Oberhuber et al. 2011](#);  
61 [Woodruff and Meinzer 2011](#)). Stored NSC<sub>s</sub> can be viewed as reservoirs  
62 refilled when C demand for growth and maintenance is low and called upon  
63 during periods of high C requirements. Under this functional interpretation of  
64 NSCs, accumulation of large C pools is inconsistent with the hypothesis of  
65 carbon supply limiting growth, and rather suggests “sink” limitations ([Hoch and  
66 Körner 2012](#); [Körner 2003](#); [Millard et al. 2007](#)). Large pools of stored C have  
67 been observed in mature coniferous and deciduous trees with, for example, C  
68 reserves in deciduous trees sufficient to replace the entire canopy several

69 times (Hoch et al. 2003). Trees growing under conditions of high  
70 environmental stress such as towards their cold thermal limits of growth  
71 (Fajardo et al. 2013; Hoch and Körner 2003; Hoch and Körner 2012; Hoch et  
72 al. 2003), following severe water stress (Breda et al. 2006; Gruber et al.  
73 2012), or defoliation (Hoch 2005; Palacio et al. 2008), also showed high levels  
74 of stored NSCs associated with, or despite, reduced growth.

75 However, in the context of on-going climate change and concurrent increases  
76 in tree mortality, the role of carbon allocation has been revisited with several  
77 studies concluding that significant amounts of mobile carbohydrates in various  
78 tree tissues may not be simply interpreted as a sink-limitation (McDowell and  
79 Sevanto 2010; Millard and Grelet 2010; Ryan 2011; Sala et al. 2010; Wiley  
80 and Helliker 2012). Data obtained by interrupting the supply of new  
81 photosynthates via phloem girdling (BhupinderpalSingh et al. 2003), or  
82 following girdling and defoliation (Hoch 2005), has revealed significant NSC  
83 pools that remain unused by trees. This may reflect a condition intermediate  
84 to the “source” and “sink” limitations. Namely, trees appear physiologically  
85 capable of incorporating stored photosynthates into permanent tissues, yet do  
86 not do so. It is still unclear to what extent these data (i) indicate the inability of  
87 trees to supply the growing tissues with carbohydrates, (ii) reflect a  
88 prioritization of resources as a safety margin in the face of environmental  
89 stochasticity, or (iii) some other underlying mechanism. Regardless of the  
90 cause, active carbon storage competing with growth has been evidenced in  
91 different species (Chapin et al. 1990; Genet et al. 2010; Silpi et al. 2007).  
92 Consequently, if a significant fraction of the C pool is actively stored (thus  
93 competing with growth) or sequestered (i.e., unavailable for any further

94 physiological processes), then the observed overabundance of C in trees is  
95 not a useful indicator for a sink limitation (Millard and Grelet 2010). Therefore,  
96 under a long-term perspective, competing requirements for NSCs (e.g.  
97 respiration, defense and export, maintenance of hydraulic integrity) might  
98 cause a source limitation (Epron et al. 2012; Hartmann et al. 2013; Sala et al.  
99 2012; Wiley and Helliker 2012).

100 Despite abundant literature on NSCs, the processes and pathways related to  
101 NSC allocation and storage within trees remain poorly understood (Epron et  
102 al. 2012; Sala et al. 2012; Wiley and Helliker 2012). Progress is hampered by  
103 the scarcity of field data necessary for model testing, with studies in natural  
104 mature forests particularly needed (Barbaroux & Breda, 2002; Hoch et al.,  
105 2003; Gough et al., 2009, Richardson et al. 2013). NSC concentrations have  
106 been measured in diverse tree organs including stems, branches, foliage and  
107 roots (Barbaroux and Bréda 2002; Damesin and Lelarge 2003; Fischer and  
108 Holl 1992; Gruber et al. 2012; Michelot et al. 2012; Oberhuber et al. 2011;  
109 Ugglá et al. 2001; Woodruff and Meinzer 2011). In these studies, a general  
110 conclusion was that high variation in intra-annual NSC content was observed  
111 nearer to sites of active growth (e.g., apical and root meristems), while low  
112 variation was recorded within the storage tissues (e.g., sapwood, coarse  
113 roots, ray parenchyma). Investigations at the primary sink location would be  
114 expected to yield better insights into the seasonal carbohydrate supply and  
115 demand in trees.

116 Due to technical challenges associated with the sampling and isolation of  
117 NSC in the cambial zone, very few studies have reported on carbon stock

118 measurements where secondary growth occurs ([Deslauriers et al. 2009](#);  
119 [Giovannelli et al. 2011](#); [Sundberg et al. 1993](#); [Uggla et al. 2001](#)). The cambial  
120 zone in tree stems is composed of a thin layer of meristematic cells and only  
121 recently has a procedure been developed to solve the challenge of separating  
122 this tissue for NSC extraction ([Giovannelli et al. 2011](#)).

123 Here we use this new procedure to contribute to the debate on carbon  
124 dynamics in trees by supplying a detailed description of the intra-annual  
125 carbon fluctuation (soluble NSCs and starch content) in the cambial zone of  
126 mature conifer trees. We aim at elucidating the mechanisms controlling  
127 growth and at better understanding the effective sink strength of the cambium  
128 and its variability over time. Therefore we perform measurements in a  
129 deciduous and an evergreen conifer species at specific phenophases of wood  
130 formation, as well as at two sites at contrasting elevations. In addition, we  
131 quantify the individual carbon sugars as glucose, fructose, sucrose, raffinose,  
132 pinitol, and starch to better assign a functional meaning to the seasonal  
133 variations in NSC concentrations. This design allows us to address specific  
134 questions related to growth limitation and carbon demand, such as (i) Is  
135 secondary growth limited by the availability of carbon or constrained by  
136 environmental conditions acting upon the sink function? (ii) How does carbon  
137 supply in the cambium respond to the carbon demand for stem growth and  
138 concurrent foliage production? (iii) Is there a species- and/or site-specific?  
139 Strategy in the production, storage and supply of NSCs/individual sugars in  
140 terms of carbon allocation to growth, management of reserves, and protection  
141 against environmental stress?

142

143 **MATERIAL AND METHODS**144 *Study sites and field activities*

145 Our study was performed in the Lötschental (46°23'40"N, 7°45'35"E), a  
146 southwest-northeast oriented inner-alpine valley in the central Swiss Alps. The  
147 valley bottom is surrounded by steep forested slopes primarily composed of  
148 mixed, evergreen Norway spruce (*Picea abies* Karst.) and deciduous  
149 European larch (*Larix decidua* Mill.). The climate of the region is cool and  
150 relatively dry, with a mean annual temperature of 6°C, ranging from -3°C  
151 (January) to 15°C (July) and a mean annual precipitation exceeding 800mm  
152 (data from MeteoSwiss for the period 1987-2006).

153 Field activities were conducted in 2010 at two sites, about 1km apart from  
154 each other, with contrasting elevations. The high elevation site is located at  
155 the upper tree line at ~2200 m asl, on a south-facing slope and consists solely  
156 of larch (this site is abbreviated S22L to reflect the aspect (**S**outh), elevation  
157 (**2200** asl), and species (**L**arch). The low elevation site is near the valley  
158 bottom close to the north-facing slope at 1300 m asl and is a mixture of larch  
159 and spruce (similarly abbreviated N13L and N13S). The mean temperature  
160 difference between sites, as monitored from April to October 2010, was 3.5°C  
161 (average maximum and minimum temperature difference of 6 and 2.5°C,  
162 respectively). Hydrological conditions are generally dryer at the low elevation  
163 site due to a combination of less precipitation and higher evaporative demand.  
164 The soils of both sites are ~60 cm deep podzolic cambisols characterized by

165 significant coarse stone content and low clay amounts. Field activities  
166 involved i) collecting stem samples to follow NSC dynamics in cambial tissue,  
167 and ii) weekly monitoring of foliar and wood formation to document the  
168 progress of growth at the time of cambial sampling.

169

#### 170 *NSC sampling and biochemical analysis*

171 NSC sampling was performed on five different dates on 45 mature trees in  
172 total, with 15 trees per species and site (S22L, N13L, and N13S). Sampling  
173 dates were selected to target five relevant phases of annual ring formation,  
174 i.e., 1) when cambial division / and earlywood cell enlargement are highly  
175 active, 2) when earlywood cells are in both phases, enlargement and wall  
176 thickening, 3) after cellular division has stopped but enlargement and wall  
177 thickening phases continue, 4) when only latewood cells are conducting  
178 secondary wall thickening and, 5) during dormancy of cambium ([Table 1](#)). The  
179 sampling dates were estimated based upon data from 2007-2009 (King et al.  
180 in. prep.; Moser et al. 2010).

181 Two samples per tree and sampling date were taken at about 50 cm height  
182 using 37 mm diameter metal punchers. The samples, comprising phloem,  
183 cambium and xylem, were kept on ice during fieldwork, stored at -22°C once  
184 in the laboratory, and freeze-dried. Subsequently the samples were prepared  
185 for biochemical analysis according to the protocol described in [Giovannelli et](#)  
186 [al. \(2011\)](#). Accordingly, samples were split along the tangential plane in the  
187 cambial zone and then the differentiating phloem and cambial cells from the



188 phloem side sample were gently scraped with a razor blade to obtain a  
189 powder from the cambial tissue. Due to the low amount of cambium powder  
190 per sample, blocks of five trees per site and species were pooled to obtain  
191 enough material for sugar extraction. After pooling and homogenizing an  
192 equal amount of cambium powder per tree, a 40 mg subsample was used for  
193 sugar extraction. NSCs were extracted from the cambial powder using  
194 chemical procedures described in [Giovannelli et al. \(2011\)](#). The sugar content  
195 was determined by High-Performance Liquid Chromatography (HPLC)  
196 analysis equipped with a SHODEX SUGAR Series SC 1011 8x300mm  
197 column (Showa Denko, Germany) preceded by a pre-column Guard Pak  
198 Insert Sugar Pak II (Waters). The mobile phase was water, Milli Q grade, at  
199 0.5 ml min<sup>-1</sup>. Soluble carbohydrate identification was verified using  
200 carbohydrate standards (Sigma, USA) and quantified by means of an internal  
201 standard. Concentrations of glucose, fructose, sucrose, pinitol and raffinose  
202 were thus obtained. The remaining pellet after soluble NSC extraction was  
203 used for starch quantification. The starch content was measured after an  
204 extraction procedure: the residual pellet was suspended in 1.5 ml acetate  
205 buffer (pH 5), brought to boil at 100°C for 1h in a sand bath, and then cooled  
206 at room temperature. After incubation at 55°C for 16h with 150 µl  
207 amyloglucosidase from *Aspergillus niger* (Fluka), samples were diluted with  
208 distilled water to 5 ml and three 0.25 ml aliquots of each sample were assayed  
209 colorimetrically using glucose oxydase (Sigma-Aldrich, Italy).

210 Seasonal changes in soluble NSCs and starch content were compared  
211 between elevation and species using repeated-measures analysis of variance  
212 (ANOVA<sub>r</sub>) ([Gumpertz and Brownie 1993](#); [von Ende 1993](#)). For the within-

213 subject analysis, a Huynh-Feldt corrected probability was used to overcome  
214 the sphericity assumption in the case of univariate repeated-measures  
215 analysis (von Ende 1993). Differences were considered significant at  $P < 0.05$ .  
216 When significant effects were found, mean comparisons by sampling date  
217 using ANOVA were performed to identify when the differences occurred. All  
218 ANOVA analyses were performed using the JMP® 8.0 software (SAS Institute  
219 Inc.).

#### 220 *Monitoring of foliar and wood formation*

221 Foliar and cambial phenology were monitored on a weekly basis over the  
222 entire growing season 2010. For this purpose, and to prevent any potential  
223 influence of the sugar sampling on growth, 12 additional trees were selected:  
224 four spruce and larch trees at N13 and four larch trees at S22. The date of  
225 budburst for each tree was defined when 50% of its buds were broken  
226 (Brügger and Vassella 2003). Exact dates were estimated by linear  
227 interpolation, with these values averaged per site and species for a site date.

228 Cambial phenology was quantified weekly on the same trees. The forming  
229 annual ring was monitored for each tree by analysing tracheid formation from  
230 microcores collected weekly between April and November after (Moser et al.  
231 2010). Microcores were collected from the stem at 1-2 m height using a  
232 Trephor (Rossi et al. 2006a) preferentially perpendicular to the slope direction  
233 to avoid reaction wood. Sampling was conducted along an oblique line and 3–  
234 5 cm apart to minimize wound reactions caused by earlier samplings (Forster  
235 et al. 2000). Microcores were placed for 24 h in a dilution of acetic acid and  
236 ethanol to preserve forming cells from degradation, and then stored in a 70%

237 alcohol solution. Samples were prepared for cellular analysis by cutting 20–30  
238  $\mu\text{m}$  thick transversal microsections using a sliding microtome. Microsections  
239 were stained with safranin and astrablue and fixed to microscope slides with  
240 Canada balsam. Ring formation was analysed at a magnification of 400–600x  
241 and the number of tracheids in the different phases of cell development (i.e.  
242 enlargement, wall thickening and maturity) assessed by averaging the  
243 counting along three radial files of each microsection. Enlarging cells were  
244 characterized by thin primary cell walls with radial diameter roughly two or  
245 more times larger than that of dividing cambial cells. Polarized light was used  
246 to discriminate between enlarging and wall thickening tracheids. Mature cells  
247 were recognized by completely lignified secondary walls and empty cell  
248 bodies (Rossi et al. 2006b; Rossi et al. 2007). Dates representing critical  
249 phenological stages of wood formation were calculated for each site and  
250 species based upon the cell counts.

251

## 252 **RESULTS**

### 253 *Growth dynamics and timing of NSC sampling*

254 At the valley bottom, larch budburst occurred on DOY  $135 \pm 3.5$  (mean  $\pm$   
255 standard deviation) and xylem formation (defined in this work as the timing of  
256 the first observed enlarging xylem cells) on DOY  $138 \pm 3$ . Budburst and xylem  
257 formation in spruce occurred later, i.e. on DOY  $149 \pm 3.5$  and DOY  $143 \pm 0$ ,  
258 respectively. Budburst in larch occurred before or close to the first  
259 observations of xylem cells entering the enlargement phase. In contrast,  
260 spruce initiated growth of new xylem tracheids before the emergence of

261 current-year needles. These observations indicate that spruce and larch adopt  
262 different sequential arrangements in the timing of foliar and xylem growth  
263 resumption. At S22L, in comparison to larch growing 900m lower or at a 3.5°C  
264 warmer site, budburst was delayed by about two weeks (DOY  $149 \pm 3.5$ ) and  
265 xylem formation onset by about three weeks (DOY  $159 \pm 3.5$ ; (Figure 1, Table  
266 2). The timing of growth resumption significantly differed (Table 3) between  
267 species ( $\sim 6$  days;  $P=0.01$ ) and elevation ( $\sim 22$  days;  $P<.0001$ ).

268 The onset of wall thickening, the formation of fully mature cells, as well as the  
269 maximum number of cells observed in the phases of enlargement and wall  
270 thickening occurred earlier at N13S than N13L (Figure 1). The first wall  
271 thickening and mature cells were both observed 8 days later for spruce.  
272 Despite their delay in the onset of wall thickening (9 days) and mature cells  
273 (12 days), S22L soon reached similar levels of cell production as found at  
274 N13L. The total number of xylem cells in the 2010 ring varied among larch  
275 (ranging from 10 to 30) and spruce (from 25 to 70), but was in general higher  
276 for spruce.

277 The differences between both species and elevation decreased toward the  
278 end of the growing season. Significant differences in the end of xylogenesis  
279 (DOY 308 in average) between N13L and N13S disappeared, although larch  
280 at the treeline stopped xylogenesis a few days earlier. Thus, the shorter total  
281 duration of xylogenesis at S22L compared to N13L (18 days,  $P=0.0004$ ) and  
282 for N13S compared to N13L (8 days,  $P=0.02$ ) was mainly due to a difference  
283 in the onset of xylem differentiation (Table 3).

284 *NSC concentrations*

285 Significant seasonal variations of NSCs in the cambial zone were observed  
286 (Figure 2). In general, the soluble fraction peaked between July and August,  
287 corresponding to the times of high rates of cell division and enlargement  
288 (period 2) and when many cells were in the wall thickening phase (period 3;  
289 Table 1). Total soluble NSC concentrations increased by more than 50%  
290 between the onset of the growing season and the period of maximum cell  
291 division. In September, during latewood cell wall thickening (period 4) the  
292 soluble carbohydrate concentrations decreased, and increased again during  
293 the subsequent dormant season (period 5). Larch and spruce displayed  
294 similar seasonal variations, with particularly high NSC concentrations during  
295 periods 2 and 3, but the NSC concentrations in spruce peaked during the  
296 dormant season. S22L and N13L showed similar seasonal patterns.

297 The starch concentrations for all sites and species were high towards the end  
298 of the growing season (periods 3 and 4) and decreased dramatically during  
299 dormancy. However, lower starch concentrations were measured at N13L  
300 during the early growing season (period 1) .

301 On average, cambium sugars consist of around 40% glucose, 35% fructose,  
302 10% starch, 10% pinitol, 5% sucrose and less than 1% raffinose (Figure 2).  
303 However, these proportions slightly vary in time, between species and  
304 elevations. Total NSC concentrations closely follow those of glucose and  
305 fructose together, showing similar patterns and accounting for nearly up to  
306 80% of the growing season soluble NSCs (Figure 2) and leading to high  
307 hexose (glucose + fructose) to sucrose ratios (in average from 15 to 30 during

308 the growing season). A fructose to glucose ratio of approximately 1 was also  
309 observed throughout the whole growing season.

310 Glucose, fructose and sucrose concentrations for spruce tend to be lower  
311 during the growing season and higher during the dormant season, however  
312 fructose concentration was not found to be as species dependant as the other  
313 sugars (Table 4). Notably, both raffinose and pinitol displayed unique intra-  
314 seasonal changes in concentrations. For all species and elevations raffinose  
315 was only found during the dormant season (period 5), whereas pinitol, in  
316 contrast to all other sugars did not show an increase during the dormant  
317 season. In addition, pinitol was the only sugar with higher concentrations in  
318 spruce during the active growing season (periods 2-3).

319 Highly significant relationships between species and the phases of cambial  
320 phenology were observed for starch, glucose, sucrose, and raffinose, while  
321 elevation was not a significant factor contributing NSC concentration  
322 variations in the cambial zone (Table 4).

323

## 324 **DISCUSSION AND CONCLUSIONS**

325 Our intra-annual NSC measurements sampled directly in the stem cambial  
326 zone of mature trees growing in the subalpine zone improve the  
327 understanding of carbohydrate variation during a complete annual cycle. The  
328 dynamics of the different mobile carbohydrates observed at both elevations  
329 and for the two species reflect the changing requirements for storage,  
330 mobilisation and use of C resources needed to sustain growth as well as

331 protecting vital tissue from harsh environmental conditions, e.g., during the  
332 winter.

333

334 *The annual cycle*

335 In temperate regions with a distinct seasonal cycle and a dormant period for  
336 vegetation in winter, the onset of wood formation is usually temperature driven  
337 (Moser et al. 2010). Photoperiod may also provide secondary control for the  
338 growth onset, at least for the primary meristem (Chuine et al. 2010; Körner  
339 and Basler 2010). Wood formation is terminated in late fall when the chain of  
340 maturation processes is completed (Rossi et al. 2012). These notions of the  
341 annual cycle are broadly reflected in the cellular developmental stages  
342 (Figure 1) and concentrations of the different types of NSCs (Figure 3) for  
343 both larch and spruce

344 The general paralleling of the NSC compounds in the cambium and the  
345 annual dynamics of wood formation has been also observed in the other few  
346 studies with comparable approaches, i.e. for poplars (Deslauriers et al. 2009;  
347 Giovannelli et al. 2011), Scots pines (Sundberg et al. 1993; Uggla et al. 2001)  
348 and eucalyptus (Stewart et al. 1973). Collectively, these studies suggest that  
349 the dynamics of NSC concentrations in the cambium of different species,  
350 habitats, and angiosperm versus gymnosperm lineages follow a similar  
351 seasonal pattern

352 During the growing season, NSCs sustain all metabolic processes involved in  
353 the formation of new cells within the cambium. We observed very high

354 amounts of soluble sugars, (primarily glucose and fructose, collectively named  
355 hexose) the cambial zone of both larch and spruce. While lower at the  
356 beginning and the end of the growing season, soluble sugar concentration  
357 increased rapidly and peaked when the resource demand was higher, i.e.  
358 when a greater number of cells were both in the enlargement and cell wall  
359 thickening phases. Such high levels of glucose and fructose are unusual  
360 compared to what is normally observed in other tissues such as stem wood  
361 ([Damesin and Lelarge \(2003\)](#), [Gruber et al. \(2011\)](#), [Streit et al. \(2013\)](#)). In our  
362 study of larch and spruce, we found similar concentrations of fructose and  
363 glucose within the cambial zones. Glucose to fructose ratios approximately  
364 equal to unity were similarly reported for Scots pine by [Uggla et al. \(2001\)](#),  
365 who also observed strongly decreasing sucrose concentration gradients (yet,  
366 strongly increasing glucose and fructose levels) from functional phloem to  
367 developing xylem. Relative to sucrose, high levels of hexose within the  
368 cambial zone are consistent with the high metabolic activity of the dividing and  
369 rapidly growing cells.

370 In addition to serving as the building blocks for growth itself, sugars play an  
371 important role as signalling molecules and/or as global regulators of gene  
372 expression ([Eveland and Jackson \(2011\)](#), [Koch \(2004\)](#)). Glucose and fructose  
373 in spruce and larch likely originated from the cleavage of sucrose as  
374 suggested by high levels of sucrose cleaving enzymes such as AI (acid  
375 invertase), and to a lesser extent Susy (sucrose synthase), in the cambial  
376 zone of Scots pine ([Uggla et al. 2001](#)). The relative ratio of hexose to sucrose  
377 concentrations are maintained by these various enzymes which collectively  
378 coordinate and fine-tune growth during key phases of development ([Eveland](#)



379 [and Jackson 2011; Koch 2004](#)). Hexose is regarded to have a greater  
380 signalling potential in promoting growth and cell proliferation whereas sucrose  
381 is typically associated with differentiation and maturation ([Eveland and](#)  
382 [Jackson 2011; Koch 2004](#)). While we did not measure enzyme  
383 concentrations, we found that particularly the hexose to sucrose ratio in the  
384 cambium zone of spruce and larch generally decreased during the growing  
385 season, as the cambial phenology shifted from rapidly dividing cells peaking  
386 around DOY 165 (10.06.2010) to the wall-thickening phases peaking  
387 approximately 2 months later (**Figure 1**).

388 Starch concentration commonly shows considerable seasonal variation in the  
389 stem and branches of temperate zone trees. Lower reserves at budburst, a  
390 late summer maximum as growth slows down, and starch hydrolysis to sugar  
391 in autumn when days are short and nights cold are all patterns previously  
392 reported ([Gruber et al. 2011; Kozłowski 1992](#)). Decreasing levels of starch  
393 during the summer in tree stem, branches and cambial zone have also been  
394 observed ([Deslauriers et al. 2009; Hoch et al. 2003; Sundberg et al. 1993](#)). In  
395 comparison to reports from poplar ([Deslauriers et al. 2009](#)) or Scots pine  
396 ([Sundberg et al. 1993](#)) cambial zones, we found starch levels in the cambial  
397 zone of spruce and larch at all three sites remained relatively constant during  
398 the growing season (**Figure 3**). Similarly, [Geisler-Lee et al. \(2006\)](#) found little  
399 expression of genes related to starch metabolism in comparison to  
400 carbohydrate-related enzymes (e.g. Susy, cellulose synthase) in the cambium  
401 of poplar during xylogenesis.

402 Large amounts of starch are consumed during cambial reactivation, with  
403 reserves replenished only sometime after the onset of xylem differentiation  
404 (Begum et al. 2013). The concentrations we observed tended to be lower than  
405 those reported for Scots pine (Sundberg et al. 1993) and higher than those  
406 from poplar (Deslauriers et al. 2009). Our first sampling campaign (DOY 155 -  
407 N13, DOY 178 - S22) was possibly too late to catch the minimum starch level  
408 as cambium reactivation, xylem differentiation, and budburst had already  
409 occurred. During reactivation of the cambium, starch is used as the main  
410 source of energy, but later on, the continuation of cambial activity seems to  
411 require a continuous supply of sucrose (Oribe et al. 2003) for cell wall  
412 biosynthesis. The low variation of starch during the growing season suggests  
413 a constant supply of fresh assimilates to the cambium of larch and spruce at  
414 our sites.

415 For all of the tree groups investigated in our study, we found starch  
416 breakdown during the cold season, and synthesis from soluble sugars in late  
417 winter-springtime (Figure 3). Our findings support previous observations of  $\alpha$ -  
418 amylase activation and starch synthase genes in dormant poplar tissues  
419 during cold periods supporting starch breakdown for cryoprotection purposes  
420 (Geisler-Lee et al. 2006). Similarly, resynthesis of starch in late winter  
421 (Kozlowski 1992), was documented in needles (Bansal and Germino 2009;  
422 Chen et al. 2012; Hansen and Beck 1994; Hoch et al. 2003) and in the trunk  
423 (Fischer and Holl 1992; Hansen and Beck 1994; Hoch et al. 2003; Michelot et  
424 al. 2012) of various tree species. Our results demonstrate that these starch  
425 dynamics also apply to the cambial region.

426 Raffinose, and pinitol are both compatible solutes (i.e. osmotically active  
427 compounds) that help cells survive osmotic stress (Bachmann et al. 1994;  
428 Bohnert and Shen 1998). While increased concentrations of raffinose and  
429 pinitol can decrease the osmotic potential of cells to maintain the water  
430 balance, their main function might be to stabilize proteins, protein complexes,  
431 or membranes by scavenging radical oxygen species (ROS) that build up  
432 during environmental stress (e.g. cold, drought, high salinity) (Bohnert and  
433 Shen 1998; Orthen et al. 1994). Levels of raffinose increased during winter  
434 (Figure 3), thereby presumably protecting cell membranes from damage  
435 during frost-induced dehydration by detoxifying ROS that accumulate at low  
436 temperatures (Nishizawa et al. 2008). Pinitol was present year-round in both  
437 species, however, concentrations peaked in the cambial zone of spruce when  
438 growth processes were most active Streit et al. (2013) observed higher pinitol  
439 concentrations in branch bark and branch wood of larch growing at another  
440 tree line location in Switzerland in comparison to larch from lowland (500 m  
441 asl) and interpreted this difference in terms of long-term adaptation to high  
442 levels of ROS in response to low temperature. Similar pinitol concentrations to  
443 Streit et al. (2013) in larch growing both at the valley bottom (1300 m asl) and  
444 the tree line (2200 m asl) suggest that long-term adaptation responses to high  
445 levels of ROS already occur at 1300 m asl and that pinitol concentrations may  
446 not increase linearly with elevation. The conclusions for greater environmental  
447 stress towards the upper elevations and the valley bottom are similarly  
448 supported by analyses of the climatic sensitivity of radial growth variations 449  
449 along an ~900 meter elevational transect (King et al. 2013b).

450

451 *Species and climatic controls on NSC*

452 The general trends in the dynamics of NSC concentrations in this study were  
453 similar between species and elevations. Differences relate primarily to the  
454 absolute concentrations: the protective sugars raffinose (during dormancy)  
455 and pinitol (during the warmest period corresponding to July/August), as well  
456 as glucose, fructose and sucrose (during dormancy) differed in spruce and  
457 larch. The species-specific concentrations are potentially explained by  
458 differing climatic sensitivities of larch and spruce, also observed in an Europe-  
459 wide multi-species tree-ring network ([Babst et al. 2013](#)) to cold (in winter) and  
460 drought (in summer). [Hinesley et al. \(1992\)](#) observed links between the  
461 raffinose content in the foliage of different conifers with their level of cold  
462 hardiness. A similar relationship was also found for the presence of other  
463 NSCs (glucose, fructose, sucrose and raffinose) in the shoots of shrubs and in  
464 tree stem ([Lee et al. 2012](#); [Morin et al. 2007](#)). Trees with thinner bark like  
465 spruce are more sensitive to frost damage in the cambium zone ([Gurskaya  
466 and Shiyatov 2006](#)) and might thus need a different strategy for additional  
467 protection. Higher levels of soluble NSCs in the cambial zone of spruce, in  
468 comparison to larch, in a period where the risk of freezing injuries is high  
469 would meet this requirement. The lower level of raffinose in particular, and  
470 total soluble NSCs in general, observed in larch might also reflect a higher  
471 degree of spring de-hardening compared to spruce in early March (sampling  
472 period 5). This would further be supported by the earlier bud break and onset  
473 of xylem differentiation of larch.

474 During June and July, pinitol concentrations were found to be higher in spruce  
475 compared to larch (in average 1.5- to 2.5-fold) at the valley bottom, while  
476 glucose levels were lower. Glucose has been described as one of the  
477 precursors of pinitol synthesis (Obendorf et al. 2008) and therefore the lower  
478 levels of glucose relative to pinitol in in spruce suggest that some glucose was  
479 directed towards pinitol synthesis. However, why the pinitol levels significantly  
480 increased in the cambial zone of spruce during the summer months, while  
481 remaining constant (and at similar levels) in larch at both the valley and tree  
482 line sites, is uncertain. These patterns might reflect a greater need to maintain  
483 turgor potential (Aranjuelo et al. 2011) within the cambial zone of spruce to  
484 sustain cell enlargement, or may result from a higher sensitivity of spruce to  
485 environmental stress leading to enhanced generation of ROS (Orthen et al.  
486 1994). A recent investigation showed that spruce growing at our study site  
487 exploit their internal stem water reserves more quickly during drier conditions  
488 compared to larch (King et al. 2013).

489 While total NSC concentrations remained similar throughout the growing  
490 season in our study, higher concentrations of the soluble sugar fraction  
491 (except pinitol) during dormancy were observed in spruce compared to larch.  
492 While winter respiration of living tissue of the dormant deciduous trees depend  
493 exclusively on reserves (starch + soluble sugars), the higher levels in our  
494 evergreen species might reflect photosynthesis (Kozłowski 1992) and phloem  
495 transport (Blechs Schmidt-Schneider 1990) during mild winter days.

496 Despite a temperature difference of 3.5 °C and difference in the length of the  
497 growing season of 30 days between the treeline and valley bottom larch sites,

498 we found similar seasonal variations in the NSC concentrations. This similarity  
499 likely has to do with our sampling keyed into expected cambial activity rather  
500 than only calendar dates. Notable differences between elevations occurred  
501 only for starch concentrations in the early growing season (period 1) although  
502 no statistical significant differences were found (Table 4). These  
503 concentrations were already near their maximum in early summer at the  
504 treeline whereas at the valley bottom they only peaked at the beginning of  
505 July. Attributing these differences to particular mechanisms remains  
506 speculative. However, higher respiration losses due to warmer temperatures  
507 at the valley bottom in addition to active growth of competing sinks while  
508 needles are still not fully photosynthetically functional ([Kozłowski 1992](#)) would  
509 be consistent with our intra-seasonal NSC data.

510

511 *Is secondary growth allocation carbon or sink limited?*

512 Our data, although focused to one growing cycle and limited to only the  
513 cambial zone, contribute to a better understanding of the potential functions  
514 on NSC allocation and storage within trees. Notable for our investigation are  
515 the direct observations of NSC concentration at the sites of secondary  
516 growth, and the ability to link these changes with cambial zone phenology.  
517 The reduced growth (i.e. number of tracheids produced) observed for larch in  
518 comparison to spruce growing at the same site and for S22L compared to  
519 N13L, cannot be explained by temporary limitation in NSC supply in the  
520 cambium. The high similarities in the NSC concentrations during the growing  
521 season of trees at different elevations, together with the delayed start of

522 cambial activity and xylem cell production at the tree line, support the  
523 hypothesis of [Körner \(2003\)](#) that tree line trees are not carbon limited. Factors  
524 affecting carbohydrate conversion to new tissues, rather than carbohydrate  
525 availability, seem to control cambial activity (Kozlowski 1992; Sundberg et al.  
526 1993). Similarly, [Hoch et al. \(2002\)](#) observed higher levels of mobile carbon  
527 pools in needles, branches, stems and roots of trees with increasing elevation  
528 and conclude that sink activity of trees growing at high elevations was limited  
529 by low temperature rather than carbon availability. Similar conclusions were  
530 reached by [Fajardo et al. \(2013\)](#) who compared deciduous and evergreen  
531 treeline species across elevational gradients. Although no significant increase  
532 of NSC concentration in the cambium zone was observed with elevation in our  
533 study, we did observe a higher contribution of starch to the total NSC pool in  
534 larch at the tree line ( $P=0.03$ , data not shown). Our data thus support growth  
535 limitation at the tree line in response to sink activity rather than inability of  
536 trees to supply growing tissues with carbohydrates.

537 In conclusion, the first study of intra-seasonal NSC dynamics at the cambial  
538 level in subalpine forests indicated that carbohydrate fluctuations at the sink  
539 level are closely linked to cambial activity (carbon sink demand) and the  
540 metabolic needs associated with cell formation, rather than to species or  
541 climate (elevation). Variation in the concentration of NSC with more specific  
542 functions such as raffinose or pinitol seemed to be, however, species  
543 dependant. Observations on larch growing across an elevational gradient of  
544 nearly 1000 m suggest that temperature limits the cambial activity of tree and  
545 not the availability of carbohydrates within the cambium.

546

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555

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751 a tall conifer. *Plant Cell and Environment*. 34:1920-1930.

752

753

754 **TABLES AND FIGURES**

755

756 **Table 1:** Days of the year (DOY) and corresponding dates of NSC sampling  
757 for each related phenological stages and sites

758

759

760 **Table 2:** Onset and duration of the growing season, and budburst for larch  
761 growing at the valley bottom and the treeline (1300 asl and 2200 m asl,  
762 respectively) and spruce growing at the valley bottom. Numbers in  
763 parenthesis are the standard deviation of the mean (n=4).

764

765 **Table 3:** ANOVA (*F*- and *P*-values) of the onset, end and duration of xylem  
766 differentiation, and budburst, for differences between species and elevation.  
767 Significance at ( $P < 0.05$ ).

768

769 **Table 4:** ANOVA (*F*- and *P*-values) for testing the differences in the  
770 carbohydrates compounds. Tests include differences between species (S),  
771 period (P) and their interaction (S\*P); and for elevation (E), period (P) and  
772 their interaction (E\*P). *P*-values for repeated-measures ANOVA are presented  
773 with Huynh-Feldt corrected probabilities. Significance at ( $P < 0.05$ ).

774

775 **Figure 1:** Number of cells in the cambial zone (CZ), enlarging phase (E),  
776 secondary wall formation and lignification phase (W) and mature xylem cells  
777 (M) of individual *Larix decidua* (blue -1300m; grey -2200m; n=4) and *Picea*  
778 *abies* (orange -1300m; n=4) at both elevations in the Lötschental as counted  
779 on a weekly basis during the growing season 2010. The average number of  
780 cells during the season is shown. The date of budburst (Bb) for both species  
781 at both sites is also indicated.

782

783 **Figure 2:** Concentration of non-structural carbohydrates (Fru=fructose,  
784 Glc=glucose, Suc=sucrose, Pin=pinitol, Raff=raffinose, Starch; mg g<sup>-1</sup> DW) in  
785 the cambial zone of larch and spruce at both elevation in the Lötschental  
786 during the growing season 2010. Mean values are presented (n=3; each  
787 representing 5 pooled trees). DW = dry weight. Stars indicate significant  
788 differences between *Larix decidua* and *Picea abies* at 1300 m.

789

790 **Figure 3:** Individual non-structural carbohydrates (glucose, fructose, sucrose,  
791 pinitol, raffinose:  $\mu\text{mol g}^{-1}$  DW; starch: mg g<sup>-1</sup> DW) concentrations in the  
792 cambial zone of larch (blue circles-full line - 1300m; grey circles-dotted line -  
793 2200m) and spruce (orange triangles-full line) at both elevations in the  
794 Lötschental during the growing season 2010. Values are mean  $\pm$  1 SE (n=3;  
795 each representing 5 pooled trees). Note the scale break for glucose and  
796 fructose. DW = dry weight. Stars indicate significant differences between *Larix*  
797 *decidua* and *Picea abies* at 1300 m.

**Table 1:** DOY (dates) of NSC sampling for each corresponding phenological stages and sites.

| Stage   | Period | N13              | S22              |
|---|--------|------------------|------------------|
|   |        | DOY (Date)       | DOY (Date)       |
| Highly active cambial division/EW cell enlargement    | 1      | 155 (31.05.2010) | 178 (23.06.2010) |
| EW cell enlargement and wall thickening               | 2      | 191 (05.07.2010) | 201 (15.07.2010) |
| No cell division but active wall thickening processes | 3      | 222 (04.08.2010) | 222 (04.08.2010) |
| LW cell wall thickening                               | 4      | 265 (15.09.2010) | 265 (15.09.2010) |
| Dormancy  | 5      | 69 (09.03.2011)  | 39 (08.02.2011)  |

*EW = earlywood, LW = latewood*

**Table 2:** Onset, duration of the growing season and budburst for *Larix decidua* growing at the valley bottom and the treeline (1300 m asl and 2200 m asl, respectively) and *Picea abies* growing at the valley bottom. Numbers in parenthesis are the standard deviation of the mean.

| Elevation (m) | Species      | Onset of xylem production (DOY) | End of xylem differentiation (DOY) | Duration of xylogenesis (days) | Budburst (DOY) |
|---------------|--------------|---------------------------------|------------------------------------|--------------------------------|----------------|
| 1300          | <i>Larix</i> | 138 (3.0)                       | 311 (0.0)                          | 173 (3.0)                      | 135 (3.5)      |
| 1300          | <i>Picea</i> | 143 (0.0)                       | 305 (6.9)                          | 162 (6.9)                      | 149 (3.5)      |
| 2200          | <i>Larix</i> | 159 (3.5)                       | 302 (6.0)                          | 143 (7.9)                      | 149 (3.5)      |



**Table 3:** ANOVA (*F*- and *P*-values) of the onset, end and duration of xylem differentiation, and budburst, for differences between species and elevation. Significance at ( $P < 0.05$ ).

| Source           | Onset of xylem differentiation |                 | End of xylem differentiation |                 | Duration of xylogenesis |                 | Budburst |                 |
|------------------|--------------------------------|-----------------|------------------------------|-----------------|-------------------------|-----------------|----------|-----------------|
|                  | <i>F</i>                       | <i>P</i> -value | <i>F</i>                     | <i>P</i> -value | <i>F</i>                | <i>P</i> -value | <i>F</i> | <i>P</i> -value |
| <b>Species</b>   | 13.44                          | *               | 3.00                         | ns              | 9.28                    | *               | 409.9    | ***             |
| <b>Elevation</b> | 85.00                          | ***             | 9.00                         | *               | 51.37                   | **              | 630.1    | ***             |

\*  $P < .05$ , \*\*  $P < .001$ , \*\*\*  $P < .0001$ , ns = non significant

**Table 4:** ANOVA (*F*- and *P*-values) for testing the differences in the carbohydrates compounds. Tests include differences between species (S), period (P) and their interaction (S\*P); and for elevation (E), period (P) and their interaction (E\*P). *P*-values for repeated-measures ANOVA are presented with Huynh-Feldt corrected probabilities. Significance at ( $P < 0.05$ ).

| Carbohydrates | Source | <i>F</i> | <i>P</i> -value | Source | <i>F</i> | <i>P</i> -value |
|---------------|--------|----------|-----------------|--------|----------|-----------------|
| Glucose       | S      | 5.85     | ns              | E      | 9.27     | ns              |
|               | P      | 29.03    | ***             | P      | 17.33    | ***             |
|               | S*P    | 7.64     | **              | E*P    | 2.81     | ns              |
| Fructose      | S      | 0.24     | ns              | E      | 0.04     | ns              |
|               | P      | 4.25     | *               | P      | 3.57     | *               |
|               | S*P    | 2.57     | ns              | E*P    | 3.56     | ns              |
| Sucrose       | S      | 2.76     | ns              | E      | 0.28     | ns              |
|               | P      | 60.53    | ***             | P      | 11.38    | **              |
|               | S*P    | 14.71    | ***             | E*P    | 1.38     | ns              |
| Raffinose     | S      | 100.76   | **              | E      | 0.24     | ns              |
|               | P      | 412.71   | ***             | P      | 59.73    | **              |
|               | S*P    | 100.76   | ***             | E*P    | 0.24     | ns              |
| Pinitol       | S      | 40.82    | *               | E      | 7.48     | ns              |
|               | P      | 8.37     | **              | P      | 1.78     | ns              |
|               | S*P    | 2.64     | ns              | E*P    | 2.9      | ns              |
| NSC total     | S      | 4.39     | ns              | E      | 0.53     | ns              |
|               | P      | 15.40    | ***             | P      | 9.74     | **              |
|               | S*P    | 3.47     | *               | E*P    | 2.49     | ns              |
| Starch        | S      | 2.11     | ns              | E      | 6.4      | ns              |
|               | P      | 20.37    | ***             | P      | 15.22    | **              |
|               | S*P    | 5.41     | **              | E*P    | 0.77     | ns              |

\*  $P < 0.05$ , \*\*  $P < 0.001$ , \*\*\*  $P < 0.0001$ , ns = non significant





