

Ionic Liquid-Based Ultrasound-Assisted Emulsification Microextraction Coupled with HPLC for Simultaneous Determination of Glucocorticoids and Sex Hormones in Cosmetics

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ABSTRACT

An effective and environmentally friendly method based on ionic liquid-based ultrasound-assisted emulsification microextraction (IL-USAEME) and high performance liquid chromatography (HPLC) has been developed for the determination of nine hormones including three glucocorticoids, one androgen and five progestogens in cosmetics. Several factors that affect the extraction efficiency, such as the kinds and volume of ionic liquid, salt concentration, ultrasonic power and time, and centrifugation time were investigated and optimized. Under the optimum extraction condition, the recoveries of analytes ranged from 85.97% to 108.84% except prednisolone (62.30%). The intraday and interday precision was below 2.51% and 3.30%, respectively.

Keywords: Ionic Liquid; Ultrasound-Assisted Emulsification Microextraction; Hormones; Cosmetics; High Performance Liquid Chromatography

1. Introduction

Cosmetics have become more and more important in our daily life for skin whitening, anti-wrinkle, removing freckle, moisture, and so on [1]. In order to achieve these effects remarkably, illicit chemicals such as hormones are added to the cosmetics. And the potential accumulation hazard of these additives is terrible [2]. Although the Hygienic Standard for Cosmetic of China and European Union Directive 76/768/EEC has banned the addition of hormones to commercial cosmetics, cosmetics containing hormones are still in the market.

At present, the high performance liquid chromatography and its combination with mass spectrometry are the most common methods for determination of trace hormones in the complex matrices. For these analytic techniques, in order to decrease the interference of matrix and concentrate the targeted compounds, a sample pretreatment is indispensable, which is often laborious and consumes a large amount of organic solvents [3-5].

Ionic liquids (ILs) composed of organic cations and various types of anions have been successfully applied to

various areas of analytical chemistry as environmental benign solvents [6-10]. Ionic liquid-based ultrasound-assisted emulsification microextraction (IL-USAEME) is an efficient, simple, rapid and cheap extraction technique, in which applying ultrasound radiation to facilitate the emulsification of a microvolume of ionic liquid extractant in an aqueous sample, and subsequent separation of the two immiscible liquid phases by centrifugation [11, 12]. This method has been used for the extraction of organic compounds in environmental samples. However, to the best of our knowledge, IL-USAEME has not been applied for the extraction of hormones from cosmetics.

In this paper, an ionic liquid-based ultrasound-assisted emulsification microextraction (IL-USAEME) combined with HPLC method for simultaneous determination of glucocorticoids and sex hormones in cosmetics will be investigated.

2. Materials and Methods

2.1. Reagents and Chemicals

The pharmaceutical standards (purity, 97.5% - 99.5%) of androstendione, hydroxyprogesterone acetate, megestrol

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acetate and melengestrol acetate were purchased from Dr. Ehrenstorfer GmbH (Augsburg Germany). Northindrone was purchased from Toronto Research Chemicals Inc. (North York, Canada). Prednisolone and Beclometasone Dipropionate were purchased from the National Institutes for Food and Drug Control (NIFDC) (Beijing, China). Dexamethasone was purchased from Zhejiang Xianju Pharmaceutical Co., Ltd. (Zhejiang, China) and 17 α -hydroxyprogesterone caproate was purchased from Tokyo Chemical Industry Co., Ltd. (Tokyo, Japan). Chromatographic grade methanol and acetonitrile were supplied by Merck (Darmstadt, Germany). Deionized water was purified by a Milli-Q water purification system (18 M Ω , Millipore, Bedford, MA, USA). [C₆MIM][PF₆] and [C₇MIM][PF₆] were gained from Chengjie Co. Ltd. (Shanghai, China) and used as received. The cosmetic products Kose were purchased from local supermarket and stored at room temperature.

2.2. Ionic Liquid-Based Ultrasound-Assisted Emulsification Microextraction

Ionic liquid-based ultrasound-assisted emulsification microextraction was performed in KQ-100 DA and KQ-500 DA ultrasonic water baths (Kunshan, Jiangsu, China). A 0.2 g cosmetic sample, 125 μ l [C₇MIM][PF₆], 62.5 μ l saturated NaCl aqueous solution and 187.5 μ l water were introduced to a 1.5 ml eppendorf centrifuge tube. And the mixed solution was ultrasonically extracted at ultrasonic power of 100 W for 15 min. The emulsification solution obtained after ultrasonication was centrifuged at 10000 rpm for 12 min by the Centrifuge 5804 (Eppendorf AG, Hamburg, Germany). After centrifugation, the IL extraction solution (~100 μ l) which was deposited at the bottom of the centrifuge tube was collected using a 200 μ l pipette and then diluted by methanol to 1ml. Finally, the solution was filtered through 0.22 μ m nylon filter membrane.

2.3. HPLC Analyses

The HPLC analysis was performed using a Waters 2695 HPLC system (Waters, Milford, MA, USA) equipped with a quaternary solvent delivery system, an auto-sampler with a 100 μ l sample loop, a PDA detector, a column oven, and a data station running the Empower data software. A CAPCELL PAK C18 column (150 mm \times 4.6 mm, 3 μ m, Chuo-ku, Tokyo, Japan) was used for separation and was maintained at 25 $^{\circ}$ C. The mobile phase was composed of water (eluent A) and acetonitrile (eluent B), and the elution was carried out in gradient mode at a flow rate of 0.7 ml/min. The gradient program was as follows: 0 min - 2 min, 30% B; 2 min - 6 min, 30% - 70% B; 6 min - 15 min, 70% B; 15 min - 16 min, 70% - 100% B; 16 min - 25 min, 100% B. Finally, the initial condition was re-introduced over 0.01 min, and the equilibration time was 5 min. The injection volume was 10 μ l. The detection wavelength was 242 nm for all the analytes except megestrol acetate and melengestrol acetate (288 nm).

3. Results and Discussion

3.1. Optimization of Extraction Conditions

To acquire the high recoveries and enrichment efficiency of the studied hormones (shown as **Figure 1**), the effects of several factors, such as kinds of ionic liquid, volume of [C₇MIM][PF₆], salt concentration, ultrasonic power and time, and centrifugation time, were examined and optimized.

3.1.1. Screening of the IL-Based Extraction Solvent

In the case of USAEME, the suitable extraction solvent should have density greater than that of water and also a low solubility in water [14]. And the structures of ILs have a significant effect on its physicochemical proper-

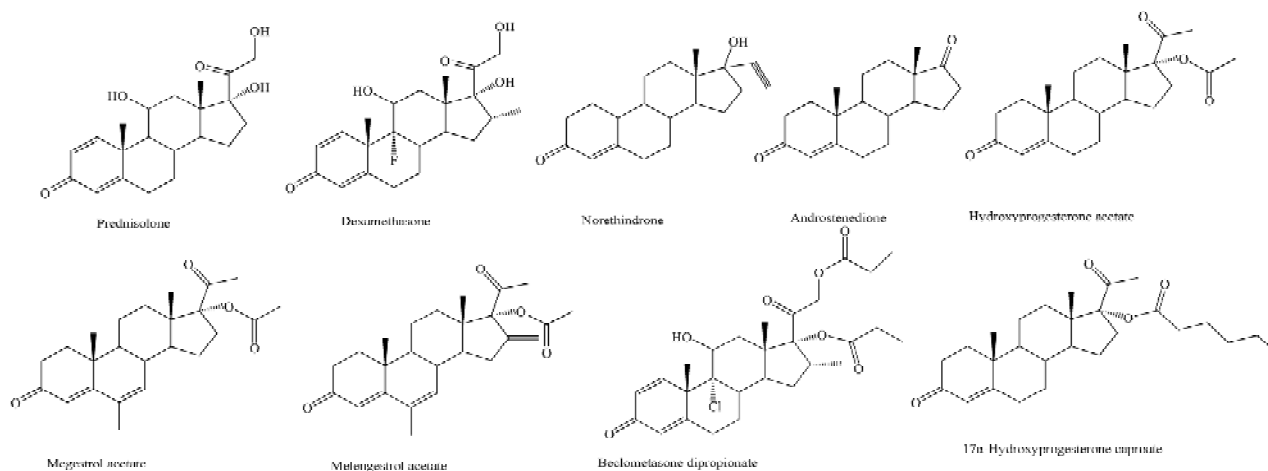


Figure 1. The chemical structures of the studied hormones.

ties, which might greatly affect the extraction efficiency of targeted compounds [15]. In this study, two types of IL, including $[C_6MIM][PF_6]$ and $[C_7MIM][PF_6]$ which are hydrophobic were examined. As shown in **Figure 2**, the $[C_7MIM][PF_6]$ extraction phase produced the better extraction performance for all hormones than that of the $[C_6MIM][PF_6]$, attributed to the stronger hydrophobic interactions between the longer alkyl chain of the $[C_7MIM][PF_6]$ ionic liquid and the hydrophobic hormones. Thus, $[C_7MIM][PF_6]$ was selected for all of the subsequent experiments.

3.1.2. Effect of the Volume of $[C_7MIM][PF_6]$

To study the influence of the volume of $[C_7MIM][PF_6]$ on the extraction efficiency, experiments were carried out by using different volumes of $[C_7MIM][PF_6]$ (from 50 to 150 μ l) as the extraction solvent. As shown in **Figure 3**, the extraction efficiency increased when the vol-

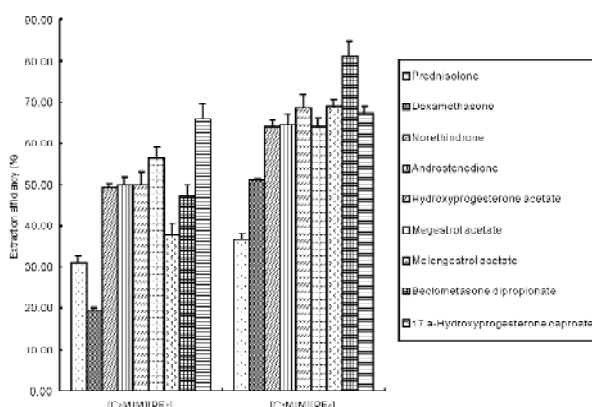


Figure 2. Effect of extraction solvent. ILs volume: 100 μ l; extraction time: 15 min; ultrasonic power: 100 W; salt addition: 0 mg/ml. centrifugation time: 10 min (10,000 rpm); spiked concentration: 10 μ g/ml.

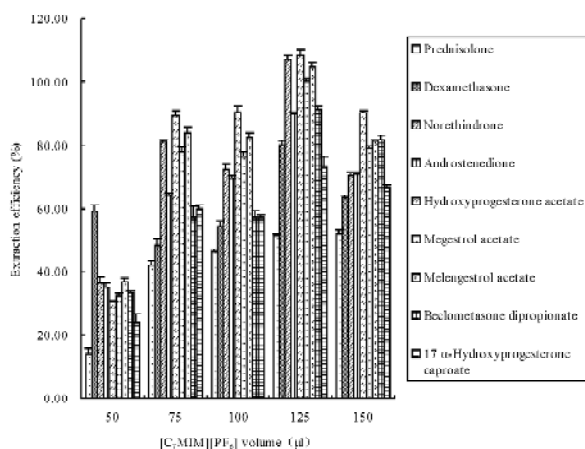


Figure 3. Effect of the volume of $[C_7MIM][PF_6]$. extraction time: 15 min; ultrasonic power: 100 W; salt addition: 0 mg/ml. centrifugation time: 10 min (10,000 rpm); spiked concentration: 10 μ g/ml.

ume of $[C_7MIM][PF_6]$ increased from 50 to 125 μ l, and decreased obviously above 125 μ l. The reason could be that the appropriate volume of $[C_7MIM][PF_6]$ could be dispersed sufficiently into sample solution under ultrasound, but with the volume of $[C_7MIM][PF_6]$ increased, the hormones could not be transferred into the IL phase sufficiently for its high viscosity and density [12]. Therefore, 125 μ l of $[C_7MIM][PF_6]$ was selected for the further study.

3.1.3. Effect of Salt Addition

The influence of salt addition on the performance of IL-USAEME was investigated by adding different amount of NaCl (0 to 9%, w/v) to solution. The results demonstrated that the salt addition had neglectful effect on the extraction efficiency of analytes. On the other hand, the adequate salt addition is helpful to the formation of IL extraction phase. Consequently, the 9% NaCl was used in subsequent experiments.

3.1.4. Effect of Ultrasonic Power and Time

In this work, the emulsification of the ionic liquid into sample solution was facilitated by ultrasound radiation. The ultrasonic power can affect the mass transfer rate and the contact area between the sample solution and the ionic liquid extractant. When the ultrasonic power was tested at two different levels: 100 W, 500 W, it was obvious that the suitable ultrasonic power should be 100 W.

Additionally, the effect of ultrasonic time on extraction efficiency was evaluated in the range of 5 min - 25 min. The results shown in **Figure 4** indicated that the extraction efficiency achieved the maximum at the ultrasonic time of 15 min. The main reason may be that adequate ultrasonic time is needed to ensure the equilibrium of extraction. However, prolonging the ultrasonic time induced the descent of extraction efficiency because of the dissolution of IL in sample solution. Consequently, the ultrasonic time of 15 min was enough.

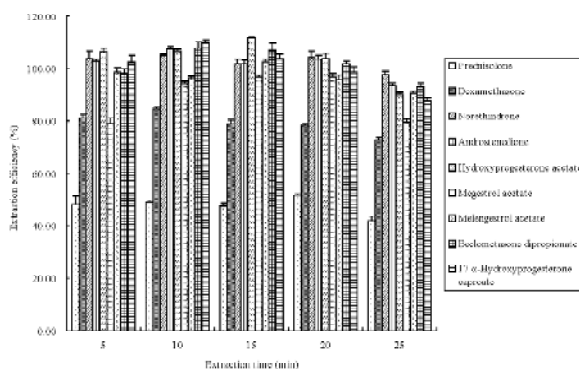


Figure 4. Effect of extraction time. $[C_7MIM][PF_6]$ volume: 125 μ l; salt addition: 9% (m/v); ultrasonic power: 100 W; centrifugation time: 10 min (10,000 rpm); spiked concentration: 10 μ g/ml.

3.1.5. Effect of Centrifugation Time

The centrifugation plays an important role on the sedimentation of IL phase. The effect of centrifugation time ranging from 9 to 21 min was tested. As observed in **Figure 5**, the extraction efficiency of the most analytes increased with the increase of the centrifugation time from 9 to 12 min. When the centrifugation time was longer than 12 min, the extraction efficiency of dexamethasone, hydroxyprogesterone acetate and beclometasone dipropionate decreased and no significant change was observed for others. Therefore, 12 min was chosen as the centrifugation time.

3.2. Evaluation of the method

The chromatogram of a spiked sample obtained by IL-USAEME under the optimum conditions was shown in **Figure 6**. Under the optimum conditions, linear range, limits of detection (LODs), recovery and precision were determined and shown in **Tables 1** and **2**. As can be seen, good linearities were observed for all analytes with the coefficient of correlation (r^2) ranging from 0.9980 to 0.9994. The LODs were calculated at a signal-to-noise (S/N) ratio of 3. The mean recoveries were 62.30% - 108.84% and the RSDs were 0.26% - 4.69%. The intraday and interday precisions varied from 0.10% to 2.51% and from 0.33% to 3.30%, respectively. The results demonstrated that the method is potential for the detection of hormones fortified in cosmetics.

4. Conclusion

In this study, a simple, cheap and green IL-USAEME combined with HPLC method was developed and successfully applied to simultaneously determine three glucocorticoids, one androgen and five progestogens in cosmetics, in which using ultrasound radiation to facilitate

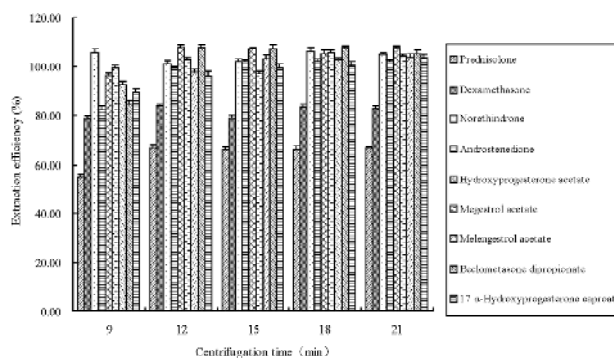


Figure 5. Effect of centrifugation time. [C₇MIM][PF₆] volume: 125 μ l; salt addition: 9% (m/v); ultrasonic power: 100 W; extraction time: 15 min; spiked concentration: 10 μ g/ml.

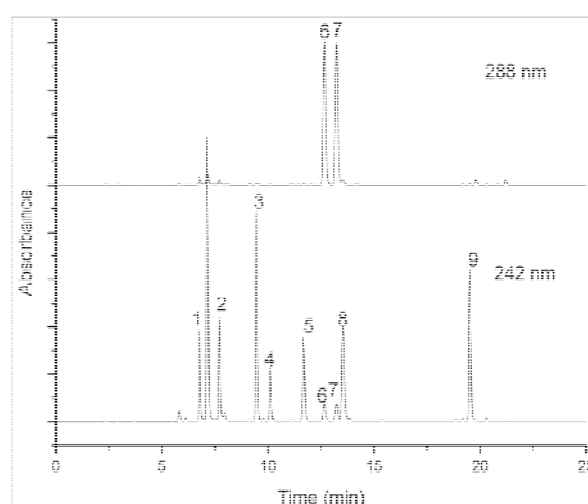


Figure 6. Chromatogram of spiked sample obtained at wavelength 242 and 288 nm. (1) prednisolone, (2) dexamethasone, (3) norethindrone, (4) androstenedione, (5) hydroxyprogesterone acetate, (6) megestrol acetate, (7) melengestrol acetate, (8) beclometasone dipropionate, (9) 17 α -hydroxyprogesterone caproate.

Table 1. Method calibration data and recoveries of analytes.

Analyte	Linear range (μ g/ml)	Correlation coefficient (r^2)	LOD (μ g/g)	Mean recovery (%) (n = 3)	
				1 (μ g/ml)	10 (μ g/ml)
Prednisolone	0.50 - 100.00	0.9994	0.15	62.30 \pm 1.63	66.45 \pm 1.23
Dexamethasone	0.60 - 120.00	0.9991	0.40	98.34 \pm 0.26	95.11 \pm 0.76
Norethindrone	0.20 - 100.00	0.9990	0.10	99.86 \pm 0.24	92.12 \pm 3.60
Androstenedione	0.10 - 50.00	0.9992	0.20	101.75 \pm 1.69	88.11 \pm 2.87
Hydroxyprogesterone acetate	0.60 - 120.00	0.9980	0.40	98.07 \pm 3.50	97.16 \pm 3.38
Megestrol acetate	0.10 - 50.00	0.9991	0.05	105.09 \pm 2.82	85.97 \pm 2.02
Melengestrol acetate	0.10 - 50.00	0.9991	0.10	100.21 \pm 2.61	92.49 \pm 1.73
Beclometasone dipropionate	1.00 - 200.00	0.9991	0.85	108.84 \pm 3.42	106.86 \pm 4.69
17 α -Hydroxyprogesterone caproate	0.30 - 150.00	0.9992	0.25	95.26 \pm 3.91	98.19 \pm 4.38

Table 2. Intraday and interday precision.

Analyte	Intraday ^a (RSD%)			Interday ^b (RSD%)		
	1 (µg/ml)	5 (µg/ml)	25 (µg/ml)	1 (µg/ml)	5 (µg/ml)	25 (µg/ml)
Prednisolone	1.91	0.45	0.57	1.54	0.94	0.68
Dexamethasone	1.82	0.88	0.61	2.01	0.40	0.70
Norethindrone	1.74	0.99	0.38	0.68	0.36	0.57
Androstenedione	1.33	0.89	0.45	0.87	0.96	0.45
Hydroxyprogesterone acetate	0.26	0.47	0.08	1.52	0.58	0.44
Megestrol acetate	0.85	0.47	0.10	1.65	0.73	0.50
Melengestrol acetate	1.05	0.09	0.64	1.97	0.87	0.42
Beclometasone dipropionate	0.60	0.52	0.34	3.30	0.69	0.57
17 α -Hydroxyprogesterone caproate	2.51	1.01	0.88	0.90	0.33	0.90

^aThe intraday precision was gained by analyzing a sample three times in one day. ^bThe interday precision was received by analyzing a sample once a day over three consecutive day.

the emulsification of a microvolume of ionic liquid instead of organic solvent into the sample solution. The experimental conditions for the extraction of these analytes have been investigated. Under the optimized conditions, satisfactory extraction efficiency of the analytes was obtained.

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