DOI: 10.1002/ajh.25554

RESEARCH ARTICLE



Is TAFRO syndrome a subtype of idiopathic multicentric Castleman disease?

Shino Fujimoto¹ | Tomoyuki Sakai¹ | Hiroshi Kawabata^{1,2} ¹ | Nozomu Kurose³ | Sohsuke Yamada³ | Kazue Takai⁴ | Sadao Aoki⁵ | Junya Kuroda⁶ ¹ | Makoto Ide⁷ | Keigo Setoguchi⁸ | Norifumi Tsukamoto⁹ | Haruka Iwao-Kawanami¹ | Takafumi Kawanami¹ | Shuichi Mizuta¹ | Toshihiro Fukushima¹ | Yasufumi Masaki¹

¹Department of Hematology and Immunology, Kanazawa Medical University, Uchinada, Japan

²Department of Hematology and Oncology, Graduate School of Medicine, Kyoto University, Kyoto, Japan

³Department of Pathology and Laboratory Medicine, Kanazawa Medical University, Uchinada, Japan

⁴Department of Hematology, Niigata City General Hospital, Niigata, Japan

⁵Department of Pathophysiology, Faculty of Pharmaceutical Sciences, Niigata University of Pharmacy and Applied Life Sciences, Niigata, Japan

⁶Division of Hematology and Oncology, Department of Medicine, Kyoto Prefectural University of Medicine, Kyoto, Japan

⁷Department of Hematology, Takamatsu Red Cross Hospital, Takamatsu, Japan

⁸Department of Systemic Immunological Diseases, Tokyo Metropolitan Cancer and Infectious Diseases Center Komagome Hospital, Tokyo, Japan

⁹Oncology Centre, Gunma University Hospital, Maebashi, Japan

Correspondence

Hiroshi Kawabata, MD, Department of Hematology and Immunology, Kanazawa Medical University, 1-1 Daigaku, Uchinada, Ishikawa-ken 920-0293, Japan. Email: hkawabat@kanazawa-med.ac.jp

Funding information

Kanazawa Medical University, Grant/Award Numbers: S2004-16, S2007-5, K2011-7, H2011-11, AR2012-06; Ministry of Education, Culture, Sports, Science and Technology, Grant/Award Numbers: 17591060, 15K09510; Ministry of Health, Labour and Welfare, Grant/Award Number: H27-28 Nanchi, etc. (Nan)- General-002; H27-Nanchi

Abstract

Castleman disease (CD) is a rare lymphoproliferative disorder that can be unicentric or multicentric. Multicentric CD (MCD) is further subdivided into human herpesvirus type-8-associated, POEMS syndrome-associated, and idiopathic (iMCD). TAFRO syndrome is a newly identified disorder of unknown etiology characterized by thrombocytopenia, anasarca, fever, reticulin myelofibrosis, renal dysfunction, and organomegaly. The TAFRO syndrome is sometimes regarded as a subtype of iMCD (TAFRO-iMCD), whereas iMCD without TAFRO syndrome is considered "not otherwise specified" (iMCD-NOS). However, a proportion of patients with TAFRO syndrome have been diagnosed without lymph node biopsies (TAFRO syndrome without proven iMCD; TAFRO-w/op-iMCD). To clarify the clinical features of iMCD-NOS, TAFRO-iMCD, and TAFRO-w/op-iMCD, we retrospectively analyzed 220 patients extracted from the database of the Multicenter Collaborative Retrospective Study for Establishing the Concept of TAFRO Syndrome. The patients included 87 with iMCD-NOS, 63 with TAFRO-iMCD, and 19 with TAFRO-w/op-iMCD. Patients in all three groups exhibited anemia, hypoalbuminemia, and elevated serum C-reactive protein and interleukin-6 levels. No significant differences in clinical, laboratory, and prognostic features were noted between the TAFRO-iMCD, and TAFRO-w/opiMCD groups. However, the iMCD-NOS group exhibited polyclonal hyper- γ -globulinemia. The five-year survival rates of patients in the iMCD-NOS and TAFRO-involved groups were 100% and 66.5%, respectively (dropping markedly during the first few months in the latter). The iMCD-NOS and the TAFRO-iMCD samples typically showed plasma cell and mixed-type histologies, respectively. Thus, iMCD can be classified into two distinct subtypes, iMCD-NOS and TAFRO-iMCD. As such, TAFRO-iMCD and TAFRO-w/op-iMCD may be considered the same entity, requiring prompt diagnosis and intensive care.

1 | INTRODUCTION

Castleman disease (CD) is a rare lymphoproliferative disorder that was originally described by Benjamin Castleman in the 1950s as "giant follicular lymph node hyperplasia".^{1,2} The originally described features were asymptomatic lymph node hyperplasia with hyaline-vascular type histologies localized in the mediastinum.² Subsequently, however, patients with enlarged lymph nodes who exhibited plasma celltype histology accompanied by systemic inflammatory symptoms were reported.³ In 1980, Mori et al reported 10 Japanese patients manifesting with generalized lymphadenopathy, plasma cell type histology, and features that indicated inflammation. They named this condition "idiopathic plasmacytic lymphadenopathy with polyclonal hyper-immunoglobulinemia (IPL)".⁴ Moreover, Mori et al posited that IPL may be a subtype of CD. After their report was published, a number of CD patients with generalized lymphadenopathy were described,⁵⁻¹⁶ based on which Frizzera et al classified CD into two categories, according to the distribution of regions of enlarged lymph nodes; the localized form (unicentric CD [UCD]), and CD with multiple lesions (multicentric CD [MCD]).^{17,18} Subsequently, a number of patients with MCD coexisting with Kaposi's sarcoma or AIDS were reported.^{19,20} In 1995, Soulier et al reported that Kaposi's sarcomaassociated herpesvirus (human herpesvirus type 8, HHV-8) was detected in the lymph nodes of all their HIV-positive MCD patients, as well as in a proportion of HIV-negative counterparts.²¹ Since HHV-8 produces viral interleukin-6 (vIL-6), and IL-6 has been identified as the key inducer of the various symptoms of most cases of CD,²² HHV-8 infection was ascribed as the etiology of MCD in HHV-8-positive patients.²³ Though no research had been done into the relative frequencies of HHV-8-positive MCD vs HHV-8-negative MCD, HHV-8-associated MCD has been recognized as a representative type of MCD in Western countries.

Another distinct MCD category is the POEMS (so-called Takatsuki's or Crow-Fukase's) syndrome-associated MCD.²⁴ The POEMS syndrome is a rare paraneoplastic condition manifesting as **p**olyneuropathy, **o**rganomegaly, **e**ndocrinopathy, **m**onoclonal plasma cell proliferation, and **s**kin changes²⁵; moreover, a proportion of patients with POEMS syndrome develop lymphadenopathy with CD histology.²⁶ Thus, the current categorization of CD includes UCD and MCD, with the latter subdivided into HHV-8-associated MCD, POEMS syndrome-associated MCD, and idiopathic MCD (iMCD), which is negative for both HHV-8 and POEMS syndrome (Figure 1).

The TAFRO syndrome is a newly proposed inflammatory disorder of unknown etiology characterized by thrombocytopenia, anasarca, fever, reticulin myelofibrosis, renal dysfunction, and organomegaly.²⁷ This syndrome was originally reported in three Japanese patients with such symptoms by Takai et al in 2010.²⁸ Two of their patients had lymphadenopathy, one of whom had an enlarged lymph node showing CD-like histology. Prior to this report's publication, patients with iMCD demonstrating all the features of TAFRO had been observed for years without receiving much attention.²⁹ However, after it was published, a number of similar patients with TAFRO syndrome,

Castleman Disease

	UCD	POEMS syndrome
	POEMS syndrome-associated	
MCD	HHV-8-associated	TAFRO syndrome
	TAFRO-IMCD	
	IMCD-NOS	

FIGURE 1 Conceptual view of the classification of Castleman disease and related diseases based on our diagnostic criteria for TAFRO syndrome. HHV-8, human herpesvirus type 8; iMCD, idiopathic MCD; MCD, multicentric Castleman disease; NOS, not otherwise specified; TAFRO-iMCD, TAFRO syndrome with iMCD; UCD, unicentric Castleman disease

together with CD-like lymph node histologies were reported. Some were successfully treated with corticosteroids and tocilizumab; the latter is an anti-IL-6 receptor antibody that is approved for iMCD treatment in Japan.³⁰⁻³⁶ Therefore, some researchers consider TAFRO syndrome a subtype of iMCD (Figure S1).^{37,38} Based on this view-point, lwaki et al proposed diagnostic criteria for this syndrome, in which lymph node histology consistent with CD is strictly required (Table S1).³⁸ Applying their diagnostic criteria, they divided iMCD into two categories; iMCD, not otherwise specified (iMCD-NOS) and TAFRO syndrome with iMCD (TAFRO-iMCD), and showed that the clinical manifestations of these two entities were notably distinct.³⁸

However, at least three caveats should be considered when applying these criteria. First, a proportion of patients with TAFRO syndrome, including one of the original patients described by Takai et al²⁸ do not show apparent lymphadenopathy, and would therefore be missed. Second, even if swollen lymph nodes are detected, anasarca and severe hemorrhagic tendencies due to thrombocytopenia and the accompanying disseminated intravascular coagulation often make biopsy difficult to perform. Third, pathological results take time to retrieve, even though they are necessary for the clinical diagnosis of TAFRO syndrome. In contrast to iMCD-NOS, which usually has a chronic clinical course, TAFRO syndrome develops sub-acutely, progresses rapidly, and can often prove fatal. Therefore, prompt diagnosis and intensive treatment with the appropriate intervention are critical for saving these patients.

To achieve a prompt diagnosis, while not excluding patients who have no apparent lymphadenopathy, we proposed another set of diagnostic criteria for TAFRO syndrome. In our criteria, lymphnode biopsy is not required, even though it remains desirable to exclude other diseases (Table S1).³⁹ Based on our diagnostic criteria,³⁹ a proportion of patients were diagnosed with TAFRO syndrome without undergoing lymph node biopsy (TAFRO syndrome without proven iMCD [TAFRO-w/op-iMCD]). As a result, iMCD could be sub-divided into iMCD-NOS and TAFRO-iMCD, and TAFRO syndrome could be sub-divided into TAFRO-iMCD and TAFRO-w/op-iMCD. However, additional research is needed to determine whether TAFRO-iMCD and TAFRO-w/op-iMCD should be considered the same disease entity. To clarify the clinical features of iMCD-NOS, TAFRO-iMCD, and TAFRO-w/op-iMCD, and to verify the validity of our diagnostic criteria, we conducted a retrospective analysis. It used the database of the Multicenter Collaborative Retrospective Study for Establishing the Concept of TAFRO Syndrome (UMIN000011809), to compare the clinical features of these three categories.

2 | METHODS

2.1 | Patient database

The Multicenter Collaborative Retrospective Study for Establishing the Concept of TAFRO Syndrome registry (UMIN000011809) includes patients with suspected MCD and TAFRO syndrome. They were documented since October 2013 at 77 collaborating centers in Japan (Table S2).⁴⁰ The database includes the clinical data obtained at onset or diagnosis, histopathological reports, treatments, and outcomes of these patients. We extracted three clinical patient groups from this registry by applying the international diagnostic criteria for iMCD²⁶ with Iwaki et al's criteria for TAFRO-iMCD (which require lymph node biopsy)³⁸ and our (Masaki et al's) diagnostic criteria for TAFRO syndrome (which do not require lymph node biopsy).³⁹ The groups comprised patients with iMCD-NOS, those with TAFRO-iMCD, and those with TAFRO-w/op-iMCD. We compared the clinical features and outcomes of these groups; most of the patients with iMCD-NOS and those with TAFRO-iMCD in the current cohort were also included in our previous study as the "iMCD" group.⁴⁰ The study protocol was approved by the institutional review board of Kanazawa Medical University and each collaborating facility.

2.2 | Statistical analysis

Two of the three aforementioned subgroups belonged to iMCD, and two belonged to TAFRO syndrome; as such, they were not independent. Therefore, we performed two sets of comparison analyses. Among patients with iMCD, we compared those with TAFRO features to those without. Among patients with TAFRO syndrome, we compared those with proven iMCD to those without; we also compared those with iMCD-NOS to those with whole TAFRO-syndrome. The Fisher's exact test was used to compare binary variables, while Student's *t*-test was used to compare continuous data between the two groups. The Kaplan-Meier method was used for survival analysis, and comparison between groups was performed using the log-rank test. All *P*-values less than .05 were considered statistically significant. All the analyses were conducted using EZR (version 1.30).⁴¹

3 | RESULTS

3.1 | Clinical features

In total, 220 patients were included in this retrospective registry of those recorded between October 2013 and December 2017.

AJH_WILEY⁹⁷⁷

Common collagen diseases, such as systemic lupus erythematosus and Sjögren's syndrome, and IgG4-related diseases were excluded by rheumatologists in each institution, and by rheumatologists (S.F. and Y.M.) at Kanazawa Medical University. This was done by carefully reviewing the provided data, including autoantibody tests and IgG4 values. To make a diagnosis of TAFRO syndrome, a minimum of whole body computed tomography (CT) scan, bone marrow biopsy, and cytological confirmation of pleural effusion/ascites were required. This was necessary to exclude leukemia. lymphoma, infectious diseases, and hemophagocytic lymphohistiocytosis. After these exclusion processes, we extracted 87 patients with iMCD-NOS, 63 with TAFRO-iMCD, and 19 with TAFRO-w/op-iMCD (Figure S2). Lymph node biopsy was not performed in patients with TAFRO-w/op-iMCD; 5 of 19 patients in this group showed no apparent lymphadenopathy on physical examinations or CT scans. All patients with iMCD-NOS fulfilled the international diagnostic criteria for iMCD.²⁶ but not lwaki et al's diagnostic criteria for TAFRO-iMCD³⁸ or Masaki et al's criteria for TAFRO syndrome.³⁹ All patients with TAFRO-iMCD fulfilled the international diagnostic criteria for iMCD and both Iwaki et al's diagnostic criteria for TAFRO-iMCD, and Masaki et al's criteria for TAFRO syndrome. Moreover, all patients with TAFRO-w/op-iMCD fulfilled Masaki et al's diagnostic criteria for TAFRO syndrome only. Clinical profiles and laboratory data at the time of diagnosis of patients in these three groups are shown in Table 1. Fever (temperatures above 37.5°) was observed in almost all patients with TAFRO syndrome, while only one-third of patients with iMCD-NOS presented with fever at the time of diagnosis. Interstitial lung lesions were observed in a guarter of patients with iMCD-NOS, while it was seldom documented in patients with TAFRO syndrome. Anasarca (pleural effusion, ascites, and/or systemic edema) was observed in all patients with TAFRO syndrome by definition, while only 11% of those with iMCD-NOS presented with the same. One-third of patients with TAFRO syndrome required hemodialysis, while only 5% of those with iMCD-NOS did. In contrast, no significant differences in these clinical features were observed between patients with TAFRO-iMCD, and those with TAFRO-w/op-iMCD.

3.2 | Laboratory features

We collected the patients' laboratory data at the time of diagnosis and compared them between groups (Table 1). There were no significant differences in these characteristics between the TAFRO-iMCD and TAFRO-w/op-iMCD groups. In contrast, the mean values of a number of laboratory parameters differed significantly between the iMCD-NOS group and the TAFRO-iMCD group, and between the iMCD-NOS and all TAFRO syndrome (TAFRO-iMCD plus TAFROw/op-iMCD) groups. Patients with all three groups commonly showed anemia, hypoalbuminemia, and elevated serum C-reactive protein (CRP), IL-6, soluble IL-2 receptor, and plasma vascular endothelial growth factor levels. Splenomegaly was frequently observed in patients in all groups. However, the leukocyte counts and CRP levels were significantly higher and the albumin levels significantly lower in the TAFRO groups than in the iMCD-NOS group. Patients in the

TABLE 1 Laboratory findings of patients with iMCD and TAFRO syndrome

		TAFRO syndrome (n = 82) (Groups B + C)		P-values		
	iMCD-NOS (n = 87)(Group A)	TAFRO-iMCD (n = 63)(Group B)	TAFRO-w/op-iMCD (n = 19)(Group C)	A vs B	B vs C	A vs B + C
Age (years)	50 (39-59)	49 (44-63)	55 (44-67)	>.1	>.1	.077
Male: female ratio	48:39	36:27	7:12	>.1	>.1	>.1
Fever (temperature > 37.5°C)	31%	97%	100%	<.001	>.1.	<.001
Splenomegaly	67%	71%	74%	>.1	>.1	>.1
Interstitial lung lesions	23%	2%	0%	<.001	>.1	<.001
Anasarca	11%	100%	100%	<.001	>.1	<.001
Hemodialysis required	5%	32%	32%	<.001	>.1	<.001
WBC (×1000/µL)	7.7 (6.0-9.1) (n = 85)	9.3 (7.1-13.0) (n = 63)	7.4 (5.0-12.8) (n = 19)	.001	>.1	.005
Hb (g/dL)	9.8 (8.3-11) (n = 85)	9.6 (7.4-11.6) (n = 63)	9.8 (7.2-11.3) (n = 19)	>.1	>.1	>.1
PLT (×1000/μL)	337 (264-413) (n = 85)	33 (17-56) (n = 63)	44 (18-74) (n = 19)	<.001	>.1	<.001
BUN (mg/dL)	12 (10-16) (n = 71)	31 (20-60) (n = 62)	38 (15-56) (n = 19)	<.001	>.1	<.001
Creatinine (mg/dL)	0.7 (0.6-0.91) (n = 85)	1.5 (1.1-2.4) (n = 63)	1.62 (0.87-2) (n = 19)	<.001	>.1	<.001
Total protein (g/dL)	9.9 (8.8-10.6) (n = 71)	5.7 (5.1-6.3) (n = 62)	5.4 (4.9-6) (n = 19)	<.001	>.1	<.001
ALB (g/dL)	2.8 (2.4-3.2) (n = 85)	2.3 (1.9-2.7) (n = 63)	2.1 (1.89-2.5) (n = 19)	<.001	>.1	<.001
CRP (mg/dL)	7.7 (4.5-11.5) (n = 82)	16.1 (6.3-21.7) (n = 63)	12.7 (6.3-26.7) (n = 19)	<.001	>.1	<.001
LDH (IU/L)	120 (97-158) (n = 84)	207 (176-280) (n = 62)	222 (185-274) (n = 19)	<.001	>.1	>.1
ALP (IU/L)	279 (219-393) (n = 84)	537 (375-1108) (n = 62)	502 (397-782) (n = 19)	<.001	>.1	<.001
γ-GTP (IU/L)	27.5 (16-59) (n = 62)	84 (40-156) (n = 60)	58 (39.5-118.2) (n = 18)	<.001	>.1	<.001
AST (IU/L)	17 (12-22) (n = 72)	23 (17-36) (n = 62)	25 (217-30) (n = 19)	<.001	>.1	<.001
ALT (IU/L)	12 (7.2-18) (n = 72)	15 (10-28) (n = 62)	14 (8-21) (n = 19)	<.001	>.1	.02
IgG (mg/dL)	4905 (3510-6113) (n = 83)	1345 (1091-1778) (n = 58)	1210 (913-1433) (n = 18)	.018	>.1	<.001
IgA (mg/dL)	632 (349-842) (n = 71)	210 (156-264) (n = 57)	214 (167-299) (n = 18)	<.001	>.1	<.001
IgM (mg/dL)	237 (169-366) (n = 71)	77 (58-99) (n = 58)	72 (40-108) (n = 18)	<.001	>.1	<.001
IL-6 (pg/mL)	21 (10-42) (n = 76)	26 (15-40) (n = 52)	23 (13-40) (n = 12)	>.1	>.1	>.1
sIL2R (U/mL)	1450 (1084-2232) (n = 80)	1669 (1070-2490) (n = 59)	1810 (1330-2455) (n = 17)	>.1	>.1	>.1
plasma VEGF (pg/mL)	435 (107-835) (n = 4)	188 (112-362) (n = 20)	172 (31-310) (n = 7)	>.1	>.1	>.1
D-dimer (µg/mL)	1 (0.7-1.2) (n = 35)	10.2 (5.2-18.5) (n = 47)	9.6 (5-16.9) (n = 15)	<.001	>.1	<.001
FDP (µg/mL)	4 (2.4-7.1) (n = 33)	23.9 (13.9-45.8) (n = 45)	21.9 (14.3-34.3) (n = 13)	<.001	>.1	<.001

Notes: For age and laboratory data, median values (25-75th percentile) were shown. For clinical manifestations, percentages of their frequencies were shown. Anasarca indicates pleural effusion, ascites, and/or generalized edema. Reference ranges: WBC, 2.97-9.13 \times 1000/µL; Hb, 12.9-9.13 g/dL; PLT, 143-333 \times 1000/µL; BUN, 8-22 mg/dL; Creatinine, 0.6-1.1 mg/dL; Total protein, 6.7-8.3 g/dL; ALB, 4.0-5.0 g/dL; CRP, 0-0.3 mg/dL; LDH, 119-229 IU/L; ALP, 115-359 IU/L; γ -GTP, 10-47 IU/L; AST, 13-33 IU/L; ALT, 8-42 IU/L; IgG, 870-1700 mg/dL; IgA, 110-410 mg/dL; IgM 86-160 mg/dL; IL-6 \leq 4 pg/mL; sIL2R 145-519 U/mL; plasma VEGF \leq 38.3 pg/mL; D-dimer \leq 1 µg/mL; FDP \leq 5 µg/mL.

Abbreviations: iMCD, idiopathic multicentric Castleman disease; NOS, not otherwise specified; TAFRO-w/op-iMCD, TAFRO syndrome without proven iMCD; WBC, white blood cell counts; Hb, hemoglobin; PLT, platelets; BUN, blood urea nitrogen; ALB, albumin; CRP, C-reactive protein; LDH, lactate dehydrogenase; ALP, alkaline phosphatase; γ -GTP, γ -glutamyl transpeptidase; AST, aspartate aminotransferase; ALT, alanine transaminase; Ig, immunoglobulin; IL-6, interleukin 6; sIL2R, soluble interleukin 2 receptor; VEGF, vascular endothelial growth factor; FDP, fibrin/fibrinogen degenerative products.

TAFRO groups exhibited thrombocytopenia by definition, whereas those with iMCD-NOS did not have thrombocytopenia (P < .001). The median serum lactate dehydrogenase (LDH) levels were not elevated in any of the 3 groups, but their levels in iMCD-NOS-group patients were significantly lower than those in the patients of the TAFRO groups. The levels of blood urea nitrogen, creatinine, alkaline phosphatase (ALP), and γ -glutamyl transpeptidase (γ -GTP) were significantly higher in TAFRO-group patients than in those of the iMCD-

NOS group (in whom these values were within the reference ranges). In contrast, polyclonal hyper-γ-globulinemia with significantly elevated serum IgA, IgG, and IgM levels was observed in the iMCD-NOS group, but not in the TAFRO groups. The median values of fibrin/fibrinogen degenerative product (FDP) and D-dimer were significantly higher in the TAFRO groups than in the iMCD-NOS group. Among patients with TAFRO-w/op-iMCD, no significant differences in any of these laboratory parameters were observed between those with detectable lymphaden opathy (n = 14) and those without (n = 5) (Table S3).

3.3 | Histological features

The lymph node histology of CD was classified into three types: hyaline-vascular, plasma cell, and mixed types. Detailed information regarding lymph node histology was available for 66 of the 87 patients with iMCD-NOS, including 4 (6%) with hyaline-vascular type, 51 (77%) with plasma cell type, and 11 (17%) with mixed-type. Moreover, detailed information regarding lymph node histology was available for 49 of the 63 patients with iMCD-TAFRO, including 6 (12%) with hyaline-vascular type, 7 (14%) with plasma cell type, and 36 (73%) with mixed-type. Lymph node biopsy was not performed in patients with TAFRO-w/o-iMCD.

3.4 | Prognostic features

Sixty-nine, 58, and 15 patients with iMCD-NOS, TAFRO-iMCD, and TAFRO-w/op-iMCD, respectively, had available follow-up data. The median follow-up period of the survivors was 63.5 months (iMCD-NOS, 117 months [range, 5-315 months]; TAFRO-iMCD, 42 months [5-161 months]; TAFRO-w/op-iMCD, 27 months [9-231 months]; and combined TAFRO syndrome, 41.5 months [5-231 months]). Among the 69 patients with iMCD-NOS, nine were observed without specific treatments, 50 received corticosteroids, 29 were treated with tocilizumab, two were treated with cyclosporine, and eight received rituximab (some patients received multiple or combination treatments). All patients with TAFRO-iMCD and TAFRO-w/op-iMCD received corticosteroids. Moreover, 24 of the 58 patients with TAFRO-iMCD received tocilizumab, 18 received cyclosporine, and 11 received rituximab. Among 19 patients



FIGURE 2 Overall survival. Statistic comparisons between iMCD-NOS and TAFRO-iMCD, and between TAFRO-iMCD and TAFRO-w/ op-iMCD, were performed using the log-rank test. iMCD-NOS, idiopathic multicentric Castleman disease-not otherwise specified; TAFRO-iMCD, TAFRO syndrome with iMCD; TAFRO-w/op-iMCD, TAFRO syndrome without proven iMCD

with TAFRO-w/op-iMCD, three received tocilizumab, nine received cyclosporine, and one received rituximab. Kaplan-Meier analyses revealed that the overall survival (OS) curves of the TAFRO syndrome groups rapidly dropped within 24 months of diagnosis, by which time a third of the patients had died. No significant differences in OS were observed between the TAFRO-iMCD and TAFRO-w/op-iMCD groups. In contrast, more than 90% of patients with iMCD-NOS were alive 10 years after diagnosis. Thus, the OS of patients with iMCD-NOS was markedly and significantly superior to that of patients with TAFRO syndrome groups (Figure 2). The five-year survival rate of patients with iMCD-NOS was 100%, while the two-year survival rates of patients with TAFRO-iMCD and TAFRO-w/op-iMCD were 67.4% and 61.7%, respectively (the survival rate was 66.5% when the TAFRO syndrome involved groups were combined).

4 | DISCUSSION

Iwaki et al analyzed the clinical and histological features of 25 patients with TAFRO-iMCD, and compared them to 19 patients with iMCD-NOS.³⁸ They reported that the TAFRO-iMCD patients frequently demonstrated abdominal pain, elevated serum ALP levels, and acute kidney failure. Twenty-eight percent of these patients required temporary hemodialysis (a proportion similar to the 32% in our cohort) without showing hyper-y-globulinemia, which is commonly observed in iMCD-NOS. All patients with iMCD-NOS showed plasma cell-type lymph node histology, while those with TAFRO-iMCD revealed completely different histologies. They included atrophic germinal centers, expansion of the interfollicular zone, proliferation of highly dense endothelial venules, and relatively few mature plasma cells³⁸ (which is now classified as hypervascular pathology²⁶). Similar findings were reported in 2008 by Kojima et al, who classified 28 iMCD patients into two subtypes: IPL and non-IPL.²⁹ They described the latter as frequently presenting with pleural effusions, thrombocytopenia, and various autoimmune features without marked hyper-y-globulinemia. Histologically, their lymph nodes exhibited small epithelioid-type follicles, and moderate-to-prominent vascularity with high endothelial venules, and various degrees of plasma cell proliferation in the interfollicular area.²⁹ This was consistent with hypervascular pathology.²⁶ We postulate that most patients with this non-IPL type of iMCD fulfilled the diagnostic criteria for TAFRO syndrome.

With our much larger cohort of TAFRO syndrome patients (n = 82), we strove to validate previous findings and obtained essentially consistent results. Anemia, hypoalbuminemia, elevation of serum CRP levels, and organomegaly were commonly observed in both the iMCD-NOS and TAFRO-iMCD groups, and unlike patients with aggressive lymphomas, serum LDH levels were normal. Patients in the iMCD-NOS group typically showed normal or slightly elevated plate-let counts, marked hyper- γ -globulinemia, and lymph node histology indicative of plasma cell type. These features are consistent with those of IPL, as previously proposed by Mori et al.⁴ In contrast, patients in the TAFRO groups (TAFRO-iMCD and TAFRO-w/op-iMCD) manifested with leukocytosis, thrombocytopenia, normal

WILEY_AJH

 γ -globulin levels, elevation of serum ALP and γ -GTP levels, and anasarca. Elevations of serum FDP and D-dimer levels, which were indicative of disseminated intravascular coagulation, were observed in the TAFRO syndrome groups but not in the iMCD-NOS group. Lymph node histology in the majority of patients with TAFRO-iMCD was classified as mixed-type by institutional diagnosis; a proportion of these cases could be reclassified as hypervascular pathology under a recent guideline.²⁶ In our recent central review of lymph nodes, plasma cell type, mixed-type, and hypervascular type histologies were observed in 67%, 33%, and 0% of iMCD-NOS patients (n = 24), and 6%, 78%, and 16% of TAFRO-iMCD patients (n = 32), respectively.⁴² On Kaplan-Meier analyses, the five-year OS in the iMCD-NOS group was 100%; this may have been attributable to the wide use of tocilizumab in Japan after 2005. In contrast, the OS curve for the TAFRO syndrome group dropped rapidly over two years; one-third of patients with TAFRO syndrome died during this period. Very similar survival curves were reported by Yu et al, who analyzed 34 patients with iMCD-NOS, and nine with TAFRO-iMCD in the United States.⁴³ Because our cohort included the largest number of patients with TAFRO syndrome analyzed to date, the survival curve shown in Figure 2 may reflect real-time rates in Japan. Taken together, iMCD can be classified into at least two types; iMCD-NOS (or IPL type), and TAFRO-iMCD (or non-IPL type); these should be considered as clearly distinct clinical entities.

In addition to iMCD-NOS and TAFRO-iMCD, we analyzed the clinical features of patients with TAFRO-w/op-iMCD as diagnosed using Masaki et al's criteria.³⁹ The clinical manifestations and laboratory data of patients in this group were similar to those of the TAFRO-iMCD group. Furthermore, the survival rates of these two groups were almost identical, and were significantly inferior to that of patients with iMCD-NOS. Notably, a majority of patients with TAFRO-w/op-iMCD had enlarged lymph nodes. Biopsy of these lymph nodes could have clearly excluded the possibilities of lymphoma and other etiologies, so we strongly recommend performing lymph node biopsy if possible.³⁹ However, it is also noteworthy that five out of 19 patients presented with no lymphadenopathy on physical examinations or imaging tests were similar to those of patients with detectable lymphadenopathy. Additional research is required to identify the TAFRO-specific clinical features among patients with TAFRO-w/op-iMCD, and overlapping disorders. We still posit that TAFRO-iMCD and TAFRO-w/op-iMCD, including those without lymphadenopathy, ought to be considered a single clinical entity, and that it is appropriate to use Masaki et al's criteria to diagnose TAFRO syndrome promptly.³⁹ When applying these criteria, we emphasize that specific diseases that are known to manifest TAFRO-like clinical features, should be carefully excluded before making a final diagnosis of TAFRO syndrome, especially of TAFRO-w/op-iMCD.

Based on data from previous studies as well as our own, it is evident that the clinicopathological features of TAFRO syndrome are different from those of iMCD-NOS, although the relationship between TAFRO syndrome and MCD remains controversial. Some researchers regard TAFRO syndrome as an aggressive subtype of iMCD, some consider it serositis with thrombocytopenia driven by autoimmune mechanisms and sometimes accompanied by lymph node histopathology that is coincidentally similar to iMCD, and others regard it as a clinical entity overlapping with iMCD (Figure S1). Based on the data from the current study, we tentatively regard iMCD-NOS and TAFRO syndrome as distinct entities, and consider that TAFRO-iMCD might represent the overlapping entity of iMCD and TAFRO syndrome.

Several limitations should be noted in our study. Because it was conducted retrospectively, patient-selection bias may exist. For example, surviving patients may be more likely to be registered than deceased patients. The periods of diagnosis varied widely (from 1990s to 2016), which may have affected the accuracy of diagnosis, and the outcomes of prognostic analyses. The pathological diagnoses and classifications were largely determined at the individual centers, which might influence their consistencies. Furthermore, not all data were available owing to the multicenter nature of the study, particularly histopathological reports and follow-up data. Nevertheless, our findings confirmed the results of previous studies that characterized the clinical features of TAFRO syndrome, and provided real-world survival rates of patients with this syndrome using the largest cohort investigated to date. We also verified the appropriateness of Masaki et al's diagnostic criteria for TAFRO syndrome.

The etiologies of iMCD-NOS and TAFRO syndrome remain completely unknown. Specific biomarkers for TAFRO syndrome are still lacking, although Iwaki et al recently reported that serum interferon γ -induced protein 10 kDa was elevated in patients with TAFROiMCD but not in those with iMCD-NOS, implicating this molecule in the pathogenesis of TAFRO-iMCD.⁴⁴ Recently, Pierson et al published an interesting study that revealed distinct proteomic profiles in patients with TAFRO-iMCD, and those with iMCD-NOS, though the number of patients was small.⁴⁵

In a study from China, only half of the patients with MCD, including those who had HHV-8 associated disease, survived beyond five years.⁴⁶ According to a retrospective study on CD from the Mayo Clinic and University of Nebraska, the five-year OS of patients with MCD was 65%.⁴⁷ Treatments with tocilizumab and siltuximab greatly improved the quality of life, and might also have improved the survival of patients with iMCD-NOS. In contrast, and despite intensive treatments with corticosteroids, tocilizumab, cyclosporine, and/or rituximab,^{31,34,35,48,49} the rate of early death in patients with TAFRO syndrome is high, signifying the importance of prompt diagnosis, and intensive treatment without delay, though the development of new therapeutic strategies remains an unmet need. We previously proposed tentative treatment strategies for TAFRO syndrome that included glucocorticoids, calcineurin inhibitors, rituximab, and tocilizumab.³⁹ Recently, the international guidelines developed for iMCD treatment cited the high response rates achieved with cyclophosphamide-based chemotherapy, rituximab, tocilizumab, and cyclosporine A, and recommended anti-IL-6 monoclonal antibody therapy with or without corticosteroids as the initial therapy.⁵⁰ However, these recommendations are essentially based on published case reports and authors' own experiences. In order to establish highquality evidence for the characteristics of iMCD and TAFRO syndrome, delineate the relationship between them, and establish

ACKNOWLEDGMENTS

The authors thank the patients, their families, all the investigators including Drs. Chisako Iriyama (Aichi Cancer Center Hospital), Taro Masunari, Kazutoshi Aoyama (Chugoku Central Hospital), Marie Nakashima (Chutoen General Medical Center), Masaru Kojima (Dokkyo Medical University), Hiroshi Doi (Ebara Hospital), Yukihiro Miyazaki (Ehime Prefectural Central Hospital), Masao Hagihara, Hua Jian, Morihiro Inoue (Eiju General Hospital), Kouichi Hirakawa (Fukuchiyama City Hospital), Hiroyuki Minemura (Fukushima Medical University), Nobuhiko Nakamura (Gifu University), Akihiko Yokohama, Takeki Mitsui (Gunma University), Takamitsu Sasaki, Takayuki Tsuji (Hamamatsu University Hospital), Tadashi Matsumura (Himeji St. Mary's Hospital), Hideki Goto, Koji Ogawa (Hokkaido University), Takayuki Fujio (Ibaraki Prefectural Central Hospital), Shigeki Hatanaka, Kazutaka Nishitarumizu, Yukiko Mori (Imamura Byoin Bun-in Hospital). Takahiro Nagashima (Japanese Red Cross Kitami Hospital). Saiko Yoshida (Japanese Red Cross Nagoya Daini Hospital), Yoko Adachi (JCHO Kobe Central Hospital), Sato Toshinobu (JCHO Sendai Hospital), Tomiko Ryu (JCHO Tokyo Yamate Hospital), Yoko Edahiro, Michiaki Koike, Yoshitaka Sunami, (Juntendo University Shizuoka Hospital), Taku Kikuchi (Keio University), Ikuko Kubokawa (Kobe University), Katsunori Kyoda (Kouseiren Takaoka Hospital), Daisuke Wakasugi, Ritsuko Seki, Koji Nagafuji, Masaki Okamoto (Kurume University), Taichi Nakamura (Kuwana City Medical Center), Hisao Nagoshi (Kyoto Prefectural University), Takeshi Matsubara (Kyoto University), Eiko Oya (Matsusaka Chuo Hospital), Kiyoyuki Ogata (Metropolitan Research and Treatment Center for Blood Disorders), Chisato Tanihashi (Minai Seikyo Hospital), Toshio Yano, Tadashi Koike (Nagaoka Red Cross Hospital), Daisuke Ogawa (Nagasaki Prefecture Shimabara Hospital), Tomoki Origuchi (Nagasaki University), Hiroyuki Kobayashi (Nasu Red Cross Hospital), Naohiro Sekiguchi (NHO Disaster Medical Center), Sachiko Suzuki, Mitsutoshi Kurosawa (NHO Hokkaido Cancer Center), Yuto Mimura (NHO Matsumoto Medical Center), Izumi Yasumori (NHO Nagasaki Medical Center), Hiroatsu lida (NHO Nagoya Medical Center), Akio Saito (NHO Nishigunma National Hospital), Jun Konishi, Kazutaka Sunami (NHO Okayama Medical Center), Koji Nikkuni (Niigata City General Hospital), Akihito Momoi (Niigata Prefectural Central Hospital), Jun Takizawa (Niigata University), Azusa Nagao (Ogikubo Hospital), Shin-ichi Nureki (Oita University), Sae Wada (Okayama Rosai Hospital), Yoshihiko Raita (Okinawa Prefectural Chubu Hospital), Keiko Yamagami, Ko lida (Osaka City General Hospital), Masanori Konayashi (Osaka City University), Yoshinobu Konishi (Osaka Red Cross Hospital), Masato Arao (Saitama Medical University), Tadashi Nakamura (Sakurajyuji Hospital), Masaya Mukai (Sapporo City General Hospital), Tsutomu Sato (Sapporo Medical University), Daisuke Shichi, Kenji Nara, Sotaro Kasukawa (Seirei Mikatahara General Hospital), Akemi Matsuo (Shinonoi General Hospital), Yoko OzawaHiroshi Yamamoto Hidekazu AJH_WILEY⁹⁸¹

Takahashi (Shinshu University), Keiki Nagaharu, Keiki Kawakami (Suzuka General Hospital), Yasuo Hoshijima (Takamatsu Municipal Hospital), Hanako Tsurumi, Shunya Uchida (Teikyo University), Noboru Hagino, Lisa Hirahara, Yusuke Takeshima (Teikyo University Chiba Medical Center), Sachiko Ando, Hajime Sakai (Teine Keijinkai Hospital), Tohru Takahashi (Tenshi Hospital), Mai Tanaka (The Jikei University), Hiroshi Fujii. Yoko Okitsu. Noriko Fukuhara (Tohoku University). Naoki Hayama (Tokai University), Yuko Toyoda, Masayoshi Yamanishi (Tokushukai Medical Cooperation Uji-Tokushukai Hospital), Yasuhiko Nishioka (Tokushima University), Tatsuhiko Shinohara, Ejima Masaru (Tokyo Medical and Dental University), Toshikazu Wada, Yume Nagaoka (Tokyo Medical University), Sayuri Motoomura (Tokyo Metropolitan Health and Medical Treatment Corporation Tama-Hokubu Medical Center), Keigo Setoguchi (Tokyo Metropolitan Komagome Hospital), Daisuke Mizuchi (Tokyo Teishin Hospital), Masakuni Tanimizu (Tottori Municipal Hospital). Satoru Kosugi (Tovonaka Municipal Hospital). Yusuke Takagi (Toyota Kosei Hospital), Yoshimasa Urasaki (University of Fukui), Jun Murakami (University of Toyama), and nurses in the participating institutions of this study.

DISCLOSURE OF INTERESTS

Dr Masaki reports grants from Astellas Pharma Inc., Kyowa Hakko Kirin Co., Ltd., Eisai Co., Ltd., Ono Pharmaceutical Co. LtD., Pfizer Seiyaku K.K. outside the submitted work; Dr Kuroda reports a grant from Chugai Pharmaceutical Co., Ltd. outside the submitted work.

AUTHOR CONTRIBUTIONS

S.F. and H.K. analyzed the data, reviewed literature, and co-wrote the manuscript. T.S., K.T., S.A., J.K., M.I., K.S., N.T., H.I.-K., T.K., S.M. and T.F. participated in the clinical enrollment/work-up of the patients and edited the manuscript. N.K. and S.Y. reviewed literature on histopathology and edited the manuscript. Y.M. designed the research and reviewed the study results.

ORCID

Hiroshi Kawabata D https://orcid.org/0000-0002-1918-8635 Junya Kuroda D https://orcid.org/0000-0001-6130-1550

REFERENCES

- Castleman B, Towne VW. CASE records of the Massachusetts General Hospital weekly Clinicopathological exercises: CASE 40011. N Engl J Med. 1954;250(1):26-30.
- Castleman B, Iverson L, Menendez VP. Localized mediastinal lymphnode hyperplasia resembling thymoma. *Cancer.* 1956;9(4): 822-830.
- Keller AR, Hochholzer L, Castleman B. Hyaline-vascular and plasmacell types of giant lymph node hyperplasia of the mediastinum and other locations. *Cancer.* 1972;29(3):670-683.
- Mori S, Mohri N, Uchida T, Shimamine T. Idiopathic plasmacytic lymphadenopathy with polyclonal hyperimmunoglobulinemia: a syndrome

982 WILEY AJH

related to giant lymph node hyperplasia of plasma cell type. *J Jpn Soc RES*. 1981;20(suppl):85-94.

- Bartoli E, Massarelli G, Soggia G, Tanda F. Multicentric giant lymph node hyperplasia. A hyperimmune syndrome with a rapidly progressive course. *Am J Clin Pathol.* 1980;73(3):423-426.
- Rizzo SC, Magrini U, Balduini CL, Ricevuti G, Storti E. Multicentric giant lymph node hyperplasia. A case report. *Haematologica*. 1981; 66(5):673-681.
- 7. Sindram MT, Fedder G. Recurrent and multicentric giant lymph node hyperplasia. *Scand J Haematol.* 1981;27(1):25-29.
- Marti S, Pahissa A, Guardia J, Moragas A, Bacardi R. Multicentric giant follicular lymph node hyperplasia. Favorable response to radiotherapy. *Cancer.* 1983;51(5):808-810.
- Tanda F, Massarelli G, Costanzi G. Multicentric giant lymph node hyperplasia: an immunohistochemical study. *Hum Pathol.* 1983;14(12): 1053-1058.
- Rywlin AM, Rosen L, Cabello B. Coexistence of Castleman's disease and Kaposi's sarcoma. Report of a case and a speculation. *Am J Dermatopathol*. 1983;5(3):277-281.
- Chen KT. Multicentric Castleman's disease and Kaposi's sarcoma. Am J Surg Pathol. 1984;8(4):287-293.
- Diebold J, Marche C, Audouin J, et al. Lymph node modification in patients with the acquired immunodeficiency syndrome (AIDS) or with AIDS related complex (ARC). A histological, immuno-histopathological and ultrastructural study of 45 cases. *Pathol Res Pract.* 1985;180(6): 590-611.
- Levo Y, Behar AJ, Blum I, Frish B. A benign course of multicentric Castleman's disease with involvement of the spleen and bone marrow. *Eur J Haematol*. 1987;39(5):471-474.
- Yago K, Kanoh T, Uchino H. Multicentric giant lymph node hyperplasia, plasma cell type, with monoclonal gammopathy. *Tohoku J Exp Med.* 1987;153(1):49-54.
- Ben-Chetrit E, Flusser D, Okon E, Ackerman Z, Rubinow A. Multicentric Castleman's disease associated with rheumatoid arthritis: a possible role of hepatitis B antigen. Ann Rheum Dis. 1989;48(4): 326-330.
- Mizutani N, Okada S, Tanaka J, et al. Multicentric giant lymph node hyperplasia with ascites and double cancers, an autopsy case. *Tohoku J Exp Med.* 1989;158(1):1-7.
- Frizzera G, Banks PM, Massarelli G, Rosai J. A systemic lymphoproliferative disorder with morphologic features of Castleman's disease. Pathological findings in 15 patients. *Am J Surg Pathol.* 1983;7(3): 211-231.
- Frizzera G, Peterson BA, Bayrd ED, Goldman A. A systemic lymphoproliferative disorder with morphologic features of Castleman's disease: clinical findings and clinicopathologic correlations in 15 patients. J Clin Oncol. 1985;3(9):1202-1216.
- Ulbright TM, Santa Cruz DJ. Kaposi's sarcoma: relationship with hematologic, lymphoid, and thymic neoplasia. *Cancer*. 1981;47(5): 963-973.
- Lachant NA, Sun NC, Leong LA, Oseas RS, Prince HE. Multicentric angiofollicular lymph node hyperplasia (Castleman's disease) followed by Kaposi's sarcoma in two homosexual males with the acquired immunodeficiency syndrome (AIDS). Am J Clin Pathol. 1985;83(1):27-33.
- Soulier J, Grollet L, Oksenhendler E, et al. Kaposi's sarcomaassociated herpesvirus-like DNA sequences in multicentric Castleman's disease. *Blood*. 1995;86(4):1276-1280.
- Yoshizaki K, Matsuda T, Nishimoto N, et al. Pathogenic significance of interleukin-6 (IL-6/BSF-2) in Castleman's disease. *Blood.* 1989;74(4): 1360-1367.
- Parravicini C, Corbellino M, Paulli M, et al. Expression of a virusderived cytokine, KSHV vIL-6, in HIV-seronegative Castleman's disease. Am J Pathol. 1997;151(6):1517-1522.

- 24. Frizzera G. Castleman's disease and related disorders. Semin Diagn Pathol. 1988;5(4):346-364.
- Dispenzieri A. POEMS syndrome: 2017 update on diagnosis, risk stratification, and management. Am J Hematol. 2017;92(8):814-829.
- Fajgenbaum DC, Uldrick TS, Bagg A, et al. International, evidencebased consensus diagnostic criteria for HHV-8-negative/idiopathic multicentric Castleman disease. *Blood.* 2017;129(12):1646-1657.
- 27. Kawabata H, Takai K, Kojima M, et al. Castleman-Kojima disease (TAFRO syndrome) : a novel systemic inflammatory disease characterized by a constellation of symptoms, namely, thrombocytopenia, ascites (Anasarca), microcytic Anemia, Myelofibrosis, renal dysfunction, and Organomegaly : a status report and summary of Fukushima (6 June, 2012) and Nagoya meetings (22 September, 2012). J Clin Exp Hematop. 2013;53(1):57-61.
- Takai K, Nikkuni K, Shibuya H, Hashidate H. Thrombocytopenia with mild bone marrow fibrosis accompanied by fever, pleural effusion, ascites and hepatosplenomegaly. *Rinsho Ketsueki*. 2010;51(5): 320-325.
- Kojima M, Nakamura N, Tsukamoto N, et al. Clinical implications of idiopathic multicentric castleman disease among Japanese: a report of 28 cases. Int J Surg Pathol. 2008;16(4):391-398.
- Iwaki N, Sato Y, Takata K, et al. Atypical hyaline vascular-type castleman's disease with thrombocytopenia, anasarca, fever, and systemic lymphadenopathy. J Clin Exp Hematop. 2013;53(1):87-93.
- Kawabata H, Kotani S, Matsumura Y, et al. Successful treatment of a patient with multicentric Castleman's disease who presented with thrombocytopenia, ascites, renal failure and myelofibrosis using tocilizumab, an anti-interleukin-6 receptor antibody. *Intern Med.* 2013;52(13):1503-1507.
- 32. Masaki Y, Nakajima A, Iwao H, et al. Japanese variant of multicentric castleman's disease associated with serositis and thrombocytopeniaa report of two cases: is TAFRO syndrome (Castleman- Kojima disease) a distinct clinicopathological entity? J Clin Exp Hematop. 2013; 53(1):79-85.
- Kubokawa I, Yachie A, Hayakawa A, et al. The first report of adolescent TAFRO syndrome, a unique clinicopathologic variant of multicentric Castleman's disease. *BMC Pediatr.* 2014;14(1):139.
- Konishi Y, Takahashi S, Nishi K, et al. Successful treatment of TAFRO syndrome, a variant of multicentric Castleman's disease, with cyclosporine a: possible Pathogenetic contribution of Interleukin-2. *Tohoku J Exp Med.* 2015;236(4):289-295.
- Tedesco S, Postacchini L, Manfredi L, et al. Successful treatment of a Caucasian case of multifocal Castleman's disease with TAFRO syndrome with a pathophysiology targeted therapy - a case report. *Exp Hematol Oncol.* 2015;4(1):3.
- 36. Fujiwara S, Mochinaga H, Nakata H, et al. Successful treatment of TAFRO syndrome, a variant type of multicentric Castleman disease with thrombotic microangiopathy, with anti-IL-6 receptor antibody and steroids. Int J Hematol. 2016;103(6):718-723.
- Carbone A, Pantanowitz L. TAFRO syndrome: an atypical variant of KSHV-negative multicentric Castleman disease. *Am J Hematol.* 2016; 91(2):171-172.
- Iwaki N, Fajgenbaum DC, Nabel CS, et al. Clinicopathologic analysis of TAFRO syndrome demonstrates a distinct subtype of HHV-8-negative multicentric Castleman disease. Am J Hematol. 2016;91(2):220-226.
- Masaki Y, Kawabata H, Takai K, et al. Proposed diagnostic criteria, disease severity classification and treatment strategy for TAFRO syndrome, 2015 version. Int J Hematol. 2016;103(6):686-692.
- 40. Fujimoto S, Koga T, Kawakami A, et al. Tentative diagnostic criteria and disease severity classification for Castleman disease: a report of the research group on Castleman disease in Japan. *Mod Rheumatol.* 2018;28(1):161-167.
- 41. Kanda Y. Investigation of the freely available easy-to-use software 'EZR' for medical statistics. *Bone Marrow Transplant*. 2013;48(3):452-458.

- 42. Kurose N, Futatsuya C, Mizutani KI, et al. The clinicopathological comparison among nodal cases of idiopathic multicentric Castleman disease with and without TAFRO syndrome. *Hum Pathol*. 2018;77:130-138.
- Yu L, Tu M, Cortes J, et al. Clinical and pathological characteristics of HIV- and HHV-8-negative Castleman disease. *Blood*. 2017;129(12): 1658-1668.
- Iwaki N, Gion Y, Kondo E, et al. Elevated serum interferon gammainduced protein 10 kDa is associated with TAFRO syndrome. *Sci Rep.* 2017;7:42316.
- 45. Pierson SK, Stonestrom AJ, Shilling D, et al. Plasma proteomics identifies a 'chemokine storm' in idiopathic multicentric Castleman disease. *Am J Hematol*. 2018;93(7):902-912.
- Zhang X, Rao H, Xu X, et al. Clinical characteristics and outcomes of Castleman disease: a multicenter study of 185 Chinese patients. *Cancer Sci.* 2018;109(1):199-206.
- 47. Dispenzieri A, Armitage JO, Loe MJ, et al. The clinical spectrum of Castleman's disease. *Am J Hematol.* 2012;87(11):997-1002.
- Jose FF, Kerbauy LN, Perini GF, et al. A life-threatening case of TAFRO syndrome with dramatic response to tocilizumab, rituximab, and pulse steroids: the first case report in Latin America. *Medicine* (*Baltimore*). 2017;96(13):e6271.

49. Hiramatsu S, Ohmura K, Tsuji H, et al. Successful treatment by rituximab in a patient with TAFRO syndrome with cardiomyopathy. *Jpn J Clin Immunol.* 2016;39(1):64-71.

AJH_WILEY

 van Rhee F, Voorhees P, Dispenzieri A, et al. International, evidencebased consensus treatment guidelines for idiopathic multicentric Castleman disease. *Blood.* 2018;132(20):2115-2124.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of this article.

How to cite this article: Fujimoto S, Sakai T, Kawabata H, et al. Is TAFRO syndrome a subtype of idiopathic multicentric Castleman disease? *Am J Hematol*. 2019;94:975–983. <u>https://</u> doi.org/10.1002/ajh.25554