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NOTE

Isohericenone, a new cytotoxic isoindolinone alkaloid from *Hericium erinaceum*

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In our continuing search for structurally interesting and cytotoxic metabolites from Korean wild mushrooms,[1–5] we have collected scores of Korean mushroom species annually and evaluated their MeOH extracts for their antitumor activity in our screening test. Among the collected wild mushrooms, the extract of *Hericium erinaceum* showed significant cytotoxicity against A549, SK-OV-3, SK-MEL-2 and HCT-15 cell lines using a sulforhodamine B (SRB) bioassay. This mushroom *H. erinaceum* (Yamabushitake in Japanese) belonging to the family Hericiaceae is widely known as edible mushroom that grows on dead trunks of hard woods in Korea, Japan, China and Europe. This mushroom has been used as a medicine for the treatment of dyspepsia, gastric ulcer and enervation in traditional Chinese medicine for a long time.[6] This medicinal mushroom is a rich source of unique metabolites. Chemical components of this mushroom have been reported to have hericenones A and B as cytotoxic constituents,[7] hericenones C, D and E as stimulators of nerve growth factor synthesis,[8] hericenone J as an endoplasmic reticulum stress-suppressive substance,[9] hericerin as a pollen growth inhibitor[10] and antitumor-active polysaccharides[11] as the bioactive components. A bioassay-guided fractionation and chemical investigation of its MeOH extract resulted in the isolation of a new isoindolinone alkaloid named isohericenone (**1**), together with nine known compounds, namely isohericerin (**2**),[6] erinacerin B (**3**),[12] hericenone A (**4**),[7] hericenone J (**5**),[9] 3,4-dihydro-5-methoxy-2-methyl-2-(4′-methyl-2′-oxo-3′-pentenyl)-9(7H)-oxo-2H-furo[3,4-h] benzopyran (**6**),[13] erinacerin A (**7**),[12] hericenone F (**8**),[14] hericenone D (**9**)[8] and hericenone E (**10**)[8] (Figure 1). Here, we describe the isolation and structural elucidation of (**1**) as well as the cytotoxic activities of compounds **1–10**.

The half dried fruiting bodies of *H. erinaceum* (5.0 kg) were extracted with 80% aqueous MeOH twice at room temperature and then filtered. The filtrate was evaporated under vacuum to afford a MeOH extract (500 g), which was partitioned with hexane, CH_2Cl_2, EtOAc and *n*-BuOH subsequently with H_2O, yielding hexane (63.3 g), CH_2Cl_2 (4.5 g), EtOAc (2.0 g) and *n*-BuOH fractions (17.5 g). Each fraction was evaluated for its cytotoxicity against A549, SK-OV-3, SK-MEL-2 and HCT-15 cell lines using a SRB bioassay. We selected the hexane-soluble and CH_2Cl_2-soluble fractions for the current phytochemical investigation, because the CH_2Cl_2-soluble fraction was the most active and hexane-soluble fraction also had significant cytotoxicity against the tested tumor cell lines. The active hexane-soluble fraction (60 g) was separated on a silica gel column with a gradient solvent system of hexane-EtOAc (50 : 1–1 : 1) to give five fractions (H1–H5). Fraction H1 (11 g) was separated on a RP-C_{18} silica gel column using a gradient solvent of increasing MeOH in H_2O from 10 to 100% to give three subfractions (H11–H13). Fraction H13 (5 g) was separated twice on a RP-C_{18} silica gel column with a gradient solvent system of MeOH-H_2O (2 : 3–7 : 3) and then purified by RP-C_{18} preparative HPLC (Econosil RP-18 10 μ column (Alltech, Nicholasville, KY, USA), 250 × 10 mm^2) using a solvent of MeOH-H_2O (1 : 1) to yield compound (**5**) (9 mg). Fraction H2 (12 g) was subjected to repeated RP-C_{18} silica gel column separation using a gradient solvent system of MeOH-H_2O (1 : 1–7 : 3) and then purified by RP-C_{18} preparative HPLC (60% MeOH) to afford compound (**6**) (4 mg). Fraction H5 (10 g) was separated on a RP-C_{18} silica gel column using a gradient solvent of increasing MeOH in H_2O from 10 to 100% to give three subfractions (H51-H53). Fraction H51 (1 g) was isolated using repeated RP-C_{18} silica gel column separation with a gradient solvent system of MeOH-H_2O (2 : 3–7 : 3) and then purified by RP-C_{18} preparative HPLC (60% MeOH) to give compound (**9**) (38 mg). Fraction H53 (1 g) was subjected to repeated RP-C_{18} silica gel column separation using a gradient solvent system of MeOH-H_2O (1 : 1–7 : 3) and then purified by RP-C_{18} preparative HPLC (50% MeOH) to afford compound (**8**) (36 mg). The most active CH_2Cl_2-soluble fraction (4 g) was separated on a silica gel column with a gradient solvent system of CH_2Cl_2-MeOH (50 : 1–1 : 1) to yield five fractions (C1–C5). Fraction C2 (1 g) was separated on a RP-C_{18} silica gel column using a gradient solvent of increasing MeOH in H_2O from

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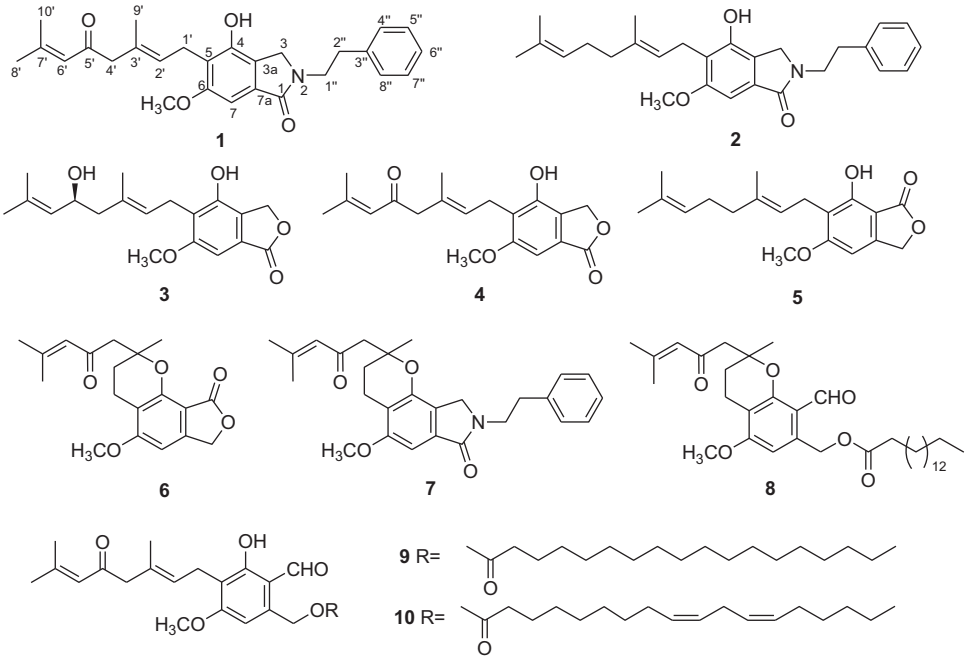

Figure 1 Structures of compounds **1**–**10**.

10 to 100% to furnish five subfractions (C21–C25). Fraction C24 (200 mg) was subjected to passage over a RP-C_{18} silica gel column using a gradient solvent system of MeOH-H_2O (1 : 1–7 : 3) and then purified by RP-C_{18} preparative HPLC (50% MeOH) to give compounds (**1**) (16 mg), (**3**) (4 mg), (**4**) (3 mg), and (**7**) (9 mg). Compounds (**2**) (50 mg) and (**10**) (7 mg) were isolated from fraction C5 (900 mg) using repeated RP-C_{18} silica gel column separation with a gradient solvent system of MeOH-H_2O (2 : 3–7 : 3) and purification with RP-C_{18} preparative HPLC (60% MeOH).

Compound (**1**) was isolated as a colorless gum. Its molecular formula was determined as $C_{27}H_{31}O_4N$ from the $[M+Na]^+$ peak at *m/z* 456.2155 (calculated for $C_{27}H_{31}O_4NNa$, 456.2151) in the positive-ion high resolution (HR)-ESI-MS spectrum. The IR spectrum of (**1**) showed the presence of a hydroxyl group (3357 cm^{-1}), a γ-lactam (1701 cm^{-1}), an α,β-unsaturated ketone (1661 cm^{-1}) and phenyl groups (1593 cm^{-1}). The physico-chemical properties of (**1**) are summarized in Supplementary Information. The ^{1}H- and ^{13}C-NMR spectral data of (**1**) are shown in Table 1.

The ^{1}H- and ^{13}C-NMR spectra (Table 1) of (**1**) were very similar to those of (**2**),[6] with an apparent difference being the presence of signal attributable to an α,β-unsaturated ketone at δ_C 200.9 in (**1**). The shifted signals for C-3′ at δ_C 129.4, for C-4′ at δ_C 54.9 and for C-7′ at δ_C 156.3 were clearly observed in the ^{13}C-NMR spectrum of (**1**), compared with those of corresponding ones in (**2**), suggesting that the α,β-unsaturated ketone in (**1**) was located at C-5′ in combination with the absence of the signal for methylene carbon (δ_C 26.5) at C-5′ of (**2**). This partial structure was confirmed by the identical ^{13}C-NMR chemical shifts of the partial structure (C-1′–C-10′) of (**1**) with those of **4**, **9** and **10**,[7,8] and HMQC and HMBC correlations of (**1**) (Figure 2). The core structure of this molecule, isoindoline-1-one substructure, was unambiguously confirmed by HMBC correlations from H-7 to C-1 and from H-3 to C-4 (Figure 2). The gross structure

Table 1 ^{1}H- and ^{13}C-NMR data of compounds (1) and (2)

	1			2
Position	δ_C	δ_H	*HMBC*	δ_C
1	169.9 s			169.3 s
3	48.6 t	4.17 (s)	C-1, 3a, 4, 7a, 1″	48.5 t
3a	121.2 s			121.3 s
4	150.1 s			150.6 s
5	120.7 s			119.4 s
6	159.3 s			158.8 s
7	96.5 d	6.86 (s)	C-1, 3a, 5, 6, 7a	97.8 d
7a	131.1 s			132.0 s
1′	22.5 t	3.45 (d, 7.5)	C-4, 5, 6, 2′, 3′	23.0 t
2′	127.3 d	5.31 (t, 7.5)	C-5, 1′, 3′, 4′, 9′	121.5 d
3′	129.4 s			138.8 s
4′	54.9 t	2.99 (s)	C-2′, 3′, 5′, 6′, 9′	39.9 t
5′	200.9 s			26.5 t
6′	122.6 d	6.13 (s)	C-4′, 5′, 7′, 8′, 10′	124.0 d
7′	156.3 s			132.1 s
8′	26.5 q	1.81 (s)	C-6′, 7′, 10′	25.8 q
9′	15.4 q	1.74 (s)	C-2′, 3′, 4′	16.3 q
10′	19.6 q	2.07 (s)	C-6′, 7′, 8′	17.8 q
1″	44.2 t	3.84 (t, 7.5)	C-1, 3, 2″, 3″	44.2 t
2″	34.4 t	2.97 (t, 7.5)	C-1″, 3″, 4″, 8″	35.0 t
3″	138.9 s			138.8 s
4″, 8″	128.5 d	7.24 (m)	C-2″, 4″, 6″, 8″	128.8 d
5″, 7″	128.4 d	7.26 (m)	C-3″, 5″, 7″	128.7 d
6″	126.3 d	7.21 (m)	C-4″, 8″	126.7 d
6-OCH_3	55.1 q	3.84 (s)	C-6	56.1 q

Abbreviations: d, doublet; m, multiplet; s, singlet; t, triplet.
NMR data were obtained in 500 MHz for ^{1}H and 125 MHz for ^{13}C in CD_3OD for (**1**) and in $CDCl_3$ for (**2**), and values in parentheses are coupling constants in Hz.

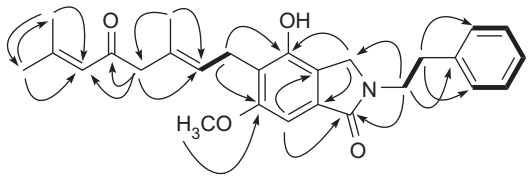

Figure 2 ^{1}H–^{1}H COSY (bold lines) correlations and key HMBC (arrows) of (**1**).

Table 2 Cytotoxic activities of compounds (1), (2), (6) and 7 isolated from *H. erinaceum*.

	IC_{50} (μM)[a]			
Compound	*A549*	*SK-OV-3*	*SK-MEL-2*	*HCT-15*
1	2.6	3.1	1.9	2.9
2	21	8.9	3.1	19
6	17	11	13	16
7	11	11	7.7	14
Doxorubicin[b]	0.001	0.003	0.002	0.081

[a]IC_{50} value of compounds against cancer cell lines, defined as the concentration (μM) that caused 50% inhibition of cell growth *in vitro*.
[b]Doxorubicin as a positive control.

of (**1**) was established by the HMBC experiment showing correlations from H-1′ to C-4 and C-6 and from H-1″ to C-1 and C-3 (Figure 2). On the basis of the above data, the structure of (**1**) was assigned as shown in Figure 1 and the compound was named isohericenone.

Compound (**2**) was isolated as an amorphous powder with the molecular formula $C_{27}H_{33}O_3N$, as determined by the HR-ESI-MS data at *m/z* 442.2352 $[M+Na]^+$ (calculated for $C_{27}H_{33}O_3NNa$, 442.2358). The full NMR assignments of (**2**) were performed by the analysis of the ^{1}H–^{1}H COSY, DEPT, HMQC and HMBC spectroscopic data (Table 1). According to the survey of literature, compound (**2**) was recently reported from this mushroom as isohericerin by Miyazawa *et al.*[6] However, the spectral data of (**2**), particularly ^{13}C-NMR data, were not completely matched with those of isohericerin.[6] We suggest that the ^{13}C-NMR data assignments at C-3a, C-7a and C-2′ of isohericerin should be corrected on the basis of our analysis of 2D-NMR data. It seems that the reported data of C-3a and C-7a should be changed with each other, similar with the case of isohericenone (**1**) and erinacerin A (**7**).[12] Here, the corrected ^{13}C-NMR data of isohericerin (**2**) are reported (Table 1).

Compounds **1–10** were evaluated for their antiproliferative activities against four human cancer cell lines, namely A549, SK-OV-3, SK-MEL-2 and HCT-15 using the SRB bioassay.[15] Compounds (**1**), (**2**), (**6**) and (**7**) showed inhibitory activity against proliferation of the tested cell lines with IC_{50} values in the range of 1.9–21 μM (Table 2). In particular, compound (**1**) exhibited the most potent cytotoxicity against A549, SK-OV-3, SK-MEL-2 and HCT-15 cell lines (IC_{50}: 2.6, 3.1, 1.9 and 2.9 μM, respectively) (Table 2). Compounds (**1**), (**2**), (**6**) and (**7**) may be promising for the development of effective drugs for various cancers. This study shows that these compounds can be considered as contributors to the antitumor activity of the mushroom *H. erinaceum*.

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Supplementary Information accompanies the paper on The Journal of Antibiotics website (http://www.nature.com/ja)