

## Kidney tissue somatomedin C and initial renal growth in diabetic and uninephrectomised rats

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**Summary.** Kidney growth after induction of experimental diabetes in rats was compared to compensatory renal growth in response to unilateral nephrectomy. After 4 days of diabetes, kidney weight had increased from  $816 \pm 21$  mg (SEM) to  $940 \pm 42$  mg (15%). In insulin-treated diabetic rats kidney weight was unchanged at the end of the study, namely  $828 \pm 15$  mg.

In unilaterally nephrectomised rats kidney weight increased from  $840 \pm 20$  mg (SEM) to  $1050 \pm 60$  mg during 4 days (24%). We observed increased kidney content of somatomedin C in both diabetic and uninephrectomised rats. In untreated diabetic rats it was maximal after 48 h, with an increase of 77% ( $3469 \pm 312$  ng/g (SEM) versus  $1961 \pm 173$  ng/g). After 4 days the somatomedin C content had returned to initial levels. In insulin-treated rats somatomedin C content did not increase during the observation period. The

somatomedin C content of the remaining kidney after unilateral nephrectomy was maximal after 24 h with an increase of 58% (from  $1340 \pm 203$  ng/g (SEM) to  $2122 \pm 214$  ng/g). The somatomedin C content returned to normal at day 4. Serum somatomedin C declined insignificantly in diabetic animals during the experimental period, but a significant decrease ( $p < 0.02$ ) was found in uninephrectomised rats. This study demonstrates that kidney somatomedin C peaks during the first or second day after uninephrectomy or induction of diabetes, respectively, and that insulin treatment sufficient to prevent kidney growth abolishes the increase. These similar rapid initial hypertrophies/hyperplasias may thus be dependent on local somatomedin C formation.

**Key words:** Kidney, hypertrophy, nephrectomy, rat, somatomedin C, streptozotocin, diabetes.

It is well-known that kidney size and function are increased in diabetes mellitus of recent onset, as demonstrated in man [1, 2] and in experimental diabetic rats [3]. Kidney size and kidney function remain normal if strict metabolic control is attained immediately [3], or return towards normal when intensive insulin treatment is started early [4].

The mechanism responsible for renal growth and hyperfunction in diabetes is unknown. Several factors have been proposed as being solitary or concurrent causes [5, 6].

Kidney growth following unilateral nephrectomy has been compared to diabetic renal growth, and in many ways the two types of kidney growth are similar, although the initial growth may follow somewhat different cellular reactions [5].

Somatomedin C (SMC) is a polypeptide with potent mitogenic activity in vitro [7, 8]. In vivo, exogenous SMC stimulates growth in hypophysectomised rats [9]. Somatomedins are synthesised at multiple sites and their actions are exerted both distant from [10, 11] and at or near the sites of production [12, 13].

Kidney growth following diabetes of recent onset or uninephrectomy is manifest within 24–48 h. We measured serum and kidney tissue concentrations of SMC each day during the first 4 days after induction of diabetes or unilateral nephrectomy.

### Materials and methods

Two studies were performed: one dealing with streptozotocin diabetic rats and the second with the effects of unilateral nephrectomy.

In the first study male Wistar rats (220–256 g) were used, purchased from Møllegaards Avlsfab. (Eiby, DK).

The rats were kept five in each separate cage in a room with a 12:12 h artificial light cycle (06.00–18.00 hours), temperature  $21 \pm 2^\circ\text{C}$  and humidity  $55 \pm 2\%$  with free access to food and water.

Streptozotocin was given i.p. in a dose of 65 mg/kg body weight at day 0. Animals with a moderate degree of diabetes were produced, having blood glucose exceeding 15 mmol/l from day 1, but there was a minimal loss of body weight in the experimental period, i.e. less than 20 g.

At day 1 the diabetic animals were separated into two groups. One of these was treated with insulin s.c. every morning. A very long-acting heat-treated non-commercial ultralente insulin (NOVO,

Bagsvaerd, DK) was used, administered after blood glucose determination.

Under sodium barbital anaesthesia (50 mg/kg body weight) five diabetic animals had their right kidneys removed at day 0 and day 1. The following 3 days five rats of each group, i.e. insulin and non-insulin treated animals, had their right kidneys removed.

The kidneys were trimmed of fat and capsule, weighed and cut into two parts, one for SMC determination and one for measuring content of DNA, RNA and protein. After weighing, the samples were immediately frozen in liquid nitrogen.

Blood was sampled (500 µl) at day 0 in all rats and from the animals taken out each day for investigation. The samples were taken from the retrobulbar venous plexus through a heparinised capillary tube. Blood was centrifuged and serum frozen for later analysis.

Blood glucose was measured daily (Haemo-Glucotest 1-44 and Reflolux II, Boehringer-Mannheim, Mannheim, FRG). Urine was tested for glucose and ketone bodies every day with Neostix-4 (Ames, Stotze Poges, Slough, UK). The animals were weighed every day.

In the second study 30 male Wistar rats (210–298 g) were used, randomised into 6 groups. Under sodium barbital anaesthesia one group underwent bilateral nephrectomy and one group was sham-operated by flank incision and gentle manipulation of left kidney.

The last four groups underwent left uninephrectomy on day 0; and on each of the following 4 days five rats had their remaining kidneys removed, which was treated as for the diabetic rats.

### Somatomedin C extraction from kidney

SMC extraction was performed as described by D'Escole [12]. The frozen kidney was pulverized in 1 mol/l acetic acid (5 ml/g tissue) on ice bath. With this procedure pH ranges from 3.6–4.2 resulting in maximal liberation of SMC from tissue. The extract was incubated on ice for 2 h, centrifuged 4000 rpm for 15 min and the supernatant decanted. The pellet was reextracted once, the supernatants pooled and then lyophilised to dryness.

The sample was redissolved in 40 mmol/l phosphate buffer, pH 8.0, at a ratio of 10 ml buffer/g tissue weight. The tissue extracts were kept at  $-20^{\circ}$  until SMC assay was performed.

### Somatomedin C radioimmunoassay

SMC was estimated using SMC antibody UB 286 (raised by L.E. Underwood and J.J. van Wyk, Paediatric Endocrinology, University of North Carolina, Chapel Hill, NC USA) donated by the National Hormone and Pituitary Program. For standards and iodination a full amino acid sequence analog (AMGEN Biologicals, Thousand Oaks, Calif, USA) was purchased from Amersham (Buckinghamshire, UK).

All constituents were made up in 40 mmol/l phosphate buffer, pH 8.0, with 0.2% bovine serum albumin (Sigma, St Louis, Mo, USA) and sodium methionate 0.6 mmol/l. Separation was achieved using (6:1) 20% polyethylene glycol 6000 with 0.5% Tween 20 (both from Merck, Darmstadt, FRG). Free and antibody-bound activities were counted.

SMC immunoactivity was measured in diluted rat serum (1:400) after previous extraction in acetic acid-methanol [14].

### RNA, DNA and protein determination

RNA and DNA were separated by a Schmidt-Tannhäuser procedure performed according to Munro and Fleck [15]. DNA was determined with diphenylamine [16].

The protein concentration of the kidney homogenate was determined according to Lowry et al. [17] with bovine albumin as standard.

### Statistical analysis

Statistics were performed using unpaired Student's t-test.

## Results

All diabetic animals had blood glucose exceeding 15 mmol/l at day 1, and a mean level of  $19 \pm 1.0$  mmol/l (SEM) which was maintained in the untreated group, while it was reduced to  $6.2 \pm 1.0$  mmol/l in insulin treated animals. In untreated diabetic rats glucosuria ranged from 14–111 mmol/l. None of the animals had ketonuria exceeding 0.5 mmol/l.

Diabetic animals showed a small initial decrease in body weight, but at end of the study it was not significant. There was no change in body weight in uninephrectomised animals.

Significant differences between untreated and insulin treated diabetic groups of rats as well as between uninephrectomised and sham operated rats are shown in Figure 1.

As seen in Figure 1A (left) kidney weight increased from  $816 \pm 21$  mg (SEM) in control rats to  $940 \pm 42$  mg at day 4 in untreated diabetic rats ( $p < 0.020$ ), while insulin treated diabetic rats had unchanged kidney weight at  $825 \pm 15$  mg after 4 days. Figure 1A (right) illustrates the kidney growth following unilateral nephrectomy increasing from  $840 \pm 20$  mg wet weight (SEM) to  $1050 \pm 60$  mg at day 4 ( $p = 0.010$ ).

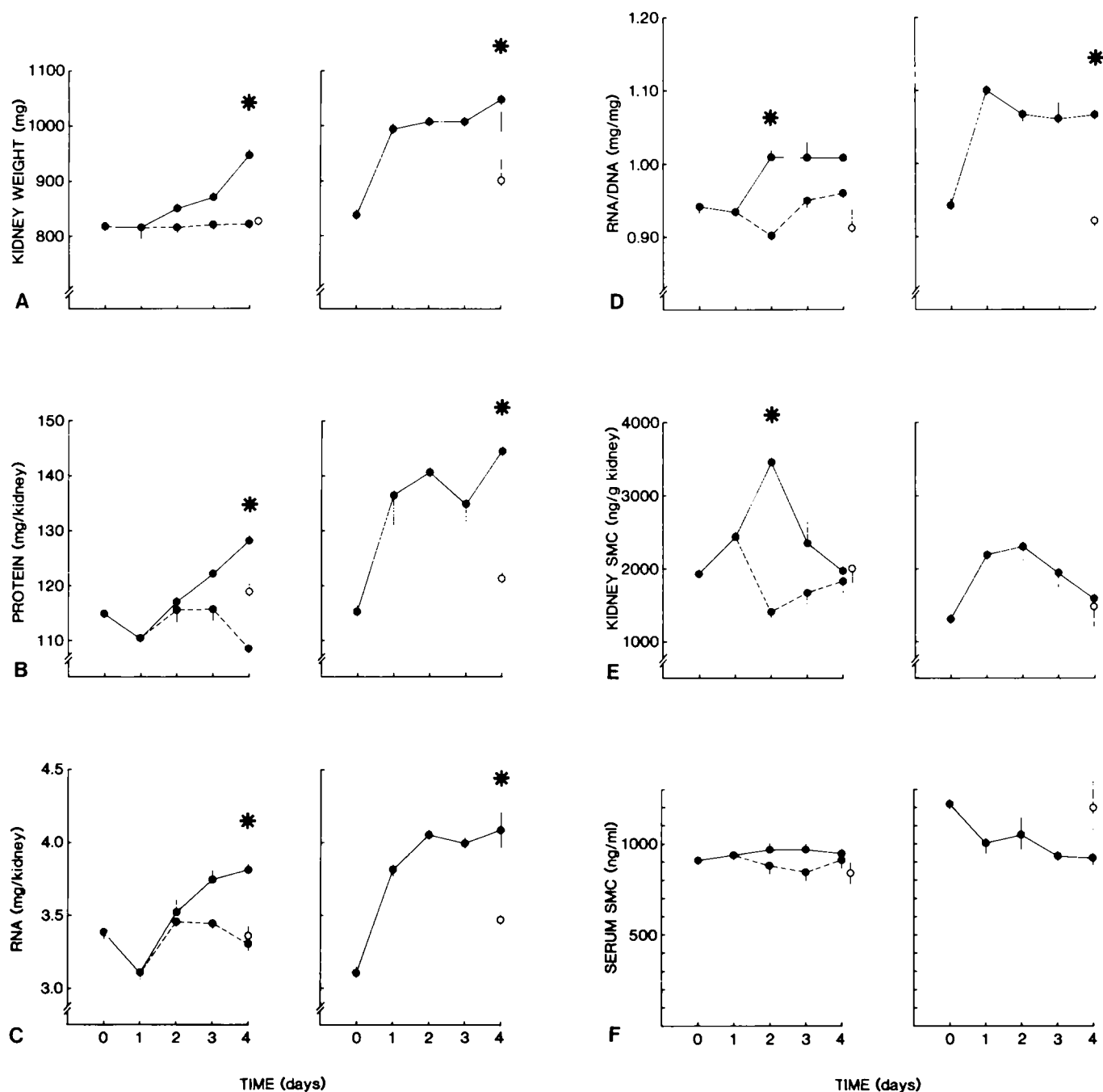
The kidney content of protein, DNA and RNA was estimated. At day 4, there was an 11% increase of protein in untreated diabetic animals ( $p = 0.009$ ) (Fig. 1B (left)) and a 25% increase in uninephrectomised rats ( $p = 0.003$ ) (Fig. 1B (right)), which paralleled the increase pattern in wet kidney weight.

Figure 1C shows an increase in RNA of 10% ( $p = 0.05$ ) in non-insulin treated diabetic rats (left) and 35% ( $p = 0.003$ ) in nephrectomised rats (right).

Untreated diabetic animals had a slight (but insignificant) increase in DNA during the observation period, while an increase of 17% was found in uninephrectomised animals ( $p = 0.042$ ). Figure 1D (left) shows an 8% increase in RNA/DNA ratio in untreated diabetic animals ( $p = 0.048$ ) and an insignificant increase during insulin treatment. In Figure 1D (right) an increase of 14% is shown in uninephrectomised rats ( $p = 0.030$ ). Finally, protein/DNA ratio increased by 9% and 7%, respectively ( $p = 0.030$  and  $p = 0.041$ ).

Assuming that DNA content is an expression of the number of cells and the protein/DNA ratio an expression of the average cell size, it can be estimated that diabetic kidney growth was mainly due to cellular hypertrophy, while postnephrectomy kidney growth was due to roughly 60% hyperplasia and 40% hypertrophy.

Figure 1E (left) illustrates the kidney content of SMC in diabetic rats. It reached maximum in untreated diabetic rats after 48 h and increased by 77% ( $1961 \pm 173$  ng/g (SEM) to  $3469 \pm 312$  ng/g) ( $p = 0.001$ ). The following two days the SMC content returned to the same level as in the non-diabetic control rats. When insulin treatment was started (one day



**Fig. 1 A-F.** Changes over 4 days in streptozotocin diabetic rats (left) and unilaterally nephrectomised rats (right) in **A** kidney weight; **B** kidney protein; **C** kidney RNA; **D** kidney RNA/DNA ratio; **E** kidney somatomedin C (SMC); and **F** serum SMC. Each point represents 5 different animals  $\pm$  SEM, except for day 0 in serum SMC, which includes all 45 rats in the diabetic study and all 30 rats in the uninephrectomy study. Untreated diabetic rats (●—●); insulin treated diabetic rats (●- - -●); non-diabetic control rats (○). Nephrectomised rats (●—●); sham operated rats (○).

\*  $p < 0.05$ , between treated and untreated diabetic groups and between nephrectomised and sham operated rats

after streptozotocin injection) serum SMC content fell after the slight elevation at day 1 and settled around the concentration in control animals.

As shown in Figure 1 E (right) kidney SMC in the remaining kidney after uninephrectomy was maximal after 24 h, with an increase of 58% (from  $1340 \pm 203$  ng/g (SEM) to  $2122 \pm 214$  ng/g ( $p = 0.025$ )). At day 4 the level was  $1575 \pm 229$  ng/g compared to sham op-

erated animals with a tissue content of  $1527 \pm 296$  ng/g.

These results, however, do not exclude the theoretical possibility that sham operated rats might have exhibited a rise in the contralateral kidney SMC content after 24 h. We therefore performed an additional short-term study comparing 5 nephrectomised to 5 sham operated rats and found a kidney weight increase in the

former ( $796 \pm 27$  vs  $700 \pm 26$  mg (SEM,  $p=0.05$ ) and no 24-h increase in the latter ( $712 \pm 13$ , which was lower than  $796 \pm 27$  mg,  $p=0.02$ ). Kidney SMC rose by 22% (from  $1271 \pm 61$  to  $1550 \pm 70$  ng/g,  $p=0.005$ ) while no 24 h increase was noted in sham operated rats ( $1148 \pm 40$  ng/g, which was lower than  $1550 \pm 70$  ng/g,  $p=0.001$ ).

Figure 1F shows serum SMC levels. In diabetic animals (insulin-treated as well as untreated) no significant changes were found (left). In uninephrectomised rats serum SMC concentrations fell from  $1221 \pm 34$  ng/ml (SEM) at day 0 to  $1000 \pm 66$  ng/ml at day 1 ( $p=0.02$ ) and remained at this lower level throughout the observation period (Fig. 1F (right)). Sham operated rats had a serum concentration of  $1200 \pm 177$  ng/ml at day 4.

## Discussion

Our study confirms previous observations that diabetes of recent onset as well as unilateral nephrectomy are followed by rapid kidney growth, involving both hypertrophy and hyperplasia [18, 19]. SMC is one of several growth factors having these effects, which would qualify it for being the "renotrophic factor". Severe experimental diabetes with general wasting, i.e. when blood glucose levels exceed 25 mmol/l, makes comparisons with control rats difficult and inhibits kidney growth [18, 20]; therefore, a moderate streptozotocin diabetes was induced in the present study.

It has been demonstrated that onset of diabetes is followed by decreased *serum levels* of SMC within one week, probably mainly due to impaired hepatic production, and that these changes are corrected by insulin treatment [21, 22]. In the present study no significant decrease was found in the untreated diabetic rats, possibly because of the short observation period and the comparatively mild degree of metabolic derangement.

Traditionally SMC was considered to originate in the liver, promoting growth at distant sites [10, 11]; but recent studies [12, 13] show that the somatomedins may act through paracrine or autocrine mechanisms, having major biological actions at or near peripheral sites of origin.

In a recent report, which addresses the potential role of local production of SMC in postnephrectomy renal growth in rats, tissue levels of SMC were increased after 5 days but no significant change was found after 24 h [23]. Serum SMC levels were unchanged.

We observed increased kidney content of SMC during initial kidney growth in diabetic as well as uninephrectomised rats. In the latter a prompt increase occurred, maximal after 24 h, in good accordance with the pronounced increase in kidney wet weight during the first 24 h following nephrectomy. In diabetic rats

the SMC content was maximal after 48 h in accord with the somewhat slower increase in kidney weight; further, the increase in kidney size and SMC content was prevented by insulin treatment. This excluded also the possibility that the kidney SMC increase could be caused by streptozotocin.

It has recently been demonstrated that exogenous SMC, given to Snell dwarf mice, increases kidney weight [24]. However, in the present study kidney growth was not induced by increased liver production of SMC, as no elevation of serum SMC was observed. Still, the rise might, theoretically, be in part due to the greater blood volume of the hypertrophying kidney. In the study referred to above Stiles et al. [23] applied a correction factor using the haemoglobin concentration in the acetic acid extract and concluded that the contribution from plasma was insignificant. In our hands the concentration of haemoglobin in extracts was too low to allow for any meaningful correction. However, it can be calculated that the contents of the simultaneously expanding intravascular space can maximally account for about 2% of the 58% and 77% increase in tissue SMC found in the present experiments.

Possible explanations for the SMC increase include a passive capture of SMC secondary to the increase in renal plasma flow (RPF) and glomerular filtration rate (GFR) occurring after uninephrectomy as well as induction of malregulated diabetes, in keeping with the fact that the kidney is a major site for removal of plasma SMC [25]. However, kidney SMC increase patterns (Fig. 1E) do not follow the known alterations in RPF and GFR in the two experimental situations [26-29].

Secondly, it could be that the kidney SMC is taken up from plasma after induction of an increased number of specific SMC receptors in kidney. This theory could be supported through binding studies *in vitro*. However, the rather stable serum SMC levels in the present experiments do not support this possibility.

Finally, and more intriguing in our view, would be an induction of augmented local formation of kidney SMC exerting paracrine effects; this could possibly also be substantiated *in vitro*.

It is well documented that glomeruli grow very quickly during the first four days of experimental diabetes, faster than the rest of the kidney [30]; this is absent in compensatory growth. In a study of  $^3\text{H}$ -thymidine incorporation in kidneys of diabetic and uninephrectomised rats it was found that tubular growth was due to hyperplasia as well as hypertrophy while initial glomerular enlargement was due to hypertrophy only [31]. Noteworthy is that the rate of  $^3\text{H}$ -thymidine incorporation was maximal 2 days after induction of diabetes or uninephrectomy, with a decline at day 4 and 6, similar to changes in the kidney SMC concentrations in the present study.

The rapid accumulation of potent mitogenic SMC in the present work and the studies of thymidine incorporation referred to above support a causal relation-

ship between kidney SMC increase and initial kidney growth. In other words, SMC may be the "renotrophic factor".

Compensatory kidney growth after uninephrectomy is relevant from a (patho)physiological point of view; but it is difficult to see any evident advantages in the similar abrupt increase in function and size after onset of imperfectly controlled diabetic metabolism. It has been suggested to be harmful; and possibly related to the development of late diabetic nephropathy [32-35].

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## References

- Mogensen CE, Andersen MJF (1973) Increased kidney size and glomerular filtration rate in early juvenile diabetes. *Diabetes* 22: 706-712
- Mogensen CE (1971) Glomerular filtration rate and renal plasma flow in short-term and long-term juvenile diabetes mellitus. *Scand J Clin Lab Invest* 28: 91-100
- Seyer-Hansen K (1976) Renal hypertrophy in streptozotocin-diabetic rats. *Clin Sci* 51: 551-555
- Mogensen CE, Andersen MJR (1975) Increased kidney size and glomerular filtration rate in untreated juvenile diabetes: normalization by insulin treatment. *Diabetologia* 11: 221-224
- Seyer-Hansen K (1983) Renal hypertrophy in experimental diabetes mellitus. *Kidney Int* 23: 643-646
- Mogensen CE (1982) Diabetes mellitus and the kidney. An introduction. *Kidney Int* 21: 673-675
- Clemmons DR, Underwood LE, Van Wyk JJ (1981) Hormonal control of immunoreactive somatomedin production by cultured human fibroblasts. *J Clin Invest* 67: 10-19
- Zapf J, Froesch ER, Humbel RE (1981) The IGF of human serum: chemical and biological characterization and aspects of their possible physiological role. *Curr Top Cell Regul* 19: 257-309
- Schoenle E, Zapf J, Humbel RE, Froesch ER (1982) Insulin-like growth factor I stimulates growth in hypophysectomized rats. *Nature* 296: 252-253
- Salmon WD Jr, Daughaday WH (1957) A hormonally controlled serum factor which stimulates sulfate incorporation by cartilage in vitro. *J Lab Clin Med* 49: 825-836
- Daughaday WH, Hall K, Raben MS, Salmon WD Jr, Van den Brande JL, Van Wyk JJ (1972) Somatomedin: proposed designation for sulfation factor. *Nature* 235: 107
- D'Ercole AJ, Stiles AD, Underwood LE (1984) Tissue concentration of somatomedin C: further evidence for multiple sites of synthesis and paracrine/autocrine mechanisms of actions. *Proc Natl Acad Sci USA* 81: 935-939
- Thorlacius-Ussing O, Flyvbjerg A, Jørgensen K, Damm, Ørskov H (1988) Growth hormone restores normal growth in selenium treated rats without increase in circulating SMC. *Acta Endocrinol* 117: 65-72
- Thorlacius-Ussing O, Flyvbjerg A, Esmann J (1987) Evidence that selenium induces growth retardation through reduced growth hormone and SMC production. *Endocrinology* 120: 659-663
- Munro HN, Fleck A (1966) The determination of nucleic acids. *Methods Biochem Anal* 14: 113-176
- Burton K (1956) A study of the conditions and mechanism of the diphenylamine reaction for a colorimetric estimation of deoxyribonucleic acid. *Biochem J* 62: 315-323
- Lowry OH, Osebrough NJ, Fara AL, Randall RL (1951) Porcine measurements with the Folin phenol reagents. *J Biol Chem* 193: 165-175
- Seyer-Hansen K (1978) Renal hypertrophy in experimental diabetes: a comparison to compensatory hypertrophy. *Diabetologia* 14: 325-328
- Preuss HG (1983) Compensatory renal growth symposium. An introduction. *Kidney Int* 23: 571-574
- Seyer-Hansen K (1977) Renal hypertrophy in experimental diabetes: relation to severity of diabetes. *Diabetologia* 13: 141-143
- Maes M, Ketelslegers JM, Underwood LE (1983) Low plasma SMC in streptozotocin-induced diabetes mellitus. *Diabetes* 32: 1060-1069
- Maes M, Underwood LE, Ketelslegers JM (1986) Low serum SMC in insulin-dependent diabetes: evidence for a postreceptor mechanism. *Endocrinology* 118: 377-382
- Stiles AD, Sosenko IRS, D'Ercole AJ, Smith BT (1985) Relations of kidney tissue somatomedin-C/insulin-like growth factor I to postnephrectomy renal growth in the rat. *Endocrinology* 117: 2397-2401
- Van Buul-Offers S, Ueda I, Van den Brande JL (1986) Biosynthetic somatomedin C (SM-C/IGF I) increases the length and weight of snell dwarf mice. *Pediatr Res* 20: 825-827
- D'Ercole AJ, Decedue CJ, Furlanetto RB, Underwood L, Van Wyk JJ (1977) Evidence that SMC is degraded by the kidney and inhibits insulin degradation. *Endocrinology* 101: 577-586
- Fleck CH, Bräunlich H (1984) Kidney function after unilateral nephrectomy. *Exp Pathol* 25: 3-18
- Michels LD, Davidman M, Keane WF (1981) Determinants of glomerular filtration and plasma flow in experimental diabetic rats. *J Lab Clin Med* 98: 869-885
- Seyer-Hansen K (1987) Renal hypertrophy in experimental diabetes: some functional aspects. *J Diabetic Complications* 1: 7-10
- Jensen PK, Christiansen JS, Steven K, Parving H-H (1981) Renal function in streptozotocin diabetic rats. *Diabetologia* 21: 409-414
- Seyer-Hansen K, Hansen J, Gundersen HJG (1980) Renal hypertrophy in experimental diabetes. A morphometric study. *Diabetologia* 18: 501-505
- Rasch R, Rytter-Nørgaard JO (1983) Renal enlargement: comparative autoradiographic studies of <sup>3</sup>H-thymidine uptake in diabetic and uninephrectomized rats. *Diabetologia* 25: 280-287
- Mogensen CE, Steffes MW, Deckert T, Christiansen JS (1981) Functional and morphological renal manifestations in diabetes mellitus. *Diabetologia* 21: 89-93
- Brenner BM, Hostetter TH, Olsen JL, Rennke HG, Venkatachalam MA (1981) The role of glomerular hyperfiltration in the initiation and progression of diabetic nephropathy. *Acta Endocrinol (Copenh)* 97: [Suppl.] 242: 7-10
- Ørskov H (1985) Growth hormone hyperproduction inducing some of the vicious circles in diabetes mellitus. *Acta Med Scand* 217: 343-346
- Mogensen CE (1986) Early glomerular hyperfiltration in insulin-dependent diabetics and late nephropathy. *Scand J Clin Lab Invest* 46: 201-206

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