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# KIF1A variants are a frequent cause of autosomal dominant hereditary spastic paraplegia 

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#### Abstract

Variants in the KIF1A gene can cause autosomal recessive spastic paraplegia 30, autosomal recessive hereditary sensory neuropathy, or autosomal (de novo) dominant mental retardation type 9. More recently, variants in KIF1A have also been described in a few cases with autosomal dominant spastic paraplegia. Here, we describe 20 KIF1A variants in 24 patients from a clinical exome sequencing cohort of 347 individuals with a mostly 'pure' spastic paraplegia. In these patients, spastic paraplegia was slowly progressive and mostly pure, but with a highly variable disease onset ( $0-57$ years). Segregation analyses showed a de novo occurrence in seven cases, and a dominant inheritance pattern in 11 families. The motor domain of KIF1A is a hotspot for disease causing variants in autosomal dominant spastic paraplegia, similar to mental retardation type 9 and recessive spastic paraplegia type 30 . However, unlike these allelic disorders, dominant spastic paraplegia was also caused by loss-of-function variants outside this domain in six families. Finally, three missense variants were outside the motor domain and need further characterization. In conclusion, KIF1A variants are a frequent cause of autosomal dominant spastic paraplegia in our cohort ( $6-7 \%$ ). The identification of KIF1A loss-of-function variants suggests haploinsufficiency as a possible mechanism in autosomal dominant spastic paraplegia.


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## Introduction

Hereditary spastic paraplegia (HSP) is a neurodegenerative disorder that affects motor control over the lower limbs and the bladder, with an incidence of $\sim 1-10$ per 100,000 individuals [1]. In the pure form, patients suffer from lower extremity spasticity and weakness and often from urinary urgency. In complicated forms, HSP may be associated with additional neurologic abnormalities, such as intellectual disability, peripheral neuropathy, ataxia, distal muscle wasting, and optic neuropathy [2]. HSP may follow a (de novo) autosomal dominant, autosomal recessive, X-linked, or mitochondrial inheritance pattern. At this moment, 79 loci and 61 corresponding genes have been associated with HSP [3]. The most common autosomal dominant HSP is spastic paraplegia (SPG) type 4, caused by a variant in the $S P A S T$ gene ( $40 \%$ ), while $S P G 7$ and SPG11 variants are the most frequent causes of recessive forms.

Variants in KIF1A have been described in three different disorders in OMIM. The first is autosomal recessive hereditary sensory neuropathy IIC (MIM \# 614213) that has been described in four families. All families had the same frameshift variant that resides in an alternative transcript (NM_001244008.1/NP_001230937.1), suggesting that absence of that alternative transcript was the cause of hereditary sensory neuropathy IIC [4]. The second is autosomal dominant mental retardation type 9 (MIM \# 614255), which is characterized by severe cognitive impairment with spastic paraparesis, axonal neuropathy, epilepsy and/or cerebellar atrophy. At least 17 de novo missense variants affecting the kinesin motor domain of KIF1A have been described in this syndrome [5-8]. Functional analyses suggested a dominantnegative effect of these missense variants on the movement of this kinesin along the microtubules. The third disorder associated with KIF1A is autosomal recessive SPG type 30 (MIM\#610357), characterized by slowly progressive pure HSP with an onset in the first or second decade. Missense variants in the kinesin domain of KIF1A are responsible for the three autosomal recessive SPG30 families described to date $[9,10]$.

A few reports of autosomal dominant SPG make up for a possible fourth disorder, not yet described in OMIM, associated with KIF1A [11-13]. Similar to dominant mental retardation type 9 and recessive SPG30, all currently reported KIF1A variants in dominant, pure HSP were also missense variants affecting the kinesin motor domain, with the exception of one variant of uncertain significance.

By exome sequencing, we identified variants in the KIF1A gene in 24 out of 347 probands with SPG. Variants are not only missense variants in the motor domain of KIF1A, but also loss-of-function variants outside this domain, expanding the spectrum of variants.

## Subjects and methods

## Editorial policies and ethical consideration

Clinical exome sequencing was approved by the Medical Review Ethics Committee, Region Arnhem-Nijmegen, Number 2011/188. All human subjects provided informed consent for this study.

## Patients

Patients with HSP from The Netherlands ( $80 \%$ ) and other European countries $(20 \%)$ were included in clinical exome sequencing as part of their diagnostic work-up. No restrictions on age at onset or inheritance pattern were made, but there had to be a clinical suspicion of a genetic etiology. All patients were counseled by a clinical geneticist or neurologist. From each family, only one patient (proband) was included for exome sequencing. Candidate variants were verified by Sanger sequencing in affected family members whenever possible.

Previous gene testing in some probands of the families presented here excluded the most common forms of autosomal dominant SPG, types 4 and 3A, caused by variants in the SPAST gene (13/24 probands) or the ATL1 gene (7/24 probands), respectively.

Available clinical and imaging data of the probands and affected family members ( 30 in total) were systematically collected and reviewed.

## Exome sequencing and data analysis

Exome sequencing was performed as previously described [14]. In summary, capture of exons was done using an Agilent SureSelect Human All Exon 50 Mb Kit (Santa Clara, CA, USA). Sequencing was performed using a LifeTechnologies 5500XL machine (Thermo Fisher, Waltham, MA, USA) or an Illumina Hiseq 2000 or 4000 (San Diego, CA, USA). Read mapping and variant calling were done using LifeScope Life Technologies (Thermo Fisher) for the 5500XL data or BWA (mapping) and GATK (calling) for the Illumina data. A filter for a 'movement disorders' gene panel was applied. This panel consists of $\sim 300$ genes implicated in various forms of cerebellar ataxia, HSP, genetic choreas, and other hyperkinetic movement disorders [15] and is available from our website (https://www.radboudumc.nl/en/patientenzorg/onderzoeken/ exome-sequencing-diagnostics/exomepanelspreviousversions/ movement-disorders). The single nucleotide variant data were filtered by [1] frequency ( $<5 \% \mathrm{dbSNP},<1 \%$ in house database, gnomAD, EXAC [16]) [2] nucleotide and amino acid conservation and [3] exon/intronic position ( -20 till +8 ). Accession number NM_004321.7 and HGVS were used for nomenclature of variants in the KIF1A gene. The likelihood of

Table 1 KIF1A variants in autosomal dominant HSP

| Family | Variant (heterozygous) DNA; protein | Domain | CADD (PHRED) | Freq in ExAC/ gnomAD | Evolutionary conservation | LOVD ID |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| P1 | c. $89 \mathrm{~T}>\mathrm{C}$; p.(Met30Thr) | Motor | 26.4 | 0/0 | M, F, DM | 00225586 |
| P2 | c.167A>G; p.(Tyr56Cys) | Motor | 28.6 | 0/0 | M, F, DM, CE | 00225587 |
| P3 | c. $206 \mathrm{C}>$ T; p.(Ser69Leu) | Motor | 16.3 | 0/0 | M | 00246956 |
| P4 | c. $221 \mathrm{~A}>\mathrm{G} ; \mathrm{p} .(\mathrm{Tyr} 74 \mathrm{Cys})^{\text {a }}$ | Motor | 25.7 | 0/0 | M, F, DM, CE | 00225589 |
| P5 | c. $232 \mathrm{G}>\mathrm{A}$; p.(Gly 78 Ser ) | Motor | 34 | 0/0 | M, F, DM, CE | 00225590 |
| P6 | c.317C>A; p.(Thr106Asn) | Motor | 26.3 | 0/0 | M, F, DM, CE | 00225591 |
| P7 | c. $500 \mathrm{G}>\mathrm{A} ; \mathrm{p} .(\operatorname{Arg} 167 \mathrm{His})$ | Motor | 34 | 0/0 | M, F, DM, CE | 00225592 |
| P8 | c.518T>C; p.(Leu173Pro) | Motor | 27 | 0/0 | M, F, DM, CE | 00225593 |
| P9/10 | c.756C>G; p.(Ser252Arg) | Motor | 25.1 | 0/0 | M, F, DM, CE | $\begin{aligned} & 00225594 \\ & 00246690 \end{aligned}$ |
| P11/12/13 | c.773C>T; p.(Thr258Met) | Motor | 28.5 | 0/0 | M, F, DM, CE | $\begin{aligned} & 00225610 \\ & 00246691 \\ & 00246692 \end{aligned}$ |
| P14 | c. $1048 \mathrm{C}>\mathrm{T}$; p. $(\mathrm{Arg} 350 \mathrm{Trp})$ | Motor | 12.95 | 0/0 | M, F, DM, CE | 00225611 |
| P15 | c. $1379 \mathrm{C}>\mathrm{G} ; \mathrm{p}$.(Ala460Gly) | Coiled coil | 25.3 | 0/0 | M, F | 00225612 |
| P16 | c.1867C>T; p.(Gln623*) | ni | 38 | 0/0 | ni | 00225613 |
| P17/18 | c. $2527 \mathrm{C}>$ T; p. (Arg843Cys) | - | 35 | 1/244238 | M, F | $\begin{aligned} & 00225614 \\ & 00246693 \end{aligned}$ |
| P19 | c. $2577 \mathrm{C}>\mathrm{A}$; p.(Asn859Lys) | - | 28.9 | 0/0 | M, F, DM, CE | 00225615 |
| P20 | c.3975C>G; p.(Tyr1325*) | ni | 29.1 | 0/0 | ni | 00225616 |
| P21 | c.4096_4103dup; (p.(Asp1369fs) | ni | 11.05 | 0/0 | ni | 00343920 |
| P22 | c.4292del; p.(Pro1431fs) | ni | 22.5 | 0/0 | M, F | 00225617 |
| P23 | c.4740dup; p.(Tyr1581fs) | ni | 35 | 0/0 | ni | 00225618 |
| P24 | Chr2[GRCh37]:g.(238475818_ 238482964)_(243037178_qter)del | ni | ni | ni | ni | 00245206 |

Variants are described according to HGVS nomenclature. Protein domains are according to UniProt consortium (https://www.uniprot.org). Due to nonsense-mediated decay domain of loss-of-function variants are not informative (ni). CADD scores (vs 1.3) and population frequencies were obtained through https://cadd.gs.washington.edu/ and http://exac.broadinstitute.org, respectively. Conservation was determined using ensemble alignments
$M$ mammals, $F$ fish, $D M$ Drosophila melanogaster, CE C.elegans, ni not informative
${ }^{\text {a }}$ This family has been described [15]
the disease causing effect of these variants was based on frequencies (absence) in control populations, evolutionary conservation, and segregation analyses. The KIF1A variants described in this paper were submitted to LOVD (https://data bases.lovd.nl/shared/genes/KIF1A).

Copy number variant (CNV) analyses from the WES data were done as described [17]. Essentially, it was done using CoNIFER (http://conifer.sourceforge.net/) [18]. CNVs with an absolute $Z$-score $>1.7$ were considered for analysis. To reduce false calls due to potential batch effects, analyses are performed using the most recent samples as controls. CNVs were annotated based on the number of RefSeq exons affected, frequency of CNVs within the cohort, and overlap with disease genes from the movement disorders gene panel. Sanger sequencing was used to confirm the presence of the identified variants if the quality score (from GATK) was not sufficient ( $<500$ ).

In addition, Sanger sequencing was used for segregation analyses and to exclude the possibility of bi-allelic KIF1A variants.

## Results

## Genetic findings

A cohort of 347 probands with a predominantly ( $>90 \%$ ) pure SPG was referred to our expert centre for 'genetic movement disorders'. Clinical exome sequencing was performed as described [14]. Twenty different heterozygous KIF1A variants were detected in 24 probands (Table 1 and Fig. 1). One of those variants (in family 24) was a chromosome 2 q 37 contiguous gene deletion including the entire KIF1A gene (Chr2 [GRCh37]:g.(238475818_238482964)_


Fig. 1 Schematic representation of the KIF1A protein with variants in dominant spastic paraplegia. Variants identified in this study are depicted above the schematic protein, isoform NP_004312.2, while previously described variants in autosomal dominant spastic
paraplegia [11-13, 15, 29-32] are shown below the schematic. Protein domains are according to UniProt consortium (https://www.uniprot. org), with kinesin motor, cc coiled coil, FHA forkhead-associated, PH pleckstrin homology domains. ${ }^{\text {a }}$ This family has been described [15]
(243037178_qter)del, Supplemental Fig.). Throughout the remainder of the paper, the variants are discussed by the protein change deduced from their DNA change. Three variants were recurrent, p.(Ser252Arg) in two, p. (Thr258Met) in three, and p.(Arg843Cys) in two families. Targeted sequencing of the KIF1A gene was subsequently performed to exclude putative bi-allelic variants, rendering a possible recessive inheritance unlikely. All KIF1A variants were absent from an in-house database of $>30,000$ clinical exomes, and from large sequencing cohorts in gnomAD [16], except for the p.(Arg843Cys) variant that was heterozygously present in one individual in gnomAD. In addition, missense variants were of evolutionary conserved amino acids (in all vertebrates and many even in lower species; Table 1). Two variants, p.(Ser69Leu) and p. (Thr258Met), were reported previously in autosomal dominant HSP [11, 13]. No overlap was seen with the KIF1A variants reported in the allelic disorders autosomal recessive hereditary sensory neuropathy IIC, autosomal dominant mental retardation type 9 , or autosomal recessive SPG30.

## De novo and inherited autosomal dominant KIF1A variants

In 7 of the 24 patients, the KIF1A variants (p.(Gly78Ser), p.(Thr106Asn), p.(Thr258Met) in three families, (Tyr1581fs) and the Chr2q37 deletion) had occurred de novo in the first affected generation (Fig. 2; P5, P6, P11, P12, P13, P23, and P24), which was revealed by parental testing including a paternity test. Eleven variants, including
one that was de novo in the first generation, were detected in multiple affected family members in multiple generations (P2, P3, P4, P8, P11, P15, P16, P17, P20, P21, and P22), though families usually were small (Fig. 2). In four families (P1, P7, P9, P14), a dominant inheritance was evident, but family members refrained from testing or were deceased. The remaining three probands did not have affected family members and parents were unavailable for testing for de novo occurrence (P10, P18, and P19).

## Variants in the KIF1A gene in- and outside the kinesin motor domain

Variants causing autosomal dominant mental retardation type 9 or SPG30 invariably are missense variants located in the kinesin motor domain of the KIF1A protein (Fig. 3), an important domain for 'walking' of the kinesin along microtubules. Functional analyses revealed a dominantnegative effect of these variants in transport along microtubules [5]. Eleven of the twenty variants in this study were also missense variants located in this motor domain (Figs. 1 and 3), suggesting that these variants cause dominant SPG. However, another nine variants were detected outside the motor domain. In contrast to the motor domain variants, six of the variants outside the domain are considered loss-of-function variants (p.(Gly623)*, p.(Tyr1325*), p.(Asp1369fs), p.(Pro1431fs), p.(Tyr1581fs), and the Chr2q37 deletion including KIF1A). Two of the loss-offunction variants were shown to be de novo occurrences (P23 and P24) and the four others (P16, P20, P21, and P22)



Fig. 3 Schematic representation of the KIF1A protein with all variants in the allelic disorders. All variants identified in spastic paraplegia are depicted above the schematic protein in red open triangles (autosomal dominant) or blue solid triangles (autosomal recessive), while variants
in mental retardation type 9 and hereditary sensory neuropathy IIC are shown below the schematic in green open diamonds and purple solid dots, respectively. Protein domains are according to UniProt consortium (https://www.uniprot.org)

However, he had a de novo chromosome 2 q 37 contiguous gene deletion including KIF1A and HDAC4 that very likely explains the more complex phenotype.

## Discussion

In conclusion, we report 24 probands with autosomal dominant SPG with 20 heterozygous variants in KIF1A. Three variants were recurrent in our cohort. SPG due to KIF1A variants was rather pure. However, two families had cognitive impairment, reflected by learning difficulties and borderline intellectual disability in one patient, whereas one additional proband had a complex phenotype and a de novo contiguous gene deletion including KIF1A. Seventeen variants in 20 families are likely to cause dominant SPG, because they are missense variants in the kinesin domain (11 variants, of which 5 had occurred de novo) or are very likely loss-of-function variants outside the motor domain (six variants, of which two were de novo). Three variants, in four probands, are considered to be of uncertain significance and would need further characterization. In addition, all variants were absent or found only once (p.(Arg843Cys)) in controls and affect evolutionary conserved amino acids in alignments of orthologs. The presence of 20 likely disease causing variants (or 24 including the variants of uncertain significance) out of 347 families (6-7\%) makes KIF1A a frequent cause of autosomal dominant pure SPG.

## Phenotypic differences in KIF1A related disease

Most variants in the allelic disorders associated with KIF1A are located in the kinase motor domain suggesting a common molecular mechanism. The differences in disease severity between autosomal dominant mental retardation type 9, with a severe neurological involvement, and the relative mild HSP in both recessive (SPG30) and dominant forms is therefore intriguing. One explanation may be a difference in the level of impairment of transport by KIF1A.

Kinesins are molecular motors involved in the transport of cargo (e.g., organelles, proteins, mRNA) to their sites of action. The motor domains of kinesins are involved in binding and movement along microtubules, whereas other
Table 2 Clinical characteristics of probands with KIF1A variants

| Patient | P1 | P2 | P3 | P4 | P5 | P6 | P7 | P8 | P9 | P10 | P11 | P12 | P13 | P14 | P15 | P16 | P17 | P18 | P19 | P20 | P21 | P22 | P23 | P24 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Family history | $+$ | + | + | $+$ | - | - | $+$ | $+$ | $+$ | - | + | - | - | + | + | + | + | ? | - | + | + | + | - | - |
| Age at onset (y) | <1 | 9 | 13 | <10 | 21 | $>18$ | 50 | <10 | 57 | <10 | <10 | <1 | 1 | <10 | ~20 | $\sim 30$ | 1 | 46 | 10 | <10 | 2 | 2 | <5 | 1 |
| Age (years) | 56 | 14 | 17 | 53 | 38 | 54 | 64 | 50 | 62 | 49 | 18 | 4 | 3 | 54 | 28 | 60 | 19 | 56 | 51 | 66 | 12 | 22 | 20 | 6,5 |
| Cognition ${ }^{\text {a }}$ | N | N | IQ80 | N | N | N | N | IQ77 | N | N | N | N | N | N | N | N | N | N | N | N | N | N | N | IQ57 |
| Walking aid | - | - | - | + | - | + | + | + | - | - | - | + | - | - | - | - | - | - | - | - | - | - | - | - |
| LL spasticity | $+$ | + | $+$ | $+$ | + | $+$ | $+$ | $+$ | $+$ | + | + | $+$ | + | + | + | + | + | + | + | + | $+$ | $+$ | $+$ | $+$ |
| LL weakness | $+$ | + | + | + | $+$ | - | - | + | $+$ | - | - | + | + | - | - | + |  | - | - | - | + | - | + | + |
| LL deep sensory disturbances | $+$ | - | + | - | $+$ | - | $+$ | + | - | - | - | + | - | + | - | + |  | - | - | - | - | - | ? | ? |
| PTR | $\uparrow$ | ? | $\uparrow$ | $\uparrow$ | $\uparrow \uparrow$ | $\uparrow$ | $\uparrow \uparrow$ | $\uparrow \uparrow$ | $\uparrow \uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ |
| ATR | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\downarrow$ | $\uparrow$ | $\uparrow$ | $\uparrow \uparrow$ | $\uparrow \uparrow$ | $\uparrow \uparrow$ | $\uparrow$ | $\uparrow$ | $\downarrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ |
| Extensor plantar response | + | + | + | + | $+$ | + | $+$ | + | $+$ | $+$ | + | + | + | + | + | + | + | + | + | + | + | + | + | - |
| UL spasticity | - | - | - | - | - | - | - | - | - | - | - | + | - | - | - | - | + | - | - | - | - | - | - | - |
| UL weakness | - | + | - | - | - | - | - | - | - | - | - | - | - | - | - | - | ? | - | - | - | - | - | - | - |
| UL sensory disturbances | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | - | ? | - | - | - | - | - | - | ? |
| UL DTR | $\uparrow$ | N | N | $\downarrow$ | N | N | ? | N | N | $\uparrow$ | $\uparrow$ | N | N | N | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | $\uparrow$ | N | N | N | N | N |
| Urinary sphincter problems | + | + | + | - | + | - | + | + | - | - | + | - | - | + | - | - | - | - | - | - | - | - | - | + |
| Brain and or spine MRI | ? | N | ? | ? | N | N | ? | ? | N | ? | N | TCC | TCC | ? | N | N | N | N | N | ? | ? | ? | N | N | $L L$ lower limb, $U L$ upper limb, $P T R$ patellar tendon reflex, ATR Achilles tendon reflex, $D T R$ deep tendon reflexes, + present, - absent, $N$ normal, $\uparrow$ increased, $\downarrow$ decreased, $L$ left, $R$ right, ? no data, TCC thin corpus callosum

${ }^{\text {a }}$ Not formally assessed in most; based on clinical impression, school performance, and/or current employment
domains of the kinesins are involved in cargo binding or cargo-induced dimerization [19, 20]. KIF1A is almost exclusively expressed in the brain and spinal cord [21] and RNA-sequencing data of human tissues reveals high expression in cerebral cortex, hippocampus, hypothalamus, cerebellum, caudate, and pituitary gland [22]. KIF1A is involved in anterograde transport of vesicles in neuronal axons [23, 24] and it is hypothesized that mild transport impairment may only affect the long axons to the legs and the bladder, resulting in SPG. In contrast, strongly impaired transport will also affect neurons more proximally, leading to more severe motor deficits, and possibly also to involvement of other neuronal populations that lead to mental impairment [5]. The difference between a dominant and recessive inheritance of SPG due to KIF1A variants may likewise be a result of subtle differences in the efficiency of the delivery of cargo to the periphery of the motor neurons.

Another possibility is that there is some redundancy in the expression of different kinesins, such as the kinesin1, $-2,-3,-4,-11$, and -13 family members [25]. In affected neuronal populations, some of these other kinesins may possibly compensate for 'mild' KIF1A variants only. In other brain regions that express KIF1A but are unaffected, even in the severe mental retardation type 9 , such compensation by other kinesins may be sufficient for normal function. These mechanisms, however, are yet speculative and need further molecular assessments.

Patients of two families had learning difficulties, and one had borderline intellectual disability. Possibly, these mild cognitive impairments may be attributed to the KIF1A variants. As such, pure SPG and mental retardation type 9 may be the two extremes of one disease spectrum, where the two families with learning disabilities are intermediates. However, mild cognitive impairment, such as learning difficulties, is relatively common in the general population and was also seen in some family members not having the KIF1A variant. More cases with KIF1A variants and this intermediate phenotype need to be discovered before a causal relationship may be assigned.

## 2q37 microdeletion syndrome/brachydactylymental retardation syndrome

The KIF1A gene is located in the cytogenetic 2 q 37.3 band, and is deleted in a subset of the $2 q 37$ microdeletion syndrome patients. This microdeletion syndrome is characterized by mental retardation (in most), brachydactyly (in $50 \%$ ), short stature, obesity, hypotonia, characteristic facial appearance, autism spectrum disorder (in 30\%), seizures (in 20-35\%), and many other congenital anomalies [26]. Inactivating variants in the HDAC4 gene in patients without the 2 q 37 microdeletion, but with brachydactyly and mental retardation, suggested that the
$H D A C 4$ gene is the cause of these two features in the 2 q 37 microdeletion syndrome [27]. Other genes are likely involved in other characteristics of this syndrome. Spasticity has been reported in decipher in one of the patients (ID 250622) having a $2 q 37$ microdeletion involving KIF1A, but not in others. This may be due to the underdiagnosis of SPG in children with severe other neurological problems, or due to a masking of SPG due to hypotonia, which is another frequent finding in the 2 q 37 microdeletion syndrome. In proband P24, the major problem was SPG, although he was also diagnosed with mild mental retardation and later developed seizures, both likely attributable to haploinsufficiency of HDAC4. Reversed phenotyping in a large set of patients with the 2 q 37 microdeletion syndrome may reveal whether SPG is a commonly overlooked feature, or whether the penetrance of SPG in this syndrome is also incomplete.

## Molecular mechanisms of missense variants in the kinesin motor domain

The anterograde transport of KIF1A motor domain mutants in dominant mental retardation type 9 and, to a lesser extent, in SPG30 was previously found to be impaired, and thereby resulted in decreased localization of KIF1A in distal axons $[5,6]$. Therefore, it is anticipated that cargo is still bound by KIF1A, but delivered with reduced speed and numbers at the neuronal distal axons, and that this mechanism is more pronounced in dominant mental retardation type 9 than in recessive SPG30 [5, 6].

The subcellular localization of the KIF1A mutants in dominant SPG has not been analyzed. Nevertheless, we speculate that the anterograde transport of motor domain mutants in autosomal dominant SPG (17 motor domain variants of 26 total, Fig. 1) is also impaired. The fact that they cause a milder disease than the variants in dominant mental retardation type 9 , suggest that the transport deficit of the dominant SPG mutants is an intermediate between dominant mental retardation type 9 and recessive SPG30, where both copies of the gene are mutated in the latter disorder. The suggestion of a variant-specific 'dose-effect' is corroborated by the fact that there is no overlap between the variants found in dominant SPG and those in dominant mental retardation type 9 or recessive SPG30. More specifically, some variants in dominant SPG (p.(Gly102Ser); p.(Arg350Trp); p.(Arg167His); Fig. 1) affect the same amino acid, but result in a different substitution compared with variants in recessive SPG (p.(Arg350Gly)) [9, 10] or dominant mental retardation type 9 (p.(Gly102Asp); p.(Arg167Cys)) [5]. This proposed relation between variants and their respective allelic disorder indicates that the phenotypic differences are caused by the KIF1A variants themselves, rather than by other genetic or environmental factors.

## Molecular mechanisms of KIF1A loss-of-function variants

Six of the KIF1A variants are likely loss-of-function variants (nonsense, frameshift, and a gene deletion (Chr2q37)) located outside the kinesin motor domain. Two of those had occurred de novo, and three others were present in multiple affected family members. The low occurrence of loss-of-function variants in KIF1A in controls has led to a 'loss-of-function intolerance' (pLI) probability score of 1.00 (on a $0.00-1.00$ scale; gnomAD [13]). Moreover, no loss-of-function variants were detected in our in-house database of $>30,000$ exomes of patients or parents without SPG. Nevertheless, even though the inheritance seems de novo dominant, additional Sanger sequencing of the KIF1A gene in these patients was performed to exclude the possibility of a recessive inheritance. Finally, heterozygous KIF1A knockout mice develop neuropathy at older age, indicating that KIF1A haploinsufficiency leads to a neurological phenotype [28]. Together, these data suggests that the six loss-of-function variants in KIF1A are likely the cause of dominant SPG in these families. In contrast to the variants in the motor domain, loss-of-function variants will not result in an impaired transport of KIF1A itself, but rather a reduced expression of KIF1A in neurons due to nonsense-mediated decay of the variant mRNA, or due to complete absence of transcription of the deleted allele. As a consequence, it is anticipated that decreased levels of KIF1A may result in reduced transport of cargo to the periphery of the neuronal axons, not due to slow transport by motor domain mutants, but because of shortage of KIF1A motors.

## Missense KIF1A variants outside the kinesin motor domain

Four KIF1A variants detected in patients with dominant SPG, three in this report and one described earlier [12], reside outside the motor domain and are more difficult to interpret. These variants are not present in a predicted protein domain, except for p.(Ala460Gly), which may be present in the major coiled-coil domain involved in dimerization of KIF1A [19, 20]. In addition, these variants are substitutions of conserved amino acids and are absent from or extremely rare in large control databases (ExAC and in-house database of $>25,000$ exomes). Unfortunately, segregation analyses were not sufficient for these four missense variants to prove their role in dominant SPG. Identification of these variants in other individuals or functional characterization of the mutant kinesins transport characteristics may determine their role in SPG in the future. In summary, we identified heterozygous causative
variants in the KIF1A gene in $6-7 \%$ of patients with autosomal dominant SPG. These variants were not only missense variants in the motor domain of KIF1A, as seen in allelic disorders, but also loss-of-function variants outside this domain. Haploinsufficiency is thus a likely pathogenic mechanism for at least some of the KIF1A variants in autosomal dominant SPG.

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## Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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