

NIH Public Access

Author Manuscript

Macromolecules. Author manuscript; available in PMC 2008 September 8

Published in final edited form as:

Macromolecules. 2007; 40(9): 2971–2973. doi:10.1021/ma070267j.

Labeling of Polymer Nanostructures for Medical Imaging: Importance of crosslinking extent, spacer length, and charge density

Jinqi Xu^{†,¥}, Guorong Sun^{†,¥}, Raffaella Rossin^{‡,¥}, Aviv Hagooly^{‡,¥}, Zicheng Li[†], Ken-ichi Fukukawa[§], Benjamin W. Messmore[§], Dennis A. Moore[⊥], Michael J. Welch^{†,‡}, Craig J. Hawker[§], and Karen L. Wooley^{*,†,‡}

[†]Center of Materials Innovation and Department of Chemistry, Washington University in Saint Louis, Saint Louis, MO 63130.

‡Mallinckrodt Institute of Radiology, Washington University School of Medicine, Saint Louis, MO 63110.

§Materials Research Laboratory, University of California at Santa Barbara, Santa Barbara, CA 93106.

⊥Mallinckrodt Inc., 675 *McDonnell Blvd.*, *P. O. Box* 5840, *Saint Louis*, *MO* 63134.

Abstract

Radiolabeling studies were employed to investigate the influence of structure on the efficiency of surface functionalization for poly(acrylic acid)-coated shell crosslinked nanoparticles (SCKs) with two types of amine-terminated DOTA chelators. An intricate interplay between the chemical and physical properties of both the DOTA derivative and the SCK nanostructures was revealed, demonstrating the importance of structural control.

Nanoscale architectures,¹ including spheres, cylinders, toroids, vesicles, discs, and other morphologies have attracted much attention. Such nanoobjects with diverse properties have promise for many aspects of nanomedicine,² for instance as carriers to deliver therapeutic agents to specific targets with release of their payloads in a controlled fashion. Great efforts have been devoted to spherical micelles,³ which derive from amphiphilic block copolymers. Above the critical micelle concentration (CMC), micelles are thermodynamically stable in aqueous environments, and are able to encapsulate hydrophobic drug molecules having poor water solubility. Meanwhile, their hydrophilic shell layer stabilizes the entire nanoobject and provides sites for addition of functional units. However, the stability of such nanoscale systems *in vivo* is of concern. To eliminate CMC restrictions, SCKs, a form of covalently-stabilized micelles, have been developed by performing intramicellar crosslinking throughout the shell layer.⁴

The modular nature of SCKs allows their tailoring for specific applications and for the targeted drug delivery to and reporting of biological activities at specific sites. SCK nanoparticles have been further functionalized *via* various methodologies to provide surface-accessible, biologically-active ligands.⁵ The conjugation of various ligands onto pre-established nanoparticles has proved to be a versatile and straightforward method that allows for multiple numbers and types of ligands to be attached onto a well-defined scaffold.⁶ In the case of SCKs, several chemistries have been explored for shell functionalization, including amidation⁷ and click chemistry.⁸ While in most cases the couplings were effective, the complexity of the

To whom correspondence should be addressed: Tel: 314-9357136; Fax: 314-9359844; Email: E-mail: klwooley@artsci.wustl.edu. [¥]These authors contributed equally to this work.

systems makes general statements about coupling difficult. As this entire field of study involving the interactions between synthetic nanomaterials and biological systems relies upon efficient functionalization strategies, it is important to acquire a better understanding of the coupling chemistry and interactions between targeting ligands and polymeric nanoparticles.

Radiolabeling of chelators conjugated onto nanostructures provides materials for *in vivo* imaging and also offers a highly sensitive quantitative measurement of the amount of accessible chelator and, therefore, can be used as a tool to determine the yield of conjugation chemistries. Herein, we report the conjugation of SCK nanoparticles with two types of DOTA (1,4,7,10-tetraazocyclododecane-*N*,*N'*,*N''*,*N'''*-tetraacetic acid) derivatives having different features, DOTAamine and DOTAlysine (Scheme 1). DOTA is a macrocyclic chelating agent widely used to chelate metal ions for diagnostic and therapeutic applications. Among these metals, copper-64 (⁶⁴Cu) has been investigated for its applications in both positron emission tomography (PET) and radiotherapy.⁹ In this study, the ⁶⁴Cu-labeling of DOTA-SCK conjugates is employed to provide new insights into the coupling chemistry, design of ligands and transformation of SCKs into functional materials.

Several SCK samples with varying extents of crosslinking, **SCK1-SCK8**, were prepared from the self assembly of amphiphilic diblock copolymers, poly(acrylic acid)-*b*-polystyrene (PAA-*b*-PS), with varying block lengths from 30 to 136 repeat units, followed by chemical crosslinking throughout the shell layer by reaction with 2,2'-(ethylenedioxy)-bis(ethylamine). These SCK nanoparticles had diameters that were in agreement with the sizes expected based upon the relative hydrophilic:hydrophobic balance and overall polymer chain lengths (Table 1).

Conjugation of each sample from **SCK1** to **SCK8** with DOTAamine molecules was performed *via* amidation chemistry, using *N*-hydroxysulfosuccinimide (sulfo-NHS)¹⁰ in aqueous media (Scheme 1). The 50% crosslinked SCKs, **SCK5** to **SCK8**, were coupled with DOTAlysine, under similar experimental conditions. Consistent with previous observations,⁷ DLS measurements of these conjugates presented similar hydrodynamic diameters to those of their respective SCK precursors. Each DOTAamine-SCK conjugate and DOTAlysine-SCK conjugate was subjected to ⁶⁴Cu radiolabeling tests, under the same experimental protocols and in the same timeframe. The specific activity of each DOTA-SCK conjugate was determined from the percent labeling efficiency, and the number of ⁶⁴Cu-accessible DOTAs per DOTA-SCK was also obtained *via* a modified isotopic dilution method (Table 2).^{7c},¹¹

The radiolabeling results indicated significant differences between the coupling yields of the DOTAamine and DOTAlysine compounds to the SCKs. It is obvious that the coupling efficiency of DOTAlysine with any of **SCK5** to **SCK8** was so low that both the specific activity $(\mu Ci/\mu g)$ and the ⁶⁴Cu-accessible DOTA per DOTA-SCK conjugate were much less than unity. In contrast, attachment of DOTAamine to SCK5-SCK8 gave products that showed high levels of radiolabeling, *i.e.*, 2 to $11 \,\mu\text{C}_i/\mu\text{g}$ of specific activity. Steric hindrance factors were first considered between a SCK nanoparticle having ca. 14-44 nm diameter with the DOTA moiety that is a macrocyclic molecule with a diameter no greater than 1 nm and a short spacer length. The nine atom spacer between the reactive amino end of DOTAamine with its cyclen ring nitrogen atom was longer than that of DOTAlysine, having five atoms (Figure 1), resulting in a distinct enhancement of the coupling by DOTAamine. It has also been found that the coupling efficiency was low (ca. < 5%) between a maleimide derivative bearing an ethylene spacer and SCKs under the same experimental conditions.^{12a} However, the negative zeta potential values of **SCK5** to **SCK8** (Table 1) indicated that the nanoparticle surface presented a high density of negatively-charged carboxylates,^{12b} while DOTAlysine and DOTAamine also contain carboxylates (Figure 1). The addition of more salt to the reaction mixture had little influence on the overall radiolabeling yield, implying that electrostatic repulsions between the SCKs and

Macromolecules. Author manuscript; available in PMC 2008 September 8.

DOTA derivatives exerted minor influences on the coupling. Taken in combination, these results confirm that it is essential that a lengthy, inert spacer exists on the designed functionalities to minimize these steric and electrostatic hindrances, improving the reaction efficiency.

Each DOTAamine-SCK conjugate prepared from 20% crosslinked SCKs exhibited *ca.* 33 to 440% higher specific activity and increased numbers of ⁶⁴Cu-accessible DOTAs per DOTA-SCK conjugate than those observed from the corresponding 50% crosslinked SCKs, derived from the same diblock copolymers (Table 2). For instance, DOTAamine-SCK2 had 45 μ C_i/ μ g, three-fold greater than the 11 μ C_i/ μ g of specific activity observed for DOTAamine-SCK6. Such remarkable improvement can be attributed to the fact that higher numbers of acrylic acid residues were present on the surface of and within the sub-surface of the SCKs with a lower degree of crosslinking, and those carboxylic acids were available for amidation with DOTAamine. Contributions from the larger number of carboxylic acids overcame the potential obstacles from steric factors and electrostatic repulsions (*vide supra*). Furthermore, it has been shown that the permeability of the crosslinked shell domains (membrane-type materials) is higher in SCKs with lower extents of crosslinking.¹³ It is hypothesized that the permeability of the reagents within the shell affects the extent to which the coupling reactions can occur.

The overall structure and composition of the SCKs also affected the coupling chemistry with DOTAamine. The order of specific activity was DOTAamine-SCK2 > DOTAamine-SCK1 > DOTAamine-SCK3 > DOTAamine-SCK4, where for each, the degree of crosslinking was 20% (Table 2). SCK2 to SCK4 contained similar lengths of PAA segments, but incrementally greater lengths of hydrophobic, glassy PS segments, which decreased the hydrophilic:hydrophobic balance and resulted in increased aggregation numbers and particle sizes. SCK2, therefore, contained the greatest proportion of PAA content, followed by SCK3 and then SCK4. The better performance of DOTAamine-SCK2 suggests that the overall composition of the nanoparticles is an important parameter for surface/shell functionalization efficiencies. A similar argument is suggested to explain the differences observed between the radiolabeling of SCK1 and SCK2, which possessed similar lengths for the PS block, but different PAA block lengths.

In summary, we have demonstrated optimized conjugation of DOTA chelators onto SCK nanoparticles in aqueous media. The coupling efficiency, determined by ⁶⁴Cu radiolabeling studies, suggests that the amidation chemistry depends on the nature of the surface and other physical properties of SCK nanoparticles, including charge density, permeability, extents of crosslinking, compositions of the domains within the SCKs and their relative contribution to the overall structure. In addition, the chemical structure and composition of the DOTA chelator is also critically important for efficient reaction. These results can have a great influence on the design and functionalization of nanoparticles, broadening their applications in advanced drug delivery. Currently, the optimized number of ⁶⁴Cu-accessible DOTA chelators per DOTA-SCK conjugate is *ca.* 22, leading to good signal-to-noise ratios in PET imaging. However, there is still a need for improvement to realize exceptionally high radiolabeling yields that will allow application of DOTA-SCKs as highly sensitive *in vivo* PET imaging agents and allow the minimum amount of imaging agent to be used. More efforts are being devoted to further improve the sensitivity and specificity of such conjugates.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgement

This material is based upon work supported by the National Heart Lung and Blood Institute of the National Institutes of Health as a Program of Excellence in Nanotechnology (HL080729). The production of 64 Cu is supported by the National Cancer Institute (M.J.W., CA86307). The authors thank Prof. Shelly E. Sakiyama-Elbert and Ms. Nicole Kohrt for their assistance with zeta potential measurements, and Mr. G. Michael Veith (Washington University Electron Microscopy Laboratory) for assistance with TEM.

References and Notes

- (a) Won Y-Y, Davis HT, Bates FS. Science 1999;283:960–963. [PubMed: 9974383] (b) Discher DE, Eisenberg A. Science 2002;297:967–973. [PubMed: 12169723] (c) Frechet JMJ. J. Polym. Sci., Part A: Polym. Chem 2003;41:3713–3725. (d) Li Z, Kesselman E, Talmon Y, Hillmyer MA, Lodge TP. Science 2004;306:98–101. [PubMed: 15459387] (e) Pochan DJ, Chen Z, Cui H, Hales K, Qi K, Wooley KL. Science 2004;306:94–97. [PubMed: 15459386] (f) Hawker CJ, Wooley KL. Science 2005;309:1200–1205. [PubMed: 16109874]
- (a) Esfand R, Tomalia DA. Drug Discov. Today 2001;6:427–436. [PubMed: 11301287] (b) Kabanov AV, Lemieux P, Vinogradov S, Alakhov V. Adv. Drug Deliv. Rev 2002;54:223–233. [PubMed: 11897147] (c) Davis ME, Brewster ME. Nat. Rev. Drug Discov 2004;3:1023–1035. [PubMed: 15573101] (d) Langer R, Tirrell DA. Nature 2004;428:487–492. [PubMed: 15057821] (e) Torchilin VP. Nat. Rev. Drug Discov 2005;4:145–160. [PubMed: 15688077] (f) Peppas NA, Hilt JZ, Khademhosseini A, Langer R. Adv. Mater 2006;18:1345–1360.
- (a) Allen C, Maysinger D, Eisenberg A. Colloid Surf. B: Biointerfaces 1999;16:3–27. (b) Rösler A, Vandermeulen GWM, Klok H-A. Adv. Drug Deliv. Rev 2001;53:95–108. [PubMed: 11733119] (c) Gaucher G, Dufresne M-H, Sant VP, Kang N, Maysingera D, Leroux J-C. J. Controlled Release 2005;109:169–188. (d) Nishiyama N, Kataoka K. Pharmacol. Ther 2006;112:630–648. [PubMed: 16815554]
- 4. (a) Thurmond KB II, Kowalewski T, Wooley KL. J. Am. Chem. Soc 1996;118:7239–7240. (b) O'Reilly RK, Hawker CJ, Wooley KL. Chem. Soc. Rev 2006;35:1068–1083. [PubMed: 17057836]
- 5. (a) Yasugi K, Nakamura T, Nagasaki Y, Kato M, Kataoka K. Macromolecules 1999;32:8024–8032.
 (b) Bes L, Angot S, Limer A, Haddleton DM. Macromolecules 2003;36:2493–2499. (c) Joralemon MJ, Murthy KS, Remsen EE, Becker ML, Wooley KL. Biomacromolecules 2004;5:903–913.
 [PubMed: 15132680] (d) Qi K, Ma Q, Remsen EE, Clark CG Jr, Wooley KL. J. Am. Chem. Soc 2004;126:6599–6607. [PubMed: 15161288] (e) Joralemon MJ, Smith NL, Holowka D, Baird B, Wooley KL. Bioconjugate Chem 2005;16:1246–1256. (f) Licciardi M, Tang Y, Billingham NC, Armes SP, Lewis AL. Biomacromolecules 2005;6:1085–1096. [PubMed: 15762681]
- 6. (a) Templeton AC, Wuelfing MP, Murray RW. Acc. Chem. Res 2000;33:27–36. [PubMed: 10639073]
 (b) Medintz IL, Uyeda HT, Goldman ER, Mattoussi H. Nat. Mater 2005;4:435–446. [PubMed: 15928695]
 (c) Bull SR, Guler MO, Bras RE, Meade TJ, Stupp SI. Nano Lett 2005;5:1–4. [PubMed: 15792402]
 (d) Frankamp BL, Fischer NO, Hong R, Srivastava S, Rotello VM. Chem. Mater 2006;18:956–959.
- (a) Becker ML, Remsen EE, Pan D, Wooley KL. Bioconjugate Chem 2004;15:699–709. (b) Pan D, Turner JL, Wooley KL. Macromolecules 2004;37:7109–7115. (c) Sun X, Rossin R, Turner JL, Becker ML, Joralemon MJ, Welch MJ, Wooley KL. Biomacromolecules 2005;6:2541–2554. [PubMed: 16153091]
- (a) Joralemon MJ, O'Reilly RK, Hawker CJ, Wooley KL. J. Am. Chem. Soc 2005;127:16892–16899. [PubMed: 16316235] (b) O'Reilly RK, Joralemon MJ, Hawker CJ, Wooley KL. J. Polym. Sci., Part A: Polym. Chem 2006;44:5203–5217.
- (a) Connett JM, Anderson CJ, Guo L-W, Schwarz SW, Zinn KR, Rogers BE, Siegel BA, Philpott GW, Welch MJ. Proc. Natl. Acad. Sci. USA 1996;93:6814–6818. [PubMed: 8692901] (b) McCarthy DW, Bass LA, Cutler PD, Shefer RE, Klinkowstein RE, Herrero P, Lewis JS, Cutler CS, Anderson CJ, Welch MJ. Nucl. Med. Biol 1999;26:351–358. [PubMed: 10382836]
- Lewis MR, Kao JY, Anderson ALJ, Shively JE, Raubitschek A. Bioconjugate Chem 2001;12:320– 324.
- Anderson CJ, Connett JM, Schwarz SW, Rocque PA, Guo LW, Philpott GW, Zinn KR, Meares CF, Welch MJ. J. Nucl. Med 1992;33:1685–1691. [PubMed: 1517844]

Macromolecules. Author manuscript; available in PMC 2008 September 8.

- 12. (a) Sun G, Wooley KL. unpublished results (b) Ma Q, Remsen EE, Kowalewski T, Wooley KL. J. Am. Chem. Soc 2001;123:4627–4628. [PubMed: 11457260]
- 13. Murthy SK, Ma Q, Clark CG Jr, Remsen EE, Wooley KL. Chem. Commun 2001:773-774.

Xu et al.



Figure 1.

Structural comparisons of DOTAlysine with DOTAamine and sulfo-NHS activated SCK nanoparticles in PBS buffer (pH 7.5).

Xu et al.



Scheme 1.

Conjugation chemistry of SCK nanoparticles with DOTAamine or DOTAlysine molecules in aqueous media.

OH

Xu et al.

NIH-PA Author Manuscript

NIH-PA Author Manuscript

NIH-PA Author Manuscript

^cComposition of amphiphilic diblock copolymer employed for micelle formation; polymer precursor numbers were defined in Supporting Information.

 d Number-average hydrodynamic diameters measured in nanopure H2O.

Macromolecules. Author manuscript; available in PMC 2008 September 8.

 $^{e}D_{\mathrm{av}}$ were measured for the SCK core.

 $f_{\rm Average}$ heights measured by tapping-mode AFM.

 8 Each aggregation number was calculated based on the particle core diameter from TEM and the contents of PS.

 $h_{\rm Z}$ zeta potential values measured in 5 mM PBS buffer (pH 7.4).

Table 2

⁶⁴Cu radiolabeling results of DOTA-SCK conjugates^a

Functionalized Particles	Specific activity $(\mu C_i \mu g^{-1})$	⁶⁴ Cu-accessible DOTA per DOTA-SCK
DOTAamine-SCK1	32	22
DOTAamine-SCK2	45	14
DOTAamine-SCK3	14	11
DOTAamine-SCK4	2	3
DOTAamine-SCK5	6	2
DOTAamine-SCK6	11	2
DOTAamine-SCK7	3	< 1
DOTAamine-SCK8	2	< 1

 a Specific activities (μ C₁ μ g⁻¹) and 64 Cu-accessible DOTA per SCK of DOTAlysine-SCK5 to DOTAlysine-SCK8 conjugates were much less than unity.

Page 9