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Lack of BAP1 protein expression in uveal melanoma is associated with increased metastatic risk and has utility in routine prognostic testing

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Background: The absence of BRCA1-associated protein 1 (BAP1) expression in uveal melanoma (UM) is associated with metastatic progression and reduced survival. In this study, we examine nuclear BAP1 (nBAP1) protein expression in primary UMs (PUMs) that show both 'typical' and 'atypical' clinical courses according to their chromosome 3 status, and secondary hepatic metastatic UM (MUM), correlating the results with histological, clinical and survival data.

Methods: Nuclear BAP1 expression was immunohistochemically assessed in tissue microarrays (TMAs) of: (a) 68 PUM patients, who had been treated surgically; and (b) 13 MUM patients, with 5 cases being paired with primary tumour tissue. All cases were fully annotated. The percentage of tumour cell nuclei staining positively for BAP1 was scored by independent observers.

Results: Nuclear BAP1 protein expression was absent in 35 out of 68 (51%) PUM patients, correlating strongly with poor prognostic clinicopathological and genetic parameters and reduced survival (Log rank, $P < 0.001$). Lack of nBAP1 expression importantly identified a subset of 'atypical' PUM patients with disomy of chromosome 3 but with unexpected metastatic relapse. Nuclear BAP1 expression was absent in 10 out of 13 (77%) MUM and expression was concordant in all paired PUM and MUM patients.

Conclusions: Absent nBAP1 protein expression is an independent survival predictor for UM patients, easily examined using immunohistochemistry.

Approximately 50% of patients with uveal melanoma (UM) develop metastatic disease, which usually involves the liver. Metastatic disease is rarely detectable at the time of treatment of the primary UM (PUM), developing varyingly months to years after diagnosis.

The development of metastatic disease correlates strongly with chromosomal abnormalities in UM. We and others have shown that disease-specific mortality is intimately linked with loss of one copy of chromosome 3 and gains of chromosome 8q (Prescher *et al*, 1996; White *et al*, 1998; Mensink *et al*, 2008; Damato *et al*, 2010; Abdel-Rahman *et al*, 2011a; Thomas *et al*, 2012). Attempts to

identify a minimal region of loss on chromosome 3, contributing to metastasis, have yielded differing results (Blasi *et al*, 1999; Tschentscher *et al*, 2001; Parrella *et al*, 2003; Zeschmick *et al*, 2003; Trolet *et al*, 2009). Mutually exclusive activating mutations in *GNAQ* and *GNA11*, located on chromosomes 9q and 19p, respectively, occur in most UM but are not associated with metastatic disease (Van Raamsdonk *et al*, 2009, 2010). In 2010, Harbour and co-workers identified inactivating mutations in the *BAP1* gene located on chromosome 3p21.1, which occurred almost exclusively in metastasising 'class 2' UM with monosomy 3 (Harbour *et al*, 2010). This finding, together with data showing

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frequent loss of heterozygosity for the *BAP1* gene in human tumours, demonstrates the importance of *BAP1* as a tumour-suppressor gene (Jensen *et al*, 1998; Bott *et al*, 2011; Testa *et al*, 2011; Wiesner *et al*, 2011; Abdel-Rahman *et al*, 2011b).

The *BAP1* gene encodes a nuclear protein belonging to a subfamily of deubiquitinating enzymes called ubiquitin C-terminal hydrolases, which was initially discovered through its interaction with the tumour-suppressor protein BRCA1 (Jensen *et al*, 1998). BRCA1-associated protein 1 (BAP1) protein is reported to be involved in a variety of cellular processes, including cell cycle regulation, DNA repair and protein trafficking. A loss of or mis-localised protein expression may result in severe deregulation of these processes contributing to tumour development and/or progression (Machida *et al*, 2009; Misaghi *et al*, 2009; Yu *et al*, 2010). In PUM, an absence of BAP1 protein was recently shown to be strongly correlated with *BAP1* gene mutations, and was significantly associated with metastatic progression and reduced survival in these patients (Koopmans *et al*, 2014). Furthermore, the two recent studies examining BAP1 protein expression in PUM have advocated the use of immunohistochemistry (IHC) for its testing in the routine work-up of UM cases where possible (Shah *et al*, 2013; Koopmans *et al*, 2014).

The purpose of this study was to examine BAP1 protein expression in 'typical' and 'atypical' cases of PUM, previously classified as such by chromosome 3 status and survival time, and in metastatic liver lesions from UM patients, and also to correlate these data with histological, clinical and survival information. To achieve this, two tissue microarrays (TMAs) of clinically, histomorphologically and genetically well-defined primary and metastatic UM (PUM-TMA and MUM-TMA, respectively) were used.

MATERIALS AND METHODS

UM specimens and TMA construction. Seventy histologically confirmed PUM treated by primary enucleation or local resection between 1996 and 2007 at the Liverpool Ocular Oncology Centre were used to construct PUM-TMA using the Manual Tissue Arrayer 1 (Beecher Instruments, Sun Prairie, WI, USA). The UM samples on PUM-TMA represented 'typical' (1 and 2) and 'atypical' (3 and 4) subgroups of patients originally selected on the basis of their chromosome 3 status and survival time as follows:

- (1) Monosomy 3 UM causing metastatic death within 6 years (yr) of diagnosis ($M3 \leq 6yr$; $n = 21$);
- (2) Disomy 3 UM with survival exceeding 6 years ($D3 > 6yr$; $n = 13$);
- (3) Monosomy 3 UM with survival exceeding 6 years ($M3 > 6yr$; $n = 17$); and
- (4) Disomy 3 UM with metastatic death within 6 years of diagnosis ($D3 \leq 6yr$; $n = 15$; Lake *et al*, 2013).

The typical UM subgroups account for >90% of all UMs (Damato *et al*, 2007). Two UM with unknown chromosome 3 status were also present on PUM-TMA and were included in the analyses. Triplicate 0.6 mm cores from each tumour were randomly distributed across the TMA alongside non-tumour reference tissues, which were obtained from tonsil, duodenum and pancreas. Whenever possible, the tumour cores were taken from different regions of the UM. Histological, clinical and survival data were available for all patients. Chromosome 3 copy number had previously been determined by fluorescence *in-situ* hybridisation as described by Damato *et al* (2007). In the case of subgroup 4 ($D3 \leq 6yr$), the disomy 3 status was also confirmed by Multiplex Ligation-Dependent Probe Amplification, as previously described (Lake *et al*, 2013).

MUM-TMA contained duplicate or triplicate 0.6 mm cores from the liver metastases of 17 UM patients, who had not received any other systemic treatment, along with tonsil and liver as non-tumour reference tissues. Seven of these cases were matched with primary tumour material on PUM-TMA.

All patients were treated in accordance with the Declaration of Helsinki. Ethical approval from the National Research Ethics Service had been provided for all work included in this study (NRES study numbers: 10/H1015/56 and 11/NW/0759).

Immunohistochemistry. Immunohistochemistry was performed on 4 μ m sections cut from the formalin-fixed, paraffin wax-embedded TMA tissue blocks. Antigen retrieval and IHC were performed using the Dako PT Link and Autostainer Plus systems according to the standard manufacturers' procedures (Dako UK Ltd, Cambridgeshire, UK). A mouse anti-human BAP1 antibody was used at a previously optimised concentration of 1 μ g ml⁻¹ (C-4, Santa Cruz, Insight Biotechnology Ltd, Middlesex, UK). The sections were then counterstained with haematoxylin, dehydrated, cleared and mounted. Positive nuclear BAP1 (nBAP1) staining in pancreas and/or tonsil cores in PUM-TMA and MUM-TMA demonstrated a 'valid' IHC run. Additional whole tonsil and pancreas sections were treated with a mouse IgG1 isotype control (Dako) at the same concentration as the BAP1 primary antibody and served as negative controls. Neither nuclear nor cytoplasmic staining was observed in mouse IgG1-treated sections.

Scoring. Stained TMA slides were evaluated for nBAP1 protein expression by three independent observers (SEC, HK and SF) with no prior knowledge of the chromosome 3 status of the individual cases. Specimens were given a final score for the percentage of tumour nuclei positive for BAP1 protein expression based on data across the TMA cores. The presence or absence of cytoplasmic BAP1 (cBAP1) staining was also recorded.

Statistical analyses. All data were processed in SPSS (ver.20.0; SPSS Science, Chicago, IL, USA) for statistical analyses. Correlation of BAP1 expression with known risk factors of UM metastatic progression was made using Chi-Square for categorical variables and either Student's *t*-test or Mann-Whitney for linear variables. Multivariate analyses were performed using Cox Regression and backward likelihood ratio analysis. Survival analyses were performed with Kaplan-Meier.

RESULTS

TMA assessment. Of the 70 PUMs included in PUM-TMA, 63 had three tumour cores that were assessable and 5 had only two assessable tumour cores, which was considered the minimum requirement for scoring. Two tumours in PUM-TMA were lost due to missing or inadequate cores following sectioning.

For the 17 liver metastases arising from a PUM included on MUM-TMA, three tumour cores were assessable in 11 cases and two tumour cores were assessable in 2 cases. Four tumours on MUM-TMA were lost due to missing or inadequate cores following sectioning. Of the 13 assessable cases, 5 were matched with a PUM on PUM-TMA.

Clinicopathological data. PUM-TMA comprised 24 males and 44 females, with a mean age at diagnosis of 60 years (median, 63 years; range, 21–89 years; Supplementary Table 1). The tumours had a mean largest basal diameter of 16.5 mm (median, 17.0 mm; range, 10.8–21.5 mm). Largest basal diameter measurements were not significantly different between the four tumour subgroups included on PUM-TMA ($P = 0.09$; one-way ANOVA), suggesting that lead time bias had not influenced the overall survival time. Histological examination previously performed on full tumour sections as part

of the routine diagnostic work-up, classified 21 UMs as predominantly spindle and 47 UMs of mixed cell type, with variable proportions of epithelioid cells. Periodic acid Schiff+ closed connective tissue loops were found in 39 UMs, and the mitotic count exceeded 5 per 40 high-power fields (HPF) in 40 tumours. Genetic analysis of all UMs examined classified 28 of these tumours as D3 and 38 as M3. In two patients, the chromosome 3 status was not known. Follow-up information was available for all patients at the close of study, 31 December 2013: 22 patients were still alive; 43 had died of metastatic melanoma; and 3 had died of other causes (i.e., ischemic heart disease, myocardial infarction and chronic obstructive pulmonary disease). After excluding these three patients, the mean survival time was 6.2 years (median 5.1 years; range 0.5–14.4 years).

Of the 13 hepatic metastases assessed on MUM-TMA, 12 were from UM patients who had undergone surgical resection, and one from a patient who had undergone staging laparoscopy and then found to have non-resectable disease. Information was not available regarding the number and size of the hepatic metastases; however, resection was incomplete following pathological examination in two cases (MUM1 and MUM4). Pathological examination of the tumour cores in MUM-TMA reported epithelioid cells to be present in nine cases. Clinical information regarding primary management of the intraocular UM was available in all cases. Details of primary tumour size, ciliary body involvement and chromosome 3 status were available in the majority of cases (Supplementary Table 2).

Expression of BAP1

PUM. BAP1 protein expression was localised predominantly to the nuclei of tumour cells in 33 out of 68 (49%) cases of PUM (Figures 1A–C). Weak to moderate cytoplasmic expression was also observed in 30 out of 68 (44%) of the UM analysed, and occurred predominantly in cases for which nBAP1 was also present. For all cases in PUM-TMA, the percentage of nBAP1-positive tumour cells in the individual cores of each UM did not differ by more than 10%, indicating no significant heterogeneity in the presence or absence of nBAP1 protein expression across any individual tumour. The percentage of nBAP1+ tumour cell nuclei was: $\geq 80\%$ across the cores in 32 cases, and was completely absent in all cores for 35 cases. A single case showed staining in 70% of tumour nuclei. For all further statistical analyses, nBAP1 protein expression was considered as either present or completely absent.

MUM. nBAP1 protein was absent in all cores of 10 out of 13 (77%) cases (Figures 1D and E). In the remaining three cases, nBAP1 staining was positive in MUM cell nuclei in every tissue core. The five pairs of matched primary and secondary MUMs showed complete concordance between the nBAP1 score in the primary tumour and the hepatic metastasis; nBAP1 was absent in four of these cases and present in one case.

Correlation of nBAP1 protein expression with clinicopathological and genetic features of metastatic risk

PUM. Univariate analysis of all cases demonstrated that lack of nBAP1 protein expression was strongly associated with clinicopathological and genetic features of increased metastatic risk, namely: increased age at primary management ($P=0.002$); ciliary body involvement ($P=0.001$); presence of periodic acid Schiff+ closed connective tissue loops ($P=0.001$); presence of epithelioid cells ($P=0.011$) and M3 ($P=0.001$; Table 1).

Multivariate analysis using the Cox regression model was performed on all UM cases ('typical' and 'atypical' subgroups, excluding only those who had died of causes other than metastatic melanoma) to determine independent risk factors associated with survival. Lack of nBAP1 protein ($P=0.002$) and an increased mitotic rate ($P=0.046$) remained in the Cox regression model as significant independent predictors of reduced survival time. Kaplan–Meier analysis further confirmed the significant association between a lack of nBAP1 protein and reduced survival time (Figure 2; Log Rank, $P<0.001$).

When only the D3 UM were considered, the atypical D3 ≤ 6 yr cases (subgroup 4) were more likely than D3 > 6 yr cases (subgroup 2) to have: ciliary body involvement ($P=0.009$); epithelioid cells present ($P=0.007$); lack of nBAP1 protein ($P=0.001$) and mitotic count $\geq 5/40$ HPF ($P=0.002$; Table 2). Nuclear BAP1 protein expression was absent in 8 out of 15 (53%) D3 ≤ 6 yr PUM; however, it was detected in all 13 D3 > 6 yr PUMs (Table 2). Multivariate Cox regression analysis of all D3 cases demonstrated the presence of epithelioid cells ($P=0.019$) and a high mitotic count ($P=0.001$) as significant independent predictors of reduced survival time (Table 3).

When only the M3 UM were considered, M3 ≤ 6 yr PUMs (subgroup 1) were less likely to have nBAP1 protein positivity ($P=0.003$) and more likely to have closed connective tissue loops ($P=0.003$) and a largest basal diameter ≥ 16 mm ($P=0.029$) when compared with M3 > 6 yr (subgroup 3) PUM (Table 2). Nuclear BAP1 protein was absent in 18 out of 21 (86%) M3 ≤ 6 yr

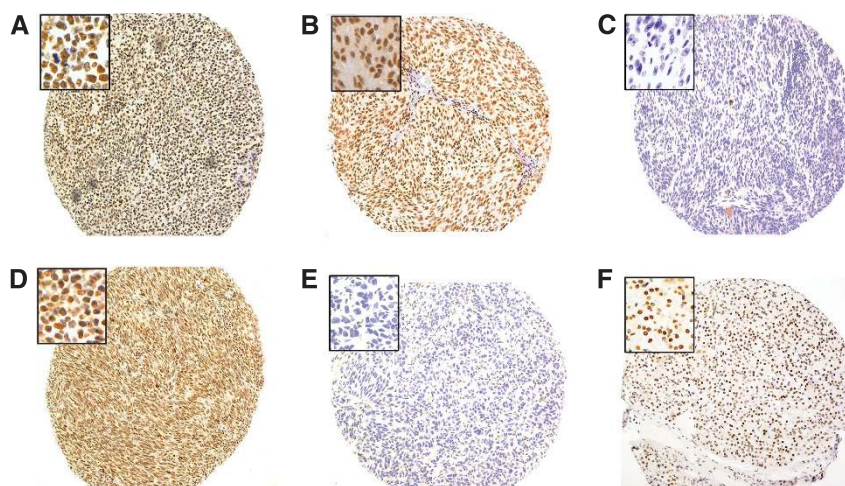


Figure 1. Representative images of BAP1 protein expression in primary tumours (A–C) and metastatic liver lesions (D and E) from patients with UM ($\times 10$ magnification). Insets show nBAP1 at $\times 40$ magnification. Images A, B and D show strong nBAP1 protein expression in 100% of the tumour cells together with weak (A) or moderate cytoplasmic staining (B and D). Images C and E show no detectable nBAP1 protein. Image F shows nBAP1 protein expression in internal positive control pancreatic tissue.

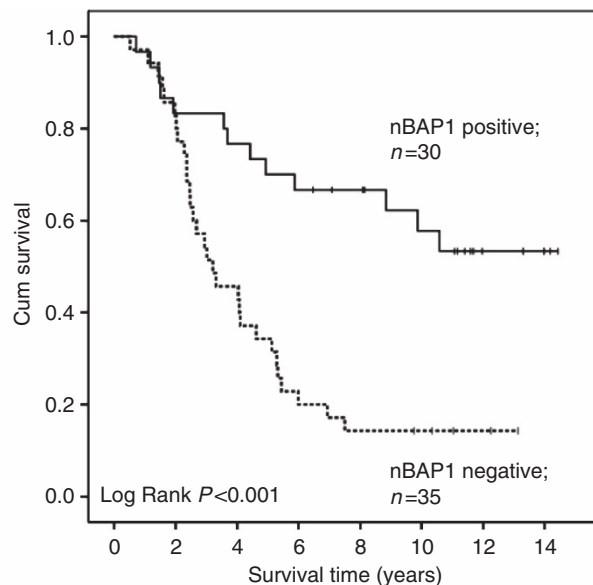
Table 1. Association of nBAP1 status with clinical, pathological and genetic risk factors in UM

Variable	nBAP1 absent	nBAP1 present	P-value
Age at PM (years)			
Mean	65.4	55.2	0.002
Median	65.0	57.0	
Range	38.0–89.0	21.0–79.0	
Gender			
Female	22 (50%)	22 (50%)	0.747
Male	13 (54%)	11 (46%)	
LBD (mm)			
Mean	16.9	16.0	0.151
Median	17.5	15.9	
Range	10.8–21.1	12.2–21.5	
UH (mm)			
Mean	9.6	8.6	0.157
Median	9.7	9.0	
Range	3.4–15.7	2.4–14.0	
Ciliary body involvement			
Yes	17 (81%)	4 (19%)	0.001
No	18 (38%)	29 (62%)	
Epithelioid cells present			
Yes	29 (62%)	18 (32%)	0.011
No	6 (29%)	15 (71%)	
Closed loops present			
Yes	27 (69%)	12 (31%)	0.001
No	8 (28%)	21 (72%)	
Mitotic count per 40 HPF			
Mean	8.3	6.7	0.235
Median	7.0	5.0	
Range	1.0–23.0	1.0–16.0	
Monosomy 3			
Yes	25 (66%)	13 (34%)	0.001
No	8 (29%)	20 (71%)	
Not available	2 (100%)	0	

Abbreviations: HPF = high-power fields; LBD = largest basal diameter; PM = primary management; nBAP1 = nuclear BRCA1-associated protein 1; UH = ultrasound height.

and 7 out of 17 (41%) M3 > 6yr PUMs (Table 2). Multivariate Cox regression analysis of all M3 cases highlighted an absence of nBAP1 as the only significant independent predictor of reduced survival time in this group ($P = 0.05$).

MUM. Of the 13 patients with assessable cores in MUM-TMA, all 10 with undetectable nBAP1 protein expression had died at the close of study. The mean survival time of this cohort of patients was 6.5 years (median, 4.2 years; range, 1.9–19.9 years). Of particular note was that three patients with nBAP1 protein staining in 100% of UM cells in the hepatic metastasis were alive at the close of study (MUM3, MUM12, MUM13; Supplementary Table 2). The time period from primary management to the development of metastatic disease tended to be longer in these individuals



BAP1SI	Total No.	No. of events	Censored		Mean survival time (years)	95% confidence interval	
			No.	%		Upper	Lower
Negative	35	30	5	14.3	4.74	3.49	6.00
Positive	30	13	17	56.7	9.97	8.05	11.90
Overall	65	43	22	33.8	7.24	5.93	8.55

Figure 2. Kaplan–Meier survival curve and table for all primary UM stratified according to nBAP1 protein expression. UM patients who had died of causes other than metastatic melanoma were excluded in the analyses. BAP1SI indicates whether the nBAP1 protein expression was scored as positive or negative. No. of events indicates the number of deaths.

(7.9, 11.4 and 6.2 years, respectively), compared with patients who had hepatic metastases in which nBAP1 protein was absent.

DISCUSSION

In this study, we found a significant independent association between lack of nBAP1 protein expression in PUM and reduced survival time, irrespective of chromosome 3 status. Moreover, we report the results of nBAP1 protein expression in MUM to the liver, demonstrating not only complete concordance between matched PUM and metastatic liver lesions, but also that the presence of nBAP1 positivity tended to be associated with increased metastatic latency in M3 tumours.

Low or absent nBAP1 protein expression is associated with a poor prognosis in non-small-cell lung carcinoma (NSCLC) and clear cell renal carcinomas with data demonstrating a shorter median survival time and increased metastatic spread to lymph nodes in NSCLC (Fan *et al*, 2012; Pena-Llopis *et al*, 2012). In UM, two recent studies similarly demonstrated that loss of BAP1 protein, examined in 30% (Koopmans *et al*, 2014) and 58% (Shah *et al*, 2013) of UM cases, is strongly associated with an aggressive metastatic phenotype and poor prognosis. Consistent with these reports, we observed absent nBAP1 protein in 51% of the PUM cases, and also demonstrated its significance as an independent predictor of reduced overall survival.

Unlike the two studies recently reported for UM, our cohort contained UM samples with both ‘typical’ (i.e., M3 UM with short survival (subgroup 1) and D3 UM with prolonged survival (subgroup 2)) and ‘atypical’ (i.e., M3 UM with prolonged survival (subgroup 3) and D3 UM with short survival (subgroup 4)) clinical

Table 2. Associations of the chromosome 3 subgroups with clinical and pathological variables

Variable	Subgroup 1 (M3 ≤ 6yr)	Subgroup 2 (D3 > 6yr)	Subgroup 3 (M3 > 6yr)	Subgroup 4 (D3 ≤ 6yr)	P-value*	P-value
Age at PM (yr)						
Mean	66.3	51.3	62.4	58.3	0.450	0.277
Gender						
Female	15	11	8	10	0.291	0.133
Male	6	2	9	5		
LBD						
< 16 mm	4	8	9	4	0.067	0.029
≥ 16 mm	17	5	8	11		
CBI						
Yes	10	0	4	6	0.009	0.133
No	11	13	13	9		
Epithelioid cells						
Yes	17	5	11	13	0.007	0.270
No	4	8	6	2		
Closed loops						
Yes	18	5	7	8	0.450	0.003
No	3	8	10	7		
Mitotic count						
< 5/40_HPF	6	10	6	3	0.002	0.668
≥ 5/40_HPF	15	3	11	12		
nBAP1						
Absent	18	0	7	8	0.001	0.003
Present	3	13	10	7		
Abbreviations: CBI = ciliary body involvement; HPF = high-power fields; LBD = largest basal diameter; PM = primary management; P-value* = subgroup 2 vs subgroup 4; P-value = subgroup 1 vs subgroup 3; yr, year.						

Table 3. Multivariate Cox regression of survival in all patients classified as D3

Variable	Hazard ratio	95% CI	P-value
Epithelioid cells	6.605	1.371–31.819	0.019
Mitotic count ≥ 5/40 HPF	10.246	2.526–41.560	0.001
'Closed' loops	—	—	—
LBD ≥ 16 mm	—	—	—
Ciliary body involvement	—	—	—
Absent nBAP1	—	—	—
Abbreviations: CI = confidence interval; HPF = high-power fields; LBD = largest basal diameter; nBAP1 = nuclear BRCA1-associated protein 1. '—' Indicates that variable did not remain in the Cox regression model.			

courses. The 'atypical' subgroups, based on chromosome 3 status and survival time following diagnosis of a PUM, represent approximately 10% of all UM (Damato *et al*, 2007, 2010): although rare, it is essential to examine these cases to fully understand the molecular pathogenesis of UM. Of particular, clinical relevance are those UM cases with D3 and 'unexpected' development of

metastatic disease (subgroup 4). Interestingly, approximately 50% of subgroup 4 patients completely lacked nBAP1 protein staining. It is possible that loss of heterozygosity of a small region containing the *BAP1* gene coupled with a mutation on the remaining *BAP1* allele in these individuals is responsible for this phenomenon, as was reported recently for three UMs analysed by SNP array and *BAP1* sequencing (Koopmans *et al*, 2014). We have previously published, however, the SNP results for these cases, which did not identify the *BAP1* locus as a common region of deletion (Lake *et al*, 2010). Similar UM cases that suggest alternative mechanisms influencing the metastatic process have been reported. Koopmans *et al* (2014) show 17% (3 out of 18) of monosomy 3 cases with short survival, for which *BAP1* mutations are absent and BAP1 protein is detected. In another study, three cases without metastases (disease-free survival of 32, 40 and 81 months) despite monosomy 3 and *BAP1* mutation are described (Dono *et al*, 2014). Newly identified driver mutations in splicing factor 3b subunit 1 (*SF3B1*) and eukaryotic translation initiation factor 1A, X-linked (*EIF1AX*) have been described in about 20–30% and 48% of UM patients with disomy 3, respectively, and are associated with a favourable prognosis (Furney *et al*, 2013; Martin *et al*, 2013). Although it is not yet clear how these mutations promote UM, it is of interest that *BAP1*, *SF3B1* and *EIF1AX* mutations occur in a largely mutually exclusive manner. Moreover, although the majority of *SF3B1* mutations detected result in a missense change

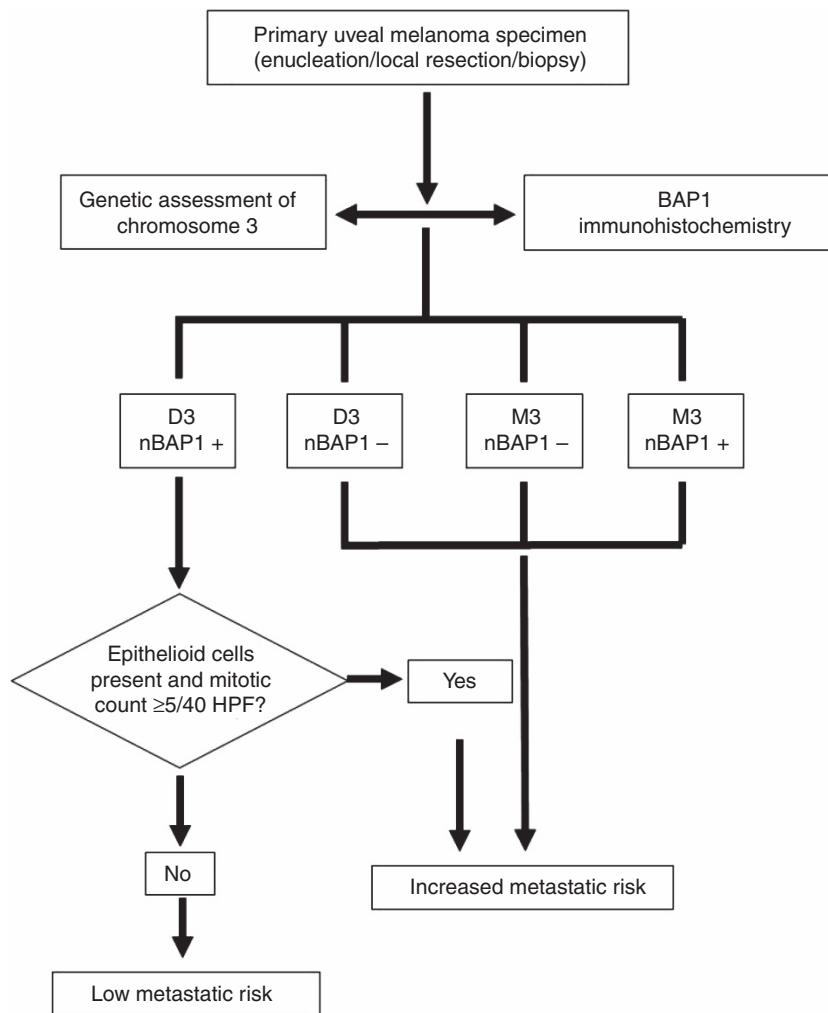


Figure 3. Proposed prognostic test workflow and risk decision outcome for primary UM specimens, based on genetic analysis of chromosome 3 and BAP1 immunohistochemistry.

at Arg625, distinct *SF3B1* mutations in 30% (3 out of 10) of disomy 3 patients who developed metastatic disease were also reported (Martin *et al*, 2013). Further subgrouping of the UM according to *SF3B1* and *EIF1AX* mutations may enhance prognostication for these patients and also be incorporated into the prognostic workflow proposed in Figure 3.

Nuclear BAP1 protein was absent in the majority of ‘typical’ M3 UMs with short survival (subgroup 1) and tended to be associated with a reduced overall survival time across all M3 cases examined (40 months) compared with tumours with positive nBAP1 staining (143 months). These data are consistent with those of Koopmans *et al* (2014) for both patient survival based on the *BAP1* mutational status (32 vs 133 months for *BAP1* mutation-positive vs *BAP1* mutation-negative tumours) or BAP1 protein expression (31 vs 133 months for tumours showing negative BAP1 expression by IHC vs those with positive BAP1 expression) and thus the proposed role of BAP1 in the pathogenesis of UM with an aggressive phenotype. Nonetheless, three patients within subgroup 1 displayed nBAP1 staining and yet died within 5 years of primary management of their UM. Clinically, at the Liverpool Ocular Oncology Centre, all patients with M3 are classified as ‘high risk’ with respect to the development of metastases, and as such would be followed-up with regular screening for metastatic disease irrespective of BAP1 protein expression (Marshall *et al*, 2013).

Previous studies have reported the presence of *BAP1* mutations in 84% of poor prognosis class 2 UMs (Harbour *et al*, 2010).

More recently, *BAP1* mutations were identified in 58% (11 out of 19) of UM patients who had developed metastasis, and loss of BAP1 immunohistochemical staining was observed in the tumour cells of all but 2 of those 11 cases (Koopmans *et al*, 2014). We were unable to conduct mutational analysis of the *BAP1* gene in this cohort of patient samples because of insufficient material remaining for this study; however, there is mounting evidence in the literature that lack of BAP1 protein expression is strongly associated with the presence of inactivating *BAP1* mutations in UM (Koopmans *et al*, 2014), mesothelioma (Yoshikawa *et al*, 2012) and Spitz melanomas (Wiesner *et al*, 2011, 2012). Moreover, work conducted in a human NSCLC cell line in nude mice, implies that both de-ubiquinating activity and nuclear localisation are necessary for the tumour-suppressive activity of BAP1 (Ventii *et al*, 2008). Our finding that BAP1 protein (both nuclear and cytoplasmic) was absent in 70% of patients who developed metastatic disease supports this concept. Our data demonstrate that such UM cases can be easily identified at primary management using BAP1 IHC, in particular to enhance the decision regarding the frequency of liver screening in a subset of ‘atypical’ disomy 3 cases, as highlighted in the proposed prognostic workflow shown in Figure 3. It is important to acknowledge, however, that in a human NSCLC cell line, missense mutations introduced into the catalytic domain of *BAP1* reduced its tumour-suppressor function, but BAP1 protein expression was unaltered (Ventii *et al*, 2008). Furthermore, in two UM samples with loss of one copy of

chromosome 3 and a *BAP1* mutation, BAP1 protein could be detected by IHC (Koopmans *et al*, 2014). Ongoing, prospective analysis of BAP1 protein expression, assessed in all UMs during routine pathological work-up, together with other clinicopathological and genetic features is therefore necessary.

The inclusion of MUM tissue in this study allowed us to demonstrate that differences in BAP1 protein expression occurring in the PUM are retained in the hepatic metastasis. Interestingly, we observed two cases of MUM that displayed positive nBAP1 staining, which is consistent with a recent report detecting BAP1 protein in 3 out of 16 metastatic lesions (Griewank *et al*, 2014). This is of key importance not only to enhance our understanding of the biology of these tumours but also when considering therapeutic intervention, which at present is likely to be determined based on the results of tumour sampling from the PUM. It should be recognised, however, that the MUM is likely to have undergone additional genetic alterations (Trolet *et al*, 2009) that could further influence the therapeutic response, as seen in other tumours (Swanton, 2012).

Although the loss of BAP1 function may be linked to metastasis, the precise mechanism of tumour dissemination in UM is still unclear. In functional studies, depletion of BAP1 protein in the 92.1 UM cell line using siRNAs led to a change in cell morphology from spindle to a more epithelioid phenotype, loss of melanocytic differentiation and transition from a 'class 1' to a 'class 2' gene expression profile (Harbour *et al*, 2010). BAP1 is also reported to interact with host cell factor-1 resulting in the regulation of cell cycle progression at the G1/S checkpoint by controlling the transcription of genes regulating cell growth and proliferation (Machida *et al*, 2009; Misaghi *et al*, 2009; Yu *et al*, 2010). Although the *in vitro* morphological changes are consistent with the significant association between absence of nBAP1 protein and the presence of epithelioid cells reported for PUM in this study, we observed no correlation between nBAP1 protein expression and changes in the mitotic count. Evidence linking BAP1 with differentiation, however, has importantly identified histone deacetylase inhibitors as therapeutic compounds that could revert UM cells with BAP1 loss to a more differentiated phenotype (Landreville *et al*, 2012). Further studies are essential to understand how these therapeutic strategies may target specific pathways in MUM.

In conclusion, our data demonstrate that the absence of nBAP1 protein expression is strongly associated with a poor outcome in patients with UM and that this can be easily (and cost efficiently) detected using IHC. Moreover, our finding that lack of BAP1 protein can identify a subgroup of 'atypical' poor prognosis D3 patients warrants the introduction of this test into the routine pathological work-up of all UM specimens. Ongoing collection and further detailed molecular analyses of all 'atypical' UM are required to fully understand the aetiology of this aggressive disease.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

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