

Large-scale sequestration of atmospheric carbon via plant roots in natural and agricultural ecosystems: why and how

Douglas B. Kell

Phil. Trans. R. Soc. B 2012 **367**, 1589-1597
doi: 10.1098/rstb.2011.0244

References

This article cites 160 articles, 30 of which can be accessed free
<http://rstb.royalsocietypublishing.org/content/367/1595/1589.full.html#ref-list-1>

EXiS Open Choice

This article is free to access

Subject collections

Articles on similar topics can be found in the following collections

[bioinformatics](#) (41 articles)
[biotechnology](#) (9 articles)
[genomics](#) (17 articles)
[molecular biology](#) (112 articles)
[plant science](#) (23 articles)
[systems biology](#) (57 articles)

Email alerting service

Receive free email alerts when new articles cite this article - sign up in the box at the top right-hand corner of the article or click [here](#)

Opinion piece

Large-scale sequestration of atmospheric carbon via plant roots in natural and agricultural ecosystems: why and how

Douglas B. Kell^{1,2,*}

¹*School of Chemistry and Manchester Interdisciplinary Biocentre, University of Manchester, 131 Princess St, Manchester M1 7DN, UK*

²*Biotechnology and Biological Sciences Research Council, Polaris House, North Star Avenue, Swindon, Wiltshire SN2 1UH, UK*

The soil holds twice as much carbon as does the atmosphere, and most soil carbon is derived from recent photosynthesis that takes carbon into root structures and further into below-ground storage via exudates therefrom. Nonetheless, many natural and most agricultural crops have roots that extend only to about 1 m below ground. What determines the lifetime of below-ground C in various forms is not well understood, and understanding these processes is therefore key to optimising them for enhanced C sequestration. Most soils (and especially subsoils) are very far from being saturated with organic carbon, and calculations show that the amounts of C that might further be sequestered (<http://dbkgroup.org/carbonsequestration/rootssystem.html>) are actually very great. Breeding crops with desirable below-ground C sequestration traits, and exploiting attendant agronomic practices optimised for individual species in their relevant environments, are therefore important goals. These bring additional benefits related to improvements in soil structure and in the usage of other nutrients and water.

Keywords: soil; carbon; sequestration; systems biology; breeding

1. INTRODUCTION: WHY SEQUESTER ATMOSPHERIC CO₂?

It is well known that atmospheric CO₂ is increasing at some 2 ppmv p.a., mainly as a result of human activities such as fossil fuel combustion, and that this has taken its values from *ca* 280 ppmv at ‘pre-industrial’ levels to more like 390 ppmv today [1]. To avoid the predicted increases in global temperature contingent upon ‘greenhouse gas’ effects we need not only to lower the emissions but preferably to find means of sequestering atmospheric CO₂ over extended periods. Similar arguments apply to all other greenhouse gases [2], such as CH₄ and N₂O.

On geological time-scales, atmospheric CO₂ levels were much (possibly 10-fold) greater than they are now [3], and the main means by which sequestration of atmospheric carbon was achieved, especially in the Devonian, Carboniferous and Cretaceous eras, was through plant photosynthesis. The question obviously arises as to whether we can drive such improved sequestration in the modern era in useful quantities and at useful rates. I believe that we can [4]. An overview of the article is given as a ‘mind map’ [5] in figure 1.

2. DYNAMICS OF CO₂ EXCHANGE BETWEEN TERRESTRIAL ECOSYSTEMS AND THE ATMOSPHERE

The first point to make is that terrestrial ecosystems including soils globally hold at least twice as much carbon (*ca* 1500–2500 Pg/Gt) as does the atmosphere (750 Pg) [6,7], so an overall increase in soil carbon by 10% implies (crudely) a decrease in (or more accurately a saving of an increase in) atmospheric carbon of at least 20%. (Note that at 750 Pg \approx 375 ppmv atmospheric CO₂, 2 Pg of C removed from or not added to the atmosphere \approx 1 ppmv removed or not added.) Overall fluxes to and from the soil are substantial, probably 60 Pg yr⁻¹ or more, albeit low in comparison to these pools [7–9]. This means that determining even the net direction of CO₂ transfer requires comparatively high-precision measurements [10]. Nevertheless, reasonably accurate estimates of net fluxes of ± 1 –2 t (ha yr)⁻¹ are easily attainable. Present agricultural ecosystems are rather depleted of soil carbon [11–14], and the existing ‘sink’ in such soils could certainly accommodate 50 t ha⁻¹. Given that world cropland and grassland each account for some 2300 Mha [15], the scope for increased sequestration in terrestrial ecosystems is clearly substantial [4], and—as with the exploitation of solar energy [16]—something we are very far from saturating.

It also needs to be recognised that there has been a certain dichotomy between most studies, that have been designed to analyse natural ecosystems—‘what is

* dbk@manchester.ac.uk

One contribution of 18 to a Theme Issue ‘Root growth and branching’.

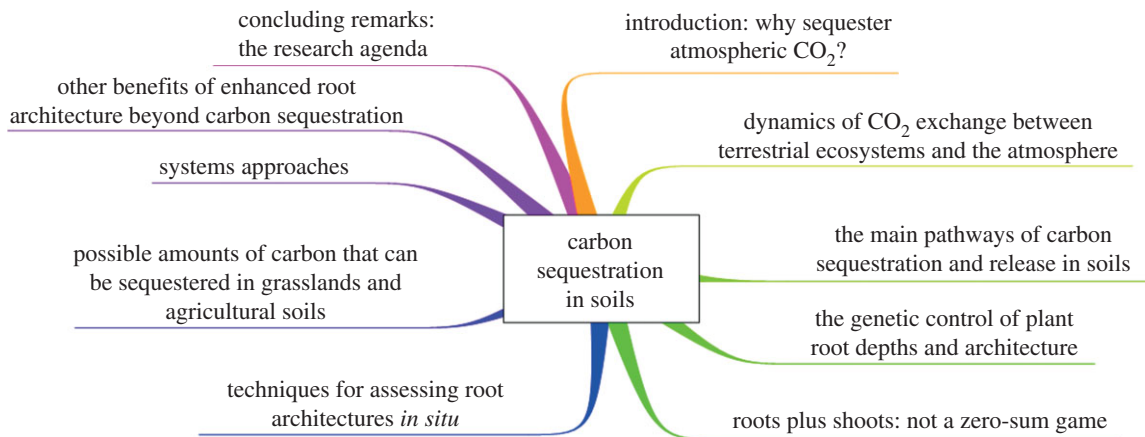


Figure 1. A 'mind map' [5] setting out the contents of the paper. To read it start at '12 o'clock' and read clockwise.

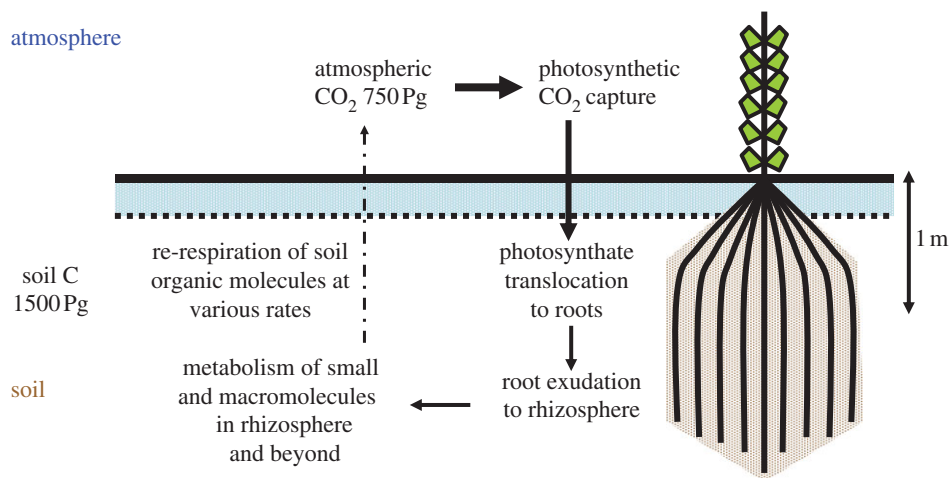


Figure 2. High-level analysis of the major processes involved in soil carbon sequestration for photosynthetically fixed CO_2 .

there'—and what could be done if we chose to breed and deploy suitable plants whether as food or non-food crops—'what might be' [4]. I shall tend to use the former as an existence proof of possibilities, while recognising that it is the latter that is the real goal.

Finally, here I note that the atmosphere is also in contact with the oceans [17] (and in pseudo- but not full equilibrium with them; if it were, the annual oscillations would tend to be much more heavily damped), and that the oceans sequester some 38 000 Pg C or 50 times that in the atmosphere [7,18]. This means that any eventual tendency to decrease atmospheric CO_2 effected by C sequestration in soil can be balanced by degassing of CO_2 from the oceans, so that what we are talking about here is stabilising values at their present levels rather than reducing them substantially (I thank Gideon Henderson for a useful discussion on this point). The liming of oceans may also offer some important opportunities there [19] (and see <http://www.cquestrate.com/>).

3. THE MAIN PATHWAYS OF CARBON SEQUESTRATION AND RELEASE IN SOILS

There are four main steps in a systems biology approach to understanding complex networks [20,21]. Steps 1 and 2 are essentially qualitative, and define the steps and the interacting partners (sources and sinks for

each step), and whether such interactions are direct and stoichiometric or indirect and 'regulatory'. Each 'step' may of course consist of multiple substeps. The third and fourth stages provide any known (or 'generalised' [22]) kinetic rate equations and the values of their parameters. Interoperable standards exist for describing such networks in XML [23], as well as for their graphical representation [24]. Armed with such information, it is then possible to develop a stochastic or ordinary differential equation model of the entire system of interest, whether based on 'lumped' compartments or involving explicit spatial differentiation.

In the spirit of step 1, figure 2 illustrates in general terms (for a more detailed version, see [25]) the main processes in soil operating to capture, sequester, transform and (in time) re-release atmospheric CO_2 . The main initial step is necessarily photosynthetic CO_2 capture, followed by its translocation below ground into plant roots [26]. Partly under genetic, nutritional and hormonal control, roots can extend to varying depths, and thereby deposit carbon as root biomass. Probably more important is the fact that roots exude all kinds of carbon-containing components into the rhizosphere [27], a complex and imperfectly characterised zone containing numerous microbes (including mycorrhiza [28,29]). From here, further transformations [30] can produce a variety of carbon-containing small and macromolecules that can exist in soil [31] and

contribute to the soil structure (not least by aggregating inorganic soil particles [11,32]). Depending on the nature of the molecule and other conditions such as pH, water activity and dissolved oxygen tension, such 'carbon' will reside in soil for a greater or lesser period (defining its 'recalcitrance' [33]). Interestingly, it is increasingly being recognised (e.g. [34,35]) that this recalcitrance may be more a property of where the molecule is sequestered than what it is chemically, and may also depend on supplies of fresh carbon [36]. At all events, eventually, most of the carbon will be re-respired to the atmosphere as CO₂. As a systems property [37], clearly the steady-state extent of sequestration depends on the topology and kinetics of all steps in the network, with the control of flux being distributed (e.g. [20,38]). Equally clearly, the relative contributions of different steps will vary in different soils [33]. However, as the step that determines the initial distribution of carbon in the soil by plant roots [26,39], it is the rooting process itself—the focus of this themed issue—on which we necessarily concentrate.

4. THE GENETIC CONTROL OF PLANT ROOT DEPTHS AND ARCHITECTURE, AND G X E INTERACTIONS

As mentioned, plant root depths and architecture are partly controlled by physical and agronomic (and hormonal, e.g. [40,41]) factors, but to a substantial degree [4,42–46] it is the genetic make-up of the organism (including genes whose products affect hormone production and distribution [47]) that determines how deep and bushy its roots can become. Some plants can indeed produce very substantial root architectures (e.g. [48–51]), and there is evidence for genetically determined variation in root architecture between plant types (e.g. [48,49,52]), between different cultivars of the same plant (e.g. [53–62]), and between different mutant strains with known genetic alleles or defects (e.g. [63–73]). In some cases, the number of genes involved in effecting substantial morphological changes may be quite small (e.g. [74,75]).

Clearly this encourages us to develop breeding programmes for plants with improved root architectures that can sequester carbon (and other nutrients, plus water) more effectively [4]. Such encouragement should be seen in broad terms as a contribution to ecosystem services [76], as well as agricultural yields [55,77], and the economic benefits derived therefrom might be enhanced by the payment of carbon credits [14,15,78–80].

I stress the breeding aspects, since we now know, especially from work with animals, that 'genomic selection' can speed genetic gain considerably [81–89], and it will soon be the norm to exploit modern whole-genome sequencing methods [90,91] to sequence every organism of interest in a breeding population.

This said, the necessary breeding will need to be assessed under a variety of agronomic conditions, since there is little doubt that agronomic practices can have a considerable impact on plant yields (there is substantial variation in yields across individual fields planted with the same crop, e.g. [92]), and in a manner that is of course dependent on the genetic make-up of the plants (G×E interactions). The

System of Rice Intensification (e.g. [93–98]) provides a particularly nice example of that.

5. ROOTS PLUS SHOOTS: NOT A ZERO-SUM GAME

It is sometimes opined that any breeding-based improvement (i.e. increase) in below-ground biomass would be balanced by an equivalent decrease in above-ground (and hence agriculturally harvestable) biomass. This is *a priori* implausible since they are more likely to feed each other than not, and most bioprocess fluxes are in fact demand-led [99]. At all events, there is plenty of evidence that the distribution of resource between root and shoot is not a zero-sum game:

- larger plant types as judged by above-ground biomass do in general tend to have larger roots—compare trees and typical crop plants, for instance [100,101];
- many mutants that have larger roots have above-ground biomass that is not smaller, and is often larger, than those of their parental wild type or 'baseline' strain (e.g. [45,77,102–112]);
- simple improvements in agronomic practices such as appropriate nutrient supply can increase the total amount of both root and shoot biomass (e.g. [113–117]), and in systems such as the System of Rice Intensification mentioned above apparently quite substantially so;
- similar behaviour (simultaneous increases in root and shoot biomass) can be induced by other non-host-genetic means that do not directly involve nutrients [118–120].

It is therefore entirely reasonable that we can improve plant root traits (and specifically to increase the size and extent of roots) in a manner that is not at the cost—and in many cases will likely be to the benefit—of above-ground traits including harvestable biomass. Both genetic and environmental (agronomic) approaches are likely to be of benefit here.

6. TECHNIQUES FOR ASSESSING ROOT ARCHITECTURES *IN SITU*

Science consists of both analysis and synthesis, and while high-throughput genomics has of course increased its throughput massively over the last decade, the same cannot be said of phenotyping [121]. Traditional (and many modern) methods for assessing the extent and nature of root phenotypes involve careful excavation and recording (e.g. [56,58,62,122]), but we need automated, non-invasive methods that likely involve some kind of spectroscopy or imaging [123] coupled to sophisticated computation. All have strengths and weaknesses, and some may be surrogates that measure properties (e.g. moisture content, or the force required to remove a plant from the soil [96]) that simply correlate with root properties, but some instrumental methods that appear promising include methods based on various kinds of impedimetry/capacitance/permittivity [124–128] (see also [129,130]) and impedance tomography [131], optical imaging [59,132–134], X-ray microtomography [57], ground-penetrating radar [131,135,136], microwave spectroscopy [137], neutron

spectroscopy and tomography [138] and magnetic resonance imaging [139] (that may be combined with positron emission tomography [140]). Fusion methods that combine multiple inputs can always [141] be expected to perform better than individual approaches.

7. POSSIBLE AMOUNTS OF C THAT CAN BE SEQUESTERED IN GRASSLANDS AND AGRICULTURAL SOILS

Calculations suggest (<http://dbkgroup.org/carbonsequestration/rootssystem.html>) [4] that the amount of C that can be stored in agricultural soils is considerably greater than is stored there now [12,14,142,143], namely in amounts similar to those that might be generated anthropogenically for the next 50 years, thereby stabilising atmospheric CO₂ at present levels. However, it is to be recognised that these calculations carry considerable uncertainty [144] as we know comparatively little about the rate and extent of root growth and in particular (e.g. [145–148]) the lifetime(s) of the various soil components before ultimately they are re-respired. The variation in sequestration time ('recalcitrance') of different forms of carbon-containing molecule can be very great, implying scope for increasing it by selective breeding (much as one can breed for enhanced degradability when this is desired for biomass crops [149]). Some analyses of existing grasslands and energy crops imply that at least 100 t ha⁻¹ of C may be sequestered in roots (or at least below ground) in the steady state [150,151], while tree forests usually sequester even more [152,153] (so deforestation [153,154] and forest drought [155] are especially damaging). One metre depth of soil containing just 1 per cent C at a bulk relative density of unity equates to 10 kg m⁻² or 100 t ha⁻¹, so estimates of 200 t ha⁻¹ in just the top metre alone [151] imply a considerably greater carrying capacity than that presently sustained, even before more recalcitrant forms of carbon such as biochar [156–158] are considered.

8. OTHER BENEFITS OF ENHANCED ROOT ARCHITECTURES BEYOND C SEQUESTRATION

This article has concentrated on the benefits to be had from improved root architectures largely in terms of carbon sequestration. However, it would be remiss not to stress that such improved root architectures also bring many other agricultural benefits [55,77], including improvements to soil structure [159], hydrology [160], drought tolerance (e.g. [161,162]) and N use efficiency [163]; in some cases these benefits may well prove to be more important overall, but can certainly be seen as additional benefits with regard to the C sequestration agenda.

9. SYSTEMS APPROACHES

As mentioned above, it is necessary to consider the whole (eco)system when undertaking studies of this type, and a strategy or intervention that seems to have a proximate benefit may have an ultimate disbenefit (or vice versa), due to ignoring important contributors to net balances or the propagation of the change via complex positive and negative feedback loops (e.g. [25], and for a more general account [164]). Thus changes in climate and

raised CO₂ may have unexpected effects on root–soil interactions [165] or any other processes, and the ability to sequester C will depend not only on the amount, extent and recalcitrance of plant roots but of the production rate and nature of root exudates, the amount of nitrogen [166] and other nutrients, and biophysical properties such as moisture content, soil compaction and the like. We also need to be ever mindful that changes affecting below-ground processes, especially if conditions are allowed to become anaerobic, might turn fixed CO₂ into much more damaging gases such as methane and nitrous oxide [2]—something to be avoided at all costs.

10. CONCLUDING REMARKS: THE RESEARCH AGENDA

This brief review purposely takes a relatively restricted and high-level approach to the problem of sequestering atmospheric carbon in soils. It recognises that (i) soils contain much carbon but are far from saturated with regard to organic matter, (ii) most soil carbon is derived from roots rather than from shoots and leaf litter, (iii) much of the carbon and most of the measurements thereof are restricted to the top 1 m of soil, and developing plants with 2 m roots could sequester considerably more C than is done presently, (iv) the transformation pathways and lifetimes of carbon components in the soil (both topsoil and subsoils), and what determines them both biologically and biophysically, are much less well understood than we would like, (v) the longer any particular form of carbon is held below ground before it is re-respired or emitted, the greater the amount that can be sequestered in the steady state, (vi) many analyses have concentrated more on 'what is there' than 'what we might do about it', and (vii) modern whole genome sequence-driven breeding offers huge opportunities for accelerating plant improvement.

As with scientific advances generally [167], we may expect to see iterative cycles, in that we may find empirically (through studying the variance between experiments [168]) that a particular cultivar treated with a particular agronomy does well with regard to soil carbon sequestration, and we may find from phenotypic (including 'omics') measurements that roots are involved mechanistically. We might then seek to apply directed breeding and agronomic practices that improve such root properties directly and then test if such crops also sequester carbon more effectively.

Thus, molecular breeding and appropriate agronomy (largely still matters for experiment), coupled to the necessary phenotyping approaches, especially non-invasive measurements of various kinds plus the attendant informatics and improved modelling [169], can lead to improved food and non-food crops that also have desirable carbon sequestration traits. Consequently, there is much to play for.

I thank many colleagues for useful discussions.

REFERENCES

- 1 Canadell, J. G. *et al.* 2007 Contributions to accelerating atmospheric CO₂ growth from economic activity, carbon intensity, and efficiency of natural sinks. *Proc.*

- Natl Acad. Sci. USA* **104**, 18 866–18 870. (doi:10.1073/pnas.0702737104)
- 2 Montzka, S. A., Dlugokencky, E. J. & Butler, J. H. 2011 Non-CO₂ greenhouse gases and climate change. *Nature* **476**, 43–50. (doi:10.1038/nature10322)
 - 3 Royer, D. L., Berner, R. A. & Beerling, D. J. 2001 Phanerozoic atmospheric CO₂ change: evaluating geochemical and paleobiological approaches. *Earth Sci. Rev.* **54**, 349–392. (doi:10.1016/S0012-8252(00)00042-8)
 - 4 Kell, D. B. 2011 Breeding crop plants with deep roots: their role in sustainable carbon, nutrient and water sequestration. *Ann. Bot.* **108**, 407–418. (doi:10.1093/aob/mcr175)
 - 5 Buzan, T. 2002 *How to mind map*. London, UK: Thorsons.
 - 6 Batjes, N. H. 1996 Total carbon and nitrogen in the soils of the world. *Eur. J. Soil. Sci.* **47**, 151–163. (doi:10.1111/j.1365-2389.1996.tb01386.x)
 - 7 Smith, P. 2004 Soils as carbon sinks: the global context. *Soil Use Manage.* **20**, 212–218. (doi:10.1079/SUM2004233)
 - 8 Bond-Lamberty, B. & Thomson, A. 2010 Temperature-associated increases in the global soil respiration record. *Nature* **464**, 579–582. (doi:10.1038/nature08930)
 - 9 Macias, F. & Arbestain, M. C. 2010 Soil carbon sequestration in a changing global environment. *Mitigation Adapt. Strateg. Glob. Change* **15**, 511–529. (doi:10.1007/s11027-010-9231-4)
 - 10 Smith, P. *et al.* 2012 Measurements necessary for assessing the net ecosystem carbon budget of croplands. *Agric. Ecosyst. Environ.* **139**, 302–315. (doi:10.1016/j.agee.2010.04.004)
 - 11 Jones, M. B. & Donnelly, A. 2004 Carbon sequestration in temperate grassland ecosystems and the influence of management, climate and elevated CO₂. *New Phytol.* **164**, 423–439. (doi:10.1111/j.1469-8137.2004.01201.x)
 - 12 Lorenz, K. & Lal, R. 2005 The depth distribution of soil organic carbon in relation to land use and management and the potential of carbon sequestration in subsoil horizons. *Adv. Agron.* **88**, 35–66. (doi:10.1016/S0065-2113(05)88002-2)
 - 13 Clay, J. 2011 Freeze the footprint of food. *Nature* **475**, 287–289. (doi:10.1038/475287a)
 - 14 Lal, R. 2011 Sequestering carbon in soils of agro-ecosystems. *Food Policy* **36**, S33–S9. (doi:10.1016/j.foodpol.2010.12.001)
 - 15 Smith, P. *et al.* 2008 Greenhouse gas mitigation in agriculture. *Phil. Trans. R. Soc. B* **363**, 789–813. (doi:10.1098/rstb.2007.2184)
 - 16 Blankenship, R. E. *et al.* 2011 Comparing photosynthetic and photovoltaic efficiencies and recognizing the potential for improvement. *Science* **332**, 805–809. (doi:10.1126/science.1200165)
 - 17 Khaliwala, S., Primeau, F. & Hall, T. 2009 Reconstruction of the history of anthropogenic CO₂ concentrations in the ocean. *Nature* **462**, 346–349. (doi:10.1038/nature08526)
 - 18 MacKay, D. J. C. 2008 *Sustainable energy: without the hot air*. Cambridge, UK: UIT Cambridge. See <http://www.withouthotair.com/>.
 - 19 Kheshgi, H. S. 1995 Sequestering atmospheric carbon dioxide by increasing ocean alkalinity. *Energy* **20**, 915–922. (doi:10.1016/0360-5442(95)00035-F)
 - 20 Kell, D. B. 2006 Metabolomics, modelling and machine learning in systems biology: towards an understanding of the languages of cells. The 2005 Theodor Bücher lecture. *FEBS J.* **273**, 873–894. (doi:10.1111/j.1742-4658.2006.05136.x)
 - 21 Herrgård, M. J. *et al.* 2008 A consensus yeast metabolic network obtained from a community approach to systems biology. *Nature Biotechnol.* **26**, 1155–1160. (doi:10.1038/nbt1492)
 - 22 Smallbone, K., Simeonidis, E., Broomhead, D. S. & Kell, D. B. 2007 Something from nothing: bridging the gap between constraint-based and kinetic modelling. *FEBS J.* **274**, 5576–5585. (doi:10.1111/j.1742-4658.2007.06076.x)
 - 23 Hucka, M. *et al.* 2003 The systems biology markup language (SBML): a medium for representation and exchange of biochemical network models. *Bioinformatics* **19**, 524–531. (doi:10.1093/bioinformatics/btg015)
 - 24 Le Novère, N. *et al.* 2009 The systems biology graphical notation. *Nat. Biotechnol.* **27**, 735–741. (doi:10.1038/nbt.1558)
 - 25 Bardgett, R. D. 2011 The root of the problem. *The Scientist*. **25**, 32–37.
 - 26 Hodge, A., Berta, G., Doussan, C., Merchan, F. & Crespi, M. 2009 Plant root growth, architecture and function. *Plant Soil* **321**, 153–187. (doi:10.1007/s11104-009-9929-9)
 - 27 Hinsinger, P., Bengough, A. G., Vetterlein, D. & Young, I. M. 2009 Rhizosphere: biophysics, biogeochemistry and ecological relevance. *Plant Soil* **321**, 117–152. (doi:10.1007/s11104-008-9885-9)
 - 28 Parniske, M. 2008 Arbuscular mycorrhiza: the mother of plant root endosymbioses. *Nat. Rev. Microbiol.* **6**, 763–775. (doi:10.1038/nrmicro1987)
 - 29 Richardson, A. E., Barea, J. M., McNeill, A. M. & Prigent-Combaret, C. 2009 Acquisition of phosphorus and nitrogen in the rhizosphere and plant growth promotion by microorganisms. *Plant Soil* **321**, 305–339. (doi:10.1007/s11104-009-9895-2)
 - 30 Rumpel, C. & Kögel-Knabner, I. 2011 Deep soil organic matter—a key but poorly understood component of terrestrial C cycle. *Plant Soil* **338**, 143–158. (doi:10.1007/s11104-010-0391-5)
 - 31 Kögel-Knabner, I. 2002 The macromolecular organic composition of plant and microbial residues as inputs to soil organic matter. *Soil Biol. Biochem.* **34**, 139–162. (doi:10.1016/S0038-0717(01)00158-4)
 - 32 Wilson, G. W., Rice, C. W., Rillig, M. C., Springer, A. & Hartnett, D. C. 2009 Soil aggregation and carbon sequestration are tightly correlated with the abundance of arbuscular mycorrhizal fungi: results from long-term field experiments. *Ecol. Lett.* **12**, 452–461. (doi:10.1111/j.1461-0248.2009.01303.x)
 - 33 von Lützow, M., Kögel-Knabner, I., Ekschmitt, K., Matzner, E., Guggenberger, G. & Marschner, B. 2006 Stabilization of organic matter in temperate soils: mechanisms and their relevance under different soil conditions: a review. *Eur. J. Soil Sci.* **57**, 426–445. (doi:10.1111/j.1365-2389.2006.00809.x)
 - 34 Rasse, D. P., Rumpel, C. & Dignac, M. F. 2005 Is soil carbon mostly root carbon? Mechanisms for a specific stabilisation. *Plant Soil* **269**, 341–356. (doi:10.1007/s11104-004-0907-y)
 - 35 Schmidt, M. W. I. *et al.* 2011 Persistence of soil organic matter as an ecosystem property. *Nature* **478**, 49–56. (doi:10.1038/nature10386)
 - 36 Fontaine, S., Barot, S., Barre, P., Bdioui, N., Mary, B. & Rumpel, C. 2007 Stability of organic carbon in deep soil layers controlled by fresh carbon supply. *Nature* **450**, 277–280. (doi:10.1038/nature06275)
 - 37 Lucas, M., Laplaze, L. & Bennett, M. J. 2011 Plant systems biology: network matters. *Plant Cell Environ.* **34**, 535–553. (doi:10.1111/j.1365-3040.2010.02273.x)
 - 38 Kell, D. B. & Westerhoff, H. V. 1986 Metabolic control theory: its role in microbiology and biotechnology. *FEMS Microbiol. Rev.* **39**, 305–320. (doi:10.1111/j.1574-6968.1986.tb01863.x)
 - 39 Orwin, K. H., Buckland, S. M., Johnson, D., Turner, B. L., Smart, S., Oakley, S. & Bardgett, R. D. 2010

- Linkages of plant traits to soil properties and the functioning of temperate grassland. *J. Ecol.* **98**, 1074–1083. (doi:10.1111/j.1365-2745.2010.01679.x)
- 40 Bishopp, A., Help, H. & Helariutta, Y. 2009 Cytokinin signaling during root development. *Int. Rev. Cell Mol. Biol.* **276**, 1–48. (doi:10.1016/S1937-6448(09)76001-0)
- 41 Fukaki, H. & Tasaka, M. 2009 Hormone interactions during lateral root formation. *Plant Mol. Biol.* **69**, 437–449. (doi:10.1007/s11103-008-9417-2)
- 42 Doussan, C., Pages, L. & Pierret, A. 2003 Soil exploration and resource acquisition by plant roots: an architectural and modelling point of view. *Agronomie* **23**, 419–431. (doi:10.1051/agro:2003027)
- 43 Kato, Y., Abe, J., Kamoshita, A. & Yamagishi, J. 2006 Genotypic variation in root growth angle in rice (*Oryza sativa*, L.) and its association with deep root development in upland fields with different water regimes. *Plant Soil* **287**, 117–129. (doi:10.1007/s11104-006-9008-4)
- 44 Osmont, K. S., Sibout, R. & Hardtke, C. S. 2007 Hidden branches: developments in root system architecture. *Annu. Rev. Plant Biol.* **58**, 93–113. (doi:10.1146/annurev.arplant.58.032806.104006)
- 45 Abberton, M. T., Marshall, A. H., Humphreys, M. W., Macduff, J. H., Collins, R. P. & Marley, C. L. 2008 Genetic improvement of forage species to reduce the environmental impact of temperate livestock grazing systems. *Adv. Agron.* **98**, 311–355. (doi:10.1016/S0065-2113(08)00206-X)
- 46 Dello Ioio, R., Nakamura, K., Moubayidin, L., Perilli, S., Taniguchi, M., Morita, M. T., Aoyama, T., Costantino, P. & Sabatini, S. 2008 A genetic framework for the control of cell division and differentiation in the root meristem. *Science* **322**, 1380–1384. (doi:10.1126/science.1164147)
- 47 Casson, S. A. & Lindsey, K. 2003 Genes and signalling in root development. *New Phytol.* **158**, 11–38.
- 48 Jackson, R. B., Canadell, J., Ehleringer, J. R., Mooney, H. A., Sala, O. E. & Schulze, E. D. 1996 A global analysis of root distributions for terrestrial biomes. *Oecologia* **108**, 389–411. (doi:10.1007/BF00333714)
- 49 Jobbágy, E. G. & Jackson, R. B. 2000 The vertical distribution of soil organic carbon and its relation to climate and vegetation. *Ecol. Appl.* **10**, 423–436. (doi:10.1890/1051-0761(2000)010[0423:TVDOSO]2.0.CO;2)
- 50 Cox, T. S., Glover, J. D., Van Tassel, D. L., Cox, C. M. & DeHaan, L. R. 2006 Prospects for developing perennial-grain crops. *Bioscience* **56**, 649–659. (doi:10.1641/0006-3568(2006)56[649:PFDPGC]2.0.CO;2)
- 51 Glover, J. D. *et al.* 2010 Increased food and ecosystem security via perennial grains. *Science* **328**, 1638–1639. (doi:10.1126/science.1188761)
- 52 Burch, G. J. & Johns, G. G. 1978 Root absorption of water and physiological responses to water deficits by *Festuca arundinacea* Schreb. and *Trifolium repens* L. *Aust. J. Plant Physiol.* **5**, 859–871. (doi:10.1071/PP9780859)
- 53 Loudet, O., Gaudon, V., Trubuil, A. & Daniel-Vedele, F. 2005 Quantitative trait loci controlling root growth and architecture in *Arabidopsis thaliana* confirmed by heterogeneous inbred family. *Theor. Appl. Genet.* **110**, 742–753. (doi:10.1007/s00122-004-1900-9)
- 54 De Smet, I., Vanneste, S., Inzé, D. & Beeckman, T. 2006 Lateral root initiation or the birth of a new meristem. *Plant Mol. Biol.* **60**, 871–887. (doi:10.1007/s11103-005-4547-2)
- 55 Lynch, J. P. 2007 Roots of the second green revolution. *Austr. J. Bot.* **55**, 493–512. (doi:10.1071/BT06118)
- 56 Danjon, F. & Reubens, B. 2008 Assessing and analyzing 3D architecture of woody root systems, a review of methods and applications in tree and soil stability, resource acquisition and allocation. *Plant Soil* **303**, 1–34. (doi:10.1007/s11104-007-9470-7)
- 57 Gregory, P. J. *et al.* 2009 Root phenomics of crops: opportunities and challenges. *Funct. Plant Biol.* **36**, 922–929. (doi:10.1071/FP09150)
- 58 Kutschera, L., Lichtenegger, E. & Sobotik, M. 2009 *Wurzelatlas der Kulturpflanzen gemäßigter Gebiete mit Arten des Feldgemüsebaues*. Frankfurt/Main: DLG Verlag.
- 59 Iyer-Pascuzzi, A. S., Symonova, O., Mileyko, Y., Hao, Y., Belcher, H., Harer, J., Weitz, J. S. & Benfey, P. N. 2010 Imaging and analysis platform for automatic phenotyping and trait ranking of plant root systems. *Plant Physiol.* **152**, 1148–1157. (doi:10.1104/pp.109.150748)
- 60 Tuberosa, R., Salvi, S., Giuliani, S., Sanguineti, M. C., Frascaroli, E., Conti, S. & Landi, P. 2010 Genomics of root architecture and functions in maize. In *Root genomics* (eds A. Costa de Oliveira & R. K. Varshney), pp. 179–204. Heidelberg, Germany: Springer.
- 61 Hund, A., Reimer, R. & Messmer, R. 2011 A consensus map of QTLs controlling the root length of maize. *Plant Soil* **344**, 143. (doi:10.1007/s11104-011-0735-9)
- 62 Trachsel, S., Kaeppler, S. M., Brown, K. M. & Lynch, J. P. 2011 Shovelomics: high throughput phenotyping of maize (*Zea mays* L.) root architecture in the field. *Plant Soil* **341**, 75–87. (doi:10.1007/s11104-010-0623-8)
- 63 Zhang, H. & Forde, B. G. 1998 An *Arabidopsis* MADS box gene that controls nutrient-induced changes in root architecture. *Science* **279**, 407–409. (doi:10.1126/science.279.5349.407)
- 64 Hardtke, C. S., Mouchel, C. F. & Briggs, G. C. 2004 Natural genetic variation in *Arabidopsis* identifies BREVIS RADIX, a novel regulator of cell proliferation and elongation in the root. *Genes Dev.* **18**, 700–714. (doi:10.1101/gad.1187704)
- 65 Levesque, M. P. *et al.* 2006 Whole-genome analysis of the SHORT-ROOT developmental pathway in *Arabidopsis*. *PLoS Biol.* **4**, e143. (doi:10.1371/journal.pbio.0040143)
- 66 Lelandais-Brière, C., Jovanovic, M., Torres, G. A. M., Perrin, Y., Lemoine, R., Corre-Menguy, F. & Hartmann, C. 2007 Disruption of AtOCT1, an organic cation transporter gene, affects root development and carnitine-related responses in *Arabidopsis*. *Plant J.* **51**, 154–164. (doi:10.1111/j.1365-313X.2007.03131.x)
- 67 Hochholdinger, F. & Tuberosa, R. 2009 Genetic and genomic dissection of maize root development and architecture. *Curr. Opin. Plant Biol.* **12**, 172–177. (doi:10.1016/j.pbi.2008.12.002)
- 68 Hochholdinger, F. 2009 The maize root system: morphology, anatomy, and genetics. In *Handbook of maize: its biology* (eds J. L. Bennetzen & S. C. Hake), pp. 145–160. Berlin, Germany: Springer.
- 69 Rebouillat, J. *et al.* 2009 Molecular genetics of rice root development. *Rice* **2**, 15–34. (doi:10.1007/s12284-008-9016-5)
- 70 Benfey, P. N., Bennett, M. & Schiefelbein, J. 2010 Getting to the root of plant biology: impact of the *Arabidopsis* genome sequence on root research. *Plant J.* **61**, 992–1000. (doi:10.1111/j.1365-313X.2010.04129.x)
- 71 Coudert, Y., Perin, C., Courtois, B., Khong, N. G. & Gantet, P. 2010 Genetic control of root development in rice, the model cereal. *Trends Plant Sci.* **15**, 219–226. (doi:10.1016/j.tplants.2010.01.008)
- 72 Scacchi, E., Salinas, P., Gujas, B., Santuari, L., Krogan, N., Ragni, L., Berleth, T. & Hardtke, C. S. 2010 Spatio-temporal sequence of cross-regulatory events in root meristem growth. *Proc. Natl Acad. Sci. USA* **107**, 22 734–22 739. (doi:10.1073/pnas.1014716108)

- 73 Lynch, J. P. 2011 Root phenes for enhanced soil exploration and phosphorus acquisition: tools for future crops. *Plant Physiol.* **156**, 1041–1049. (doi:10.1104/pp.111.175414)
- 74 Thomas, H., Thomas, H. M. & Ougham, H. 2000 Annuality, perenniality and cell death. *J. Exp. Bot.* **51**, 1781–1788. (doi:10.1093/jexbot/51.352.1781)
- 75 Melzer, S., Lens, F., Gennen, J., Vanneste, S., Rohde, A. & Beeckman, T. 2008 Flowering-time genes modulate meristem determinacy and growth form in *Arabidopsis thaliana*. *Nat. Genet.* **40**, 1489–1492. (doi:10.1038/ng.253)
- 76 Millennium Ecosystem Assessment 2005 *Ecosystems and human well-being*. Washington, DC: Island Press. See <http://www.millenniumassessment.org/documents/document.356.aspx.pdf>.
- 77 Den Herder, G., Van Isterdael, G., Beeckman, T. & De Smet, I. 2010 The roots of a new green revolution. *Trends Plant Sci.* **15**, 600–607. (doi:10.1016/j.tplants.2010.08.009)
- 78 Rokityanskiy, D., Benitez, P. C., Kraxner, F., McCallum, I., Obersteiner, M., Rametsteiner, E. & Yamagata, Y. 2007 Geographically explicit global modeling of land-use change, carbon sequestration, and biomass supply. *Technol. Forecasting Soc. Change* **74**, 1057–1082. (doi:10.1016/j.techfore.2006.05.022)
- 79 MacLeod, M. *et al.* 2010 Developing greenhouse gas marginal abatement cost curves for agricultural emissions from crops and soils in the UK. *Agric. Syst.* **103**, 198–209. (doi:10.1016/j.agsy.2010.01.002)
- 80 Smith, P. & Olesen, J. E. 2010 Synergies between the mitigation of, and adaptation to, climate change in agriculture. *J. Agric. Sci.* **148**, 543–552. (doi:10.1017/S0021859610000341)
- 81 Meuwissen, T. H., Hayes, B. J. & Goddard, M. E. 2001 Prediction of total genetic value using genome-wide dense marker maps. *Genetics* **157**, 1819–1829.
- 82 Gianola, D., Perez-Enciso, M. & Toro, M. A. 2003 On marker-assisted prediction of genetic value: beyond the ridge. *Genetics* **163**, 347–365.
- 83 Xu, S. 2003 Estimating polygenic effects using markers of the entire genome. *Genetics* **163**, 789–801.
- 84 Schaeffer, L. R. 2006 Strategy for applying genome-wide selection in dairy cattle. *J. Anim. Breed. Genet.* **123**, 218–223. (doi:10.1111/j.1439-0388.2006.00595.x)
- 85 Goddard, M. E. & Hayes, B. J. 2007 Genomic selection. *J. Anim. Breed. Genet.* **124**, 323–330. (doi:10.1111/j.1439-0388.2007.00702.x)
- 86 Luan, T., Woolliams, J. A., Lien, S., Kent, M., Svendsen, M. & Meuwissen, T. H. 2009 The accuracy of genomic selection in Norwegian red cattle assessed by cross-validation. *Genetics* **183**, 1119–1126. (doi:10.1534/genetics.109.107391)
- 87 Ødegård, J., Yazdi, M. H., Sonesson, A. K. & Meuwissen, T. H. 2009 Incorporating desirable genetic characteristics from an inferior into a superior population using genomic selection. *Genetics* **181**, 737–745. (doi:10.1534/genetics.108.098160)
- 88 Fahrenkrug, S. C. *et al.* 2010 Precision genetics for complex objectives in animal agriculture. *J. Anim. Sci.* **88**, 2530–2539. (doi:10.2527/jas.2010-2847)
- 89 Maenhout, S., De Baets, B. & Haesaert, G. 2010 Graph-based data selection for the construction of genomic prediction models. *Genetics* **185**, 1463–1475. (doi:10.1534/genetics.110.116426)
- 90 Edwards, D. & Batley, J. 2010 Plant genome sequencing: applications for crop improvement. *Plant Biotechnol. J.* **8**, 2–9. (doi:10.1111/j.1467-7652.2009.00459.x)
- 91 Meuwissen, T. & Goddard, M. 2010 Accurate prediction of genetic values for complex traits by whole-genome resequencing. *Genetics* **185**, 623–631. (doi:10.1534/genetics.110.116590)
- 92 Lark, R. M. 2001 Some tools for parsimonious modeling and interpretation of within-field variation of soil and crop systems. *Soil Tillage Res.* **58**, 99–111. (doi:10.1016/S0167-1987(00)00161-6)
- 93 Stoop, W. A., Uphoff, N. & Kassam, A. 2002 A review of agricultural research issues raised by the system of rice intensification (SRI) from Madagascar: opportunities for improving farming systems for resource-poor farmers. *Agric. Syst.* **71**, 249–274. (doi:10.1016/S0308-521X(01)00070-1)
- 94 Mishra, A. & Salokhe, V. M. 2010 The effects of planting pattern and water regime on root morphology, physiology and grain yield of rice. *J. Agron. Crop. Sci.* **196**, 368–378. (doi:10.1111/j.1439-037X.2010.00421.x)
- 95 Thakur, A. K., Uphoff, N. & Antony, E. 2010 An assessment of physiological effects of System of Rice Intensification (SRI) practices compared with recommended rice cultivation practices in India. *Exp. Agric.* **46**, 77–98. (doi:10.1017/S0014479709990548)
- 96 Barison, J. & Uphoff, N. 2011 Rice yield and its relation to root growth and nutrient-use efficiency under SRI and conventional cultivation: an evaluation in Madagascar. *Paddy Water Environ.* **9**, 65–78. (doi:10.1007/s10333-010-0229-z)
- 97 Mishra, A. & Salokhe, V. M. 2011 Rice root growth and physiological responses to SRI water management and implications for crop productivity. *Paddy Water Environ.* **9**, 41–52. (doi:10.1007/s10333-010-0240-4)
- 98 Uphoff, N., Kassam, A. & Harwood, R. 2011 SRI as a methodology for raising crop and water productivity: productive adaptations in rice agronomy and irrigation water management. *Paddy Water Environ.* **9**, 3–11. (doi:10.1007/s10333-010-0224-4)
- 99 Hofmeyr, J. S. & Cornish-Bowden, A. 2000 Regulating the cellular economy of supply and demand. *FEBS Lett.* **476**, 47–51. (doi:10.1016/S0014-5793(00)01668-9)
- 100 Gregory, P. J. 2006 *Plant roots: growth, activity and interaction with soils*. Oxford, UK: Blackwell.
- 101 Mokany, K., Raison, R. J. & Prokushkin, A. S. 2006 Critical analysis of root:shoot ratios in terrestrial biomes. *Glob. Change Biol.* **12**, 84–96. (doi:10.1111/j.1365-2486.2005.001043.x)
- 102 Bingham, I. J. 2001 Soil-root-canopy interactions. *Ann. Appl. Biol.* **138**, 243–51. (doi:10.1111/j.1744-7348.2001.tb00108.x)
- 103 Price, A. H., Steele, K. A., Gorham, J., Bridges, J. M., Moore, B. J., Evans, J. L., Richardson, P. & Jones, R. G. W. 2002 Upland rice grown in soil-filled chambers and exposed to contrasting water-deficit regimes I. Root distribution, water use and plant water status. *Field Crops Res.* **76**, 11–24. (doi:10.1016/S0378-4290(02)00012-6)
- 104 Bonos, S. A., Rush, D., Hignight, K. & Meyer, W. A. 2004 Selection for deep root production in tall fescue and perennial ryegrass. *Crop Sci.* **44**, 1770–1775. (doi:10.2135/cropsci2004.1770)
- 105 Passioura, J. 2006 Increasing crop productivity when water is scarce—from breeding to field management. *Agric. Water Manage.* **80**, 176–196. (doi:10.1016/j.agwat.2005.07.012)
- 106 Wang, H., Inukai, Y. & Yamauchi, A. 2006 Root development and nutrient uptake. *Crit. Rev. Plant. Sci.* **25**, 279–301. (doi:10.1080/07352680600709917)
- 107 Hund, A., Richner, W., Soldati, A., van Fracheboud, Y. & Stamp, P. 2007 Root morphology and

- photosynthetic performance of maize inbred lines at low temperature. *Eur. J. Agron.* **27**, 52–61. (doi:10.1016/j.eja.2007.01.003)
- 108 Sanguineti, M. C., Li, S., Maccaferri, M., Corneti, S., Rotondo, F., Chiari, T. & Tuberosa, R. 2007 Genetic dissection of seminal root architecture in elite durum wheat germplasm. *Ann. Appl. Biol.* **151**, 291–305. (doi:10.1111/j.1744-7348.2007.00198.x)
- 109 Crush, J. R., Nichols, S. N., Easton, H. S., Ouyang, L. & Hume, D. E. 2009 Comparisons between wild populations and bred perennial ryegrasses for root growth and root/shoot partitioning. *N Z J. Agric. Res.* **52**, 161–169. (doi:10.1080/00288230909510500)
- 110 Hammer, G. L., Dong, Z. S., McLean, G., Doherty, A., Messina, C., Schusler, J., Zinselmeier, C., Paszkiewicz, S. & Cooper, M. 2009 Can changes in canopy and/or root system architecture explain historical maize yield trends in the US corn belt? *Crop Sci.* **49**, 299–312. (doi:10.2135/cropsci2008.03.0152)
- 111 Zhang, H., Yang, J., Xue, Y. G., Wang, Z. Q. & Zhang, J. H. 2009 Morphological and physiological traits of roots and their relationships with shoot growth in 'super' rice. *Field Crops Res.* **113**, 31–40. (doi:10.1016/j.fcr.2009.04.004)
- 112 Peng, Y. F., Niu, J. F., Peng, Z. P., Zhang, F. S. & Li, C. J. 2010 Shoot growth potential drives N uptake in maize plants and correlates with root growth in the soil. *Field Crops Res.* **115**, 85–93. (doi:10.1016/j.fcr.2009.10.006)
- 113 Ainsworth, E. A. *et al.* 2002 A meta-analysis of elevated [CO₂] effects on soybean (*Glycine max*) physiology, growth and yield. *Glob. Change Biol.* **8**, 695–709. (doi:10.1046/j.1365-2486.2002.00498.x)
- 114 Gastal, F. & Lemaire, G. 2002 N uptake and distribution in crops: an agronomical and ecophysiological perspective. *J. Exp. Bot.* **53**, 789–799. (doi:10.1093/jxb/53.370.789)
- 115 Linkohr, B. I., Williamson, L. C., Fitter, A. H. & H. M. O., Leyser 2002 Nitrate and phosphate availability and distribution have different effects on root system architecture of *Arabidopsis*. *Plant J.* **29**, 751–760. (doi:10.1046/j.1365-313X.2002.01251.x)
- 116 Grechi, I., Vivin, P., Hilbert, G., Milin, S., Robert, T. & Gaudillère, J. P. 2007 Effect of light and nitrogen supply on internal C : N balance and control of root-to-shoot biomass allocation in grapevine. *Env. Exp. Bot.* **59**, 139–149. (doi:10.1016/j.envexpbot.2005.11.002)
- 117 Huang, H. C. & Erickson, R. S. 2007 Effect of seed treatment with *Rhizobium leguminosarum* on *Pythium* damping-off, seedling height, root nodulation, root biomass, shoot biomass, and seed yield of pea and lentil. *J. Phytopathol.* **155**, 31–37. (doi:10.1111/j.1439-0434.2006.01189.x)
- 118 Rodriguez, R. J., Freeman, D. C., McArthur, E. D., Kim, Y. O. & Redman, R. S. 2009 Symbiotic regulation of plant growth, development and reproduction. *Commun. Integr. Biol.* **2**, 141–143.
- 119 Tokuhisa, D., Shinano, T., Watanabe, T., Yamamura, T. & Osaki, M. 2010 Promotion of root growth by the application of inosine. *Soil Sci. Plant Nutr.* **56**, 272–280. (doi:10.1111/j.1747-0765.2010.00452.x)
- 120 Redman, R. S., Kim, Y. O., Woodward, C. J. D. A., Greer, C., Espino, L., Doty, S. L. & Rodriguez, R. J. 2011 Increased fitness of rice plants to abiotic stress via habitat adapted symbiosis: a strategy for mitigating impacts of climate change. *PLoS ONE* **6**, e14823. (doi:10.1371/journal.pone.0014823)
- 121 Tester, M. & Langridge, P. 2010 Breeding technologies to increase crop production in a changing world. *Science* **327**, 818–822. (doi:10.1126/science.1183700)
- 122 Pagès, L., Serra, V., Draye, X., Doussan, C. & Pierret, A. 2010 Estimating root elongation rates from morphological measurements of the root tip. *Plant Soil* **328**, 35–44. (doi:10.1007/s11104-009-0079-x)
- 123 Zhu, J. M., Ingram, P. A., Benfey, P. N. & Elich, T. 2011 From lab to field, new approaches to phenotyping root system architecture. *Curr. Opin. Plant Biol.* **14**, 310–317. (doi:10.1016/j.pbi.2011.03.020)
- 124 Chloupek, O. 1977 Evaluation of size of a plant's root system using its electrical capacitance. *Plant Soil* **48**, 525–532. (doi:10.1007/BF02187258)
- 125 Nadezhdina, N. & Čermák, J. 2003 Instrumental methods for studies of structure and function of root systems of large trees. *J. Exp. Bot.* **54**, 1511–1521. (doi:10.1093/jxb/erg154)
- 126 McBride, R. A., Preston, G. M., Bryan, J. & Candido, M. 2004 Estimating root mass in young hybrid poplar trees using the electrical capacitance method. *Agroforestr. Syst.* **60**, 305–309. (doi:10.1023/B:AGFO.0000024439.41932.e2)
- 127 McBride, R., Candido, M. & Ferguson, J. 2008 Estimating root mass in maize genotypes using the electrical capacitance method. *Arch. Agron. Soil Sci.* **54**, 215–226. (doi:10.1080/03650340701790658)
- 128 Pitre, F. E., Brereton, N. J. B., Audoire, S., Richter, G. M., Shield, I. & Karp, A. 2010 Estimating root biomass in *Salix viminalis* × *Salix schwerinii* cultivar 'Olof' using the electrical capacitance method. *Plant Biosyst.* **144**, 479–483.
- 129 Harris, C. M., Todd, R. W., Bungard, S. J., Lovitt, R. W., Morris, J. G. & Kell, D. B. 1987 The dielectric permittivity of microbial suspensions at radio frequencies: a novel method for the estimation of microbial biomass. *Enzyme Microbial. Technol.* **9**, 181–186. (doi:10.1016/0141-0229(87)90075-5)
- 130 Pethig, R. & Kell, D. B. 1987 The passive electrical properties of biological systems: their significance in physiology, biophysics and biotechnology. *Phys. Med. Biol.* **32**, 933–970. (doi:10.1088/0031-9155/32/8/001)
- 131 Zenone, T. *et al.* 2008 Preliminary use of ground-penetrating radar and electrical resistivity tomography to study tree roots in pine forests and poplar plantations. *Funct. Plant Biol.* **35**, 1047–1058. (doi:10.1071/FP08062)
- 132 Durham Brooks, T. L., Miller, N. D. & Spalding, E. P. 2010 Plasticity of *Arabidopsis* root gravitropism throughout a multidimensional condition space quantified by automated image analysis. *Plant Physiol.* **152**, 206–216. (doi:10.1104/pp.109.145292)
- 133 French, A., Ubeda-Tomas, S., Holman, T. J., Bennett, M. J. & Pridmore, T. 2009 High-throughput quantification of root growth using a novel image-analysis tool. *Plant Physiol.* **150**, 1784–1795. (doi:10.1104/pp.109.140558)
- 134 Vegapareddy, M., Richter, G. M. & Goulding, K. W. T. 2010 Using digital image analysis to quantify the architectural parameters of roots grown in thin rhizotrons. *Plant Biosyst.* **144**, 499–506.
- 135 Stover, D. B., Day, F. P., Butnor, J. R. & Drake, B. G. 2007 Effect of elevated CO₂ on coarse-root biomass in Florida scrub detected by ground-penetrating radar. *Ecology* **88**, 1328–1334. (doi:10.1890/06-0989)
- 136 Cui, X. H., Chen, J., Shen, J. S., Cao, X., Chen, X. H. & Zhu, X. L. 2011 Modeling tree root diameter and biomass by ground-penetrating radar. *Sci. China Earth Sci.* **54**, 711–719. (doi:10.1007/s11430-010-4103-z)

- 137 Jackson, T. J., Le Vine, D. M., Hsu, A. Y., Oldak, A., Starks, P. J., Swift, C. T., Isham, J. D. & Haken, M. 1999 Soil moisture mapping at regional scales using microwave radiometry: the Southern Great Plains Hydrology Experiment. *IEEE Trans. Geosci. Remote Sens.* **37**, 2136–2151. (doi:10.1109/36.789610)
- 138 Esser, H. G., Carminati, A., Vontobel, P., Lehmann, E. H. & Oswald, S. E. 2010 Neutron radiography and tomography of water distribution in the root zone. *J. Plant Nutr. Soil Sci.* **173**, 757–764. (doi:10.1002/jpln.200900188)
- 139 Segal, E., Kushnir, T., Mualem, Y. & Shani, U. 2008 Microsensing of water dynamics and root distributions in sandy soils. *Vadose Zone J.* **7**, 1018–1026. (doi:10.2136/vzj2007.0121)
- 140 Nagel, K. A. *et al.* 2009 Temperature responses of roots: impact on growth, root system architecture and implications for phenotyping. *Funct. Plant Biol.* **36**, 947–959. (doi:10.1071/FP09184)
- 141 Hastie, T., Tibshirani, R. & Friedman, J. 2001 *The elements of statistical learning: data mining, inference and prediction*. Berlin, Germany: Springer.
- 142 Lal, R. 2004 Soil carbon sequestration to mitigate climate change. *Geoderma* **123**, 1–22. (doi:10.1016/j.geoderma.2004.01.032)
- 143 Thomson, A. M., Izaurrealde, R. C., Smith, S. J. & Clarke, L. E. 2008 Integrated estimates of global terrestrial carbon sequestration. *Glob. Environ. Change Hum. Policy Dimens.* **18**, 192–203. (doi:10.1016/j.gloenvcha.2007.10.002)
- 144 Chapin, F. S., McFarland, J., McGuire, A. D., Euskirchen, E. S., Ruess, R. W. & Kielland, K. 2009 The changing global carbon cycle: linking plant-soil carbon dynamics to global consequences. *J. Ecol.* **97**, 840–850. (doi:10.1111/j.1365-2745.2009.01529.x)
- 145 Amelung, W., Brodowski, S., Sandhage-Hofmann, A. & Bol, R. 2008 Combining biomarker with stable isotope analyses for assessing the transformation and turnover of soil organic matter. *Adv. Agron.* **100**, 155–250. (doi:10.1016/S0065-2113(08)00606-8)
- 146 Marschner, B. *et al.* 2008 How relevant is recalcitrance for the stabilization of organic matter in soils? *J. Plant Nutr. Soil Sci.* **171**, 91–110. (doi:10.1002/jpln.200700049)
- 147 Blagodatskaya, E., Yuyukina, T., Blagodatsky, S. & Kuzyakov, Y. 2011 Turnover of soil organic matter and of microbial biomass under C(3)–C(4) vegetation change: consideration of (13)C fractionation and preferential substrate utilization. *Soil Biol. Biochem.* **43**, 159–166. (doi:10.1016/j.soilbio.2010.09.028)
- 148 Spence, A., Simpson, A. J., McNally, D. J., Moran, B. W., McCaul, M. V., Hart, K., Paull, B. & Kelleher, B. P. 2011 The degradation characteristics of microbial biomass in soil. *Geochim. Cosmochim. Acta* **75**, 2571–2581. (doi:10.1016/j.gca.2011.03.012)
- 149 Mortimer, J. C. *et al.* 2010 Absence of branches from xylan in *Arabidopsis gux* mutants reveals potential for simplification of lignocellulosic biomass. *Proc. Natl Acad. Sci. USA* **107**, 17 409–17 414. (doi:10.1073/pnas.1005456107)
- 150 Dondini, M., Hastings, A., Saiz, G., Jones, M. B. & Smith, P. 2009 The potential of *Miscanthus* to sequester carbon in soils: comparing field measurements in Carlow, Ireland to model predictions. *Glob. Change Biol. Bioenergy* **1**, 413–425. (doi:10.1111/j.1757-1707.2010.01033.x)
- 151 Silver, W. L., Ryals, R. & Eviner, V. 2010 Soil carbon pools in California's annual grassland ecosystems. *Rangeland Ecol. Manage.* **63**, 128–136. (doi:10.2111/REM-D-09-00106.1)
- 152 Malhi, Y., Baldocchi, D. D. & Jarvis, P. G. 1999 The carbon balance of tropical, temperate and boreal forests. *Plant Cell Environ.* **22**, 715–740. (doi:10.1046/j.1365-3040.1999.00453.x)
- 153 Pan, Y. 2011 A large and persistent carbon sink in the world's forests. *Science* **333**, 988–993. (doi:10.1126/science.1201609)
- 154 Le Quéré, C. *et al.* 2009 Trends in the sources and sinks of carbon dioxide. *Nature Geoscience* **2**, 831–836. (doi:10.1038/ngeo689)
- 155 Phillips, O. L. *et al.* 2009 Drought sensitivity of the Amazon rainforest. *Science* **323**, 1344–1347. (doi:10.1126/science.1164033)
- 156 Atkinson, C. J., Fitzgerald, J. D. & Hipps, N. A. 2010 Potential mechanisms for achieving agricultural benefits from biochar application to temperate soils: a review. *Plant Soil* **337**, 1–18. (doi:10.1007/s11104-010-0464-5)
- 157 Sohi, S. P., Krull, E., Lopez-Capel, E. & Bol, R. 2010 A review of biochar and its use and function in soil. *Adv. Agron.* **105**, 47–82. (doi:10.1016/S0065-2113(10)05002-9)
- 158 Woolf, D., Amonette, J. E., Street-Perrott, F. A., Lehmann, J. & Joseph, S. 2010 Sustainable biochar to mitigate global climate change. *Nat. Commun.* **1**, 56.
- 159 Gregory, A. S. *et al.* 2010 Soil management and grass species effects on the hydraulic properties of shrinking soils. *Soil Sci. Soc. Am. J.* **74**, 753–761. (doi:10.2136/sssaj2009.0284)
- 160 Macleod, C. J. A., Binley, A., Hawkins, S. L., Humphreys, M. W., Turner, L. B., Whalley, W. R. & Haygarth, P. M. 2007 Genetically modified hydrographs: what can grass genetics do for temperate catchment hydrology? *Hydrol. Process.* **21**, 2217–2221. (doi:10.1002/hyp.6780)
- 161 Kamoshita, A., Babu, R. C., Boopathi, N. M. & Fukai, S. 2008 Phenotypic and genotypic analysis of drought-resistance traits for development of rice cultivars adapted to rainfed environments. *Field Crops Res.* **109**, 1–23. (doi:10.1016/j.fcr.2008.06.010)
- 162 McKenzie, B. M., Bengough, A. G., Hallett, P. D., Thomas, W. T. B., Forster, B. & McNicol, J. W. 2009 Deep rooting and drought screening of cereal crops: a novel field-based method and its application. *Field Crops Res.* **112**, 165–171. (doi:10.1016/j.fcr.2009.02.012)
- 163 Trachsel, S., Messmer, R., Stamp, P. & Hund, A. 2009 Mapping of QTLs for lateral and axile root growth of tropical maize. *Theor. Appl. Genet.* **119**, 1413–1424. (doi:10.1007/s00122-009-1144-9)
- 164 Levitt, S. D. & Dubner, S. J. 2005 *Freakonomics: a rogue economist explores the hidden side of everything*. London, UK: Allen Lane.
- 165 De Deyn, G. B., Cornelissen, J. H. C. & Bardgett, R. D. 2008 Plant functional traits and soil carbon sequestration in contrasting biomes. *Ecol. Lett.* **11**, 516–531. (doi:10.1111/j.1461-0248.2008.01164.x)
- 166 Dijkstra, F. A., Hobbie, S. E. & Reich, P. B. 2006 Soil processes affected by sixteen grassland species grown under different environmental conditions. *Soil Sci. Soc. Am. J.* **70**, 770–777. (doi:10.2136/sssaj2005.0088)
- 167 Kell, D. B. & Oliver, S. G. 2004 Here is the evidence, now what is the hypothesis? The complementary roles of inductive and hypothesis-driven science in the post-genomic era. *Bioessays* **26**, 99–105. (doi:10.1002/bies.10385)
- 168 Broadhurst, D. & Kell, D. B. 2006 Statistical strategies for avoiding false discoveries in metabolomics and related experiments. *Metabolomics* **2**, 171–196. (doi:10.1007/s11306-006-0037-z)
- 169 Fourcaud, T., Zhang, X., Stokes, A., Lambers, H. & Körner, C. 2008 Plant growth modelling and applications: The increasing importance of plant architecture in growth models. *Ann. Bot.* **101**, 1053–1063. (doi:10.1093/aob/mcn050)