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1 **Title**

2 Large-scale seroepidemiologic surveillance of COVID-19 - Cross-sectional study in Hyogo  
3 prefecture of Japan in August, 2021

4

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26

27 **Abbreviations**

28 COVID-19: Coronavirus disease 2019

29 SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2

30 ECLIA: Electrochemiluminescence immunoassay

31 ELISA: Enzyme-linked immunosorbent assay

32 N: Nucleocapsid

33 S: Spike

34 PBS: phosphate buffered saline

35 COI: cut-off index

**NOTE: This preprint reports new research that has not been certified by peer review and should not be used to guide clinical practice.**

36 ABTS: 2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid

37

38 **Abstract**

39 The situation of the COVID-19 pandemic in Japan is drastically changing in the 2<sup>nd</sup> year, 2021,  
40 due to the appearance of SARS-CoV-2 variants of concern and the roll-out of mass vaccination.  
41 In addition to PCR diagnosis, periodic seroepidemiologic surveillance is important to analyze  
42 the epidemic situation. In this study, we analyzed the rate of seropositivity for the SARS-CoV-2  
43 N and S antigens in Hyogo prefecture, Japan in August 2021. Sera collected from people who  
44 received a health check-up in a clinic of the Hyogo Prefecture Health Promotion Association  
45 were subjected to analysis of reactivity to the SARS-CoV-2 N and S antigens by  
46 electrochemiluminescence immunoassay (ECLIA) and enzyme-linked immunosorbent assay  
47 (ELISA), respectively. For a total 1,000 sera, the positive rates to N and S antigens were 2.1%  
48 and 38.7%, respectively. The infectious rate estimated by serological analysis based on the  
49 presence of the anti-N antibody was 2.5-fold higher than the value reported based on PCR-based  
50 analysis, and it increased five-fold compared to the rate determined by our previous  
51 seroepidemiologic study in October, 2020. The anti-S positive rate was almost consistent with  
52 the vaccination rate in this area. The observed high anti-S antibody level in the seropositive  
53 population may indicate that the mass vaccination in Japan is being performed smoothly at this  
54 time point, although the infectious rate has also increased.

55

56 **Introduction**

57 The COVID-19 pandemic, which first emerged in December 2019, has undergone  
58 several turning points, introducing drastic changes in its progress. One of the most important  
59 factors is the appearance of the SARS-CoV-2 variants of concern (VOCs) replacing the original  
60 variant. In Japan, the Alpha variant (B.1.1.7) replaced the existing strain by around April, 2021;  
61 then, the Delta variant (B.1.617.2) began spreading rapidly throughout the country from July to  
62 August, 2021 (2,16,22). The other critical event has been the launch of anti-COVID-19  
63 vaccines. Accelerated development of several vaccine platforms was based on the components  
64 of the original SARS-CoV-2 strain as the template, and these vaccines were shown to be

65 effective for reducing the COVID-19 outbreak (17, 20). Although the COVID-19 vaccine  
66 rollout in Japan was delayed compared to that in other countries such as Israel and United States  
67 (19), the Comirnaty (BNT162b2 mRNA, BioNTech-Pfizer, Mainz, Germany/New York, United  
68 States) vaccine (25) was first approved in Japan and administered to health-care workers  
69 starting in February 2021, and elderly persons aged  $\geq 65$  years old starting April 2021,  
70 expanding to other populations from May, 2021. The mRNA vaccines developed by Moderna  
71 (7) were also approved and have been widely used in Japan from May 2021. The number of  
72 vaccinated people is increasing rapidly in Japan, as monitored by the government system (1).  
73 On September 13th, the government announced that 50.9% of the population in Japan has  
74 completed vaccination (<https://covid19.who.int/region/wpro/country/jp>). These two factors, that  
75 is, propagation of SARS-CoV-2 VOCs and the progress of the vaccination, may balance each  
76 other out, making predictions of the COVID-19 situation more difficult.

77 In October 2020, we conducted seroepidemiologic surveillance in Hyogo prefecture,  
78 population 5.5 million, located in the southern-central region of Japan, the so-called Kansai area  
79 (11). In our 1st seroepidemiologic surveillance, we reported that 0.15% of 10,377 sera had  
80 neutralizing activity against SARS-CoV-2 infection *in vitro*. The value 0.15% was interpreted to  
81 be the true infectious rate, and it was three times higher than the value (0.05%) estimated based  
82 on the PCR diagnosis at that time point (33), probably because the serological analysis could  
83 also detect persons with histories of asymptomatic infection.

84 In previous COVID-19 seroepidemiologic analyses, both the Nucleocapsid (N) and  
85 Spike (S) antigens of the SARS-CoV-2 were used as markers of infection; reactivity of serum  
86 antibodies against one or two of these antigens was detected (9, 21, 27, 33). However, currently  
87 the anti-N antibody is the major marker because the vaccines used in Japan protect individuals  
88 by inducing anti-S immunities including anti S-antibodies, making it is impossible to distinguish  
89 the S antibodies due to SARS-CoV-2 infection from those induced by vaccination. Nevertheless,  
90 analysis of the anti-S antibodies is of great importance because they are the key immune  
91 components for counteracting SARS-CoV-2 infection, and can serve as a marker of an

92 individual's potential to be protected from the infection (29, 31).

93 Under the current situation where infected and vaccinated people are both prevalent in  
94 society, seroepidemiologic surveillance can be used to accurately measure the proportion of  
95 individuals who have acquired SARS-CoV-2-specific antibodies by infection, vaccination or  
96 both. Seroepidemiologic surveillance is frequently conducted to understand and predict  
97 epidemics (8, 23, 26, 28, 35).

98 In this study, we reported the 2<sup>nd</sup> seroepidemiologic survey of COVID-19 in Hyogo  
99 prefecture from July to August 2021, which overlaps with the period of the 2020 Tokyo  
100 Olympics, delayed one year and held in Japan in the summer of 2021 (30). The anti-N antibody  
101 positivity rate revealed that the estimated infection rate was slightly higher than that reported by  
102 PCR diagnosis, whereas the anti-S antibody positive rate was consistent with the current  
103 vaccination rate in Hyogo prefecture. This report will be a milestone in the era during which the  
104 vaccine and SARS-CoV-2 VOCs are struggling for dominance.

105

## 106 **Materials and methods**

### 107 **Samples**

108 Serum samples were collected from people receiving health check-ups at the clinics of Hyogo  
109 Prefecture Health Promotion Association, Kobe, Japan from 19<sup>th</sup> July to 6<sup>th</sup> August. This  
110 retrospective observational study was explained on the website of Kobe University Hospital  
111 along with the opportunity to opt-out. Each serum was heat-treated at 56°C for 30 min and  
112 stored in 4°C until use.

113

### 114 **Detection of anti-N antibody by electrochemiluminescence immunoassay (ECLIA)**

115 An electrochemiluminescence immunoassay (ECLIA) was conducted using the cobas  
116 e801 module (Roche Diagnostics, Rotkreuz, Switzerland) as in our previous study (11). The  
117 Elecsys Anti-SARS-CoV-2 assay kit (Roche Diagnostics, Rotkreuz, Switzerland) is based on a  
118 double-antigen sandwich assay, which detects antibodies against the SARS-CoV-2 nucleocapsid

119 (N). The measurement was performed according to the manufacturer's instructions, and samples  
120 with a cut-off index (COI) > 1.0 were diagnosed as positive.

121

### 122 **Expression and purification of SARS-CoV-2 spike protein**

123 The Spike protein used for the ELISA assay was prepared by a recombinant  
124 expression system similar to that previously reported by another research group (15). Briefly,  
125 the gene sequence of the SARS-CoV-2 Spike ectodomain (amino acids 1-1213) was subcloned  
126 into a pCAGGS vector (24) with a puromycin-resistant gene. The Spike sequence used here  
127 includes following mutations: D614G, R682del, R683del, R685del, F817P, A892P, A899P,  
128 A942P, K986P, and V987P, and additional sequences: namely, an HRV-3c recognition site, T4  
129 foldon, and a His-tag at the C-terminal side. The sequence was confirmed by the capillary  
130 electrophoresis sequencer DS3000 (Hitachi High-Tech).

131 Spike protein was expressed using the Expi293 expression system (ThermoFisher  
132 Scientific) according to the manufacturer's instruction. The culture supernatant was collected at  
133 four days post transfection. The His-tagged Spike protein was purified by Ni-NTA agarose  
134 (Qiagen) and size exclusion chromatography using the Superose 6 increase column and AKTA  
135 pure system (Cytiva).

136

### 137 **Detection of anti-Spike antibodies by enzyme-linked immunosorbent assay (ELISA)**

138 Anti-Spike antibodies in the human sera were detected by an enzyme-linked  
139 immunosorbent assay with the purified Spike ectodomain described above. Each well of the 96-  
140 well ELISA plate (Corning) was coated with 100 ng of purified Spike protein dissolved in a  
141 carbonate buffer at 4°C overnight. After washing the plate with PBS containing 0.1% Tween 20  
142 (PBST) to remove the antigen, PBST supplemented with 1% bovine serum albumin was added  
143 as a blocking buffer followed by incubation at 4°C for 2 hours. Individual sera samples serially  
144 diluted from 1:40 to 1:5120 in the blocking buffer were added to the antigen-coated plate, and  
145 then incubated at 37°C for 1 hour. After washing with PBST, a goat anti-human IgG with

146 conjugated horseradish peroxidase (abcam) diluted 1:10000 with PBST was added as a  
147 secondary antibody followed by incubation at 37°C for 1 hour. After washing with PBST, 100 µl  
148 per well of ABTS solution (Roche) was added as substrate, and the plate was incubated at room  
149 temperature for 40 minutes in the dark. The reaction was stopped by adding 100 µl of 1.5%  
150 (w/v) oxalic acid dehydrate solution. The optical density at wavelength 405 nm ( $OD_{405}$ ) was  
151 measured using the plate reader Multiskan FC (Thermofisher Scientific). Based on a preliminary  
152 experiment using sera from healthy volunteers (average 0.14; standard deviation: 0.022, n=12),  
153 we set a cut-off value of 0.3 for the 1:40 dilution to define positivity, with the aim of avoiding  
154 false positive detection. The value of the area under the curve (AUC) was used to evaluate the  
155 anti-S antibody amount (6, 12). The AUCs were calculated for the plot of  $OD_{405}$  values  
156 according to dilution factors, and an arbitrary value of 1 was given as the width for a two-fold  
157 dilution step.

158

### 159 **Ethics statement**

160 This study was approved by the ethical committees of Kobe University Graduate School of  
161 Medicine (approval codes: B2156702). Hyogo Prefecture Health Promotion Association was  
162 also granted approval under the ethical committee of Kobe University Graduate School of  
163 Medicine. Information about this retrospective observational study was published on the  
164 website of Kobe University Hospital, along with the opportunity to opt out. To validate the  
165 ELISA, sera from COVID-19 patients and healthy volunteers were used under approval of the  
166 ethical committee of Kobe University Graduate School of Medicine (approval code B200200),  
167 and written informed consent was obtained from the donors.

168

### 169 **Results**

#### 170 **Summary of the samples**

171 We collected sera from 1,000 persons who received a health check-up at a clinic of the  
172 Hyogo Prefecture Health Promotion Association in Hyogo prefecture. A summary of the



173 samples is shown in Table 1. The gender ratio was nearly equal: males, 58.7%; females, 41.3%.  
174 The age distribution was widely spread with a median of 48 (Table 1 and Figure 1A). The  
175 history of vaccination and infection were not included as selection or exclusion criteria; thus, no  
176 prior information was referred to in this study.

177

### 178 **Reactivity of sera against the SARS-CoV-2 N and S antigens**

179 The SARS-CoV-2 anti-N antibodies were detected by the electrochemiluminescence  
180 immunoassay (ECLIA) method (10, 11), and 21 of the 1,000 samples (2.1%) were deemed  
181 positive given the cut-off of 1.0 (Table 2). The age distribution of positive cases is shown in  
182 Figure 1B and Table 2. The positive rate for the age groups 30-39 yrs and 40-49 yrs were  
183 especially high, at 3.4% (6/179) and 4.1% (10/243), respectively. The positive rates for males  
184 and females were 2.7% (16/587) and 1.2% (5/413), respectively.

185 The anti-S antibodies were detected by the enzyme-linked immunosorbent assay  
186 (ELISA) method using a purified SARS-CoV-2 Spike protein. As a result, 38.7% (387/1,000)  
187 were positive given the cut-off of 0.3 (See the Materials and Methods). The age distribution of  
188 anti-S positive cases is shown in Figure 1C and Table 2. The positive rate for those in their 60s  
189 and 70s showed an apparently high rate, 70.7% (123/174) and 90.2% (37/41), respectively. On  
190 the other hand, the anti-S positive rates for younger age groups were relatively low; 18.9%  
191 (24/127), 31.3% (56/179), 27.6% (67/243), and 33.9% (80/236), for the age groups  $\leq 29$  y, 30-39  
192 y, 40-49 y, and 50-59 y, respectively.

193 The correlation of measured scores for anti-N and anti-S antibodies was analyzed in  
194 N- positive cases and is shown in Figure 2. The Spearman's correlation factor was 0.42,  
195 indicating little correlation if any, probably due to the mixed effect of infection and vaccination  
196 for the value of the anti-S antibody. As expected, all the anti-N positive sera also were anti-S  
197 positive with one exception for a serum in which the N antibody detection value was also  
198 relatively low, probably due to a weak immune response

199

## 200 **Distribution of anti-S antibody amount in sera**

201 We further analyzed the distribution of the anti-S positive reactivity of the sera  
202 because it represents the amount of anti-S antibodies related to the neutralizing antibodies  
203 against SARS-CoV-2. The anti-S positive sera were further evaluated by serial dilution.  
204 Representative curves are shown in Figure 3A. The area under the curve (AUC), which  
205 indicates the amount of anti-S antibodies, was calculated for the 387 anti-S positive samples,  
206 and the distribution is plotted in Figure 3B. The median was 13.75, and the higher and lower  
207 quartile of the values were 17.45 and 8.79, respectively. In the same analysis, the AUC values  
208 for the serum of COVID-19 patient and that of a healthy control were 11.56 and 1.66,  
209 respectively. Thus, most of the anti-S-positive sera contained comparable amounts of anti-S  
210 antibodies to that of an actual COVID-19 patient. The anti-S AUC values for the anti-N positive  
211 sera were plotted in Figure 3C by extracting from the data of Figure 3B. The values are widely  
212 distributed within the range of whole AUC values, consistent with the result of Figure 2.

213

## 214 **Discussion**

215 Seroepidemiologic surveillance is a powerful approach to understanding the spread of  
216 infectious diseases in combination with other methods such as PCR diagnosis and antigen tests.  
217 Our surveillance in August 2021 revealed that the anti-N-positive rate, which represents the  
218 SARS-CoV-2 infectious rate, was 2.1%. At the end of the surveillance, the reported infectious  
219 rate based on PCR diagnosis was 0.85% and 0.80% in the Hyogo prefecture and in Japan,  
220 respectively (33), indicating approximately a 2.5-fold difference compared with the result in this  
221 study. The difference is accounted for by the existence of asymptotically or mildly infected  
222 individuals whose cases may not have been detected because they never underwent testing and  
223 PCR analysis. Indeed, a substantial portion of SARS-CoV-2 infected individuals are  
224 asymptomatic (4), leading to the inaccuracy of surveillance rates determined solely on PCR  
225 diagnosis. Although it is difficult to extrapolate our data directly to the whole population, they  
226 suggest that the infection is more widespread in Japan than the current PCR test results suggest

227 and indicate a need for more systematic testing. Nonetheless, the difference between the  
228 seroepidemiologic analysis and the PCR diagnosis was not overly large; i.e., the two sets of data  
229 are mutually validating.

230 The infectious rate of 2.1% revealed in this study in August 2021 is 14-fold higher  
231 than the rate 0.15% we reported in our 1st seroepidemiologic analysis in October 2020 (11). The  
232 infectious rate of the 1st surveillance was based on the neutralizing antibody titer; however,  
233 vaccination has made it difficult to analyze the relationship between the infection and the  
234 neutralizing antibody titer of the sera. In contrast to the anti-S antibody, the anti-N antibody is  
235 expected to be induced only by the SARS-CoV-2 infection as long as only S-targeted vaccines  
236 are used (13, 32). The anti-N analysis by the ECLIA method of the 1st surveillance for samples  
237 collected at the same clinic of the Hyogo Prefecture Health Promotion Association was 0.4%  
238 (4/1,000) (11), indicating a 5-fold increase at the same area in these 10 months. This result was  
239 not surprising considering the fact that after the 1<sup>st</sup> surveillance in October 2020, Japan  
240 including Hyogo prefecture experienced two additional waves of COVID-19 infection around  
241 January 2021 and May 2021; currently, we face the so-called fifth wave, which started in July,  
242 2021 (3, 33).

243 As of August, 2021 vaccination in Japan with mRNA vaccines that induce anti-S-  
244 based immunity has been proceeding. As reported by our previous research and others, the  
245 vaccination actually induces neutralizing antibody in sera (14, 34), (Furukawa et al., manuscript  
246 submitted). According to the Japanese government system, the vaccination rate in the Hyogo  
247 prefecture as of August 6<sup>th</sup>, 2021 was 32.79% and 42.05% for single- or two-dose vaccinations,  
248 respectively (1). The revealed anti-S positive rate was 38.7% in this study; this result well  
249 represents the current progress in vaccination in Hyogo prefecture, Japan.

250 The observed high anti-S positive rate for the age groups 60-69 yrs and  $\geq 70$  yrs is  
251 explained by the priority vaccination for elderly people (more than 65 years old) in Japan  
252 (Figure 1C). The high anti-S positive rate might be related to the relatively low infection rate in  
253 these age groups (Figure 1B). However, the data for the age groups 50-59 yrs and  $< 29$  yrsr, for

254 which both the anti-S and anti-N positive rates were relatively low, indicated that the low  
255 infectious rate could not be simply explained by the high anti-S positive rate of the age group.  
256 To understand the relatively high infectious rate for the age groups 30-39 yrs and 40-49 yrs  
257 shown in our data (Figure 1B), social reasons such as work commitments and social activities  
258 may need to be taken into consideration.

259 The relationship between the anti-S positive and the anti-N positive rate has become  
260 unclear because of the effect of vaccination. Actually, we could not find a correlation between  
261 the anti-N and the anti-S scores (Figure 2). Although all anti-N-positive sera were also positive  
262 for the anti-S antibody, with one exception, some of the sera showed high anti-S values with  
263 relatively low anti-N scores (Figure 2). Breakthrough infection for vaccinees may induce anti-N  
264 antibodies; however, the titers of the antibodies may be expected to be decreased owing to the  
265 prophylactic immunity induced by vaccination, preventing infection and viral replication.  
266 Actually, Allen et al. has reported that anti-N antibodies were hardly detected by the ECLIA  
267 methods we used in this study in cases of breakthrough infection (5). The cut-off value for the  
268 anti-N assays should be reconsidered in future seroepidemiologic studies in light of the high  
269 vaccination rate that is expected soon in Japan.

270 The COVID-19 situations in Japan and all over the world are rapidly changing day by  
271 day. This study was conducted during the build-up of social immunity by vaccination and the  
272 rapid spread of the SARS-CoV-2 Delta strain in Japan. It will become an important reference for  
273 future studies including our third epidemiological surveillance, which we anticipate will show a  
274 drastically altered situation of COVID-19.

275

#### 276 **Conflict of interest**

277 The authors declare no conflict of interest in this research.

278

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281

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287 Internal Medicine, Kobe University Graduate School of Medicine and Hyogo Prefectural  
288 Kakogawa Medical Center.

289

## 290 **Figure legends**

### 291 **Figure 1**

292 Age distribution of the samples and seropositive rate. (A) Age distribution of the volunteers who  
293 provided the sera. Subjects were divided into six age groups as follows: 29 years or less ( $\leq 29$   
294 years), 30-39 years, 40-49 years, 50-59 years, 60-69 years, 70 years or more ( $\geq 70$  yrs). The anti-  
295 N positive rate (B) and anti-S positive rate (C) were calculated by age groups. The bars were  
296 painted according to the genders. For the anti-N, the measurement was performed once for all,  
297 and confirmed by a repeating measurement for the anti-N positive samples and anti-N negative  
298 samples with relatively high values ( $>0.25$ ). For anti-S, all the positive samples were measured  
299 twice, and samples for which consistent results were obtained were counted as positive.

300

### 301 **Figure 2**

302 Correlation between the observed scores of anti-N and anti-S analysis. For the anti-N analysis,  
303 the cut of index value (COI) recorded by the device was used. For the anti-S analysis, the  
304 observed  $OD_{405}$  value of the ELISA was used.

305

### 306 **Figure 3**

307 Qualitative analysis of the anti-S antibody amount. (A) Three representative curves plotting the

308 observed OD<sub>405</sub> values along the serial serum dilution factors are shown. The serum from a  
309 COVID-19 patient and that from a healthy donor were used as positive and negative controls,  
310 respectively. The areas under the curve were calculated from the individual curves and used as a  
311 reference of the anti-S amount. The calculated AUCs for the curves shown in (A) were as  
312 follows: Sample 1, 20.35; Sample 2, 3.59; Sample 3, 7.87; patient serum, 11.56; and healthy  
313 control: 1.66. (B) The distribution of AUC values from the anti-S ELISA ELISA curves are  
314 shown as a violin plot. The thick horizontal line stands for the median, 13.75, and the thin  
315 horizontal lines indicated the lower and higher quartile values, 8.79 and 17.45, respectively. The  
316 values for the patient sera and sera of healthy controls are indicated as filled and open  
317 arrowheads, respectively. (C) Distribution of anti-S AUC values for the anti-N positive sera. The  
318 exceptional sample determined as anti-S negative is shown as an open circle, and has the lowest  
319 value, as expected.  
320  
321 .

322 **Tables**

323 **Tables 1. Information of the sample numbers and the age distribution**

	All	Sex	
		Male	Female
<b>Sample number, n (%)</b>	1,000 (100%)	587 (58.7%)	413 (41.3%)
<b>Age, yrs, median, (range)</b>	48 (19-83)	51 (20-83)	44 (19-78)
<b>Numbers by age group, n</b>			
≤29 years	127	41	86
30-39 years	179	97	82
40-49 years	243	134	109
50-59 years	236	153	83
60-69 years	174	133	41
≥70 years	41	29	12

324

325 **Tables 2. Numbers of anti-N and anti-S positive samples by age groups**

	All (n=1,000)	Sex	
		Male (n=587)	Female (n=413)
<b>SARS-CoV-2 anti-N antibody (ECLIA)</b>			
<b>Positive No. by age group, n</b>			
≤29 years (n=127)	1	0	1
30-39 years (n=179)	6	5	1
40-49 years (n=243)	10	7	3
50-59 years (n=236)	3	3	0
60-69 years (n=174)	1	1	0
≥70 years (n=41)	0	0	0
<b>Positive No. in all participants, n (%)</b>	21 (2.1%)	16 (2.7%)	5 (1.2%)
<b>SARS-CoV-2 anti-S antibody (ELISA)</b>			
<b>Positive No. by age group, n</b>			
≤29 years (n=127)	24	4	20
30-39 years (n=179)	56	30	26
40-49 years (n=243)	67	28	39
50-59 years (n=236)	80	48	32
60-69 years (n=174)	123	92	31
≥70 years (n=41)	37	26	11
<b>Positive No. in all participants, n (%)</b>	387 (38.7%)	228 (38.8%)	159 (38.5%)

326



327 **References**

- 328 1. Government CIOs' portal, Japan. COVID-19 Vaccination in Japan.  
329 <https://cio.go.jp/en/index.php>, [https://cio.go.jp/c19vaccine\\_dashboard](https://cio.go.jp/c19vaccine_dashboard).
- 330 2. Ministry of Health, Labour and Welfare. [https://www.mhlw.go.jp/stf/covid-](https://www.mhlw.go.jp/stf/covid-19/open-data_english.html)  
331 [19/open-data\\_english.html](https://www.mhlw.go.jp/stf/covid-19/open-data_english.html).
- 332 3. World Health Organization. WHO Coronavirus (COVID-19) Dashboard. .  
333 <https://covid19.who.int/region/wpro/country/jp>. Accessed 15 September,  
334 2021).
- 335 4. **Alene, M., L. Yismaw, M. A. Assemie, D. B. Ketema, B. Mengist, B. Kassie, and**  
336 **T. Y. Birhan.** 2021. Magnitude of asymptomatic COVID-19 cases throughout  
337 the course of infection: A systematic review and meta-analysis. PLoS One  
338 16:e0249090.
- 339 5. **Allen, N., M. Brady, A. I. Carrion Martin, L. Domegan, C. Walsh, L. Doherty,**  
340 **U. N. Riain, C. Bergin, C. Fleming, and N. Conlon.** 2021. Serological markers of  
341 SARS-CoV-2 infection; anti-nucleocapsid antibody positivity may not be the  
342 ideal marker of natural infection in vaccinated individuals. J Infect.
- 343 6. **Amanat, F., D. Stadlbauer, S. Strohmeier, T. H. O. Nguyen, V. Chromikova, M.**  
344 **McMahon, K. Jiang, G. A. Arunkumar, D. Jurczynszak, J. Polanco, M. Bermudez-**

- 345 Gonzalez, G. Kleiner, T. Aydillo, L. Miorin, D. S. Fierer, L. A. Lugo, E. M.
- 346 Kojic, J. Stoeber, S. T. H. Liu, C. Cunningham-Rundles, P. L. Felgner, T.
- 347 Moran, A. Garcia-Sastre, D. Caplivski, A. C. Cheng, K. Kedzierska, O.
- 348 Vapalahti, J. M. Hepojoki, V. Simon, and F. Krammer. 2020. A serological assay  
349 to detect SARS-CoV-2 seroconversion in humans. *Nat Med* **26**:1033-1036.
- 350 7. Baden, L. R., H. M. El Sahly, B. Essink, K. Kotloff, S. Frey, R. Novak, D.
- 351 Diemert, S. A. Spector, N. Roupheal, C. B. Creech, J. McGettigan, S. Khetan, N.
- 352 Segall, J. Solis, A. Brosz, C. Fierro, H. Schwartz, K. Neuzil, L. Corey, P. Gilbert,
- 353 H. Janes, D. Follmann, M. Marovich, J. Mascola, L. Polakowski, J. Ledgerwood,
- 354 B. S. Graham, H. Bennett, R. Pajon, C. Knightly, B. Leav, W. Deng, H. Zhou, S.
- 355 Han, M. Ivarsson, J. Miller, and T. Zaks. 2021. Efficacy and Safety of the  
356 mRNA-1273 SARS-CoV-2 Vaccine. *N Engl J Med* **384**:403-416.
- 357 8. Batool, H., O. Chughtai, M. D. Khan, A. S. Chughtai, S. Ashraf, and M. J. Khan.
- 358 2021. Seroprevalence of COVID-19 IgG antibodies among healthcare workers of  
359 Pakistan: a cross-sectional study assessing exposure to COVID-19 and  
360 identification of high-risk subgroups. *BMJ Open* **11**:e046276.
- 361 9. den Hartog, G., R. M. Schepp, M. Kuijer, C. GeurtsvanKessel, J. van Beek, N.
- 362 Rots, M. P. G. Koopmans, F. R. M. van der Klis, and R. S. van Binnendijk. 2020.

- 363 SARS-CoV-2-Specific Antibody Detection for Seroepidemiology: A Multiplex  
364 Analysis Approach Accounting for Accurate Seroprevalence. *J Infect Dis*  
365 **222**:1452-1461.
- 366 10. **Egger, M., C. Bundschuh, K. Wiesinger, C. Gabriel, M. Clodi, T. Mueller, and**  
367 **B. Dieplinger.** 2020. Comparison of the Elecsys(R) Anti-SARS-CoV-2  
368 immunoassay with the EDI enzyme linked immunosorbent assays for the  
369 detection of SARS-CoV-2 antibodies in human plasma. *Clin Chim Acta* **509**:18-  
370 21.
- 371 11. **Furukawa, K., J. Arie, M. Nishimura, L. H. Tjan, A. Lystia Poetranto, Z. Ren, S.**  
372 **Aktar, J. R. Huang, S. Sutandhio, Y. Kurahashi, A. Nishino, S. Shigekuni, Y.**  
373 **Takeda, K. Uto, K. Matsui, I. Sato, Y. Inui, K. Endo, Y. Kosaka, T. Oota, J.**  
374 **Saegusa, and Y. Mori.** 2021. Seroepidemiological Survey of the Antibody for  
375 Severe Acute Respiratory Syndrome Coronavirus 2 with Neutralizing Activity at  
376 Hospitals: A Cross-sectional Study in Hyogo Prefecture, Japan. *JMA J* **4**:41-49.
- 377 12. **Gaebler, C., Z. Wang, J. C. C. Lorenzi, F. Muecksch, S. Finkin, M. Tokuyama,**  
378 **A. Cho, M. Jankovic, D. Schaefer-Babajew, T. Y. Oliveira, M. Cipolla, C. Viant,**  
379 **C. O. Barnes, Y. Bram, G. Breton, T. Hagglof, P. Mendoza, A. Hurley, M.**  
380 **Turroja, K. Gordon, K. G. Millard, V. Ramos, F. Schmidt, Y. Weisblum, D. Jha,**

- 381 M. Tankelevich, G. Martinez-Delgado, J. Yee, R. Patel, J. Dizon, C. Unson-  
382 O'Brien, I. Shimeliovich, D. F. Robbani, Z. Zhao, A. Gazumyan, R. E. Schwartz,  
383 T. Hatzioannou, P. J. Bjorkman, S. Mehandru, P. D. Bieniasz, M. Caskey, and  
384 M. C. Nussenzweig. 2021. Evolution of antibody immunity to SARS-CoV-2.  
385 Nature 591:639-644.
- 386 13. Harris, R. J., H. J. Whitaker, N. J. Andrews, F. Aiano, Z. Amin-Chowdhury, J.  
387 Flood, R. Borrow, E. Linley, S. Ahmad, L. Stapley, B. Hallis, G. Amirthalingam,  
388 K. Hoschler, B. Parker, A. Horsley, T. J. G. Brooks, K. E. Brown, M. E. Ramsay,  
389 and S. N. Ladhani. 2021. Serological surveillance of SARS-CoV-2: Six-month  
390 trends and antibody response in a cohort of public health workers. J Infect  
391 82:162-169.
- 392 14. Hirotsu, Y., K. Amemiya, H. Sugiura, M. Shinohara, M. Takatori, H. Mochizuki,  
393 and M. Omata. 2021. Robust Antibody Responses to the BNT162b2 mRNA  
394 Vaccine Occur Within a Week After the First Dose in Previously Infected  
395 Individuals and After the Second Dose in Uninfected Individuals. Front  
396 Immunol 12:722766.
- 397 15. Hsieh, C. L., J. A. Goldsmith, J. M. Schaub, A. M. DiVenere, H. C. Kuo, K.  
398 Javanmardi, K. C. Le, D. Wrapp, A. G. Lee, Y. Liu, C. W. Chou, P. O. Byrne, C.

- 399           **K. Hjorth, N. V. Johnson, J. Ludes-Meyers, A. W. Nguyen, J. Park, N. Wang, D.**  
400           **Amengor, J. J. Lavinder, G. C. Ippolito, J. A. Maynard, I. J. Finkelstein, and J. S.**  
401           **McLellan.** 2020. Structure-based design of prefusion-stabilized SARS-CoV-2  
402           spikes. *Science* **369**:1501-1505.
- 403    16.   **Ito, K., C. Piantham, and H. Nishiura.** 2021. Predicted dominance of variant  
404           Delta of SARS-CoV-2 before Tokyo Olympic Games, Japan, July 2021. *Euro*  
405           *Surveill* **26**.
- 406    17.   **Izda, V., M. A. Jeffries, and A. H. Sawalha.** 2021. COVID-19: A review of  
407           therapeutic strategies and vaccine candidates. *Clin Immunol* **222**:108634.
- 408    18.   **Izumo, T., N. Kuse, N. Awano, M. Tone, K. Sakamoto, K. Takada, Y. Muto, K.**  
409           **Fujimoto, A. Saiki, Y. Ito, H. Matsumoto, and M. Inomata.** 2021. Side effects  
410           and antibody titer transition of the BNT162b2 messenger ribonucleic acid  
411           coronavirus disease 2019 vaccine in Japan. *Respir Investig* **59**:635-642.
- 412    19.   **Kosaka, M., T. Hashimoto, A. Ozaki, T. Tanimoto, and M. Kami.** 2021. Delayed  
413           COVID-19 vaccine roll-out in Japan. *Lancet* **397**:2334-2335.
- 414    20.   **Lopez Bernal, J., N. Andrews, C. Gower, C. Robertson, J. Stowe, E. Tessier, R.**  
415           **Simmons, S. Cottrell, R. Roberts, M. O'Doherty, K. Brown, C. Cameron, D.**  
416           **Stockton, J. McMenemy, and M. Ramsay.** 2021. Effectiveness of the Pfizer-

- 417 BioNTech and Oxford-AstraZeneca vaccines on covid-19 related symptoms,  
418 hospital admissions, and mortality in older adults in England: test negative case-  
419 control study. *BMJ* **373**:n1088.
- 420 21. **Megasari, N. L. A., T. Utsumi, L. N. Yamani, Juniastuti, E. Gunawan, K.**  
421 **Furukawa, M. Nishimura, M. I. Lusida, and Y. Mori.** 2021. Seroepidemiological  
422 study of SARS-CoV-2 infection in East Java, Indonesia. *PLoS One* **16**:e0251234.
- 423 22. **Murayama, H., T. Kayano, and H. Nishiura.** 2021. Estimating COVID-19 cases  
424 infected with the variant alpha (VOC 202012/01): an analysis of screening data  
425 in Tokyo, January-March 2021. *Theor Biol Med Model* **18**:13.
- 426 23. **Naesens, R., H. Mertes, J. Clukers, S. Herzog, C. Brands, P. Vets, I. De Laet, P.**  
427 **Bruynseels, P. De Schouwer, S. van der Maas, K. Bervoets, N. Hens, and P. Van**  
428 **Damme.** 2021. SARS-CoV-2 seroprevalence survey among health care providers  
429 in a Belgian public multiple-site hospital. *Epidemiol Infect* **149**:e172.
- 430 24. **Niwa, H., K. Yamamura, and J. Miyazaki.** 1991. Efficient selection for high-  
431 expression transfectants with a novel eukaryotic vector. *Gene* **108**:193-9.
- 432 25. **Polack, F. P., S. J. Thomas, N. Kitchin, J. Absalon, A. Gurtman, S. Lockhart, J.**  
433 **L. Perez, G. Perez Marc, E. D. Moreira, C. Zerbini, R. Bailey, K. A. Swanson, S.**  
434 **Roychoudhury, K. Koury, P. Li, W. V. Kalina, D. Cooper, R. W. Frenck, Jr., L. L.**

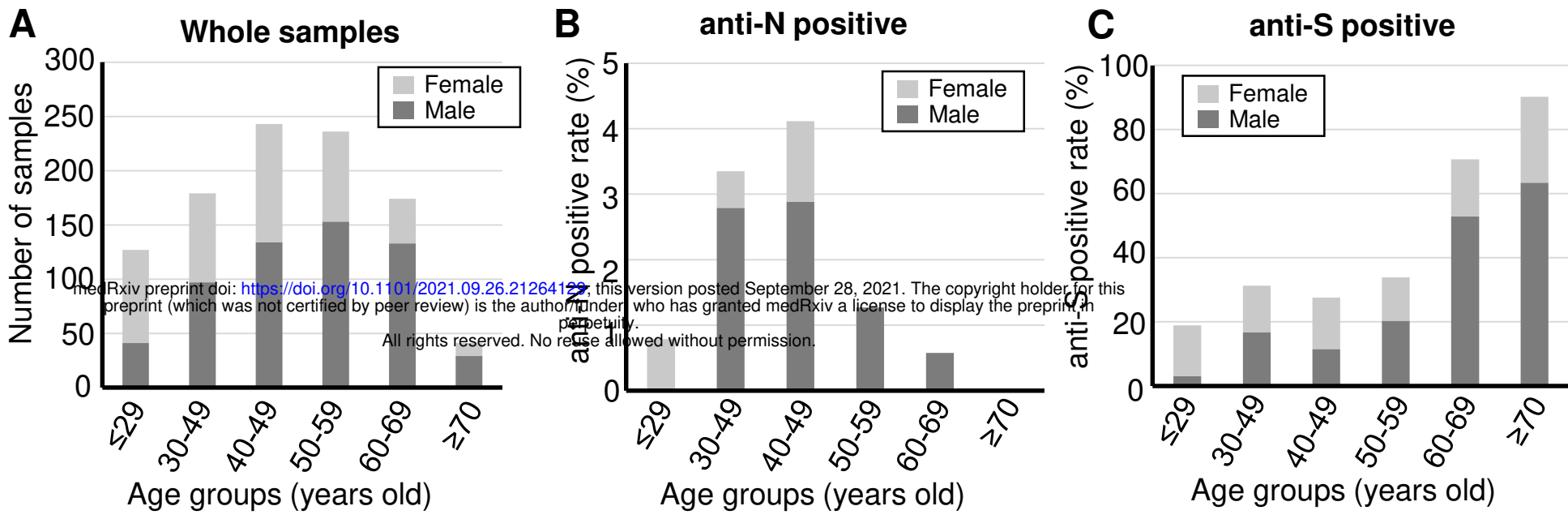
- 435 **Hammitt, O. Tureci, H. Nell, A. Schaefer, S. Unal, D. B. Tresnan, S. Mather, P.**  
436 **R. Dormitzer, U. Sahin, K. U. Jansen, and W. C. Gruber.** 2020. Safety and  
437 Efficacy of the BNT162b2 mRNA Covid-19 Vaccine. *N Engl J Med* **383**:2603-  
438 2615.
- 439 26. **Santosh, C. S., D. P. Tukaram, K. Maharudra, A. Balkrishna, G. Chinmay, I.**  
440 **Aniket, K. Kinge, and J. Nilam.** 2021. Seroprevalence of SARS-CoV-2  
441 Antibodies and Associated Factors in Health Care Workers. *J Assoc Physicians*  
442 *India* **69**:11-12.
- 443 27. **Selvaraju, S., M. S. Kumar, J. W. V. Thangaraj, T. Bhatnagar, V. Saravanakumar,**  
444 **C. P. G. Kumar, K. Sekar, E. Ilayaperumal, R. Sabarinathan, M. Jagadeesan, M.**  
445 **S. Hemalatha, and M. V. Murhekar.** 2021. Population-Based Serosurvey for  
446 Severe Acute Respiratory Syndrome Coronavirus 2 Transmission, Chennai,  
447 India. *Emerg Infect Dis* **27**:586-589.
- 448 28. **Smigelskas, K., K. Petrikonis, V. Kasiulevicius, R. Kalediene, A. Jakaitiene, S.**  
449 **Kaseliene, S. Sauliune, A. Berzanskyte, and M. Stankunas.** 2021. SARS-CoV-2  
450 Seroprevalence in Lithuania: Results of National Population Survey. *Acta Med*  
451 *Litu* **28**:48-58.
- 452 29. **Steensels, D., N. Pierlet, J. Penders, D. Mesotten, and L. Heylen.** 2021.

- 453 Comparison of SARS-CoV-2 Antibody Response Following Vaccination With  
454 BNT162b2 and mRNA-1273. JAMA.
- 455 30. Tokuda, Y., and T. Kuniya. 2021. Prediction of COVID-19 cases during Tokyo's  
456 Olympic and Paralympic Games. *J Gen Fam Med* **22**:171-172.
- 457 31. Wang, Z., F. Schmidt, Y. Weisblum, F. Muecksch, C. O. Barnes, S. Finkin, D.  
458 Schaefer-Babajew, M. Cipolla, C. Gaebler, J. A. Lieberman, T. Y. Oliveira, Z.  
459 Yang, M. E. Abernathy, K. E. Huey-Tubman, A. Hurley, M. Turroja, K. A. West,  
460 K. Gordon, K. G. Millard, V. Ramos, J. Da Silva, J. Xu, R. A. Colbert, R. Patel, J.  
461 Dizon, C. Unson-O'Brien, I. Shimeliovich, A. Gazumyan, M. Caskey, P. J.  
462 Bjorkman, R. Casellas, T. Hatziioannou, P. D. Bieniasz, and M. C. Nussenzweig.  
463 2021. mRNA vaccine-elicited antibodies to SARS-CoV-2 and circulating  
464 variants. *Nature* **592**:616-622.
- 465 32. Whitaker, H. J., S. Elgohari, C. Rowe, A. D. Otter, T. Brooks, E. Linley, I.  
466 Hayden, S. Ribeiro, J. Hewson, A. Lakhani, E. Clarke, C. Tsang, C. N. Campbell,  
467 M. Ramsay, K. Brown, and G. Amirthalingam. 2021. Impact of COVID-19  
468 vaccination program on seroprevalence in blood donors in England, 2021. *J*  
469 *Infect* **83**:237-279.
- 470 33. Yazaki, S., T. Yoshida, Y. Kojima, S. Yagishita, H. Nakahama, K. Okinaka, H.

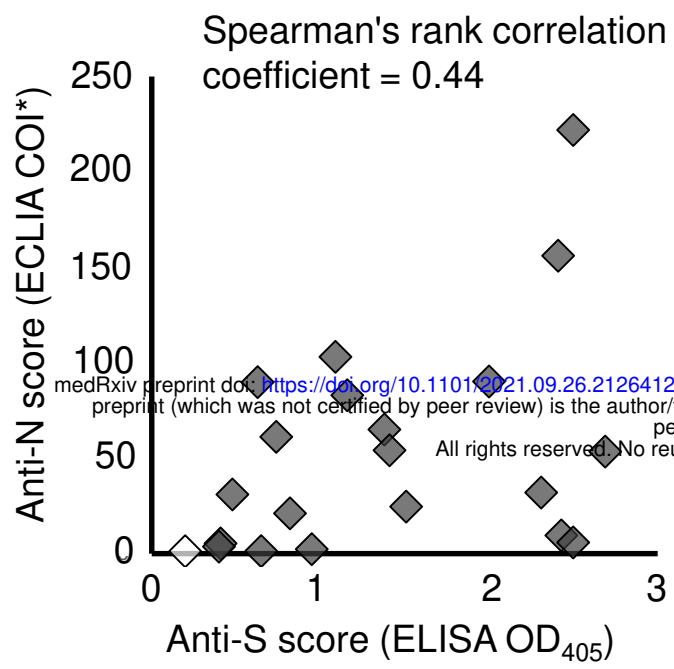


- 471           **Matsushita, M. Shiotsuka, O. Kobayashi, S. Iwata, Y. Narita, A. Ohba, M.**
- 472           **Takahashi, S. Iwasa, K. Kobayashi, Y. Ohe, A. Hamada, T. Doi, and N.**
- 473           **Yamamoto.** 2021. Difference in SARS-CoV-2 Antibody Status Between Patients
- 474           With Cancer and Health Care Workers During the COVID-19 Pandemic in
- 475           Japan. *JAMA Oncol* 7:1141-1148.
- 476    34.   **Yoshimura, Y., H. Sasaki, N. Miyata, K. Miyazaki, and N. Tachikawa.** 2021.
- 477           Antibody response after COVID-19 vaccine BNT162b2 on health care workers
- 478           in Japan. *J Infect Chemother.*
- 479    35.   **Zurcher, K., C. Mugglin, F. Suter-Riniker, P. M. Keller, M. Egger, S. Muller, M.**
- 480           **Fluri, M. Hoffmann, and L. Fenner.** 2021. Seroprevalence of SARS-CoV-2 in
- 481           healthcare workers from outpatient facilities and retirement or nursing homes in
- 482           a Swiss canton. *Swiss Med Wkly* 151.
- 483
- 484

**Figure 1**

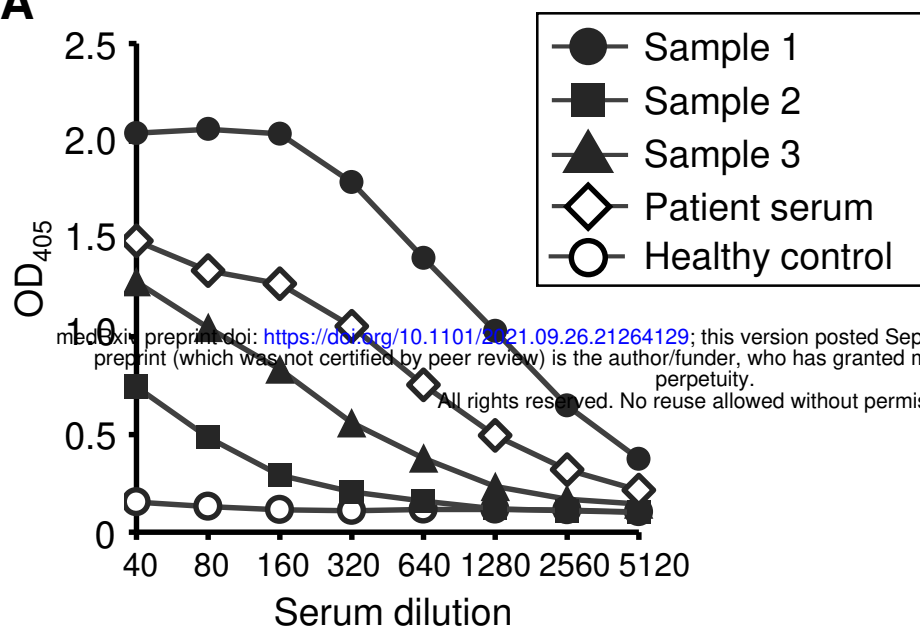


**Figure 2**

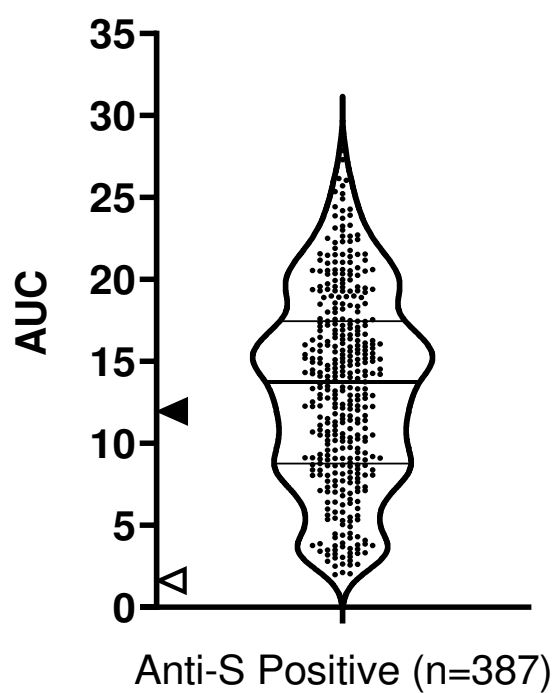


**Figure 3**

**A**



**B**



**C**

