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Laser microfluidics: fluid actuation by light

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Abstract

The development of microfluidic devices is still hindered by the lack of robust fundamental building blocks that constitute any fluidic system. An attractive approach is optical actuation because light field interaction is contactless and dynamically reconfigurable, and solutions have been anticipated through the use of optical forces to manipulate microparticles in flows. Following the concept of an 'optical chip' advanced from the optical actuation of suspensions, we propose in this survey new routes to extend this concept to microfluidic two-phase flows. First, we investigate the destabilization of fluid interfaces by the optical radiation pressure and the formation of liquid jets. We analyze the droplet shedding from the jet tip and the continuous transport in laser-sustained liquid channels. In the second part, we investigate a dissipative light-flow interaction mechanism consisting in heating locally two immiscible fluids to produce thermocapillary stresses along their interface. This opto-capillary coupling is implemented in adequate microchannel geometries to manipulate two-phase flows and propose a contactless optical toolbox including valves, droplet sorters and switches, droplet dividers or droplet mergers. Finally, we discuss radiation pressure and opto-capillary effects in the context of the 'optical chip' where flows, channels and operating functions would all be performed optically on the same device.

Keywords: optical radiation pressure, thermocapillary effects, microfluidics

(Some figures in this article are in colour only in the electronic version)

1. Introduction

Fully integrated microfluidic laboratories [1], operating at nanoliter fluid volumes, are now recognized to have the potential for managing high-throughput massively multiplexed arrays of chemical microreactors or biological assays [2] by significantly reducing the material consumption, increasing data collecting rates and allowing many different operations to be processed in serial or in parallel. Conversely, recent advances in the manipulation of colloidal particles within microfluidic devices [3, 4] have shown that optical forces may represent a new and promising route towards microactuation. Indeed, beyond the fact that light fields can easily be focused to micrometric spots, they are contactless, dynamically reconfigurable and do not impose any dedicated microfabrication. Contrary to classical total micro-analysis systems, which are generally constrained to perform predesigned tasks, this optical flexibility proved to be very powerful in manipulating suspensions in a single fluid phase

via optical forces, including pumping [5–7], sorting [8–11], conveying [12] and propelling [13, 14]. Parallel actuation was also demonstrated [15]. Moreover, this combination of optics and microfluidics, called optofluidics [16-18], also became an emergent research field offering an unprecedented level of integration for building a new generation of optically actuated microdevices, merging optical reconfigurability, the softness of fluid, smoothness of fluid interfaces and compactness. Along this line, great achievements were realized in optical switching [19] and splitting [20], adaptive lensing [21, 22], interferometry [23] and lasing [24]. Then, using all these new ideas, we can address the following question: are there routes to build a 'total-optical-lab-on-a-chip', where flows, channels and operating functions would all be performed optically on the same device? Glückstad [25] already advanced the concept of an 'optical chip', thinking of a chip where a suspension would be entirely actuated by light. The present survey aims at extending this concept to the optical manipulation of fluids and digital microfluidics, i.e. droplet microfluidics. It is indeed appealing to investigate how far light can (or cannot) be used to build channels, induce flows, drive droplet formation and finally actuate droplets in channels.

Up to now, two types of devices have emerged for the manipulation of drops at a microscale: open substrate and closed microchannels. In the first case, droplets are formed and manipulated on a surface or interface through the modulation of surface stresses by thermal [26], electrical [27], optical [12], photoresponsive [28] or chemical means [29]. Heat carried by a focused laser beam was also used to move droplets [30] or continuous fluid interfaces [31] on liquid or solid substrates, thus simplifying microfabrication since actuators are not embedded in the substrate anymore. This approach nevertheless suffers a major drawback for application purposes: since devices are open, reliability requires a well-controlled environment to prevent crosscontamination between the droplet and the atmosphere. Manipulation on solid substrates also leads to limitations in droplet velocity [32]. Conversely, in microchannels, drops are formed and transported in a closed chamber using a carrier fluid, usually oil for wetting purposes. In this case droplets are classically in the nanoliter range. A list of fundamental necessary operations was recently enumerated [33], including the need to block, merge, divide or sort droplets individually. Many solutions were proposed to divide or merge drops, to mix their contents and demonstrate their use as microreactors [34, 35]. However, due to the robustness of the environment of a microchannel, actuation functions are usually predetermined by the channel geometry and almost no further control is possible on individual droplets. Thus, a general and reliable solution for actively controlling droplets in microchannels still remains unavailable. The implementation of these functions would open the way to more complex operations being performed which may then be combined into a droplet-based lab-on-a-chip. As illustrated above by the accurate manipulation of suspensions by lasers, an attractive route could be optical actuation.

The extension of particle manipulation to multiphase flows in bulk is nevertheless at a very early stage because bulk

and surface optical forces are usually too weak to counteract hydrodynamic forces acting on fast moving droplets. The deformation of fluid interfaces by the optical radiation pressure is weak as well because interfacial tension generally behaves as a very efficient stabilizing effect. However, since droplet generation always involves fluid interfaces and since emulsification in microchannels always involves a surfactant, it becomes necessary to re-investigate this coupling between liquid interface and laser waves. To do so and decrease the relative importance of interfacial tension versus optical radiation pressure, we considered a model system and performed experimental investigations on the meniscus of phase-separated liquid mixtures close to a liquid-liquid critical point. Indeed, near-critical interfaces become highly deformable because interfacial tension vanishes close to a critical point. In these conditions, huge stationary interface deformations of several hundreds of microns can be generated with low beam power c.w. lasers. The induced hump can become unstable as well at sufficiently large beam intensities and lead to the formation of stationary beam-centered liquid micro-jets emitting droplets, anticipating the basis of our totaloptical approach of microfluidics. However, as far as flowing microdroplets are used as tight fluid reactors or carriers, new fluid-handling micro-technologies are also required to convey, synchronize and more generally to manipulate accurately these droplets within a microchannel. To overcome optical radiation pressure limitations, we used instead the thermocapillary (or Marangoni) effect [36] induced by a laser. By locally heating the drop's surface, an interfacial tension gradient is set up, creating an imbalance in the surface stresses acting on the drop. In the low Reynolds number regime of microfluidics, this affects the bulk flow very rapidly and allows for the implementation of an optical toolbox for total control of digital microfluidics.

This paper is organized into three main parts. After a brief survey on optical manipulation of liquid interfaces, we show in section 2 how a laser wave can couple to a liquid interface and drive its bending and how this bending can become unstable and trigger a liquid jet emitting microdroplets continuously. We investigate this liquid flow and demonstrate that it originates from bulk radiation pressure effects. Finally, we show how this 'opto-hydrodynamic' instability can be used to form and stabilize large aspect ratio liquid channels to carry liquids. In section 3, we switch to the optical actuation of droplets by thermocapillary forces in microchannels and demonstrate its potential to create various drop-handling functions. In section 4, an attempt at combining radiation pressure and thermocapillary effects is discussed in order to dream about the 'total-optical-labon-a-chip' concept for further appealing investigations of contactless digital microfluidics.

2. Optical manipulation of fluid interfaces and light-induced jetting

2.1. Mechanical aspects

The first experiment on deformation of a liquid interface by the radiation pressure of a laser wave [37] is contemporary to

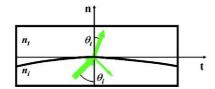


Figure 1. Schematic representation used for the calculation of the radiation pressure exerted by a laser beam on an interface separating two dielectric fluids characterized by the indices of refraction n_i and n_t . The angle of incidence and transmission are, respectively, θ_i and θ_t . **t** and **n** are the tangent and the normal directions at the location where the beam impinges the interface.

the optical levitation demonstration (they were both performed by the same authors). Considering some earlier theoretical developments [38], the first motivation of Ashkin and Dziedzic was to determine in which direction the bending of a fluid interface separating two dielectric liquids of different refractive indices occurs. Their result was sufficiently non-intuitive to briefly summarize here.

Let us consider two dielectric media, of different refractive indices $n_{i,t}$, separated by an interface (figure 1). The subscripts i and t refer to incidence and transmission and θ_i and θ_t are, respectively, the incident and the transmitted angles. Since the momentum carried by the incident propagating light depends on the refractive index, it is not conserved when the beam travels from one to the other dielectric and the resulting discontinuity gives birth to a radiation pressure applied to the interface. Considering the Minkowski formalism [39], its expression is given by

$$\Pi_{\text{Rad}} = n_{\text{i}} \cos^2 \theta_{\text{i}} \left[1 + R(\theta_{\text{i}}, \theta_{\text{t}}) - \frac{\tan \theta_{\text{i}}}{\tan \theta_{\text{t}}} T(\theta_{\text{i}}, \theta_{\text{t}}) \right] \frac{I}{c} \mathbf{n}, \quad (1)$$

where I is the beam intensity, and $R(\theta_i, \theta_t)$ and $T(\theta_i, \theta_t) = 1 - 1 - 1$ $R(\theta_i, \theta_t)$ are the classical reflection and transmission Fresnel coefficients in electromagnetic energy. Consequently, Π_{Rad} is always normal to the interface. Moreover, by considering the expressions of the reflection and transmission coefficients $R(\theta_i, \theta_t)$ and $T(\theta_i, \theta_t)$ [40], it can be easily shown that the optical radiation force Π_{Rad} is always directed toward the dielectric medium of the lowest index of refraction. This means that interface bending does not depend on beam propagation, and the main reason for that is that a photon gains momentum when passing from a low to a large refractive index medium. Ashkin and Dziedzic [37] verified this remarkable property, which was further generalized to the liquid–liquid case [41]. A quantitative interpretation was nevertheless difficult due to the spatial and temporal profiles of the laser pulses used [42]. For instance, Brevik [43] had to consider a larger beam waist and to use a time delay to retrieve the temporal behavior of the induced lens. In fact, very few results related to the deformation of fluid interfaces by the optical radiation forces have been published. Except for a brief letter [44], we are aware of a second set of experiments realized at the liquid/air interface [45] using a ruby laser. As interface bending was still very weak, detection was performed holographically.

Nevertheless, the emergence of the so-called soft matter physics in the 1980s brought a sort of renewal of laser

radiation pressure by opening new horizons, particularly in the metrology of soft interfaces. For instance, a Japanese team exploited the bending of an interface by a laser wave to measure interfacial tensions [46] in a non-contact manner. In a second work [47], they focused their attention on the difficult case of ultra-low interfacial tension, for which most classical techniques fail. Finally, they showed the pertinence of radiation pressure to measure shear viscosities [48]. The characterization of the mechanical properties of lipidic membranes is also of fundamental interest in biophysics. Since optical dynamometry is limited to large vesicles, Wang's group used radiation pressure to quantitatively measure curvature rigidity of small vesicles [49] and in vivo cells [50] at a nanometric length scale. Compared to optical dynamometry using a beam-trapped bead as a probe [51], these experiments show that the main interests in using radiation pressure are the strong localization of excitation as well as its contactless character. In the meantime, Käs' group patented a new tool, called 'Optical Stretcher', to probe the elasticity of red blood cells using radiation pressure [52]. Due to its sensitivity, one of the ultimate goals of this technique is to differentiate healthy from malignant cells from their difference in elastic response [53, 54].

Conversely, Zhang and Chang [55] performed the first direct observations of laser-induced nonlinear interface deformation on spherical water droplets using a high speed camera. After the passage of a 100 mJ pulse, a first series of images showed that interface deformation is directed toward the air on both sides of the drop, with a small asymmetry on the exit face as a result of the intensity increase due to the ball lens behavior of the drop. A quantitative interpretation of this weakly nonlinear regime of deformation was published by Poon and coworkers [56] and further developed by Brevik and Kluge [57]. Even more surprisingly, Zhang and Chang showed that 200 mJ pulses lead to droplet disruption and the formation of a long liquid filament on the exit face. Note that nonlinear interface deformations were also observed on vesicles using c.w. lasers [53].

2.2. Principles of the experiment

As discussed above, deformations induced by a continuous laser beam on classical interfaces remain weak. Thus, direct visualization of deformations induced by a c.w. laser obviously requires very soft interfaces. We thus choose to work with near-critical interfaces because the interfacial tension σ vanishes close to a critical point as $\sigma = \sigma_0 |T/T_{\rm C} - 1|^{1.26}$, where T and $T_{\rm C}$ are, respectively, the actual and critical temperatures. Moreover, given a temperature shift to criticality, the use of supramolecular liquids leads to even smaller interfacial tension. This is due to the fact that the amplitude σ_0 is inversely proportional to the square of the characteristic molecular length scale ξ_0 according to a universal ratio R^+ , defined as $R^+ = (\sigma_0 \xi_0^2 / k_B T_C) = 0.39$ [58] resulting from renormalization group theory. Consequently, in order to enhance radiation pressure effects on fluid interfaces, we performed experiments in a near-critical phase-separated micellar phase of microemulsions (assembly of surfactantcoated water nanodroplets suspended in oil) [41]. As they

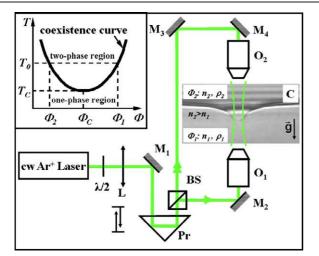


Figure 2. (Adapted from [60]): experimental set-up. $\lambda/2$: half-wave plate, L: lens, $M_{i=1,4}$: mirrors, Pr: prism; BS: beamsplitter; $O_{i=1,2}$ microscope objectives $10 \times .$ C denotes the experimental cell in which an example of a deformed near-critical interface separating the phases Φ_1 and Φ_2 for $T_0 > T_C$ is presented. Inset: schematic phase diagram of the micellar phase of microemulsion used. *T* is temperature and Φ is the volume fraction of micelles.

belong to the same universality class (d = 3, n = 1) of the Ising model, like all isotropic liquids, results obtained from our 'exotic' choice of fluid can be transposed to any interface separating liquids belonging to the same universality class. We used a quaternary mixture of sodium dodecyl sulfate (surfactant), water, toluene (oil) and n-butan-1-ol (alcohol). Its mass composition in weight fraction is 4% sodium dodecyl sulfate, 9% water, 70% toluene and 17% n-butan-1-ol. This composition was chosen so as to be critical at a temperature $T_{\rm C} = 35 \,^{\circ}{\rm C}$ [59]. As illustrated in the inset of figure 2, for a temperature $T > T_{\rm C}$, the mixture phase separates into two micellar phases of different concentrations Φ , called hereafter Φ_1 and Φ_2 . For this composition, the micelle radius is $\xi_0 =$ 40 ± 2 Å, i.e. one order of magnitude larger than the classical molecular length scale, and small enough to get a suspension transparent in the visible optical wavelength window. Due to the amplitude of this ξ_0 value, the interfacial tension is expected to be extremely weak compared to that of liquidvapor interfaces. We found σ to be one million times smaller than that of the water-free surface ($\sigma = 72 \text{ mN m}^{-1}$) at 1.5 K from the critical point. This weakness clearly justifies the reason why we expect large interface deformations using c.w. lasers. As discussed below, other quantities either vanish (density and refractive index contrast) or diverge (correlation length of density fluctuations and osmotic compressibility) close to the critical point.

Finally, the optical absorption of our micellar phases at the wavelength used is $\alpha_a = 3 \times 10^{-4} \text{ cm}^{-1}$. Thus thermal heating and associated side effects do not disturb interface deformation and will be discarded.

The experimental set-up is presented in figure 2. As beam intensity depends on both power and beam radius, the main point is to locate the beam waist ω_0 on the meniscus of the phase-separated mixture and to vary its size. This was done by using the lens L to form a first intermediate waist. The variation of the beam waist ω_0 is performed by moving the prism Pr to increase the optical path between L and the focusing long working distance microscope objectives, either O_1 or O_2 (10×). As the corresponding beam waist position varies within the cell, C is mounted on translation Moreover, our set-up allows for ascending and stages. descending beams by using the half-wave plate $\lambda/2$ and the beamsplitter BS. Interface deformations are induced by the radiation pressure of a continuous TEM₀₀ argon-ion laser (Innova 90, Coherent, working wavelength $\lambda_0 = 0.514 \ \mu m$). Using this procedure accessible values are $\omega_0 = 4.8$ –32.1 μ m (resp. $\omega_0 = 3.5 - 11.3 \ \mu m$) for ascending (resp. descending) beams. Considering weakly focused beams, we assume the beam divergence to be negligible around the focus and write the intensity profile as

$$I(r,z) \approx I(r) = \frac{2P}{\pi\omega_0^2} \exp\left(-\frac{2r^2}{\omega_0^2}\right),$$
 (2)

where *P* is the incident beam power. Strictly speaking, this approximation is correct as far as $\lambda_0 h/(n\pi\omega_0^2) < 1$, where *h* and *n* are, respectively, the height of the deformation and the index of refraction. Observations of deformations are performed transversally using a white light source for illumination and a microscope for imaging on a CCD video camera coupled to a frame grabber. A spectral filter is placed between the microscope and the CCD to eliminate residual laser light scattered by the micellar phases.

2.3. Light-induced jetting

The general expression governing the stationary interface shape h(r) under radiation pressure effects is given in cylindrical coordinates by [41]

$$(\rho_1 - \rho_2) gh(r) - \sigma \kappa(r) = \Pi_{\text{Rad}}(r), \qquad (3)$$

where $\kappa(r)$ is the interface double mean curvature. Equation (3) shows that the height of the deformation results from the balance between radiation pressure $\Pi_{\text{Rad}}(r)$, given by equation (1), and both buoyancy $(\rho_1 - \rho_2)gh(r)$ and Laplace pressure $\Pi_{\text{Laplace}}(r) = -\sigma\kappa(r)$. It leads to

$$(\rho_1 - \rho_2) gh(r) - \frac{\sigma}{r} \frac{\mathrm{d}}{\mathrm{d}r} \left(\frac{rh'(r)}{\sqrt{1 + h'(r)^2}} \right)$$
$$= \frac{n_\mathrm{i} I(r)}{c} \cos^2 \theta_\mathrm{i} \left(1 + R - \frac{\tan \theta_\mathrm{i}}{\tan \theta_\mathrm{t}} T \right). \tag{4}$$

The incidence and transmission angles θ_i and θ_t can be related to the shape of the deformation. For a downward beam, i.e. propagating from the large to the small refractive index medium, they are given by

$$\cos \theta_{\rm i} = \frac{1}{\sqrt{1 + h'(r)^2}},$$

$$\cos \theta_{\rm t} = \sqrt{1 - \left(\frac{n_2}{n_1}\right)^2 \frac{h'(r)^2}{1 + h'(r)^2}}.$$
(5)

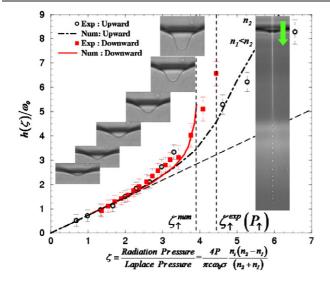


Figure 3. (Adapted from [62]): experimental and numerical evolution of the deformation height $h(r = 0)/\omega_0$ versus the rescaled downward and upward radiation pressure ζ for $(T - T_C) = 3$ K and $\omega_0 = 5.3 \ \mu\text{m}$. The dashed line indicates the linear regime in deformation and the measured threshold P_{\uparrow} , above which the interface becomes unstable, is indicated by two vertical dashed lines, the experimental and the predicted one. Snapshots show the downward evolution for beam power increasing from left to right as P = 190, 250, 280, 310, 340, 370 and $P_{\uparrow} = 400$ mW for the stationary jet. The total height of right picture is 1 mm.

Note that one can use the expression of R and T at normal incidence for weak deformations, and linearize curvature. In these conditions equation (4) reduces to

$$(\rho_1 - \rho_2) gh(r) - \sigma \Delta_r h(r) = \frac{2n_2}{c} \left(\frac{n_1 - n_2}{n_1 + n_2} \right) I(r).$$
(6)

While for an upward beam we observe a continuous transition from a bell to a nipple-like shape of increasing pedestal [61], the morphology is totally different when the beam propagates downwards, from the large to the small refractive index fluid. As illustrated in figure 3, the deformation height deviates from the linear regime (shown by the dashed line) for increasing incident beam power and diverges at some well-defined beam power threshold P_{\uparrow} . Above this instability threshold a beamcentered stationary liquid jet forms, which emits droplets at its tip.

In fact, interface instability can only be triggered by light if the incident fluid has the largest index of refraction. Indeed, in this case, total internal reflection of light can occur within the deformed interface which in turn increases light intensity and radiation pressure at the tip of the deformation, and so on [63]. Once the jet is formed, droplets are continuously shed at the tip. Since the index of refraction of the droplet is larger than that of the surrounding fluid, the beam automatically traps them. This brings directionality in the emission and transport of droplets that can be actuated by tilting the beam. Moreover, beyond the continuous emission of liquid droplets, the mean jet length can be continuously tuned by increasing beam power [60]. Finally, at fixed $(T - T_C)$ and ω_0 , the droplet flow rate and the drop emission frequency can be controlled by the incident

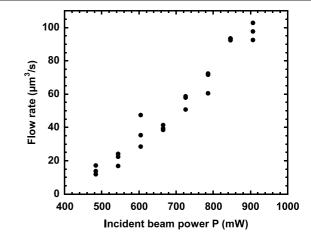


Figure 4. Flow rate variation within the jet versus beam power *P* for $(T - T_c) = 12$ K and $\omega_0 = 4.5 \ \mu$ m.

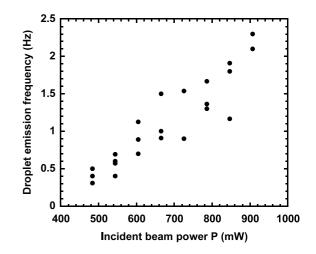


Figure 5. Droplet emission frequency variation at the jet tip versus beam power *P* for $(T - T_c) = 12$ K and $\omega_0 = 4.5 \ \mu$ m.

beam power, as illustrated respectively in figures 4 and 5 for $(T - T_{\rm C}) = 12$ K and $\omega_0 = 4.5 \,\mu$ m. Flow rates of hundreds of μ m³ s⁻¹ are easily achieved with a shedding droplet frequency of several Hz. Consequently, contrary to other methods (flow-mediated: [64, 65] or electrically mediated [66]), this optical approach of microfluidics provides droplets that are directly produced at a chosen micrometric size in a contactless way without further processing. Also, as droplets are produced in three dimensions from a jet size controlled by the beam, neither a particular microfluidic device nor dedicated equipment is required to manage the droplet flow.

2.4. Light-induced liquid channel

Classically, liquid columns of large aspect ratio are not stable. Indeed, beyond a certain aspect ratio, they are known to break up into droplets due to the Rayleigh–Plateau instability [67]. Noticeable is the fact that laser waves are able to stabilize such structures that would be unstable otherwise. This can be easily demonstrated by removing the optical excitation; Rayleigh–Plateau rupture of stationary jets occurs

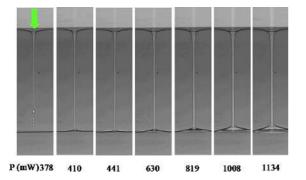


Figure 6. Formation of a liquid channel and increase in its diameter with beam power. Parameters are $\omega_0 = 3.47 \ \mu m$ and $(T - T_{\rm C}) = 4$ K. The liquid channel length is 334 μ m.

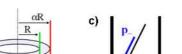
instantaneously. This optical stabilization of jets advances the possibility of stabilizing real liquid columns under intense illumination. We can indeed form liquid columns between the liquid interface and the bottom of the experimental cell. Thus, as the experimental optical cell is 1 or 2 mm high and the meniscus is located in the middle due to near-criticality, it should be possible to connect the liquid jet to the bottom of the cell and to form a liquid column of super-large aspect ratio. This is illustrated in figure 6 in a 1 mm thick cell. The corresponding aspect ratio is $\Lambda = L/2R \approx 70$, well above the Rayleigh–Plateau onset value $\Lambda = \pi$; Λ values largely above 100 are easily achieved in 2 mm cells. Finally, as already demonstrated by the droplets shedding at the jet tip, hydrodynamic flow still persists within liquid channels. as illustrated by the fluid accumulation at their feet.

Since the optical specificity of liquid channels is to intrinsically behave as waveguides, as illustrated in figure 7(a), we suspected this guiding to be at the origin of the observed optical stabilization well above the Rayleigh-Plateau threshold. To check this hypothesis, we used, in a first step, a ray optics description of the light trapped in the bridge to investigate the balance between the radiation pressure of the guided photons and the Laplace pressure [68]. The model consists in counting the number of photons trapped within the channel by total reflection and to calculate the corresponding radiation pressure. Due to the funnel shape of the entrance of the channel, we assumed that the total power trapped in the channel, denoted P_A , equals the power shone on the interface between the channel radius R and αR ($\alpha > 1$). As illustrated in figure 7(b), the parameter α is defined such that rays impinging on the funnel at radius αR have an impact angle of θ_{TIR} , the total internal reflection angle; in a geometrical optics approach, the light shining directly into the bridge, i.e. at a radius smaller than R, will not interact with the walls.

In experiments, we found $\alpha_{exp} = 2.13$ for $\omega_0 = 6.95 \ \mu m$, a value which remains almost constant within the investigated range of waists. Following this scheme, we find

$$P_A = P \exp\left(-\frac{2R^2}{\omega_0^2}\right) \times \left[1 - \exp\left(-\frac{2\left(\alpha^2 - 1\right)R^2}{\omega_0^2}\right)\right].$$
(7)

Once the reflected light enters the bridge, it will bounce down the channel at some angle $\theta > \theta_{\text{TIR}}$ to the normal of the



a) b)

Figure 7. (Adapted from [68]): geometrical model for optical stabilization of liquid channels. (a) Light guiding by the channel observed by scattering; P = 460 mW, $\omega_0 = 3.47 \mu \text{m}$ and $(T - T_{\rm C}) = 5$ K. (b) Rays participating in the stabilization of a liquid channel. (c) Linear momentum transfer Δp of a photon that undergoes total internal reflection inside the bridge.

Since the optical indices of the two fluids are interface. close together in the vicinity of the critical point, θ_{TIR} almost equals $\pi/2$. More precisely, at $(T - T_C) = 4$ K, we have $\theta_{\text{TIR}} = 81.4^{\circ}$. Thus, a reasonable approximation consists in assuming that all the guided light reflects at the same mean angle ϕ and, further, that this angle can be approximated as $\phi = \frac{1}{2}(\theta_{\text{TIR}} + \frac{\pi}{2})$. As each guided photon of energy E_{γ} reflects off of the channel interface, its linear momentum changes by $\Delta p = \|\mathbf{p}^+ - \mathbf{p}^-\| = 2 \frac{n_2}{c} E_{\gamma} \cos \bar{\phi}$. The channel interface consequently receives an equal impulse in the outward normal direction (see figure 7(c)). Then, the total radiation pressure on the channel surface is the product of the momentum change per photon, Δp , by the number of reflections per unit area of the channel wall. This reflection density is given by the rate of photons P_A/E_{γ} divided by the area per reflection for a single photon, $4\pi R^2 \tan \bar{\phi}$. The resulting radiation pressure is

$$\Pi_{\text{Rad}} = \frac{P_A/E_{\gamma}}{4\pi R^2 \tan \bar{\phi}} \Delta p = \frac{n_2}{2c} \frac{\cos \bar{\phi}}{\tan \bar{\phi}} \frac{P_A}{\pi R^2}.$$
 (8)

We can now deduce the bridge radius as a function of the laser power by balancing the radiation pressure Π_{Rad} , which tends to expand a cylindrical channel outwards, against the inward constricting stresses exerted by the Laplace pressure $\Pi_{\text{Laplace}} = \sigma/R$:

$$\frac{\sigma}{R} = \frac{n_2 - n_1}{4\pi c} \frac{P_A}{R^2}.$$
(9)

Figure 8 shows the predictions of the model compared to experimental data for $(T - T_{\rm C}) = 4$ K and $\omega_0 = 6.95 \ \mu {\rm m}$. For an easy comparison, we have rescaled the channel diameter d = 2R and the beam power P, respectively, by the beam waist ω_0 and the laser power $P_{\rm C}$ below which no bridge can exist. This simple geometric model successfully captures all the qualitative features of laser-stabilized liquid channel formation. In particular, it demonstrates that a liquid column can be stabilized by light above a beam power threshold and retrieves the gradual widening of the channel radius with P. However, even if the right orders of magnitude are found, there is, as illustrated in figure 8, a quantitative mismatch in both the size of the bridge and the value of $P_{\rm C}$. These variations can be explained by the use of a ray optics approach and

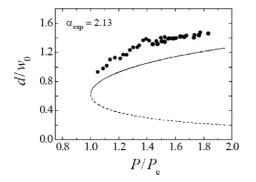


Figure 8. (Adapted from [68]): comparison between the prediction from a geometric model and experimental results obtained for $(T - T_{\rm C}) = 4$ K and $\omega_0 = 6.95 \,\mu$ m. Solid (resp. dashed) line corresponds to stable (resp. unstable) states and circles are experimental results.

its subsequent ray selection mechanism of the photons at the entrance of the channel which represents a partial picture of a more complicated propagation problem.

The mismatch as well as refined predictions were solved using a fully developed electromagnetic description [69, 70]. To do so, we consider the liquid channel as a semi-infinite cylinder of radius R = d/2 perfectly aligned with the incident beam, the inner and outer indices of refraction being, respectively, n_2 and n_1 . Due to the use of near-critical fluids, n_2 and n_1 are very close together and the modes propagating within the channel can be satisfactorily described by the linearly polarized LP_{lm} modes [71]. Moreover, since $n_2 \simeq n_1$, only the first-order terms in $n_2 - n_1$ are retained in calculations. Since the incident beam is Gaussian, there is no coupling with l > 1 modes and only the LP_{0m} modes are excited, each of them carrying a power $P_m = T_M P$, where T_M is the coupling coefficient. The transmission for the *m*th mode is calculated from the normalized overlap integral between the incident Gaussian field and the propagating LP_{0m} field:

$$T_m = \frac{8}{\omega_0^2 R^2} \frac{\left| \int_0^\infty \Re_m(r) \exp(-r^2/\omega_0^2) r \, dr \right|^2}{\frac{J_1^2(\kappa_m R)}{J_0^2(\kappa_m R)} + \frac{K_1^2(\gamma_m R)}{K_0^2(\gamma_m R)}}, \quad (10)$$

where J_m and K_m are the usual *m*th-order Bessel functions and κ_m and γ_m are the *m*th roots of the characteristic equation defining the LP_{0m} mode, $\kappa_m \frac{J_1(\kappa_m R)}{J_0(\kappa_m R)} = \gamma_m \frac{K_1(\gamma_m R)}{K_0(\gamma_m R)}$, with $\kappa_m^2 + \gamma_m^2 = (2\pi/\lambda_0)^2 (n_2^2 - n_1^2)$ [71]. The function $\Re_m(r)$ is associated with the radial profile of the electric field of the *m*th mode:

$$\mathfrak{R}_{m}(r) = \left(\left. \frac{J_{0}\left(\kappa_{m}r\right)}{J_{0}\left(\kappa_{m}R\right)} \right|_{r \leqslant R}, \left. \frac{K_{0}\left(\gamma_{m}r\right)}{K_{0}\left(\gamma_{m}R\right)} \right|_{r \geqslant R} \right).$$
(11)

For steady and perfectly cylindrical liquid channels in weightless conditions, the interface equilibrium condition is, as before, $\Pi_{\text{Rad}} = \Pi_{\text{Laplace}}$, since the viscous stress due to a possible flow within a perfect cylinder (see section 2.5) is tangential to the interface. By definition, the radiation pressure Π_{Rad} corresponds to the radial discontinuity of the electromagnetic stress tensor [72] across the normal to the

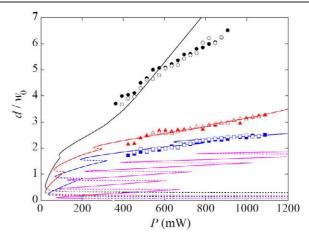


Figure 9. (Adapted from [69]): power dependence of the rescaled liquid channel diameter for different beam waist values: $\omega_0 = 1.8 \ \mu m$ (circles), $\omega_0 = 2.7 \ \mu m$ (triangles), $\omega_0 = 3.5 \ \mu m$ (squares) and $\omega_0 = 7 \ \mu m$ (no symbols). Solid and open symbols refer to experimental data for increasing and decreasing beam power variations, respectively. Solid and dashed lines correspond, respectively, to stable and unstable states predicted by the model.

interface. In the present situation involving nonmagnetic fluids, Π_{Rad} is, at the lowest order in $n_2 - n_1$,

$$\Pi_{\text{Rad}} = \frac{1}{2} \varepsilon_0 \bar{n} (n_2 - n_1) \sum_m |\mathbf{E}^{(m)}|_{r=R}^2, \quad (12)$$

where $\bar{n} = (n_1 + n_2)/2$ is the averaged refractive index, ε_0 is the permittivity in vacuum and $\mathbf{E}^{(m)}$ is the complex electric field of the *m*th mode. Then, since the flux of the Poynting vector across a plane perpendicular to the propagation axis *z* equals the power carried by the *m*th mode of power $P_m = T_M P$, we deduce

$$|\mathbf{E}^{(m)}|_{r=R}^{2} = \frac{2P_{m}}{\varepsilon_{0}\pi c\bar{n}R^{2}} \left[\frac{J_{1}^{2}(\kappa_{m}R)}{J_{0}^{2}(\kappa_{m}R)} + \frac{K_{1}^{2}(\gamma_{m}R)}{K_{0}^{2}(\gamma_{m}R)} \right]^{-1}.$$
 (13)

Finally Π_{Rad} is explicitly obtained by combining equations (10)–(13) and the liquid channel diameter is further found by numerically solving the equilibrium equation $\Pi_{\text{Rad}} = \Pi_{\text{Laplace}}$. As illustrated in figure 9, the model predicts that there is no equilibrium radius below a critical power and the emergence of the set of discrete solutions $\{R_{\text{eq}}^{(n)}\}$. The stability criterion is obtained from a standard static stability analysis at $R = R_{\text{eq}}^{(n)}$ and leads to $\partial \Pi_{\text{Rad}}/\partial R < \partial \Pi_{\text{Laplace}}/\partial R$. Further details in the calculation procedure can be found in [70].

Results are displayed in figure 9, where the solid (resp. dashed) line refers to stable (resp. unstable) solutions. For smallest waists, a good quantitative agreement with experimental data is obtained, particularly if we recall that there is no adjustable parameter. A more complex sequence of stable and unstable states is predicted at low beam powers, but this region cannot be explored experimentally because the large aspect ratio column detaches from the wetting film at the bottom of the cell. Note finally that a single and welldefined diameter does not imply a monomodal behavior. For example, the numbers of modes involved in the range of parameters presented in figure 9 are $m_{\rm max}$ = 6 for ω_0 = 1.8 μ m and $m_{\text{max}} = 4$ for $\omega_0 = 2.7 \ \mu$ m. Conversely, when the beam waist is larger, the model predicts instead multistable states, as illustrated in figure 9 for $\omega_0 = 7 \ \mu m$. There are indeed several stable radii at fixed power that should be characterized experimentally. Then, one would expect to observe metastable states as well as a hysteretic behavior of d(P). We rather found a diameter which varies along the column and with time. In fact, the column explores different discrete diameter values at a single time, which is also a signature of a bistable (or multistable) process [69]. Since the mean amplitude of thermally activated interface fluctuations $(k_{\rm B}T/\sigma)^{1/2}$ [73], where $k_{\rm B}T$ is the thermal energy, is within the range 0.1–0.2 μ m, i.e. one order of magnitude smaller than the interplateaux distance, we eliminate their direct contribution to the observed spatial and temporal radius variations. However, the coupling between these thermal fluctuations with the light-induced flow inside the liquid column, discussed in the following section 2.5, can also give birth to a normal viscous stress allowing exploration of multivalued fiber radii at large beam diameters. In conclusion, figure 9 shows that the mechanism by which liquid channels can be sustained by light is clearly identified.

2.5. Light-induced bulk flow

We showed above how liquid jets can be driven and stabilized by light pressure. However, this surface force always applies normally to interfaces and thus is unable to explain the fluid flow inside the structure at the origin of the droplet shedding at the jet tip or the fluid accumulation at the bottom of liquid channels. A bulk effect is required in the absence of tangential forcing. Radiation pressure is nonetheless known to push suspended particles, as demonstrated by optical levitation [74], because light momentum changes direction when it is elastically scattered by the particles. This momentum change results in a scattering force applied to the particle. As a macroscopic consequence, the photon momentum lost during the beam propagation in a suspension of nanoparticles is transferred to the fluid by momentum conservation. Light is thus able to drive bulk flows in liquid suspensions and emulsions. In a critical fluid, the correlation length of density fluctuations ξ^+ , which diverges close to the critical point, plays this role of refractive index nonhomogeneities [75], as schematically illustrated in the left inset of figure 10. The length scale of these density fluctuations being smaller than 100 nm in experiments, we assumed them to behave as Rayleigh scatterers and to undergo individually a scattering force in the same manner as suspended particles. If we neglect the resulting intensity attenuation due to the associated fluid turbidity, the scattering force density is

$$\mathbf{F}_{\text{Scat}}(r) = \frac{n_{\text{i}}I(r)}{\upsilon c} \oint_{\Omega} (\hat{\mathbf{k}}_{0} - \hat{\mathbf{k}}_{\text{Scat}}) \Sigma(\mathbf{k}_{\text{Scat}}) \, \mathrm{d}\Omega$$
$$= \frac{\pi^{3}}{\lambda_{0}^{4}} \frac{n_{\text{i}}I(r)}{c} \left(\Phi \frac{\partial \varepsilon}{\partial \Phi}\right)^{2} k_{\text{B}}T \chi_{T} f(\beta) \mathbf{z}, \tag{14}$$

where $\hat{\mathbf{k}}_0$ and $\hat{\mathbf{k}}_{Scat}$ are the directions of incident and scattered wavevector, υ^{-1} is the number of scatterers per unit volume,

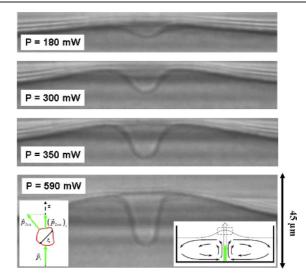


Figure 10. (Adapted from [75]): interface deformations when the laser shines upwards for $(T - T_{\rm C}) = 1.5$ K and $\omega_0 = 8.9 \,\mu$ m. The interface forms a downward deformation (due to radiation pressure) and an upward, broad hump along the beam axis. The left inset illustrates the scattering mechanism for momentum transfer to the fluid and the right inset presents the toroidal recirculation produced in the lower fluid in a one-layer model. The height and radial extent of the hump have been exaggerated. The dotted box indicates the area seen in the photographs.

 $\varepsilon = n^2$, $\Sigma(\mathbf{k}_{\text{Scat}})$ is the scattering cross section, Ω is the solid angle, χ_T is the osmotic compressibility which also diverges close to the critical point and $f(\beta)$ is a function of $\beta = 2(2\pi n\xi^+/\lambda_0)^2$ [76]. Assimilating a jet to a fluid cylinder, and assuming that the low Reynolds number flow within this jet is driven by the scattering force, the Poiseuille equation leads to the following flow rate $Q = \frac{\pi}{128\eta} |\mathbf{F}_{\text{Scat}}| d^4$ for flows bounded by solid cylindrical boundaries. Despite this last approximation, the right order of magnitude is retrieved compared to the measured droplet shedding [75], illustrated in figure 4.

In order to characterize quantitatively this scattering force, we also investigated interface deformation with an upward beam close to the critical point. An example is illustrated in figure 10. Associated with a downward deformation due to radiation pressure effects, the interface also deflects upwards, i.e. in the propagation direction, forming a hump whose lateral length scale is much larger than the beam width. Light scattering produces now an upward body force on the liquid within the laser beam, driving an upward flow within the light region. By conservation of fluid mass, this flow is replenished by a downward flow. Since the Reynolds number is small, as illustrated in figure 4, inertial effects are negligible and purely viscous flows minimize dissipation [77]. The replenishing flow takes the form of a toroidal recirculation whose size is set by the container (see the right inset of figure 10 for a schematic). Consequently, viscous stresses associated with the recirculation deform the interface upwards and creates a hump whose width is ultimately determined by the size of the cell which is therefore much wider than the laser beam. Assuming a Stokes flow, we can deduce $F_{\text{Scat}} \sim \mu \nabla^2 u_0$, where μ is

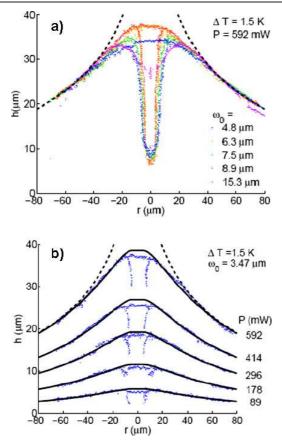


Figure 11. (Adapted from [75]): interface deformations when the laser shines upwards for $(T - T_c) = 1.5$ K. The dashed lines represent the pure buoyancy solution, when the Laplace pressure is ignored. (a) Variation of the hump profile versus the beam waist ω_0 . Away from the beam axis, all the profiles fall onto the same shape, demonstrating that only the laser power, and not ω_0 , affects the large scale hump shape. (b) Calculated (solid line) and experimental (dots) hump profile for $(T - T_c) = 1.5$ K and $\omega_0 = 4.8 \,\mu$ m.

the viscosity and u_0 the induced fluid velocity. Using ω_0 as the optically forced length scale, this leads to $u_0 \sim \chi_T P/\mu$, which shows that the flow should be independent of the beam waist. Figure 11(a) experimentally demonstrates this property. Finally, taking into account buoyancy and the Laplace pressure, as for the description of radiation pressure effects on interfaces, and adding the scattering force, it becomes possible to fit the experimental deformation profiles as shown in figure 11(b).

2.6. Summary

Within a 'total-optical-lab-on-a-chip' framework, we demonstrated in section 2 how the radiation pressure of a laser wave is able to drive large scale flows with flow rates of several hundreds of $\mu m^3 s^{-1}$ using scattering forces. Such light-induced flows should exist whenever fluids have spatial variation in the refractive index, as in nanocolloidal suspensions [78]. Moreover, we showed that surface effects of radiation pressure are able to create soft channels to guide this flow whose diameter can be tuned by light. This ability to optically actuate liquid flow dynamically in three dimensions, without prefabricated channels, opens a new avenue for building microfluidic

devices. Finally, we have presented an electromagnetic instability mechanism of fluid interfaces, still driven by the optical radiation pressure, which leads to optical streaming of fluid interfaces and droplet shedding. The stability of the induced jetting, the actuation of the droplet emission frequency, as well as the monodispersity of the produced microdroplets, also offer new perspectives in digital microfluidics. Since these droplets are optically trapped by the exciting beam, they may further be manipulated by light, if required. Consequently, as far as the involved interfacial tensions are weak enough, mechanical effects of light seem very promising to build microfluidic devices or, conversely, to anticipate new optical microfluidicbased micro-systems for fully developed optofluidics applications. In the next section, we propose a different alternative in order to actuate droplets when interfacial tensions are large enough to prevent efficient actuation by optical forces.

3. Laser actuation for digital microfluidics

3.1. Mechanical aspects

As far as fast flowing nanoliter droplets are used as tight fluid reactors or carriers, new reliable fluid-handling microtechnologies are required to form, convey, synchronize and more generally manipulate accurately these droplets within the robust environment of microchannels. To integrate all these functions onto a single microchip, the method generally consists in plugging in different fluid-handling modules, each of these modules accomplishing one of the required tasks. However, most of these modules need either moving mechanical parts or special micromachining and micropatterning, thus preventing any further spatial reconfiguration in real time. Moreover, each task needs its solution preventing, up to now, the emergence of a sort of unified approach. Since optical forces are within the pN range, which is often weak, to efficiently manipulate fast flowing droplets, we propose an optical alternative based on the production of localized thermocapillary stresses (also known as the Marangoni effect [79]) on the droplet interface. Indeed, in the presence of surfactant, the magnitude of thermocapillary forces can reach the μN range [80], becoming on the way orders of magnitude more efficient than optical forces. Thermocapillary forcing is associated with the temperature dependence of the interfacial tension $\sigma(T)$ between two immiscible fluids. If a droplet flows in a carrying fluid in the presence of a thermal gradient, then this thermal gradient will produce an interfacial tension gradient which will induce in turn a viscous stress in both fluids. It results in an interfacial flow, directed toward the area of largest interfacial tension, which develops both inside and outside the drop. Considering mechanical momentum conservation, the droplet will move in the opposite direction. This mechanism is illustrated in figure 12.

In an unbounded fluid, the thermocapillary migration of a droplet is given by the following expression [79]:

$$\mathbf{U}_{\mathrm{Th}} = -\frac{2}{2\mu_{\mathrm{o}} + 3\mu_{\mathrm{i}}} \left(\frac{\partial\sigma}{\partial T}\right) \frac{R}{2 + \Lambda_{\mathrm{i}}/\Lambda_{\mathrm{o}}} \nabla T, \qquad (15)$$

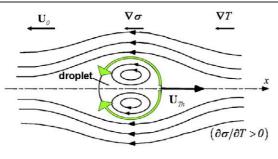


Figure 12. Droplet migration under thermocapillary stresses in a flow field \vec{U}_0 when $\partial \sigma / \partial T > 0$. See text for notation.

where R is the droplet radius, $\mu_{o,i}$ and $\Lambda_{o,i}$ are shear viscosities and thermal conductivities, and the subscripts i, o denote the fluids inside and outside the drop. Equation (15) shows that U_{Th} depends on the thermal gradient and not the temperature directly. Then, even weak, an overheating may drive efficiently thermocapillary flows if the thermal gradient is large, as usually happens with laser light. Moreover, since thermocapillary flows have an interfacial origin, they are particularly suitable to drive flows at small scales where surface effects dominate bulk behaviors, as in microchannels. Such a thermal gradient can be induced by laser, when light is partially absorbed by one of the two fluids. For instance, it was shown that a floating drop [30] or a non-wetting drop deposited on a substrate [81] can move under laser-driven thermocapillary stresses. Thermocapillary actuation inside microchannels was also demonstrated [82, 83], although the amplitude of the velocity is slightly reduced by wall friction [84]. Finally, note that, depending on the sign of $(\partial \sigma / \partial T)$, a laser beam will attract $(\partial \sigma / \partial T < 0)$ or repel $(\partial \sigma / \partial T > 0)$ a drop. Below, we illustrate the generality of this optical approach by showing how it provides a large set of tools for the contactless manipulation of drops in microchannels when $\partial \sigma / \partial T > 0$ [85].

3.2. Experimental set-up

The experimental set-up, presented in figure 13, is composed of an inverted microscope (IX 71, Olympus), a continuous TEM₀₀ argon-ion laser (Innova 300, Coherent) and different types of polydimethylsiloxane (PDMS) microchannels fabricated by soft lithography techniques. Water (viscosity $\eta_{\rm W} = 10^{-3}$ Pa s, thermal conductivity $\Lambda_W = 0.6 \text{ W} \text{ m}^{-1} \text{ K}^{-1}$ at 20 °C) and hexadecane, called hereafter oil ($\eta_{\rm O} = 3.34 \times 10^{-3}$ Pa s, $\Lambda_{\rm O} = 0.14 \text{ W m}^{-1} \text{ K}^{-1}$) are pumped using syringe pumps; hydrostatic pressure was also used incidentally. A surfactant, sorbitan monooelate (Span 80), is added to the oil at 2 wt%. Such an amount of Span 80, which is quite classical in microfluidics, leads to $(\partial \sigma / \partial T) > 0$ [86], yielding to a thermocapillary force that pushes drops away from the beam. Water droplets are formed in the oil either at T or crossjunctions in a central channel; the typical microchannel height is $h = 30 \pm 5 \,\mu\text{m}$. The flowing water is locally heated with the laser by adding 0.1 wt% of fluorescein. The corresponding optical absorption of the aqueous phase is $\alpha_a = 1.18 \text{ cm}^{-1}$. The beam, injected by the rear port of the microscope and a

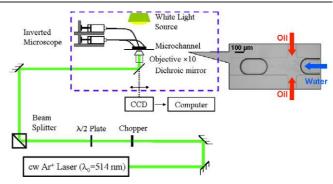


Figure 13. Experimental set-up: an argon-ion laser is focused through the microscope objective into the microchannel. Syringe pumps control the flow rates of oil and water. Right image: close-up view of the water drop formation in a cross-microchannel.

dichroic mirror (Omega), is focused inside the channel using standard microscope objectives (Olympus) of magnification $2.5 \times$, $5 \times$ and $10 \times$ to, respectively, form beam waists of value $\omega_0 = 10.3$, 5.2 and $2.6 \ \mu$ m. In these focusing conditions, the beam can be assumed as cylindrical over the microchannel height. Finally, images of the droplet traffic are acquired with a high speed CMOS camera (Lightning RTD, DRS).

3.3. Optofluidic mixer: a first step

Fluid mixing in miniaturized systems is a difficult task because diffusion dominates low Reynolds number flows. An appealing method is to use the so-called chaotic advection concept, which is based on the fact that chaotic fluid streamlines can be forced to disperse fluid species effectively, even in smooth and regular flow fields [87]. However, such a mixing method often requires the use of complex three-dimensional microstructures. Active mixing, still based on chaotic advection, is usually achieved by periodic perturbation of the flow fields. Several fluid actuation methods were demonstrated using heat convection, differential pressure, ultrasonic actuation, magnetic actuation or electrokinetic pressure; see [88] for a recent review. One important advantage of these mixers is that they can be activated on demand. However, compared to passive ones, they need external power sources and hence require more complex packaging. Local heating by laser could bring new insights in active mixing [89] for at least four reasons: (i) laser waves can easily be focused over spatial scales in the typical range of microchannels; (ii) laser heating is contactless and thus does not need any complex packaging or patterning of heating elements; (iii) laser heating is totally reconfigurable in terms of input energy (beam power), spatial extension (beam waist), duration (chopping) and positioning within the channel in a very fast way (compared to hydrodynamic timescales) and finally (iv) multi-actuation is also easy to implement using spatial light modulators and/or galvanometric mirrors.

A first step towards thermocapillary mixing in microchannels is illustrated in figure 14. A water drop flowing in a 140 μ m wide channel at a few mm s⁻¹ is blocked by the laser due to thermocapillary stresses; the blocking mechanism is discussed in the following subsection. Using tracer particles in both fluids, we observe convection rolls inside and outside the

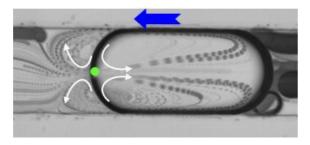


Figure 14. (Adapted from [80]): superposition of 100 frames from a video sequence showing the thermocapillary motion of particle tracers inside and outside a beam-blocked drop. The laser position is indicated by the dot and the main flow by the wide arrow. The tracer motion is directed toward the hot spot (indicated by the thin arrows), thus demonstrating $(\frac{\partial \sigma}{\partial T}) > 0$. Water flow rate

 $Q_{\rm W} = 0.01 \ \mu l \ {\rm min}^{-1}$, oil flow rate $Q_{\rm O} = 0.1 \ \mu l \ {\rm min}^{-1}$, frame rate 200 Hz, beam power $P = 60 \ {\rm mW}$, beam waist $\omega_0 = 5.2 \ \mu {\rm m}$, microchannel section $140 \times 35 \ \mu {\rm m}^2$, carbon tracers: $\emptyset \approx 1 \ \mu {\rm m}$.

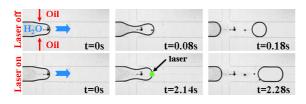


Figure 15. (Adapted from [80]): microfluidic valve: in a cross-shaped microchannel, the oil and water flows, respectively, enter from the lateral and the central channels. In the absence of laser excitation, drops are shed with a typical break-off time of 100 ms. When the laser is applied, the interface is blocked for several seconds, producing a larger drop. Water flow rate

 $Q_{\rm W} = 0.08 \ \mu l \ {\rm min}^{-1}$, oil flow rate $Q_{\rm O} = 0.9 \ \mu l \ {\rm min}^{-1}$, beam power $P = 80 \ {\rm mW}$, beam waist $\omega_0 = 5.2 \ \mu {\rm m}$, main channel width 200 $\mu {\rm m}$ and lateral channel width 100 $\mu {\rm m}$.

drop due to the presence of the laser beam. In the present experiment the tracer velocity at the beam location is 11 mm s^{-1} . The presence of rolls inside and outside the drop illustrates the procedure to perform thermocapillary mixing in both fluids. Indeed, a single set of vortices is obviously not sufficient, but chaotic mixing should emerge by using three independent beams, each producing its set of vortices.

3.4. Optofluidic valve

Experiments were performed in a cross-microchannel. In the example illustrated in figure 15, water droplets are formed by hydrodynamic focusing and are emitted at a frequency of 0.5 Hz. Their mean length and velocity are, respectively, $L = 200 \ \mu\text{m}$ and $U_0 = 1.2 \ \text{mm s}^{-1}$. When the front of the forming drop reaches the beam spot position it is pushed away from the laser by thermocapillary forces; see equation (15) using $(\partial \sigma / \partial T) > 0$. Since it is constrained by the channel walls, the interface cannot turn around the beam, which thus behaves as a sort of 'soft' wall. When the beam power reaches a threshold (such as $|U_{\text{Th}}| \ge U_0$), the drop is blocked in the microchannel.

When the microchannel is pumped at constant flow rates, the blocking time T_b increases with the incident beam

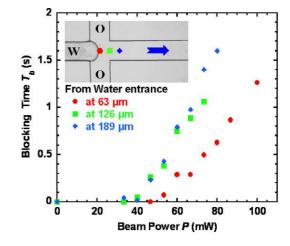


Figure 16. (Adapted from [80]): dependence of the blocking time $T_{\rm b}$ on laser power and position indicated in the picture. Water flow rate $Q_{\rm W} = 0.03 \ \mu \rm l \ min^{-1}$, oil flow rate $Q_{\rm O} = 0.1 \ \mu \rm l \ min^{-1}$, beam waist $\omega_0 = 2.6 \ \mu \rm m$ and channel widths 125 $\mu \rm m$.

power [80]. This is illustrated in figure 16 for different blocking positions. T_b also decreases with the beam waist at a given beam power and significantly increases when flows are controlled by pressure instead of flow rate [90].

3.5. Optofluidic sampler

Another way to calibrate droplets consists in using as a drain a T junction with two arms of different length [91]. At the T junction, the flowing drop divides into two daughter drops whose lengths are imposed by the asymmetry of the two exit channels. While very efficient and reliable, this type of sampler is passive and the asymmetry in daughter droplets can only be varied by switching to channels with different asymmetry. This type of asymmetry can be optically actuated at a classical T junction by positioning the laser close to one of the outlets. This is illustrated in figure 17.

By momentarily blocking the drop flow close to one outlet, the drop preferentially flows into the other exit. We then obtained daughter droplets of different sizes. This asymmetry, discussed in [85], is characterized from the mean aspect ratio $\Lambda = (L_1 - L_2)/(L_1 + L_2)$, where L_1 and L_2 are the lengths of the two daughter droplets. The extreme values $\Lambda = 0, 1,$ respectively, correspond to the symmetric case and a situation where the mother drop is entirely diverted toward a single outlet. Both situations are illustrated in figure 18. When the beam power is smaller than a threshold, still defined by $|U_{\rm Th}| \ge U_0$, the mother drop divides into two daughters whose aspect ratio increases with P. Above threshold, equal to P = 100 mW for the experiment presented in figure 17, the mother does not divide anymore and flows toward the unblocked outlet. Below the beam power threshold the device behaves as a sampler while above it, it becomes a droplet switcher.

3.6. Optofluidic switcher

A second approach to microfluidic switching can be devised using a slightly different channel geometry. Instead of blocking

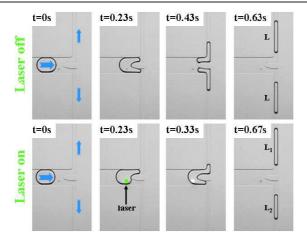


Figure 17. (Adapted from [85]): microfluidic sampler. In the absence of a laser the mother drop divides into two daughters of the same size. Laser forcing allows for breaking this symmetry in a controlled fashion. Water flow rate $Q_W = 0.02 \ \mu l \ min^{-1}$, oil flow rate $Q_O = 0.2 \ \mu l \ min^{-1}$, beam power $P = 87 \ m$ W, beam waist $\omega_0 = 5.2 \ \mu m$ and main channel width 200 μm .

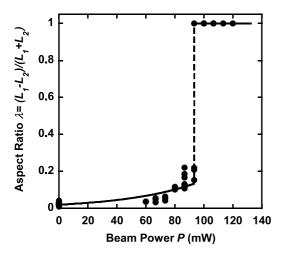


Figure 18. (Adapted from [85]): microfluidic sampler and switcher: dependence of the asymmetry in daughter droplet sizes on laser power ($\Lambda = 0$ is for symmetric drops and $\Lambda = 1$ is for drop switching). Lines are guides for the eye. The operating conditions are those of figure 17.

droplets at the exit, we can just switch their trajectories at an enlargement of the main channel. This is illustrated in figure 19 using a Y-type channel. The main advantage of this approach is that the droplet can be manipulated at velocities as large as cm s^{-1} [92] because blocking is not required anymore. On the one hand, without a laser, water drops formed upstream arrive in the enlarged part of the Y junction and flow asymmetrically downstream towards the exit channel of smaller hydrodynamic resistance (the down one in pictures). This weak asymmetry is forced by shortening one of the outlets, as in [91]. On the other hand, when a flowing drop is illuminated below its equator by the laser, it is suddenly pushed out in the opposite direction above a beam power threshold. Once again, the beam behaves as an active wall whose stiffness is optically actuated.

The beam power variation of the droplet switching efficiency was investigated in [92]. It presents a very steep

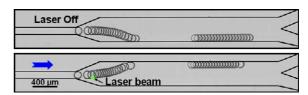


Figure 19. (Adapted from [92]): microfluidic switcher: superposition of successive frames illustrating drop switching by local thermocapillary actuation. The dot indicates the laser location observed by the fluorescence of the water–dye solution. Water flow rate $Q_W = 0.05 \ \mu l \ min^{-1}$, oil flow rate $Q_O = 5 \ \mu l \ min^{-1}$, $U_0 = 2.6 \ mm \ s^{-1}$, $R = 60 \ \mu m$, $P = 58 \ mW$, frame rate 100 fps. Channel width: inlet 100 μ m, enlargement 400 μ m and exits 200 μ m.

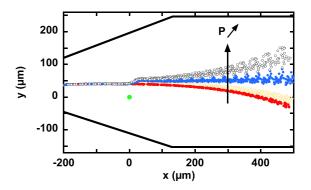


Figure 20. (Adapted from [92]): trajectory of the drop center of mass for various beam powers (from bottom to top: P = 0, 48, 53, 58 mW); switching occurs at P = 53 mW. The beam location is indicated by the dot at (x = 0, y = 0). Water flow rate $Q_{\rm W} = 0.05 \ \mu \rm l \ min^{-1}$, oil flow rate $Q_{\rm O} = 5 \ \mu \rm l \ min^{-1}$, $U_0 = 2.6 \ mm \ s^{-1}$, $R = 60 \ \mu m$.

behavior. Below a power threshold, all drops exit from the asymmetrically forced outlet while above threshold they all deviate toward the opposite outlet, as illustrated by the beam power variation of the drop trajectories in the microchannel (figure 20).

Due to the expression of the thermocapillary velocity (equation (15)), the beam power threshold varies with the drop velocity and radius. Generally speaking, it appears that switching occurs when the mean deviation angle between the natural and the beam-modified trajectory reaches some threshold value associated with the forced asymmetry in the *y* direction for the droplet traffic in the absence of a laser. In the above experiment, the mean deviation angle over the interaction time is 3° ; this mean critical angle and the related beam power required for switching could be even lowered by locating the beam closer to the channel centerline.

3.7. Optofluidic sorter

To demonstrate the reliability of the above laser switching process for sorting droplets on chips, we used the same Y-type microchannel with two upstream T junctions (seen on the left of figure 21) instead of one. The very left one is used for the generation of pure water droplets and the other produces water–dye droplets as in the above switching experiment. An

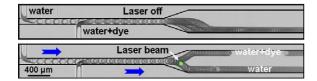


Figure 21. (Adapted from [92]): microfluidic sorting: top picture: frames superposition showing three drops (water-dye + water + water-dye) which are naturally drawn toward the lower exit channel without a laser. Bottom picture: frames superposition showing sorting of a pure water and a water-dye drop when the laser shines on the droplets. The dot indicates the laser location observed by fluorescence of the water-dye solution. Water flow rate $Q_W = (0.03 + 0.03) \ \mu l \ min^{-1}$, oil flow rate $Q_O = 5 \ \mu l \ min^{-1}$, $U_0 = 2.2 \ mm \ s^{-1}$, $R = 59 \ \mu m$ and $P = 159 \ mW$, frame rate 100 fps. Channel width: inlets 100 $\ \mu m$, enlargement 400 $\ \mu m$ and exits (not shown) 200 $\ \mu m$.

asymmetry in outlets, designed as in [91], still forces the drops to naturally flow toward the lower outlet, whatever their composition. Since the residual optical absorption of water at $\lambda_0 = 514.5$ nm is negligible ($\approx 3 \times 10^{-3}$ cm⁻¹), pure water droplets are rather insensitive to the beam while water-dye droplets are spontaneously deviated and exit from the other outlet at beam powers above threshold, as illustrated in figure 21.

3.8. Optofluidic merger

Finally, beyond mixing inside and outside flowing drops, there is a second type of mixing process required for digital microfluidics. As far as flowing droplets are used as microreactors, the coalescence of at least two daughter droplets, supporting different reagents, is necessary. This coalescence stage is difficult to accomplish passively in microchannels because surfactant molecules are known to stabilize drops against merging, even if recent work has shown a very interesting alternative to circumvent this difficulty [93]. Active control can be efficient too, as illustrated by recent investigations on charged microdroplets [94].

The thermocapillary coupling may also circumvent the lubrication film problem by evacuating the surfactant molecules, as illustrated in figure 22. Here, the downstream drop is held stationary by thermocapillary blocking until a second one collides with it (figure 22, t = 0 s). At this point, the two drops advance until the laser intercepts the adjacent interfaces, and we observe that the lubrication film between the drops is rapidly evacuated, leading to merging [85].

3.9. Summary

Within a 'total-optical-lab-on-a-chip' framework, we proposed in section 3 a method, thermocapillary actuation by laser, to optically manipulate droplets inside a microchannel. The method seems promising because it takes advantage of the dominance of surface effects in microfluidics, as opposed to dipolar couplings used by optical trapping standards. Within this framework, we developed a basic optical toolbox for digital microfluidics, including valves, samplers, switchers,

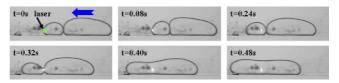


Figure 22. (Adapted from [85]): time sequence showing droplet fusion by laser-induced thermocapillary stresses. Half of a 200 μ m wide channel is presented, droplets flowing close to one edge. Coalescence occurs when the beam intercepts the touching interfaces. Water flow rate $Q_{\rm W} = 0.2 \ \mu \rm I \ min^{-1}$, oil flow rate $Q_{\rm O} = 0.9 \ \mu \rm I \ min^{-1}$, beam power $P = 67 \ m$ W, beam waist $\omega_0 = 2.6 \ \mu \rm m$.

sorters, mergers and a first step towards mixers. These drophandling functions can also be used in combination; a first example is illustrated in [85]. As in any optical technique, our method is still contactless, allowing droplet manipulation without moving parts or dedicated microfabrication. Optical forcing also allows reconfigurability in real time, either spatially or temporally, with one or many beams. Finally, extension to near-IR excitation would make the method even more flexible and amenable for applications since the optical absorption of pure water at $\lambda_0 = 1500$ nm is $\alpha_a \approx 20$ cm⁻¹, thus eliminating polluting dyes for heating and reducing the required beam power range by a factor of 20.

4. Conclusion and prospects

The main purpose of this survey was to theoretically and experimentally explore fluid actuation by light within a 'total-optical-lab-on-a-chip' framework. To implement such a strategy in microchannels, we need interface forcing mechanisms. In the first part (section 2), we demonstrated how the optical radiation pressure of a laser wave was able to induce large scale flows, to create soft channels for guiding these flows and to drive optical streaming of fluid interfaces and droplet shedding. In section 3, we implemented an indirect optical interface coupling, opto-capillarity, to build an optical toolbox for digital microfluidics. The method shows appealing prospects to form valves, samplers, switchers, sorters, mergers and mixers, and to even develop further these actuators and to couple them. Drops were nevertheless formed by flow focusing instead of light.

These experimental results bring us back to the Glückstad concept of an 'optical chip' [25], where flows, channels and operating functions on microdroplets would all be performed optically on the same device, extending by the way the optofluidic approach [16, 17] to digital microfluidics.

On the one hand, the mechanisms at the origin of flows driven by bulk radiation pressure effects, using the scattering force, are analogous to those involved in the recent development of the so-called optical chromatography of microparticles, which is a relatively new technique of laser separation [95] still under development [96, 97]. Since in our experiment in near-critical fluids the particle-like fluctuations are only 100 nm large with a very weak refraction index contrast, and since refractive index contrast in real colloids is usually much larger, we expect an enhancement of scattering forces in classical suspensions of nanoparticles. Thus, bulk radiation forces should be able to push collectively a suspension as a whole and this pushing effect should be still very efficient at nanometric scales. While previous calculations performed in the geometric optics approximation (which is not very accurate for these sizes) gave 100 nm as the smallest size for separation by optical chromatography [95], using instead the Rayleigh cross section, we find $F_{\text{Scat}} = 0.5$ pN for a glass bead of radius 100 nm which may lead to a water flow of mean velocity 0.2 mm s⁻¹. We can therefore conclude that the scattering force should initiate a flow in a suspension of colloidal particles of a few tens of nm and would become particularly efficient in the 100 nm range to initiate and sustain microfluidics flows.

The interfacial component of the radiation pressure efficiency would nevertheless suffer from a drawback: interfacial tension must be weak enough to allow significant mechanical effects of light. Hopefully, digital microfluidics frequently uses surfactants at concentrations generally above the critical micellar concentration (CMC). Is it sufficient to stabilize liquid channels? Since the resulting interfacial tensions are within the range of $0.1-1 \text{ mN m}^{-1}$, radiation pressure may stabilize small radius channels as far as these radii are reachable (remember the subcritical behavior of the channel radius versus the incident beam power, figures 8 and 9). Conversely, is it sufficient to destabilize fluid interfaces? Up to now, the answer is probably no due to the fact that even more power will be required to induce a liquid jet. A significant power increase also means an increase of the unavoidable side effects such as those triggered by laser heating. On the other hand, the major limitation of the optocapillary approach is its dissipative nature (use of thermal gradients triggered by optical absorption). Its application viability would need an extension towards near-IR in order to (i) eliminate polluting dyes and (ii) significantly reduce the required beam powers in order to directly integrate the laser on the chip. These requirements can easily be achieved. However, we will not be able to use the same laser (multiplexed as much as needed) to control the flow and simultaneously actuate the droplet shedding because the required beam power ranges are incompatible to each other at the same wavelength. Typically, too much IR light, to trigger surface effects of radiation pressure will vaporize fluids while visible light will be inefficient for actuating droplets. Consequently up to now the best compromise, if it exists, would probably be the use of different lasers, each accomplishing his task. The subject clearly deserves further investigations!

As a preliminary conclusion, the present survey illustrates another step in the exploration of the coupling between optics and microfluidics. Associated with much more exhaustive reviews [16–18] and to the increasing number of publications in this domain, it illustrates the emergence of a new and very active research field. In fact, the development of optofluidics, and in our case optohydrodynamics, is obviously in its infancy, probably because it intimately mixes disciplines which have been considered far away for a long time. Right now, everything is open since we do not even feel the limits of our framework. The constant progress in microfabrication always pushes these limits, engaging more and more fundamental and applied investigations. Our presentation participates in this dash, giving one more view of the synergy between optics and fluidics and allowing us to dream of 'optical chips'.

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References

- [1] Squires T M and Quake S R 2005 Rev. Mod. Phys. 77 977
- [2] Song H, Chen D L and Ismagilov R F 2006 Angew. Chem. Int. Edn 45 7336
- [3] Grier D 2003 Nature 424 810
- [4] Dholakia K, MacDonald M P, Zemanek P and Cizmar T 2007 Methods Cell Biol. 82 467
- [5] Galajda P and Ormos P 2001 Appl. Phys. Lett. 78 249
- [6] Neale S L, MacDonald M P, Dholakia K and Krauss T F 2005 Nat. Mater. 4 530
- [7] Leach J, Mushfique H, di Leonardo R, Padgett M and Cooper J 2006 Lab Chip 6 735
- [8] Terray A, Oakey J and Marr D W M 2002 Science **296** 1841
- [9] MacDonald M P, Spalding G C and Dholakia K 2003 Nature 426 421
- [10] Ozkan M, Wang M M, Ozkan C, Flynn R A and Esener S 2003 Biomed. Microdev. 5 61
- [11] Applegate R W Jr, Squier J, Vestad T, Oakey J, Marr D W M, Bado P, Dugan M A and Said A A 2006 Lab Chip 6 422
- [12] Grujic K, Hellesø O G, Hole J P and Wilkinson J S 2005 Opt. Express 13 1
- [13] Renn M J, Pastel R and Lawandowski H J 1999 Phys. Rev. Lett. 82 1574
- [14] Hart S J, Terray A V, Arnold J and Leski T A 2007 Opt. Express 15 2724
- [15] Chiou P Y, Ohta A T and Wu M C 2005 Nature 436 370
- [16] Psaltis D, Quake S R and Yang C 2006 Nature 442 381
- [17] Hunt H C and Wilkinson J S 2008 Microfluid. Nanofluid. 4 53
- [18] Monat C, Domachuk P, Grillet C, Collins M, Eggleton B J, Cronin-Golomb M, Mutzenich S, Mahmud T, Rosengarten G and Mitchell A 2008 *Microfluid. Nanofluid.* 4 81
- [19] Campbell K, Groisman A, Levy U, Pang L, Mookherjea S, Psaltis D and Fainman Y 2004 Appl. Phys. Lett. 85 6119
- [20] Nguyen N T, Kong T F, Goh J H and Low C L N 2007 J. Micromech. Microeng. 17 2169
- [21] Zhang D Y, Justis N and Lo Y H 2004 Appl. Phys. Lett. 84 4194
- [22] Casner A and Delville J P 2001 Opt. Lett. 26 1418
- [23] Grillet C, Domachuk P, Ta'eed V, Mägi E, Bolger J, Eggleton B, Rodd L and Cooper-White J 2004 Opt. Express 12 5440
- [24] Li Z, Zhang Z, Emery T, Scherer A and Psaltis D 2006 Opt. Express 14 696
- [25] Glückstad J 2004 Nat. Mater. 39

- [26] Darhuber A and Troian S 2005 Annu. Rev. Fluid Mech. 37 425
- [27] Velev O, Prevo B and Bhatt K 2003 Nature 426 515
- [28] Ichimura K, Oh S K and Nakagawa M 2000 Science 288 1624
- [29] Ito Y, Heydari M, Hashimoto A, Konno T, Hirasawa A, Hori S, Kurita K and Nakajima A 2007 Langmuir 23 1845
- [30] Rybalko S, Magome N and Yoshikawa K 2004 Phys. Rev. E 70 046301
- [31] Garnier N, Grigoriev R and Schatz M F 2003 Phys. Rev. Lett. 91 054501
- [32] Darhuber A, Valentino J, Davis J, Troian S and Wagner S 2003 Appl. Phys. Lett. 82 657
- [33] Joanicot M and Ajdari A 2005 Science **309** 887
- [34] Günther A and Jensen K F 2006 Lab Chip 6 1487
- [35] Teh S Y, Lin R, Hung L H and Lee A P 2008 *Lab Chip* 8 198
- [36] Schatz M F and Neitzel G P 2001 Annu. Rev. Fluid Mech. 33 93
- [37] Ashkin A and Dziedzic J M 1973 Phys. Rev. Lett. 30 139
- [38] Kats A V and Kontorovich V M 1975 Sov. Phys.-JETP 41 346
- [39] Gordon J P 1973 Phys. Rev. A 8 14
- [40] Born M and Wolf E 1980 *Principles of Optics* 6th edn (Oxford: Pergamon)
- [41] Casner A and Delville J P 2001 Phys. Rev. Lett. 87 054503
- [42] Lai H M and Young K 1976 Phys. Rev. A 14 2329
- [43] Brevik I 1979 Phys. Rep. 52 133
- [44] Chernov S P, Ivanova O N and Shepelev V A 1975 Sov. J. Quantum Electron. 4 1161
- [45] Ostrovskaya G V, Komissarova I I and Shedova E N 1987 Opt. Commun. 66 15
- [46] Sakai K, Mizuno D and Takagi K 2001 Phys. Rev. E 63 046302
- [47] Mitani S and Sakai K 2002 Phys. Rev. E 66 031604
- [48] Yoshitake Y, Mitani S, Sakai K and Takagi K 2005 J. Appl. Phys. 97 024901
- [49] Lee C H, Guo C L and Wang J 1998 Opt. Lett. 23 307
- [50] Lee C H, Lin W C and Wang J 2001 *Phys. Rev. E* 64 020901[51] Richert A, Hénon S, Le Normand G and Gallet F 1999
- Biophys. J. 76 1145
- [52] Guck J, Ananthakrishnan R, Moon T J, Cunningham C C and Käs J 2000 Phys. Rev. Lett. 84 5451
- [53] Guck J, Ananthakrishnan R, Mahmood H, Moon T J, Cunningham C C and Käs J 2001 *Biophys. J.* 81 767
- [54] Guck J et al 2005 Biophys. J. 88 3689
- [55] Zhang J Z and Chang R K 1988 Opt. Lett. **13** 916
- [56] Poon K L, Lai H M, Leung P T and Young K 1989 J. Opt. Soc. Am. B 6 2430
- [57] Brevik I and Kluge R 1999 J. Opt. Soc. Am. B 16 976
- [58] Moldover M R 1985 Phys. Rev. A 31 1022
- [59] Meunier J, Cazabat A M, Langevin D and Pouchelon A 1982 J. Physique Lett. 43 89
- [60] Wunenburger R, Casner A and Delville J P 2006 Phys. Rev. E 73 036314
 - Wunenburger R, Casner A and Delville J P 2006 *Phys. Rev.* E **73** 036315
- [61] Casner A, Delville J P and Brevik I 2003 J. Opt. Soc. Am. B 20 2355
- [62] Chraibi H, Lasseux D, Arquis E, Wunenburger R and Delville J P 2008 *Eur. J. Mech.* B **27** 419
- [63] Casner A and Delville J P 2003 Phys. Rev. Lett. 90 144503
- [64] Thorsen T, Roberts R W, Arnold F H and Quake S R 2001 Phys. Rev. Lett. 86 4163

- [65] Anna S L, Bontoux N and Stone H A 2003 Appl. Phys. Lett. 82 364
- [66] Link D R, Grasland-Mongrain E, Duri A, Sarrazin F, Cheng Z, Cristobal G, Marquez M and Weitz D A 2006 Angew. Chem. Int. Edn 45 2556
- [67] Eggers J 1997 Rev. Mod. Phys. 69 865
- [68] Schroll R D, Brasselet E, Zhang W W and Delville J P 2008 Eur. Phys. J. E 26 405
- [69] Brasselet E, Wunenburger R and Delville J P 2008 Phys. Rev. Lett. 101 014501
- [70] Brasselet E and Delville J P 2008 Phys. Rev. A 78 013835
- [71] Okamoto K 2006 *Fundamentals of Optical Waveguides* 2nd edn (Amsterdam: Elsevier)
- [72] Landau L and Lifschitz E 1984 *Electrodynamics of Continuous Media* (Oxford: Pergamon)
- [73] Aarts D G A L, Schmidt M and Lekkerkerker H N W 2004 Science 304 847
- [74] Ashkin A 1997 Proc. Natl Acad. Sci. USA 94 4853
- [75] Schroll R D, Wunenburger R, Casner A, Zhang W W and Delville J P 2007 Phys. Rev. Lett. 98 133601
- [76] Puglielli V G and Ford N C Jr 1970 Phys. Rev. Lett. 25 143
- [77] Batchelor G K 1967 An Introduction to Fluid Dynamics (Cambridge: Cambridge University Press)
- [78] Mitani S and Sakai K 2005 Faraday Discuss. 129 141
- [79] Barton K D and Subramanian R S 1989 J. Colloid Interface Sci. 133 211
- [80] Baroud C N, Delville J P, Gallaire F and Wunenburger R 2007 Phys. Rev. E 75 046302
- [81] Kotz K T, Noble K A and Faris G W 2004 Appl. Phys. Lett.
 85 2658
- [82] Otha A T, Jamshidi A, Valley J K, Hsu H Y and Wu M C 2007 *Appl. Phys. Lett.* **91** 074103
- [83] Ting T H, Yap Y F, Nguyen N T, Wong T N, Chai J C K and Yobas L 2006 Appl. Phys. Lett. 89 234101
- [84] Bratukhin Y K, Kostarev K G, Viviani A and Zuev A L 2005 Exp. Fluids 38 594
- [85] Baroud C, Robert de Saint Vincent M and Delville J P 2007 Lab Chip 7 1029
- [86] Sloutskin E, Bain C D, Ocko B M and Deutsch M 2005 Faraday Discuss. 129 1
- [87] Stroock A D, Dertinger S K W, Ajdari A, Mezic I, Stone H A and Whitesides G M 2002 Science 295 647
- [88] Nguyen N T and Wu Z 2005 J. Micromech. Microeng. 15 R1
- [89] Grigoriev R O 2005 Phys. Fluids 17 033601
- [90] Robert de Saint Vincent M and Delville J P 2008 unpublished results
- [91] Link D R, Anna S L, Weitz D A and Stone H A 2004 Phys. Rev. Lett. 92 054503
- [92] Robert de Saint Vincent M, Wunenburger R and Delville J P 2008 Appl. Phys. Lett. 92 154105
- [93] Bremond N, Thiam A R and Bibette J 2008 Phys. Rev. Lett. 100 024501
- [94] Ahn K, Agresti J, Chong H, Marquez M and Weitz D A 2006 Appl. Phys. Lett. 88 264105
- [95] Kaneta T, Ishidzu Y, Mishima N and Imasaka T 1997 Anal. Chem. 69 2701
- [96] Terray A V, Arnold J and Hart S J 2005 Opt. Express 13 10406
- [97] Hart S J, Terray A V and Arnold J 2007 Appl. Phys. Lett. 91 171121