

Proceedings

Open Access

## Linkage analysis of anti-CCP levels as dichotomized and quantitative traits using GAW15 single-nucleotide polymorphism scan of NARAC families

Xiaohong R Yang\*, Kimberly F Kerstann, Andrew W Bergen, Alisa M Goldstein and Lynn R Goldin

Address: Division of Cancer Epidemiology and Genetics, National Cancer Institute, 6120 Executive Boulevard, MSC 7236, Bethesda, Maryland 20892, USA

Email: Xiaohong R Yang\* - royang@mail.nih.gov; Kimberly F Kerstann - kkerstann@mail.nih.gov; Andrew W Bergen - bergena@mail.nih.gov; Alisa M Goldstein - goldstea@mail.nih.gov; Lynn R Goldin - goldinl@mail.nih.gov

\* Corresponding author

from Genetic Analysis Workshop 15  
St. Pete Beach, Florida, USA. 11–15 November 2006

Published: 18 December 2007

BMC Proceedings 2007, 1(Suppl 1):S107

This article is available from: <http://www.biomedcentral.com/1753-6561/1/S1/S107>

© 2007 Yang et al; licensee BioMed Central Ltd.

This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

Rheumatoid arthritis is a clinically and genetically heterogeneous disease. Anti-cyclic citrullinated (anti-CCP) antibodies have a high specificity for rheumatoid arthritis and levels correlate with disease severity. The focus of this study was to examine whether analyzing anti-CCP levels could increase the power of linkage analysis by identifying a more homogeneous subset of rheumatoid arthritis patients. We also wanted to compare linkage signals when analyzing anti-CCP levels as dichotomized (CCP\_binary), categorical (CCP\_cat), and continuous traits, with and without transformation (log\_CCP and CCP\_cont). Illumina single-nucleotide polymorphism scans of the North American Rheumatoid Arthritis Consortium families were analyzed for four chromosomes (6, 7, 11, 22) using nonparametric linkage (NPL) (rheumatoid arthritis and CCP\_binary), regress (CCP\_cat and Log\_CCP), and deviates (CCP\_cont) analysis options as implemented in Merlin. Similar linkage results were obtained from analyses of rheumatoid arthritis, CCP\_binary, and CCP\_cont. The only exception was that we observed improved linkage signals and a narrower region for CCP\_binary as compared to a clinical diagnosis of rheumatoid arthritis alone on chromosome 7, a region which previously showed variation in linkage results with rheumatoid arthritis according to anti-CCP levels. Analyses of CCP\_cat and Log\_CCP had little power to detect linkage. Our data suggested that linkage analyses of anti-CCP levels may facilitate identification of rheumatoid arthritis genes but quantitative analyses did not further improve power. Our study also highlighted that quantitative trait linkage results are highly sensitive to phenotype transformation and analytic approaches.

## Introduction

Rheumatoid arthritis (RA) is a chronic inflammatory autoimmune disease affecting about 1% of the population. A genetic component for RA has been well established, with the MHC region being the largest single contributing component. Other chromosome regions (11q, 10q, 14q, 6p, 6q, 16q, 12p, etc.) and candidate genes (*PTPN22*, *CTLA4*, *PADI4*) have been identified by whole-genome linkage scans and association studies [1-5]. Most recently, a high-density SNP analysis of 642 families affected with RA collected by the North American Rheumatoid Arthritis Consortium (NARAC), the largest single linkage study of RA, identified two new linkage regions, 11p and 2q [6]. These findings reflect the genetic complexity of the disease and suggest that analysis of a more homogeneous RA phenotype might increase the power of linkage analysis. In addition, most previous studies have analyzed RA as a dichotomous trait, which could lead to a power loss if RA is a naturally quantitative trait [7].

Anti-cyclic citrullinated (anti-CCP) antibodies have a high specificity for RA [8] and the levels are correlated with disease severity [9,10]. To examine whether the power of linkage analysis could be improved by analyzing a more homogeneous phenotype and by quantitative characterization of the trait, we performed linkage analysis of anti-CCP antibody levels for selected chromosome regions previously linked to RA using NARAC data.

## Methods

### Data set

Illumina SNP scans of the NARAC families were analyzed. We chose anti-CCP antibody levels as the phenotype of interest, and evaluated the effect of covariates including sex, age of onset, year of birth, ever/never smoking, and current smoking. Anti-CCP antibody levels were analyzed in three ways: dichotomized, categorical, and continuous. An antibody titer of 20 was used as a cut-off value to dichotomize anti-CCP levels into positive ( $>20$ ) and negative ( $\leq 20$ ). In addition, anti-CCP levels were characterized into multiple (four) categories (negative, 0–19.9; low, 20–49.9; medium, 50–99.9; high,  $\geq 100$ ). Anti-CCP levels were also analyzed as continuous measurements. Because the assay for anti-CCP antibody titer has an upper limit of 210, we recoded all measurements exceeding 210 to 210. A log transformation was applied to approximate normality of anti-CCP levels because the raw data were highly skewed.

### Chromosome regions

Because the purpose of this study was to compare methods rather than search for a new locus for anti-CCP, we limited our linkage analyses to selected regions. Chromosomes were selected based on findings from a previous

SNP scan of NARAC families for RA [6]. Chromosome 6, which contains human lymphocyte antigen (HLA) locus, was chosen as a positive control region. Chromosome 22, which did not show evidence for linkage with RA, was selected as the negative control. Chromosomes 7 and 11, which showed suggestive and significant evidence for linkage with RA, respectively, were included in our linkage analyses as test regions. In particular, it has been shown that chromosome 7 might harbor a susceptibility locus that was more closely linked to anti-CCP positive disease [6].

### Linkage analysis

SNPs on chromosomes 6, 7, 11, and 22 were analyzed for all four anti-CCP phenotypes (CCP\_binary, CCP\_cat, Log\_CCP, and CCP\_cont) as well as RA affection status. Linkage disequilibrium (LD) between markers was calculated and markers in LD defined by  $D' > 0.7$  were removed using SNPLINK [11]. CCP\_binary and RA affection were analyzed by nonparametric (NPL) linkage analysis using Merlin. CCP\_cat and Log\_CCP were analyzed by regression analysis implemented in Merlin Regress, which uses trait-squared sums and differences to predict IBD sharing between sib pairs [12]. To run Merlin Regress, it was necessary to specify some trait distribution parameters, such as mean, variance, and heritability in the general population. We did not use the sample mean and variance because the families were affected with RA and therefore had higher frequencies and levels of anti-CCP positives. Instead, we estimated the mean (0 for CCP\_cat, 0.78 for Log\_CCP) and variance (0.0088 for CCP\_cat, 0.11 for Log\_CCP) among individuals who were anti-CCP negative ( $\leq 20$ ) to approximate the distribution in the general population. Heritability of 0.6 was estimated based on variance-component analysis using Merlin. Untransformed continuous anti-CCP levels were also analyzed using the *deviates* option implemented in Merlin, which makes no assumptions about the trait distribution. Again, we chose 0 to approximate the population mean for anti-CCP levels.

## Results

A total of 746 families containing 1794 affected individuals with RA were used for the linkage analyses. There were only 11 individuals in this data set who were unaffected with RA. Anti-CCP data was available for 1499 individuals. Among the 823 sib pairs with available anti-CCP data, there were 77 concordant negative, 228 discordant, and 518 concordant positive for anti-CCP. The distribution of anti-CCP levels was highly skewed toward high positive, with  $>50\%$  of the individuals having high anti-CCP levels when using cut points suggested by the data provider (Table 1). Male sex and ever smoking were associated with significantly increased anti-CCP values ( $t$ -test  $p < 0.001$  and 0.004, respectively) (Table 2).

**Table 1: Distribution of anti-CCP levels among the 1499 individuals included in the linkage analyses**

| CCP category | N   | %     |
|--------------|-----|-------|
| Negative     | 341 | 22.75 |
| Low          | 167 | 11.14 |
| Medium       | 181 | 12.07 |
| High         | 810 | 54.04 |

Results obtained from NPL analysis of RA are consistent with those shown previously by Amos et al. [6]. Overall, similar linkage signals were obtained from analyses with RA and CCP\_binary, with weaker evidence for linkage with CCP\_binary on chromosomes 6 and 11 (Table 3, Figure 1). On the other hand, higher LOD scores and improved definition of a linkage peak for CCP\_binary were observed on chromosome 7 (Figure 1), which has been suggested to be more closely linked to anti-CCP compared to RA. Among quantitative anti-CCP levels, only analysis of untransformed anti-CCP levels showed comparable results with that of RA. Analyses of either categorical or log-transformed anti-CCP levels resulted in significantly reduced linkage signals for both positive control (chromosome 6) and test regions (chromosomes 7 and 11). Adjustment of gender and ever smoking in the regression-based linkage analyses did not significantly change the results (data not shown). As expected, chromosome 22 did not show evidence for linkage to any of the phenotypes.

## Discussion

In this study, we analyzed anti-CCP levels as dichotomized, categorical, and continuous traits in linkage analyses. On chromosome 7, we observed improved linkage signals and a narrower linkage peak for dichotomized anti-CCP levels as compared to a clinical diagnosis of RA alone, although neither analysis reached statistical signif-

icance. Chromosome 7 previously showed stronger evidence for linkage for RA with positive anti-CCP [6]. However, for the three chromosomes we selected that demonstrated evidence for linkage with RA, analyzing anti-CCP levels as quantitative traits did not further improve power to detect linkage compared to RA. In addition, categorization and log transformation of the continuous phenotype resulted in significantly reduced power to detect linkage.

Mapping susceptibility genes is challenging for clinically and genetically heterogeneous diseases such as RA. Examining a more homogenous disease phenotype might increase the power of linkage analysis. Anti-CCP is highly specific for RA and its levels are correlated with disease severity [9,10]. A previous study suggested genetic variability according to anti-CCP status on chromosomes 4, 5, 6, and 7 [6]. Our data on chromosome 7 supports the hypothesis that anti-CCP status might represent a more genetically homogeneous phenotype of RA. In addition, although linkage signals on chromosome 11 were less significant for the CCP\_binary phenotype than RA, linkage peak regions on both chromosomes 7 and 11 were narrower, suggesting that analyzing anti-CCP levels as a dichotomous phenotype might be helpful in narrowing candidate regions in fine mapping.

For naturally occurring continuous or polychotomous traits, dichotomization could lead to power loss for linkage analysis [7]. In particular, if the underlying gene confers not only disease susceptibility but also disease severity, treating disease phenotypes as quantitative traits could provide additional information in linkage analysis. We hypothesized that performing quantitative linkage analyses might increase the power to detect linkage for anti-CCP levels. However, analyses of none of the three quantitative phenotypes provided improved linkage signals compared to NPL analysis of RA. In particular, regres-

**Table 2: Anti-CCP levels by gender, age, year of birth, and smoking status**

| Covariate     | Values          | N    | anti-CCP mean | SD     | t test P        |
|---------------|-----------------|------|---------------|--------|-----------------|
| Sex           | Female          | 1153 | 112.19        | 102.58 | <0.001          |
|               | Male            | 346  | 143.24        | 100.19 |                 |
| Age of onset  | <40             | 758  | 119.70        | 97.53  | ns <sup>a</sup> |
|               | ≥40             | 722  | 119.89        | 108.01 |                 |
| Year of birth | <1944           | 754  | 119.34        | 97.53  | ns              |
|               | ≥1944           | 738  | 119.50        | 108.01 |                 |
| Smoking       | Never           | 618  | 110.54        | 100.38 | 0.004           |
|               | Ever            | 832  | 126.13        | 104.91 |                 |
|               | Current         | 244  | 119.84        | 105.32 |                 |
|               | Never or former | 1211 | 119.44        | 102.79 |                 |

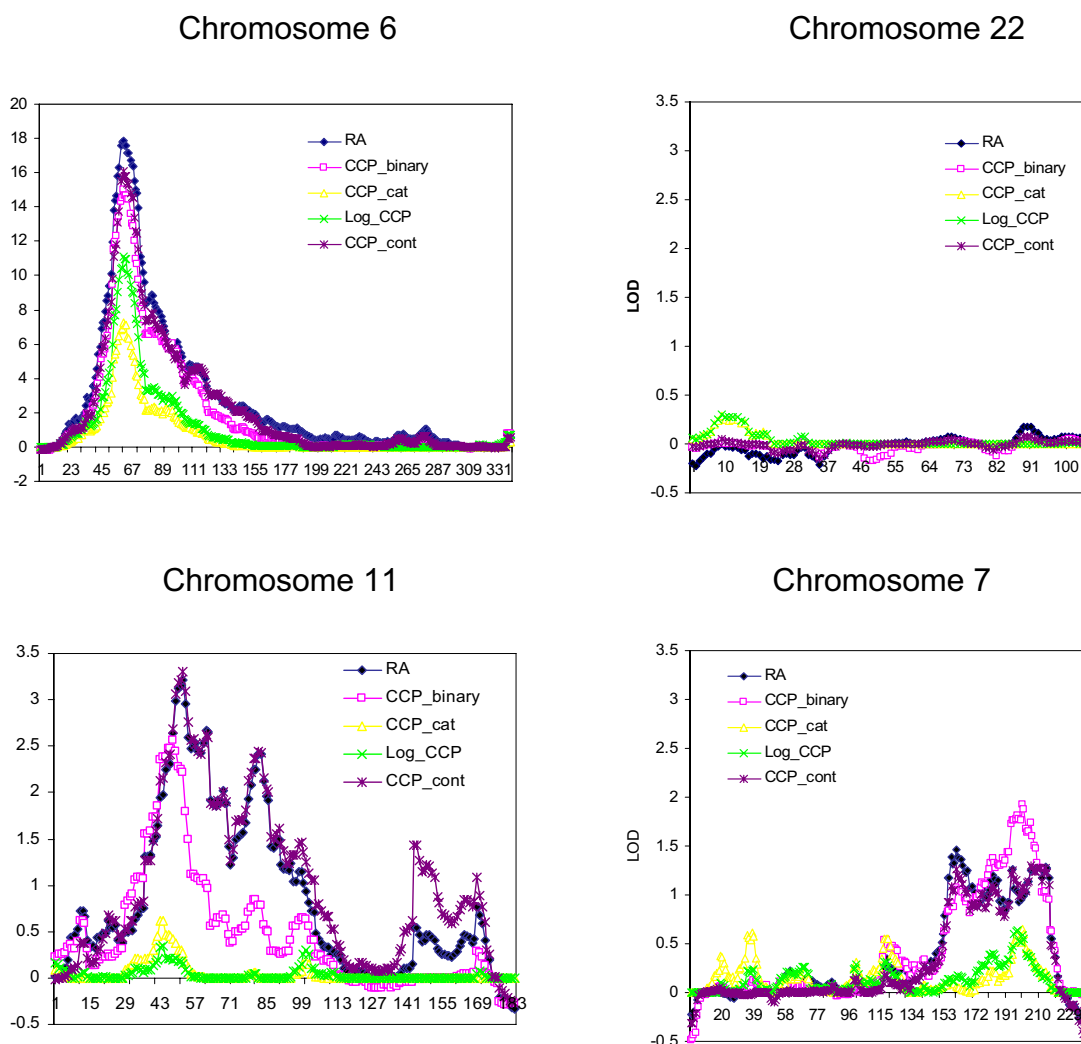
<sup>a</sup>ns, not significant

**Table 3: Maximal LOD scores for RA and anti-CCP phenotypes in the four chromosome regions**

| Chromosome | RA    | CCP binary | CCP categorical | Log CCP | CCP continuous |
|------------|-------|------------|-----------------|---------|----------------|
| 6          | 17.86 | 15.01      | 7.26            | 11.10   | 15.77          |
| 7          | 1.46  | 1.93       | 0.65            | 0.64    | 1.30           |
| 11         | 3.22  | 2.56       | 0.62            | 0.34    | 3.30           |
| 22         | 0.08  | 0.05       | 0.29            | 0.31    | 0.08           |

sion-based quantitative linkage analyses of log-transformed or ordinal anti-CCP levels, with or without covariate adjustment, appeared to have reduced power to

detect linkage compared to NPL analysis of dichotomized anti-CCP status. We also used the variance components and quantitative trait locus analysis options implemented



**Figure 1**  
**Linkage analyses of 4 chromosomes for RA and anti-CCP levels using SNPs.** LOD scores for binary outcomes (RA, CCP\_binary), transformed quantitative traits (CCP\_cat, Log\_CCP), and untransformed quantitative trait (CCP\_cont) were calculated using Merlin NPL, Merlin Regress, Merlin --deviates, respectively. The scale of the Y-axis for chromosome 6 is different.

in Merlin for CCP\_cat and Log\_CCP phenotypes and observed similar results. For this particular trait, transforming the phenotype to achieve normality reduced the signal due to the major locus. Alternate approaches to handle non-normality are needed for quantitative trait linkage analysis in selected samples. Consistent with another report [13], we found loss of power when treating CCP\_cat as continuous. Novel approaches, such as the recently proposed proportional odds latent variable model by Feng et al. [14], need to be further explored. Finally, all these quantitative linkage analyses are very sensitive to the specification of population parameters, such as population mean and variance, chosen for the *regress* and *deviates* analyses. Replacement of the estimated population mean by the sample mean or default values set by the programs resulted in completely different LOD scores. These results further highlight the importance of precise estimates and appropriate analytical approaches in linkage analysis of quantitative traits.

## Conclusion

Linkage analyses of anti-CCP levels may facilitate identification of RA genes by making the phenotype more homogeneous. However, quantitative analyses of anti-CCP levels did not improve power to detect linkage over analysis of RA. Furthermore, log transformation and categorization of anti-CCP levels resulted in reduced power to detect linkage.

## Competing interests

The author(s) declare that they have no competing interests.

## Acknowledgements

This article has been published as part of *BMC Proceedings* Volume 1 Supplement 1, 2007: Genetic Analysis Workshop 15: Gene Expression Analysis and Approaches to Detecting Multiple Functional Loci. The full contents of the supplement are available online at <http://www.biomedcentral.com/1753-6561/1?issue=S1>.

## References

- Jawaheer D, Seldin MF, Amos CI, Chen WV, Shigeta R, Monteiro J, Kern M, Criswell LA, Albani S, Nelson JL, Clegg DO, Pope R, Schroeder HW, Bridges SL, Pisetsky DS, Ward R, Kastner DL, Wilder RL, Pincus T, Callahan LF, Flemming D, Wener MH, Gregersen PK: **A genome-wide screen in multiplex rheumatoid arthritis families suggests genetic overlap with other autoimmune diseases.** *Am J Hum Genet* 2001, **68**:927-936.
- Jawaheer D, Seldin MF, Amos CI, Chen WV, Shigeta R, Etzel C, Damle A, Xiao X, Chen D, Lum RF, Monteiro J, Kern M, Criswell LA, Albani S, Nelson JL, Clegg DO, Pope R, Schroeder HW, Bridges SL, Pisetsky DS, Ward R, Kastner DL, Wilder RL, Pincus T, Callahan LF, Flemming D, Wener MH, Gregersen PK, North American Rheumatoid Arthritis Consortium: **Screening the genome for rheumatoid arthritis susceptibility genes. A replication study and combined analysis of 512 multicase families.** *Arthritis Rheum* 2003, **48**:906-916.
- Osorio Y, Fortéa J, Bukulmez H, Petit-Teixeira E, Michou L, Pierlot C, Cailleau-Moindrault S, Lemaire I, Lasbleiz S, Alibert O, Quillet P, Bardin T, Prum B, Olson JM, Cornélis F: **Dense genome-wide linkage analysis of rheumatoid arthritis, including covariates.** *Arthritis Rheum* 2004, **50**:2757-2765.
- Tamiya G, Shinya M, Imanishi T, Ikuta T, Makino S, Okamoto K, Furugaki K, Matsumoto T, Mano S, Ando S, Nozaki Y, Yukawa W, Nakashige R, Yamaguchi D, Ishibashi H, Yonekura M, Nakami Y, Takayama S, Endo T, Saruwatari T, Yagura M, Yoshikawa Y, Fujimoto K, Oka A, Chiku S, Linsen SE, Giphart MJ, Kulski JK, Fukazawa T, Hashimoto H, Kimura M, Hoshina Y, Suzuki Y, Hotta T, Mochida J, Minezaki T, Komai K, Shiozawa S, Taniguchi A, Yamanaka H, Kamatani N, Gojobori T, Bahram S, Inoko H: **Whole genome association study of rheumatoid arthritis using 27,039 microsatellites.** *Hum Mol Genet* 2005, **14**:2305-2321.
- Oliver JE, Worthington J, Silman AJ: **Genetic epidemiology of rheumatoid arthritis.** *Curr Opin Rheumatol* 2006, **18**:141-146.
- Amos CI, Chen WV, Lee A, Li W, Kern M, Lundsten R, Batliwalla F, Wener M, Remmers E, Kastner DA, Criswell LA, Seldin MF, Gregersen PK: **High-density SNP analysis of 642 Caucasian families with rheumatoid arthritis identifies two new linkage regions on 11p12 and 2q33.** *Genes Immun* 2006, **7**:277-286.
- Corbett J, Gu CC, Rice JP, Reich T, Province MA, Rao DC: **Power loss for linkage analysis due to the dichotomization of trichotomous phenotypes.** *Hum Hered* 2004, **57**:21-27.
- Rantapaa-Dahlqvist S, de Jong BAW, Berglin W, Hallmans G, Wadell G, Stenlund H, Sundin U, van Venrooij WJ: **Antibodies against cyclic citrullinated peptide and IgA rheumatoid factor predict the development of rheumatoid arthritis.** *Arthritis Rheum* 2003, **48**:2741-2749.
- Kastbom A, Strandberg G, Lindroos A, Skogh T: **Anti-CCP antibody test predicts the disease course during 3 years in early rheumatoid arthritis (the Swedish TIRA project).** *Ann Rheum Dis* 2004, **63**:1085-1089.
- del Val Del Amo N, Ibanez Bosch R, Fito Manteca C, Gutierrez Polo R, Loza Cortina E: **Anti-cyclic citrullinated peptide antibody in rheumatoid arthritis: relation to disease aggressiveness.** *Clin Exp Rheumatol* 2006, **24**:281-286.
- Webb El, Sellick GS, Houlston RS: **SNPLINK: multipoint linkage analysis of densely distributed DNP data incorporating automated linkage disequilibrium removal.** *Bioinformatics* 2005, **21**:3060-3061.
- Sham PC, Purcell S, Cherny SS, Abecasis GR: **Powerful regression-based quantitative-trait linkage analysis of general pedigrees.** *Am J Hum Genet* 2002, **71**:238-253.
- Wang X, Ye Y, Zhang H: **Family-based association tests for ordinal traits adjusting for covariates.** *Genet Epidemiol* 2006, **30**:728-736.
- Feng R, Leckman JF, Zhang H: **Linkage analysis of ordinal traits for pedigree data.** *Proc Natl Acad Sci USA* 2004, **101**:16739-16744.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

