



## Links Between Inflammatory Bowel Disease and Chronic Obstructive Pulmonary Disease

#### April L. Raftery, Evelyn Tsantikos, Nicola L. Harris and Margaret L. Hibbs\*

Department of Immunology and Pathology, Central Clinical School, Monash University, Melbourne, VIC, Australia

Inflammatory bowel disease (IBD) and chronic obstructive pulmonary disease (COPD) are chronic inflammatory diseases of the gastrointestinal and respiratory tracts, respectively. These mucosal tissues bear commonalities in embryology, structure and physiology. Inherent similarities in immune responses at the two sites, as well as overlapping environmental risk factors, help to explain the increase in prevalence of IBD amongst COPD patients. Over the past decade, a tremendous amount of research has been conducted to define the microbiological makeup of the intestine, known as the intestinal microbiota, and determine its contribution to health and disease. Intestinal microbial dysbiosis is now known to be associated with IBD where it impacts upon intestinal epithelial barrier integrity and leads to augmented immune responses and the perpetuation of chronic inflammation. While much less is known about the lung microbiota, like the intestine, it has its own distinct, diverse microflora, with dysbiosis being reported in respiratory disease settings such as COPD. Recent research has begun to delineate the interaction or crosstalk between the lung and the intestine and how this may influence, or be influenced by, the microbiota. It is now known that microbial products and metabolites can be transferred from the intestine to the lung via the bloodstream, providing a mechanism for communication. While recent studies indicate that intestinal microbiota can influence respiratory health, intestinal dysbiosis in COPD has not yet been described although it is anticipated since factors that lead to dysbiosis are similarly associated with COPD. This review will focus on the gut-lung axis in the context of IBD and COPD, highlighting the role of environmental and genetic factors and the impact of microbial dysbiosis on chronic inflammation in the intestinal tract and lung.

## OPEN ACCESS

#### Edited by:

Gislane Lelis Vilela de Oliveira, São Paulo State University, Brazil

#### Reviewed by:

Helioswilton Sales-Campos, Universidade Federal de Goiás, Brazil Elena Vikström, Linköping University, Sweden

> \*Correspondence: Margaret L. Hibbs Margaret.Hibbs@monash.edu

#### Specialty section:

This article was submitted to Mucosal Immunity, a section of the journal Frontiers in Immunology

Received: 01 July 2020 Accepted: 07 August 2020 Published: 11 September 2020

#### Citation:

Raftery AL, Tsantikos E, Harris NL and Hibbs ML (2020) Links Between Inflammatory Bowel Disease and Chronic Obstructive Pulmonary Disease. Front. Immunol. 11:2144. doi: 10.3389/fimmu.2020.02144 Keywords: inflammatory bowel disease, Crohn's disease metabolites, chronic obstructive pulmonary disease, microbial dysbiosis, gut-lung axis

## **INTRODUCTION**

Inflammatory bowel disease (IBD) and chronic obstructive pulmonary disease (COPD) are chronic inflammatory diseases that affect the gastrointestinal tract and respiratory system, respectively, with both being characterized by recurrent disease cycles that result in tissue damage and worsening of disease symptoms. As mucosal epithelial sites, the gastrointestinal and respiratory tracts share structural similarities which may result in part from common embryonic origin in the primitive foregut (1). Its hypothesized that these structural similarities may account for inherent parallels in the immune responses at these two sites and contribute to the dynamic involvement of the gut-lung axis in inflammation.

1

## **CHRONIC INFLAMMATORY DISEASES**

## **Inflammatory Bowel Disease**

IBD is an umbrella term that describes chronic relapsing inflammation of the gastrointestinal tract; the major types being Crohn's disease metabolites (CDM) and ulcerative colitis (UC). CD is characterized by transmural, non-continuous, and noncaseating granulomatous inflammation that can occur at any point along the entirety of the gastrointestinal tract, however, inflammation most commonly manifests in the terminal ileum (2, 3). UC is characterized by continuous inflammation that originates in the rectum and progresses proximally. Unlike CD, the inflammation in UC only affects the mucosa and submucosa and solely manifests in the colon (4, 5). The etiology of IBD has not been fully elucidated, however, a complex interplay of genetic susceptibility, environmental risk factors, inappropriate immune responses directed against the microbiota, intestinal barrier hyperpermeability, and dysbiosis of commensal microbiota of the intestines are thought to contribute to pathogenesis (6) (Figure 1).

Presently there are no curative treatments for IBD. With current management strategies, 10–35% of CD patients will require surgery within the first year of diagnosis, and up to 60% will require surgery within a decade of initial diagnosis (7). For UC, 30% of patients will require a colectomy within 10 years of diagnosis (8). Further research into the mechanisms driving IBD is needed to identify novel therapeutic targets, and a fuller understanding of the role of intestinal microbiota in IBD could provide some valuable insights in pursuit of this.

## **Chronic Obstructive Pulmonary Disease**

COPD is a progressive and largely irreversible disease that is characterized by prolonged inflammation, tissue destruction, and airflow obstruction leading to the reduced functional capacity

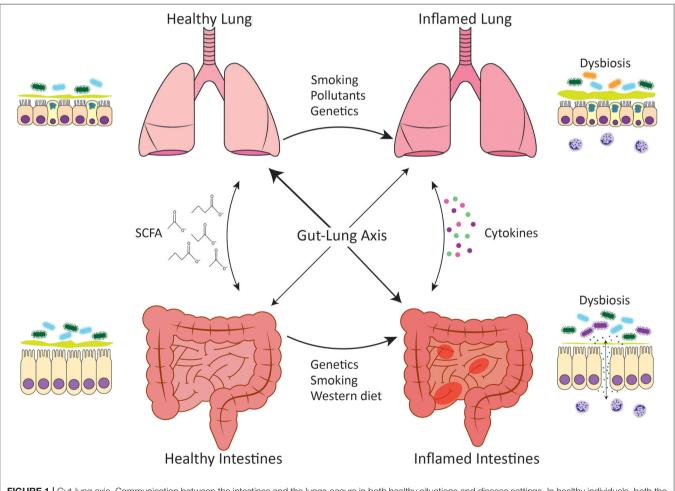


FIGURE 1 | Gut-lung axis. Communication between the intestines and the lungs occurs in both healthy situations and disease settings. In healthy individuals, both the intestines and the lungs harbor diverse microbial communities that have evolved to complement the host and predominately comprise bacteria of the Bacteroidetes and Firmicutes phyla. The gut microbiota performs key functions such as the generation of SCFA from the host's diet, which play an important role in homeostatic maintenance. Microbial dysbiosis occurs in association with chronic inflammatory diseases such as IBD and COPD and leads to loss of epithelial barrier integrity and inappropriate immune responses directed against the microbiota. Dysbiosis is characterized by reduced diversity of *Firmicutes* spp. in IBD and the expansion of *Proteobacteria* spp. in COPD. Genetic variations as well as environmental stimuli such as cigarette smoke or a Western diet have been linked to intestinal and lung microbial dysbiosis. A healthy, fiber-rich diet promotes intestinal, and respiratory health.

of the lungs. The disease is driven by chronic exaggerated inflammatory responses in the airways and parenchyma of the lungs in response to a noxious insult such as cigarette smoke or environmental pollutants or genetic factors such as alpha-1 antitrypsin deficiency (Figure 1). Airway inflammation drives airway remodeling leading to mucus metaplasia and airway obstruction as well as tissue destruction that results in the enlargement of the alveoli, also known as emphysema. Smoking is a major risk factor for the development of COPD, however, other factors such as chronic asthma, low birth weight, childhood respiratory tract infections, pulmonary tuberculosis, and occupational exposures to dusts have also been associated with COPD (9). The prevalence of COPD has been reported to be as high as 20% in never-smokers suggesting that other risk factors for COPD have been overlooked due to the focus on cigarette smoke (10). COPD is one of the leading causes of mortality worldwide accounting for 3 million deaths annually (11), and much like IBD, there is no curative treatment beyond lung transplantation although this is still up for debate (12).

## Linking IBD and COPD

Population based studies have identified an increased prevalence of IBD in patients with COPD, and an increased risk of mortality in patients with both COPD and CD (13-15). Furthermore, the risk of COPD patients developing either CD or UC is increased by 2.72 and 1.83, respectively, compared to healthy controls (13), results that have been confirmed in additional populations (16). Reciprocally, a retrospective study found that CD patients were at increased risk of dying from COPD (17). Additionally, a study in Quebec residents found that asthmatic and COPD patients had increased incidences of CD of 27 and 55%, respectively, compared to the general population in that province (18). There is also evidence that up to 60% of IBD patients have some degree of subclinical lung disease (19). COPD and CD share environmental risks, with cigarette smoke exposure being a major risk factor for both (20). However, this does not explain the increased prevalence of UC amongst COPD patients as cigarette smoke has been proposed to be protective against the development of UC in the general population (21). CD patients without chronic lung disease have reduced pulmonary function that correlates with the presence of sputum lymphocytosis and eosinophilia and inversely with CD activity index (22). This suggests that shared environmental risk factors alone are not sufficient to induce these comorbidities and that alternative mechanisms that link the intestine and lung are responsible.

The epithelium of the respiratory and gastrointestinal systems are derived embryonically from the primitive foregut (1). This shared origin likely underlies the ability of these two mucosal surfaces to act similarly as selective barriers, allowing for the translocation of gases or nutrients, whilst maintaining mutualistic relationships with the microbiota and keeping pathogens at bay. These similarities in function suggest that these tissues may also respond to disease-causing stimuli in the same way, which could account for the increased risk of CD development in COPD patients. Indeed, parallels exist in the immune systems at both mucosal sites and this is mirrored in the immunopathology of IBD and COPD. Both disease settings are characterized by increases in myeloid cells such as neutrophils, eosinophils and macrophages, as well as innate lymphoid cells (ILCs) and unconventional T cells such as  $\gamma\delta$ T cells, all of which are important in microbial interactions and maintenance of epithelial barriers. While this is beyond the scope of this review, the reader is directed to the following reviews for more information on the pathophysiology and immunopathology of these diseases (23-26). It is well-known that the intestinal epithelium is readily damaged in IBD (27), and increased intestinal permeability has now also been reported in COPD (28, 29). Whilst the association between IBD and COPD has been largely investigated at an epidemiological level, further research is required to elucidate the inflammatory mechanisms that link the intestine and the lung. This review will focus on the mechanisms linking IBD and COPD that pertain to mucosal microbial communities and dysbiosis in disease settings (Figure 1).

## MUCOSAL MICROBIAL COMMUNITIES AND DYSBIOSIS IN DISEASE SETTINGS

## **Healthy Intestinal Microbiota**

The gastrointestinal tract is one of the largest surfaces of the body that is constantly in contact with environmental factors. At mucosal sites, such as the intestine, the immune system is primed by interactions with the microbiota and environmental antigens, and a fine homeostatic balance needs to be maintained to ensure quiescence toward harmless microbes whilst being able to promote a proinflammatory response against invading pathogens. The intestine is the most densely colonized surface of the human body with Bacteroidetes and Firmicutes representing the two most abundant bacterial phyla making up approximately 90% of all microorganisms of the gastrointestinal tract (30, 31). A mutualistic relationship exists between the intestinal microbiota and the human host. The intestine provides a nutrient-rich niche for the microbiota to inhabit, whilst the host benefits from the increased digestive capacity that the microbiota provides, from its ability to prime the immune system, and from a reduction in the available niche for potentially pathogenic micro-organisms (32). In addition, the intestinal microbiome contains more than 3.3 million non-redundant genes, 150-fold greater than the human gene complement, and these provide both metabolic and health benefits to the host (33).

The microbiota is not stagnant, and its composition can be altered by a variety of factors including diet, infection, inflammation, or antibiotics; and this shift in microbial communities is referred to as dysbiosis. Dysbiosis is often associated with IBD (34, 35) but is also observed in a variety of chronic inflammatory and autoimmune diseases including rheumatoid arthritis, psoriasis, neurodegenerative diseases, diabetes, allergic diseases, and asthma (36–42). At present, essentially nothing is known about the gut microbiota in COPD.

## Healthy Lung Microbiota

Historically, due to a reliance on culture-dependent techniques, the lungs were thought to be sterile. With the emergence

Links Between IBD and COPD

of culture-independent techniques utilizing genetic sequencing, pulmonary microbial communities have been found in the healthy lung and characterization of the lung microbiota in the context of respiratory diseases has become possible (43). However, progress in our understanding of the lung microbiota is still limited by factors such as low microbial loads in the lower airways, contamination of samples from the upper airways and oral cavity, and sampling methods that over-represent the upper airways over the lower airways. This is important since the upper and lower airways harbor distinct microbial communities (44). A study comparing the microbial communities of both the oral cavity and the lower airways in healthy non-smokers and smokers identified bacteria that comprised the healthy lung microbiota and showed that lung bacterial communities were not significantly altered by smoking, although differences were apparent in the oral microbiota. Similar to the intestines, the most abundant bacterial phyla present in the lungs were Bacteroidetes and Firmicutes and they did not derive entirely from the mouth as the microbiota of the oral cavity was dominated by Firmicutes, Proteobacteria, and Bacteroidetes (44).

### **Dysbiosis of Intestinal Microbiota in IBD**

Dysbiosis has been associated with the development of IBD, although whether this is cause or effect is yet to be elucidated since most research to date has been correlative. Cross-sectional studies are the most common for assessing the microbiota of IBD patients but these only provide a snapshot in time (45). Instituting longitudinal studies could assist in establishing when dysbiosis occurs relative to the onset of intestinal inflammation and how this may change with the course of disease but this would require early and frequent sampling. Furthermore, the microbiome tends to be sequenced from fecal samples and while this is non-invasive for the IBD patient, this method could provide inaccurate measures, especially for CD patients where inflammation is often localized to the ileum or other portions of the gastrointestinal tract. This sampling method can also overlook mucosa-associated microbiota, and due to the variations in microbial composition that are apparent across the gastrointestinal tract, sampling methods need to be carefully considered when formulating studies (46). Nonetheless, the research that has been conducted to date has provided a solid foundation upon which future studies can expand to improve our understanding of dysbiosis in IBD.

Broadly, dysbiosis in IBD patients is associated with shifts promoting an increase in potentially proinflammatory bacteria and a decrease in protective bacteria. In CD patients, intestinal dysbiosis has been characterized by a reduction in diversity of species belonging to the Firmicutes phyla, a change that has been suggested to occur prior to disease onset (34, 47, 48). Furthermore, a specific reduction in relative abundance in *Dialister invisus* and *Faecalibacterium prausnitzii* species of the Firmicutes phylum and *Bifidobacterium adolescentis* of the Actinobacteria phylum, together with an increase in the mucolytic species *Ruminococcus gnavus* and *Ruminococcus torques* of the Firmicutes phylum is observed in CD patients compared to healthy controls (34, 49). The importance of intestinal microbiota in development of IBD is supported by common experimental mouse models, such as nucleotidebinding oligomerization domain-containing protein 2 (NOD2)deficiency and IL-10-deficiency which ordinarily develop ileitis when maintained under standard housing conditions but display reduced disease penetrance in specific pathogen-free environments (50-53).

Adherent-invasive *Escherichia coli* (AIEC) is an organism that is commonly associated with CD. This bacterium is able to take advantage of gaps in host defense such as impaired bacterial recognition and defective intracellular killing, allowing AIEC to expand (54). AIEC have been found to induce granuloma formation in the inflamed ilea (55), a hallmark pathological feature of CD. These granulomas consist of multinucleated giant cells and epithelioid cells, both phagocytic cells of the macrophage lineage that activate T cells in an antigenspecific manner, that are surrounded by a B cell corona (56). Furthermore, it has been found that AIEC are able to survive and replicate within macrophages without inducing cell death, a process that also promotes increased secretion of proinflammatory tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) (57), which is involved in the pathogenesis of CD.

A study that examined dysbiosis in an inducible model of ileitis using Toxoplasma gondii together with high dose indomethacin, found that severe ileitis was associated with a shift in microbial communities from populations dominated by species belonging to the Firmicutes phyla to those largely represented by species belonging to the Proteobacteria phyla (58). This dysbiosis was accompanied by translocation of AIEC and was similar to that observed in CD patients. While these data suggest that inflammation is sufficient to induce dysbiosis, it is also clear that genetic susceptibility plays a role since T. gondii induced heightened dysbiosis and AIEC invasion in mice lacking the ileitis susceptibility gene NOD2, while disease was significantly muted in mice lacking the proinflammatory C-C chemokine receptor 2 (CCR2), which are a model of ileitis resistance (58). Furthermore, it has been demonstrated in an alternative genetic knock out model of ileitis susceptibility that dysbiosis precedes the onset of ileitis (59).

Similar to CD, dysbiosis in UC is characterized by a reduction in species belonging to Firmicutes and Bacteroidetes phyla, and a concomitant increase in species belonging to Proteobacteria and Actinobacteria phyla (60). However, at the species level, the dysbiotic signature of UC is distinct from CD. UC patients exhibit a reduction in relative abundance of *Roseburia hominis* and *Faecalibacterium prausnitzii* (35), both butyrate-producing bacteria of the Firmicutes phylum whose abundance is inversely correlated with disease severity (35). A similar mechanism involving the interaction of genetic susceptibility, inflammation and microbial dysbiosis contributes to UC.

# Genetics of IBD and Its Role in Intestinal Dysbiosis

To date, more than 150 susceptibility genes have been identified for IBD, most of which are involved in the detection and clearance of microbial compounds (61). Three common variants of NOD2 are associated with an increased risk of developing

CD, with a 2-4-fold increase for heterozygous mutations and a 20-40-fold increase for homozygous mutations (62, 63). NOD2 variants are the most common mutations in CD patients of Caucasian descent, being identified in 30% of patients (64). Furthermore, certain variants in NOD2 have been associated with more severe disease phenotypes including early-onset disease, ileitis, and strictures caused by fibrosis (65-67). NOD2 is a cytoplasmic molecule that senses the pattern-associated molecular pattern muramyl dipeptide (MDP) of gram positive and negative bacteria, stimulating the immune response via activation of the transcription factor NF-KB or induction of apoptosis (68). The three common variants found in CD patients all affect NOD2 binding to MDP resulting in loss of function (62). This is thought to cause diminished secretion of antimicrobial peptides, which can lead to dysbiosis and promote a mucosal immune response (69). Functional loss of NOD2 in macrophages may also induce IL-12 and IL-1β expression which leads to the promotion of type 1 immune responses and inflammation (51). Interestingly, not everyone harboring homozygous or compound heterozygous NOD2 variants develop CD (70). Furthermore, NOD2-deficient mice do not spontaneously develop CD-like intestinal inflammation but require a second genetic mutation and specific microbiota (71). This suggests that additional factors, be those microbial, genetic or other environmental factors, are required to promote disease onset in susceptible individuals.

Autophagy-related 16-like protein 1 (ATG16L1) is crucial for normal autophagy function in cells, which is a standard cellular recycling process for protein and organelle turnover that is upregulated during nutrient deprivation or cellular stress signals such as microbial infection. It has been reported that 33.2% of Caucasian CD patients are homozygous for the CDassociated polymorphism rs2241880 (T300A) increasing their risk of developing CD by 2.38-fold (72, 73). This single nucleotide polymorphism is a missense mutation near a caspase cleavage site, making ATG16L1 more sensitive to degradation resulting in diminished autophagy, impaired ability of monocytes to clear invading pathogens and increased production of IL-1β in response to MDP (74). Normal ATG16L1 activity in intestinal epithelial cells (IEC) is important for maintaining the intestinal barrier. The rs2241880 variant in CD patients has been associated with Paneth cell abnormalities, specifically an impaired secretory granule pathway and increased production of proinflammatory mediators (75). Atg16l1-deficiency in IEC has been associated with increased susceptibility to colitis in mice, with increased CD4<sup>+</sup> T cells and increased secretion of proinflammatory cytokines such as TNF- $\alpha$ , interferon- $\gamma$  (IFN- $\gamma$ ) and IL-1β. Thus, diminished autophagy may induce susceptibility through alterations to both immune cell activity and intestinal barrier function.

Changes in *NOD2* and *ATG16L1*, as well as other genes involved in the intestinal epithelial barrier, microbial sensing, and antimicrobial activity in IBD, demonstrate the impact of genetics on the intestinal microbiota. Deficiencies in such genes are a mechanism by which dysbiosis can precede development of IBD and drive intestinal inflammation.

## Dysbiosis in COPD

### Lung Dysbiosis Is Observed in COPD

Chronic inflammatory lung diseases such as COPD and asthma have been associated with dysbiosis of the lung microbiota with the outgrowth of pathogenic bacteria. Mucus hypersecretion and lower respiratory tract infections in COPD have been associated with accelerated decline in lung function, indicating that the lung microbiota plays an important role in COPD pathogenesis (76, 77). The microbiota of the bronchial secretions from COPD patients predominately comprises members of the Proteobacteria, Firmicutes, and Actinobacteria phyla (78). Specifically, studies that have assessed the lung microbiota of COPD patients suggest that their bacterial communities differ from those of healthy individuals, with an expansion of Hemophilus spp. Afipia, Brevundimonas, Curvibacter, Moraxella, Neisseria and Undibacterium spp. of the Proteobacteria phylum, Corynebacterium spp. of the Actinobacteria phylum, Capnocytophaga spp. of the Bacteroidetes phylum, and Leptolyngbya spp. of the Cyanobacteria phylum, as well as a reduction in microbial community diversity compared to healthy individuals (79-82). Patients with more severe COPD have a less diverse lung microbiota but expansion of more pathogenic microbes (78). Furthermore, during acute exacerbations of COPD, the lung microbiota are more unstable; these exacerbations tend to be associated with reduced species diversity, an increased relative abundance of Proteobacteria mainly due to increased Moraxella spp. and a decreased relative abundance of species belonging to the Firmicutes phyla (82, 83). Changes in the core microbiota during acute exacerbations of COPD allow for an expansion of respiratory pathogens including Acinetobacter spp. and Klebsiella spp. highlighting the important role of the commensal lung microbiota in protecting against the colonization of pathogenic microbes (83). It is worth highlighting that the exact nature of dysbiosis within the lung microbiota that occurs during acute exacerbations of COPD is dependent on the cause of the exacerbation, with specific differences noted between bacterial and eosinophilic exacerbations (characterized by bacterial dysbiosis and elevated sputum eosinophils respectively). The characteristic decrease in species diversity and relative abundance of species belonging to the Firmicutes phyla, alongside an increase in species belonging to the Proteobacteria phyla is more pronounced in bacterial exacerbations (82). Bacterial exacerbations also have a significant decrease in Streptococcus spp. and an increase in Hemophilus spp. whilst eosinophilic exacerbations exhibit a decrease in the Proteobacteria:Firmicutes ratio (82). Changes in the composition of the lung microbiota are associated with changes in local inflammatory responses, the most significant being the negative correlation between species diversity and CXCL8, which indicates reduced species diversity is associated with an influx in neutrophils (82). Interestingly, in lung transplants, the microbiota influences the immune response with Firmicutesdominated and Proteobacteria-dominated dysbiosis being proinflammatory, Bacteroidetes-dominated dysbiosis being associated with tissue remodeling, and a balanced microbial community being associated with homeostasis (84). Collectively, these studies show that respiratory microbial communities can regulate inflammation in the lungs.

#### Intestinal Dysbiosis Influences Lung Health

Recent research has shown that the intestinal microbiota is important in reducing the risk of lung inflammation by supporting mucosal immunity. Studies have shown that depletion of intestinal bacteria through antibiotic treatment, renders mice more susceptible to Pneumonia infection and respiratory inflammation (85, 86). Reciprocally, viral and bacterial respiratory infections are able to drive dysbiosis of the intestinal microbiota demonstrating that respiratory inflammation can influence the intestine (87, 88). In mice lacking an intestinal microbiota, their alveolar macrophages have an altered transcriptome which results in decreased phagocytic activity and bacterial killing (85, 86). Dysbiosis of the intestinal microbiota has been shown to influence the composition of the respiratory microbiota through changes in circulating inflammatory cytokines and translocation of intestinal microbiota to the airways Figure 1, although this has only been demonstrated in severe sepsis models and has not yet been established in COPD. It has been shown that segmented filamentous bacteria (SFB) in the intestines induced Th17 responses and IL-22 production in the lungs and protected against respiratory infection with S. pneumoniae with reduced bacterial burden and lung inflammation (89). In a murine model of sepsis, an increased abundance of gut-specific bacteria in the lungs has been observed, with there being greater similarities between the communities found in the intestines and lungs than in sham mice (90). A meta-analysis of sixteen studies found that infection with H. pylori, which colonizes the human gastric mucosa, was associated with an increased risk of COPD (91). During acute exacerbations of COPD there is a significant decrease in categories of bacteria as defined by operational taxonomic units (OTUs) in the intestinal microbiota (83). Beyond assessing H. pylori infection in COPD patients, and a recent study examining the intestinal microbiota in acute exacerbations of COPD in a small group of patients (83), the intestinal microbiota has not been investigated in COPD. This would be of particular interest since cigarette smoking, which is strongly associated with COPD, has been linked with dysbiosis of fecal microbiota in CD patients, characterized by an increase in the relative abundance of Bacteroides and Prevotella (92).

# Cigarette Smoke Exposure Is a Risk Factor for Both COPD and CD

Cigarette smoke is the most important risk factor in COPD, with approximately 80% of COPD patients being past or current smokers. Smoking can have prolonged effects on lung inflammation which can persist years after smoking cessation, despite the slowed decline in lung function and better survival (93). In addition, active smoking is also associated with higher mortality rates in COPD patients (94). Both active and passive smoking is the most well recognized environmental risk factor for CD being associated with a 2-fold increased risk of disease (95). It is linked with early onset of disease, as well as more aggressive disease progression with an increase in the occurrence

of strictures and fistulas and the increased likelihood of a need for surgical intervention (96–98). Smoking may also influence the locus of inflammation, increasing occurrence in the ileum as opposed to the colon (99). An intervention study investigated the effect of quitting smoking on CD severity and found that patients who stopped for at least a year were less likely to relapse (100). The association between smoking and CD highlights the potential crosstalk between the lungs and intestines, although, the possibility that noxious agents from cigarettes can reach the intestines via the oral route cannot be ignored as an additional CD risk mechanism (101).

Smoking also alters the composition of the intestinal microbiota not only in CD patients but also in smokers without IBD (102). Following smoking cessation, the fecal microbiota is altered in non-IBD individuals with an increased relative abundance of species belonging to Firmicutes and Actinobacteria phyla and a decreased relative abundance of species belonging to Bacteroidetes and Proteobacteria phyla (103). CD patients who smoke exhibit intestinal dysbiosis characterized by a higher Bacteroides:Prevotella ratio compared to non-smokers and healthy smokers (92). Animal studies that examined the impact of smoke exposure on the intestinal microbiota found that smoking increased the relative abundance of Clostridium clostridiforme with a decreased relative abundance of Lactoccoci spp. and Ruminococcus albus of the Firmicutes phylum and Enterobacteriaceae spp. in the cecum compared to control mice (3, 104). Collectively, these studies suggest that dysbiosis of the intestinal microbiota could be another mechanism by which cigarette smoke might increase the risk of CD development.

A study of chronic smoke exposure in mice compared the microbiota across the ileum, cecum, and distal colon finding changes in microbiota (105). The authors described increased activity of *Lachnospiraceae* spp. in the cecum and colon, which is of particular interest since it has been reported that *Lachnospiraceae* spp. can promote macrophage recruitment to the colon (106). This study also demonstrated that chronic smoke exposure may impact the intestinal microbiota by altering mucus profiles and the local immune environment. They found that cigarette smoke increased the secretion of the two major ileal mucins, Muc2 and Muc3, and enhanced the cell surface expression of the anti-adhesive Muc4 (105). However, at present it is unclear if these changes are a result of dysbiosis, or of cigarette smoke itself.

Cigarette smoke has been found to increase intestinal barrier permeability in the ileum, but not the colon (107). These changes are associated with intestinal villi atrophy, bacterial translocation and abnormal tight junction proteins, with evidence that they were mediated through NF- $\kappa$ B signaling (107, 108). Cigarette smoke has also been found to alter Paneth cell function in mice through reduced antimicrobial peptide expression and reduced bactericidal capacity, which leaves mice more susceptible to bacterial infection (108). Changes in the ileum, but not the colon, may explain why cigarette smoke increases the risk of developing CD whilst conceivably offering protection against UC (95). Certain susceptibility genes have been associated with epithelial barrier defects in CD patients who smoke. A promoter variant in the gene encoding the aryl hydrocarbon receptor (AHR) has been linked to increased risk of intestinal hyperpermeability, with cigarette smoking further increasing this risk (109). From an immunological perspective, cigarette smoke induces an IL-17 response with increases in Th17 cells and neutrophils in the lungs and circulation (110). This enhances intestinal Th17 cells and neutrophils, as well as IL-17-producing type 3 innate lymphoid cells (ILC3s), in a manner that is dependent on neutrophil recruitment via IL-17A (110). Thus, in addition to changes in the intestinal epithelial barrier, cigarette smoke can promote intestinal inflammation which is already augmented in individuals who are genetically susceptible to IBD.

## Non-bacterial Microbiota in IBD and COPD

To date most studies examining the microbiota in IBD and COPD have focused on bacteria, however, the microbiota also encompasses fungi and viruses. Whilst this review primarily discusses the role of bacterial dysbiosis in IBD and COPD these other microorganisms cannot be ignored. There is evidence to suggest that there are shifts in the intestinal viral and fungal communities during IBD and alterations to pulmonary communities in COPD. However, considerably more research is needed to determine the role of viruses and fungi in the gut-lung axis especially in the context of IBD and COPD.

#### The Virome and Mycobiome May Contribute to IBD

The intestinal virome is predominately comprised of bacteriophages (111), therefore the interactions between viruses and bacteria during IBD could play a role in dysbiosis and disease pathogenesis (112). The most abundant bacteriophages of the intestine include the Caudovirales order and Microviridae family (113–115) and perhaps not surprisingly, viral dysbiosis in IBD patients is characterized by an increase in Caudovirales species (115, 116). This expansion is associated with reduced bacterial diversity and does not appear to occur secondary to changes in bacterial populations suggesting that the virome may contribute to bacterial dysbiosis and inflammation in IBD (115). However, virome research in IBD is in its infancy and more studies are required to elucidate how changes in intestinal viruses may impact upon other intestinal microorganisms and intestinal inflammation.

A potential role for fungi in IBD pathogenesis was first proposed in 1988 when antibodies directed against Saccharomyces cerevisiae were identified in the blood of CD patients (117). Furthermore, several IBD susceptibility genes are involved in anti-fungal immune responses such as CARD9 and CLEC7A (61). Fungal dysbiosis in IBD has been characterized by an increased Basidiomycota:Ascomycota ratio, decreased proportion of Saccharomyces cerevisiae and an increased proportion of Candida albicans (118). Similar to the virome, further research is needed to understand how changes in the mycobiome during IBD may impact upon other microorganisms and inflammation.

### The Virome and Mycobiome in COPD

Similar to the gastrointestinal tract the respiratory tract consists of bacteriophages and eukaryotic viruses (119, 120). COPD

patients have a heightened viral load in their lungs with an increased abundance of influenza, cytomegalovirus, and Epstein-Barr virus, the latter of which has been associated with pulmonary fibrosis, a feature of COPD (121–123). More non-targeted approaches are required to define other viruses that may be involved in COPD pathogenesis and dysbiosis.

The most abundant fungi in healthy lungs are of the Davidellaceae family and the genera Cladosporium, Eurotium, Penicillium, and Aspergillus, although many other species including Candida spp. are present as well (124). Compared to healthy individuals, COPD patients have an increased relative abundance of Candida spp. in their lungs (125). Furthermore, the enhanced abundance of Aspergillus, Candida, Phialosimplex, Penicillium, Cladosporium, and Eutypella has been associated with severe exacerbations of COPD (126). Of the chronic pulmonary diseases, COPD is one of the least studied in the context of the mycobiome and more research is required to understand how the mycobiome is altered in COPD patients. Furthermore, the relationship between the nonbacterial microbes of the lung and gut and their role in the gut-lung axis have been poorly considered and are an area for future research.

## FACTORS LINKING IBD AND COPD Dietary-Derived Metabolites Are Protective in IBD and COPD

Certain macro- and micro-nutrients have been inversely associated with the development of CD (127), with dietary fiber being the most extensively researched. Soluble-fiber from fruits and vegetables as opposed to insoluble fiber from whole grains and cereals has shown protection against CD (128). Non-digestible carbohydrates are fermented by saccharolytic bacteria in the gastrointestinal tract into metabolites known as short-chain fatty acids (SCFA), which include acetate, propionate, and butyrate. Acetate and propionate are produced by Bacteroidetes and butyrate by Firmicutes and these SCFA can be immunomodulatory by preventing the transcription of proinflammatory mediators. Butyrate in particular is an energy source for IEC thereby promoting intestinal barrier integrity (129), and thus it is not surprising that a decrease in butyrateproducing bacteria is a characteristic of intestinal dysbiosis in IBD (35). Experimental models of IBD have shown that dietary SCFA reduce inflammation, specifically via decreases in proinflammatory mediators such as  $TNF-\alpha$  and nitric oxide synthase which correlates with increased concentrations of butyrate and propionate in the luminal contents of the intestines (130, 131). However, the efficacy of SCFA in IBD patients has been brought into question due to the reduced responsiveness of their peripheral blood mononuclear cells to n-butyrate following toll-like receptor-2 (TLR-2) activation (132). Furthermore, for UC patients, butyrate enemas have shown no clinical benefit (133). This could be due to the finding that monocarboxylate transporter 1, which is responsible for the uptake of butyrate in the intestine, is downregulated in response to proinflammatory cytokines and its expression is reduced in the inflamed mucosa of IBD patients and in a rat model of colitis (134). Interestingly, studies examining the fecal contents of CD patients found that disease activity or localization was not affected by SCFA concentration, although CD disease activity was inversely correlated to levels of the medium-chain fatty acid hexanoate (135, 136). Contradictory to these studies, a clinical trial examining the effects of soluble fiber supplements showed that high fiber could reduce disease activity index in CD patients (137). Discrepancies in research may reflect the heterogeneity of CD as well as differences in diets, and thus, further research is required to elucidate the effects of the different components of diet on intestinal inflammation.

It is now known that SCFA can have immunomodulatory effects beyond the intestines where they are produced, promoting anti-inflammatory responses elsewhere in the body. Dysregulated SCFA production and absorption has been implicated in a variety of neurological, metabolic, allergic, and autoimmune diseases (138-141). In the context of the lungs, increased dietary fiber intake is associated with improved lung function in the general population and a reduced risk of developing COPD (142, 143). Increased intake of vegetables, which are high in soluble fiber, is associated with improved COPD symptoms such as breathlessness, as well as reduced risk of developing COPD (144). Similar to vegetables, high dietary fruit intake has also been associated with improved COPD outcomes and reduced incidence of COPD (145-148). Despite this correlation between increased dietary fiber intake and protection against COPD, there have been few reports on the efficacy of SCFA specifically in COPD pathogenesis highlighting that this is an understudied area worthy of further research. High fiber dietproducing SCFA have been shown to be immunomodulatory in asthma responses by enhancing the production of dendritic cells that seed the lungs but have an impaired ability to promote pathogenic type 2 immune responses (149). A more recent study building on these findings has shown that high fiber diets protect against influenza by enhancing the generation of Ly6c<sup>-</sup> patrolling monocytes from progenitors. This led to an increase in alternatively activated macrophages in lungs and restrained neutrophil recruitment while simultaneously enhancing influenza-specific CD8 T cells responses (150). Similar mechanisms may be associated with the protection created by high fiber diets in COPD where neutrophils play a key pathogenic role in the inflamed lungs. COPD patients exhibit poor responses to influenza vaccination (151, 152) and thus, diet modulation may be a mechanism to improve vaccination outcomes for this susceptible population. In all, modulation of intestinal microbiota with high fiber diets might be beneficial to IBD and COPD patients (Figure 1), however, further research is required to determine how efficacious this strategy would be.

## Dietary Fat Can Alter the Gut Microbiota and Influence Disease

Foods high in saturated fat or "Western" diets have been associated with a variety of autoimmune and chronic

inflammatory disease including IBD and COPD (142, 153, 154). A "Western" diet can influence the composition of the intestinal microbiota, promote intestinal barrier permeability, and enhance inflammation (155-157). Generally, fat intake is able to induce proinflammatory responses through the increase in cytokines, including TNF- $\alpha$  and IL-6, and neutrophils in circulation, all of which play a pathogenic role in IBD and COPD (155, 158). Intestinal dysbiosis induced by fat intake is characterized by an increased Firmicutes:Bacteroidetes ratio and the promotion of endotoxemia, which induces intestinal barrier hyperpermeability (156, 157, 159). In CD patients, the changes induced by a high fat diet that contribute to dysbiosis include increased intestinal barrier permeability, reduced mucus layer thickness and increased NOD2, TLR5, and carcinoembryonic antigen-related cell adhesion molecule 6 (CEABAC6) expression, all of which allow for AIEC colonization (49, 160). Generally, a high fat diet tends to be associated with a low fiber diet, and thus, a "Western" diet may contribute to IBD and COPD pathogenesis not only through direct proinflammatory mechanisms but also indirectly through a reduction in the anti-inflammatory benefits of SCFA Figure 1.

## Vitamin D Alters the Microbiota and May Have Therapeutic Benefits in CD and COPD

Vitamin D deficiency commonly occurs in IBD patients and has been associated with diagnosis and the need for surgical intervention (161). In keeping with this, colitis-prone IL-10deficient mice exhibit a decline in vitamin D receptor (VDR) expression that correlates with colitis symptoms (162). In addition, mouse models unable to produce the active form of vitamin D, 1,25-dihydroxycholecalciferol [1,25(OH)<sub>2</sub>D<sub>3</sub>], or lacking the VDR are more susceptible to DSS-induced colitis and this is associated with intestinal dysbiosis characterized by an increase in species of the Proteobacteria phylum and a decrease in species of the Firmicutes phylum (163, 164). VDR signaling regulates numerous antimicrobial processes including the expression of  $\beta$ -defensins, cathelicidin antimicrobial peptides, and ATG16L1 (162). In IBD patients, reduced ATG16L1 expression due to deficiency in VDR signaling promotes an overrepresentation of intestinal Bacteroides and a decrease in butyrate-producing bacteria (162). Interestingly, treating human IEC with butyrate upregulates VDR expression, a phenomenon that also translated to IL-10-deficient mice that were given butyrate, suggesting a close link between the microbiota and vitamin D signaling (162). Vitamin D supplementation in CD patients results in an increase in Firmicutes species correcting some of the dysbiosis that occurs in CD (165). In COPD patients, increased vitamin D intake is positively associated with improved lung function, and like IBD, vitamin D deficiency is associated with COPD (166, 167). This could relate to the effect of vitamin D on the intestinal microbiota. Additionally, vitamin D plays a role in macrophage activation and shaping the lung microbiota promoting reduced bacterial richness (168, 169). VDR-deficient mice exhibit increased inflammation in the lungs with up-regulation of matrix metalloproteinase-2 (MMP-2), MMP-9, and MMP-12, the development of emphysema and a decline in lung function mimicking COPD in humans (170). Studies where mice are fed vitamin D have shown that vitamin D reduces the abundance of respiratory pathobionts, such as *Pseudomonas*, and increases the secretion of murine  $\beta$ -defensin-2 in the lungs (171). To date, very little has been done examining the impact of vitamin D supplementation on the lung microbiota. One study in cystic fibrosis patients found that the sputum of vitamin D insufficient patients was enriched for Bacteroides and there was a significant difference between the lung microbiota of these patients and those who were vitamin D sufficient (172). It is clear that the immunomodulatory and microbiota-regulating effects of vitamin D can strongly influence inflammation in both the intestine and the lungs. Given that vitamin D deficiency is associated with both IBD and COPD, vitamin D supplementation should be trialed more extensively in these patient cohorts, specifically in patients with comorbid IBD and COPD.

# THERAPEUTIC TARGETING OF THE INTESTINAL MICROBIOTA

## Antibiotics for IBD: the Yin and Yang

Early-life antibiotic treatment has been associated with early onset CD via enhanced pathogenicity of helper T cells (173). This link is particularly strong in children who have received multiple doses of antibiotics or antibiotics during their first year of life (174, 175). Antibiotics cause dysbiosis of intestinal microbiota and a reduction in bacterial diversity, and this may be a potential mechanism by which antibiotic therapy in early life could result in the development of CD in genetically susceptible individuals (176). Furthermore, short-term antibiotic treatment may have prolonged effects, up to at least 2 years post-therapy (177). In infancy, the intestinal microbiota between individuals can be highly variable, before converging to more similar phyla in adulthood (178). This may in part explain the profound impact of antibiotic treatment on the microbiota in early childhood.

While antibiotics given in early life may promote CD, they have been used to treat CD with varying levels of success, and this appears to be dependent on disease location and severity as well as the type of antibiotic (179). Differences in populations of commensal bacteria between the ileum and colon most likely contribute to the lack of response of ileitis patients to a variety of antibiotics (179, 180). Patients who had undergone ileal resection and were treated with metronidazole, an antibiotic that is usually ineffective in patients with ileitis (180), exhibited a delay in symptomatic recurrence (181). A systematic review of antibiotic therapy in CD patients found that antibiotics likely have a modest effect that may not be clinically relevant (182). Furthermore, to maintain antibiotic treatment efficacy and prevent relapse, longterm treatment is required, as with all therapies for CD (183, 184). All in all, antibiotics in early life may increase the risk of CD development in susceptible individuals but may be beneficial as a therapeutic in established disease.

Unlike CD, antibiotic exposure is not associated with an increased risk of developing UC (173) and indeed antibiotics are

effective as an adjunctive to conventional therapies, including corticosteroids and 5-aminosalycilic acid (185). This emphasizes the pathogenic role that bacterial dysbiosis plays in UC and suggests that more therapeutics targeting both inflammation and dysbiosis could benefit a large proportion of UC patients.

# Antibiotics for COPD Are Used to Manage Disease Exacerbations

Persistent and recurrent infections contribute to the progression of COPD through the induction of further chronic inflammation. Antibiotics are commonly used to treat acute exacerbations of COPD, however, the efficacy of antibiotic treatment in mild to moderate exacerbations is still in debate (186, 187). Treatment of COPD patients with antibiotics enhances respiratory microbiota diversity, decreases the relative abundance of Proteobacteria species and increases the relative abundance of Firmicutes species. These changes somewhat correct the dysbiosis associated with COPD, an effect that is maintained post-therapy (80, 82). Contrary to antibiotic therapy, corticosteroids are associated with a decrease in species diversity and an increase of Proteobacteria over Firmicutes corresponding to an increase in Hemophilus spp. and Moraxella spp. and a decrease in Streptococcus spp. changes that are associated with COPD pathogenesis (82). The differing effects of antibiotics and corticosteroids on the lung microbiota suggest that antibiotics are able to partially restore lung microbial communities whilst corticosteroids may further promote dysbiosis. Macrolides are the most commonly prescribed antibiotics for COPD due to both anti-inflammatory and immunomodulatory effects, however, the mechanisms by which macrolides exert these effects have not been elucidated (188). Relatively few COPD patients are treated with antibiotics long-term (189), however, the few studies that have been conducted suggest that prophylactic antibiotic treatment can reduce exacerbations in COPD patients (190). Nonetheless, further studies are required to understand whether long-term antibiotics are efficacious and the impact they have on the lung and intestinal microbiota.

## Fecal Microbiota Transplants, a Possible Treatment Strategy

Fecal microbial transplants are effective in the treatment of Clostridium difficile infection, an intestinal disease that is linked to intestinal bacterial dysbiosis (191). Success is associated with an expansion in bacterial diversity including increases in Bacteroidetes, Firmicutes and other butyrate-producing bacteria and a decrease in Proteobacteria (192). In patients with UC, fecal microbial transplants have some initial benefit by promoting a change in colonic microbiota at the phylum level with a decrease in the relative abundance of Proteobacteria species and an increase in Bacteroidetes species. While this partially corrects the dysbiotic changes that are associated with UC, these changes were not prolonged and did not translate to a vast clinical improvement (193). Another study where patients were treated with antibiotics prior to fecal microbial transplant had better clinical outcomes resulting in remission (194). Patients who respond to fecal microbial transplants have been characterized by distinct microbial profiles compared to non-responders. Alterations in microbiota in responders is characterized by an increase in bacterial diversity as well as a shift in composition toward that of the donor feces (195, 196). Lack of response is associated with the presence of *Fusobacterium* spp. and *Sutterella* spp. suggesting that patients should be screened prior to treatment (196). Studies examining the efficacy of fecal microbial transplants for UC have yielded variable results, which could be due to a lack of consistency in methodology as well as the heterogeneous nature of UC and more work is required to establish if this could be an efficacious treatment strategy.

In the context of CD, there has been far less research into the efficacy of fecal microbial transplants and at present it is not clear if this could be a management strategy for CD patients (197, 198). With respect to COPD, no fecal or respiratory microbiota transplant studies have been conducted and thus it is not yet known if these strategies could be a viable option for this disease.

## CONCLUSION

Microbial dysbiosis has a pivotal role in the development of IBD and COPD impacting on the intestinal and respiratory epithelial barriers and promoting damaging immune responses. Circulating microbial products and their metabolites are altered during dysbiosis and likely represent a significant component of the gut-lung axis. Shifts in these factors, where they may be produced at one site and act at another, provides a mechanism for organ crosstalk in disease and the comorbid presentation of IBD and COPD. Currently there are no curative treatments for either disease. However, elucidating the mechanisms by which the intestinal and respiratory microbiota drive inflammation and promote changes in mucosal epithelial barriers could provide new insights into disease pathogenesis and help

### REFERENCES

- Girosi D, Bellodi S, Sabatini F, Rossi GA. The lung and the gut: common origins, close links. *Paediatr Respir Rev.* (2006) 7:S235–9. doi: 10.1016/j.prrv.2006.04.192
- Gajendran M, Loganathan P, Catinella AP, Hashash JG. A comprehensive review and update on Crohn's disease. *Dis Mon.* (2018) 64:20–57. doi: 10.1016/j.disamonth.2017.07.001
- Savin Z, Kivity S, Yonath H, Yehuda S. Smoking and the intestinal microbiome. Arch Microbiol. (2018) 200:677–84. doi: 10.1007/s00203-018-1506-2
- Head KA, Jurenka JS. Inflammatory bowel disease part I: ulcerative colitis– pathophysiology and conventional and alternative treatment options. *Altern Med Rev.* (2003) 8:247–83.
- Khor B, Agnès G, Ramnik JX. Genetics and pathogenesis of inflammatory bowel disease. *Nature*. (2011) 474:307–17. doi: 10.1038/nature 10209
- McGovern DPB, Jones MR, Taylor KD, Marciante K, Yan X, Dubinsky M, et al. Fucosyltransferase 2 (FUT2) non-secretor status is associated with Crohn's disease. *Hum Mol Genet*. (2010) 19:3468–76. doi: 10.1093/hmg/ddq248
- Bernstein CN, Loftus EV, Ng SC, Lakatos PL, Moum B. Hospitalisations and surgery in Crohn's disease. *Gut.* (2012) 61:622–9. doi: 10.1136/gutjnl-2011-301397

to improve current treatment strategies and identify novel therapeutic targets. These may include approaches that target the microbiota such as diet, antibiotics or fecal microbiota transplants which may be able to modulate inflammation in the intestines and lungs. Most microbiota studies have focused on either IBD or COPD and have largely ignored patients with both diseases. Additionally, there are no reports of gut microbiota alterations in COPD, with published studies focused solely on the lung metagenome. Future studies into patients that harbor COPD, as well as patients with both mucosal inflammatory diseases will provide a more complete understanding of the microbiota in the gut-lung axis in health and disease. Furthermore, increasing awareness and understanding of the links between IBD and COPD will improve clinical management and more timely detection of comorbid disease in affected patients.

## **AUTHOR CONTRIBUTIONS**

AR, NH, and MH conceived and planned the review. AR wrote the first draft of the manuscript. ET, NH, and MH provided editorial comment. All authors contributed to the article and approved the submitted version.

## FUNDING

This work was supported by grant funding from the National Health and Medical Research Council of Australia (APP1147267 to MH, APP1141208 to MH, and ET and APP1140313 to NH). AR was supported by a scholarship from the Australia Government administered by Monash University. NH is supported by a National Health and Medical Research Council (NHMRC) of Australia SRF-B fellowship.

- Cottone M, Scimeca D, Mocciaro F, Civitavecchia G, Perricone G, Orlando A. Clinical course of ulcerative colitis. *Dig Liver Dis.* (2008) 40:S247–52. doi: 10.1016/S1590-8658(08)60533-2
- Barnes PJ. Inflammatory mechanisms in patients with chronic obstructive pulmonary disease. J Allergy Clin Immunol. (2016) 138:16–27. doi: 10.1016/j.jaci.2016.05.011
- Lamprecht B, McBurnie MA, Vollmer WM, Gudmundsson G, Welte T, Nizankowska-Mogilnicka E, et al. COPD in never smokers: results from the population-based burden of obstructive lung disease study. *Chest.* (2011) 139:752–63. doi: 10.1378/chest.10-1253
- Lopez A. Measuring the global burden of disease. N Engl J Med. (2013) 369:448–57. doi: 10.1056/NEJMra1201534
- Siddiqui FM, Diamond JM. Lung transplantation for chronic obstructive pulmonary disease: past, present, and future directions. *Curr Opin Pulm Med.* (2018) 24:199–204. doi: 10.1097/MCP.0000000000 000452
- Ekbom A, Brandt L, Granath F, Löfdahl C-G, Egesten A. Increased risk of both ulcerative colitis and crohn's disease in a population suffering from COPD. *Lung.* (2008) 186:167–72. doi: 10.1007/s00408-008-9080-z
- Duricova D, Pedersen N, Elkjaer M, Gamborg M, Munkholm P, Jess T. Overall and cause-specific mortality in Crohn's disease: a metaanalysis of population-based studies. *Inflamm Bowel Dis.* (2010) 16:347–53. doi: 10.1002/ibd.21007

- Vutcovici M, Bitton A, Ernst P, Kezouh A, Suissa S, Brassard P. Inflammatory bowel disease and risk of mortality in COPD. *Eur Respir J.* (2016) 47:1357–64. doi: 10.1183/13993003.01945-2015
- Lee J, Im JP, Han K, Park S, Soh H, Choi K, et al. Risk of inflammatory bowel disease in patients with chronic obstructive pulmonary disease: a nationwide, population-based study. *World J Gastroenterol.* (2019) 25:6354– 64. doi: 10.3748/wjg.v25.i42.6354
- Jess T, Loftus EV, Harmsen WS, Zinsmeister AR, Tremaine WJ, Melton LJ, et al. Survival and cause specific mortality in patients with inflammatory bowel disease: a long term outcome study in Olmsted County, Minnesota, 1940-2004. *Gut.* (2006) 55:1248–54. doi: 10.1136/gut.2005.079350
- Brassard P, Vutcovici M, Ernst P, Patenaude V, Sewitch M, Suissa S, et al. Increased incidence of inflammatory bowel disease in québec residents with airway diseases. *Eur Respir J.* (2015) 45:962–8. doi: 10.1183/09031936.00079414
- Vutcovici M, Brassard P, Bitton A. Inflammatory bowel disease and airway diseases. World J Gastroenterol. (2016) 22:7735–41. doi: 10.3748/wjg.v22.i34.7735
- Keely S, Talley NJ, Hansbro PM. Pulmonary-intestinal cross-talk in mucosal inflammatory disease. *Mucosal Immunol.* (2011) 5:7–18. doi: 10.1038/mi.2011.55
- Van Der Heide KF, Dijkstra AA, Weersma MJR, Albersnagel NF, Van Der Logt JE, Faber HK, et al. Effects of active and passive smoking on disease course of Crohn's disease and ulcerative colitis. *Inflamm Bowel Dis.* (2009) 15:1199–207. doi: 10.1002/ibd.20884
- Ji XQ, Ji YB, Wang SX, Zhang CQ, Lu DG. Alterations of pulmonary function in patients with inflammatory bowel diseases. *Ann Thorac Med.* (2016) 11:249–53. doi: 10.4103/1817-1737.191877
- Caramori G, Casolari P, Barczyk A, Durham AL, Di Stefano A, Adcock I. COPD immunopathology. *Semin Immunopathol.* (2016) 38:497–515. doi: 10.1007/s00281-016-0561-5
- Pickard JM, Zeng MY, Caruso R, Núñez G. Gut microbiota: Role in pathogen colonization, immune responses, and inflammatory disease. *Immunol Rev.* (2017) 279:70–89. doi: 10.1111/imr.12567
- Borger JG, Lau M, Hibbs ML. The influence of innate lymphoid cells and unconventional T cells in chronic inflammatory lung disease. *Front Immunol.* (2019) 10:1597. doi: 10.3389/fimmu.2019.01597
- Friedrich M, Pohin M, Powrie F. Cytokine networks in the pathophysiology of inflammatory bowel disease. *Immunity*. (2019) 50:992–1006. doi: 10.1016/j.immuni.2019.03.017
- Martini E, Krug SM, Siegmund B, Neurath MF, Becker C. Mend your fences: the epithelial barrier and its relationship with mucosal immunity in inflammatory bowel disease. *Cell Mol Gastroenterol Hepatol.* (2017) 4:33–46. doi: 10.1016/j.jcmgh.2017.03.007
- Rutten EPA, Lenaerts K, Buurman WA, Wouters EFM. Disturbed intestinal integrity in patients with COPD: effects of activities of daily living. *Chest.* (2014) 145:245–52. doi: 10.1378/chest.13-0584
- Sprooten RTM, Lenaerts K, Braeken DCW, Grimbergen I, Rutten EP, Wouters EFM, et al. Increased small intestinal permeability during severe acute exacerbations of COPD. *Respiration*. (2018) 95:334–42. doi: 10.1159/000485935
- Gill S, Pop M, Deboy R, Eckburg P, Turnbaugh P, Samuel B, et al. Metagenomic analysis of the human distal gut microbiome. *Science*. (2006) 312:1355–9. doi: 10.1126/science.1124234
- Peterson D, Frank D, Pace N, Gordon J. Metagenomic approaches for defining the pathogenesis of inflammatory bowel diseases. *Cell Host Microbe*. (2008) 3:417–27. doi: 10.1016/j.chom.2008.05.001
- Maynard CL, Elson CO, Hatton RD, Weaver CT. Reciprocal interactions of the intestinal microbiota and immune system. *Nature*. (2012) 489:231–41. doi: 10.1038/nature11551
- Qin J, Li R, Raes J, Arumugam M, Burgdorf KS, Manichanh C, et al. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature*. (2010) 464:59–65. doi: 10.1038/nature08821
- Joossens M, Huys G, Cnockaert M, De Preter V, Verbeke K, Rutgeerts P, et al. Dysbiosis of the faecal microbiota in patients with Crohn's disease and their unaffected relatives. *Gut.* (2011) 60:631–7. doi: 10.1136/gut.2010.223263
- 35. Machiels K, Joossens M, Sabino J, De Preter V, Arijs I, Eeckhaut V, et al. A decrease of the butyrate-producing species *Roseburia hominis* and

Faecalibacterium prausnitzii defines dysbiosis in patients with ulcerative colitis. Gut. (2014) 63:1275–83. doi: 10.1136/gutjnl-2013-304833

- Kaufman A, Rosenstreich DL. Mast cell heterogeneity in chronic idiopathic urticaria. Ann Allergy. (1990) 65:367–73.
- Collins SM, Surette M, Bercik P. The interplay between the intestinal microbiota and the brain. *Nat Rev Microbiol.* (2012) 10:735. doi: 10.1038/nrmicro2876
- Abrahamsson TR, Jakobsson HE, Andersson AF, Björkstén B, Engstrand L, Jenmalm MC. Low gut microbiota diversity in early infancy precedes asthma at school age. *Clin Exp Allergy*. (2014) 44:842–50. doi: 10.1111/cea.12253
- Hua X, Goedert JJ, Pu A, Yu G, Shi J. Allergy associations with the adult fecal microbiota: analysis of the american gut project. *EBioMedicine*. (2016) 3:172–9. doi: 10.1016/j.ebiom.2015.11.038
- Maeda Y, Kurakawa T, Umemoto E, Motooka D, Ito Y, Gotoh K, et al. Dysbiosis contributes to arthritis development via activation of autoreactive T cells in the intestine. *Arthritis Rheumatol.* (2016) 68:2646–61. doi: 10.1002/art.39783
- Li X, Watanabe K, Kimura I. Gut microbiota dysbiosis drives and implies novel therapeutic strategies for diabetes mellitus and related metabolic diseases. *Front Immunol.* (2017) 8:1882. doi: 10.3389/fimmu.2017.018820
- Hidalgo-Cantabrana C, Gómez J, Delgado S, Requena-López S, Queiro-Silva R, Margolles A, et al. Gut microbiota dysbiosis in a cohort of patients with psoriasis. *Br J Dermatol.* (2019) 181:1287–95. doi: 10.1111/bjd.17931
- Beck JM, Young VB, Huffnagle GB. The microbiome of the lung. *Transl Res.* (2012) 160:258–66. doi: 10.1016/j.trsl.2012.02.005
- 44. Morris A, Beck J, Schloss P, Campbell T, Crothers K, Curtis J, et al. Comparison of the respiratory microbiome in healthy nonsmokers and smokers. *Am J Respir Crit Care Med.* (2013) 187:1067–75. doi: 10.1164/rccm.201210-1913OC
- Prosberg M, Bendtsen F, Vind I, Petersen AM, Gluud LL. The association between the gut microbiota and the inflammatory bowel disease activity: a systematic review and meta-analysis. *Scand J Gastroenterol.* (2016) 51:1407– 15. doi: 10.1080/00365521.2016.1216587
- Zhang S, Cao X, Huang H. Sampling strategies for three-dimensional spatial community structures in IBD microbiota research. Front Cell Infect Microbiol. (2017) 7:51. doi: 10.3389/fcimb.2017.00051
- Manichanh C, Rigottier-Gois L, Bonnaud E, Gloux K, Pelletier E, Frangeul L, et al. Reduced diversity of faecal microbiota in Crohn's disease revealed by a metagenomic approach. *Gut.* (2006) 55:205–11. doi: 10.1136/gut.2005.073817
- Kang ES, Denman SS, Morrison SM, Yu SZ, Dore SJ, Leclerc SM, et al. Dysbiosis of fecal microbiota in Crohn's disease patients as revealed by a custom phylogenetic microarray. *Inflamm Bowel Dis.* (2010) 16:2034–42. doi: 10.1002/ibd.21319
- Martinez-Medina LM, Aldeguer LX, Gonzalez-Huix LF, Acero LD, Garcia-Gil LJ. Abnormal microbiota composition in the ileocolonic mucosa of Crohn's disease patients as revealed by polymerase chain reactiondenaturing gradient gel electrophoresis. *Inflamm Bowel Dis.* (2006) 12:1136– 45. doi: 10.1097/01.mib.0000235828.09305.0c
- Kim SC, Tonkonogy SL, Albright CA, Tsang J, Balish EJ, Braun J, et al. Variable phenotypes of enterocolitis in interleukin 10-deficient mice monoassociated with two different commensal bacteria. *Gastroenterology*. (2005) 128:891–906. doi: 10.1053/j.gastro.2005.02.009
- Biswas A, Liu YJ, Hao L, Mizoguchi A, Salzman NH, Bevins CL, et al. Induction and rescue of Nod2-dependent Th1-driven granulomatous inflammation of the ileum. *Proc Natl Acad Sci USA*. (2010) 107:14739–44. doi: 10.1073/pnas.1003363107
- Maharshak N, Packey CD, Ellermann M, Manick S, Siddle JP, Huh EY, et al. Altered enteric microbiota ecology in interleukin 10-deficient mice during development and progression of intestinal inflammation. *Gut Microbes*. (2013) 4:316–24. doi: 10.4161/gmic.25486
- Al Nabhani Z, Lepage P, Mauny P, Montcuquet N, Roy M, Le Roux K, et al. Nod2 deficiency leads to a specific and transmissible mucosa-associated microbial dysbiosis which is independent of the mucosal barrier defect. J Crohn's Colitis. (2016) 10:1428–36. doi: 10.1093/ecco-jcc/jjw095
- Mukhopadhya I, Hansen R, El-Omar EM, Hold GL. IBD-what role do Proteobacteria play? Nat Rev Gastroenterol Hepatol. (2012) 9:219–30. doi: 10.1038/nrgastro.2012.14

- Meconi S, Vercellone A, Levillain F, Payré B, Al Saati T, Capilla F, et al. Adherent-invasive *Escherichia coli* isolated from Crohn's disease patients induce granulomas *in vitro*. *Cell Microbiol*. (2007) 9:1252–61. doi: 10.1111/j.1462-5822.2006.00868.x
- Janssen CEI, Rose CD, De Hertogh G, Martin TM, Bader Meunier B, Cimaz R, et al. Morphologic and immunohistochemical characterization of granulomas in the nucleotide oligomerization domain 2–related disorders Blau syndrome and Crohn disease. *J Allergy Clin Immunol.* (2012) 129:1076– 84. doi: 10.1016/j.jaci.2012.02.004
- 57. Glasser A, Boudeau J, Barnich N, Perruchot M, Colombel J, Darfeuille-Michaud A. Adherent invasive *Escherichia coli* strains from patients with crohn's disease survive and replicate within macrophages without inducing host cell death. *Infect Immun.* (2001) 69:5529–37. doi: 10.1128/IAI.69.9.5529-5537.2001
- Craven M, Egan CE, Dowd SE, McDonough SP, Dogan B, Denkers EY, et al. Inflammation drives dysbiosis and bacterial invasion in murine models of ileal Crohn's Disease (inflammation drives ileal dysbiosis). *PLoS ONE*. (2012) 7:e41594. doi: 10.1371/journal.pone.0041594
- Dobranowski PA, Tang C, Sauvé JP, Menzies SC, Sly LM. Compositional changes to the ileal microbiome precede the onset of spontaneous ileitis in SHIP deficient mice. *Gut Microbes.* (2019) 10:578–98. doi: 10.1080/19490976.2018.1560767
- Frank DN, St. Amand AL, Feldman RA, Boedeker EC, Harpaz N, Pace NR. Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. *Proc Natl Acad Sci USA*. (2007) 104:13780–5. doi: 10.1073/pnas.0706625104
- Jostins L, Ripke S, Weersma RK, Duerr RH, McGovern DP, Hui KY, et al. Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature*. (2012) 491:119–24. doi: 10.1038/nature11582
- Hugot JP, Chamaillard M, Zouali H, Lesage S, Cézard JP, Belaiche J, et al. Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature*. (2001) 411:599–603. doi: 10.1038/35079107
- Ogura Y, Lala S, Xin W, Smith E, Dowds TA, Chen FF, et al. Expression of NOD2 in paneth cells: a possible link to Crohn's ileitis. *Gut.* (2003) 52:1591-7. doi: 10.1136/gut.52.11.1591
- Yazdanyar S, Nordestgaard BG. NOD2/CARD15 genotype and common gastrointestinal diseases in 43,600 individuals. *J Intern Med.* (2010) 267:228– 36. doi: 10.1111/j.1365-2796.2009.02137.x
- 65. Büning C, Genschel J, Bühner S, Krüger S, Kling K, Dignass A, et al. Mutations in the NOD2/CARD15 gene in Crohn's disease are associated with ileocecal resection and are a risk factor for reoperation. *Aliment Pharmacol Ther.* (2004) 19:1073–8. doi: 10.1111/j.1365-2036.2004.01967.x
- 66. Seiderer J, Schnitzler F, Brand S, Staudinger T, Pfennig S, Herrmann K, et al. Homozygosity for the CARD15 frameshift mutation 1007fs is predictive of early onset of Crohn's disease with ileal stenosis, entero-enteral fistulas, and frequent need for surgical intervention with high risk of re-stenosis. *Scand J Gastroenterol.* (2006) 41:1421–32. doi: 10.1080/00365520600 703900
- Gazouli M, Pachoula I, Panayotou I, Mantzaris G, Chrousos G, Anagnou N, et al. NOD2/CARD15, ATG16L1 and IL23R gene polymorphisms and childhood-onset of Crohn's disease. *World J Gastroenterol.* (2010) 16:1753–8. doi: 10.3748/wjg.v16.i14.1753
- Rogler G. The effects of NOD2/CARD15 mutations on the function of the intestinal barrier. J Crohn's Colitis. (2007) 1:53–60. doi: 10.1016/j.crohns.2007.08.008
- Wehkamp J, Harder J, Weichenthal M, Schwab M, Schäffeler E, Schlee M, et al. NOD2 (CARD15) mutations in Crohn's disease are associated with diminished mucosal α-defensin expression. *Gut.* (2004) 53:1658–64. doi: 10.1136/gut.2003.032805
- Brant SR, Wang MH, Rawsthorne P, Sargent M, Datta LW, Nouvet F, et al. A population-based case-control study of CARD15 and other risk factors in crohn's disease and ulcerative colitis. *Am J Gastroenterol.* (2007) 102:313–23. doi: 10.1111/j.1572-0241.2006.00926.x
- Caruso R, Mathes T, Martens EC, Kamada N, Nusrat A, Inohara N, et al. A specific gene-microbe interaction drives the development of Crohn's disease-like colitis in mice. *Sci Immunol.* (2019) 4:eaaw4341. doi: 10.1126/sciimmunol.aaw4341

- 72. Okazaki WT, Wang YMH, Rawsthorne NP, Sargent RM, Datta RL, Shugart RY, et al. Contributions of IBD5, IL23R, ATG16L1, and NOD2 to Crohn's disease risk in a population-based case-control study: evidence of gene–gene interactions. *Inflamm Bowel Dis.* (2008) 14:1528–41. doi: 10.1002/ibd.20512
- Altshuler DM, Gibbs RA, Peltonen L, Dermitzakis E, Schaffner SF, Yu F, et al. Integrating common and rare genetic variation in diverse human populations. *Nature*. (2010) 467:52–8. doi: 10.1038/nature09298
- Murthy A, Li Y, Peng I, Reichelt M, Katakam AK, Noubade R, et al. A Crohn's disease variant in Atg1611 enhances its degradation by caspase 3. *Nature*. (2014) 506:456–62. doi: 10.1038/nature13044
- Cadwell K, Liu JY, Brown SL, Miyoshi H, Loh J, Lennerz JK, et al. A key role for autophagy and the autophagy gene Atg16l1 in mouse and human intestinal paneth cells. *Nature*. (2008) 456:259–63. doi: 10.1038/nature07416
- Vestbo J, Prescott E, Lange P. Association of chronic mucus hypersecretion with FEV1 decline and chronic obstructive pulmonary disease morbidity. Copenhagen city heart study group. *Am J Respir Crit Care Med.* (1996) 153:1530–5. doi: 10.1164/ajrccm.153.5.8630597
- Kanner RE, Anthonisen NR, Connett JE. Lower respiratory illnesses promote FEV 1 decline in current smokers but not ex-smokers with mild chronic obstructive pulmonary disease: results from the lung health study. Am J Respir Crit Care Med. (2001) 164:358–64. doi: 10.1164/ajrccm.164.3.2010017
- Garcia-Nuñez M, Millares L, Pomares X, Ferrari R, Pérez-Brocal V, Gallego M, et al. Severity-related changes of bronchial microbiome in chronic obstructive pulmonary disease. J Clin Microbiol. (2014) 52:4217–23. doi: 10.1128/JCM.01967-14
- Erb-Downward J, Thompson D, Han M, Freeman C, McCloskey L, Schmidt L, et al. Analysis of the lung microbiome in the "healthy" smoker and in COPD. *PLoS ONE*. (2011) 6:e16384. doi: 10.1371/journal.pone.0016384
- Huang YJ, Sethi S, Murphy T, Nariya S, Boushey HA, Lynch SV. Airway microbiome dynamics in exacerbations of chronic obstructive pulmonary disease. J Clin Microbiol. (2014) 52:2813–23. doi: 10.1128/JCM.00035-14
- Millares L, Ferrari R, Gallego M, Garcia-Nunez M, Perez-Brocal V, Espasa M, et al. Bronchial microbiome of severe COPD patients colonised by *Pseudomonas aeruginosa. Eur J Clin Microbiol Infect Dis.* (2014) 33:1101–11. doi: 10.1007/s10096-013-2044-0
- Wang Z, Bafadhel M, Haldar K, Spivak A, Mayhew D, Miller B, et al. Lung microbiome dynamics in COPD exacerbations. *Eur Resp J.* (2016) 47:1082–92. doi: 10.1183/13993003.01406-2015
- Sun Z, Zhu QL, Shen Y, Yan T, Zhou X. Dynamic changes of gut and lung microorganisms during chronic obstructive pulmonary disease exacerbations. *Kaohsiung J Med Sci.* (2020) 36:107–13. doi: 10.1002/kjm2.12147
- Bernasconi E, Pattaroni C, Koutsokera A, Pison C, Kessler R, Benden C, et al. Airway microbiota determines innate cell inflammatory or tissue remodeling profiles in lung transplantation. *Am J Respir Crit Care Med.* (2016) 194:1252–63. doi: 10.1164/rccm.201512-2424OC
- Chen LW, Chen PH, Hsu CM. Commensal microflora contribute to host defense against *Escherichia coli* pneumonia through toll-like receptors. *Shock*. (2011) 36:67–75. doi: 10.1097/SHK.0b013e3182184ee7
- Schuijt TJ, Lankelma JM, Scicluna BP, de Sousa e Melo F, Roelofs JJTH, de Boer JD, et al. The gut microbiota plays a protective role in the host defence against pneumococcal pneumonia. *Gut.* (2016) 65:575–83. doi: 10.1136/gutjnl-2015-309728
- Sze MA, Tsuruta M, Yang SWJ, Oh Y, Man SFP, Hogg JC, et al. Changes in the bacterial microbiota in gut, blood, and lungs following acute LPS instillation into mice lungs (bacterial microbiota in gut, blood, and lungs after LPS. *PLoS ONE*. (2014) 9:e111228. doi: 10.1371/journal.pone.0111228
- Samuelson DR, Charles TP, de La Rua NM, Taylor CM, Blanchard EE, Luo M, et al. Analysis of the intestinal microbial community and inferred functional capacities during the host response to Pneumocystis pneumonia. *Exp Lung Res.* (2016) 42:425–39. doi: 10.1080/01902148.2016.1258442
- Gauguet S, D'Ortona S, Ahnger-Pier K, Duan B, Surana NK, Lu R, et al. Intestinal microbiota of mice influences resistance to *Staphylococcus aureus* pneumonia. *Infect Immun.* (2015) 83:4003–14. doi: 10.1128/IAI.00037-15
- Robert PD, Benjamin HS, Michael WN, Nicole RF, John RE-D, Theodore JS, et al. Enrichment of the lung microbiome with gut bacteria in sepsis and the acute respiratory distress syndrome. *Nat Microbiol.* (2016) 1:16113. doi: 10.1038/nmicrobiol.2016.113

- Wang F, Liu J, Zhang Y, Lei P. Association of helicobacter pylori infection with chronic obstructive pulmonary disease and chronic bronchitis: a meta-analysis of 16 studies. *Infect Dis.* (2015) 47:597–603. doi: 10.3109/00365548.2014.989539
- Benjamin LJ, Hedin RHC, Koutsoumpas CA, Ng ES, McCarthy JN, Prescott GN, et al. Smokers with active Crohn's disease have a clinically relevant dysbiosis of the gastrointestinal microbiota. *Inflamm Bowel Dis.* (2012) 18:1092–100. doi: 10.1002/ibd.21864
- 93. Anthonisen NR, Skeans MA, Wise RA, Manfreda J, Kanner RE, Connett JE. The effects of a smoking cessation intervention on 14.5-year mortality: a randomized clinical trial. (Author Abstract). Ann Intern Med. (2005) 142:233–9. doi: 10.7326/0003-4819-142-4-200502150-00005
- 94. Office of the Surgeon General, Office on Smoking and Health (US). (2004). Reports of the surgeon general. In: *The Health Consequences of Smoking: A Report of the Surgeon General*. Atlanta, GA: Centers for Disease Control and Prevention (US).
- Mahid SS, Minor KS, Soto RE, Hornung CA, Galandiuk S. Smoking and inflammatory bowel disease: a meta-analysis. *Mayo Clin Proc.* (2006) 81:1462–71. doi: 10.4065/81.11.1462
- Sutherland L, Ramcharan S, Bryant H, Fick G. Effect Of cigarettesmoking on recurrence of crohns-disease. *Gastroenterology*. (1990) 98:1123– 8. doi: 10.1016/0016-5085(90)90324-T
- Cosnes J, Carbonnel F, Carrat F, Beaugerie L, Cattan S, Gendre J. Effects of current and former cigarette smoking on the clinical course of Crohn's disease. *Aliment Pharmacol Ther.* (1999) 13:1403–11. doi: 10.1046/j.1365-2036.1999.00630.x
- Picco MF, Bayless TM. Tobacco consumption and disease duration are associated with fistulizing and stricturing behaviors in the first 8 years of Crohn's disease. Am J Gastroenterol. (2003) 98:363–8. doi: 10.1111/j.1572-0241.2003.07240.x
- Lindberg E, Järnerot G, Huitfeldt B. Smoking in Crohn's disease: effect on localisation and clinical course. *Gut.* (1992) 33:779–82. doi: 10.1136/gut.33.6.779
- Cosnes J, Beaugerie L, Carbonnel F, Gendre J-P. Smoking cessation and the course of Crohn's disease: an intervention study. *Gastroenterology*. (2001) 120:1093–9. doi: 10.1053/gast.2001.23231
- 101. Lindell G, Farnebo LO, Chen D, Nexø E, Madsen JR, Bukhave K, et al. Acute effects of smoking during modified sham feeding in duodenal ulcer patients: an analysis of nicotine, acid secretion, gastrin, catecholamines, epidermal growth factor, prostaglandin E2, and bile acids. *Scand J Gastroenterol*. (1993) 28:487–94. doi: 10.3109/00365529309098254
- 102. Biedermann L, Zeitz J, Mwinyi J, Sutter-Minder E, Rehman A, Ott S, et al. Smoking cessation induces profound changes in the composition of the intestinal microbiota in humans. *PLoS ONE.* (2013) 8:e59260. doi: 10.1371/journal.pone.0059260
- 103. Biedermann RL, Brülisauer JK, Zeitz JJ, Frei JP, Scharl JM, Vavricka JS, et al. Smoking cessation alters intestinal microbiota: insights from quantitative investigations on human fecal samples using FISH. *Inflamm Bowel Dis.* (2014) 20:1496–501. doi: 10.1097/MIB.000000000000129
- 104. Wang H, Zhao JX, Hu N, Ren J, Du M, Zhu MJ. Side-stream smoking reduces intestinal inflammation and increases expression of tight junction proteins. World J Gastroenterol. (2012) 18:2180–7. doi: 10.3748/wjg.v18. i18.2180
- 105. Allais L, Kerckhof FM, Verschuere S, Bracke KR, De Smet R, Laukens D, et al. Chronic cigarette smoke exposure induces microbial and inflammatory shifts and mucin changes in the murine gut. *Environ Microbiol.* (2016) 18:1352–63. doi: 10.1111/1462-2920.12934
- Nakanishi Y, Sato T, Ohteki T. Commensal Gram-positive bacteria initiates colitis by inducing monocyte/macrophage mobilization. *Mucosal Immunol.* (2014) 8:152–60. doi: 10.1038/mi.2014.53
- 107. Zuo L, Li Y, Wang H, Wu R, Zhu W, Zhang W, et al. Cigarette smoking is associated with intestinal barrier dysfunction in the small intestine but not in the large intestine of mice. J Crohn's Colitis. (2014) 8:1710–22. doi: 10.1016/j.crohns.2014.08.008
- Berkowitz L, Pardo-Roa C, Salazar GA, Salazar-Echegarai F, Miranda JP, Rama-rez G, et al. (2019). Mucosal exposure to cigarette components induces intestinal inflammation and alters antimicrobial response in mice. *Front Immunol.* 10:2289. doi: 10.3389/fimmu.2019.02289

- 109. Prager M, Büttner J, Grunert P, Ellinghaus D, Büning C. A promoter variant within the aryl hydrocarbon receptor gene is associated with an epithelial barrier defect in smokers with crohn's disease. *Inflamm Bowel Dis.* (2016) 22:2356–68. doi: 10.1097/MIB.00000000000910
- 110. Kim M, Gu B, Madison MC, Song HW, Norwood K, Hill AA, et al. Cigarette smoke induces intestinal inflammation via a Th17 cellneutrophil axis. (report)(brief article). *Front Immunol.* (2019) 10:75. doi: 10.3389/fimmu.2019.00075
- 111. Breitbart M, Hewson I, Felts B, Mahaffy JM, Nulton J, Salamon P, et al. Metagenomic analyses of an uncultured viral community from human feces. *J Bacteriol.* (2003) 185:6220. doi: 10.1128/JB.185.20.6220-6223.2003
- 112. Duerkop BA, Clements CV, Rollins D, Rodrigues JLM, and Hooper LV. A composite bacteriophage alters colonization by an intestinal commensal bacterium. *Proc Natl Acad Sci.* (2012) 109:17621. doi: 10.1073/pnas.1206136109
- Reyes A, Haynes M, Hanson N, Angly FE, Heath AC, Rohwer F, et al. Viruses in the faecal microbiota of monozygotic twins and their mothers. *Nature*. (2010) 466:334–8. doi: 10.1038/nature09199
- 114. Minot S, Sinha R, Jun C, Hongzhe L, Keilbaugh SA, Wu GD, et al. The human gut virome: inter-individual variation and dynamic response to diet.(Report). *Genome Res.* (2011) 21:1616–25. doi: 10.1101/gr.122705.111
- 115. Norman, JM, Handley SA, Baldridge MT, Droit L, Liu CY, Keller BC, et al. Disease-specific alterations in the enteric virome in inflammatory bowel disease. *Cell.* (2015) 160:447–60. doi: 10.1016/j.cell.2015.01.002
- 116. Wagner J, Maksimovic J, Farries G, Sim WH, Bishop RF, Cameron DJ, et al. Bacteriophages in gut samples from pediatric crohn's disease patients: metagenomic analysis using 454 pyrosequencing. *Inflamm Bowel Dis.* (2013) 19:1598–608. doi: 10.1097/MIB.0b013e318292477c
- 117. Main J, McKenzie H, Yeaman GR, Kerr MA, Robson D, Pennington CR, et al. Antibody to saccharomyces cerevisiae (bakers' yeast) in Crohn's disease. *BMJ*. (1988) 297:1105–6. doi: 10.1136/bmj.297.6656.1105
- 118. Sokol H, Leducq V, Aschard H, Pham H-P, Jegou S, Landman C, et al. Fungal microbiota dysbiosis in IBD. *Gut.* (2017) 66:1039. doi: 10.1136/gutjnl-2015-310746
- 119. Willner D, Furlan M, Haynes M, Schmieder R, Angly FE, Silva J, et al. Metagenomic analysis of respiratory tract DNA viral communities in cystic fibrosis and non-cystic fibrosis individuals. *PLOS ONE*. (2009) 4:e7370. doi: 10.1371/journal.pone.0007370
- 120. Young JC, Chehoud C, Bittinger K, Bailey A, Diamond JM, Cantu E, et al. Viral metagenomics reveal blooms of anelloviruses in the respiratory tract of lung transplant recipients. *Am J Transplant.* (2015) 15:200–209. doi: 10.1111/ajt.13031
- 121. McManus TE, Marley AM, Baxter N, Christie SN, Elborn JS, Neill HJ, et al. High levels of Epstein-Barr virus in COPD. *Eur Respir J.* (2008) 31:1221. doi: 10.1183/09031936.00107507
- 122. Utokaparch S, Sze MA, Gosselink JV, McDonough JE, Elliott WM, Hogg JC, et al. Respiratory Viral Detection and Small Airway Inflammation in Lung Tissue of Patients with Stable, Mild COPD. COPD. (2014) 11:197–203. doi: 10.3109/15412555.2013.836166
- 123. Tan DBA, Amran FS, Teo T-H, Price P, and Moodley YP. Levels of CMVreactive antibodies correlate with the induction of CD28null T cells and systemic inflammation in chronic obstructive pulmonary disease (COPD). *Cell Mol Immunol.* (2016) 13:551–3. doi: 10.1038/cmi.2015.4
- 124. Charlson ES, Diamond JM, Bittinger K, Fitzgerald AS, Yadav A, Haas AR, et al. Lung-enriched organisms and aberrant bacterial and fungal respiratory microbiota after lung transplant. *Am J Respir Crit Care Med.* (2012) 186:536– 45. doi: 10.1164/rccm.201204-0693OC
- 125. Martinsen EMH, Eagan TML, Leiten EO, Haaland I, Husebø GR, Sanseverino W, et al. A study of the airway mycobiome in COPD patients and controls. *Eur Respir J.* (2019) 54:OA1601. doi: 10.1183/13993003.congress-2019.OA1601
- 126. Su J, Liu H-y, Tan X-l, Ji Y, Jiang Y-X, Prabhakar M, et al. Sputum bacterial and fungal dynamics during exacerbations of severe COPD. *PLOS ONE*. (2015) 10:e0130736. doi: 10.1371/journal.pone.0130736.e0130736
- 127. Ananthakrishnan A. Epidemiology and risk factors for IBD. Nat Rev Gastroenterol Hepatol. (2015) 12:205–17. doi: 10.1038/nrgastro.2015.34
- 128. Ananthakrishnan AN, Khalili H, Konijeti GG, Higuchi LM, de Silva P, Korzenik JR, et al. A prospective study of long-term intake of dietary fiber

and risk of crohn's disease and ulcerative colitis. *Gastroenterology*. (2013) 145:970-7. doi: 10.1053/j.gastro.2013.07.050

- 129. Roberts CL, Keita ÅV, Duncan SH, O'Kennedy N, Söderholm JD, Rhodes JM, et al. Translocation of Crohn's disease *Escherichia coli* across M-cells: contrasting effects of soluble plant fibres and emulsifiers. *Gut.* (2010) 59:1331–9. doi: 10.1136/gut.2009.195370
- 130. Rodríguez-Cabezas ME, Gálvez J, Lorente MD, Concha A, Camuesco D, Azzouz S, et al. Dietary fiber down-regulates colonic tumor necrosis factor α and nitric oxide production in trinitrobenzenesulfonic acid-induced colitic rats. J Nutr. (2002) 132:3263–71. doi: 10.1093/jn/132.11.3263
- 131. Rodriguez-Cabezas ME, Gálvez J, Camuesco D, Lorente MD, Concha A, Martinez-Augustin O, et al. Intestinal anti-inflammatory activity of dietary fiber (*Plantago ovata* seeds) in HLA-B27 transgenic rats. *Clin Nutr.* (2003) 22:463–71. doi: 10.1016/S0261-5614(03)00045-1
- 132. Kovarik JJ, Tillinger W, Hofer J, Hölzl MA, Heinzl H, Saemann MD, et al. Impaired anti-inflammatory efficacy of n-butyrate in patients with IBD. *Eur J Clin Invest.* (2011) 41:291–8. doi: 10.1111/j.1365-2362.2010.02407.x
- 133. Breuer RI, Soergel KH, Lashner BA, Christ ML, Hanauer SB, Vanagunas A, et al. Short chain fatty acid rectal irrigation for left-sided ulcerative colitis: a randomised, placebo controlled trial. *Gut.* (1997) 40:485–91. doi: 10.1136/gut.40.4.485
- 134. Thibault R, De Coppet P, Daly K, Bourreille A, Cuff M, Bonnet C, et al. Down-regulation of the monocarboxylate transporter 1 is involved in butyrate deficiency during intestinal inflammation. *Gastroenterology*. (2007) 133:1916–27. doi: 10.1053/j.gastro.2007.08.041
- Hove H, Mortensen P. Influence of intestinal inflammation (IBD) and small and large bowel length on fecal short-chain fatty acids and lactate. *Dig Dis Sci.* (1995) 40:1372–80. doi: 10.1007/BF02065554
- 136. De Preter V, Machiels K, Joossens M, Arijs I, Matthys C, Vermeire S, et al. Faecal metabolite profiling identifies medium-chain fatty acids as discriminating compounds in IBD. *Gut.* (2015) 64:447–58. doi: 10.1136/gutjnl-2013-306423
- 137. Joossens M, De Preter V, Ballet V, Verbeke K, Rutgeerts P, Vermeire S. Effect of oligofructose-enriched inulin (OF-IN) on bacterial composition and disease activity of patients with Crohn's disease: results from a double-blinded randomised controlled trial. *Gut.* (2012) 61:958. doi: 10.1136/gutjnl-2011-300413
- Unger MM, Spiegel J, Dillmann K-U, Grundmann D, Philippeit H, Bürmann J, et al. Short chain fatty acids and gut microbiota differ between patients with Parkinson's disease and age-matched controls. *Parkinsonism Relat Disord*. (2016) 32:66–72. doi: 10.1016/j.parkreldis.2016.08.019
- Mariño E, Richards J, McLeod K, Stanley D, Yap Y, Knight J, et al. Gut microbial metabolites limit the frequency of autoimmune T cells and protect against type 1 diabetes. *Nat Immunol.* (2017) 18:552–62. doi: 10.1038/ni.3713
- 140. McKenzie C, Tan J, Macia L, Mackay CR. The nutrition-gut microbiomephysiology axis and allergic diseases. *Immunol Rev.* (2017) 278:277–95. doi: 10.1111/imr.12556
- 141. Sanna S, van Zuydam N, Mahajan A, Kurilshikov A, Vila A, Vōsa U, et al. Causal relationships among the gut microbiome, short-chain fatty acids and metabolic diseases. *Nat Genet.* (2019) 51:600–3. doi: 10.1038/s41588-019-0350-x
- Varraso R, Willett WC, Camargo CA. Prospective study of dietary fiber and risk of chronic obstructive pulmonary disease among US women and men. *Am J Epidemiol.* (2010). 171:776–84. doi: 10.1093/aje/kwp455
- 143. Fonseca Wald ELA, van Den Borst B, Gosker HR, Schols AMWJ. Dietary fibre and fatty acids in chronic obstructive pulmonary disease risk and progression: a systematic review. *Respirol.* (2014) 19:176–184. doi: 10.1111/resp.12229
- 144. Hirayama F, Lee AH, Binns CW, Zhao Y, Hiramatsu T, Tanikawa Y, et al. Do vegetables and fruits reduce the risk of chronic obstructive pulmonary disease? A case–control study in Japan. *Prev Med.* (2009) 49:184– 9. doi: 10.1016/j.ypmed.2009.06.010
- 145. Tabak C, Smit HA, Heederik D, Ocké MC, Kromhout D. Diet and chronic obstructive pulmonary disease: independent beneficial effects of fruits, whole grains, and alcohol (the MORGEN study. *Clin Exp Allergy*. (2001) 31:747–55. doi: 10.1046/j.1365-2222.2001.01064.x
- 146. Watson L, Margetts B, Howarth P, Dorward M, Thompson R, Little P. The association between diet and chronic obstructive pulmonary disease

in subjects selected from general practice. Eur Resp J. (2002) 20:313-8. doi: 10.1183/09031936.02.00256402

- Butler L, Koh WP, Lee H, Yu MC, London SJ. Dietary fiber and reduced cough with phlegm - a cohort study in Singapore. *Am J Respir Crit Care Med.* (2004) 170:279–87. doi: 10.1164/rccm.200306-789OC
- Celik F, Topcu F. Nutritional risk factors for the development of chronic obstructive pulmonary disease (COPD) in male smokers. *Clin Nutr.* (2006) 25:955–61. doi: 10.1016/j.clnu.2006.04.006
- 149. Trompette A, Gollwitzer ES, Yadava K, Sichelstiel AK, Sprenger N, Ngom-Bru C, et al. Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. *Nat Med.* (2014) 20:159–66. doi: 10.1038/nm.3444
- 150. Trompette A, Gollwitzer ES, Pattaroni C, Lopez-Mejia IC, Riva E, Pernot J, et al. Dietary fiber confers protection against flu by shaping Ly6c- patrolling monocyte hematopoiesis and CD8+ T cell metabolism. *Immunity*. (2018) 48:992–1005.e1008. doi: 10.1016/j.immuni.2018.04.022
- 151. Nath K, Burel J, Shankar V, Pritchard A, Towers M, Looke D, et al. Clinical factors associated with the humoral immune response to influenza vaccination in chronic obstructive pulmonary disease. *Int J Chron Obstruct Pulmon Dis.* (2013) 9:51–6. doi: 10.2147/COPD. S53590
- 152. Parpaleix A, Boyer L, Wiedemann A, Lacabaratz C, Margarit L, Enouf V, et al. Impaired humoral and cellular immune responses to influenza vaccination in chronic obstructive pulmonary disease patients. *J Allergy Clin Immunol.* (2017) 140:1754–7.e1756. doi: 10.1016/j.jaci.2017. 07.038
- Chapman-Kiddell CA, Davies PSW, Gillen L, Radford-Smith GL. Role of diet in the development of inflammatory bowel disease. *Inflamm Bowel Dis.* (2010) 16:137–51. doi: 10.1002/ibd.20968
- 154. Manzel A, Muller D, Hafler D, Erdman S, Linker R, Kleinewietfeld M. Role of "western diet" in inflammatory autoimmune diseases. *Curr Allergy Asthma Rep.* (2014) 14:404. doi: 10.1007/s11882-013-0404-6
- 155. Nappo F, Esposito K, Cioffi M, Giugliano G, Molinari AM, Paolisso G, et al. Postprandial endothelial activation in healthy subjects and in type 2 diabetic patients: role of fat and carbohydrate meals. J Am Coll Cardiol. (2002) 39:1145–50. doi: 10.1016/S0735-1097(02)01741-2
- 156. Wu G, Chen J, Hoffmann C, Bittinger K, Ying-Yu C, Keilbaugh S, et al. Linking long-term dietary patterns with gut microbial enterotypes. *Science*. (2011) 334:105–8. doi: 10.1126/science.1208344
- 157. Pendyala S, Walker JM, Holt PR. A high-fat diet is associated with endotoxemia that originates from the gut. *Gastroenterology*. (2012) 142:1100–01.e1102. doi: 10.1053/j.gastro.2012.01.034
- van Oostrom A, Sijmonsma TP, Verseyden C, Jansen E, de Koning E, Rabelink T, et al. Postprandial recruitment of neutrophils may contribute to endothelial dysfunction. J Lipid Res. (2003) 44:576–83. doi: 10.1194/jlr.M200419-JLR200
- Turnbaugh PJ, Bäckhed F, Fulton L, Gordon JI. Diet-induced obesity is linked to marked but reversible alterations in the mouse distal gut microbiome. *Cell Host Microbe.* (2008) 3:213–23. doi: 10.1016/j.chom.2008.02.015
- 160. Martinez-Medina M, Denizot J, Dreux N, Robin F, Billard E, Bonnet R, et al. Western diet induces dysbiosis with increased *E. coli* in CEABAC10 mice, alters host barrier function favouring AIEC colonisation. *Gut.* (2014) 63:116–24. doi: 10.1136/gutjnl-2012-304119
- 161. Hassan V, Hassan S, Seyed-Javad P, Ahmad K, Asieh H, Maryam S, et al. Association between serum 25 (OH) vitamin D concentrations and inflammatory bowel diseases (IBDs) activity. *Med J Malaysia*. 68:34–8. doi: 10.1371/journal.pone.0132036
- 162. Wu S, Zhang YG, Lu R, Xia Y, Zhou D, Petrof EO, et al. Intestinal epithelial vitamin D receptor deletion leads to defective autophagy in colitis. *Gut.* (2015) 64:1082–94. doi: 10.1136/gutjnl-2014-307436
- 163. Ooi JH, Li Y, Rogers CJ, Cantorna MT. Vitamin D regulates the gut microbiome and protects mice from dextran sodium sulfateinduced colitis. J Nutr. (2013) 143:1679–86. doi: 10.3945/jn.113. 180794
- 164. Jin D, Wu S, Zhang YG, Lu R, Xia Y, Dong H, et al. Lack of vitamin D receptor causes dysbiosis and changes the functions of the murine intestinal microbiome. *Clin Ther.* (2015) 37:996–1009.e1007. doi: 10.1016/j.clinthera.2015.04.004

- 165. Schäffler H, Herlemann DP, Klinitzke P, Berlin P, Kreikemeyer B, Jaster R, et al. Vitamin D administration leads to a shift of the intestinal bacterial composition in Crohn's disease patients, but not in healthy controls. *J Dig Dis*. (2018) 19:225–34. doi: 10.1111/1751-2980.12591
- 166. Black PN, Scragg R. Relationship between serum 25-hydroxyvitamin D and pulmonary function in the third national health and nutrition examination survey. *Chest.* (2005) 128:3792–8. doi: 10.1378/chest.128.6.3792
- 167. Janssens W, Bouillon R, Claes B, Carremans C, Lehouck A, Buysschaert I, et al. Vitamin D deficiency is highly prevalent in COPD and correlates with variants in the vitamin D-binding gene. *Thorax.* (2010) 65:215–20. doi: 10.1136/thx.2009.120659
- Liu P, Stenger S, Li H, Wenzel L, Tan B, Krutzik S, et al. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science*. (2006) 311:1770–3. doi: 10.1126/science.1123933
- 169. Toivonen L, Hasegawa K, Ajami N, Celedón J, Mansbach J, Petrosino J, et al. Circulating 25-hydroxyvitamin D, nasopharyngeal microbiota, and bronchiolitis severity. *Pediatr Allergy Immunol.* (2018) 29:877–80. doi: 10.1111/pai.12977
- 170. Sundar IK, Hwang JW, Wu S, Sun J, Rahman I. Deletion of vitamin D receptor leads to premature emphysema/COPD by increased matrix metalloproteinases and lymphoid aggregates formation. *Biochem Biophys Res Commun.* (2011) 406:127–33. doi: 10.1016/j.bbrc.2011.02.011
- Balcells ME, Yokobori N, Hong B-Y, Corbett J, Cervantes J. The lung microbiome, vitamin D, and the tuberculous granuloma: a balance triangle. *Microb Pathog.* (2019) 131:158–63. doi: 10.1016/j.micpath.2019.03.041
- 172. Kanhere RM, He AJ, Chassaing AB, Ziegler TT, Alvarez TJ, Ivie TE, et al. Bolus weekly vitamin D3 supplementation impacts gut and airway microbiota in adults with cystic fibrosis: a double-blind, randomized, placebo-controlled clinical trial. *J Clin Endocrinol Metab.* (2018) 103:564–74. doi: 10.1210/jc.2017-01983
- 173. Ungaro NR, Bernstein PC, Gearry PR, Hviid PA, Kolho PKL, Kronman PM, et al. Antibiotics associated with increased risk of new-onset crohn's disease but not ulcerative colitis: a meta-analysis. *Am J Gastroenterol.* (2014) 109:1728–38. doi: 10.1038/ajg.2014.246
- 174. Shaw YS, Blanchard FJ, Bernstein NC. Association between the use of antibiotics in the first year of life and pediatric inflammatory bowel disease. *Am J Gastroenterol.* (2010) 105:2687–92. doi: 10.1038/ajg.2010.398
- 175. Hviid A, Svanstrom H, Frisch M. Antibiotic use and inflammatory bowel diseases in childhood. *Gut.* (2011) 60:49–54. doi: 10.1136/gut.2010.219683
- 176. Hansen KR, Russell HR, Reiff WC, Louis RP, McIntosh MF, Berry JS, et al. Microbiota of *De-Novo* pediatric IBD: increased *faecalibacterium prausnitzii* and reduced bacterial diversity in crohn's but not in ulcerative colitis. *Am J Gastroenterol.* (2012) 107:1913–22. doi: 10.1038/ajg. 2012.335
- Jernberg C, Löfmark S, Edlund C, Jansson J. Long-term ecological impacts of antibiotic administration on the human intestinal microbiota. *ISME J.* (2013) 7:456. doi: 10.1038/ismej.2012.91
- 178. Palmer C, Bik EM, DiGiulio DB, Relman DA, Brown PO. Development of the human infant intestinal microbiota (microbiota of the infant intestine). *PLoS Biol.* (2007) 5:e177. doi: 10.1371/journal.pbio.0050177
- 179. Greenberg GR. Antibiotics should be used as first-line therapy for Crohn's disease. *Inflamm Bowel Dis.* (2004) 10:318–20. doi: 10.1097/00054725-200405000-00021
- 180. Steinhart AH, Feagan BG, Wong CJ, Vandervoort M, Mikolainis S, Croitoru K, et al. Combined budesonide and antibiotic therapy for active Crohn's disease: A randomized controlled trial. *Gastroenterology*. (2002) 123:33–40. doi: 10.1053/gast.2002.34225
- 181. Rutgeerts P, Hiele M, Geboes K, Peeters M, Penninckx F, Aerts R, et al. Controlled trial of metronidazole treatment for prevention of crohn's recurrence after ileal resection. *Gastroenterology*. (1995) 108:1617–21. doi: 10.1016/0016-5085(95)90121-3
- 182. Townsend CM, Parker CE, Macdonald J, Nguyen TM, Jairath V, Feagan B, et al. Antibiotics for induction and maintenance of remission in Crohn's disease. *Cochrane Database Syst Rev.* (2019) 2:CD012730. doi: 10.1002/14651858.CD012730.pub2
- Arnold GL, Beaves MR, Pryjdun VO, Mook WJ. Preliminary study of ciprofloxacin in active Crohn's disease. *Inflamm Bowel Dis.* (2002) 8:10–5. doi: 10.1097/00054725-200201000-00002

- 184. Bamias G, Marini M, Moskaluk CA, Odashima M, Ross WG, Rivera-Nieves J, et al. Down-regulation of intestinal lymphocyte activation and Th1 cytokine production by antibiotic therapy in a murine model of Crohn's disease. J Immunol. (2002) 169:5308–14. doi: 10.4049/jimmunol.169.9.5308
- 185. Rahimi R, Nikfar S, Rezaie A, Abdollahi M. A meta-analysis of antibiotic therapy for active ulcerative colitis. *Dig Dis Sci.* (2007) 52:2920–5. doi: 10.1007/s10620-007-9760-1
- 186. Puhan MA, Vollenweider D, Steurer J, Bossuyt P, Ter Riet G. Where is the supporting evidence for treating mild to moderate chronic obstructive pulmonary disease exacerbations with antibiotics? A systematic review. *BMC Med.* (2008) 6:28. doi: 10.1186/1741-7015-6-28
- 187. Laue J, Reierth E, Melbye H. When should acute exacerbations of COPD be treated with systemic corticosteroids and antibiotics in primary care:a systematic review of current COPD guidelines. NPJ Prim Care Respir Med. (2015) 25:15002. doi: 10.1038/npjpcrm.2015.2
- 188. Marjanović N, Bosnar M, Michielin F, Willé DR, Anić-Milić T, Culić O, et al. Macrolide antibiotics broadly and distinctively inhibit cytokine and chemokine production by COPD sputum cells *in vitro*. *Pharmacol Res.* (2011) 63:389–97. doi: 10.1016/j.phrs.2011.02.001
- 189. James GDR, Petersen I, Nazareth I, Wedzicha JA, Donaldson GC. Use of long-term antibiotic treatment in COPD patients in the UK: a retrospective cohort study. *Prim Care Res J.* (2013) 22:271–7. doi: 10.4104/pcrj.2013.00061
- Herath SC, Normansell R, Maisey S, Poole P. Prophylactic antibiotic therapy for chronic obstructive pulmonary disease (COPD). *Cochrane Database Syst Rev.* (2018) 10:CD009764. doi: 10.1002/14651858.CD009764.pub3
- 191. Guo B, Harstall C, Louie T, Veldhuyzen van Zanten S, Dieleman LA. Systematic review: faecal transplantation for the treatment of clostridium difficile-associated disease. *Aliment Pharmacol Ther.* (2012) 35:865–75. doi: 10.1111/j.1365-2036.2012.05033.x
- 192. Shahinas D, Silverman M, Sittler T, Chiu C, Kim P, Allen-Vercoe E, et al. Toward an understanding of changes in diversity associated with fecal microbiome transplantation based on 16S rRNA gene deep sequencing. *MBio.* (2012) 3:e00338-12. doi: 10.1128/mBio.00338-12
- 193. Kump KP, Gröchenig MH-P, Lackner HS, Trajanoski JS, Reicht JG, Hoffmann JK, et al. Alteration of intestinal dysbiosis by fecal microbiota transplantation does not induce remission in patients with chronic active ulcerative colitis. *Inflamm Bowel Dis.* (2013) 19:2155–65. doi: 10.1097/MIB.0b013e31829ea325
- Borody TJ, Warren EF, Leis S, Surace R, Ashman O. Treatment of ulcerative colitis using fecal bacteriotherapy. J Clin Gastroenterol. (2003) 37:42–7. doi: 10.1097/00004836-200307000-00012
- 195. Rossen NG, Fuentes S, van Der Spek MJ, Tijssen JG, Hartman JHA, Duflou A, et al. Findings from a randomized controlled trial of fecal transplantation for patients with ulcerative colitis. *Gastroenterology*. (2015) 149:110–8.e114. doi: 10.1053/j.gastro.2015.03.045
- 196. Paramsothy S, Kamm MA, Kaakoush NO, Walsh AJ, van Den Bogaerde J, Samuel D, et al. Multidonor intensive faecal microbiota transplantation for active ulcerative colitis: a randomised placebo-controlled trial. *Lancet*. (2017) 389:1218–28. doi: 10.1016/S0140-6736(17)30182-4
- 197. Anderson JL, Edney RJ, Whelan K. Systematic review: faecal microbiota transplantation in the management of inflammatory bowel disease. *Aliment Pharmacol Ther.* (2012) 36:503–16. doi: 10.1111/j.1365-2036.2012.05220.x
- 198. Paramsothy S, Paramsothy R, Rubin DT, Kamm MA, Kaakoush NO, Mitchell HM, et al. Faecal microbiota transplantation for inflammatory bowel disease: a systematic review and meta-analysis. J Crohn's Colitis. (2017) 11:1180–99. doi: 10.1093/ecco-jcc/jjx063

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Raftery, Tsantikos, Harris and Hibbs. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.