



# Logging and soil nutrients independently explain plant trait expression in tropical forests

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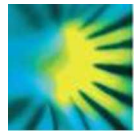
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1 **Logging and soil nutrients independently explain plant trait expression in tropical forests**

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For Peer Review

40 **Summary**

- 41 • Plant functional traits regulate ecosystem functions but little is known about how co-occurring  
42 gradients of land use and edaphic conditions influence their expression. We test how gradients  
43 of logging disturbance and soil properties relate to community-weighted mean traits in logged  
44 and old-growth tropical forests in Borneo.
- 45 • In eight 1 ha plots, we studied 32 physical, chemical and physiological traits from 284 tree  
46 species and measured long-term soil nutrient supplies and plant-available nutrients.
- 47 • Logged plots had greater values for traits that drive carbon capture and growth, whilst old-  
48 growth forests had greater values for structural and persistence traits. Although disturbance was  
49 the primary driver of trait expression, soil nutrients explained a statistically independent axis of  
50 variation linked to leaf size and nutrient concentration. Soil characteristics influenced trait  
51 expression via nutrient availability, and through nutrient pools and pH.
- 52 • Our finding, that traits have contrasting responses to land use and soil resource availability,  
53 provides robust evidence for the need to consider the abiotic context of logging when predicting  
54 plant functional diversity across human-modified tropical forests. The detection of two  
55 independent axes was made possible by the measurement of many more functional traits than  
56 have been examined in previous studies.

57  
58 **Keywords:** anthropogenic disturbance, Borneo, functional diversity, functional traits, land use, Rao's  
59 Q, tropical rainforest, variance partitioning

60

## 61 Introduction

62 The differential expression of plant functional traits influences key ecosystem functions (Cornwell *et al.* 2008; De Deyn *et al.* 2008; Fortunel *et al.* 2009; Finegan *et al.* 2015). Trait expression varies across  
63 landscapes as a result of anthropogenic disturbance, soil characteristics, and other abiotic factors  
64 such as climate (Ordoñez *et al.* 2009; Baraloto *et al.* 2012; Fortunel *et al.* 2014a; Dent & Burslem  
65 2016). Fertile soils are associated with traits conferring rapid nutrient acquisition and use, which  
66 support fast growth rates, whereas nutrient-poor soils are often associated with conservative  
67 strategies for the maintenance of long-lived tissues (Aerts & Chapin 2000; Ordoñez *et al.* 2009; Jager  
68 *et al.* 2015). Anthropogenic disturbances pervade ecosystems worldwide (Hansen *et al.* 2013) and  
69 can affect trait expression in seeds, leaves and woody tissue (Gómez-González *et al.* 2011; Baraloto  
70 *et al.* 2012; Carreño-Rocabado *et al.* 2012). Substantial effort has been dedicated to characterising  
71 the independent effects of human disturbance (McIntyre *et al.* 1999; Mouillot *et al.* 2013a; Buzzard  
72 *et al.* 2016) and environmental gradients (Fortunel *et al.* 2014b; Fyllas *et al.* 2017) on plant trait  
73 expression and ecosystem function. However, in practice, communities are influenced by multiple  
74 factors simultaneously, and the effects of disturbance may vary along environmental gradients such  
75 as nutrient availability.  
76

77 Comprehensively analysing trait expression in response to multiple gradients is challenging but  
78 essential for predicting the ecosystem-level consequences of anthropogenic disturbance. The leaf  
79 economics spectrum (Wright *et al.* 2004; Díaz *et al.* 2016) suggests that the increase in resource  
80 availability associated with disturbance and soil fertility will select for similar leaf trait syndromes.  
81 Therefore, functional traits of plant communities should converge at the extremes of environmental  
82 gradients. Evidence from tropical tree communities suggests that foliar concentrations of N and P  
83 and specific leaf area increase in response to gradients of both disturbance (Baraloto *et al.* 2012;  
84 Carreño-Rocabado *et al.* 2012; Carreño-Rocabado *et al.* 2016) and soil nutrient availability (Fyllas *et al.*  
85 2009; Fortunel *et al.* 2014a; Apaza *et al.* 2015; Jager *et al.* 2015; Turnbull *et al.* 2016; Van der  
86 Sande *et al.* 2016). Similarly, leaf dry matter content and branch and stem wood density decrease  
87 with both disturbance (Verburg & van Eijk-Bos 2003; Baraloto *et al.* 2012; Carreño-Rocabado *et al.*  
88 2012; Carreño-Rocabado *et al.* 2016) and soil nutrients (Ordoñez *et al.* 2009; Fortunel *et al.* 2014b;  
89 Jager *et al.* 2015). All these studies, however, share two weaknesses: they did not examine the  
90 influence of multiple factors on trait expression, and they used a limited set of traits. Moreover,  
91 many did not consider traits associated with ecologically important processes, such as structural and  
92 defence compounds (important for herbivory and hence trophic interactions); photosynthetic  
93 activity (essential for biomass production); and leaf  $\delta^{15}\text{N}$  values (provides insight into sources and  
94 use of nitrogen). Our understanding about the links between trait sensitivity to anthropogenic

95 disturbance, soil properties, and ecosystem processes in tropical forests therefore remains  
96 incomplete.

97         The consequences of changes in community structure and diversity for ecosystem service  
98 provision are determined by the impacts of disturbance on community-level trait expression, which  
99 is a function of the type and intensity of disturbance. For example, in South American tropical  
100 forests, disturbance due to logging and silvicultural activity increases light availability and triggers  
101 the recruitment of species with traits that promote rapid growth rates (Baraloto *et al.* 2012;  
102 Carreño-Rocabado *et al.* 2012). We expect a similar response to logging in Southeast Asian tropical  
103 forest but additionally aim to determine the influence of soil properties including nutrient  
104 availability.

105         Tropical lowland forests in Southeast Asia are amongst the most species-rich communities  
106 worldwide, but are also the most threatened by intensive logging and conversion to agriculture  
107 (Hansen *et al.* 2013; Edwards *et al.* 2014; Stibig *et al.* 2014). The high density of commercially  
108 valuable species explains the high intensity of logging in Southeast Asian forests (Osman *et al.* 2012;  
109 Gaveau *et al.* 2014), which suffer rates of extraction that far exceed those in tropical forests  
110 elsewhere (Asner *et al.* 2005). Logging creates a spatially patchy disturbance, with gaps and skid  
111 trails characterised by high light and temperature distributed among fragments of relatively  
112 unmodified forest (Johns 1997). The selective removal of target species, logging-induced mortality  
113 and recruitment of pioneer species in disturbed areas affects tree species composition and the pools  
114 and fluxes of biomass and nutrients (Cannon *et al.* 1998; Verburg & van Eijk-Bos 2003; Pfeifer *et al.*  
115 2016; Riutta *et al.* 2018). Although the magnitude of anthropogenic disturbance is much greater in  
116 forests in Southeast Asia than in South America, they have received significantly less attention  
117 regarding the modification of plant functional traits. Specifically, intense logging has the potential to  
118 override the effects of other environmental gradients including soil properties.

119         Here, we measured 32 leaf, wood and physiological traits of 284 tropical tree species to  
120 capture community level trait expression in response to selective logging across a gradient of soil  
121 properties in species-rich tropical rainforest in Sabah, northern Borneo. We assess traits reflecting  
122 nutrient status, light capture and photosynthesis, and allocation to structure and defence to provide  
123 a whole-plant perspective and avoid overlooking traits that contribute to functional diversity. We  
124 tested the hypothesis that CWM values of functional traits and functional diversity shift in response  
125 to anthropogenic disturbance and soil properties. We used CWM traits to quantify average trait  
126 values, and a multi-trait index of functional diversity (FD) to quantify trait variation, which can occur  
127 independent of variation in CWM values of trait (Ricotta & Moretti 2011).

128 Specifically, we predict that (1) increasing disturbance and soil nutrient availability will  
129 increase CWM traits related to tissue nutrient concentrations and carbon assimilation rates, but  
130 reduce tissue densities and investments in structural defences. Given the high logging intensity in  
131 Southeast Asia, we further hypothesise that (2) a greater proportion of the variance in CWM traits  
132 will be explained by logging than by soil properties, emphasizing the pervasive impact of  
133 anthropogenic land use on functional trait expression. Finally, due to the high overall tree species  
134 richness we predict that (3) functional diversity will remain high in response to disturbance, despite  
135 shifts in overall CWM traits.

136

## 137 **Material and Methods**

### 138 *Study sites*

139 Sampling was conducted in eight 1 ha plots in Sabah, Malaysian Borneo. The plots are part of the  
140 Global Ecosystems Monitoring (GEM) network of permanent sample plots with intensive, regular  
141 carbon cycle measurements (Malhi *et al.* 2015; Riutta *et al.* 2018) and were selected to capture  
142 variation in logging intensity across a range of lowland tropical forests in northern Borneo. Old-  
143 growth lowland mixed dipterocarp forest plots (hereafter OG) were located in the Maliau Basin  
144 Conservation Area (MBCA, two plots) and the Danum Valley Conservation Area (DVCA, two plots),  
145 while the other four plots were distributed between these two areas in the selectively logged  
146 Kalabakan Forest Reserve (hereafter SL). The four logged plots are part of the Stability of Altered  
147 Forest Ecosystem (SAFE) project (Ewers *et al.* 2011). This area has been logged two to four times  
148 with the first round of logging in the mid-1970s and subsequent repeated logging during 1990-2008.  
149 Approximately 150–179 m<sup>3</sup> ha<sup>-1</sup> of timber was removed over this time period (Struebig *et al.* 2013),  
150 bracketing the mean extraction volume across Sabah (152 m<sup>3</sup> ha<sup>-1</sup>, Fisher *et al.* 2011).

151 All three areas are part of the Yayasan Sabah Forest Management Area and belong to a  
152 formerly connected area of lowland dipterocarp rainforest characterised by high species richness  
153 and many tall, emergent trees. The region has a moist tropical climate with an annual daily mean  
154 temperature of 26.7 °C and annual precipitation of approximately 2600-2700 mm (Walsh & Newbery  
155 1999). Although the climate is aseasonal there are occasional droughts and dry spells associated  
156 with supra-annual El Niño Southern Oscillation events (Walsh & Newbery 1999; Newbery &  
157 Lingenfelder 2009). The forest soils in Sabah are mostly orthic Acrisols or Ultisols (for more details  
158 see Marsh & Greer 1992; Nainar *et al.* 2015).

159

### 160 *Sampling design*



161 Thirty-two functional traits were measured on 651 individual trees  $\geq 10$  cm diameter at breast height  
162 (dbh) representing 284 species during an intensive sampling campaign from July to December 2015.  
163 We combined two strategies to sample the functional trait values in each plot. In the first, weighted  
164 basal area strategy, we sampled species that most contributed to the total plot basal area. This  
165 approach assumes that species accounting for a larger proportion of plot basal area also make a  
166 greater contribution to ecosystem functioning. Species were ranked based on their contribution to  
167 total basal area at the most recent census for each plot (2011 to 2015), which ranged from 10.9–  
168 41.8  $\text{m}^2 \text{ha}^{-1}$ . All species that contributed to 70% of plot basal area (in decreasing order of species  
169 basal area) were identified for sampling. In 57% of cases only one individual per species occurred;  
170 otherwise, the individual with the greatest dbh within a species was sampled. This strategy  
171 disproportionately sampled large-statured and abundant species. To ensure that smaller and  
172 potentially rare species were also represented, we adopted a second strategy: stratified random and  
173 taxon-independent sampling of all trees  $\geq 10$  cm dbh in three randomly selected  $20 \times 20$  m subplots  
174 within each 1 ha plot. As this strategy allowed for repeated samples of the same species, as well as  
175 sampling from all height strata, it contained understory and shaded trees. This combination of  
176 different sampling strategies provided a comprehensive representation of the tree community  
177 (Paine *et al.* 2015) and resulted in an overall representation of  $> 90\%$  of the total basal area per plot  
178 (except one plot with 65%) and 51–71% of the species  $\geq 10$  cm dbh (Fig. S1).

179

#### 180 *Soil properties*

181 We measured total nutrients and exchangeable nutrient pools to estimate both long-term nutrient  
182 supply as well as plant-available forms. Two randomly located soil cores were taken per plot in 2014–  
183 2015. Surface soil (0–10 cm) was analysed for pH, cation exchange capacity and total concentrations  
184 of P, C, N, Mg, K and Ca using the protocols of Quesada *et al.* (2010, 2012). To assess availability of K,  
185 Ca, Mg, P,  $\text{NO}_3^-$  and  $\text{NH}_4^+$  we measured nutrient supply rates using in-situ ion exchange membranes  
186 (PRSTM Probes, Western AG, Saskatoon, Canada). To account for spatial variability we installed four  
187 probe pairs (each composed of one cation and anion probe) vertically at corners of three  $50 \times 50$  cm  
188 quadrats to 10 cm depth within each of the three subplots used in the stratified random sampling.  
189 These were collected after two weeks, washed with distilled water and sent to the manufacturer for  
190 analysis. They pooled the four probe pairs from each quadrat prior to elution with 0.5M HCl for 1 hr,  
191 yielding 72 samples.  $\text{NO}_3^-$  and  $\text{NH}_4^+$  were measured colorimetrically using automated flow injection  
192 analysis (FIA). All other elements were analysed using Inductively Coupled Plasma Mass  
193 Spectrometry (ICP-MS). Results are reported as supply rates over the burial period (micrograms/10  
194  $\text{cm}^2/14$  days).

195

196 *Trait measurements*

197 From each target tree, we attempted to sample a fully sunlit canopy branch and a fully shaded  
198 branch; however, branches of only one type were available for most trees (91%) because it was  
199 uncommon for large canopy trees to possess fully shaded branches and for small understorey trees  
200 to have fully sunlit branches. Branch samples were collected by tree climbing or by cutting from the  
201 ground with telescopic branch cutters. Target tree height ranged from 2.3 to 78.1 m, and sample  
202 height ranged from 2.3 m to 53 m. Branches were approximately 2-4 cm in diameter and provided  
203 sufficient leaf material for all analyses. Photosynthetic activity was only measured on trees selected  
204 by the basal area sampling strategy due to time constraints (N = 298), whereas all other functional  
205 traits were determined on all trees. Undamaged mature leaves were collected and cleaned with  
206 water for subsequent analyses. Fresh and dry leaf weight, (specific) leaf area, leaf thickness, leaf dry  
207 matter content (LDMC), (specific) force to punch and branch wood density were determined in a  
208 field laboratory. Dried bulked and milled leaf material was used for determination of Ca, K, Mg, P, C  
209 and N concentrations,  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  stable isotope composition, cellulose, hemicellulose and lignin  
210 concentrations. Analyses of pigments (chlorophyll a, chlorophyll b, and carotenoids), phenols and  
211 tannins were conducted on 0.7 cm diameter leaf discs punched from fresh leaves immediately after  
212 field collection and frozen in liquid nitrogen. Herbarium voucher specimens were taken for  
213 identification of trees and were deposited in the herbarium at Danum Valley Field Centre. All trait  
214 measurements follow standardised protocols (Pérez-Harguindeguy *et al.* 2013), and detailed  
215 methods and an overview of sampling and replication are provided in Table S1.

216

217 *Statistical analyses*

218 Replicated leaf-level functional traits were averaged for sun and shade leaves for the few individuals  
219 that possessed both to generate a tree-level dataset. Our results were identical for analyses using a  
220 data-set comprising sun leaves alone, where available, or otherwise shade leaves, so henceforth we  
221 use both sun and shade leaves together. Leaf chemical properties that are most relevant for  
222 photosynthetic activity were expressed as mass-based as well as area-based values (chlorophyll a,  
223 chlorophyll b, bulk carotenoids, N, and P). For all analyses, if necessary, leaf traits were log-  
224 transformed to improve the normality of residuals. A CWM was calculated for each trait, weighted  
225 by the abundance of each species in each plot (Pla *et al.* 2012). Values of dark respiration fluxes and  
226  $\delta^{13}\text{C}$  were converted to positive values for ease of interpretation. To characterise soil properties in  
227 relation to land use, we performed a principal component analysis (PCA) of soil chemical properties  
228 across the eight plots with the measurements of total concentrations of P, C, N, Mg, K, Ca,

229 exchangeable Mg, K, Ca,  $\text{NH}_4^+$  and  $\text{NO}_3^-$ , extractable P, pH and cation exchange capacity (CEC). To  
230 visualise the distribution of CWM traits across forest types and test hypothesis 1, we conducted a  
231 PCA using centred and standardised CWM trait values for each study plot.

232 In order to test hypothesis 2, site scores from the first two PCA axes of soil properties, along with  
233 a binary variable representing logging history, were used as predictors in multiple regression models  
234 to partition the variance in each CWM trait. The first two principal components from the soil PCA  
235 were both statistically independent of logging history (linear model:  $p \geq 0.48$ ). Nevertheless, because  
236 the three predictors were weakly but non-significantly inter-correlated ( $R^2 \leq 0.29$ ), we used the  
237 hierarchical partitioning method of Chevan and Sutherland (1991), as implemented in the hier.part  
238 library of R, to estimate the variance in functional traits explained by each. This technique calculates  
239 the  $R^2$  of every possible model, then averages over this set of models to allocate the variance  
240 explained by each predictor variable. It thereby overcomes the effect of the order that inter-  
241 correlated variables enter a model on the inferred variance explained by each variable (Chevan &  
242 Sutherland 1991).

243 To address hypothesis 3, we computed plot-level values of functional diversity (FD) as Rao's  
244 quadratic entropy (Rao's Q), which is the sum of the pairwise distances between species in  
245 multidimensional trait space weighted by their relative abundance (Rao 1982) and compared forest  
246 types with ANOVA.

247 All analyses were performed using R 3.4.0 (R Core Team 2017).

248

## 249 Results

### 250 *Forest structure, species composition and soil properties of plots*

251 The study plots varied substantially in basal area (BA) and stem density, with BA varying fourfold  
252 ( $10.9 \text{ m} - 41.8 \text{ m}^2 \text{ ha}^{-1}$ , Table 1). Basal area was significantly lower in the SL plots but the range  
253 among plots within each forest type was similar (means: SL  $17.8 \pm 12.7$ , OG  $34.8 \pm 14.9 \text{ m}^2 \text{ ha}^{-1}$ ;  
254 ANOVA:  $F_{1,6} = 15.26$ ,  $p = 0.008$ ). Stem density ranged from 331 to 565  $\text{ha}^{-1}$ , peaking in plots with  
255 intermediate BA but was not associated with logging ( $F_{1,6} = 0.02$ ,  $p = 0.90$ ). Species richness was  
256 similar in OG and SL plots, ranging from 124 to 211 tree species  $\text{ha}^{-1}$  ( $F_{1,6} = 0.55$ ,  $p = 0.49$ ).

257 A PCA of soil chemical properties explained 69.5% on the first two principal axes (Fig. 1). The  
258 strongest gradient was defined by variation in total Mg, total P and CEC, whereas the second axis  
259 represented a gradient of total C, total N and exchangeable Ca to exchangeable K and  $\text{NH}_4^+$ . Nutrient  
260 concentrations varied markedly among plots, including 10-fold and 5-fold variation in total Mg and  
261 total P concentrations along axis 1, and 9-fold and 10-fold variation in exchangeable K and  $\text{NH}_4^+$

262 along axis 2 (Table 1). Soil properties differed among plots, but were independent of logging history,  
 263 indicated by the overlapping distribution of OG and SL plots in the PCA (Fig. 1).

264

#### 265 *Community-weighted mean traits*

266 Major gradients in CWM trait expression were visualised by PCA, with the first two axes  
 267 explaining 77.7% of the variance in functional traits (Fig. 2). There was a clear differentiation of  
 268 functional composition between OG and SL plots along the first principal component, indicated by a  
 269 distinct clustering of the study plots. Tree communities in OG plots were characterised by greater  
 270 investment in defence and tissue density, whereas SL tree communities expressed higher  
 271 photosynthetic activity and reduced investment into structural components (Table 2). Old-growth  
 272 forests were characterized by denser wood and tougher leaves. These traits reflect enhanced  
 273 structural investment, implying longer leaf life span and slower growth rates. Tree communities in SL  
 274 forest had higher photosynthetic activity represented by higher CWM values of  $A_{max}$  and  $A_{sat}$  and  
 275 higher  $R_d$ . These higher rates of gas exchange were supported by the expression of higher CWM  
 276 area-based pigment concentrations in SL communities and higher  $N_a$ ,  $N_m$  and  $P_a$  concentrations. Tree  
 277 communities in SL were enriched in  $^{13}C$  compared to OG communities, indicating greater water-use  
 278 efficiency.

279 The second axis of functional trait space represented tissue nutrient concentrations and leaf  
 280 area, but was independent of logging history (Fig. 2). This axis reflects covariation among CWM  
 281 values of leaf area, leaf  $P_m$ ,  $N_m$ ,  $Mg_m$  and  $Ca_m$  concentrations, and a negative association of these  
 282 traits with leaf  $C_m$  and tannin concentrations. Variability of these traits within both logged and  
 283 unlogged forests was high, which suggests that the expression of these traits is driven by underlying  
 284 soil properties rather than logging history.

285

#### 286 *Variance partitioning*

287 We grouped the functional traits based on their main association with leaf nutrients, photosynthesis  
 288 and structure. Partitioning the CWM response of traits to logging and the first two principal  
 289 components of soil properties showed that these factors explained up to 90% of the variation in  
 290 traits. Overall, the proportion of variance explained was on average 74.4%. (Fig. 3, Table S4).  
 291 Variation in mass-based concentrations of leaf  $Ca_m$ ,  $P_m$ ,  $N_m$  and  $K_m$  and to a lesser extent  $Mg_m$  were  
 292 associated with variation in soil properties. However, expressed on area basis, foliar  $P_a$  and  $N_a$   
 293 concentrations were mainly explained by logging. Variation in SLA and leaf thickness appears to  
 294 underlie the contrasting response of mass and area based traits. Community-weighted mean values  
 295 of  $C_m$ ,  $P_m$  and  $Ca_m$  were most strongly and significantly explained by the first principal component of  
 296 soil properties, reflecting the underlying gradients of soil total P and Ca concentrations along this

297 axis (Fig. 1). For leaf traits related to photosynthesis, 33.5–78.6% of variance was explained by  
298 logging and a much smaller proportion by soil PC1 (1.5–21.8%) or soil PC2 (0.9–35.6%), see Table S4.  
299 Structural traits were explained by a combination of both logging history and the independent  
300 effects of soil properties. Logging explained on average 39.2% of variance in traits reflecting tissue  
301 density and structural investment, such as specific force to punch and branch wood density, which  
302 had consistently lower values in logged forest plots. Community-weighted mean LDMC was unusual  
303 in that it was poorly explained by the predictor variables. In contrast, leaf size, expressed as CWM  
304 leaf area and leaf mass, increased significantly with increasing values of soil PC1, which was linked to  
305 plots with higher total N and exchangeable Ca concentrations (Fig. 1). There was a significant  
306 increase in CWM tannin concentrations in logged forest plots and at higher values of soil PC1.

307

### 308 *Functional diversity*

309 Functional diversity, expressed as Rao's Q, was similar between forest types (Fig. 4;  $F_{1,6} = 0.16$ ,  $p =$   
310 0.70), and neither logging nor soil properties explained a significant proportion of its variance (Fig.  
311 3). Variance in FD was greater among SL than OG forests (Fig. 4), indicating the heterogeneous  
312 conditions resulting from logging.

313

### 314 **Discussion**

315 Logging profoundly affected the expression of plant functional traits in Bornean tropical forests.  
316 Logging was the primary driver of variation in CWM values of functional traits (Fig. 2), and explained  
317 more variation than soil properties for 20 of 32 traits (Fig. 3). Similar impacts of logging on functional  
318 trait expression have been demonstrated in Neotropical forests (Baraloto *et al.* 2012; Carreño-  
319 Rocabado *et al.* 2012), although those studies did not analyse soil-related factors. Together, these  
320 studies confirm that CWM traits are highly sensitive to land-use change in tropical forests, but  
321 results from our study additionally highlight the context dependency of trait expression. Logging has  
322 impacted over half of all tropical forests globally, and over 70% of forests in Sabah (Bryan *et al.* 2013;  
323 Potapov *et al.* 2017). Therefore, the effect of logging on the expression of functional traits is likely to  
324 pervade tropical forest landscapes and impact ecosystem processes. Effects on other trophic levels  
325 are also likely. For example, herbivorous insects are sensitive to leaf traits such as LDMC and phenol  
326 concentrations (Hevia *et al.* 2017). Moreover, logging-associated changes in forest structure and  
327 CWM traits drive altered patterns of productivity in tropical forests (Pfeifer *et al.* 2016; Riutta *et al.*  
328 2018).

329 In our study, the principal axis of functional trait space defined a clear gradient from values of  
330 traits that maximise carbon capture and growth, which were predominantly expressed in selectively

331 logged forests, to greater allocation to tissue persistence and stability, which were predominantly  
332 expressed in old-growth forests. This strong signal of anthropogenic disturbance is partly congruent  
333 with the leaf economics spectrum, which differentiates species along a gradient based on leaf traits  
334 contributing to resource acquisition and conservation (Wright *et al.* 2004; Díaz *et al.* 2016). Thus,  
335 trees in logged forest communities expressed higher CWM values of area-based measures of N, P  
336 and pigments, whereas old-growth forest communities expressed low CWM values of these traits  
337 and higher values of traits conferring structural stability and resistance to herbivory, such as branch  
338 wood density and leaf toughness. Supporting hypothesis 1, disturbance enhanced the recruitment of  
339 species possessing traits that confer rapid carbon capture and investment in fast growth rates  
340 (Baraloto *et al.* 2012; Carreño-Rocabado *et al.* 2012, 2016). We amplify previous results by  
341 additionally demonstrating that logged forest communities expressed higher CWM values of  $A_{\text{sat}}$ ,  
342  $A_{\text{max}}$  and  $R_d$ , whereas old-growth forest communities were characterised by higher values of traits  
343 conferring structural stability of wood and leaves.

344 We observed lower CWM values of SLA in logged forests, in contrast to results from French  
345 Guiana (Baraloto *et al.* 2012), and contrary to the expectation that SLA scales positively with  $A_{\text{max}}$   
346 and foliar  $N_a$  and  $P_a$  concentrations among species (Wright *et al.* 2004). The decoupling of SLA from  
347 other leaf-economic traits may be explained by the abundance of pioneer species, which recruit  
348 following disturbance and are adapted to resist the more exposed and potentially desiccating  
349 conditions created by logging (Hardwick *et al.* 2015). A reduction in SLA may contribute to  
350 photosynthetic water-use efficiency, especially when combined with enhanced investment in  
351 photosynthetic enzymes, to ensure draw-down of internal  $\text{CO}_2$  concentrations at a given stomatal  
352 conductance (Reich *et al.* 2003). The greater enrichment of CWM  $\delta^{13}\text{C}$  of logged forest tree  
353 communities demonstrates lower discrimination for the heavier  $^{13}\text{C}$  isotope and provides  
354 independent evidence of enhanced integrated water-use efficiency for trees in this hotter and drier  
355 environment (Farquhar *et al.* 1989; Rumman *et al.* 2018). The absence of a shift in SLA in response to  
356 logging in French Guianan forests (Baraloto *et al.* 2012) suggests that logging imposes a more  
357 extreme environmental contrast for trees growing in the less seasonal climate of Borneo than in  
358 seasonal French Guiana. The impact of logging may be more severe in tree communities not adapted  
359 to drought and emphasises the potential sensitivity of Bornean forests to future climatic change.

360 We show that trait expression responded independently to logging disturbance and soil  
361 properties. Variation in soil properties can be attributed to interactions between underlying soil  
362 texture and mineralogy, on one hand, and the impacts of logging disturbances including soil  
363 inversion, removal, and compaction on the other (Pinard *et al.* 2000). However, soil properties did  
364 not differ systematically between old-growth and selectively logged forests (Fig. 1), owing to

365 variation in underlying soil types and the heterogeneous nature of logging. This allowed us to assess  
366 their independent effects on the expression of CWM functional traits. For most of the mass-based  
367 nutrient concentrations, variation in soil properties explained more variation in CWM trait values  
368 than did logging, whereas traits linked to photosynthesis and carbon capture were more sensitive to  
369 logging than to variation in soil properties (Fig. 3). The mechanisms underlying these associations  
370 deserve further study.

371 Moreover, we provide clear evidence that soil properties act on trait expression in two  
372 independent ways; the first axis reflected total pools of nutrients, and the second reflected nutrient  
373 availability (Fig. 1). Community-weighted mean values of leaf area, leaf dry mass and foliar  
374 concentrations of  $P_m$ ,  $P_a$ ,  $N_m$  and  $Ca_m$  all increased significantly in response to the first axis of  
375 variation in soil properties, whereas leaf  $C_m$  and tannin concentrations decreased along this gradient  
376 (Fig. 3). The second component of soil variation also influenced some leaf traits, particularly  $\delta^{13}C$ ,  $N_a$   
377 and  $P_a$  concentrations (Table 2, Table S4). Moreover, most traits were influenced either by the first  
378 or second axis of variation in soil properties, but rarely both. This finding may reflect a trade-off in  
379 how plants interact with local edaphic conditions, as suggested in other systems (Laliberte *et al.*  
380 2015); a key future challenge will be to disentangle the mechanisms underpinning these contrasting  
381 responses of functional traits to the soil environment. Our results highlight the need to consider the  
382 context dependency of drivers of variation in functional traits. Landscape-level predictions of change  
383 in functional trait expression in response to anthropogenic disturbance will need to account for the  
384 additional effects of soil properties.

385 Despite the large variation in CWM traits, FD did not differ between logged and old-growth  
386 forest, which is consistent with our third hypothesis and results from Neotropical forests (Fig. 4;  
387 Baraloto *et al.* 2012; Carreño-Rocabado *et al.* 2012). This finding emphasises that forests can retain  
388 species richness and trait variation, yielding similar FD, despite logging. In line with Mayfield *et al.*  
389 (2010), logging did not result in loss of FD but in shifts of numerous CWM trait values, indicating a  
390 lower sensitivity of multi-trait FD to these changes (Ricotta & Moretti 2011). FD was more variable  
391 among logged forests than old-growth forests, probably owing to variation in logging history and  
392 intensity, which affect forest structure (Cannon *et al.* 1994; Berry *et al.* 2008), microclimatic  
393 conditions (Hardwick *et al.* 2015) and ecosystem functions (Mayfield *et al.* 2006; Both *et al.* 2017;  
394 Riutta *et al.* 2018). The substantial variance in FD among the disturbed plots highlights the challenge  
395 of predicting the impacts of anthropogenic modification on FD in environments where the outcomes  
396 may be highly context-dependent (Costantini *et al.* 2016).

397 Assessments of functional composition and diversity at a community scale are critical as  
398 human-modified landscapes become more extensive and exert an increasing role in the provision of



399 ecosystem services (Berry *et al.* 2010; Gibson *et al.* 2011). However, uncertainty remains over how  
400 changes in community trait expression will affect ecosystem functioning and resilience after  
401 selective logging and other forms of disturbance (Laliberté *et al.* 2010; Mayfield *et al.* 2010; Edwards  
402 *et al.* 2014). Part of that uncertainty arises because rare tree species may contribute substantially to  
403 resilience (Mouillot *et al.* 2013b), but tend to be under-represented in traditional sampling designs  
404 adopted for measuring ecosystem functions. Our nested sampling design explicitly resolved this  
405 issue by selecting both common and rare species across the full range of size classes. We therefore  
406 advocate this approach in future assessments of trait expression at the community scale.

407 We demonstrate a consistent shift in community-level trait expression in response to logging,  
408 reflecting a transition from an old-growth forest dominated by individuals with resource conserving,  
409 structurally persistent tissues to logged forests manifesting greater capacity for carbon assimilation  
410 and vegetative growth. Strikingly, there was a second, independent, axis of functional trait variation  
411 reflecting variation in soil properties (i.e. nutrient availability and chemistry including pH), which  
412 explained variation in leaf size and mass-based foliar nutrient concentrations. The elucidation of  
413 these orthogonal dimensions of plant trait variation was made possible by the measurement of  
414 numerous functionally relevant traits and their consideration at the community level, as well as by  
415 the inclusion of rare species. These results provide a basis for predicting how pervasive logging  
416 disturbance combines with natural gradients to determine trait expression and ecosystem  
417 functioning across human-modified tropical landscapes.

418

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434

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440

For Peer Review

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613 **Figure captions**

614

615 Figure 1: Principal component analysis of plot-level soil properties. Notably, plots with similar logging  
616 histories do not cluster together. The highest loadings on the first axis are CEC (12.6%), total Mg  
617 (12.3%) and total P (12.2%). The highest loadings on the second axis are exchangeable K (12.0%),  
618 total N (11.1%) and exchangeable Ca (11.1%). See Table S2 for all PCA loadings.

619

620 Figure 2: PCA of plot-level community-weighted mean functional traits. Plots cluster by logging  
621 history, with increased values of traits that maximise carbon capture and growth in logged forest  
622 communities and greater allocation to tissue persistence and structural stability in old-growth  
623 forests. The highest loadings on the first axis are chlorophyll  $b_m$  (4.44%),  $N_a$  (4.37%), branch wood  
624 density (4.25%),  $A_{sat}$  (4.25%),  $A_{max}$  (4.21%) and SLA (4.17%). The highest loadings on second axis are  
625  $P_m$  (7.45%), LA (7.02%), tannins (6.22%),  $C_m$  (5.88%) and leaf dry weight (5.75%). Mass-based  
626 nutrients are denoted by superscript "m" and area-based values by superscript "a". See Table S3 for  
627 all PCA loadings.

628

629 Figure 3: Proportion of variance in community-weighted mean functional trait values explained by  
630 forest type and the first two principal components of soil properties (Fig. 1). Functional traits are  
631 grouped by the ecosystem function to which they most contribute. Statistical significance is derived  
632 from linear regression models, asterisks indicate significance level, '+' and '-' indicate the direction of  
633 the relationship. For forest type '+' indicates that trait values were greater in selectively logged than  
634 old-growth forests (i.e. positive with first PC axis). For variance explained by soil, '+' indicates  
635 positive relationship with the respective PC axis. See Table 2 and Table S4 for detailed results.

636

637 Figure 4: Box-and-whisker plots showing the median, upper and lower quartile of functional diversity  
638 calculated as Rao's Q with no significant difference between forest type (ANOVA:  $F_{1,6} = 0.16$ ,  $p =$   
639  $0.70$ ).

640



641 **Tables**

642 Table 1: Study plot description and soil properties, plots are listed with decreasing basal area. Basal area, stem density and number of tree species refers to  
 643 all tree individuals  $\geq 10$  cm dbh.

Plot name	Location	Total basal area [m <sup>2</sup> ha <sup>-1</sup> ]	Stem density	Number of species	C [%]	N [%]	Total P [mg kg <sup>-1</sup> ]	Total Ca [mg kg <sup>-1</sup> ]	Total K [mg kg <sup>-1</sup> ]	Total Mg [mg kg <sup>-1</sup> ]	eCEC [mmol <sup>+</sup> kg <sup>-1</sup> ]	pH (H <sub>2</sub> O)	NO <sub>3</sub> <sup>-</sup> [μg 10 cm <sup>2</sup> 14 days <sup>-1</sup> ]	NH <sub>4</sub> <sup>+</sup> [μg 10 cm <sup>2</sup> 14 days <sup>-1</sup> ]	Exchangable Ca [μg 10 cm <sup>2</sup> 14 days <sup>-1</sup> ]	Exchangable Mg [μg 10 cm <sup>2</sup> 14 days <sup>-1</sup> ]	Exchangable K [μg 10 cm <sup>2</sup> 14 days <sup>-1</sup> ]	Extractable P [μg 10 cm <sup>2</sup> 14 days <sup>-1</sup> ]	
<b>Old-growth forest</b>																			
MLA-01	MBCA	41.8	396	141	1.67	0.14	163.01	50.06	101.03	39.67	36.44	3.95	59.6 ± 45.2	10.5 ± 3.11	237 ± 130	152 ± 80.6	265 ± 154	0.23 ± 0.262	
MLA-02	MBCA	37.3	484	170	2.00	0.17	266.44	194.68	80.62	99.41	43.18	4.25	184 ± 103	6.37 ± 1.52	417 ± 476	303 ± 215	151 ± 80.2	0.433 ± 0.325	
DAN-04	DVCA	30.8	456	128	1.80	0.18	557.21	603.28	16.12	389.63	63.88	5.81	75.2 ± 57.2	11.9 ± 12.7	480 ± 596	235 ± 187	268 ± 158	0.177 ± 0.134	
DAN-05	DVCA	29.1	413	144	1.84	0.17	255.59	140.50	77.03	180.37	44.90	4.46	154 ± 164	5 ± 1.88	737 ± 682	271 ± 183	305 ± 269	1.03 ± 1.89	
<b>Selectively logged forest</b>																			
SAF-03	SAFE project	25.8	565	211	1.89	0.16	231.05	40.42	115.55	38.65	35.66	3.68	83.8 ± 63.7	26.4 ± 44.9	392 ± 265	318 ± 201	332 ± 308	8.51 ± 12.4	
SAF-04	SAFE project	19.5	465	188	7.15	0.32	117.07	107.71	67.21	41.36	14.55	3.90	83.4 ± 60.5	2.68 ± 4.32	1.35*10 <sup>3</sup> ± 621	310 ± 94.1	45.1 ± 58	1.18 ± 1.21	
SAF-02	SAFE project	14.8	416	124	1.51	0.12	137.33	160.50	105.86	41.80	49.38	4.04	340 ± 239	23.9 ± 33.2	182 ± 187	191 ± 173	401 ± 239	0.464 ± 0.214	
SAF-01	SAFE project	10.9	331	129	3.08	0.28	375.15	887.03	96.37	266.36	70.76	4.66	20 ± 17.5	16.8 ± 18.7	212 ± 269	79 ± 67.6	245 ± 90.6	3.03 ± 2.48	

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646 Table 2: Results from linear regression models from which the explained variance was generated,  
 647 factors are the categorical ‘forest type’ (OG – old-growth, SL – selectively logged), and continuous  
 648 ‘soil PC1’ and ‘soil PC2’. For analyses values of dark respiration  $R_d$  fluxes and  $\delta^{13}C$  were converted to  
 649 positive values for ease of interpretation, here untransformed values are shown. For abbreviations  
 650 and description of the functional traits, see table S1.

Functional trait	CWM trait value (and 95% confidence intervals)		Forest type		soil PC1		soil PC2	
	OG	SL	F-value	p-value	F-value	p-value	F-value	p-value
Rao's Q	0.791 (0.735 - 0.847)	0.789 (0.736 - 0.842)	0.197	0.680 ns	0.119	0.748 ns	3.060	0.155 ns
$\delta^{15}N$ [‰]	1.64 (0.966 - 2.32)	0.846 (0.204 - 1.49)	5.032	0.088 ns	0.185	0.689 ns	0.527	0.508 ns
$Ca_m$ [mg g <sup>-1</sup> ]	7.09 (6.12 - 8.2)	6.6 (5.75 - 7.59)	6.114	0.069 ns	17.555	0.014 *	1.578	0.278 ns
$Mg_m$ [mg g <sup>-1</sup> ]	2.57 (2.06 - 3.2)	2.41 (1.96 - 2.97)	0.724	0.443 ns	1.275	0.322 ns	0.490	0.523 ns
$K_m$ [mg g <sup>-1</sup> ]	9.99 (8.4 - 11.9)	10.8 (9.13 - 12.7)	0.923	0.391 ns	0.321	0.601 ns	2.758	0.172 ns
$N_m$ [%]	1.83 (1.74 - 1.91)	1.97 (1.89 - 2.05)	9.191	0.039 *	7.746	0.050 *	4.523	0.101 ns
$P_m$ [mg g <sup>-1</sup> ]	0.998 (0.912 - 1.09)	0.99 (0.909 - 1.08)	1.484	0.290 ns	13.219	0.022 *	0.444	0.542 ns
$C_m$ [%]	44.6 (44.3 - 44.9)	44.7 (44.4 - 45)	12.266	0.025 *	89.717	0.001 ***	0.240	0.650 ns
$\delta^{13}C$ [‰]	-32.4 (-32.1 - (-32.8))	-31.4 (-31.1 - (-31.8))	28.775	0.006 **	8.747	0.042 *	11.356	0.028 *
$P_a$ [mg mm <sup>-2</sup> ]	$6.62 \times 10^{-5}$ ( $6.21 \times 10^{-5}$ - $7.06 \times 10^{-5}$ )	$8.04 \times 10^{-5}$ ( $7.57 \times 10^{-5}$ - $8.54 \times 10^{-5}$ )	33.208	0.005 **	12.267	0.025 *	10.391	0.032 *
$N_a$ [mg mm <sup>-2</sup> ]	0.128 (0.12 - 0.136)	0.166 (0.158 - 0.174)	97.699	0.001 ***	0.520	0.511 ns	19.704	0.011 *
$R_d$ [ $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ]	-1.03 (-0.871 - (-1.19))	-1.25 (-1.09 - (-1.4))	7.708	0.050 ns	1.546	0.282 ns	5.759	0.074 ns
$A_{\text{max}}$ [ $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ]	11.7 (8.94 - 14.5)	18 (15.4 - 20.6)	23.237	0.009 **	0.878	0.402 ns	4.268	0.108 ns
$A_{\text{sat}}$ [ $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ ]	4.08 (2.66 - 5.5)	7.03 (5.69 - 8.38)	20.267	0.011 *	0.284	0.622 ns	4.495	0.101 ns
Carotenoids <sub>a</sub> [mg mm <sup>-2</sup> ]	$4.74 \times 10^{-5}$ ( $4.47 \times 10^{-5}$ - $5.02 \times 10^{-5}$ )	$5.54 \times 10^{-5}$ ( $5.28 \times 10^{-5}$ - $5.79 \times 10^{-5}$ )	47.726	0.002 **	1.707	0.261 ns	7.957	0.048 *
Carotenoids <sub>m</sub> [mg g <sup>-1</sup> ]	0.687 (0.667 - 0.708)	0.667 (0.647 - 0.687)	6.353	0.065 ns	1.555	0.281 ns	0.264	0.635 ns
Chl b <sub>a</sub> [mg mm <sup>-2</sup> ]	$7.42 \times 10^{-5}$ ( $6.79 \times 10^{-5}$ - $8.04 \times 10^{-5}$ )	$7.95 \times 10^{-5}$ ( $7.36 \times 10^{-5}$ - $8.55 \times 10^{-5}$ )	5.552	0.078 ns	1.678	0.265 ns	1.333	0.313 ns
Chl b <sub>m</sub> [mg g <sup>-1</sup> ]	1.09 (1.05 - 1.13)	0.97 (0.931 - 1.01)	40.379	0.003 **	0.005	0.945 ns	2.605	0.182 ns
Chl a <sub>a</sub> [mg mm <sup>-2</sup> ]	$1.8 \times 10^{-4}$ ( $1.69 \times 10^{-4}$ - $1.92 \times 10^{-4}$ )	$2.06 \times 10^{-4}$ ( $1.95 \times 10^{-4}$ - $2.17 \times 10^{-4}$ )	29.141	0.006 **	1.457	0.294 ns	7.060	0.057 ns
Chl a <sub>m</sub> [mg g <sup>-1</sup> ]	2.62 (2.54 - 2.71)	2.49 (2.41 - 2.57)	13.853	0.020 *	0.967	0.381 ns	0.020	0.894 ns
SLA [mm <sup>2</sup> mg <sup>-1</sup> ]	16.3 (15.3 - 17.4)	13.2 (12.2 - 14.1)	52.069	0.002 **	3.036	0.156 ns	5.338	0.082 ns
LA [mm <sup>2</sup> ]	$1.37 \times 10^4$ ( $1.13 \times 10^4$ - $1.65 \times 10^4$ )	$1.31 \times 10^4$ ( $1.1 \times 10^4$ - $1.57 \times 10^4$ )	1.678	0.265 ns	11.116	0.029 *	1.898	0.240 ns
Leaf dry weight [mg]	922 (784 - $1.08 \times 10^3$ )	$1.08 \times 10^3$ (930 - $1.27 \times 10^3$ )	2.077	0.223 ns	10.600	0.031 *	6.120	0.069 ns
Leaf thickness [mm]	0.221 (0.209 - 0.233)	0.236 (0.224 - 0.249)	7.196	0.055 ns	0.213	0.668 ns	2.319	0.203 ns
Specific force to punch [N mm <sup>-2</sup> ]	1.23 (1.03 - 1.47)	0.889 (0.751 - 1.05)	13.209	0.022 *	0.431	0.547 ns	0.013	0.915 ns
Force to punch [N mm <sup>-1</sup> ]	0.266 (0.227 - 0.311)	0.212 (0.183 - 0.247)	7.611	0.051 ns	0.300	0.613 ns	0.260	0.637 ns
LDMC [mg g <sup>-1</sup> ]	416 (391 - 440)	410 (387 - 433)	0.143	0.725 ns	0.332	0.595 ns	0.079	0.793 ns
Branch density [g cm <sup>-3</sup> ]	0.564 (0.528 - 0.599)	0.493 (0.46 - 0.526)	15.309	0.017 *	2.442	0.193 ns	1.522	0.285 ns
Phenol <sub>m</sub> [mg g <sup>-1</sup> ]	36.4 (33.7 - 39.1)	42.7 (40.2 - 45.3)	29.960	0.005 **	5.004	0.089 ns	2.762	0.172 ns
Tannin <sub>m</sub> [mg g <sup>-1</sup> ]	8.56 (7.7 - 9.41)	9.41 (8.6 - 10.2)	8.888	0.041 *	8.069	0.047 *	0.004	0.951 ns
Lignin & recalitrants [%]	19.4 (17.3 - 21.4)	17.3 (15.3 - 19.2)	2.707	0.175 ns	3.333	0.142 ns	0.080	0.791 ns
Cellulose [%]	22.5 (20.9 - 24)	20.9 (19.4 - 22.4)	3.343	0.142 ns	0.313	0.606 ns	0.995	0.375 ns
Hemicellulose [%]	12.3 (11.4 - 13.1)	11.8 (11 - 12.6)	1.728	0.259 ns	0.767	0.431 ns	0.270	0.631 ns

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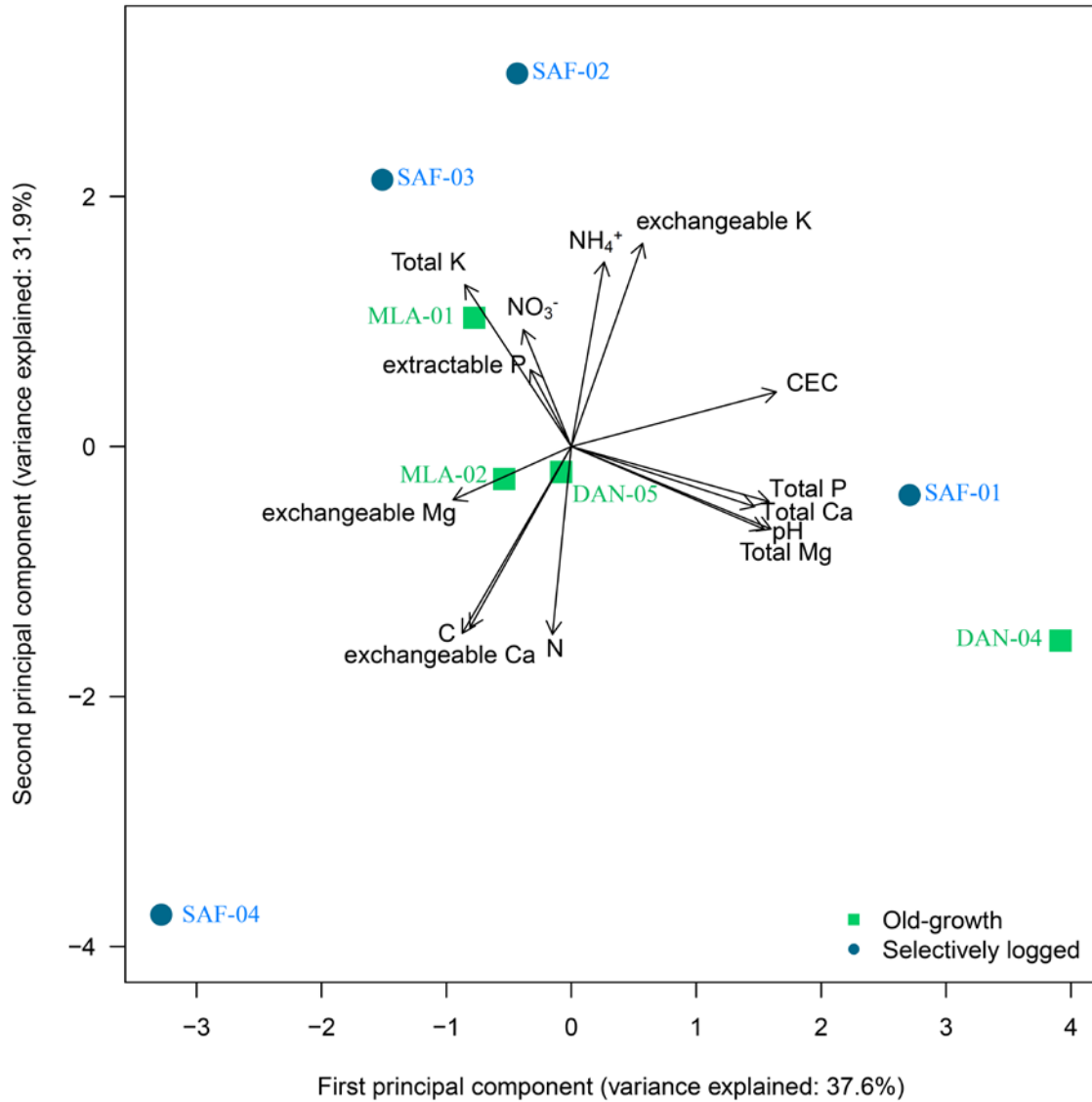


Figure 1: Principal component analysis of plot-level soil properties. Notably, plots with similar logging histories do not cluster together. The highest loadings on the first axis are CEC (12.6%), total Mg (12.3%) and total P (12.2%). The highest loadings on the second axis are exchangeable K (12.0%), total N (11.1%) and exchangeable Ca (11.1%). See Table S2 for all PCA loadings.

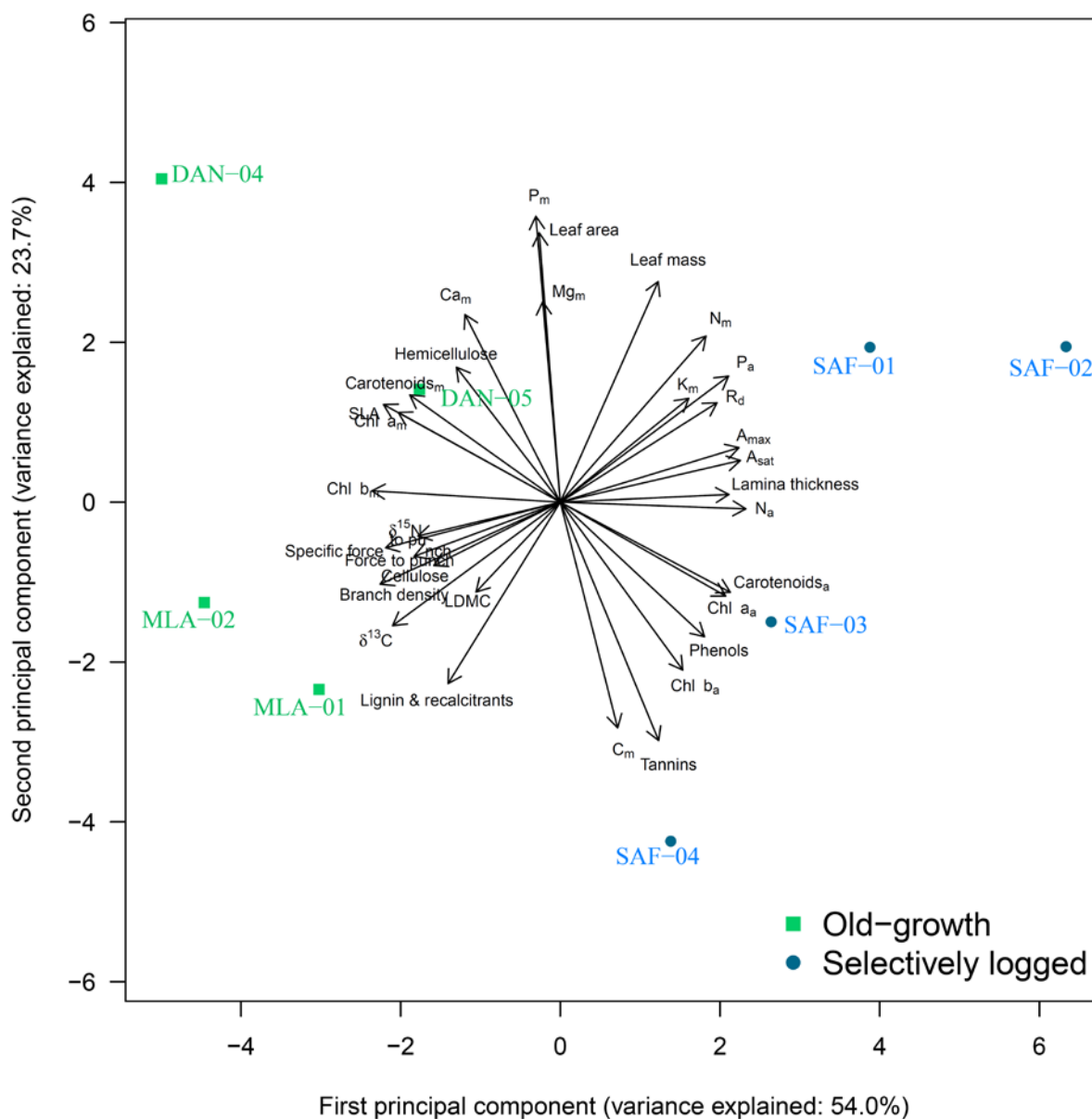


Figure 2: PCA of plot-level community-weighted mean functional traits. Plots cluster by logging history, with increased values of traits that maximise carbon capture and growth in logged forest communities and greater allocation to tissue persistence and structural stability in old-growth forests. The highest loadings on the first axis are chlorophyll  $b_m$  (4.44%),  $N_a$  (4.37%), branch wood density (4.25%),  $A_{sat}$  (4.25%),  $A_{max}$  (4.21%) and SLA (4.17%). The highest loadings on second axis are  $P_m$  (7.45%), LA (7.02%), tannins (6.22%),  $C_m$  (5.88%) and leaf dry weight (5.75%). Mass-based nutrients are denoted by superscript “m” and area-based values by superscript “a”. See Table S3 for all PCA loadings.

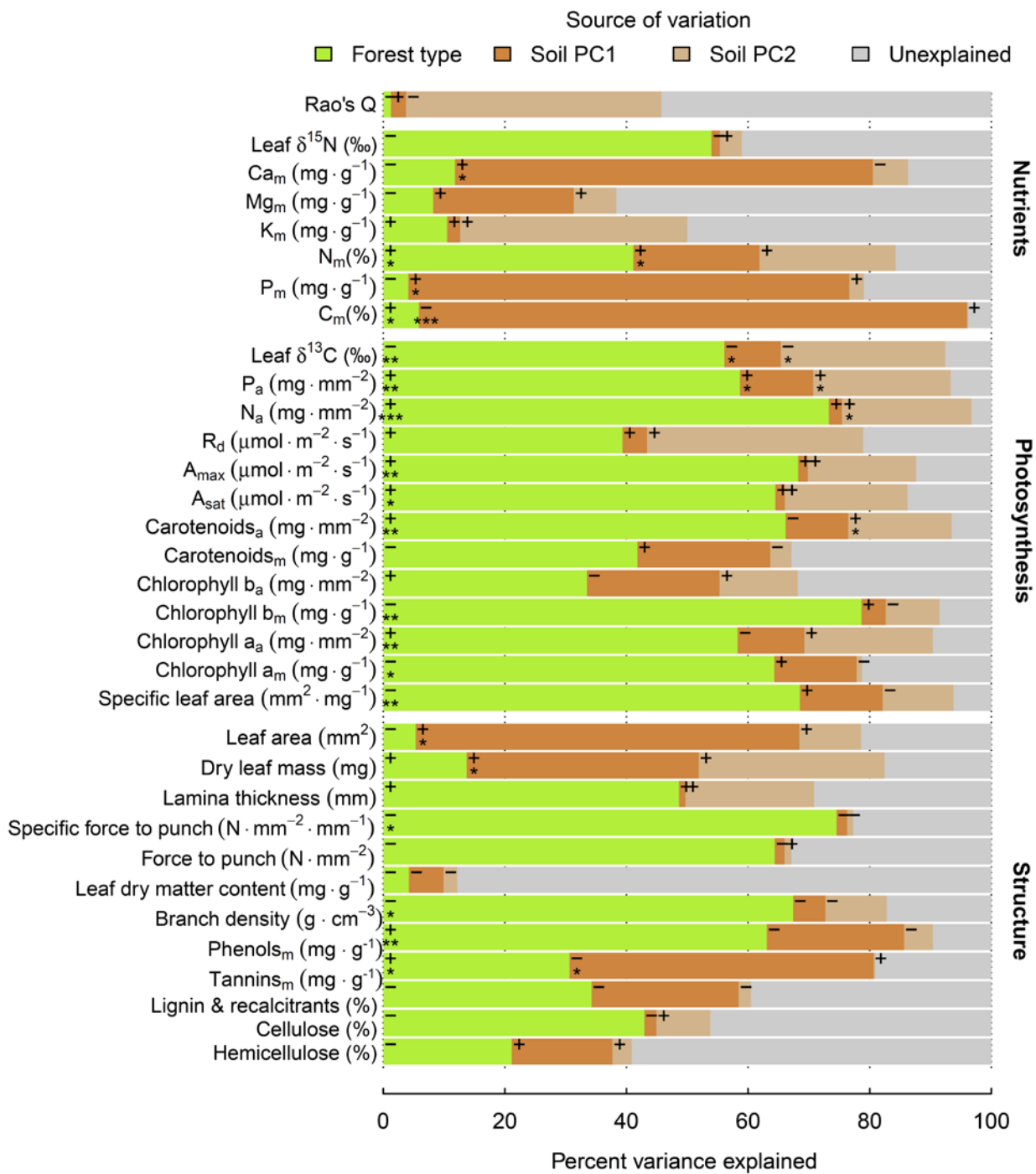


Figure 3: Proportion of variance in community-weighted mean functional trait values explained by forest type and the first two principal components of soil properties (Fig. 1). Functional traits are grouped by the ecosystem function to which they most contribute. Statistical significance is derived from linear regression models, asterisks indicate significance level, '+' and '-' indicate the direction of the relationship. For forest type '+' indicates that trait values were greater in selectively logged than old-growth forests (i.e. positive with first PC axis). For variance explained by soil, '+' indicates positive relationship with the respective PC axis. See Table 2 and Table S4 for detailed results.

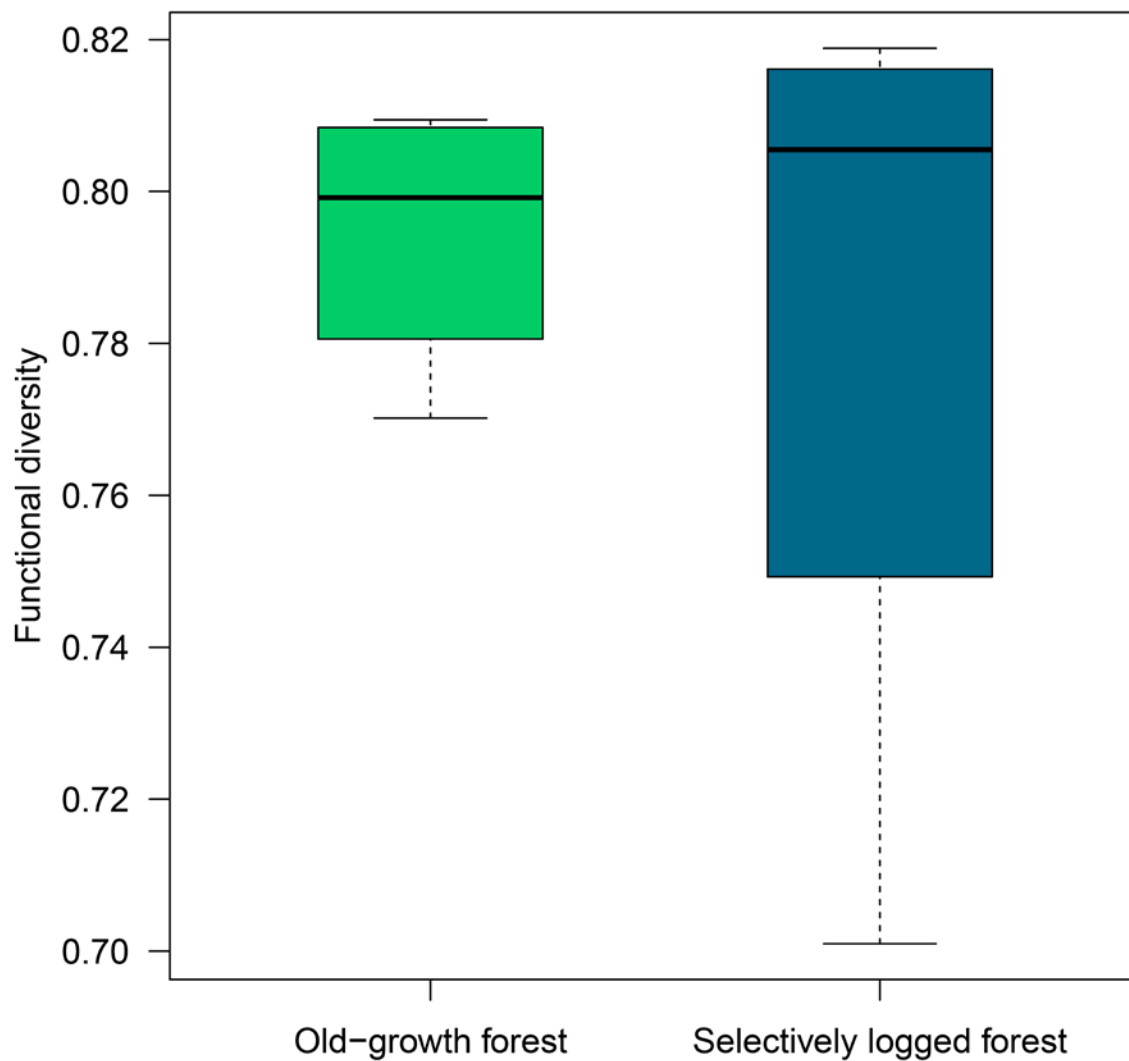


Figure 4: Box-and-whisker plots showing the median, upper and lower quartile of functional diversity calculated as Rao's Q with no significant difference between forest type (ANOVA:  $F_{1,6} = 0.16$ ,  $p = 0.70$ ).