REVIEW

Long Non-Coding Small Nucleolar RNA Host Genes (SNHGs) in Endocrine-Related Cancers

This article was published in the following Dove Press journal: OncoTargets and Therapy

Yuan Qin (1)*
Wei Sun*
Zhihong Wang (1)
Wenwu Dong (1)
Liang He
Ting Zhang
Hao Zhang (1)

Department of Thyroid Surgery, The First Hospital of China Medical University, Shenyang 110001, Liaoning Province, People's Republic of China

*These authors contributed equally to this work

Abstract: Long non-coding RNAs (lncRNAs) are emerging regulators of a diverse range of biological processes through various mechanisms. Genome-wide association studies of tumor samples have identified several lncRNAs, which act as either oncogenes or tumor suppressors in various types of cancers. Small nucleolar RNAs (snoRNAs) are predominantly found in the nucleolus and function as guide RNAs for the processing of transcription. As the host genes of snoRNAs, lncRNA small nucleolar RNA host genes (SNHGs) have been shown to be abnormally expressed in multiple cancers and can participate in cell proliferation, tumor progression, metastasis, and chemoresistance. Here, we review the biological functions and emerging mechanisms of SNHGs involved in the development and progression of endocrine-related cancers including thyroid cancer, breast cancer, pancreatic cancer, ovarian cancer and prostate cancer.

Keywords: endocrine, cancers, lncRNA, SNHG

Introduction

Long non-coding RNAs (lncRNAs, >200 nucleotides in length) are emerging regulators of gene transcription. The human genome estimated to encode >28,000 lncRNAs, but only 15,778 lncRNAs are annotated in the current GENECODE version 27. Therefore, more lncRNAs are yet to be discovered. Moreover, the known lncRNAs have not been studied in depth.

Accumulating evidence suggests lncRNAs play key roles in the development and progression of several cancers, acting as either oncogenes or tumor suppressors. LncRNAs can regulate transcription, translation, protein modification, and the formation of RNA-protein or protein-protein complexes, depending on the cellular location. For example, lncRNAs primarily located in the nucleus are involved in transcriptional regulation and mRNA processing, while cytoplasmic lncRNAs play roles in modulating mRNA translation by competing with proteins or in miRNA-mediated mRNA decoy. 5,6

Small nucleolar RNAs (snoRNAs, 60–300 nucleotides in length) are more well-characterized than lncRNAs and are predominantly found in the nucleolus. Most snoRNAs function as guide RNAs for the post-transcriptional modification of ribosomal RNAs and some spliceosomal RNAs, with some involved in the nucleolytic processing of the original rRNA transcript. As shown in Figure 1, the majority of snoRNAs are encoded (hosted) in the introns of protein-coding and non-protein-coding genes, termed small nucleolar RNA host genes (SNHGs). Primary RNA transcripts of host genes (including all exons and introns with their snoRNAs) are cut into different exons and introns. Exons are then re-spliced and function in the cytoplasm, while the introns are further processed into snoRNAs and play roles in the nucleolus.

Correspondence: Hao Zhang Department of Thyroid Surgery, The First Hospital of China Medical University, 155 Nanjing Bei Street, Shenyang, Liaoning 110001, People's Republic of China Email haozhang@cmu.edu.cn

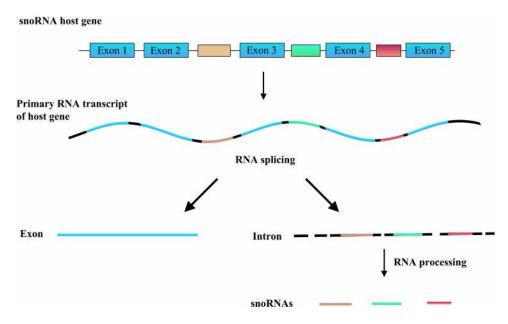


Figure I The synthetic pathway of snoRNAs.

Currently, there are 22 members of SNHG family (SNHG1 to SNHG22) that have been shown to regulate proliferation, apoptosis, invasion, and migration in multiple cancers, including endocrine-related cancers (as summarized in Tables 1 and 2). These 22 SNHGs have diverse activities and mechanisms of action. For example, SNHG1 has been shown to promote colorectal cancer cell growth by modulating histone methylation of gene promoters of the Kruppel Like Factor 2 (KLF2, a member of the KLF family, also exerts tumor-suppressive roles) and the cyclin-dependent kinase 4 inhibitor B (CDKN2B, a tumor suppressor). 12 SNHG1 can also act as a sponge for miR-154-5p to upregulate expression of G1/S-specific cyclin-D2 (CCND2, which is involved in cell cycle progression). 12 Meanwhile, SNHG13 serves as a competing endogenous RNA (ceRNA) of miR-34a-5p, leading to the derepression of Jagged 1 (JAGI) expression, which eventually triggers resistance to docetaxel in prostate cancer. 13

This review aims to provide an overview on the current understanding of the regulation and function of SNHGs in endocrine-related cancers that arise from the endocrine glands or neuroendocrine tissues, including thyroid cancer, breast cancer, pancreatic cancer, ovarian cancer, and prostate cancer. 14

Thyroid Cancer

Thyroid cancer is the most common malignancy of the endocrine system with enormous heterogeneity in terms of morphological features and prognosis. 15 Although the

majority of cases of thyroid cancer tend to be biologically indolent and have an excellent prognosis, some are associated with more aggressive clinical behavior. 16

SNHG1 may act as an oncogene in thyroid cancer by competing with miR-199a-5p and upregulating the expression of its target gene, the transcription factor (TF) SP1. In turn, SP1 targets the promoter region of SNHG1 and promote its transcription, forming a positive feedback loop to promote cancer cell proliferation and invasion.¹⁷ Conversely, low expression of SNHG2, also known as growth arrest specific transcript 5 (GAS5), is associated with poor prognosis of patients with thyroid cancer. 18 Mechanistically, GAS5 acts as a sponge for miR-222-3p, thereby modulating the expression of the phosphatase and tensin homolog (PTEN), leading to PTEN/protein kinase B (AKT) pathway activation and the suppression of thyroid cancer cell proliferation.¹⁹

SNHG7 is also markedly upregulated in thyroid cancer samples, with high SNHG7 expression associated with shorter survival times.²⁰ Indeed, SNHG7 knockdown leads to a suppression of thyroid cancer cell proliferation and migration, and induction of apoptosis via downregulating the acyl-CoA synthetase long chain family member 1 (ACSL1) and the brain-derived neurotrophic factor (BDNF). 21,22 In addition, bioinformatics analysis showed SNHG7 was associated with the processes of "protein translation", "viral life cycle", "RNA processing", "mRNA splicing", "histone ubiquitination", "endoplasmic reticulum-to-Golgi vesicle-mediated transport", "sister chromatid cohesion", "DNA damage checkpoint

Table I Characteristics of SNHG Members

SNHG Member	Aliases	Chromosomal Location	GENE ID	Associated Endocrine-Related Cancers
SNHGI	LINC00057, NCRNA00057, U22HG, UHG, IncRNA16	11q12.3	23642	Thyroid Cancer, Breast Cancer, Pancreatic Cancer, Ovarian Cancer, Prostate Cancer
GAS5	NCRNA00030, SNHG2	1q25.1	60674	Thyroid Cancer, Breast Cancer, Pancreatic Cancer, Ovarian Cancer, Prostate Cancer
SNHG3	NCRNA00014, RNU17C, RNU17D, U17HG, U17HG-A, U17HG-AB	Гр35.3	8420	Breast Cancer, Ovarian Cancer
SNHG4	NCRNA00059, UI9H	5q31.2	724102	Prostate Cancer
SNHG5	C6orf160, LINC00044, NCRNA00044, U50HG	6q14.3	387066	Breast Cancer, Ovarian Cancer
SNHG6	HBII-276HG, NCRNA00058, U87HG	8q13.1	641638	Breast Cancer, Prostate Cancer
SNHG7	NCRNA00061	9q34.3	84973	Thyroid Cancer, Breast Cancer, Pancreatic Cancer, Prostate Cancer
SNHG9	NCRNA00062	16p13.3	735301	Pancreatic Cancer
SNHG12	ASLNC04080, C1orf79, LINC00100, NCRNA00100, PNAS-123	Ip35.3	85028	Thyroid Cancer, Breast Cancer, Ovarian Cancer, Prostate Cancer
DANCR	AGU2, ANCR, KIAA0114, SNHG13, IncRNA-ANCR	4q12	57291	Thyroid Cancer, Breast Cancer, Pancreatic Cancer, Ovarian Cancer, Prostate Cancer
SNHG14	I I 5HG, IC-SNURF-SNRPN, LNCAT, NCRNA002 I 4, U-UBE3A-ATS, UBE3A-AS, UBE3A-AS I, UBE3A-ATS, UBE3AATS	15q11.2	104472715	Breast Cancer, Pancreatic Cancer, Ovarian Cancer, Prostate Cancer
SNHG15	C7orf40, Linc-Myo1g, MYO1GUT	7p13	285958	Thyroid Cancer, Breast Cancer, Pancreatic Cancer, Ovarian Cancer, Prostate Cancer
SNHG16	Nbla10727, Nbla12061, ncRAN	17q25.1	100507246	Thyroid Cancer, Breast Cancer, Pancreatic Cancer, Ovarian Cancer
SNHG17	-	20q11.23	388796	Breast Cancer
SNHG20	C17orf86, LINC00338, NCRNA00338, SCARNA16HG	17q25.2	654434	Breast Cancer, Ovarian Cancer, Prostate Cancer
SNHG22	-	18q21.1	103091864	Ovarian Cancer

regulation", "translation", and "the spliceosome", suggesting further research directions for this lncRNA. ²⁰

SNHG12 is also upregulated (by 3.8-fold) in papillary thyroid carcinoma (PTC) tissues compared to normal adjacent tissue samples.²³ High *SNHG12* was associated with poorer progression in PTC in terms of tumor node metastasis (TNM) staging and lymph node metastasis (LNM).²⁴ *SNHG12* likely acts as a sponge for *miR-16-5p*, thereby inducing PTC cell proliferation, migration, and invasion,

as well as inhibiting apoptosis.²⁵ SNHG12 also promotes the proliferation and migration of PTC cells via the Wnt/β-catenin signaling pathway.²³ Meanwhile, SNHG13, also known as differentiation antagonizing non-protein coding RNA (DANCR), acts as a tumor suppressor in PTC: down-regulation of DANCR is associated with more aggressive clinical features of PTC.²⁶ DANCR is also a potential biomarker for PTC diagnosis, showing a sensitivity of 85.29% and a specificity of 66.18%.²⁶

Table 2 SNHG Members in Endocrine-Related Cancers

SNHG Member	Mechanism	Related Signaling Pathway	Related Clinicopathological Characteristics	Prognostic Significance	Related Cell Biofunctions	Role	In vivo
Thyroid C	ancer						
SNHGI	Positive feedback loop and ceRNA: SPI/SNHG1/miR-199a-5p/SPI		Tumor size		Proliferation, invasion	Oncogene	NO
GAS5	ceRNA: GAS5/miR-222-3p/ PTEN Regulation: GAS5/p-AKT	PI3K/AKT signaling pathway	TNM stage, LNM, multiple cancer foci	DFS, OS	Proliferation	Antioncogene	YES
SNHG7	ceRNA: SNHG7/miR-449a/ ACSL1 Regulation: SNHG7/BDNF	BDNF/TrkB signaling pathway	Tumor size, TNM stage	DFS	Proliferation, migration, cell cycle, apoptosis	Oncogene	NO
SNHG12	ceRNA: SNHG12/miR-16-5p; Regulation: SNHG12/β- catenin, MMP2, Cyclin D1	Wnt/β-catenin signaling pathway	TNM stage, LNM		Proliferation, migration, invasion, apoptosis, cell cycle	Oncogene	YES
DANCR	-	-	T grade, TNM stage		-	Antioncogene	NO
SNHG15	ceRNA: SNHG15/miR-200a- 3p/YAP1, SNHG15/miR-510- 5p; Regulation: SNHG15/β- catenin, E-cadherin, N-cadherin, Vimentin, MST1, LATS1	YAP I -Hippo signaling pathway	Gender, tumor size, TNM stage, LNM, distant metastasis	DFS, OS	Proliferation, migration, invasion, apoptosis, EMT	Oncogene/ Antioncogene	YES
SNHG16	ceRNA: SNHG16/miR-497; Regulation: SNHG16/BDNF		TNM stage, LNM		Proliferation, migration, invasion, apoptosis	Oncogene	NO
Breast Ca	ncer			I	1	l	
SNHGI	ceRNA: SNHG1/miR-382-5p, SNHG1/miR-448/IDO; Regulation: SNHG1/ E-cadherin, N-cadherin, Vimentin, ZEB1		TNM stage	OS	Treg cell differentiation, immune escape, proliferation, migration, invasion, EMT	Oncogene	YES
GAS5	ceRNA: GAS5/miR-196a-5p, GAS5/miR-23a/ATG3, GAS5/ miR-21/PTEN, GAS5/miR- 378a-5p/SUFU, GAS5/miR- 221-3p/DKK2; Regulation: GAS5/β-catenin, c-Myc, Cyclin D1, FOXO1, p-PJ3K, p-AKT, miR-221/GAS5, miR-222/GAS5	PI3K/AKT signaling pathway, Notch signaling pathway, Wnt/ β-Catenin signaling pathway	Tumor size, TNM stage, histological grade, LNM, ER-	OS	Proliferation, invasion, apoptosis, autophagy, cell cycle, Chemosensitivity: DNC, Trastuzumab, Imatinib, PTX, CIS, Adriamycin	Antioncogene	YES
SNHG3	ceRNA: SNHG3/miR-330-5p/ PKM, SNHG3/miR-384/HDGF		Histological grade, LNM, TNM stage, ER, Her-2		Glycolysis metabolism, proliferation, invasion, migration	Oncogene	NO
SNHG5	ceRNA: SNHG5/miR-154-5p/ PCNA; Regulation: SNHG5/ Cyclin D1, p16			OS	Proliferation, apoptosis, cell cycle	Oncogene	YES

Table 2 (Continued).

SNHG Member	Mechanism	Related Signaling Pathway	Related Clinicopathological Characteristics	Prognostic Significance	Related Cell Biofunctions	Role	In vivo
SNHG6	ceRNA: SNHG6/miR-26a/ VASP, SNHG6/miR-26a/MAPK6		Tumor size, TNM stage, distant metastasis		Proliferation, migration, invasion, cell cycle, apoptosis, EMT	Oncogene	YES
SNHG7	Activated by TF and ceRNA: c-Myc/SNHG7/miR-34a-5p/ LDHA; ceRNA: SNHG7/miR- 186, SNHG7/miR-381; Regulation: SNHG7/Ki67, MMP-2, MMP-7, E-cadherin, Vimentin, Snail, Notch-1, Survivin, Cyclin D1	Notch-I signaling pathway	T grade, LNM, distant metastasis	OS	Proliferation, glycolysis metabolism, migration, invasion, EMT	Oncogene	YES
SNHG12	Activated by TF: c-MYC/ SNHG12		Tumor size, LNM		Proliferation, migration, apoptosis	Oncogene	NO
DANCR	ceRNA: DANCR/miR-216a-5p; Methylation: DANCR/EZH2/ SOCS3&CD44&ABCG2 Phosphorylation: DANCR/ RXRA/PIK3CA; Regulation: DANCR/Snail, Slug. MMP-2, MMP-9, E-cadherin, Vimentin, CD133, OCT3/4, NANOG, p-p65, p65, p-STAT3, STAT3, SOX2, ABCG2, ALDH1	PI3K/AKT signaling pathway	TNM stage, histologic grade, LNM	OS	Proliferation, invasion, migration, EMT	Oncogene	YES
SNHG14	ceRNA: SNHG14/miR-193a- 3p; Acetylation: SNHG14/ PABPC1/Nr/2/HO-1; Regulation: SNHG14/c-PARP, c-Caspase-3	Nfr2 signaling pathway	LNM, distant metastasis, cardiac toxicity		Proliferation, invasion, cell cycle, Chemosensitivity: Trastuzumab	Oncogene	YES
SNHG15	ceRNA: SNHG15/miR-411- 5p/VASP, SNHG15/miR-381, SNHG15/miR-211-3p; Regulation: SNHG15/Bcl-2, Bax, VEGF, MMP-2, MMP-9, MMP-14, PCNA, Cyclin D1, c-Caspase-3, Snail, Vimentin, E-Cadherin		Tumor size, TNM stage, LNM	OS	Proliferation, migration, invasion, apoptosis, cell cycle, Chemosensitivity: DDP	Oncogene	YES
SNHG16	ceRNA: SNHG16/miR-30a/ RRM2, SNHG16/miR-98/E2F5, RRM2-let-7a-5p-SNHG16 /MAL2			DFS, OS	Proliferation, invasion, migration	Oncogene	NO
SNHG17	ceRNA: SNHG17/miR-124-3p		TNM stage, LNM	OS	Proliferation, migration, invasion	Oncogene	YES
	ceRNA: SNHG20/miR-495/				Proliferation,	Oncogene	YES

Table 2 (Continued).

SNHG Member	Mechanism	Related Signaling Pathway	Related Clinicopathological Characteristics	Prognostic Significance	Related Cell Biofunctions	Role	In vivo
SNHGI	ceRNA: SNHGI/miR-195/ Cyclin D1; Regulation: SNHGI/p21, Vimentin, E-Cadherin, N-Cadherin, Notch- I, Hes-1, PI3K, p-AKT, t-AKT, BcI-2, Bax	PI3K/AKT signaling pathway, Notch-I signaling pathway	Tumor size, TNM stage	OS	Proliferation, apoptosis, cell cycle, migration, invasion	Oncogene	YES
GAS5	ceRNA: GAS5/miR-221/ SOCS3, GAS5/miR-32-5p/ PTEN, GAS5/181c-5p; Regulation: GAS5/Vimentin, E-Cadherin, N-Cadherin, Snail, OCT4, CD133, Nanog, SOX2, CDK6	PI3K/AKT signaling pathway			Proliferation, migration, invasion, cell cycle, EMT, Chemosensitivity: Gemcitabine	Antioncogene	YES
SNHG7	ceRNA: SNHG7/miR-342-3p/ ID4		Tumor size, LNM, TNM stage, tumor differentiation	OS	Proliferation, migration, invasion	Oncogene	YES
SNHG9	-		N grade, distant metastasis	OS		Antioncogene	NO
DANCR	ceRNA: DANCR/miR-33b/ MMP16, DANCR/miR-135a/ NLRP37, DANCR/miR-214-5p/ E2F2, DANCR/miR-33a-5p/ AXL; Regulation: DANCR/ E-Cadherin, N-Cadherin, NLRP3		Tumor size, T grade, N grade, TNM stage, LNM, vascular invasion, recurrence rates	PFS, OS	Proliferation, invasion	Oncogene	YES
SNHG14	ceRNA: SNHG14/miR-101, SNHG14/miR-613/ANXA2; Bind and regulation: SNHG14/EZH2; Regulation: SNHG14/Vimentin, E-Cadherin, RAB5A, ATG4D		LNM	DFS, OS	Proliferation, invasion, apoptotic, EMT, autophagy, Chemosensitivity: Gemcitabine	Oncogene	YES
SNHG15	Methylation: SNHG15/EZH2/ P15	&KLF2 Regulation: SNHG15/ CDK2, CDK4, c-Caspase-3, c-Caspase-9		Tumor size, TNM stage, LNM		Proliferation, apoptosis, cell cycle	
SNHG16	Oncogene ceRNA: SNHG16/miR-200a- 3p, SNHG16/miR-195/ SREBP2, SNHG16/miR-218-5p	YES	TNM stage, LNM, distant metastasis, tumor differentiation	OS	Proliferation, migration, invasion	Oncogene	YES
Ovarian C	ancer				•	•	
SNHGI	Regulation: SNHG1/β-catenin, Bax, BcI-2, Caspase-9, c-Caspase-9, PARP, Vimentin, E-Cadherin, N-Cadherin, MMP- 2, MMP-9, Lamin A, Cyclin D1, c-myc	Wnt/β-catenin signaling pathway	Pathological grade, TNM stage	OS	Proliferation, migration, invasion, apoptosis, EMT	Oncogene	YES

Table 2 (Continued).

SNHG Member	Mechanism	Related Signaling Pathway	Related Clinicopathological Characteristics	Prognostic Significance	Related Cell Biofunctions	Role	In vivo
GAS5	ceRNA: GAS5/miR-196a-5p/ HOXA5, GAS5/miR-21/SPRY2; Bind and regulation: GAS5/ E2F4/PARP1/MAPK; Regulation: GAS5/c-Caspase-3, Caspase-3, c-Caspase-7, Caspase-7, CDK4, CDK6, Cyclin D, ERK1/2, p-ERK, p-JNK, P38MAPK, GDF15, ASC, Cas-1, p-Cas-1, IL-1β, p-IL-1β, IL-18, p-IL-18, APAF1, p21	MAPK signaling pathway	Tumor size, invasive depth, FIGO stage, histological type	DFS, OS	Proliferarion, cell cycle, apoptosis, migration, invasion, Chemosensitivity:	Antioncogene	YES
SNHG3	Regulation: SNHG3/GSK3β, Cyclin D1, CDK1, MMP-9, MMP-3, β-catenin	GSK3\$ \$- catenin signaling pathway	FIGO stage, LNM	OS	Proliferation, invasion	Oncogene	NO
SNHG5	ceRNA: SNHG5/miR-23a		Tumor grade, FIGO stage, LNM	OS	Proliferation, apoptosis, Chemosensitivity: PTX	Antioncogene	YES
SNHG12	ceRNA: SNHG12/miRNA- 129/SOX4			OS	Proliferate, migration	Oncogene	
DANCR	ceRNA: DANCRmiR-145/ VEGF; Regulation: DANCR/ UPF1, IGF2		Tumor stage, accompanied by metastatic loci		Proliferation, invasion, migration, angiogenesis	Oncogene	YES
SNHG14	ceRNA: SNHG14/miR-219a- 5p, SNHG14/miR-125a-5p/ DHX33; Regulation: SNHG14/DGCR8			OS	Proliferation, migration, invasion, cell cycle	Oncogene	NO
SNHG15	-		Cancer type, ascites, FIGO stage	PFS, OS	Proliferation, migration, invasion, Chemosensitivity: DDP	Oncogene	NO
SNHG16	Regulation: SNHG16/p-AKT, AKT, MMP9	PI3K/AKT signaling pathway	Clinical stage, tumor size, LNM, distant metastasis	OS	Proliferation, invasion, migration	Oncogene	NO
SNHG20	Regulation: SNHG201/β-catenin, GSK-3β, p-GSK-3β, cyclin D1, c-myc, E-cadherin, P21, Vimentin	Wnt/β-catenin signaling pathway	Histological grade, LNM	OS	Proliferation, migration, invasion, EMT	Oncogene	NO
SNHG22	ceRNA: SNHG22/miR-2467/ Gal-1		Tumor size, CA125 expression	OS	Chemosensitivity: DDP, PTX	Oncogene	NO
Prostate (Cancer	•			•	•	•
SNHGI	ceRNA: SNHG1/miR-377-3p/ AKT2, SNHG1/miR-199a-3p/ CDK7		Gleason score, T grade	Biochemical RFS, OS	Proliferation, apoptosis, cell cycle	Oncogene	NO

Dovepress

Table 2 (Continued).

SNHG Member	Mechanism	Related Signaling Pathway	Related Clinicopathological Characteristics	Prognostic Significance	Related Cell Biofunctions	Role	In vivo
GAS5	ceRNA: GAS5/miR-21/ PDCD4/PTEN, GAS5/miR- 1284/AKT, GAS5/miR-1284/ HMGB1, GAS5/miR-103; Regulation: GAS5/ ₂ -H2AX, H2AX, p-mTOR, mTOR, S6K1, p-S6K1; SNP: rs55829688, rs14520427	AKT/mTOR signaling pathway	Clinical T stage, pathologic N stage, seminal vesicle invasion, lymphovascular invasion	DFS, OS	Proliferation, migration, invasion, apoptosis, cell cycle, radiosensitivity	Oncogene/ Antioncogene	YES
SNHG4	ceRNA: SPI/SNHG4/miR- 377/ZIC5		Tumor stage, LNM	OS	Proliferation, invasion, migration	Oncogene	NO
SNHG6	-		Gleason score, T grade	DFS		Oncogene	NO
SNHG7	ceRNA: SNHG7/miR-324-3p/ WNT2B, SNHG7miR-503/ Cyclin D1; Regulation: E-cadherin, N-cadherin, CDK4, CDK6		T grade, TNM stage, Gleason score, bone metastasis, pelvic LNM	OS	Proliferation, migration, invasion, cell cycle, EMT	Oncogene	YES
SNHG12	ceRNA: SNHG12/miR-195/ CCNE1, SNHG12/miR-195, SNHG12/miR-133b; Regulation: Bcl-2, Bax, Caspase-3, c-Caspase-3, Caspase-9, c-Caspase-9, LC3, Beclin-1, p62, PTEN, PI3K, p-PI3K, AKT, p-AKT, mTOR, p-mTOR, β-catenin, c-Myc	Wnt/β-catenin signaling pathway	Gleason score, clinical stage, bone metastasis, disease recurrence, serum PSA, LNM, new tumor event after treatment, lymph nodes examined, PSA value, residual tumor	OS	Proliferation, invasion, apoptosis, autophagy, cell cycle	Oncogene	YES
DANCR	ceRNA: DANCR/miR-135a, DANCR/miR-34a-5p/JAG1; Methylation: DANCR/EZH2/ TIMP2/3; Regulation: DANCR/ PCNA, Ki-67, c-Caspase-3, Bax, LRP, p-gp, MRP1	Notch signaling pathway			Proliferation, apoptosis, migration, invasion, Chemosensitivity: PTX, Docetaxel	Oncogene	YES
SNHG14	ceRNA: SNHG14/miR-613				Proliferation	Oncogene	YES
SNHG15	ceRNA: SNHG15/miR-338- 3p/FKBP1A; Regulation: E-cadherin, N-cadherin				Proliferation, invasion, migration, EMT	Oncogene	NO
SNHG20	ceRNA: SNHG20/miR-6516- 5p/SCGB2A1				Proliferation, invasion, apoptosis	Oncogene	NO

Abbreviations: LNM, lymph node metastasis; ER-, estrogen receptor-negative; Her-2, human epidermal growth factor receptor 2; FIGO, International Federation of Gynaecology and Obstetrics; DFS, disease-free survival; OS, overall survival; PFS, progression-free survival; RFS, recurrence-free survival; DNC, dendrosomal curcumin; PTX, paclitaxel; DDP, cisplatin.

The role of *SNHG15* in thyroid cancer remains controversial. *SNHG15* is upregulated in human PTC tissues and cell lines compared to controls, and was associated with gender, larger tumor size, LNM, advanced TNM stage, and poorer overall survival (OS).²⁷ Meanwhile, *SNHG15* downregulation attenuated cell proliferation, migration, and epithelial—

mesenchymal transition (EMT) in PTC cells, as well as inducing apoptosis.²⁷ Mechanistically, *SNHG15* acts as a sponge for *miR-200a-3p*, thereby upregulating the Yes-associated protein 1 (*YAP1*) signaling pathway.²⁷ Alternatively, another study showed SNHG15 was downregulated in thyroid cancer tissues and cell lines and suppressed tumor progression,

indicating *SNHG15* may act as a tumor suppressor.²⁸ Moreover, inhibition of *SNHG15* by *miR-510-5p* promoted cell proliferation, migration, and invasion in thyroid cancer.²⁹ These diverse functions of *SNHG15* found in different studies may reflect the different subtypes of thyroid cancer; however, further research is required.

Finally, *SNHG16*, which functions as an endogenous sponge for *miR-497*, was upregulated in both PTC tissues and cell lines and shown to induce proliferation, migration, and invasion of thyroid cancer cells, while inhibiting apoptosis.³⁰ High expression of *SNHG16* was also positively associated with advanced TNM stage and LNM.³⁰

In summary, SNHG1, GAS5, SNHG7, SNHG12, DANCR, SNHG15, and SNHG16 all appear to play essential roles in thyroid cancer; although the function of SNHG15 requires further confirmation.

Breast Cancer

Breast cancer is the most commonly diagnosed cancer worldwide and the leading cause of cancer-related death for women. Although advances in early detection and cancer therapeutics have led to a decrease in mortality rates, breast cancer remains a significant public health concern. Some classes of breast cancer, such as triplenegative breast cancer (characterized by a lack of expression of the progesterone receptor, estrogen receptor, and Her-2), have a poor prognosis. Many lncRNAs have been implicated in breast cancer development in recent years, which may eventually lead to better outcomes for these patients. 33

The downregulation of *SNHG1* can suppress the proliferation and invasion of breast cancer cells by regulating *miR-382*.³⁴ In addition, *SNHG1* may inhibit the differentiation of regulatory T cells, promote *miR-448* expression, and reduce indoleamine 2,3 dioxygenase (*IDO*) levels in breast cancer.³⁵ Therefore, *SNHG1* may be a useful target in breast cancer treatment.

GAS5 was first reported to be a tumor suppressor in breast cancer in 2009.³⁶ Since then, studies have shown low GAS5 expression is closely related to a more aggressive tumor phenotype, enhanced proliferation, and attenuated apoptosis in breast cancer cells.^{37–39} GAS5 can bind to miR-196a-5p, thereby partially alleviating its tumor-promoting effects, including invasion and downstream forkhead box O1 (FOXO1)/phosphatidylinositol 3-kinase (PI3K)/AKT signal pathway activation.³⁷ Notch-1 also promotes breast cancer cell proliferation by downregulating GAS5.⁴⁰ GAS5 can also act as a sponge for miR-23a to promote autophagy via the

GAS5-miR-23a-ATG3 axis in breast cancer.³⁸ Moreover, in drug-resistant breast cancer cells, GAS5 overexpression increases chemosensitivity (eg to trastuzumab, imatinib, paclitaxel, cisplatin, among others), especially in triplenegative breast cancer cells.^{39,41-46} Another study showed miR-221/222 suppresses GAS5 expression and enhances tumor growth in a mouse model of breast cancer xenografts.⁴⁷ Moreover, lower plasma GAS5 levels were found in patients with a high Ki67 proliferation index before surgery and in those with LNM after surgery.⁴⁸ Finally, bioinformatics analysis showed GAS5 plays a role in "proliferation" and the "cell cycle", although the molecular mechanisms related to these regulatory pathways are unclear.⁴⁹

There is evidence that IncRNA secreted in exosomes from cancer cells can regulate gene expression and signaling pathways in other niche cells. For example, breast cancer-derived cancer-associated fibroblasts can secrete increased amounts of *SNHG3* than healthy breast tissue cells, which in turn promotes the growth of breast cancer cells by regulating *miR-330-5p*/Pyruvate Kinase M1/M2 (*PKM*). SNHG3 can also act as a sponge for *miR-384*/hepatoma-derived growth factor (*HDGF*) to drive breast cancer cell proliferation, migration, and invasion. ⁵¹

SNHG5 is an oncogene and acts as a sponge for miR-154-5p, reducing its ability to repress proliferating cell nuclear antigen (PCNA), thus promoting breast cancer proliferation, cell cycle progression, and inhibiting apoptosis. 52 SNHG6 was also found to be highly expressed in breast cancer tissues and cell lines, and is associated with poorer clinicopathologic features. 53 Indeed, SNHG6 knockdown inhibits breast cancer cell proliferation, migration, invasion, and G1 cell cycle arrest by acting as a sponge for miR-26a-5p, which regulates expression of the vasodilator-stimulated phosphoprotein $(VASP)^{54}$ and mitogen-activated protein kinase 6 (MAPK6). 55

The expression of *SNHG7* is also upregulated in breast cancer tissues and cells compared to healthy tissues, with high *SNHG7* expression strongly related to tumor stage, distant metastasis, LNM, and OS. ^{56–58} Knocking down *SNHG7* inhibited breast cancer cell proliferation, invasion, and EMT. ^{56–58} Further mechanistic studies revealed *SNHG7* could act as a sponge to repress *miR-34a*, ⁵⁷ *miR-186*, ⁵⁸ and *miR-381*, ⁵⁶ thereby activating the *Notch-1* pathway and glycolysis in breast cancer. Additionally, *c-Myc* (a TF) can bind to the *SNHG7* promoter and positively regulate its expression in breast cancer. ⁵⁹

Increased expression of SNHG12 has been observed in triple-negative breast cancer. 60 SNHG12 upregulation positively correlated with advanced tumor stage and size, and negatively correlated with OS.60 SNHG12 is a direct transcriptional target of c-Myc, and the c-Myc-induced upregulation of SNHG12 enhances the proliferation of breast cancer cells and inhibits apoptosis. 60 SNHG12 may also promote the migration of breast cancer cells by regulating the expression of matrix metalloproteinase 13 $(MMP13).^{60}$

High DANCR levels can lead to shorter OS in triplenegative breast cancer, by acting as a sponge for miR-216a-5p and thereby promoting the proliferation and invasion of tumor cells. 61 DANCR can mediate protein assembly and modification in triple-negative breast cancer. For example, DANCR can bind to the phosphorylation site of retinoid X receptor alpha (RXRA) and suppresses its interaction with the phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha (PIK3CA) promoter. 62 This leads to the activation of the P13K/AKT pathway, which in turn, promotes the proliferation and growth of triple-negative breast cancer cells. 62 DANCR may also participate in the enhancer of zeste homolog 2 (EHZ2)-mediated epigenetic repression of the suppressor of cytokine signaling 3 (SOCS3) in breast cancer cells.63 Sha et al64 proposed DANCR knockdown was associated with increased binding of EZH2 to the promoters of CD44 and ABCG2 (two triplenegative breast cancer stem cell markers), and the concomitant reduction of expression of these genes decreased cancer cell proliferation and invasion. Furthermore, nanoparticle-mediated RNAi of DANCR was shown to be an effective therapy for triple-negative breast cancer.65

Upregulation of SNHG14 in breast cancer tissues may also promote cancer cell proliferation and invasion. 66 In particular, SNHG14 upregulates polyadenylate-binding protein 1 (PABPCI) expression by modulating H3K27 acetylation (H3K27ac) in the promoter of PABPC1 gene, resulting in the activation of the nuclear factor E2-related factor 2 (NRF2) signaling pathway, which is involved in cell defense and survival against chemotherapy drugs.⁶⁶ Besides histone methylation, acetylation is another important form of histone modification.

Indeed, exosomal SNHG14 was upregulated in trastuzumab-resistant human epidermal growth factor receptor 2 (HER2) breast cancer cells compared with parental breast cancer cells, and SNHG14 knockdown re-sensitized breast cancer cells to trastuzumab treatment. 67 These results indicate SNHG14 may be a promising therapeutic target for patients with HER2+ breast cancer. In addition, SNHG14 may enhance breast cancer cell proliferation and invasion by acting as a sponge for miR-193a-3p.68

SNHG15 has also been shown to be highly expressed in breast cancer tissues and cell lines and is positively associated with larger tumor size, LNM, advanced TMN stage, and worse survival. 69,70 SNHG15 primarily acts as a sponge for $miR-411-5p^{69}$ and $miR-211-3p^{70}$ leading to the proliferation, migration, and invasion of breast cancer cells. Additionally, SNHG15 knockdown enhances the cisplatin sensitivity of breast cancer cells by acting as a sponge for miR-381.⁷¹ Moreover, bioinformatics analysis showed SNHG16 might be associated with the prognosis of breast cancer. 72,73 In particular, SNHG16 may interact with miR-30a to regulate the expression of ribonucleosidediphosphate reductase subunit M2 (RRM2)⁷⁴ and competitively bind miR-98 and the E2F Transcription Factor 5 $(E2F5)^{75}$ to promote the proliferation and invasion of breast cancer cells. Finally, SNHG17⁷⁶ and SNHG20⁷⁷ may also drive breast cancer progression by sponging miR-124-3p and miR-495, respectively.

In general, multiple SNHGs, including SNHG1, GAS5, SNHG3, SNHG5, SNHG6, SNHG7, SNHG12, DANCR, SNHG14, SNHG15, SNHG16 and SNHG20, play a role in breast cancer. Targeting SNHGs, especially the treatment of drug-resistant breast cancer, is the future research direction.

Pancreatic Cancer

Pancreatic cancer is one of the most devastating human tumors, with high invasiveness, early metastasis, lack of specific symptoms, and high mortality. According to the most recent statistical data, the 5-year survival of pancreatic cancer is 9%, which is the lowest among all types of cancers and continues to increase (by 0.3% per year) in men. 78 The high fatality rate in pancreatic cancer is attributed to late diagnosis and resistance to current therapies. Recent studies demonstrate lncRNAs are critical in the pathogenesis of pancreatic cancer and are therefore potential biomarkers or drug targets.⁷⁹

SNHG1 acts as an oncogene in pancreatic cancer and accelerates cancer cell growth. 80 In addition, SNHG1 overexpression can promote cyclin D1-mediated pancreatic cancer proliferation by regulating the cell cycle.81 Meanwhile, SNHG1 downregulation inhibits the proliferation, migration, and invasion of pancreatic cancer cells by

suppressing the *Notch-1* signaling pathway.⁸⁰ Similarly, *SNHG1* downregulation inhibits the *PI3K/AKT* signaling pathway in pancreatic ductal adenocarcinoma (PDAC).⁸²

By acting as a sponge for *miR-32-5p*, *GAS5* can promote the expression of *PTEN* and stop the activation of the *PI3K/AKT* signaling pathway, thereby inhibiting pancreatic cancer cell proliferation and survival. ⁸³ *GAS5* also inhibits the expression of the oncogene cyclin-dependent kinase 6 (*CDK6*), although the underlying mechanisms have not been determined. ⁸⁴ Studies also show *GAS5* reduces the chemoresistance of pancreatic cancer cells by downregulating *miR-181c-5p* and *miR-221*. ^{85,86}

SNHG7 is highly expressed in pancreatic cancer tissues and positively correlates with reduced OS. Meanwhile, *SNHG7* knockdown suppresses cell proliferation, migration, and invasion of pancreatic cancer cells by modulating the *miR-342-3p*/inhibitor of DNA binding 4 (*ID4*) axis. ⁸⁷ Zhang et al⁸⁸ showed low expression of *SNHG9* in pancreatic cancer tissues and serums, while those with high *SNHG9* expression had significantly higher survival rates. This data indicates *SNHG9* may be a novel prognostic marker for pancreatic cancer.

High *DANCR* expression correlates with vascular invasion, advanced T grade, LNM, and advanced TNM stage, and is an independent risk factor for poor OS and progression-free survival (PFS) in PDAC.^{89,90} Mechanistically, *DANCR* acts as an miRNA sponge, affecting the *miRNA-33a-5p*/Anexelekto (*AXL*) axis, ⁸⁹ the *miRNA-33b/MMP16* axis, ⁹¹ the *miR-135a/NLRP3* axis, ⁹² and the *miR-214-5p/E2F2* axis ⁹⁰ to promote cell proliferation, migration, invasion, and metastasis in pancreatic cancer.

The *SNHG14* oncogene also potentiates pancreatic cancer cell proliferation through modulation of annexin A2 (*ANXA2*) expression by acting as a ceRNA for *miR-613*. ⁹³ It also acts as a sponge for *miR-10*, thereby enhancing autophagy, which underlies the chemoresistance of PDAC cells to gemcitabine. ^{94,95} Finally, *SNHG15* and *SNHG16* are upregulated in pancreatic cancer samples and are associated with progression in pancreatic cancer patients. ^{96,97} *SNHG15* may help repress *P15* and *KLF2* expression, ⁹⁶ while *SNHG16* promotes cell proliferation, migration, and invasion of pancreatic cancer by sponging *miR-200a-3p* ⁹⁸ and *miR-218-5p*. ⁹⁷ *SNHG16* may also promote pancreatic cancer lipogenesis by directly regulating the *miR-195/SREBP2* axis. ⁹⁹

In short, many SNHGs have a significant predictive effect on the survival of pancreatic cancer patients, and

can be used as a clinical prognostic marker in pancreatic cancer.

Ovarian Carcinoma

Ovarian cancer is the most lethal gynecological cancer in women globally. Despite recent improvements in cytor-eductive surgery and chemotherapy, the 5-year survival rate of ovarian cancer is still approximately 40–50% owing to its late diagnosis and the development of chemoresistance. Therefore, understanding the molecular mechanisms of ovarian carcinogenesis may help improve diagnosis, therapy, and prevention.

Expression of SNHG1 is increased in human epithelial ovarian cancer tissues and cell lines compared to normal healthy tissues, and promotes the proliferation and invasion of ovarian carcinoma cells through the regulation of EMT and the Wnt/β-catenin pathway. 101,102 Meanwhile, GAS5 acts as a tumor suppressor and is expressed in low levels epithelial ovarian cancer samples. 103,104 Indeed. GAS5 expression correlates with prognosis in epithelial ovarian cancer, including International Federation of Gynecology and Obstetrics (FIGO) stage, histological type, OS, and disease-free survival (DFS). 103,104 In terms of its mechanism of action, GAS5 may block CCAAT/ enhancer-binding protein beta (CEBPB)-mediated transcription of the growth/differentiation factor 15 (GD15), leading to decreased viability and increased apoptosis of ovarian cancer cells. 105 GAS5 may also suppress the proliferation of ovarian cancer cells by sponging miR-21106 and miR-196a-5p, 107 which regulate sprouty homolog 2 (SPRY2) and homeobox A5 (HOXA5) expression, respectively. GAS5 is also implicated in inflammasome formation and pyroptosis, but the underlying mechanism is unclear. 108 Finally, GAS5 has been linked to chemoresistance; in particular, GAS5 overexpression control the expression of poly(ADP-ribose) polymerase 1 (PARPI) by recruiting the transcription factor *E2F4* to its promoter, which subsequently affects the mitogen-activated protein kinase (MAPK) pathway activity, further enhance the cisplatin sensitivity of ovarian cancer cells. 109

Upregulation of *SNHG3* expression is associated with poor prognosis in ovarian cancer (including FIGO stage and LNM) and promotes proliferation and invasion by activating the *GSK3β/-catenin* signaling pathway.¹¹⁰ Bioinformatics analysis has shown *SNHG3* is related to energy metabolism in the "glycolysis", "Kreb's cycle", and "oxidative phosphorylation" pathways, and to "drug resistance".¹¹¹ Similarly, *SNHG5* has been implicated in

OncoTargets and Therapy 2020:13 submit your manuscript | www.dovepress.com 7709

chemoresistance: paclitaxel-resistant ovarian cancer tissues and cell lines have lower levels of SNHG5, while SNHG5 overexpression can enhance paclitaxel sensitivity (likely by sponging miR-23a). 112

SNHG12 is also upregulated in ovarian cancer tissues and enhances the proliferative and migratory capacity of cells via sponging miR-129 and upregulating expression of SOX4 (a TF). 113 In addition, DANCR levels are higher in ovarian cancer patients with worse tumor stage and accompanied by metastatic loci. 114 DANCR binds directly to miR-145 and regulates vascular endothelial growth factor (VEGF) expression. 115 Indeed, knockdown of DANCR impairs ovarian tumor growth by inhibiting tumor angiogenesis. 115 In addition, DANCR may enhance the proliferation, migration, and invasion capacities of ovarian cancer cells by upregulating expression of the insulin-like growth factor 2 (IGF2)¹¹⁶ and downregulating UPF1 RNA Helicase And ATPase (UPFI) expression. 114

Like SNHG12, SNHG14 is highly expressed in ovarian cancer tissues and associated with poorer OS. 117,118 SNHG14 may promote ovarian cancer cell progression by sponging $miR-125a-5p^{117}$ and $miR-219a-5p^{118}$ or directly regulating the expression of DiGeorge syndrome chromosomal region 8 (DGCR8). 119 SNHG15 and SNHG16 may also serve as oncogenes in epithelial ovarian cancer. SNHG16 has been shown to promote the proliferation, invasion, and migration of cancer cells via activation of the PI3K/AKT signaling pathway, 120 while the role of SNHG15 is unclear. 121 SNHG20 is also upregulated in ovarian cancer and is associated with shorter OS. 122 SNHG20 knockdown suppresses Wnt/β-catenin signaling activity and EMT-associated gene expression, thereby inhibiting ovarian cancer cell proliferation, migration, and invasion. 123 Finally, the SNHG22 oncogene may regulate the miR-2467/Gal-1 axis to promote cisplatin- and paclitaxel-resistance of ovarian cancer cells. 124

In a word, compared with other SNHGs, GAS5 regulates the progression of ovarian cancer through various mechanisms, indicating its key role in the development of ovarian cancer.

Prostate Cancer

Prostate cancer is the most common malignancy in males and accounts for 10% of cancer-related deaths.⁷⁸ Androgen deprivation therapy (ADT) is the standard treatment for patients with biochemical recurrence after primary treatment, or with locally-advanced or metastatic disease. However, the majority of cancers will eventually

acquire ADT resistance and progress to castration-resistant prostate cancer (CRPC). 125 Aberrantly expressed lncRNAs can be indicative of certain stages of prostate cancer progression, and may predict early progression or efficiently sustain tumor-related signaling pathways. Thus, lncRNAs may be applicable for the diagnosis of prostate cancer, as well as being potential criteria in the choice of therapy and new therapeutic targets of CRPC. 126

SNHG1 upregulation in prostate cancer correlates with the Gleason score, T stage, and a short biochemical recurrence-free survival time. 127 SNHG1 may promote prostate cancer cell proliferation by regulating the miR-199a-3p/ CDK7 axis¹²⁸ and the miR-377-3p/AKT2 axis.¹²⁹ Conversely, GAS5 levels are reduced in prostate cancer tissues and cell lines. 130-132 Low GAS5 levels are associated with prostate-specific antigen level, Gleason score, and pathological stage. 130-132

Most studies indicate that GAS5 inhibits the proliferation, migration, and invasion of prostate cancer cells, and promotes apoptosis. 130-132 In terms of its mechanism of action, GAS5 may act as a sponge for miR-103, which in turn, inactivates the AKT/mTOR signaling pathway, thus inhibiting prostate cancer cell proliferation. 131 In addition, two single nucleotide polymorphisms (SNPs) located in the chromosomal segment of GAS5 (rs55829688 and rs145204276) can increase GAS5 expression, 133-135 and are associated with improved survival in prostate cancer. 133 Patients with prostate cancer and the GAS5 rs145204276 polymorphism are associated with a low risk of pathologic N stage and seminal vesicle invasion. 135 Furthermore, patients with prostate cancer aged >65 years who carry the GAS5 rs145204276 polymorphism show decreased risk of clinical T stage, pathologic N stage, and lymphovascular invasion. 135 Differential expression of GAS5 due to these SNPs likely affects the miR-21/programmed cell death 4 (PDCD4)/PTEN axis, 133 as well as the miR-1284/AKT¹³³ and miR-1284/high mobility group box 1 (HMGB1)¹³⁴ pathways. In addition, overexpression of miR-145 can upregulate GAS5 expression, although GAS5 overexpression (or silencing) has no effect on miR-145 levels. 132

Enhancing GAS5 expression may be particularly useful in androgen-sensitive prostate cancers. 136 Indeed, mTOR inhibitors enhance GAS5 transcript levels in androgensensitive prostate cancer cell lines but have no effect on androgen-independent cell lines (which exhibit low endogenous levels of GAS5). 136 As further evidence of its tumor-suppressing role, GAS5 is implicated in improving the radiosensitivity of prostate cancer cells. In particular,

GAS5 can enhance the α -Solanine-induced radiosensitivity of prostate cancer cells by negatively regulating miR-18a. 137

Despite available evidence showing that *GAS5* acts as a tumor suppressor, some studies report *GAS5* may exist as an oncogene in prostate cancer. For example, Zhang and Chen et al. ^{138,139} found *GAS5* expression was higher in prostate cancer tissues than normal healthy tissues in both public databases and human tissue samples. In addition, functional analysis showed *GAS5* knockdown inhibited the proliferation and cell cycle progression of prostate cancer cells, while promoting apoptosis. ¹³⁸ A bioinformatics analysis also showed high expression of *GAS5* correlated with poorer DFS in prostate cancer, and other studies show *GAS5* may be involved in regulating translational elongation, protein biosynthesis, transcription, protein translation, and proliferation. ^{138–140}

SP1-mediated upregulation of SNHG4 can facilitate prostate cancer progression via the miR-377/zic family

member 5 (ZIC5) axis. 141 Similarly, SNHG6 overexpression was associated with shorter DFS in the Cancer Genome Atlas (TCGA) and Taylor datasets, with bioinformatics analysis revealing SNHG6 is associated with "translation", "nuclear-transcribed mRNA catabolic processes", "ribosomal RNA processing", and "mRNA splicing". 142 SNHG7 is also significantly upregulated in prostate cancer tissue and cell lines, 143,144 and correlates with the Gleason score, bone metastasis, pelvic LNM, TNM stage, and OS. 145 In terms of its mechanism of action, SNHG7 knockdown was found to inhibit proliferation and promote CCND1-induced cell cycle arrest at the G0/G1 phase. 144 SNHG7 can also promote EMT via regulating miR-324-3p and WNT2B, an important protein in the Wnt signaling pathway. 143 Therefore, targeting the SNHG7/miR-324-p/WNT2B axis may represent a novel therapeutic strategy for prostate cancer treatment.

As *SNHG12* acts as an oncogene, it may be a useful predictor of poor prognosis in prostate cancer. Indeed,

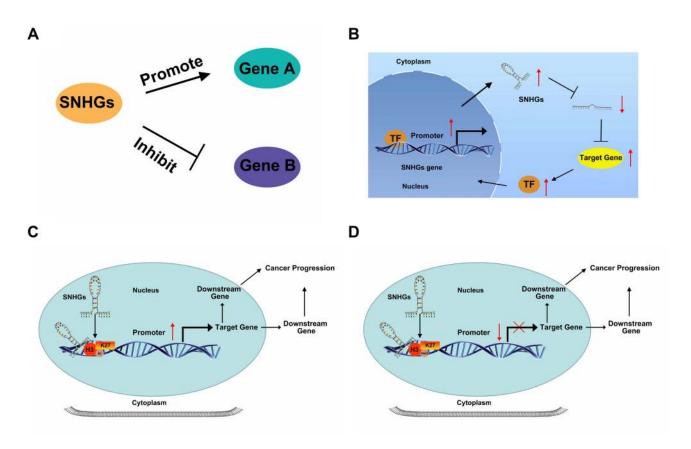


Figure 2 Schematic diagram of the functional mechanism of SNHGs. (A) SNHGs can promote or inhibit expression of downstream target genes. (B) Transcription factors (TF) bind to the promoter and activate transcription of SNHGs. SNHGs can then act as competing endogenous RNA sponges to regulate transcription of downstream target genes (ie TF), forming a positive feedback loop. SNHGs regulate promoter methylation (C) or acetylation (D) of downstream target genes and regulate tumor progression.

a study showed SNHG12 acts as a sponge for miR-195 and can activate the Wnt/β-catenin signaling pathway. ¹⁴⁶ SNHG12 can also promote cell viability and inhibit apoptosis and autophagy of prostate cancer cells via regulating the expression of the G1/S-specific cyclin-E1 (CCNE1) by sponging miR-195.¹⁴⁷ Bioinformatic analysis revealed higher expression of SNHG12 was enriched in the "P53 signaling pathway", "cell cycle", "regulation of cell migration", "cellular metabolic process", "gene expression", and "Notch signaling pathway", and that SNHG12 may target miR-133b. 148

The oncogene DANCR has also been shown to promote the invasion and migration of prostate cancer cells in vitro and the metastasis of tumor xenografts in nude mice. 149 Mechanistically, DANCR works synergistically with EZH2 to downregulate the expression of the tissue inhibitor of metalloproteinases (TIMP) 2/3. 149 Furthermore, downregulation of DANCR can increase the paclitaxel sensitivity of prostate cancer cells by negatively regulating the expression of miR-135a. 150 In addition, stimulation of the DANCR/miR-34a-5p axis enhanced docetaxel-resistance in prostate cancer via targeting JAG1, which in turn activates the Notch signaling pathway.¹³ Finally, SNHG14, ¹⁵¹ SNHG15, ¹⁵² and SNHG20¹⁵³ may all act as oncogenes in prostate cancer via targeting miR-613, miR-338-3p, and miR-6516-5p to promote cell proliferation, migration, and invasion.

In conclusion, SNHGs plays an important role in the process and embody diversified treatment strategies in prostate cancer, especially in CRPC.

Conclusion

This review highlights that the abnormal expression of SNHGs is significantly related to poor prognosis (eg TNM stage, LNM, OS, DFS) and function (eg proliferation, invasion, migration, apoptosis, autophagy, and chemoresistance) in multiple endocrine-related cancers. Some SNHGs played similar roles in different tumors. For example, SNHG1, SNHG3, SNHG4, SNHG6, SNHG7, SNHG12, SNHG14, SNHG16, SNHG17, SNHG20 and SNHG22 promotes tumor growth as oncogenes, while GAS5 and SNHG9 played the role of tumor suppressor genes. In addition, SNHG5, DANCR, SNHG15 played a dual role, which have attracted more scholars' attention. SNHGs could regulate the tumor process via various mechanisms, including direct regulation (promotion or inhibition) (Figure 2A), binding and being activated by TFs, acting as a ceRNA, activating different signaling pathways (Figure 2B), and regulating promoter methylation (Figure 2C) or acetylation of downstream target

genes (Figure 2D). Both methylation and acetylation were histone modifications and their mechanisms were similar. The difference between them was that they bound and modified different histones, and then promoted or inhibited the expression of downstream genes. However, the SNHGs described in this review are only just the tip of the iceberg, and further mechanistic will be required as more SNHG family members are uncovered.

Abbreviations

ACSL1, acyl-CoA synthetase long chain family member 1; ADT, androgen deprivation therapy; AKT, protein kinase B; ANXA2, annexin A2; AXL, Anexelekto; BDNF, brain-derived neurotrophic factor; CCND1/2, cyclin-D1, cyclin-D2; CCNE1, cyclin-E1; CDK6/7, cyclin-dependent kinase 6, cyclin-dependent kinase 7; CDKN2B, cyclin-dependent kinase 4 inhibitor B; CEBPB, CCAAT/enhancer-binding protein beta; CeRNA, competing endogenous RNA; CRPC, castration-resistant prostate cancer; DANCR, Differentiation antagonizing non-protein coding RNA; DFS, disease-free survival; DGCR8, DiGeorge syndrome chromosomal region 8; E2F5, E2F Transcription Factor 5; EHZ2, enhancer of zeste homolog 2; EMT, epithelial-mesenchymal transition; FIGO, International Federation of Gynecology and Obstetrics; FOXO1, forkhead box O1; GAS5, growth arrest specific transcript 5; GD15, growth/differentiation factor 15; HDGF, hepatoma-derived growth factor; HER2, human epidermal growth factor receptor 2; HMGB1, high mobility group box 1; HOXA5, homeobox A5; IDO, indoleamine 2,3 dioxygenase; IGF2, insulin-like growth factor 2; JAG1, Jagged 1; KLF2, Kruppel Like Factor 2; LncRNA, long non-coding RNA; LNM, lymph node metastasis; MAPK6, mitogen-activated protein kinase 6; MMP13, matrix metalloproteinase 13; NRF2, nuclear factor E2-related factor 2; OS, overall survival; PABPC1, polyadenylate-binding protein 1; PARP1, poly (ADP-ribose) polymerase 1; PCNA, proliferating cell nuclear antigen; PDAC, pancreatic ductal adenocarcinoma; PDCD4, programmed cell death 4; PFS, progression-free survival; PI3K, phosphatidylinositol 3-kinase; PIK3CA, phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha; PKM, pyruvate Kinase M1/M2; PTC, papillary thyroid carcinoma; PTEN, phosphatase and tensin homolog; RRM2, ribonucleoside-diphosphate reductase subunit M2; RXRA, retinoid X receptor alpha; SNHG, small nucleolar RNA host genes; SnoRNA, small nucleolar RNA; SNP, single nucleotide polymorphisms; SOCS3, suppressor of cytokine signaling 3; SPRY2, sprouty homolog 2; TCGA, the Cancer Genome Atlas; TF, transcription Factor; TIMP, tissue inhibitor of

metalloproteinases; TNM, tumor node metastasis; *UPF1*, *UPF1* RNA Helicase And ATPase; *VASP*, vasodilatorstimulated phosphoprotein; *VEGF*, Vascular endothelial growth factor; *YAP1*, Yes-associated protein 1; *ZIC5*, zic family member 5.

Data Sharing Statement

The data supporting the findings of this study are available from the corresponding author upon reasonable request.

Funding

This project was funded by the National Natural Science Foundation of China (grant number 81902726), the China Postdoctoral Science Foundation (grant number 2018M641739), and the Natural Science Foundation of Liaoning Province (grant number 20180530090).

Disclosure

The authors report no conflicts of interest for this work.

References

- Qin Y, Sun W, Zhang H, et al. LncRNA GAS8-AS1 inhibits cell proliferation through ATG5-mediated autophagy in papillary thyroid cancer. *Endocrine*. 2018;59(3):555–564. doi:10.1007/s12020-017-1520-1
- Tragante V, Moore JH, Asselbergs FW. The ENCODE project and perspectives on pathways. *Genet Epidemiol*. 2014;38 (4):275–280. doi:10.1002/gepi.21802
- Klinge CM. Non-coding RNAs: long non-coding RNAs and microRNAs in endocrine-related cancers. *Endocr Relat Cancer*. 2018;25(4):R259–R282. doi:10.1530/ERC-17-0548
- Peng WX, Koirala P, Mo YY. LncRNA-mediated regulation of cell signaling in cancer. *Oncogene*. 2017;36(41):5661–5667. doi:10.1038/onc.2017.184
- Tang Y, Cheung BB, Atmadibrata B, et al. The regulatory role of long noncoding RNAs in cancer. *Cancer Lett.* 2017;391:12–19. doi:10.1016/j.canlet.2017.01.010
- Rashid F, Shah A, Shan G. Long non-coding RNAs in the cytoplasm. Genomics Proteomics Bioinformatics. 2016;14 (2):73–80. doi:10.1016/j.gpb.2016.03.005
- Lestrade L, Weber MJ. snoRNA-LBME-db, a comprehensive database of human H/ACA and C/D box snoRNAs. *Nucleic Acids Res*. 2006;34(Database issue):D158–62. doi:10.1093/nar/gki002
- Tollervey D, Kiss T. Function and synthesis of small nucleolar RNAs. Curr Opin Cell Biol. 1997;9(3):337–342. doi:10.1016/ S0955-0674(97)80005-1
- Tycowski KT, Shu MD, Steitz JA. A mammalian gene with introns instead of exons generating stable RNA products. *Nature*. 1996;379(6564):464–466. doi:10.1038/379464a0
- Smith CM, Steitz JA. Classification of gas5 as a multi-smallnucleolar-RNA (snoRNA) host gene and a member of the 5'terminal oligopyrimidine gene family reveals common features of snoRNA host genes. *Mol Cell Biol*. 1998;18(12):6897–6909. doi:10.1128/MCB.18.12.6897

 Williams GT, Farzaneh F. Are snoRNAs and snoRNA host genes new players in cancer. *Nat Rev Cancer*. 2012;12(2):84–88. doi:10.1038/nrc3195

- Xu M, Chen X, Lin K, et al. The long noncoding RNA SNHG1 regulates colorectal cancer cell growth through interactions with EZH2 and miR-154-5p. *Mol Cancer*. 2018;17(1):141. doi:10.1186/s12943-018-0894-x
- Ma Y, Fan B, Ren Z, Liu B, Wang Y. Long noncoding RNA DANCR contributes to docetaxel resistance in prostate cancer through targeting the miR-34a-5p/JAG1 pathway. Onco Targets Ther. 2019;12:5485–5497. doi:10.2147/OTT.S197009
- 14. Anik A, Abaci A. Endocrine cancer syndromes: an update. *Minerva Pediatr*. 2014;66(6):533–547.
- Nikiforov YE, Nikiforova MN. Molecular genetics and diagnosis of thyroid cancer. *Nat Rev Endocrinol*. 2011;7(10):569–580. doi:10.1038/nrendo.2011.142
- LiVolsi VA. Papillary thyroid carcinoma: an update. Mod Pathol. 2011;24(Suppl 2):S1–9. doi:10.1038/modpathol.2010.129
- Ding W, Zhao S, Shi Y, Chen S. Positive feedback loop SP1/ SNHG1/miR-199a-5p promotes the malignant properties of thyroid cancer. *Biochem Biophys Res Commun.* 2020;522 (3):724–730. doi:10.1016/j.bbrc.2019.11.075
- Guo LJ, Zhang S, Gao B, et al. Low expression of long non-coding RNA GAS5 is associated with poor prognosis of patients with thyroid cancer. *Exp Mol Pathol*. 2017;102 (3):500–504. doi:10.1016/j.yexmp.2017.05.008
- Zhang XF, Ye Y, Zhao SJ. LncRNA Gas5 acts as a ceRNA to regulate PTEN expression by sponging miR-222-3p in papillary thyroid carcinoma. *Oncotarget*. 2018;9(3):3519–3530. doi:10. 18632/oncotarget.23336
- Chen L, Zhu J, Zhang LJ. Long non-coding RNA small nucleolar RNA host gene 7 is upregulated and promotes cell proliferation in thyroid cancer. *Oncol Lett.* 2019;18(5):4726–4734. doi:10.3892/ ol.2019.10782
- Wang YH, Huo BL, Li C, Ma G, Cao W. Knockdown of long noncoding RNA SNHG7 inhibits the proliferation and promotes apoptosis of thyroid cancer cells by downregulating BDNF. Eur Rev Med Pharmacol Sci. 2019;23(11):4815–4821. doi:10.26355/ eurrev_201906_18067
- Guo L, Lu J, Gao J, Li M, Wang H, Zhan X. The function of SNHG7/miR-449a/ACSL1 axis in thyroid cancer. *J Cell Biochem*. 2020. doi:10.1002/jcb.29569
- Ding S, Qu W, Jiao Y, Zhang J, Zhang C, Dang S. LncRNA SNHG12 promotes the proliferation and metastasis of papillary thyroid carcinoma cells through regulating wnt/β-catenin signaling pathway. *Cancer Biomark*. 2018;22(2):217–226. doi:10.3233/ CBM-170777
- Liu J, Tang X, Lv J, et al. LncRNAs SNHG12 and LINC00152 were associated with progression of patients with papillary thyroid carcinoma. *Future Oncol*. 2019;15(36):4167–4179. doi:10.2217/fon-2019-0016
- Feng X, Dong X, Wu D, Zhao H, Xu C, Li H. Long noncoding RNA small nucleolar RNA host gene 12 promotes papillary thyroid carcinoma cell growth and invasion by targeting miR-16-5p. *Histol Histopathol*. 2020;35(2):217–224. doi:10. 14670/HH-18-155
- Zhang K, Lv J, Peng X, et al. Down-regulation of DANCR acts as a potential biomarker for papillary thyroid cancer diagnosis. *Biosci Rep.* 2019;39(4).
- 27. Wu DM, Wang S, Wen X, et al. LncRNA SNHG15 acts as a ceRNA to regulate YAP1-Hippo signaling pathway by sponging miR-200a-3p in papillary thyroid carcinoma. *Cell Death Dis.* 2018;9(10):947. doi:10.1038/s41419-018-0975-1
- Liu Y, Li J, Li F, Li M, Shao Y, Wu L. SNHG15 functions as a tumor suppressor in thyroid cancer. J Cell Biochem. 2019;120 (4):6120–6126. doi:10.1002/jcb.27899

OncoTargets and Therapy 2020:13 submit your manuscript | www.dovepress.com 7713

Qin et al **Dove**press

29. Liu Y, Li J, Li M, Li F, Shao Y, Wu L. microRNA-510-5p promotes thyroid cancer cell proliferation, migration, and invasion through suppressing SNHG15. J Cell Biochem. 2019.

- 30. Wen Q, Zhao L, Wang T, et al. LncRNA SNHG16 drives proliferation and invasion of papillary thyroid cancer through modulation of miR-497. Onco Targets Ther. 2019;12:699-708. doi:10.2147/OTT.S186923
- 31. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394-424. doi:10.3322/caac.21492
- 32. Bombonati A, Sgroi DC. The molecular pathology of breast cancer progression. J Pathol. 2011;223(2):307-317. doi:10.1002/path.2808
- 33. Lo PK, Wolfson B, Zhou X, Duru N, Gernapudi R, Zhou Q. Noncoding RNAs in breast cancer. Brief Funct Genomics. 2016;15(3):200-221. doi:10.1093/bfgp/elv055
- 34. Zheng S, Li M, Miao K, Xu H. SNHG1 contributes to proliferation and invasion by regulating miR-382 in breast cancer. Cancer Manag Res. 2019;11:5589-5598. doi:10.2147/CMAR.S198624
- 35. Pei X, Wang X, Li H. LncRNA SNHG1 regulates the differentiation of Treg cells and affects the immune escape of breast cancer via regulating miR-448/IDO. Int J Biol Macromol. 2018;118(Pt A):24-30. doi:10.1016/j.ijbiomac.2018.06.033
- 36. Mourtada-Maarabouni M, Pickard MR, Hedge VL, Farzaneh F, Williams GT. GAS5, a non-protein-coding RNA, controls apoptosis and is downregulated in breast cancer. Oncogene. 2009;28 (2):195-208. doi:10.1038/onc.2008.373
- 37. Li S, Zhou J, Wang Z, Wang P, Gao X, Wang Y. Long noncoding RNA GAS5 suppresses triple negative breast cancer progression through inhibition of proliferation and invasion by competitively binding miR-196a-5p. Biomed Pharmacother. 2018;104:451-457. doi:10.1016/j.biopha.2018.05.056
- 38. Gu J, Wang Y, Wang X, et al. Effect of the LncRNA GAS5-MiR-23a-ATG3 axis in regulating autophagy in patients with breast cancer. Cell Physiol Biochem. 2018;48(1):194-207. doi:10.1159/000491718
- 39. Li J, Li L, Yuan H, Huang XW, Xiang T, Dai S. Up-regulated lncRNA GAS5 promotes chemosensitivity and apoptosis of triple-negative breast cancer cells. Cell Cycle. (16):1965–1975. doi:10.1080/15384101.2019.1635870
- 40. Pei J, Wang B. Notch-1 promotes breast cancer cells proliferation by regulating LncRNA GAS5. Int J Clin Exp Med. 2015;8 (8):14464-14471.
- 41. Pickard MR, Williams GT. Regulation of apoptosis by long non-coding RNA GAS5 in breast cancer cells: implications for chemotherapy. Breast Cancer Res Treat. 2014;145(2):359-370. doi:10.1007/s10549-014-2974-y
- 42. Esmatabadi M, Motamedrad M, Sadeghizadeh M. Down-regulation of lncRNA, GAS5 decreases chemotherapeutic effect of dendrosomal curcumin (DNC) in breast cancer cells. Phytomedicine. 2018;42:56-65. doi:10.1016/j.phymed.2018.03.022
- 43. Li W, Zhai L, Wang H, et al. Downregulation of LncRNA GAS5 causes trastuzumab resistance in breast cancer. Oncotarget. 2016;7(19):27778-27786. doi:10.18632/oncotarget.8413
- 44. Gu J, Wang Y, Wang X, et al. Downregulation of lncRNA GAS5 confers tamoxifen resistance by activating miR-222 in breast cancer. Cancer Lett. 2018;434:1-10. doi:10.1016/j.canlet.2018.06.039
- 45. Zheng S, Li M, Miao K, Xu H. lncRNA GAS5-promoted apoptosis in triple-negative breast cancer by targeting miR-378a-5p/ SUFU signaling. J Cell Biochem. 2020;121(3):2225-2235. doi:10.1002/jcb.29445
- 46. Chen Z, Pan T, Jiang D, et al. The lncRNA-GAS5/miR-221-3p/ DKK2 axis modulates ABCB1-mediated adriamycin resistance of breast cancer via the Wnt/β-catenin signaling pathway. Mol Ther Nucleic Acids. 2020;19:1434–1448. doi:10.1016/j.omtn.2020. 01.030

47. Zong Y, Zhang Y, Sun X, Xu T, Cheng X, Qin Y. miR-221/222 promote tumor growth and suppress apoptosis by targeting lncRNA GAS5 in breast cancer. Biosci Rep. 2019;39(1). doi:10.1042/BSR20181859

- 48. Han L, Ma P, Liu SM, Zhou X. Circulating long noncoding RNA GAS5 as a potential biomarker in breast cancer for assessing the surgical effects. Tumour Biol. 2016;37(5):6847–6854. doi:10.1007/s13277-015-4568-7
- 49. Tokgun PE, Tokgun O, Kurt S, Tomatir AG, Akca H. MYCdriven regulation of long non-coding RNA profiles in breast cancer cells. Gene. 2019;714:143955. doi:10.1016/j.gene.2019. 143955
- 50. Li Y, Zhao Z, Liu W, Li X. SNHG3 functions as miRNA sponge to promote breast cancer cells growth through the metabolic reprogramming. Appl Biochem Biotechnol. 2020.
- 51. Ma Q, Qi X, Lin X, Li L, Chen L, Hu W. LncRNA SNHG3 promotes cell proliferation and invasion through the miR-384/ hepatoma-derived growth factor axis in breast cancer. Hum Cell. 2020;33(1):232-242. doi:10.1007/s13577-019-00287-9
- 52. Chi JR, Yu ZH, Liu BW, et al. SNHG5 promotes breast cancer proliferation by sponging the miR-154-5p/PCNA Axis. Mol Ther Nucleic Acids. 2019;17:138–149. doi:10.1016/j. omtn.2019.05.013
- 53. Jafari-Oliayi A, Asadi MH. SNHG6 is upregulated in primary breast cancers and promotes cell cycle progression in breast cancer-derived cell lines. Cell Oncol (Dordr). 2019;42 (2):211-221. doi:10.1007/s13402-019-00422-6
- 54. Li K, Ma YB, Tian YH, et al. Silencing lncRNA SNHG6 suppresses proliferation and invasion of breast cancer cells through miR-26a/VASP axis. Pathol Res Pract. 2019;215(10):152575. doi:10.1016/j.prp.2019.152575
- 55. Lv P, Qiu X, Gu Y, Yang X, Xu X, Yang Y. Long non-coding RNA SNHG6 enhances cell proliferation, migration and invasion by regulating miR-26a-5p/MAPK6 in breast cancer. Biomed Pharmacother. 2019;110:294-301. doi:10.1016/j.biopha.2018.11.
- 56. Gao YT, Zhou YC. Long non-coding RNA (lncRNA) small nucleolar RNA host gene 7 (SNHG7) promotes breast cancer progression by sponging miRNA-381. Eur Rev Med Pharmacol Sci. 2019;23(15):6588–6595. doi:10.26355/eurrev 201908 185
- 57. Sun X, Huang T, Liu Z, Sun M, Luo S. LncRNA SNHG7 contributes to tumorigenesis and progression in breast cancer by interacting with miR-34a through EMT initiation and the notch-1 pathway. Eur J Pharmacol. 2019;856:172407. doi:10.1016/j.ejphar.2019.172407
- 58. Luo X, Song Y, Tang L, Sun DH, Ji DG. LncRNA SNHG7 promotes development of breast cancer by regulating microRNA-186. Eur Rev Med Pharmacol Sci. 2018;22 (22):7788-7797. doi:10.26355/eurrev 201811 16403
- 59. Zhang L, Fu Y, Guo H. c-Myc-induced long non-coding RNA small nucleolar RNA host gene 7 regulates glycolysis in breast cancer. J Breast Cancer. 2019;22(4):533-547. doi:10.4048/ jbc.2019.22.e54
- 60. Wang O, Yang F, Liu Y, et al. C-MYC-induced upregulation of lncRNA SNHG12 regulates cell proliferation, apoptosis and migration in triple-negative breast cancer. Am J Transl Res. 2017;9(2):533-545
- 61. Tao W, Wang C, Zhu B, Zhang G, Pang D. LncRNA DANCR contributes to tumor progression via targetting miR-216a-5p in breast cancer: lncRNA DANCR contributes to tumor progression. Biosci Rep. 2019;39(4). doi:10.1042/BSR20181618
- 62. Tang J, Zhong G, Zhang H, et al. LncRNA DANCR upregulates PI3K/AKT signaling through activating serine phosphorylation of RXRA. Cell Death Dis. 2018;9(12):1167. doi:10.1038/s41419-018-1220-7

 Zhang KJ, Tan XL, Guo L. The long non-coding RNA DANCR regulates the inflammatory phenotype of breast cancer cells and promotes breast cancer progression via EZH2-dependent suppression of SOCS3 transcription. *Mol Oncol*. 2020;14(2):309–328. doi:10.1002/1878-0261.12622

- Sha S, Yuan D, Liu Y, Han B, Zhong N. Targeting long non-coding RNA DANCR inhibits triple negative breast cancer progression. *Biol Open*. 2017;6(9):1310–1316. doi:10.1242/ bio.023135
- 65. Vaidya AM, Sun Z, Ayat N, et al. Systemic delivery of tumor-targeting siRNA nanoparticles against an oncogenic LncRNA facilitates effective triple-negative breast cancer therapy. *Bioconjug Chem.* 2019;30(3):907–919. doi:10.1021/acs. bioconjchem.9b00028
- 66. Dong H, Wang W, Mo S, et al. Long non-coding RNA SNHG14 induces trastuzumab resistance of breast cancer via regulating PABPC1 expression through H3K27 acetylation. *J Cell Mol Med*. 2018;22(10):4935–4947. doi:10.1111/jcmm.13758
- Dong H, Wang W, Chen R, et al. Exosome-mediated transfer of lncRNA-SNHG14 promotes trastuzumab chemoresistance in breast cancer. *Int J Oncol.* 2018;53(3):1013–1026. doi:10.3892/ ijo.2018.4467
- Xie SD, Qin C, Jin LD, et al. Long noncoding RNA SNHG14 promotes breast cancer cell proliferation and invasion via sponging miR-193a-3p. *Eur Rev Med Pharmacol Sci.* 2019;23 (6):2461–2468. doi:10.26355/eurrev_201903_17393
- Liu LB, Jiang ZJ, Jiang XL, Wang S. Up-regulation of SNHG15 facilitates cell proliferation, migration, invasion and suppresses cell apoptosis in breast cancer by regulating miR-411-5p/VASP axis. Eur Rev Med Pharmacol Sci. 2020;24(4):1899–1912. doi:10.26355/eurrev_202002_20368
- Kong Q, Qiu M. Long noncoding RNA SNHG15 promotes human breast cancer proliferation, migration and invasion by sponging miR-211-3p. *Biochem Biophys Res Commun*. 2018;495(2):1594–1600. doi:10.1016/j.bbrc.2017.12.013
- Mi H, Wang X, Wang F, et al. SNHG15 contributes to cisplatin resistance in breast cancer through sponging miR-381. *Onco Targets Ther.* 2020;13:657–666. doi:10.2147/OTT.S223321
- Li J, Gao C, Liu C, et al. Four lncRNAs associated with breast cancer prognosis identified by coexpression network analysis. *J Cell Physiol*. 2019;234(8):14019–14030. doi:10.1002/jcp.28089
- Zhong G, Lou W, Yao M, Du C, Wei H, Fu P. Identification of novel mRNA-miRNA-lncRNA competing endogenous RNA network associated with prognosis of breast cancer. *Epigenomics*. 2019;11(13):1501–1518. doi:10.2217/epi-2019-0209
- Du SM. The SNHG16/miR-30a axis promotes breast cancer cell proliferation and invasion by regulating RRM2. *Neoplasma*. 2020;67(03):567–575. doi:10.4149/neo_2020_190625N550
- Cai C, Huo Q, Wang X, Chen B, Yang Q. SNHG16 contributes to breast cancer cell migration by competitively binding miR-98 with E2F5. *Biochem Biophys Res Commun*. 2017;485 (2):272–278. doi:10.1016/j.bbrc.2017.02.094
- Du Y, Wei N, Hong J, Pan W. Long non-coding RNASNHG17 promotes the progression of breast cancer by sponging miR-124-3p. Cancer Cell Int. 2020;20(1):40. doi:10.1186/s12935-020-1129-v
- Guan YX, Zhang MZ, Chen XZ, Zhang Q, Liu SZ, Zhang YL. Lnc RNA SNHG20 participated in proliferation, invasion, and migration of breast cancer cells via miR-495. *J Cell Biochem*. 2018;119(10):7971–7981. doi:10.1002/jcb.26588
- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2019. CA Cancer J Clin. 2019;69(1):7–34. doi:10.3322/caac.21551
- Saluja A, Maitra A. Pancreatitis and pancreatic cancer. Gastroenterology. 2019;156(7):1937–1940. doi:10.1053/j.gastro. 2019.03.050

 Cui L, Dong Y, Wang X, et al. Downregulation of long noncoding RNA SNHG1 inhibits cell proliferation, metastasis, and invasion by suppressing the notch-1 signaling pathway in pancreatic cancer. *J Cell Biochem*. 2019;120(4):6106–6112. doi:10.1002/jcb.27897

- Li D, Zhang X, Yang Y, et al. Long non-coding RNA SNHG1 promotes cyclin D1-mediated proliferation in pancreatic cancer by acting as a ceRNA of miR-195. *Int J Clin Exp Pathol*. 2019;12 (3):730-739.
- Zhang Y, Zhang R, Luo G, Ai K. Long noncoding RNA SNHG1 promotes cell proliferation through PI3K/AKT signaling pathway in pancreatic ductal adenocarcinoma. *J Cancer*. 2018;9 (15):2713–2722. doi:10.7150/jca.26207
- Gao ZQ, Wang JF, Chen DH, et al. Long non-coding RNA GAS5 suppresses pancreatic cancer metastasis through modulating miR-32-5p/PTEN axis. *Cell Biosci.* 2017;7(1):66. doi:10.1186/s13578-017-0192-0
- 84. Lu X, Fang Y, Wang Z, et al. Downregulation of gas5 increases pancreatic cancer cell proliferation by regulating CDK6. *Cell Tissue Res*. 2013;354(3):891–896. doi:10.1007/s00441-013-1711-x
- 85. Gao Z-Q, Wang J-F, Chen D-H, et al. Long non-coding RNA GAS5 antagonizes the chemoresistance of pancreatic cancer cells through down-regulation of miR-181c-5p. *Biomed Pharmacother*. 2018;97:809–817. doi:10.1016/j.biopha.2017.10.157
- Liu B, Wu S, Ma J, et al. lncRNA GAS5 reverses EMT and tumor stem cell-mediated gemcitabine resistance and metastasis by targeting miR-221/SOCS3 in pancreatic cancer. *Mol Ther Nucleic Acids*. 2018;13:472–482. doi:10.1016/j.omtn.2018.09.026
- 87. Cheng D, Fan J, Ma Y, et al. LncRNA SNHG7 promotes pancreatic cancer proliferation through ID4 by sponging miR-342-3p. *Cell Biosci*. 2019;9(1):28. doi:10.1186/s13578-019-0290-2
- Zhang B, Li C, Sun Z. Long non-coding RNA LINC00346, LINC00578, LINC00673, LINC00671, LINC00261, and SNHG9 are novel prognostic markers for pancreatic cancer. Am J Transl Res. 2018;10(8):2648–2658.
- 89. Chen L, Liu J, Tang T, et al. lncRNA differentiation antagonizing nonprotein coding RNA overexpression accelerates progression and indicates poor prognosis in pancreatic ductal adenocarcinoma. *Onco Targets Ther.* 2018;11:7955–7965. doi:10.2147/OTT.S167065
- Yao Z, Chen Q, Ni Z, et al. Long non-coding RNA differentiation antagonizing nonprotein coding RNA (DANCR) promotes proliferation and invasion of pancreatic cancer by sponging miR-214-5p to regulate E2F2 expression. *Med Sci Monit*. 2019;25:4544–4552. doi:10.12659/MSM.916960
- Luo Y, Wang Q, Teng L, et al. LncRNA DANCR promotes proliferation and metastasis in pancreatic cancer by regulating miRNA-33b. FEBS Open Bio. 2020;10(1):18–27. doi:10.1002/ 2211-5463.12732
- Tang Y, Cao G, Zhao G, Wang C, Qin Q. LncRNA differentiation antagonizing non-protein coding RNA promotes proliferation and invasion through regulating miR-135a/NLRP37 axis in pancreatic cancer. *Invest New Drugs*. 2019.
- Deng PC, Chen WB, Cai HH, et al. LncRNA SNHG14 potentiates pancreatic cancer progression via modulation of annexin A2 expression by acting as a competing endogenous RNA for miR-613. *J Cell Mol Med*. 2019;23(11):7222–7232. doi:10.1111/jcmm.14467
- 94. Zhang X, Zhao P, Wang C, Xin B. SNHG14 enhances gemcitabine resistance by sponging miR-101 to stimulate cell autophagy in pancreatic cancer. *Biochem Biophys Res Commun.* 2019;510 (4):508–514. doi:10.1016/j.bbrc.2019.01.109
- Xie F, Huang Q, Wang C, et al. Downregulation of long noncoding RNA SNHG14 suppresses cell proliferation and invasion by regulating EZH2 in pancreatic ductal adenocarcinoma (PDAC). Cancer Biomark. 2020;27(3):357–364. doi:10.3233/CBM-190908

OncoTargets and Therapy 2020:13 submit your manuscript | www.dovepress.com 7715

Qin et al Dovepress

 Ma Z, Huang H, Wang J, et al. Long non-coding RNA SNHG15 inhibits P15 and KLF2 expression to promote pancreatic cancer proliferation through EZH2-mediated H3K27me3. *Oncotarget*. 2017;8(48):84153–84167. doi:10.18632/oncotarget.20359

- Liu S, Zhang W, Liu K, Liu Y. LncRNA SNHG16 promotes tumor growth of pancreatic cancer by targeting miR-218-5p. *Biomed Pharmacother*. 2019;114:108862. doi:10.1016/j. biopha.2019.108862
- Guo JQ, Yang ZJ, Wang S, Wu ZZ, Yin LL, Wang DC. LncRNA SNHG16 functions as an oncogene by sponging miR-200a-3p in pancreatic cancer. *Eur Rev Med Pharmacol Sci.* 2020;24 (4):1718–1724. doi:10.26355/eurrev_202002_20347
- Yu Y, Dong JT, He B, et al. LncRNA SNHG16 induces the SREBP2 to promote lipogenesis and enhance the progression of pancreatic cancer. *Future Oncol*. 2019;15(33):3831–3844. doi:10.2217/fon-2019-0321
- 100. Prat J. New insights into ovarian cancer pathology. *Ann Oncol.* 2012;23(Suppl 10):x111–7. doi:10.1093/annonc/mds300
- 101. Ge J, Wu XM, Yang XT, Gao JM, Wang F, Ye KF. Role of long non-coding RNA SNHG1 in occurrence and progression of ovarian carcinoma. *Eur Rev Med Pharmacol Sci.* 2018;22 (2):329–335. doi:10.26355/eurrev 201801 14176
- 102. Wang S, Jiang J, Wang Z, Xie Y, Wu X. Long non-coding RNA SNHG1 is an unfavorable prognostic factor and promotes cell proliferation and migration by Wnt/β-catenin pathway in epithelial ovarian cancer. *Int J Clin Exp Pathol*. 2017;10 (9):9284–9292.
- 103. Gao J, Liu M, Zou Y, et al. Long non-coding RNA growth arrest-specific transcript 5 is involved in ovarian cancer cell apoptosis through the mitochondria-mediated apoptosis pathway. Oncol Rep. 2015;34(6):3212–3221. doi:10.3892/ or.2015.4318
- 104. Li J, Huang H, Li Y, Li L, Hou W, You Z. Decreased expression of long non-coding RNA GAS5 promotes cell proliferation, migration and invasion, and indicates a poor prognosis in ovarian cancer. *Oncol Rep.* 2016;36(6):3241–3250. doi:10.3892/ or.2016.5200
- 105. Guo LL, Wang SF. Downregulated long noncoding RNA GAS5 fails to function as decoy of CEBPB, resulting in increased GDF15 expression and rapid ovarian cancer cell proliferation. *Cancer Biother Radiopharm.* 2019;34(8):537–546. doi:10.1089/cbr.2019.2889
- 106. Ma N, Li S, Zhang Q, Wang H, Qin H, Wang S. Long non-coding RNA GAS5 inhibits ovarian cancer cell proliferation via the control of microRNA-21 and SPRY2 expression. *Exp Ther Med*. 2018;16(1):73–82. doi:10.3892/etm.2018.6188
- 107. Zhao H, Yu H, Zheng J, et al. Lowly-expressed lncRNA GAS5 facilitates progression of ovarian cancer through targeting miR-196-5p and thereby regulating HOXA5. *Gynecol Oncol*. 2018;151(2):345–355. doi:10.1016/j.ygyno.2018.08.032
- Li J, Yang C, Li Y, Chen A, Li L, You Z. LncRNA GAS5 suppresses ovarian cancer by inducing inflammasome formation. *Biosci Rep.* 2018;38(2).
- 109. Long X, Song K, Hu H, et al. Long non-coding RNA GAS5 inhibits DDP-resistance and tumor progression of epithelial ovarian cancer via GAS5-E2F4-PARP1-MAPK axis. *J Exp Clin Cancer Res.* 2019;38(1):345. doi:10.1186/s13046-019-1329-2
- Hong L, Chen W, Wu D, Wang Y. Upregulation of SNHG3 expression associated with poor prognosis and enhances malignant progression of ovarian cancer. *Cancer Biomark*. 2018;22 (3):367–374. doi:10.3233/CBM-170710
- 111. Li N, Zhan X, Zhan X. The lncRNA SNHG3 regulates energy metabolism of ovarian cancer by an analysis of mitochondrial proteomes. *Gynecol Oncol.* 2018;150(2):343–354. doi:10.1016/j. ygyno.2018.06.013

 Lin H, Shen L, Lin Q, et al. SNHG5 enhances Paclitaxel sensitivity of ovarian cancer cells through sponging miR-23a. *Biomed Pharmacother*. 2020;123:109711. doi:10.1016/j.biopha.2019. 109711

- 113. Sun D, Fan XH. LncRNA SNHG12 accelerates the progression of ovarian cancer via absorbing miRNA-129 to upregulate SOX4. Eur Rev Med Pharmacol Sci. 2019;23(6):2345–2352. doi:10.26355/eurrev_201903_17378
- 114. Pei CL, Fei KL, Yuan XY, Gong XJ. LncRNA DANCR aggravates the progression of ovarian cancer by downregulating UPF1. Eur Rev Med Pharmacol Sci. 2019;23(24):10657–10663. doi:10.26355/eurrev_201912_19763
- 115. Lin X, Yang F, Qi X, et al. LncRNA DANCR promotes tumor growth and angiogenesis in ovarian cancer through direct targeting of miR-145. *Mol Carcinog*. 2019;58(12):2286–2296. doi:10.1002/mc.23117
- 116. Gao YQ, Cheng HY, Liu KF. Long non-coding RNA DANCR upregulates IGF2 expression and promotes ovarian cancer progression. *Eur Rev Med Pharmacol Sci.* 2019;23 (9):3621–3626. doi:10.26355/eurrev_201905_17785
- Zhao YL, Huang YM. LncSNHG14 promotes ovarian cancer by targeting microRNA-125a-5p. Eur Rev Med Pharmacol Sci. 2019;23(8):3235–3242. doi:10.26355/eurrev 201904 17683
- 118. Li L, Zhang R, Li SJ. Long noncoding RNA SNHG14 promotes ovarian cancer cell proliferation and metastasis via sponging miR-219a-5p. *Eur Rev Med Pharmacol Sci.* 2019;23 (10):4136–4142. doi:10.26355/eurrev_201905_17915
- 119. Zhao JL, Wang CL, Liu YL, Zhang GY. Long noncoding RNA SNHG14 enhances migration and invasion of ovarian cancer by upregulating DGCR8. Eur Rev Med Pharmacol Sci. 2019;23 (23):10226–10233. doi:10.26355/eurrev 201912 19659
- 120. Yang XS, Wang GX, Luo L. Long non-coding RNA SNHG16 promotes cell growth and metastasis in ovarian cancer. Eur Rev Med Pharmacol Sci. 2018;22(3):616–622.
- 121. Qu C, Dai C, Guo Y, Qin R, Liu J. Long noncoding RNA SNHG15 serves as an oncogene and predicts poor prognosis in epithelial ovarian cancer. *Onco Targets Ther.* 2019;12:101–111. doi:10.2147/OTT S182657
- 122. Wang D, Dai J, Hou S, Qian Y. LncRNA SNHG20 predicts a poor prognosis and promotes cell progression in epithelial ovarian cancer. *Biosci Rep.* 2019;39(4).
- 123. He S, Zhao Y, Wang X, et al. Up-regulation of long non-coding RNA SNHG20 promotes ovarian cancer progression via Wnt/β-catenin signaling. *Biosci Rep.* 2018;38(1). doi:10.1042/BSR20170681.
- 124. Zhang PF, Wu J, Luo JH, et al. SNHG22 overexpression indicates poor prognosis and induces chemotherapy resistance via the miR-2467/Gal-1 signaling pathway in epithelial ovarian carcinoma. *Aging (Albany NY)*. 2019;11(19):8204–8216. doi:10. 18632/aging.102313
- Misawa A, Takayama KI, Inoue S. Long non-coding RNAs and prostate cancer. Cancer Sci. 2017;108(11):2107–2114.
- 126. Kirby M, Hirst C, Crawford ED. Characterising the castration-resistant prostate cancer population: a systematic review. *Int J Clin Pract.* 2011;65(11):1180–1192. doi:10.1111/j.1742-1241.2011.02799.x
- 127. Wan X, Huang W, Yang S, et al. Identification of androgen-responsive lncRNAs as diagnostic and prognostic markers for prostate cancer. *Oncotarget*. 2016;7(37):60503–60518. doi:10.18632/oncotarget.11391
- 128. Li J, Zhang Z, Xiong L, et al. SNHG1 lncRNA negatively regulates miR-199a-3p to enhance CDK7 expression and promote cell proliferation in prostate cancer. *Biochem Biophys Res Commun.* 2017;487(1):146–152. doi:10.1016/j. bbrc.2017.03.169

- Xie M, Zhang Z, Cui Y. Long noncoding RNA SNHG1 contributes to the promotion of prostate cancer cells through regulating miR-377-3p/AKT2 axis. *Cancer Biother Radiopharm*. 2020;35 (2):109–119. doi:10.1089/cbr.2019.3177
- 130. Pickard MR, Mourtada-Maarabouni M, Williams GT. Long non-coding RNA GAS5 regulates apoptosis in prostate cancer cell lines. *Biochim Biophys Acta*. 2013;1832(10):1613–1623. doi:10.1016/j.bbadis.2013.05.005
- 131. Xue D, Zhou C, Lu H, Xu R, Xu X, He X. LncRNA GAS5 inhibits proliferation and progression of prostate cancer by targeting miR-103 through AKT/mTOR signaling pathway. *Tumour Biol.* 2016;37(12):16187–16197. doi:10.1007/s13277-016-5429-8
- 132. Xie X, Dai J, Huang X, Fang C, He W. MicroRNA-145 inhibits proliferation and induces apoptosis in human prostate carcinoma by upregulating long non-coding RNA GAS5. *Oncol Lett.* 2019;18(2):1043–1048. doi:10.3892/ol.2019.10419
- 133. Zhu L, Zhu Q, Wen H, Huang X, Zheng G. Mutations in GAS5 affect the transformation from benign prostate proliferation to aggressive prostate cancer by affecting the transcription efficiency of GAS5. *J Cell Physiol.* 2019;234(6):8928–8940.
- 134. Deng ZH, Yu GS, Pan B, et al. Rs145204276 and rs4759314 affect the prognosis of prostate cancer by modulating the GAS5/miR-1284/HMGB1 and HOTAIR/miR-22/HMGB1 signalling pathways. *Artif Cells Nanomed Biotechnol*. 2020;48(1):435–442. doi:10.1080/21691401.2019.1709859
- 135. Lin CY, Wang SS, Yang CK, et al. Impact of GAS5 genetic polymorphism on prostate cancer susceptibility and clinicopathologic characteristics. *Int J Med Sci.* 2019;16(11):1424–1429. doi:10.7150/ijms.38080
- Yacqub-Usman K, Pickard MR, Williams GT. Reciprocal regulation of GAS5 lncRNA levels and mTOR inhibitor action in prostate cancer cells. *Prostate*. 2015;75(7):693–705. doi:10.1002/pros.22952
- 137. Yang J, Hao T, Sun J, Wei P, Zhang H. Long noncoding RNA GAS5 modulates α-Solanine-induced radiosensitivity by negatively regulating miR-18a in human prostate cancer cells. Biomed Pharmacother. 2019;112:108656. doi:10.1016/j. biopha.2019.108656
- Zhang Y, Su X, Kong Z, et al. An androgen reduced transcript of LncRNA GAS5 promoted prostate cancer proliferation. *PLoS One*. 2017;12(8):e0182305. doi:10.1371/journal.pone.0182305
- 139. Chen X, Yang C, Xie S, Cheung E. Long non-coding RNA GAS5 and ZFAS1 are prognostic markers involved in translation targeted by miR-940 in prostate cancer. *Oncotarget*. 2018;9 (1):1048–1062. doi:10.18632/oncotarget.23254
- 140. Romanuik TL, Wang G, Morozova O, Delaney A, Marra MA, Sadar MD. LNCaP atlas: gene expression associated with in vivo progression to castration-recurrent prostate cancer. *BMC Med Genomics*. 2010;3:43. doi:10.1186/1755-8794-3-43
- 141. Wang ZY, Duan Y, Wang P. SP1-mediated upregulation of lncRNA SNHG4 functions as a ceRNA for miR-377 to facilitate prostate cancer progression through regulation of ZIC5. *J Cell Physiol*. 2020;235(4):3916–3927. doi:10.1002/jcp.29285

- 142. Yan Y, Chen Z, Xiao Y, Wang X, Qian K. Long non-coding RNA SNHG6 is upregulated in prostate cancer and predicts poor prognosis. *Mol Biol Rep.* 2019;46(3):2771–2778. doi:10.1007/s11033-019-04723-9
- 143. Han Y, Hu H, Zhou J. Knockdown of LncRNA SNHG7 inhibited epithelial-mesenchymal transition in prostate cancer though miR-324-3p/WNT2B axis in vitro. *Pathol Res Pract*. 2019;215 (10):152537. doi:10.1016/j.prp.2019.152537
- 144. Qi H, Wen B, Wu Q, et al. Long noncoding RNA SNHG7 accelerates prostate cancer proliferation and cycle progression through cyclin D1 by sponging miR-503. *Biomed Pharmacother*. 2018;102:326–332. doi:10.1016/j.biopha.2018. 03.011
- 145. Xia Q, Li J, Yang Z, Zhang D, Tian J, Gu B. Long non-coding RNA small nucleolar RNA host gene 7 expression level in prostate cancer tissues predicts the prognosis of patients with prostate cancer. *Medicine (Baltimore)*. 2020;99(7):e18993. doi:10.1097/ MD.0000000000018993
- 146. Song J, Wu X, Ma R, Miao L, Xiong L, Zhao W. Long noncoding RNA SNHG12 promotes cell proliferation and activates Wnt/βcatenin signaling in prostate cancer through sponging microRNA-195. J Cell Biochem. 2019;120(8):13066–13075. doi:10.1002/ jcb.28578
- 147. Wang X, He C, Yang Z, Li S, Qiao L, Fang L. Dysregulation of long non-coding RNA SNHG12 alters the viability, apoptosis, and autophagy of prostate cancer cells by regulating miR-195/CCNE1 axis. Int J Clin Exp Pathol. 2019;12(4):1272–1283.
- 148. Cheng G, Song Z, Liu Y, et al. Long noncoding RNA SNHG12 indicates the prognosis of prostate cancer and accelerates tumorigenesis via sponging miR-133b. *J Cell Physiol*. 2020;235 (2):1235–1246. doi:10.1002/jcp.29039
- 149. Jia J, Li F, Tang XS, et al. Long noncoding RNA DANCR promotes invasion of prostate cancer through epigenetically silencing expression of TIMP2/3. *Oncotarget*. 2016;7 (25):37868–37881. doi:10.18632/oncotarget.9350
- 150. Zhao HF, Zhang ZC, Shi BK, Jiang XZ. DANCR sponges miR-135a to regulate paclitaxel sensitivity in prostate cancer. Eur Rev Med Pharmacol Sci. 2019;23(16):6849–6857. doi:10.26355/eurrev_201908_18724
- 151. Sun B, Ke KB, Liu DF, et al. Long noncoding RNA SNHG14 acts as an oncogene in prostate cancer via targeting miR-613. Eur Rev Med Pharmacol Sci. 2020;24(2):633–638. doi:10.26355/ eurrev_202001_20039
- 152. Zhang Y, Zhang D, Lv J, Wang S, Zhang Q. LncRNA SNHG15 acts as an oncogene in prostate cancer by regulating miR-338-3p/FKBP1A axis. Gene. 2019;705:44–50. doi:10.1016/j.gene.2019.04.033
- 153. Wu X, Xiao Y, Zhou Y, Zhou Z, Yan W. IncRNA SNHG20 promotes prostate cancer migration and invasion via targeting the miR-6516-5p/SCGB2A1 axis. Am J Transl Res. 2019;11 (8):5162–5169.

OncoTargets and Therapy

Publish your work in this journal

OncoTargets and Therapy is an international, peer-reviewed, open access journal focusing on the pathological basis of all cancers, potential targets for therapy and treatment protocols employed to improve the management of cancer patients. The journal also focuses on the impact of management programs and new therapeutic

agents and protocols on patient perspectives such as quality of life, adherence and satisfaction. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit http://www.dovepress.com/testimonials.php to read real quotes from published authors.

 $\textbf{Submit your manuscript here:} \ \texttt{https://www.dovepress.com/oncotargets-and-therapy-journal}$

Dovepress