Longitudinal analysis of the behavioral phenotype in a novel transgenic rat model of early stages of Alzheimer's disease

Pablo Galeano^{1,2}, Pamela V. Martino Adami¹, Sonia Do Carmo³, Eduardo Blanco⁴, Cecilia Rotondaro¹, Francisco Capani², Eduardo M. Castaño¹, A. Claudio Cuello³ and Laura Morelli¹*

- ¹ Laboratorio de Amiloidosis y Neurodegeneración, Instituto de Investigaciones Bioquímicas de Buenos Aires, Fundación Instituto Leloir, CONICET, Ciudad Autónoma de Buenos Aires, Argentina
- ² Instituto de Investigaciones Cardiológicas "Prof. Dr. Alberto C. Taquini" (ININCA), Universidad de Buenos Aires and CONICET, Ciudad Autónoma de Buenos Aires, Argentina
- ³ Department of Pharmacology and Therapeutics, McGill University, Montreal, QC, Canada
- ⁴ Departament de Pedagogia i Psicologia, Facultatd'Educació, Psicologia i Treball Social, Universitat de Lleida, Lleida, Spain

Edited by:

Johannes Gräff, Ecole Polytechnique Fédérale de Lausanne. Switzerland

Reviewed by:

Deborah Suchecki, Universidade Federal de São Paulo, Brazil Fiona Hollis, Ecole Polytechnique Fédérale de Lausanne, Switzerland Luis Javier Santín, University of Malaga, Spain

*Correspondence:

Laura Morelli, Laboratorio de Amiloidosis y Neurodegeneración, Fundación Instituto Leloir, IIBBA (CONICET), Av. Patricias Argentinas 435, C1405BWE, Ciudad Autónoma de Buenos Aires, Argentina e-mail: Imorelli@leloir.org.ar

Intraneuronal accumulation of amyloid β (iAβ) has been linked to mild cognitive impairment that may precede Alzheimer's disease (AD) onset. This neuropathological trait was recently mimicked in a novel animal model of AD, the hemizygous transgenic McGill-R-Thy1-APP (Tg^{+/-}) rat. The characterization of the behavioral phenotypes in this animal model could provide a baseline of efficacy for earlier therapeutic interventions. The aim of the present study was to undertake a longitudinal study of AB accumulation and a comprehensive behavioral evaluation of this transgenic rat model. We assessed exploratory activity, anxiety-related behaviors, recognition memory, working memory, spatial learning and reference memory at 3, 6, and 12 months of age. In parallel, we measured Aβ by ELISA, Western blots and semiquantitative immunohistochemistry in hippocampal samples. SDS-soluble Aβ peptide accumulated at low levels (~9 pg/mg) without differences among ages. However, Western blots showed SDS-resistant Aβ oligomers (~30 kDa) at 6 and 12 months, but not at 3 months. When compared to wild-type (WT), male $Tq^{+/-}$ rats exhibited a spatial reference memory deficit in the Morris Water Maze (MWM) as early as 3 months of age, which persisted at 6 and 12 months. In addition, $Tg^{+/-}$ rats displayed a working memory impairment in the Y-maze and higher anxiety levels in the Open Field (OF) at 6 and 12 months of age, but not at 3 months. Exploratory activity in the OF was similar to that of WT at all-time points. Spatial learning in the MWM and the recognition memory, as assessed by the Novel Object Recognition Test, were unimpaired at any time point. The data from the present study demonstrate that the hemizygous transgenic McGill-R-Thy1-APP rat has a wide array of behavioral and cognitive impairments from young adulthood to middle-age. The low Aß burden and early emotional and cognitive deficits in this transgenic rat model supports its potential use for drug discovery purposes in early AD.

Keywords: Alzheimer's disease, cognitive impairment, anxiety, amyloid β , transgenic rat models

INTRODUCTION

Alzheimer's disease (AD) is a neurodegenerative disorder that frequently begins with memory complaints progressing to a severe deterioration of multiple memory systems (Eustache et al., 2006). In addition to cognitive impairment, AD patients exhibit several neuropsychiatric symptoms with the progression of the disease (Geda et al., 2013). Some of the classical neuropathological changes in AD are the presence of amyloid β (A β) deposits and neurofibrillary tangles which initiate in the entorhinal cortex and hippocampus, and spread to the medial temporal lobe (Scahill et al., 2002). This progression of neuropathology leads to episodic and semantic memory deficits that can be detected at early stages of the disease (Bondi et al., 1999, 2008; Collie and Maruff, 2000;

Schmitt et al., 2000; Smith et al., 2007). As the disease progresses, other cognitive domains, such as attention and executive function, among others, are affected (Bondi et al., 2008).

Recent reports suggest that intraneuronal accumulation of $A\beta$ (iA β) takes place with the aging of the human brain many years prior to $A\beta$ plaques accumulation and Alzheimer's disease onset (Gouras et al., 2000; D'Andrea et al., 2001). Aging is the predominant risk factor for AD and there is biochemical evidence supporting that the disease has an extensive preclinical phase (Sperling et al., 2011; Bateman et al., 2012). In this context, preventive medicine combined with early diagnosis may allow a substantial proportion of aged people to escape from suffering AD. The relevance of the early stage of AD is further supported

by the failure of recent clinical trials aimed at reducing extracellular A β levels and plaque formation (Laurijssens et al., 2013; Vellas et al., 2013). Some of the reasons proposed for the failure may be the initiation of the trials in demented patients with serious brain damage together with unforeseen design flaws in the studies based on results from inadequate animal models (Laurijssens et al., 2013; Vellas et al., 2013).

To test potential novel drug therapies specific for early stages of AD it is necessary to develop an animal model at a pre-plaque stage with a progressive decline in emotional and cognitive function. The phenotype should be sensitive to iAβ accumulation and aging. Transgenic mouse models of AD have been widely used to model the impairments in some cognitive domains shown in patients with AD, such as semantic, working and recognition memory (Webster et al., 2014). Semantic memory refers to a conscious recollection and retrieval of factual information and in rodents is most closely correlated with reference memory. The latter implies the learning of certain aspects of a task that remain constant throughout the behavioral test (Webster et al., 2014). Working memory is a system that holds information for a limited amount of time, allowing its use and manipulation to guide behaviors (Baddeley, 2003). Recognition memory belongs to the long-term declarative memory system and consists in the ability to recognize individuals, events and objects that were already encountered (Ally, 2012).

Available transgenic mice of AD have a general difficulty in recreating the characteristics of early AD as most of them show an aggressive neuropathological phenotype characterized by a massive plaque formation (Do Carmo and Cuello, 2013). Transgenesis in rats offers great potential to decipher subtle and early aspects of AD pathology. The rat is, in many aspects, closer to humans than mice. Because of its complex CNS and its predictable and multi-faceted behavioral display, this species is of great value for accurate cognitive assessment (Whishaw et al., 2001; Abbott, 2004). Different transgenic rat models of AD have been developed in the last 10 years (Do Carmo and Cuello, 2013) however, only two models, the UKUR25 (Echeverria et al., 2004) and the hemizygous McGill-R-Thy1-APP (Leon et al., 2010) show exclusively iAB accumulation in cortex and hippocampus without amyloid plaques or tangles at later stages. The difference between both models is that McGill-R-Thy1-APP rats show progressive amyloid pathology from postnatal week one while UKUR25 rats from 6 months of age (Echeverria et al., 2004; Leon et al., 2010). In contrast to other transgenic rat models of AD, such as UKUR25 and TgF344-AD in which iAβ accumulation is sufficient to increase p-Tau levels, available bibliographic reports do not indicate that McGill-R-Thy1-APP rat model develops Tau pathology (Do Carmo and Cuello, 2013). This discrepancy can be explained by differences in the genetic background and the transgene expressed.

The McGill-R-Thy1-APP model was previously evaluated in the Morris Water Maze (MWM) test (Leon et al., 2010). MWM is the most widely employed spatial learning and memory task systematically impaired in all of the different strains of AD transgenic mice (Webster et al., 2014). Recently, it was also reported that the hemizygous transgenic McGill-R-Thy1-APP ($Tg^{+/-}$) rat, the least genetically aggressive AD transgenic model developed so

far, also showed impairments in auditory fear conditioning and in the Novel Object Recognition and Location (NORL) task as early as 3 months of age (Iulita et al., 2014). However, it was not indicated if by this age $Tg^{+/-}$ rats accumulate soluble A β peptides as compared to wild-type littermates (Iulita et al., 2014). In addition, working memory and the temporal progression of AD-like non-cognitive behavioral abnormalities, which are affected early during the course of AD, were not determined in this novel transgenic animal model.

Multiple behavioral tasks should ideally be used to characterize the phenotypes of potential animal models of AD since in the clinical setting, a wide range of neuropsychological tests are employed to determine the profile of the cognitive impairment (Gainotti et al., 2014; Webster et al., 2014). In the present study, by using a longitudinal assessment procedure, we sought to determine the levels and accumulation of soluble A β and iA β peptides and the development of impairments in the exploratory activity, emotional behavior and working and spatial reference memory in Tg^{+/-} rats. In this way, we will test if this model recapitulates some of the biochemical, emotional and cognitive alterations seen in pre-symptomatic AD. Our results could be relevant to define in this animal model the critical window for therapeutic interventions that may be applied to early stages of AD.

MATERIALS AND METHODS

ETHICAL STATEMENTS

The study was carried out in strict accordance with the guidelines of the OLAW-NIH (Office Laboratory Animal Welfare). The protocol was approved by the local Animal Care Committee of Fundación Instituto Leloir (FIL) Assurance# A5168-01.

ANIMALS

Hemizygous transgenic McGill-R-Thy1-APP (Tg^{+/-}) rats, harboring the human APP751 transgene with the Swedish and Indiana mutation under the control of the murine Thy1.2 promoter (Leon et al., 2010), were provided to Fundación Instituto Leloir (FIL) by The Royal Institution for the Advancement of Learning/McGill University, Montreal, Quebec (Canada).

HUSBANDRY AND GENOTYPING

An in-house breeding colony was established at FIL by crossbreeding Tg^{+/-} males with wild-type Wistar females rats. Each litter was culled to 10 pups (up to 8 males per litter were left). At weaning (21 post-natal days), dams were removed from the litters and pups were genotyped as described below. When the results of the genotyping were available (between 22 and 23 post-natal days), rats were pseudorandomly assigned to experimental groups. To avoid the litter effect, groups were made up of pups from 3 to 4 different litters. Animals were maintained on a 12-h light/12-h dark cycle, group-housed 2-4 per cage and had ad libitum access to water and a standard rodent diet. Tissue samples were obtained by ear punches from the litters to determine the genotype of each rat. Genotyping was performed using a validated protocol. Briefly, genomic DNA was prepared from ear biopsy tissue using proteinase K digestion, followed by phenol/chloroform/isoamyl alcohol extraction (Life Technologies). DNA quantification was determined

in a Nanodrop 2000 spectrofotometer. Samples with a ratio of absorbance at 260/280 and 260/230 nm above 1.80 were used for PCR to recognize whether or not the rat carried the human APP (hAPP) gene. The following sets of primers were used: APP-F 5'-AGGACTGACCACTCGACCAG-3'; APP-R 5'-CGGGGGTCTAGTTCTGGAT-3'; CyclophilinG-F 5'- TACAA CAGTAGAACAAGGGAGCGAAG-3' and CyclophilinG-R 5'-ATCCCTCCTTCTTCTCCTCCTATCTTT-3'. Cycling conditions were: 95°C (15 min) and 40 cycles of 94°C (30 s), 54°C (1 min), 72°C (1 min) followed by one step at 72°C (2 min) and kept at 4°C until processed. PCR products were resolved by 2% agarose mini-gel electrophoresis in TBE buffer and visualized with ethidium bromide (10 mg/ml) staining under UV light. Cyclophilin G amplicon (248 bp) was used as a housekeeping gene to verify DNA integrity and PCR conditions. The hAPP amplicon (387 bp) indicates the presence of the human transgene. To verify the hemizygocity of the transgenic rats, quantitative PCR (qPCR) was performed by using the same set of hAPP primers as described above and the following set of GAPDH (used as housekeeping gene) primers: GAPDH-F: 5'-GGGGAAGGACGCTGTACGGG-3' and GAPDH-R: 5'-AAGGGGAGC AACAGCTGGGGT-3'. A total of 1 ng of genomic DNA was used per well (5 μl of a 0.2 ng/μl dilution). Samples were plated in triplicate. SYBR-Green qPCR was performed by using the KAPA SYBR® FAST Universal 2X qPCR Master Mix (Kapa Biosystems). Reactions were run in a Stratagene Mx3005P cycler and results analyzed by the MxPro software in a Comparative Quantitation mode. In all of the runs a genomic DNA sample of a hemizygous animal was included and used as an internal calibrator. If the unknown sample is hemizygous, the relative quantity (dRn) value should be between 0.8 and 1.3; if the unknown sample is homozygous the result is close to a value of 2 (between 1.8 and 2.3).

EXPERIMENTAL DESIGN

For time-course analysis of the behavioral phenotype, 3 cohorts of male rats consisting in $Tg^{+/-}$ and wild-type (WT) littermates were used. Rats were divided in 6 experimental groups as follows: group 1 (3 months old $Tg^{+/-}$ rats; n=10); group 2 (3 months old WT rats; n=9); group 3 (6 months old $Tg^{+/-}$ rats; n=9); group 4 (6 months old WT rats; n=7); group 5 (12 months old $Tg^{+/-}$ rats; n=11); group 6 (6 months old WT rats; n=11) (see **Figure 2**). Only male rats were used to avoid any potential effects of changes in the female estrus cycle on behavioral performance.

HIPPOCAMPAL ISOLATION

Three groups (n = 3/group) of naïve Tg^{+/-} male rats of 3, 6, and 12 months of age, and one group (n = 3) of naïve WT male rats of 12 months of age, were anesthetized with ketamine (50 mg/kg) and xylacine (10 mg/kg), perfused transaortically with 0.9% sodium chloride with 200 U/l heparine and brains were quickly removed as described previously (Leal et al., 2006). Brains were divided into left and right hemispheres. Hippocampus was isolated from left hemisphere and retained for biochemical studies. The right hemisphere was post-fixed in 4% paraformaldehyde in 0.1 M phosphate buffer (PB), pH 7.4 overnight at 4°C, and finally transferred to a solution of 30% sucrose in 0.1 M PB for 5 days, or until sectioned for immunohystochemistry into 40 μ m

coronal sections with a freezing sledge microtome (SM 2000R, Leica) at -20° C. None of the animals assigned to immunohistochemical and biochemical experiments were submitted to behavioral testing.

SDS-SOLUBLE AB PEPTIDE QUANTITATION

To estimate the levels of hippocampal SDS-soluble Aβ peptides, 100 mg of hippocampus were homogenized with a manual Teflon glass homogenizer in 1 ml of RIPA buffer (150 mM NaCl, 0.5% sodium deoxycolate, 1% Triton X-100, 2% SDS) containing proteases inhibitors (5 mM EDTA, 5 mM EGTA, 5 µg/ml leupeptin, 10 µg/ml aprotinin, 1 µg/ml pepstatin) and phosphatases inhibitors (50 mM sodium fluoride and 5 mM sodium orthovanadate) and centrifuged at $20,000 \times g$ at 4° C for 45 min using a Eppendorf microcentrifuge. Supernatants were aliquoted and stored at -80°C. Individual samples were analyzed to assess SDS-soluble human A\u00e440 or A\u00e442 levels by using sandwich ELISA tests commercially available (Invitrogen). Results were expressed as pg of Aβ/mg of hippocampal homogenate proteins as determined with a bicinchoninic acid assay (Pierce, Rockford, IL) and were corrected by the signal of WT protein background. Determinations were performed in duplicate samples in three independent experiments. To evaluate SDS-resistant oligomeric Aβ levels, samples (250 µg/lane) were run in SDS-Tricine 12% gels and processed as previously described (Leon et al., 2010). Immunoreactivity was detected by Western blot using an enhanced chemiluminescence detection system (ECL, Thermo Scientific) and membranes analyzed and quantitated with a STORM 840 Phosphor Imager (GE Healthcare). Group values were obtained simultaneously and normalized with respect to APP immunoreactivity.

ESTIMATION OF THE VOLUME FRACTION OF THE HIPPOCAMPUS AFFECTED BY $ia\beta$ deposition

Brain sections were analyzed by using systematic random sampling. Every sixth brain section (240 μm apart) was stained using a free-floating immunohistochemistry procedure as previously described (Leon et al., 2010). iA β label was detected with McSA1 (1:4000) mouse monoclonal antibody and sections were analyzed at 40× magnification by using an Olympus BX50 microscope. Immunoreactivity was assessed with Image J software (NIH, USA) in sections between the coordinates -2.30 and -3.8 mm from bregma for dorsal hippocampus and between -4.16 and -6.04 mm from bregma for ventral hippocampus (Paxinos and Watson, 2009) following a standard protocol. Stereological estimation of the hippocampal volume affected by iA β is described below in *Image analysis* Section.

NEUROLOGICAL SCREENING AND BEHAVIORAL TESTING

One week before starting with the behavioral battery each cohort was transported to ININCA (School of Medicine, Universidad de Buenos Aires) where a behavioral core facility is available. Throughout the study period, animals were housed in a controlled environment (21 \pm 2°C, 50–60% relative humidity, 12 h light/12 h dark schedule) and all efforts were made to minimize discomfort. After acclimation, rats were handled daily (5 min/day) for three consecutive days and subjected to a basic

neurological examination which consisted of a battery of neurological reflex tests: righting response after being placed on the dorsal side; eye blink (response to light touch with a small camel hair brush), ear twitch, and limb withdrawal in response to tactile stimuli (light touch with a gloved finger); orienting response to a visual stimuli (flashlight) and startle response following an auditory stimulus (a metal clicker) as previously reported (Cohen et al., 2013). Responses were scored as being present or absent. The maximum time period in which behavioral tests were carried out was between 8:00 a.m. and 5 p.m. Since there is a major circadian influence on EPM and OF performance (Verma et al., 2010), these tests were conducted between 8:00 a.m. and 12:00 p.m. Moreover, testing order of the groups was counterbalanced to avoid the confounding effect of time of the day at which animals were tested. The order of tests were as follows: Elevated Plus Maze (EPM) test, Open Field (OF) test, Novel Object Recognition Test (NORT), Y-maze spontaneous alternation (Y-Maze) test, and Morris Water Maze (MWM) test. The interval between tests was at least 24 h. After each animal was tested, the entire apparatus and objects in the NORT were cleaned with 70% ethanol to prevent a bias due to olfactory cues. All testing sessions were digitally recorded (JVC Everio GZ-HD620) and analyzed using a computerized video-tracking system (Ethovision XT, version 7, Noldus Information Technology, Wageningen, The Netherlands) or the ethological observation software JWatcher V1.0. All the experimenters were blinded to the genotype of the animals.

Elevated plus maze test

The EPM test assesses anxiety-like behaviors (Pellow et al., 1985). The apparatus consisted of a black melamine central square platform (11 × 11 cm) from which four black melamine arms radiate $(50 \times 11 \text{ cm})$ separated from each other by 90° . Two of the arms ("closed arms") have a wall (40 cm in height) all around its perimeter but not in the entrance and the other two arms ("open arms") do not have any wall but with raised edges (0.25 cm) around its perimeter. The maze was elevated one meter from the floor by five legs. Light intensity in the open arms was 80–90 lux. Each rat was placed onto the central platform facing an open arm and allowed to freely explore the maze for 5 min. An arm entry was counted when rat introduced its four paws into an arm. The following dependent variables were recorded: total distance moved, number of closed arm entries, percentage of open arm entries $\left(\frac{N^{\circ} \text{ open arm entries}}{\text{Total entries}} \times 100\right)$ and percentage of time spent in open arms $\left(\frac{Time\ spent\ in\ open\ arms}{300\ s}\times 100\right)$.

Open field test

The OF test was used to assess spontaneous exploratory activity and anxiety-related behaviors (Walsh and Cummins, 1976). The apparatus consisted of a black melamine square (60×60 cm) surrounded by walls of 40 cm in height. A central area was arbitrarily defined as a square of 30×30 cm and it was drawn over the image of the OF in the video-tracking system. A rat was considered to be into the central area when its four paws were on it. Light intensity in the center of the OF was 70–80 lux. Each rat was placed in the center of the maze and its behavior was analyzed for 5 min. The following dependent variables were recorded: total distance

moved, number of rears, number of entries into the central area and time spent in central area.

Novel object recognition test

The NORT evaluates recognition memory of one previously explored object (familiar) compared with one novel object (nonfamiliar) (Ennaceur and Delacour, 1988; Ennaceur, 2010). NORT was performed in the same OF apparatus 24 h after the OF test. Rats were tested during two trials (sample and retention trials) separated by an interval of 1 h during which they were returned to their cages. In the sample trial (duration: 5 min), each rat was faced with two identical objects (called familiar objects) placed in a symmetrical position and the time exploring freely each object was recorded. In the retention trial (duration: 3 min), one of the two familiar objects was replaced by a novel object and the time exploring each was recorded. Sets composed of three copies of the same object were used to prevent odor cues and all combinations and location of objects were used to prevent bias due to preference for a particular object or location. Exploration time was computed when the snout pointed to the object at a distance <2 cm. Discrimination index (d1) and discrimination ratio (d2) scores were calculated using the following formulas: $d1 = t_n - t_f$ and $d2 = \frac{t_n - t_f}{t_n + t_f}$, where $t_n =$ the amount of time rats explored the novel object and t_f = the amount of time rats explored the familiar object.

Y-maze spontaneous alternation test

The Y-maze test was used to measure spatial working memory as previously described with minor modifications (Sierksma et al., 2014). The apparatus consisted of three identical arms $(45 \times 12 \times 35 \text{ cm})$ diverging at 120° angle one to the other and an equilateral triangular central area. Each animal was placed in the center of the Y-maze and was free to explore the arena for 8 min. Rats tend to explore the least recently visited arm, and thus tend to alternate visits between the three arms. For efficient alternation, rats need to use working memory, and thus, they should maintain an ongoing record of most recently visited arms, and continuously update such record (Wietrzych et al., 2005). An arm entry was scored when rat placed the four paws within that arm. The following dependent variables were registered: total number of arm entries, number of triads (sequence of three consecutive visits to different arms), and percentage of alternation. An alternation was defined as an entry into three different arms on consecutive choices. The percentage of alternation was calculated as the ratio of actual to maximum number of alternations. The maximum number of possible alternations was defined as the total number of arm entries minus 2. A low percentage of alternation is indicative of an impaired spatial working memory because the rat cannot remember which arm it has just visited, and thus shows decreased spontaneous alternation.

Morris water maze test

The apparatus used was previously described in detail by Galeano et al. (2011). Briefly, it consisted of a circular galvanized steel pool (180 cm in diameter and 60 cm in height), painted black, filled with water at $22 \pm 1^{\circ}$ C to a depth of 36–40 cm, and imaginarily divided into four quadrants: North (N), East (E), South (S), and

West (W). A cylindrical platform (10 cm in diameter and made of transparent acrylic plastic) was placed 2 cm above (visible escape platform) or beneath the water surface (hidden escape platform) in the center of one of the quadrants. To enhance the visibility of the platform during the cued learning training, a "flag" was attached to the platform. The pool was located in the center of an experimental room with multiple extra-maze visual geometric cues hanging on the wall. Indirect illumination was provided by four spiral compact fluorescent lamp in each corner facing the walls. The following dependent variables were recorded: latency to the platform, distance swum to the platform, swimming speed, time spent in each quadrant, the number of crossings over an annulus zone of 20 cm in diameter (defined as a zone surrounding the location where the platform was located during the spatial learning training), and time spent in the outer ring of the pool (25 cm wide).

Cued learning. During cued learning the platform protruded 2 cm above water surface and a "flag" was attached to it (visible escape platform). The maze was surrounded by black curtains to minimize the availability of extra-maze cues. For each of the four trials conducted on each day, the platform was moved to a different quadrant and a different start location was used. If a rat had not located the platform before 120 s elapsed, it was gently guided to the platform location and was allowed to remain there for 15 s. Inter-trial interval duration was 20–30 min. Two days of cued training were conducted.

Spatial learning and reference memory. We used procedures described previously with some modifications (Vorhees and Williams, 2006; Galeano et al., 2011). Black curtains were removed and the spatial learning phase of the task was performed over five consecutive days with four trials per day. In each session, rats were released into the pool from one of the four starting positions (one per each quadrant) and the order of the sequence was changed pseudorandomly between days. Animals were able to escape from the water using the hidden escape platform that was kept in the same location throughout the five spatial learning sessions. A trial was finished when the animal found the escape platform or when 120 s had elapsed, whichever occurred first. If a rat failed to find the platform, the experimenter guided to it by hand. Rats remained on the platform for 15 s with an intertrial interval of 20-30 min. Twenty four hour after the last trial of the learning phase, spatial reference memory was assessed with a probe trial in which the escape platform was removed from the pool and rats were released from a new starting position not used during the learning phase. Time spent in each quadrant, the number of annulus crossings and the time spent in the outer ring of the pool (thigmotactic swimming) was recorded. After each trial rats were dried and when the session finished they were returned to their colony room.

IMAGE ANALYSIS

For stereological estimations the contours of ventral and dorsal hippocampus showing immunoreactivity for iA β was drawn in each section with Image J software (NIH, USA) tool to determine the affected area. The Cavalieri's principle (Gundersen

and Jensen, 1987) was used to calculate the affected volume by using the following formula: $V = \sum AxNxE$, where $V = \text{volume } (\text{mm}^3)$; $\sum A = \text{sum of area counted } (\text{mm}^2)$; N = number of sections analyzed; E = thickness of each section (mm).

STATISTICAL ANALYSIS

All data are shown as the mean \pm s.e.m. Data were analyzed by Two-Way ANOVA or by Two-Way mixed ANOVA tests followed by Tukey's HSD *post-hoc* tests for multiple comparisons, unless noted otherwise. The significance level was set at 5%. SPSS 15.0 for Windows software (Chicago, IL, USA) was used to perform all statistical analyses.

RESULTS

PROGRESSION OF SOLUBLE A β AND iA β ACCUMULATION IN Tg+/- RATS

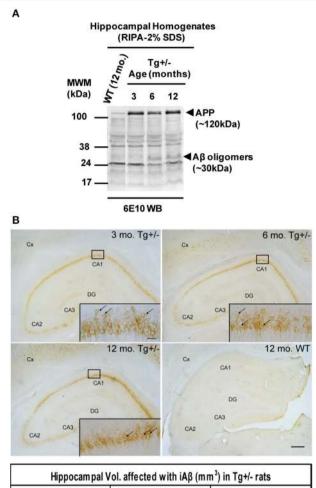
In order to better characterize Aβ-mediated neuropathology at early age, we quantified the levels of SDS-soluble and SDS-resistant peptide AB immunoreactivity in hippocampal homogenates from 3, 6, and 12 months old animals by a commercially available ELISA test and Western blotting, respectively. By ELISA, we detected the presence of SDS-soluble human Aβ40 in $Tg^{+/-}$ hippocampal homogenates early at the age of 3 months $(9.16 \pm 1.42 \,\mathrm{pg/mg})$. The levels of SDS-soluble human A β 40, however, did not differ significantly between Tg^{+/-} rats at 6 months (9.84 \pm 1.30 pg/mg) or 12 months (8.00 \pm 3.19 pg/mg) of age (F < 1), with an average of 9.00 \pm 1.92 pg of A β 40/mg of total hippocampal proteins. By contrast, we were not able to detect SDS-soluble human Aβ42 peptide in RIPA homogenates of $Tg^{+/-}$ rats at any age analyzed. In addition, SDS-resistant A β oligomers appeared in hippocampal homogenates of 6 and 12 months old Tg^{+/-} rats, but not in 3 months old Tg^{+/-} rats (see Figure 1A). In accordance with previous reports, our data show that in $Tg^{+/-}$ rats the neuropathology was restricted to iA β (see Figure 1B, upper panel) and no significant differences were obtained in the volume fraction affected in dorsal or ventral hippocampus among different ages analyzed (F < 1 for both areas analyzed; see Figure 1B, lower panel). Our data strongly suggest a subtle phenotype in $Tg^{+/-}$ rats with an "age-independent" accumulation for SDS-soluble monomeric AB40 and iAB but an "age-dependent" accumulation for SDS-resistant Aβ oligomers.

GENERAL HEALTH AND NEUROLOGICAL SCREENING

Gross inspection of each rat prior to the behavioral experiments did not reveal any visible differences between WT and $Tg^{+/-}$ animals. All animals appeared healthy and neurological reflexes were all scored as being present.

ELEVATED PLUS MAZE TEST

Two-Way ANOVA tests revealed significant main effects of age for "total distance moved" and "number of closed arm entries" $[F_{(2, 51)} = 8.36, p = 0.001; F_{(2, 51)} = 9.29, p < 0.001, respectively]. By contrast, neither the main effects of genotype nor the age × genotype interactions were significant (<math>F < 1$ for all cases). These results suggest that locomotor activity changed with age, regardless of the genotype. *Post-hoc* analysis confirmed that both 6- and 12-month-old WT and $Tg^{+/-}$ rats moved less distance and



Hippocampal Vol. affected with iAβ (mm³) in Tg+/- rats					
3 mo.		6 mo.		12 m o.	
Dorsal	Ventral	Dorsal	Ventral	Dorsal	Ventral
102.4 ± 13.4	296.3 ± 10.7	119.6 ± 20.6	303.2 ± 28.2	97.4 ± 10.2	310.7 ± 7.0

FIGURE 1 | SDS-resistant Aβ **oligomers and intracellular A**β **(iAβ) accumulation in Tg**^{+/-} **rats. (A)** Representative Western blotting showing APP expression and Aβ oligomers by using 6E10 monoclonal antibody. Hippocampal samples of 6 and 12-month-old $Tg^{+/-}$ rats contained Aβ oligomers (~30 kDa) in contrast to 3-month-old $Tg^{+/-}$ or 12-month-old wild-type (WT) littermates. (B) iAβ is present in the hippocampus and cortex (Cx) of $Tg^{+/-}$ animals at 3, 6, and 12 months of age (upper panels). Scale bar, 300 μm. Inset, magnification of CA1 region decorated with iAβ (arrows) in the neurons of granular layer. Scale bar, 30 μm. Hippocampal volume affected with iAβ is similar among different ages analyzed (lower panel). Three groups (n = 3/group) of naïve $Tg^{+/-}$ male rats of 3, 6, and 12 months of age, and one group (n = 3) of naïve WT male rats of 12 months of age were assessed. These animals were not submitted to behavioral testing.

had a lower number of entries into closed arms than 3-monthold groups (p < 0.05 for all comparisons; see **Figures 3A,B**). Regarding "percentage of open arm entries" and "percentage of time spent in open arms," neither the main effects of age nor the main effects of genotype nor the age × genotype interactions were significant (F < 1 for all cases; see **Figures 3C,D**), indicating that levels of anxiety were similar among all experimental groups.

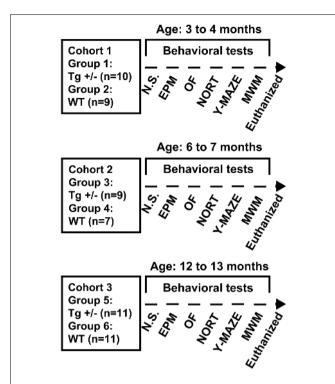
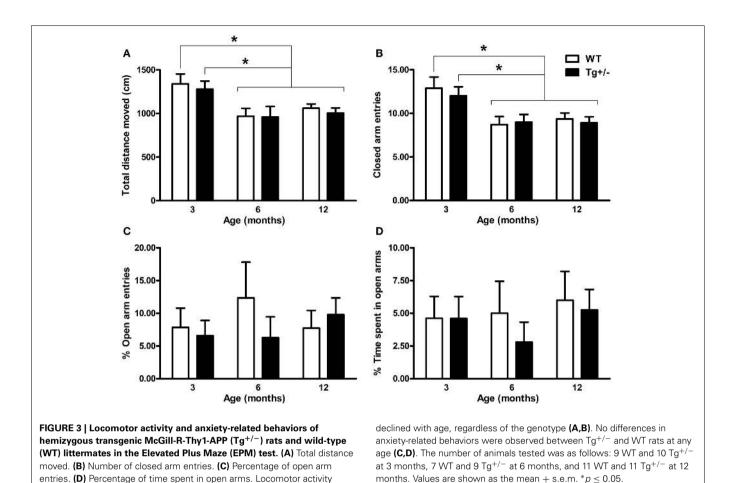


FIGURE 2 | Experimental design, cohorts' characteristics and behavioral tests employed. Three cohorts (1, 2, and 3) of 3, 6, and 12 months old hemizygous transgenic McGill-R-Thy1-APP (Tg+/-) rats and their wild-type (WT) littermates were assessed in a behavioral test battery including: Elevated Plus Maze (EPM) test, Open Field (OF) test, Novel Object Recognition Test (NORT), spontaneous alternation in the Y-maze test (Y-maze), and cued and spatial reference memory versions of the Morris Water Maze (MWM) test. Prior to behavioral assessment, a neurological screening (N.S.) was performed and animals were euthanized after completion of behavioral testing.

OPEN FIELD TEST

When "total distance moved" and "number of rears" were analyzed by Two-Way ANOVA tests, the main effects of age were significant $[F_{(2, 51)} = 13.98, p < 0.001; F_{(2, 51)} = 11.10, p < 0.001,$ respectively] but neither the main effects of genotype nor the age \times genotype interactions were significant ($F \le 1$ for all cases). Post-hoc multiple comparisons confirmed that locomotor and rearing activity declined with age, since 6- and 12-month-old WT and Tg^{+/-} rats moved less distance and presented a lower rearing frequency than 3-month-old groups (p < 0.05 for all comparisons; see Figures 4A,B). Relative to anxiety levels, the Two-Way ANOVA tests for "number of entries into the center" and "time spent in central area" indicated that neither the main effects of age nor the age \times genotype interactions were significant (F < 1for all cases). However, the main effects of genotype showed significant differences $[F_{(2,51)} = 8.46, p = 0.005; F_{(2,51)} = 13.87,$ p < 0.001, respectively]. Post-hoc analyses revealed that 6- and 12-month-old Tg^{+/-} rats showed a lower number of entries and spent less time in the center of the OF than WT groups (p < 0.05for all comparisons; see Figures 4C,D). In contrast, 3-month-old Tg^{+/-} rats did not differ from their WT littermates in none of the variables analyzed (see Figures 4C,D). Since the interactions



age \times genotype did not reach statistical significance, the lack of differences between 3 month-old WT and $Tg^{+/-}$ groups should be taken with caution (Tybout et al., 2001).

NOVEL OBJECT RECOGNITION TEST

The discrimination index (d1) and discrimination ratio (d2) were not statistically different between genotypes or ages [Two-Way ANOVA tests. d1. Genotype: $F_{(1, 51)} = 2.25$, p = n.s.; Age: $F_{(2, 51)} < 1$; Genotype × age: $F_{(2, 51)} < 1$. d2. Genotype: $F_{(1, 51)} < 1$; Age: $F_{(2, 51)} = 1.25$, p = n.s.; Genotype × age: $F_{(2, 51)} < 1$]. One sample t-tests indicated that d1 and d2 scores were significantly different from those expected by chance for all experimental groups (p < 0.05 for all cases; see **Figures 5A,B**). These results demonstrate that WT and $Tg^{+/-}$ rats were able to recognize the novel object from the familiar one at all ages tested.

Y-MAZE TEST

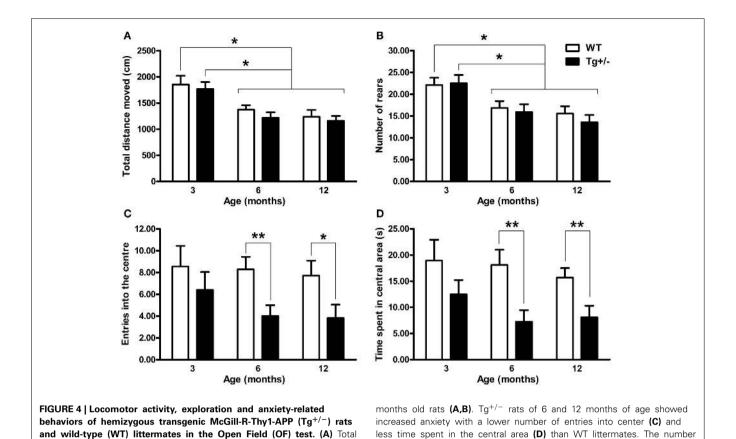
A significant main effect of age for "total number of arm entries" $[F_{(2, 51)} = 17.52, p < 0.001]$ was revealed by a Two-Way ANOVA test. Neither the main effect of genotype nor the age × genotype interaction were significant (F < 1 for both cases). Regardless of the genotype, *post-hoc* analysis showed that 12-month-old WT and $Tg^{+/-}$ rats displayed a lower number of arm entries than 3- and 6-month-old groups (p < 0.01 for all comparisons; see **Figure 6A**). When the "percentage of alternation" was also

analyzed by a Two-Way ANOVA test, the main effects of age and genotype and the interaction were all significant $[F_{(2, 51)} = 4.48, p = 0.016; F_{(2, 51)} = 7.81, p = 0.007; F_{(2, 51)} = 3.13, p = 0.05, respectively].$ *Post-hoc*tests indicated that 6- and 12-monthold Tg^{+/-} rats showed a lower percentage of alternation than their WT littermates (<math>p = 0.020 and p = 0.006, respectively; see **Figure 6B**), while 3-month-old groups displayed similar levels of alternation (see **Figure 6B**). These results indicate that Tg^{+/-} rats displayed a spontaneous alternation impairment at 6 and 12 months of age.

MORRIS WATER MAZE TEST

Cued learning

At the three ages studied, *post-hoc* multiple comparisons showed that, during the second day, both genotypes significantly reduced their latency to reach the visible platform to similar levels (p < 0.01 for all cases; see **Figures 7A–C**) [Two-Way mixed ANOVA test. 3 months old. Day: $F_{(1, 17)} = 66.57$, p < 0.001; Genotype: $F_{(1, 17)} < 1$; Day × genotype: $F_{(1, 17)} < 1$. 6 months old. Day: $F_{(1, 14)} = 145.87$, p < 0.001; Genotype: $F_{(1, 14)} = 1.44$, p = n.s.; Day × genotype: $F_{(1, 14)} < 1$. 12 months old. Day: $F_{(1, 20)} = 400.14$, p < 0.001; Genotype: $F_{(1, 20)} < 1$; Day × genotype: $F_{(1, 20)} = 8.27$, p < 0.01]. These results suggest that transgenic groups were not visually impaired and showed no deficit in swimming ability.



Spatial learning

Two-Way mixed ANOVA tests, with day as within-subject factor and genotype as between-subject factor, indicated that 3, 6, and 12 months old rats significantly decreased their escape latencies and distances swum to reach the hidden platform across days, regardless of the genotype [Escape Latency. 3 months old. Day: $F_{(4, 68)} = 40.36$, p < 0.001; Genotype: $F_{(1, 17)} < 1$; Day \times genotype: $F_{(4, 68)} = 1.6$, p = n.s. 6 months old. Day: $F_{(4, 56)} = 20.42, p < 0.001$; Genotype: $F_{(1, 14)} < 1$; Day × genotype: $F_{(4, 56)} < 1$. 12 months old. Day: $F_{(4, 80)} = 61.36$, p < 10.001; Genotype: $F_{(1, 20)} < 1$; Day × genotype: $F_{(4, 80)} < 1$. Distance swum. 3 months old. Day: $F_{(4, 68)} = 48.24$, p < 0.001; Genotype: $F_{(1, 17)} < 1$; Day × genotype: $F_{(4, 68)} < 1$. 6 months old. Day: $F_{(4, 56)} = 21.14$, p < 0.001; Genotype: $F_{(1, 14)} < 1$; Day × genotype: $F_{(4, 56)}$ < 1. 12 months old. Day: $F_{(4, 80)}$ = 50.67, p < 0.001; Genotype: $F_{(1, 20)} < 1$; Day × genotype: $F_{(4,80)} < 1$]. Post-hoc analyses confirmed that, at any age, escape latencies and distances swum did not differ between WT and Tg^{+/-} groups on any day of the learning phase (p = n.s. for all comparisons), indicating that all experimental groups were able to learn the task at an equivalent rate (see Figures 8A-F). Moreover, both genotypes displayed similar swimming speeds at all ages (see Figure 1 in Supplementary Material), suggesting similar levels of motivation to solve the task.

distance moved. (B) Number of rears. (C) Entries into the center. (D) Time spent in the central area. Regardless of the genotype, 6 and 12

months old rats showed less locomotion and rearing activity than 3

Spatial reference memory

Two-Way mixed ANOVA tests, with quadrant as within-subject factor and genotype as between-subject factor, were performed to analyze the amount of time that experimental groups spent in each one of the quadrant of the MWM when platform was removed. At the three ages analyzed, results showed that the main effects of quadrant and the quadrant × genotype interactions were significant, in contrast to the main effects of genotype [3 months old. Quadrant: $F_{(3, 51)} = 5.49$, p = 0.02; Genotype: $F_{(1, 17)} < 1$; Quadrant x genotype: $F_{(3, 51)} = 2.73$, p = 0.04. 6 months old. Quadrant: $F_{(3, 42)} = 3.74$, p = 0.02; Genotype: $F_{(1, 14)} < 1$; Quadrant × genotype: $F_{(3, 42)} = 2.70$, p = 0.05. 12 months old. Quadrant: $F_{(3, 60)} = 2.87$, p = 0.04; Genotype: $F_{(1, 20)} < 1$; Quadrant × genotype: $F_{(3, 60)} = 2.74$, p = 0.04]. Post-hoc tests revealed that 3, 6, and 12 months old WT rats spent significantly more time in the quadrant where the platform was located (target quadrant) than in the remaining quadrants (p < 0.05 for all comparisons; see Figures 9A,C,E). On the contrary, Tg^{+/-} rats spent similar amount of time in each one of the quadrant at any age (p = n.s. for all comparisons; see **Figures 9A,C,E**). Furthermore, Student's *t*-tests revealed that 3 and 12 months old WT rats crossed the annulus zone significantly more time than $Tg^{+/-}$ rats (p < 0.05 for both cases; see **Figures 9B,F**). A strong tendency was also detected for the 6 months old WT group (see **Figure 9D**). Overall, these results indicate that $Tg^{+/-}$ rats had a

of animals tested was as follows: 9 WT and 10 Tg+/- at 3 months, 7

WT and 9 Tg^{+/-} at 6 months, and 11 WT and 11 Tg^{+/-} at 12 months. Values are shown as the mean + s.e.m. * $p \le 0.05$; ** $p \le 0.01$.

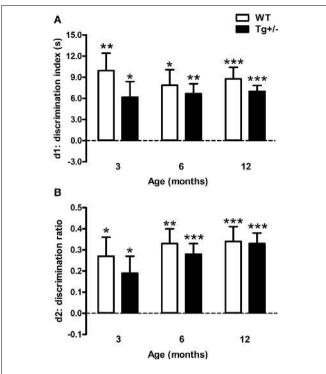


FIGURE 5 | Performance of hemizygous transgenic McGill-R-Thy1-APP (Tg+/-) rats and wild-type (WT) littermates in the Novel Object Recognition Test (NORT). (A) Discrimination index (d1). (B) Discrimination ratio (d2). At all the ages tested, both genotypes recognized the novel object. The dashed lines indicate an equivalent exploration time for the two objects (chance level). The number of animals tested was as follows: 9 WT and 10 Tg+/- at 3 months, 7 WT and 9 Tg+/- at 6 months, and 11 WT and 11 Tg+/- at 12 months. Values are shown as the mean + s.e.m. * $p \le 0.05$; ** $p \le 0.01$; *** $p \le 0.001$ vs. chance (dashed lines).

significant impairment to remember the location of the platform as compared to WT animals. Regarding swimming speed and thigmotactic swimming, no differences were detected in probe trials (see Figures 1, 2 in Supplementary Material).

DISCUSSION

Hemizygous McGill-R-Thy1-APP transgenic rats represent an interesting model of early AD pathology as the presence of AB is limited to the intracellular compartment. While the longitudinal accumulation of AB in its homozygous counterpart has been recently documented (Iulita et al., 2014), little is known regarding the progression of the Aβ intracellular accumulation in the heterozygous counterpart and of the behavioral outcomes. In order to address this issue, we investigated the occurrence of human Aβ40 peptides after 2% SDS solubilization in hippocampal homogenates from 3-, 6-, and 12-month-old $Tg^{+/-}$ rats by using an ELISA specific for human AB peptide. We were able to detect accumulation of Aβ40 immunoreactive material which remained at similar levels from 3 to 12 months of age. This finding is in agreement with the data reported in homozygous McGill-R-Thy1-APP rats (Iulita et al., 2014). The impossibility to detect Aβ42 peptide may be explained by the low amount of this peptide in the 2% SDS homogenates and the dynamic range of the

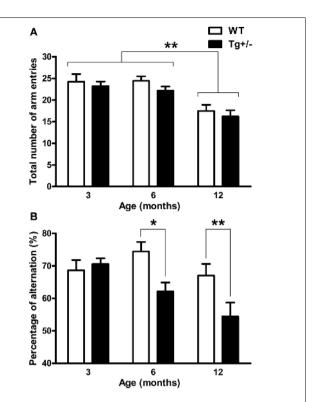


FIGURE 6 | Spontaneous alternation in the Y-maze test. (A) Total number of arm entries. (B) Percentage of alternation. Regardless of the genotype, 12 months old rats showed less total number of arm entries than 3 and 6 months old rats (A). At 6 and 12 months of age, hemizygous transgenic McGill-R-Thy1-APP ($Tg^{+/-}$) rats exhibited an impairment in spontaneous alternation compared with wild-type (WT) groups (B). The number of animals tested was as follows: 9 WT and 10 $Tg^{+/-}$ at 3 months, 7 WT and 9 $Tg^{+/-}$ at 6 months, and 11 WT and 11 $Tg^{+/-}$ at 12 months. Values are shown as the mean + s.e.m. * $p \le 0.05$; ** $p \le 0.01$.

ELISA test used in this experiment (between 1.56 and 100 pg/ml). Yet, the presence of SDS-resistant AB oligomers was detected by Western blot in 6- and 12-month-old animals. Since an aminoterminal anti-Aß was used it was not possible to determine if these oligomers were composed of Aβ40, Aβ42 or both. To complement the structural and neurochemical characterization, we performed a longitudinal behavioral analysis of male hemizygous transgenic McGill-R-Thy1-APP $(Tg^{+/-})$ rats, in tightly controlled conditions. To avoid high variability already reported in Tg^{+/-} groups (Leon et al., 2010) we followed 2 strategies: (1) exclusion of females rats, which are known to introduce confounding factors in behavioral tests (Mehta et al., 2013) and (2) the evaluation of at least 9 animals in each $Tg^{+/-}$ group to increase statistical power. As it was mentioned in the "Materials and Methods" Section, the groups of animals that underwent the behavioral assessment were not the same used to study Aβ accumulation.

EMOTIONAL BEHAVIOR

There are several reports on murine models of AD, aimed at understanding the impact of amyloid formation on learning and memory but few are focused on emotional behavior. In this study, we analyzed the effect of genetic modification of APP on

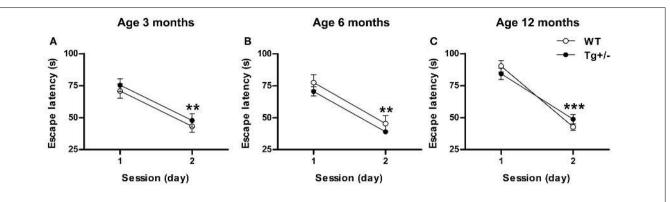


FIGURE 7 | Hemizygous transgenic McGill-R-Thy1-APP ($Tg^{+/-}$) rats showed an unimpaired cued learning in the Morris Water Maze (MWM) test. Escape latency of wild-type (WT) and $Tg^{+/-}$ groups at 3 (A), 6 (B) and 12 (C) months of age. At all ages tested, both genotypes reduced their

escape latencies to similar levels from day 1 to day 2. The number of animals tested was as follows: 9 WT and 10 Tg+/- at 3 months, 7 WT and 9 Tg+/- at 6 months, and 11 WT and 11 Tg+/- at 12 months. Values are shown as the mean \pm s.e.m. ** $p \leq 0.01$; *** $p \leq 0.001$ vs. day 1.

EPM and OF tests in $Tg^{+/-}$ rats in view of the tendency of AD patients to be either apathetic or agitated (Chung and Cummings, 2000; Senanarong et al., 2004). We were unable to detect differences in anxiety-related behaviors between Tg^{+/-} and WT rats in the EPM test in the present study. Unchanged anxiety levels assessed by EPM were described in transgenic mice lacking amyloid plaques (Moran et al., 1995; Lalonde et al., 2002; Lee et al., 2004; Boon et al., 2010) and in two models with Aß plaques (Arendash et al., 2001; Le Cudennec et al., 2008; Blanchard et al., 2009). It is important to note that behavior in EPM was not assessed in any of the transgenic rat models of AD described so far (Do Carmo and Cuello, 2013) and that our results in Tg^{+/-} rats, lacking extracellular AB deposition even at late stages, are in agreement with the concept that changes of EPM behavior appear only in transgenic models of AD with an extensive neuropathological phenotype (Savonenko et al., 2003; Lee et al., 2006; Lalonde et al., 2012). It is important to note that the levels of exploration of open arms seem to be low as compared to other reports (see Figures 3C,D). This effect could be attributable to the levels of illumination used (80-90 lux in the open arms). To rule out the possibility of a floor effect that could be masking an anxiety phenotype in Tg^{+/-} rats in the EPM, future studies are needed to investigate the behavior of these animals in this test under different experimental conditions.

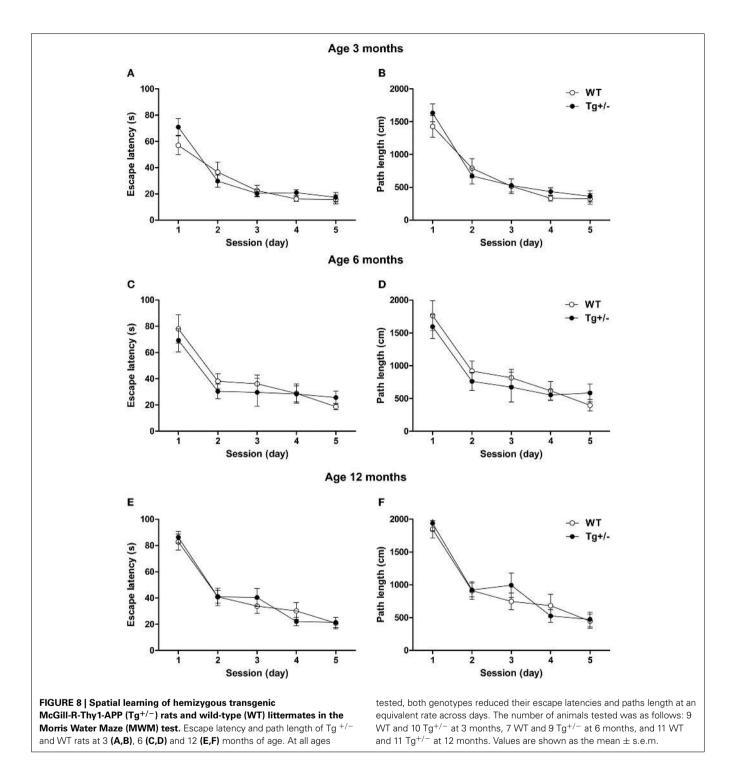
Only one transgenic rat model of AD (the Tg6590) has been previously characterized for its performance in the OF. This transgenic rat model showed increased A β precursor protein (APP), cerebrovascular deposits, few diffuse plaques and increased phosphorylated tau at 15 months of age. At 9 months old these animals showed no differences either in the number of entries into center or in the time spent in the central area of the OF, as compared to control group (Kloskowska et al., 2010). By contrast, here we show that Tg^{+/-} rats are more anxious in the OF test at 6 months of age. This behavior was also observed at 12 months of age, suggesting the robustness of this phenotype. It is worth noting that this anxious phenotype was observed in the OF test but not in the EPM test. Previously, it was reported that the OF and the EPM operationalize different aspects of emotional behavioral (Trullas and Skolnick, 1993; Ramos et al., 1998). This lack of inter-test

correlations has been attributed to the multidimensional nature of emotional behavior (Ramos, 2008). Therefore, it could be possible that the higher anxiety levels detected in Tg^{+/-} rats in the present study, reflect an specific dimension of emotional behavior linked to OF activity. Finally, it is important to note that both tests (EPM and OF) were sensitive to the age-related decline in spontaneous locomotor activity and exploration (see **Figures 3A,B**, **4A,B**).

In summary, even if the emergency (Holmes, 2001) or the light/dark box (Pinton et al., 2011) tests were not assessed in this study, our report is the first analysis of the anxiety disturbances in a transgenic rat model of early AD that does not develop A β plaques. It is of interest that the anxiety pattern described here for the Tg^{+/-} rats appear to closely mimic neuropsychiatric symptoms described in some pre-clinical AD patients.

COGNITIVE BEHAVIOR

Ideally, genetically modified animal models of AD not only have to replicate some of the hallmark neuropathology, such as plaquelike amyloid accumulations and tau deposition, but also should reproduce some of the cognitive impairments relevant to the disease. In this context, near all of the transgenic mice models with excessive Aβ either via transgenic overexpression of mutant hAPP and/or its processing enzymes, typically present static cognitive deficits that are evident early in life and progressive deficits that are age-related and associated with increased concentrations of deposited or unbound assemblies of Aβ (Kobayashi and Chen, 2005). By contrast to transgenic mice in which a wide battery of cognitive tests were performed, a narrower battery of tests were used in transgenic rat models of AD. These included MWM, Fear Conditioning, and Novel Object Recognition and Location (NORL) test (Leon et al., 2010; Iulita et al., 2014). Early description of the cognitive performance of hemizygous transgenic McGill-R-Thy1-APP rat in the MWM showed that at 3 months of age (n = 4) the learning curve was undistinguishable from that of WT (n = 8) rats while at 13 months of age, $Tg^{+/-}$ rats (n = 4) exhibited a mild spatial learning deficit reflected by a significantly increased escape latency, compared to WT (n = 12), during the third day of the learning phase.



Moreover, spatial reference memory, assessed during the probe trial, was not impaired at both ages, although a trend could be seen in the 3 month-old group (Leon et al., 2010). To further characterize the cognitive abilities of $Tg^{+/-}$ rats, in this study we analyzed the performance of these animals in three different tasks including MWM, Novel Object Recognition (NOR) and Y-maze tests at 3, 6 and 12 months old. Using a larger number of $Tg^{+/-}$ animals, we were able to detect, in the MWM, a spatial reference

memory deficit early at 3 months of age, which persisted at 6 and 12 months. This deficit cannot be explained by a slower learning curve, as in contrast to the original report (Leon et al., 2010), we did not find in $Tg^{+/-}$ rats any impairment in the learning phase of the MWM test. These discrepancies can be explained both by the number of animals used and by the gender of the animals utilized (males and females in the first report while males only in our study).

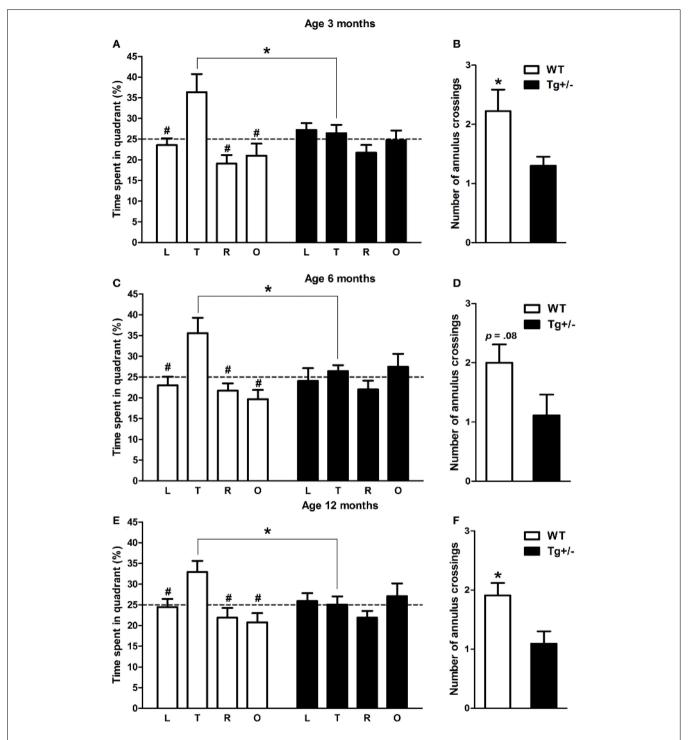


FIGURE 9 | Spatial reference memory impairment in hemizygous transgenic McGill-R-Thy1-APP ($Tg^{+/-}$). Percentage of time spent in each quadrant of the Morris water maze and number of annulus crossings during the probe trial at 3 (A,B), 6 (C,D) and 12 (E,F) months of age. At the three ages tested, WT groups spent significantly more time in the quadrant where the platform was previously located (target quadrant) (A,C,D). By contrast, $Tg^{+/-}$ groups spent similar amount of time in each one of the four quadrants (A,C,D). Furthermore, $Tg^{+/-}$ groups crossed the annulus zone significantly less

times than WT groups (except at 6 months of age when a strong tendency was detected; **B,D,F**), indicating a spatial reference memory impairment. Dashed lines indicate the percentage of time spent in each one of the quadrant predicted by chance (25%). L, left quadrant; T, target quadrant; R, right quadrant; O, opposite quadrant. The number of animals tested was as follows: 9 WT and 10 Tg $^{+/-}$ at 3 months, 7 WT and 9 Tg $^{+/-}$ at 6 months, and 11 WT and 11 Tg $^{+/-}$ at 12 months. Values are shown as the mean \pm s.e.m. $^{\#}p \leq 0.05$ vs. percentage of time spent in target quadrant; $^{*}p \leq 0.05$.

Memory tasks sensitive to hippocampal functions such as contextual fear conditioning (FC), NORL and Y-maze alternation have been described in different transgenic mice models of AD (Corcoran et al., 2002; Dineley et al., 2002; Ohno et al., 2004; Devi and Ohno, 2012; Mitani et al., 2013; Nagahara et al., 2013). However, none of these tests were assessed in transgenic rats until a recent report in which hemizygous transgenic McGill-R-Thy1-APP (Tg^{+/-}) rats showed reduced auditory Fear Conditioning (FC) and reduced contextual and cued FC recall (Iulita et al., 2014). In the same study, $Tg^{+/-}$ rats displayed an impairment in the Novel Object Recognition and Location (NORL) task (Iulita et al., 2014). The impairment described in the NORL test was not reproduced by Tg^{+/-} in the NOR task performed in our study. This could be due to the differential stringency of the tasks as five objects were used in the previous report (Iulita et al., 2014). Although both tests are hippocampal-dependent, the NORL is a more complex task that includes the spatial component that is absent in the classic NORT test, and may explain the discrepancy between results. While in the former test one unfamiliar object had to be discriminated among a total of five objects during the novel object recognition phase of the task, we employed a more classical approach with two dissimilar objects (one unfamiliar and one familiar) that had to be discriminated during the retention trial. Discrepancies in results reveal that when the complexity of the task is increased, recognition memory may be also impaired in this transgenic rat model.

Spontaneous alternation in the Y-maze test estimates the willingness to explore novel stimuli, or to avoid familiar stimuli, and is an appropriate paradigm to asses working memory and/or behavioral disinhibition, signs also manifest in AD. Also, the total number of arm entries in the Y-maze is a parameter used to assess sensoriomotor capacities. In this study, we did not find significant differences in the total number of arm entries between Tg^{+/-} and WT rats. However, a significant decline was detected in the percentage of alternation in $Tg^{+/-}$ rats at 6 and 12 months old. We could not ascribe this reduced rate in spontaneous alternation to behavioral disinhibition since performance of Tg^{+/-} rats in locomotor and exploratory parameters in the EPM and OF did not differ from WT. Therefore, it is likely that reduced spontaneous alternation reflects spatial working memory impairment. Poor performance in Y-maze appears in young mice of several single transgenic AD models at 2-4 months, when there are no plaque-like deposits, but these mice have had high levels of soluble AB in their nervous systems from early postnatal life (Lalonde et al., 2012). These observations are in agreements with our results, since Tg^{+/-} rats lack parenchymal Aβ plaques but thus accumulate soluble oligomers intraneuronally.

CONCLUDING REMARKS

In conclusion, in the present time-course study, we investigated emotional and cognitive symptoms in $Tg^{+/-}$ rats. The most relevant result is that transgenic rats showed, from adulthood (6-month-old) to middle age (12-month-old), increased anxiety levels and working memory impairment. Moreover, as early as 3 months of age, transgenic rats showed an impaired spatial reference memory in comparison to WT rats. This deficit was also observed at 6 and 12 months of age. At all ages, recognition

memory and spatial learning were spared. Given that $Tg^{+/-}$ rats capture many behavioral features of early stages of AD, this new model may represent an important tool for the neuroscience community to enable future studies in basic and translational AD research.

ACKNOWLEDGMENTS

This work was supported by grants from Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET-PIP 693) and Fundación Florencio Fiorini (to Laura Morelli), Alzheimer's Association (IIRG-11-205127) (to Eduardo M. Castaño) and Canadian Institute for Health and Research (MOP-97776) (to A. Claudio Cuello). Pablo Galeano and Pamela V. Martino Adami are supported by CONICET fellowships. Sonia Do Carmo is holder of a Charles E. Frosst-Merck post-doctoral fellowship. Eduardo M. Castaño, Francisco Capani and Laura Morelli are investigators of CONICET. A. Claudio Cuello is holder of a McGill University Charles E. Frosst-Merck Chair in Pharmacology. We would like to thank Tamara Logica Tornatore, Mariana Inés Holubiec and Juan Ignacio Romero for their helpful assistance during the behavioral tests.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: http://www.frontiersin.org/journal/10.3389/fnbeh.2014. 00321/abstract

REFERENCES

Abbott, A. (2004). Laboratory animals: the Renaissance rat. Nature 428, 464–466. doi: 10.1038/428464a

Ally, B. A. (2012). Using pictures and words to understand recognition memory deterioration in amnestic mild cognitive impairment and Alzheimer's disease: a review. Curr. Neurol. Neurosci. Rep. 12, 687–694. doi: 10.1007/s11910-012-0310-7

Arendash, G. W., King, D. L., Gordon, M. N., Morgan, D., Hatcher, J. M., Hope, C. E., et al. (2001). Progressive, age-related behavioral impairments in transgenic mice carrying both mutant amyloid precursor protein and presenilin-1 transgenes. *Brain Res.* 891, 42–53. doi: 10.1016/S0006-8993(00)03186-3

Baddeley, A. (2003). Working memory: looking back and looking forward. Nat. Rev. Neurosci. 4, 829–839. doi: 10.1038/nrn1201

Bateman, R. J., Xiong, C., Benzinger, T. L., Fagan, A. M., Goate, A., Fox, N. C., et al. (2012). Clinical and biomarker changes in dominantly inherited Alzheimer's disease. N. Engl. J. Med. 367, 795–804. doi: 10.1056/NEJMoa1202753

Blanchard, J., Decorte, L., Nogues, X., and Micheau, J. (2009). Characterization of cognition alteration across the course of the disease in APP751SL mice with parallel estimation of cerebral Abeta deposition. *Behav. Brain Res.* 201, 147–157. doi: 10.1016/j.bbr.2009.02.005

Bondi, M. W., Jak, A. J., Delano-Wood, L., Jacobson, M. W., Delis, D. C., and Salmon, D. P. (2008). Neuropsychological contributions to the early identification of Alzheimer's disease. *Neuropsychol. Rev.* 18, 73–90. doi: 10.1007/s11065-008-9054-1

Bondi, M. W., Salmon, D. P., Galasko, D., Thomas, R. G., and Thal, L. J. (1999). Neuropsychological function and apolipoprotein E genotype in the preclinical detection of Alzheimer's disease. *Psychol. Aging* 14, 295–303. doi: 10.1037/0882-7974.14.2.295

Boon, W. C., Van Den Buuse, M., Wegener, N., Martin, S., Chua, H. K., Bush, A. I., et al. (2010). Behavioural phenotype of APPC100.V717F transgenic mice over-expressing a mutant Abeta-bearing fragment is associated with reduced NMDA receptor density. *Behav. Brain Res.* 209, 27–35. doi: 10.1016/j.bbr.2010. 01.013

Chung, J. A., and Cummings, J. L. (2000). Neurobehavioral and neuropsychiatric symptoms in Alzheimer's disease: characteristics and treatment. *Neurol. Clin.* 18, 829–846. doi: 10.1016/S0733-8619(05)70228-0

Cohen, R. M., Rezai-Zadeh, K., Weitz, T. M., Rentsendorj, A., Gate, D., Spivak, I., et al. (2013). A transgenic Alzheimer rat with plaques, tau pathology, behavioral impairment, oligomeric Aβ, and frank neuronal loss. *J. Neurosci.* 33, 6245–6256. doi: 10.1523/JNEUROSCI.3672-12.2013

- Collie, A., and Maruff, P. (2000). The neuropsychology of preclinical Alzheimer's disease and mild cognitive impairment. *Neurosci. Biobehav. Rev.* 24, 365–374. doi: 10.1016/S0149-7634(00)00012-9
- Corcoran, K. A., Lu, Y., Turner, R. S., and Maren, S. (2002). Overexpression of hAPPswe impairs rewarded alternation and contextual fear conditioning in a transgenic mouse model of Alzheimer's disease. *Learn. Mem.* 9, 243–252. doi: 10.1101/lm.51002
- D'Andrea, M. R., Nagele, R. G., Wang, H. Y., Peterson, P. A., and Lee, D. H. (2001). Evidence that neurones accumulating amyloid can undergo lysis to form amyloid plaques in Alzheimer's disease. *Histopathology* 38, 120–134. doi: 10.1046/j.1365-2559.2001.01082.x
- Devi, L., and Ohno, M. (2012). Mitochondrial dysfunction and accumulation of the β-secretase-cleaved C-terminal fragment of APP in Alzheimer's disease transgenic mice. *Neurobiol. Dis.* 45, 417–424. doi: 10.1016/j.nbd.2011. 09.001
- Dineley, K. T., Xia, X., Bui, D., Sweatt, J. D., and Zheng, H. (2002). Accelerated plaque accumulation, associative learning deficits, and up-regulation of alpha 7 nicotinic receptor protein in transgenic mice co-expressing mutant human presenilin 1 and amyloid precursor proteins. *J. Biol. Chem.* 277, 22768–22780. doi: 10.1074/jbc.M200164200
- Do Carmo, S., and Cuello, A. C. (2013). Modeling Alzheimer's disease in transgenic rats. Mol. Neurodegener. 8:37. doi: 10.1186/1750-1326-8-37
- Echeverria, V., Ducatenzeiler, A., Alhonen, L., Janne, J., Grant, S. M., Wandosell, F., et al. (2004). Rat transgenic models with a phenotype of intracellular Abeta accumulation in hippocampus and cortex. *J. Alzheimers Dis.* 6, 209–219.
- Ennaceur, A. (2010). One-trial object recognition in rats and mice: methodological and theoretical issues. *Behav. Brain Res.* 215, 244–254. doi: 10.1016/j.bbr.2009.12.036
- Ennaceur, A., and Delacour, J. (1988). A new one-trial test for neurobiological studies of memory in rats. 1: behavioral data. *Behav. Brain Res.* 31, 47–59. doi: 10.1016/0166-4328(88)90157-X
- Eustache, F., Giffard, B., Rauchs, G., Chételat, G., Piolino, P., and Desgranges, B. (2006). Alzheimer's disease and human memory. Rev. Neurol. (Paris) 162, 929–939. doi: 10.1016/S0035-3787(06)75102-5
- Gainotti, G., Quaranta, D., Vita, M. G., and Marra, C. (2014). Neuropsychological predictors of conversion from mild cognitive impairment to Alzheimer's disease. J. Alzheimers Dis. 38, 481–495. doi: 10.3233/JAD-130881
- Galeano, P., Blanco Calvo, E., Madureira De Oliveira, D., Cuenya, L., Kamenetzky, G. V., Mustaca, A. E., et al. (2011). Long-lasting effects of perinatal asphyxia on exploration, memory and incentive downshift. *Int. J. Dev. Neurosci.* 29, 609–619. doi: 10.1016/j.ijdevneu.2011.05.002
- Geda, Y. E., Schneider, L. S., Gitlin, L. N., Miller, D. S., Smith, G. S., Bell, J., et al. (2013). Neuropsychiatric symptoms in Alzheimer's disease: past progress and anticipation of the future. *Alzheimers Dement.* 9, 602–608. doi: 10.1016/j.jalz.2012.12.001
- Gouras, G. K., Tsai, J., Naslund, J., Vincent, B., Edgar, M., Checler, F., et al. (2000). Intraneuronal Abeta42 accumulation in human brain. Am. J. Pathol. 156, 15–20. doi: 10.1016/S0002-9440(10)64700-1
- Gundersen, H. J., and Jensen, E. B. (1987). The efficiency of systematic sampling in stereology and its prediction. J. Microsc. 147, 229–263. doi: 10.1111/j.1365-2818.1987.tb02837.x
- Holmes, A. (2001). Targeted gene mutation approaches to the study of anxiety-like behavior in mice. *Neurosci. Biobehav. Rev.* 25, 261–273. doi: 10.1016/S0149-7634(01)00012-4
- Iulita, M. F., Allard, S., Richter, L., Munter, L. M., Ducatenzeiler, A., Weise, C., et al. (2014). Intracellular Aβ pathology and early cognitive impairments in a transgenic rat model overexpressing human amyloid precursor protein: a multidimensional study. Acta Neuropathol. Commun. 2:61. doi: 10.1186/2051-5960-2-61
- Kloskowska, E., Pham, T. M., Nilsson, T., Zhu, S., Oberg, J., Codita, A., et al. (2010). Cognitive impairment in the Tg6590 transgenic rat model of Alzheimer's disease. J. Cell. Mol. Med. 14, 1816–1823. doi: 10.1111/j.1582-4934.2009.00809.x
- Kobayashi, D. T., and Chen, K. S. (2005). Behavioral phenotypes of amyloid-based genetically modified mouse models of Alzheimer's disease. *Genes Brain Behav.* 4, 173–196. doi: 10.1111/j.1601-183X.2005.00124.x

- Lalonde, R., Dumont, M., Fukuchi, K., and Strazielle, C. (2002). Transgenic mice expressing the human C99 terminal fragment of betaAPP: effects on spatial learning, exploration, anxiety, and motor coordination. *Exp. Gerontol.* 37, 1401–1412. doi: 10.1016/S0531-5565(02)00123-7
- Lalonde, R., Fukuchi, K., and Strazielle, C. (2012). APP transgenic mice for modelling behavioural and psychological symptoms of dementia (BPSD). *Neurosci. Biobehav. Rev.* 36, 1357–1375. doi: 10.1016/j.neubiorev.2012.02.011
- Laurijssens, B., Aujard, F., and Rahman, A. (2013). Animal models of Alzheimer's disease and drug development. *Drug Discov. Today Technol.* 10, e319–327. doi: 10.1016/j.ddtec.2012.04.001
- Leal, M. C., Dorfman, V. B., Gamba, A. F., Frangione, B., Wisniewski, T., Castaño, E. M., et al. (2006). Plaque-associated overexpression of insulindegrading enzyme in the cerebral cortex of aged transgenic tg2576 mice with Alzheimer pathology. J. Neuropathol. Exp. Neurol. 65, 976–987. doi: 10.1097/01.inen.0000235853.70092.ba
- Le Cudennec, C., Faure, A., Ly, M., and Delatour, B. (2008). One-year longitudinal evaluation of sensorimotor functions in APP751SL transgenic mice. *Genes Brain Behav.* 7, 83–91. doi: 10.1111/j.1601-183X.2007.00374.x
- Lee, K. W., Im, J. Y., Song, J. S., Lee, S. H., Lee, H. J., Ha, H. Y., et al. (2006). Progressive neuronal loss and behavioral impairments of transgenic C57BL/6 inbred mice expressing the carboxy terminus of amyloid precursor protein. *Neurobiol. Dis.* 22, 10–24. doi: 10.1016/j.nbd.2005.09.011
- Lee, K. W., Lee, S. H., Kim, H., Song, J. S., Yang, S. D., Paik, S. G., et al. (2004). Progressive cognitive impairment and anxiety induction in the absence of plaque deposition in C57BL/6 inbred mice expressing transgenic amyloid precursor protein. J. Neurosci. Res. 76, 572–580. doi: 10.1002/jnr.20127
- Leon, W. C., Canneva, F., Partridge, V., Allard, S., Ferretti, M. T., Dewilde, A., et al. (2010). A novel transgenic rat model with a full Alzheimer's-like amyloid pathology displays pre-plaque intracellular amyloid-beta-associated cognitive impairment. J. Alzheimers Dis. 20, 113–126. doi: 10.3233/JAD-2010-1349
- Mehta, N. S., Wang, L., and Redei, E. E. (2013). Sex differences in depressive, anxious behaviors and hippocampal transcript levels in a genetic rat model. *Genes Brain Behav.* 12, 695–704. doi: 10.1111/gbb.12063
- Mitani, Y., Yarimizu, J., Akashiba, H., Shitaka, Y., Ni, K., and Matsuoka, N. (2013). Amelioration of cognitive deficits in plaque-bearing Alzheimer's disease model mice through selective reduction of nascent soluble Aβ42 without affecting other Aβ pools. *J. Neurochem.* 125, 465–472. doi: 10.1111/jnc.12125
- Moran, P. M., Higgins, L. S., Cordell, B., and Moser, P. C. (1995). Age-related learning deficits in transgenic mice expressing the 751-amino acid isoform of human beta-amyloid precursor protein. *Proc. Natl. Acad. Sci. U.S.A.* 92, 5341–5345. doi: 10.1073/pnas.92.12.5341
- Nagahara, A. H., Mateling, M., Kovacs, I., Wang, L., Eggert, S., Rockenstein, E., et al. (2013). Early BDNF treatment ameliorates cell loss in the entorhinal cortex of APP transgenic mice. *J. Neurosci.* 33, 15596–15602. doi: 10.1523/JNEUROSCI.5195-12.2013
- Ohno, M., Sametsky, E. A., Younkin, L. H., Oakley, H., Younkin, S. G., Citron, M., et al. (2004). BACE1 deficiency rescues memory deficits and cholinergic dysfunction in a mouse model of Alzheimer's disease. *Neuron* 41, 27–33. doi: 10.1016/S0896-6273(03)00810-9
- Paxinos, G., and Watson, C. (2009). The Rat Brain in Stereotaxic Coordinates: Compact, 6th Edn. New York, NY: Academic Press.
- Pellow, S., Chopin, P., File, S. E., and Briley, M. (1985). Validation of open:closed arm entries in an elevated plus-maze as a measure of anxiety in the rat. *J. Neurosci. Methods* 14, 149–167. doi: 10.1016/0165-0270(85)90031-7
- Pinton, S., Da Rocha, J. T., Gai, B. M., and Nogueira, C. W. (2011). Sporadic dementia of Alzheimer's type induced by streptozotocin promotes anxiogenic behavior in mice. *Behav. Brain Res.* 223, 1–6. doi: 10.1016/j.bbr.2011.04.014
- Ramos, A. (2008). Animal models of anxiety: do I need multiple tests? Trends Pharmacol. Sci. 29, 493–498. doi: 10.1016/j.tips.2008.07.005
- Ramos, A., Mellerin, Y., Mormède, P., and Chaouloff, F. (1998). A genetic and multifactorial analysis of anxiety-related behaviours in Lewis and SHR intercrosses. *Behav. Brain Res.* 96, 195–205. doi: 10.1016/S0166-4328(98) 00023-0
- Savonenko, A. V., Xu, G. M., Price, D. L., Borchelt, D. R., and Markowska, A. L. (2003). Normal cognitive behavior in two distinct congenic lines of transgenic mice hyperexpressing mutant APP SWE. Neurobiol. Dis. 12, 194–211. doi: 10.1016/S0969-9961(02)00012-8
- Scahill, R. I., Schott, J. M., Stevens, J. M., Rossor, M. N., and Fox, N. C. (2002). Mapping the evolution of regional atrophy in Alzheimer's disease:

unbiased analysis of fluid-registered serial MRI. *Proc. Natl. Acad. Sci. U.S.A.* 99, 4703–4707. doi: 10.1073/pnas.052587399

- Schmitt, F. A., Davis, D. G., Wekstein, D. R., Smith, C. D., Ashford, J. W., and Markesbery, W. R. (2000). "Preclinical" AD revisited: neuropathology of cognitively normal older adults. *Neurology* 55, 370–376. doi: 10.1212/WNL.55.3.370
- Senanarong, V., Cummings, J. L., Fairbanks, L., Mega, M., Masterman, D. M., and O'Connor, S. M. (2004). Agitation in Alzheimer's disease is a manifestation of frontal lobe dysfunction. *Dement. Geriatr. Cogn. Disord.* 17, 14–20. doi: 10.1159/000074080
- Sierksma, A. S., Van Den Hove, D. L., Pfau, F., Philippens, M., Bruno, O., Fedele, E., et al. (2014). Improvement of spatial memory function in APPswe/PS1dE9 mice after chronic inhibition of phosphodiesterase type 4D. *Neuropharmacology* 77, 120–130. doi: 10.1016/j.neuropharm.2013.09.015
- Smith, G. E., Pankratz, V. S., Negash, S., Machulda, M. M., Petersen, R. C., Boeve, B. F., et al. (2007). A plateau in pre-Alzheimer memory decline: evidence for compensatory mechanisms? *Neurology* 69, 133–139. doi: 10.1212/01.wnl.0000265594.23511.16
- Sperling, R. A., Aisen, P. S., Beckett, L. A., Bennett, D. A., Craft, S., Fagan, A. M., et al. (2011). Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement. 7, 280–292. doi: 10.1016/j.jalz.2011.03.003
- Trullas, R., and Skolnick, P. (1993). Differences in fear motivated behaviors among inbred mouse strains. *Psychopharmacology (Berl.)* 111, 323–331. doi: 10.1007/BF02244948
- Tybout, A., Sternthal, B., Keppel, G., Verducci, J., Meyers-Levy, J., Barnes, J., et al. (2001). Analysis of Variance. J. Consum. Psychol. 10, 5–35. doi: 10.1207/S15327663JCP1001&2_03
- Vellas, B., Carrillo, M. C., Sampaio, C., Brashear, H. R., Siemers, E., Hampel, H., et al. (2013). Designing drug trials for Alzheimer's disease: what we have learned from the release of the phase III antibody trials: a report from the EU/US/CTAD Task Force. Alzheimers Dement. 9, 438–444. doi: 10.1016/j.jalz.2013.03.007
- Verma, P., Hellemans, K. G., Choi, F. Y., Yu, W., and Weinberg, J. (2010). Circadian phase and sex effects on depressive/anxiety-like behaviors and HPA axis responses to acute stress. *Physiol. Behav.* 99, 276–285. doi: 10.1016/j.physbeh.2009.11.002

- Vorhees, C. V., and Williams, M. T. (2006). Morris water maze: procedures for assessing spatial and related forms of learning and memory. *Nat. Protoc.* 1, 848–858. doi: 10.1038/nprot.2006.116
- Walsh, R. N., and Cummins, R. A. (1976). The Open-Field Test: a critical review. *Psychol. Bull.* 83, 482–504. doi: 10.1037/0033-2909. 83.3.482
- Webster, S. J., Bachstetter, A. D., Nelson, P. T., Schmitt, F. A., and Van Eldik, L. J. (2014). Using mice to model Alzheimer's dementia: an overview of the clinical disease and the preclinical behavioral changes in 10 mouse models. *Front. Genet.* 5:88. doi: 10.3389/fgene.2014.00088
- Whishaw, I. Q., Metz, G. A., Kolb, B., and Pellis, S. M. (2001). Accelerated nervous system development contributes to behavioral efficiency in the laboratory mouse: a behavioral review and theoretical proposal. *Dev. Psychobiol.* 39, 151–170. doi: 10.1002/dev.1041
- Wietrzych, M., Meziane, H., Sutter, A., Ghyselinck, N., Chapman, P. F., Chambon, P., et al. (2005). Working memory deficits in retinoid X receptor gamma-deficient mice. *Learn. Mem.* 12, 318–326. doi: 10.1101/lm. 89805

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 11 July 2014; accepted: 29 August 2014; published online: 16 September 2014. Citation: Galeano P, Martino Adami PV, Do Carmo S, Blanco E, Rotondaro C, Capani F, Castaño EM, Cuello AC and Morelli L (2014) Longitudinal analysis of the behavioral phenotype in a novel transgenic rat model of early stages of Alzheimer's disease. Front. Behav. Neurosci. 8:321. doi: 10.3389/fnbeh.2014.00321

This article was submitted to the journal Frontiers in Behavioral Neuroscience.

Copyright © 2014 Galeano, Martino Adami, Do Carmo, Blanco, Rotondaro, Capani,
Castaño, Cuello and Morelli. This is an open-access article distributed under the terms
of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are
credited and that the original publication in this journal is cited, in accordance with
accepted academic practice. No use, distribution or reproduction is permitted which
does not comply with these terms.