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REVIEW

Macrophage activation syndrome as part of systemic juvenile idiopathic arthritis: diagnosis, genetics, pathophysiology and treatment

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Macrophage activation syndrome (MAS) is a severe, frequently fatal complication of systemic juvenile idiopathic arthritis (sJIA) with features of hemophagocytosis leading to coagulopathy, pancytopenia, and liver and central nervous system dysfunction. MAS is overt in 10% of children with sJIA but occurs subclinically in another 30–40%. It is difficult to distinguish sJIA disease flare from MAS. Development of criteria for establishing MAS as part of sJIA are under way and will hopefully prove sensitive and specific. Mutations in cytolytic pathway genes are increasingly being recognized in children who develop MAS as part of sJIA. Identification of these mutations may someday assist in MAS diagnosis. Defects in cytolytic genes have provided murine models of MAS to study pathophysiology and treatment. Recently, the first mouse model of MAS not requiring infection but rather dependent on repeated stimulation through Toll-like receptors was reported. This provides a model of MAS that may more accurately reflect MAS pathology in the setting of autoinflammation or autoimmunity. This model confirms the importance of a balance between pro- and anti-inflammatory cytokines. There has been remarkable progress in the use of anti-pro-inflammatory cytokine therapy, particularly against interleukin-1, in the treatment of secondary forms of MAS, such as in sJIA.

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INTRODUCTION

Macrophage activation syndrome (MAS) is a serious, potentially fatal complication of rheumatic diseases, which is seen most frequently in systemic juvenile idiopathic arthritis (sJIA) and in its adult equivalent, adult-onset Still disease,¹⁻³ although it is increasingly reported in other pediatric inflammatory disorders, namely juvenile systemic lupus erythematosus⁴ and Kawasaki disease.⁵⁻⁷ In recent years, this condition has also been observed in periodic fever syndromes.^{8,9} MAS may occur spontaneously, as a complication of active underlying disease, or may be triggered by an infection, a change in drug therapy or a toxic effect of a medication, including biologics.¹⁰ Clinically, patients with MAS present with non-remitting high fever, pancytopenia, hepatosplenomegaly, hepatic dysfunction, encephalopathy, coagulation abnormalities and sharply increased levels of ferritin (Table 1). The pathognomonic feature of the syndrome is seen on bone marrow examination, which reveals numerous morphologically benign macrophages exhibiting hemophagocytic activity (Figure 1). Such cells may also be found in lymph nodes and spleen, but they may infiltrate almost any organ in the body and may account for many of the systemic features of the syndrome. As MAS bears a close resemblance to a group of histiocytic disorders collectively known as hemophagocytic lymphohistiocytosis (HLH), it is currently classified among the secondary, or acquired, forms of HLH. $^{11,12}\,$

DIAGNOSTIC CRITERIA FOR MAS IN sJIA

MAS is a severe condition that can pursue a rapidly fatal course. Prompt recognition of its clinical and laboratory features and immediate therapeutic intervention are, therefore, imperative. However, diagnosis of MAS can be difficult and hard to distinguish from sepsis-like syndromes, although it may also be associated with sepsis, especially in cases of intraphagocytic pathogen infections. In addition, the recognition of subclinical forms of MAS in sJIA^{13,14} underscores the importance of establishing criteria sensitive enough to distinguish MAS from routine disease flare. Differentiation of MAS from these conditions is critical to select the appropriate therapeutic approach. The difficulties in making the diagnosis and recent therapeutic advances (see below) emphasize the need of diagnostic tools and wellestablished diagnostic guidelines. Diagnostic criteria will also be important for research purposes and use in literature reports.

The recognition that MAS is clinically similar to HLH has led some to propose the use of the HLH diagnostic guidelines for establishing a diagnosis of MAS. However, HLH criteria developed primarily for homozygous genetic disorders leading to hemopha-gocytosis¹⁵ are not necessarily of timely use in defining/ identifying MAS in the setting of sJIA. The main shortcoming of HLH criteria in MAS is due to the fact that certain criteria may not apply to patients with sJIA. Owing to the prominent inflammatory expression of the latter disease, the occurrence of a relative

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 Table 1. Main clinical, laboratory and pathological features

 of macrophage activation syndrome

Clinical features Non-remitting high fever Hepatomegaly Splenomegaly Lymphadenopathy Hemorrhages Central nervous system dysfunction

Laboratory features Cytopenia Abnormal liver function tests Coagulopathy Decreased erythrocyte sedimentation rate Hypertriglyceridemia Hypoalbuminemia Hypoefferritinemia Elevated sCD25 and sCD163

Histopathological features

Macrophage hemophagocytosis in the bone marrow Increased CD163 staining of the bone marrow

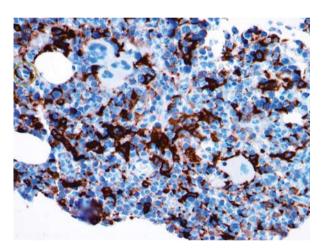


Figure 1. CD163 staining of histiocytes/macrophages in the bone marrow of a patient with MAS. A bone marrow biopsy from a child with MAS was stained with anti-CD163 (haptoglobin receptor) antibody and analyzed under high power magnification. Numerous activated histiocytes/macrophages are present revealing hemophagocytic activity. A particularly phagoctyic histiocyte is circled in green in the upper left hand corner of the figure.

decrease in white blood cell count, platelets or fibrinogen, rather than the absolute decrease required by the HLH criteria, may be more relevant in making an early diagnosis. Indeed, sJIA patients often have increased white blood cell and platelet counts as well as increased serum levels of fibrinogen as part of the inflammatory response seen in this disease. Therefore, when these patients develop MAS, they may only reach the degree of cytopenia and hypofibrinogenemia seen in HLH at the late stages of the syndrome, when their management becomes challenging. Another problem is that the minimum threshold level for hyperferritinemia required for the diagnosis of HLH (500 μ g I⁻¹) is not suitable to detect MAS in children with sJIA. It is well known that many patients with active sJIA, in the absence of MAS, have ferritin levels above that threshold.¹⁶ In the acute phase of MAS, ferritin levels generally peak to more than 5000 μ g I⁻¹. Thus, use of

a 500- μ g l⁻¹ threshold may not help to discriminate MAS from a flare of sJIA. Other HLH criteria that are not readily applicable to MAS are the demonstration of low or absent natural killer cell activity or soluble interleukin-2 (IL-2) receptor alpha chain (CD25) above normal limits for age, as these tests are not routinely performed in pediatric rheumatology settings. We recently found that the diagnostic guidelines for HLH were highly specific in patients with sJIA-associated MAS, but lacked sensitivity (manuscript in preparation).

Previously, preliminary diagnostic guidelines for MAS complicating sJIA were published.¹⁷ Laboratory criteria include decreased platelet count ($\leq 262 \times 10^9 l^{-1}$), elevated levels of aspartate aminotransferase ($>59UI^{-1}$), decreased white blood cell count $(\leq 4.0 \times 10^9 l^{-1})$ and hypofibrinogenemia $(\leq 2.5 g l^{-1})$. Clinical criteria include hepatomegaly, hemorrhagic manifestations and central nervous system dysfunction. The diagnosis of MAS requires the presence of any two or more laboratory criteria, or any two or three or more clinical and laboratory criteria. The demonstration of macrophage hemophagocytosis in the bone marrow aspirate is required only in doubtful cases. As noted in patients with HLH¹⁸ and in several instances of MAS,¹⁷ the bone marrow aspirate does not always show hemophagocytosis, and, furthermore, hemophagocytosis is not always demonstrable in the initial stages of the disease. Repeat bone marrow aspirate over time may eventually demonstrate hemophagocytosis. These criteria have the advantage of being data-driven and not merely based on expert consensus. However, the study underlying their development has several limitations, including the lack of several laboratory measurements in a number of patients and insufficient data for some of the laboratory parameters evaluated. Moreover, the criteria have yet to be validated.

In recent years, an international collaborative effort was started, which is aimed at developing a new and robust set of diagnostic criteria for MAS complicating sJIA, based on the combination of expert consensus and the analysis of real patient data. The first step of the project, with a specific aim to identify candidate items using international consensus formation through the Delphi survey technique, has just been accomplished.¹⁹ A total of 505 pediatric rheumatologists belonging to three large networks were sent a questionnaire that listed 28 clinical, laboratory and histopathological features that were thought most likely to be helpful and relevant in the diagnosis of MAS complicating sJIA indentified through literature review. Respondents were asked to select the 10 features that they felt were most important and to rank-order the selected features by assigning 10 to the most important one, and end with 1 as the least important. The following nine features were selected by more than 50% of the 232 respondents and were given most frequently the highest ranks: falling platelet count, hyperferritinemia, evidence of macrophage hemophagocytosis in the bone marrow, increased liver enzymes, falling leukocyte count, persistent continuous fever \geq 38 °C, falling ESR, hypofibrinogenemia and hypertriglyceridemia. These features may be the best candidates to be part of the final set of diagnostic criteria for the syndrome.

Questionnaire respondents were intentionally not asked to indicate the threshold level for each laboratory test that they felt was optimal for early identification of MAS. This objective is intended to be pursued in the subsequent phase of the project, which is under way and is aimed at collecting real data from patients with sJIA-associated MAS and patients with conditions that may be confused with MAS, including sJIA flare without MAS and febrile systemic infection controls. This process is also meant to enable a data-driven assessment of the relative sensitivity and specificity of clinical, laboratory and histopathological features in discriminating MAS from the confusable conditions. Notably, the data collection is structured is such a way that it may establish whether laboratory criteria for the syndrome are better assessed in terms of absolute threshold values, or percentage change over the



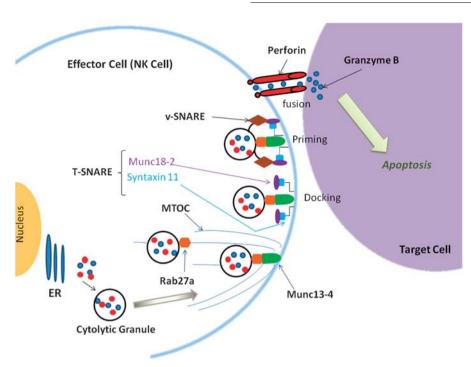


Figure 2. Cytolytic pathway proteins mutated in MAS. A diagram of the immunologic synapse between a cytolytic (to the left) lymphocyte and an APC (to the right) is portrayed. Proteins involved in the cytolytic pathway that can be found mutated in MAS and HLH include Rab27a, Munc13-4, Syntaxin 11, Munc18-2 and perforin.

preceding days or both. The ultimate goal of the project is to develop a core set of criteria for the diagnosis MAS in sJIA that is timely, feasible and both highly sensitive and specific.

GENETIC FACTORS FOR MAS IN sJIA

The etiology of MAS remains elusive. However, MAS bears strong resemblance to a group of histiocytic disorders collectively known as HLH.^{11,20} HLH is a term that describes a spectrum of disease processes characterized by the accumulations of well-differentiated mononuclear cells with a macrophage phenotype exhibiting hemophagocytic activity. HLH is further subdivided into primary or familial (FHLH) and secondary or reactive (ReHLH).^{21,22} FHLH is a constellation of rare autosomal recessive immune disorders resulting from homozygous deficiency in cytolytic pathway proteins.¹⁵ Its clinical symptoms usually become evident within the first 2 months of life. ReHLH tends to occur in older children and more often is associated with an identifiable infectious episode or autoimmune condition.²¹ However, distinctions between primary and secondary HLH are becoming increasingly blurred as new genetic causes are identified, some of which are associated with less severe and somewhat distinct clinical presentations occurring later in life.²³ Some of these may present later in life due to heterozygous or compound heterozygous mutations in cytolytic pathway genes that confer a partial dominant negative effect on cytolysis.

The pathological mechanisms of HLH are not fully understood. In primary HLH, the uncontrolled expansion of T cells and macrophages has been linked to decreased NK cell and cytotoxic T-cell function^{21,24} due to mutations in various genes whose products are involved in the cytolytic pathway.^{21,25-27} The cytotoxic activity of these cells is mediated by the release of specialized cytotoxic granules that contain several classes of proteins expressed only in cytotoxic cells, including perforin and granzymes. Once cytotoxic cells are activated, these granules are delivered to the surface of the cells and the contents are released at the immunologic synapse with the target cell (Figure 2). Perforin aids in delivering of the contents of the granules into the cytoplasm of the target cell, while granzymes trigger apoptosis once in the cytoplasm of the target cell. In 15-40% of patients with FHLH, cytolytic dysfunction is due to mutations in the gene encoding perforin.²⁶ Mutations in another gene, MUNC13-4, have been implicated in the development of hemophagocytosis in about 10-30% of patients with inherited HLH.²⁵ The protein encoded by the MUNC13-4 gene is important for docking and fusion of the cytotoxic granules with the cytoplasmic membrane (Figure 2). Although the cytolytic cells of the patients with FHLH caused by MUNC13-4 mutations produce sufficient amounts of perforin, the poor ability to deliver the content of the cytolytic granules to the immunologic synapse with the target cell leads to profoundly decreased cytolytic activity. More recently, mutations in two other genes encoding proteins that facilitate granule fusion in intracellular trafficking events have been linked to the development of primary HLH: Syntaxin 11, a member of the SNARE protein family,²⁸ and syntaxin binding protein 2 (STXBP2, also known as MUNC18-2)²⁷ (Figure 2).

Defects in the granule-dependent cytotoxic functions of lymphocytes have also been implicated in two other genetic diseases associated with the hemophagocytic syndrome. Thus, mutations in the gene encoding Rab27a, one of the MUNC13-4 effector molecules (Figure 2), have been linked to the development of Griscelli syndrome type 2.²⁹ Mutations in the *Lyst* gene have been identified as a cause of Chediak–Higashi syndrome. Both disorders can be complicated by the development of HLH.³⁰

HLH following exposure to Epstein-Barr virus (EBV) and occasionally other viruses, termed fulminant infectious mononucleosis, is the most frequent life-threatening complication of X-linked lymphoproliferative syndrome (XLP). XLP1 is caused by hemizygous mutations in the *SH2D1A* gene encoding SAP (SLAMassociated protein), which leads to abnormal NK cell responses and invariant NKT cell deficiency.³¹ XLP2 is caused by mutations in *BIRC4*, which encodes XIAP, and has been described as an X-linked form of FHLH.³² Recent observations suggest that lymphocytes from patients with both types of XLP demonstrate decreased activation-induced apoptosis that contributes to the uncontrolled lymphoproliferation. Taken all together, these genetic disorders still account for less than half of the diagnosed cases of HLH in children, including many familial cases still awaiting molecular definition.²¹

The presence of the defects in the granule-dependent cytotoxic activity of lymphocytes in several diseases associated with hemophagocytic syndromes highlights the importance of this function in restraining some inflammatory responses.33-36 The exact mechanisms that link deficient NK cell and cytotoxic T-cell functions with expansion of activated macrophages are not clear. One possible explanation is related to the fact that poor cytolytic activity seen in HLH patients may lead to diminished ability to control some infections. More specifically, NK cells and cytotoxic T lymphocytes fail to kill infected cells and, thus, to remove the antigen-presenting cell source of antigenic stimulation. Such persistent antigen stimulation leads, in turn, to persistent antigendriven activation and proliferation of T cells associated with escalating production of cytokines that stimulate macrophages. It has also been shown that abnormal cytotoxic cells may fail to provide appropriate apoptotic signals for removal of activated macrophages and T cells during the contraction stage of some immune responses leading to persistent expansion of T cells and macrophages secreting proinflammatory cytokines.34,35,37 As a result of continuous stimulation with proinflammatory cytokines (most notably, interferon gamma (IFN γ)), macrophages will become hemophagocytic.^{33,38}

Although familial cases of MAS in sJIA have not been reported, as in FHLH, sJIA/MAS patients may also have functional defects in the exosome degranulation pathway.³⁹⁻⁴¹ Furthermore, these functional abnormalities are associated with SNPs in the FHLHassociated genes, including PRF1⁴⁰ and MUNC13-4.^{42,43} In addition, sJIA/MAS patients with bi-allelic mutations in the MUNC13-4 gene reported in FHLH have been described,⁴³ suggesting that there is likely a genetic component that overlaps between MAS and FHLH. Interestingly, mutations in MUNC13-4 also affect the degranulation of neutrophils⁴⁴ and platelets.⁴⁵ Not surprisingly, FHLH patients with bi-allelic mutations in MUNC13-4 often have atypical clinical features with some overlap with sJIA. Preliminary sequencing of the FHLH-associated genes in a small cohort of sJIA/MAS patients identified new sequence variants, as well as known heterozygous mutations, in several genes, most notably MUNC13-4, STXBP2 and BIRC4 (Grom, Cron unpublished observations). The exact prevalence of such sequence variants and their impact on the cytolytic function of CD8 T or NK cells and ultimately on the clinical phenotype still need to be clarified. One possibility is that these heterozygous mutations still affect the cytolytic function through a gene dosage effect. Alternatively, the defects may be in multiple genes. Consistent with this idea, FHLH in patients who are heterozygous for the A91V perforin variation is often associated with other heterozygous genetic defects.⁴⁶

Another recent study from Japan identified *IRF5* (interferon regulatory factor 5) gene polymorphisms as risk factors for MAS development in patients with sJIA.⁴⁷ Given the presumed role that IFN γ has in the pathophysiology of MAS (see further), this observation is very intriguing and needs to be confirmed in other ethnic groups. Thus, the genetics of MAS in children with sJIA likely shares many of the same defects as in infants who present with FHLH.

PROPOSED PATHOPHYSIOLOGY OF MAS IN CHILDREN WITH sJIA

Because of the rarity of MAS and the difficulty in performing mechanistic human immunology studies, much of our knowledge regarding the immunologic pathoetiology of MAS has relied on

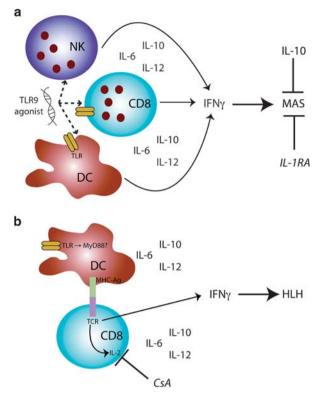


Figure 3. Proposed pathophysiology contributing to MAS/HLH and associated cytokine storm. (**a**) MAS and (**b**) HLH result from the inability of a CD8 T cell to lyse an APC due to cytolytic pathway defects resulting in sustained immune cell activation and a cytokine storm of pro-inflammatory cytokines, including IFN γ . Blockade of IL-1 (with IL-1RA, anakinra) can help control the cytokine storm, and IL-10 expression can help prevent hemophagocytosis in a model of MAS. CsA can also help prevent the cytokine storm in models of MAS/HLH. CD8, CD8 T cells; DC, dendritic cell; MHC-Ag, major histocompatibility complex coupled to antigen; NK, natural killer cell; TCR, T-cell receptor; TLR, Toll-like receptor.

animal studies. Initially, animal models of primary HLH were used to dissect the mechanisms behind this related disease. Jordan *et al.*³³ described the first such model, a mouse deficient in the gene for perforin that developed a primary HLH syndrome upon infection with lymphochoriomeningitic virus. Depletion or neutralization of various cytokines and cellular populations revealed a critical role for both IFN γ and CD8 T cells in the development of disease. These results lead to the proposition of a model whereby defects in cytotoxic granule function prevent the ability of immune cells to clear infected, activated antigen-presenting cells. The continued presence of these antigen-presenting cells results in the repeated, uninterrupted stimulation of CD8 T cells and in turn, the production of IFN γ . The accumulation of large amounts of IFN γ in this process leads to cytokine toxicity, and the various elements of the disease (Figure 3).

These results, and the predictions made by this model, have subsequently been confirmed in other mouse models that are defective in many of the other human genes associated with primary HLH. Rab27a is important in docking the perforin laden vesicle at the cell membrane for release.⁴⁸ Munc13-4 is likely important in activating Syntaxin 11 that in turn is important for vesicle fusion.^{25,28} Absence of any these molecules prevents the release of perforin and therefore destruction of target antigenpresenting cells (Figure 2). Accordingly, mice deficient in the genes encoding Munc13-4⁴⁹ and Rab27a^{50,51} also develop HLH upon infection with lymphochoriomeningitic virus in an IFN_γ-dependent manner.

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In many instances of sJIA/MAS, no such defects in cytotoxic cell function have been identified, or have only variable penetrance. This has lead to a search for alternate mechanisms to explain the origin of the syndrome in this context as the above described model does not account for these situations. Recently, a new murine model of MAS induced by repeated stimulation of Toll-like receptor (TLR) 9 may provide some insight.⁵² The rationale for this model involves multiple observations. Gene signatures consistent with chronic TLR/IL-1 β signaling are present in sJIA patients. 53 Lupus, another rheumatic condition associated with MAS,⁴ has also long been associated with hyperactive TLR9 function.⁵⁴ EBV. perhaps the most common infectious trigger of secondary HLH, is a DNA virus that triggers TLR9.55 Furthermore, as opposed to patients with EBV-mononucleosis, patients with EBV-HLH have very high titers of viral particles in the blood,⁵⁶ meaning that there is an excess of TLR9 ligand present in these patients. As described above, polymorphisms of the TLR9 signaling molecule IRF5 that result in the hyperactivation have been strongly associated with sJIA-related MAS.⁴⁷ Taken together, these observations suggest that situations of repeated activation of TLR9 will replicate the environment that allows MAS to develop in the susceptible genetically predisposed host.

Indeed, mice given repeated TLR9 stimulation develop many of the features of MAS. Furthermore, the disease in this model is also IFN γ dependent, drawing a connection back to primary HLH⁵² (Figure 3). Unlike primary HLH, this IFN γ is arising from different sources than CD8 T cells, including dendritic cells and NK cells. These results suggest that pathogenic IFN γ can arise from multiple initial immunologic insults, and, that while the end stage clinical syndrome may be similar, the origin of the hypercytokinemia may be different in different disease settings. Another observation made from these studies was the important regulatory role of IL-10 in controlling disease. Mice given repeated TLR9 stimulation concordant with blockade of the IL-10 receptor developed much more fulminant disease. This is consistent with human data where gene signatures of IL-10 responses are prominent in patients with HLH and sJIA/MAS, suggesting the importance of IL-10 in the immune response.⁵⁷ Intriguingly, polymorphisms of IL-10 associated with decreased function of IL-10 are associated with sJIA.58,59 It is possible that the combined immunologic insult of hyperactive TLR/IL-1 β signaling in combination with decreased IL-10 function may result in a predisposition to MAS. It will be of interest to explore the combined haplotype of IRF5 and IL-10 gene polymorphisms to see if this provides increased power to identify at risk patients. It is also possible that IL-10 function serves to explain the phenomenon of occult or subclinical MAS, in that these patients may have higher IL-10 function than those developing fulminant MAS. This increase in IL-10 function would be able to maintain their state of relative compensation (Figure 3). Interestingly, some of these ideas may apply to primary HLH as well, as a recent report shows the critical need for the TLR signaling adaptor MyD88 in the development of disease in lymphochoriomeningitic virus-infected MUNC13-4-deficient mice.60

The immunological mechanisms behind other animal models of HLH/MAS are less well defined. Infection of rabbits with Herpesvirus papio results in a clinical syndrome similar to EBV-HLH.^{61,62} These animals develop anti-red blood cell antibodies, which were suggested to contribute to hemophagocytosis. This is unlike other models and human disease, where anti-red blood cell antibodies are usually not found, making this model difficult to interpret. Mice deficient in asparaginyl endopeptidase (AEP), an enzyme important for lysosomal degradation, were also reported to develop an HLH-like syndrome including fever, cytopenia, splenomegaly and erythrophagocytosis.⁶³ Different from human disease, the phagocytozing histiocytes in this model were only consuming red blood cells and not other hematopoetic cells. This was likely due to the fact that these red blood cells had deformed

membranes as a result of the *AEP* deficiency. Additionally different from human disease, these mice do not display a hypercytokinemia. Thus, the *AEP*-deficient mice may represent a distinct pathologic process from MAS, perhaps more similar to a lysosomal storage disease. As in humans, mice deficient in the gene encoding *SAP* develop hypogammaglobulinemia and are susceptible to HLH secondary to herpesvirus infections.⁶⁴ The precise immune mechanisms that lead to HLH in *SAP* deficiency are not clear. The most obvious link is that *SAP*-deficient NK cells also have impaired cytotoxicity.⁶⁵ Infection of Sv12956 mice with *Salmonella enterica* serotype Typhimurium led to a syndrome of secondary HLH consistent with human disease.⁶⁶ This model has not yet been carefully immunologically dissected to provide insight into the mechanisms of disease.

There is much for room for continued exploration of the immunology underlying MAS. What is the source and nature of the IL-10 response? Almost every immune cell in the mouse has been described to produce IL-10 under the correct stimulus. T cells themselves might be a source of the IL-10, but this might be surprising under the very strong Th1 milieu of HLH/MAS. Recently, B cells have been noted to be an important source of regulatory IL-10.⁶⁷ A role for these cells in HLH/MAS might provide novel targets for therapeutic options. Myeloid derived IL-10, from dendritic cells and macrophages may also have a role in regulating the potentially hyperactive antigen presentation process that drives HLH.³⁵

Although CD8 T cells are clearly the source of the pathogenic IFN γ in perforin-deficient HLH, the IFN γ producing cell is less clear in other forms. Which cells are producing IFN γ in EBV-HLH or sJIAassociated MAS remain untested. In the TLR9-induced mouse model of MAS, a mixed population of dendritic cells, T cells and NK cells produced IFN γ . Ultimately, as the identity of the IFN γ producing cell is likely to be different depending on the initiating stimulus, the producer may be less important than the cell upon which the pathogenic IFN γ is acting. It has been recently reported that IFN γ is sufficient to induce hemophagocytes, and that the $\text{IFN}\gamma$ receptor is required on macrophages for this effect in a STAT1-dependent manner.⁶⁸ Whether the IFN γ responding macrophage becomes a hemophagocyte, or whether it induces another cell type to become a hemophagocyte is unclear. Furthermore, although the presence of hemophagocytosis is correlated with development of anemia⁶⁸ and worsened disease activity,⁵² there is of yet no direct demonstration that hemophagocytes are directly pathogenic. In fact, some authors have found potentially anti-inflammatory/regulatory properties for hemophagocytes.⁶⁹ Certainly CD163, highly expressed on human hemophagocytes, is a marker strongly associated with regulatory M2 macrophage differentiation. Better characterization of the function of the hemophagocyte is clearly needed to understand these syndromes.

What other cytokines are having a role? What other immunologic insults can lead to a similar hyper-IFN γ state? How can we modulate the IFN γ and IL-10 axes to treat disease? Answers to these questions have relevance not only for MAS but also for 'cytokine storm' syndromes in general, such as sepsis or systemic inflammatory response syndrome. Continued work in animal models, as well as in translating these findings back to the human syndromes both remain important goals.

TRADITIONAL THERAPY FOR MAS IN THE SETTING OF SJIA

Although animal models of MAS may lead to novel and improved therapies, treatment of MAS as part of sJIA has traditionally developed from anecdotal experience. The first reported therapy, and still used today, for MAS as part of sJIA is high dose corticosteroids. Hadchouel *et al.*⁷⁰ in France described seven children with JIA (six with sJIA) complicated by acute hemorrhagic, hepatic and neurologic manifestations (later recognized as MAS).

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The seven JIA patients were treated with high dose steroids and five survived.⁷⁰ A similar benefit was reported for high dose steroids in nine children with MAS (seven with sJIA) in England.³ More recently, glucocorticoids, along with other immunosuppressive therapy, were reported to be effective in 13 children with sJIA in China⁷¹ and in 6 children with sJIA in India.⁷² Hence, high dose steroids are routinely used to treat MAS as part of sJIA.

In addition to corticosteroids, cyclosporine A (CsA) has become a staple in the therapy of MAS as a complication of sJIA. After some initial cases reported in the mid-1990's⁷³⁻⁷⁵ in 2001, a report from France highlighted the utility of treating 24 patients (18 with sJIA) with MAS using CsA in pulse steroid failures or in 5 patients as a first line therapy.⁷⁶ Similarly, the protocol for treating primary or FHLH employs both corticosteroids (although at much lower doses) and CsA as part of the treatment protocol.¹⁵ CsA is thought to preferentially target lymphocytes by inhibiting the NFAT family of transcription factors that are critical for the activation of a wide array of cytokine genes.⁷⁷ One of the likely benefits of CsA is dampening of the cytokine storm that occurs during MAS,³⁷ and, thus, lymphocyte-targeted immunosuppression is a key component of treating MAS as part of sJIA.

Another relatively lymphocyte specific therapy that has been explored for treating severe sJIA and MAS is cyclophosphamide. Anecdotal success has been reported with this chemotherapeutic agent^{3,78} but it has not received a lot of attention of late for treating MAS as part of sJIA. Another chemotherapeutic agent used to treat MAS is etoposide, which is part of the protocol developed for treating FHLH.¹⁵ This protocol carries a not insignificant risk of mortality both pre- and post-bone marrow transplantation, and may not be appropriate as first-line therapy for MAS as part of sJIA.⁷⁹ Autologous hematopoetic stem cell transplantation has been used specifically for treating cases of severe sJIA,⁸⁰ but the occurrence of several fatal complications due to MAS has suggested better control of systemic disease before transplantation is necessary.⁸¹ Whether or not a less aggressive use of etoposide for treating secondary forms of MAS, such as those complicating sJIA, will be beneficial, remains unclear. Meanwhile, the advent and use of a variety of biologic agents is beginning to replace cytotoxic agents as first-line therapy for secondary forms of HLH/MAS.

One of the first biologic agents to be used for MAS is intravenous immunoglobulin. Success of intravenous immunoglobulin requires treatment early in the course of MAS.⁸² Although often useful for the treatment of infection-triggered MAS^{83,84} and a few children with sJIA,^{72,85} children with MAS as part of sJIA are often refractory to intravenous immunoglobulin.⁸⁶ A more aggressive approach using anti-thymocyte globulin has been proposed for FHLH⁸⁷ and has been used successfully in two patients with probable MAS.⁸⁸ Nevertheless, there is a significant risk of serious infection and mortality with anti-thymocyte globulin use,⁸⁷ and less aggressive but effective biologic therapies have found recent success in treating MAS as part of sJIA.

A relative explosion of novel biologic therapies is now available for the treatment of rheumatic disorders. Rituximab, a B-cell depleting anti-CD20 antibody developed for the treatment of B-cell lymphoma, has found a variety of indications to treat rheumatic disorders ranging from arthritis to lupus to vasculitis and beyond.⁸⁹ Rituximab has also recently been reported to achieve remission in a high percentage of children with refractory sJIA.⁹⁰ In addition, rituximab has anecdotally been used to effectively treat EBV-associated HLH/MAS in the setting of EBV infection.^{91,92} To date, no cases of rituximab therapy for MAS associated with sJIA have been reported. However, anti-cytokine therapies are finding a niche in the treatment of MAS as part of sJIA.

The first cytokine inhibitors used to treat MAS as part of sJIA were TNF blockers. Initially, there was some excitement as TNF inhibition was reported to effectively treat many cases of MAS

including several children with sJIA.⁹³⁻¹⁰¹ This early enthusiasm has been tempered by the notion that TNF inhibitors may trigger MAS in some instances.¹⁰²⁻¹⁰⁹ Although cause and effect is certainly difficult to formally prove in these circumstances, the fact that MAS can develop in the setting of TNF inhibition is concerning.^{110,111} While TNF inhibition may not be the ideal therapy for MAS, particularly in the setting of a child with sJIA, therapy directed at two other pro-inflammatory cytokines, IL-1 and IL-6, seems promising.

IL-1 BLOCKADE FOR THE TREATMENT OF MAS IN CHILDREN WITH sJIA

The first reported use of the IL-1 receptor antagonist, anakinra, to purposely target MAS was in a critically ill child with a severe inflammatory condition and resulted in a remarkable improve-ment in a relatively brief time frame.¹¹² Although not formally defined as having MAS, two severely ill children with sJIA also received substantial benefit from anakinra treatment.¹¹³ Soon after, anakinra was shown to markedly improve both systemic and arthritic features in a cohort of conventional treatment refractory sJIA patients.¹¹⁴ Two independent groups concluded that, in addition to the \sim 10% risk of developing overt MAS as part of sJIA, another 30-40% of sJIA patients may have occult or subclinical MAS during sJIA disease flare that can eventually lead to overt MAS.^{13,14} This, along with the similarity of many clinical and laboratory features shared by MAS and sJIA flare, led to the concept that MAS may be an inherent aspect of sJIA pathophysiology in a large percentage of patients. The fact that anakinra was shown to be highly effective for sJIA^{114,115} suggested anakinra would also be a valuable treatment for MAS as part of sJIA.

Several cases of sJIA-associated MAS dramatically benefiting from anakinra after inadequate response to corticosteroids and CsA have now been reported^{86,115-118} (Table 2). For those severely ill children, IL-1 blockade has been remarkably effective in a relative brief time frame. Similarly, treatment refractory adult Still disease (likely related or identical to sJIA but in adults) with MAS has been described as having miraculously responded to anakinra therapy by several groups.¹¹⁹⁻¹²⁸ Although anakinra appears to be extremely safe, as a recombinant human protein with a short half-life (\sim 3 hours),¹²⁹ and a large therapeutic window (1-48 mg kg⁻¹ per day),^{118,130} there has been a report of hepatitis attributed to anakinra in children with sJIA.¹³¹ Moreover, there has also been the suggestion that anakinra triggered MAS in two children with sJIA^{132,133} but once again, cause and effect is difficult to establish. In a large case series of 46 sJIA patients treated with anakinra at disease onset, anakinra was a potential MAS trigger in five children at doses of $1-2 \text{ mg kg}^{-1}$ per day.¹¹⁵ However, dose escalation of anakinra often seemed to help control MAS, and permanent discontinuation of anakinra was unnecessary for any of the children.¹¹⁵ The overall published experience for the use of IL-1 blockade therapy for the treatment of refractory MAS as part of sJIA has been highly favorable (Table 2).

Like IL-1 inhibition, IL-6 blockade, via an anti-IL-6 receptor monoclonal antibody (tocilizumab), has proven highly efficacious in treating sJIA.¹³⁴ Whether tocilizumab will be similarly helpful in treating MAS in sJIA remains unclear at present, as there has been a case of MAS attributed to IL-6 blockade.¹³⁵ However, because many children with sJIA flare of disease are in a state of MAS^{13,14} and respond favorably to IL-6 blockade, it is likely that blocking the actions of this cytokine will also benefit MAS in children with sJIA. Co-stimulatory blockade with CTLA-4-Ig has been anecdotally beneficial in children with severe sJIA¹¹⁸ but its place in treating MAS is unknown. Nevertheless, there is building evidence that biologic therapies, particularly IL-1 inhibitors, are a welcome addition to corticosteroids and CsA in treating MAS associated with sJIA.⁸⁶ Time will tell if novel biologic therapies in the pipeline also hold promise for the treatment of MAS in children with sJIA.

2	9	5

Patient no.	Age (yrs) ^a	Sex	Dx	Immuno-suppression	Anakinra dose (mg kg ⁻¹ per d)	Ferritin (ng ml ⁻¹) ^b	Outcome of MAS	First Author	Reference no.
1	13	F	sJIA	St, CsA, IVIg	1	132 206	Resolved	Kelly	117
2	17	М	sJIA	St, CsA, IVIg	2	4787	Resolved	Miettunen	86
3	9	F	sJIA	St, CsA, IVIg	2	2279	Resolved	Miettunen	86
4	12	М	sJIA	St, CsA, IVIg	2	3141	Resolved	Miettunen	86
5	8	F	sJIA	St, CsA, IVIg	2	423	Resolved	Miettunen	86
6	1	F	sJIA	St, CsA, IVIg	2	438	Resolved	Miettunen	86
7	15	М	sJIA	St, CsA, IVIg	2	1055	Resolved	Miettunen	86
8	13	М	sJIA	St, CsA, IVIg	2	1285	Resolved	Miettunen	86
9	8	F	sJIA	St, CsA, Et	2	>10000	Resolved	Miettunen	86
10	4	F	sJIA	St, CsA, MTX, Ab	11	869	Resolved	Record	118
11	10	F	sJIA	St, CsA, MTX, Ab, Cyc, IVIg	9	15 693	Resolved	Record	118
12	8	М	sJIA	St	2	19674	Resolved	Bruck	116
13	12	F	sJIA	St	2	28 000	Resolved	Bruck	116

Abbreviations: Ab, abatacept; CsA, cyclosporine A; Cyc, cyclosphophamide; Dx, diagnosis; Et, etoposide; F, female; IVIg, intravenous immunoglobulin; M, male; MAS, macrophage activation syndrome; MTX, methotrexate; sJIA, systemic juvenile idiopathic arthritis; St, steroids. ^aAge when MAS treated with anakinra. ^bLevel before anakinra treatment.

SUMMARY

MAS is becoming an increasingly recognized, sometimes fatal, complication of sJIA disease flare. Because of the urgency in making the diagnosis of MAS and instituting appropriate therapy in a timely fashion, efforts are ongoing to establish timely, practical, and highly sensitive and specific criteria for recognizing MAS in children with sJIA. Progress is also being made in understanding the genetics and pathophysiology of MAS in children with sJIA. Preliminary studies suggest that children with sJIA and MAS frequently possess heterozygous mutations/ polymorphisms in cytolytic pathway genes, which are disrupted in a homozygous fashion in infants with the related condition, FHLH. Knocking out these genes in mice has provided a tool for studying infection triggered MAS/HLH, and recently a novel murine model of non-infectious triggered MAS, which is dependent on repeated TLR stimulation, is imparting added insights into the pro- and anti-inflammatory milieu of the MAS cytokine storm. Understanding the balance of cytokines in the cytokine storm of MAS is crucial to the efforts in developing and utilizing the available cytokine-targeted biologic therapies. The rapid and dramatic benefits seen with the use of the recombinant human protein, anakinra, is generating substantial enthusiasm for treating MAS in the setting of sJIA with IL-1 blockade. Future therapeutic protocols for the treatment of MAS as part of sJIA will likely include a combination of high dose corticosteroids, CsA and anti-pro-inflammatory cytokine treatment, such as blockade of IL-1.

CONFLICT OF INTEREST

Drs Cron and Behrens who serve as consultants to Genentech for the use of Actemra (tocilizumab) to treat sJIA. Dr Cron also serves as a consultant to Novartis on the use of canakinumab to treat sJIA. Other authors declare no conflicts of interest.

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REFERENCES

- 1 Grom AA, Passo M. Macrophage activation syndrome in systemic juvenile rheumatoid arthritis. J Pediatr 1996; **129**: 630-632.
- 2 Prieur AM, Stephan JL. [Macrophage activation syndrome in rheumatic diseases in children]. *Rev Rhum Ed Fr* 1994; **61**: 447-451.
- 3 Sawhney S, Woo P, Murray KJ. Macrophage activation syndrome: a potentially fatal complication of rheumatic disorders. *Arch Dis Child* 2001; **85**: 421-426.
- 4 Parodi A, Davi S, Pringe AB, Pistorio A, Ruperto N, Magni-Manzoni S et al. Macrophage activation syndrome in juvenile systemic lupus erythematosus: a multinational multicenter study of thirty-eight patients. Arthritis Rheum 2009; 60: 3388 - 3399.
- 5 Avcin T, Tse SM, Schneider R, Ngan B, Silverman ED. Macrophage activation syndrome as the presenting manifestation of rheumatic diseases in childhood. *J Pediatr* 2006; **148**: 683-686.
- 6 Latino GA, Manlhiot C, Yeung RS, Chahal N, McCrindle BW. Macrophage activation syndrome in the acute phase of Kawasaki disease. J Pediatr Hematol Oncol 2010; 32: 527-531.
- 7 Simonini G, Pagnini I, Innocenti L, Calabri GB, De Martino M, Cimaz R. Macrophage activation syndrome/hemophagocytic lymphohistiocytosis and Kawasaki disease. *Pediatr Blood Cancer* 2010; **55**: 592.
- 8 Rigante D, Capoluongo E, Bertoni B, Ansuini V, Chiaretti A, Piastra M et al. First report of macrophage activation syndrome in hyperimmunoglobulinemia D with periodic fever syndrome. Arthritis Rheum 2007; 56: 658-661.
- 9 Rossi-Semerano L, Hermeziu B, Fabre M, Kone-Paut I. Macrophage activation syndrome revealing familial Mediterranean fever. *Arthritis Care Res (Hoboken)* 2010; **63**: 780-783.
- 10 Buoncompagni A, Loy A, Sala I, Ravelli A. The paradox of macrophage activation syndrome triggered by biologic medications. *Pediatr Rhematol Online J* 2005; 3: 70-73.
- 11 Athreya BH. Is macrophage activation syndrome a new entity? *Clin Exp Rheumatol* 2002; **20**: 121-123.
- 12 Ramanan AV, Schneider R. Macrophage activation syndrome-what's in a name!. *J Rheumatol* 2003; **30**: 2513-2516.
- 13 Behrens EM, Beukelman T, Paessler M, Cron RQ. Occult macrophage activation syndrome in patients with systemic juvenile idiopathic arthritis. *J Rheumatol* 2007; **34**: 1133 1138.
- 14 Bleesing J, Prada A, Siegel DM, Villanueva J, Olson J, Ilowite NT *et al.* The diagnostic significance of soluble CD163 and soluble interleukin-2 receptor alpha-chain in macrophage activation syndrome and untreated new-onset systemic juvenile idiopathic arthritis. *Arthritis Rheum* 2007; **56**: 965-971.
- 15 Henter JI, Horne A, Arico M, Egeler RM, Filipovich AH, Imashuku S et al. HLH-2004: diagnostic and therapeutic guidelines for hemophagocytic lymphohistiocytosis. *Pediatr Blood Cancer* 2007; 48: 124-131.
- 16 Pelkonen P, Swanljung K, Siimes MA. Ferritinemia as an indicator of systemic disease activity in children with systemic juvenile rheumatoid arthritis. Acta Paediatr Scand 1986; 75: 64-68.

- 17 Ravelli A, Magni-Manzoni S, Pistorio A, Besana C, Foti T, Ruperto N et al. Preliminary diagnostic guidelines for macrophage activation syndrome complicating systemic juvenile idiopathic arthritis. J Pediatr 2005; 146: 598-604.
- 18 Gupta A, Tyrrell P, Valani R, Benseler S, Weitzman S, Abdelhaleem M. The role of the initial bone marrow aspirate in the diagnosis of hemophagocytic lymphohisticytosis. *Pediatr Blood Cancer* 2008; **51**: 402 - 404.
- 19 Davi S, Consolaro A, Guseinova D, Pistorio A, Ruperto N, Martini A *et al.* An international consensus survey of diagnostic criteria for macrophage activation syndrome in systemic juvenile idiopathic arthritis. *J Rheumatol* 2011; **38**: 764-768.
- 20 Ramanan AV, Baildam EM. Macrophage activation syndrome is hemophagocytic lymphohistiocytosis-need for the right terminology. *J Rheumatol* 2002; 29: 1105; author reply 1105.
- 21 Filipovich AH. Hemophagocytic lymphohistiocytosis (HLH) and related disorders. Hematology Am Soc Hematol Educ Program 2009: 127-131.
- 22 Favara BE, Feller AC, Pauli M, Jaffe ES, Weiss LM, Arico M et al. Contemporary classification of histiocytic disorders. The WHO Committee on Histiocytic/ Reticulum Cell Proliferations. Reclassification Working Group of the Histiocyte Society. Med Pediatr Oncol 1997; 29: 157-166.
- 23 Zhang K, Jordan MB, Marsh RA, Johnson JA, Kissell D, Meller J *et al.* Hypomorphic mutations in PRF1, MUNC13-4, and STXBP2 are associated with adult-onset familial hemophagocytic lymphohistiocytosis. *Blood* 2011; **118**: 5794-5798.
- 24 Sullivan KE, Delaat CA, Douglas SD, Filipovich AH. Defective natural killer cell function in patients with hemophagocytic lymphohistiocytosis and in first degree relatives. *Pediatr Res* 1998; **44**: 465 468.
- 25 Feldmann J, Callebaut I, Raposo G, Certain S, Bacq D, Dumont C et al. Munc13-4 is essential for cytolytic granules fusion and is mutated in a form of familial hemophagocytic lymphohistiocytosis (FHL3). Cell 2003; 115: 461-473.
- 26 Stepp SE, Dufourcq-Lagelouse R, Le Deist F, Bhawan S, Certain S, Mathew PA *et al.* Perforin gene defects in familial hemophagocytic lymphohistiocytosis. *Science* 1999; **286**: 1957-1959.
- 27 zur Stadt U, Rohr J, Seifert W, Koch F, Grieve S, Pagel J *et al.* Familial hemophagocytic lymphohistiocytosis type 5 (FHL-5) is caused by mutations in Munc18-2 and impaired binding to syntaxin 11. *Am J Hum Genet* 2009; **85**: 482-492.
- 28 zur Stadt U, Schmidt S, Kasper B, Beutel K, Diler AS, Henter JI et al. Linkage of familial hemophagocytic lymphohistiocytosis (FHL) type-4 to chromosome 6q24 and identification of mutations in syntaxin 11. *Hum Mol Genet* 2005; 14: 827-834.
- 29 Menasche G, Pastural E, Feldmann J, Certain S, Ersoy F, Dupuis S *et al.* Mutations in RAB27A cause Griscelli syndrome associated with haemophagocytic syndrome. *Nat Genet* 2000; 25: 173 - 176.
- 30 Barbosa MD, Nguyen QA, Tchernev VT, Ashley JA, Detter JC, Blaydes SM *et al.* Identification of the homologous beige and Chediak-Higashi syndrome genes. *Nature* 1996; **382**: 262-265.
- 31 Coffey AJ, Brooksbank RA, Brandau O, Oohashi T, Howell GR, Bye JM et al. Host response to EBV infection in X-linked lymphoproliferative disease results from mutations in an SH2-domain encoding gene. Nat Genet 1998; 20: 129-135.
- 32 Marsh RA, Madden L, Kitchen BJ, Mody R, McClimon B, Jordan MB *et al.* XIAP deficiency: a unique primary immunodeficiency best classified as X-linked familial hemophagocytic lymphohistiocytosis and not as X-linked lymphopro-liferative disease. *Blood* 2010; **116**: 1079-1082.
- 33 Jordan MB, Hildeman D, Kappler J, Marrack P. An animal model of hemophagocytic lymphohisticocytosis (HLH): CD8+ T cells and interferon gamma are essential for the disorder. *Blood* 2004; **104**: 735-743.
- 34 Kagi D, Odermatt B, Mak TW. Homeostatic regulation of CD8+ T cells by perforin. *Eur J Immunol* 1999; **29**: 3262-3272.
- 35 Lykens JE, Terrell CE, Zoller EE, Risma K, Jordan MB. Perforin is a critical physiologic regulator of T-cell activation. *Blood* 2011; **118**: 618-626.
- 36 Menasche G, Feldmann J, Fischer A, de Saint Basile G. Primary hemophagocytic syndromes point to a direct link between lymphocyte cytotoxicity and homeostasis. *Immunol Rev* 2005; **203**: 165 179.
- 37 Behrens EM. Macrophage activation syndrome in rheumatic disease: what is the role of the antigen presenting cell? *Autoimmun Rev* 2008; **7**: 305-308.
- 38 Billiau AD, Roskams T, Van Damme-Lombaerts R, Matthys P, Wouters C. Macrophage activation syndrome: characteristic findings on liver biopsy illustrating the key role of activated, IFN-gamma-producing lymphocytes and IL-6- and TNF-alpha-producing macrophages. *Blood* 2005; **105**: 1648-1651.
- 39 Grom AA, Villanueva J, Lee S, Goldmuntz EA, Passo MH, Filipovich A. Natural killer cell dysfunction in patients with systemic-onset juvenile rheumatoid arthritis and macrophage activation syndrome. J Pediatr 2003; 142: 292-296.
- 40 Vastert SJ, van Wijk R, D'Urbano LE, de Vooght KM, de Jager W, Ravelli A *et al.* Mutations in the perforin gene can be linked to macrophage activation syndrome in patients with systemic onset juvenile idiopathic arthritis. *Rheumatology (Oxford)* 2010; **49**: 441-449.

- 41 Villanueva J, Lee S, Giannini EH, Graham TB, Passo MH, Filipovich A *et al.* Natural killer cell dysfunction is a distinguishing feature of systemic onset juvenile rheumatoid arthritis and macrophage activation syndrome. *Arthritis Res Ther* 2005; **7**: R30-R37.
- 42 Zhang K, Biroschak J, Glass DN, Thompson SD, Finkel T, Passo MH *et al.* Macrophage activation syndrome in patients with systemic juvenile idiopathic arthritis is associated with MUNC13-4 polymorphisms. *Arthritis Rheum* 2008; **58**: 2892-2896.
- 43 Hazen MM, Woodward AL, Hofmann I, Degar BA, Grom A, Filipovich AH et al. Mutations of the hemophagocytic lymphohistiocytosis-associated gene UNC13D in a patient with systemic juvenile idiopathic arthritis. *Arthritis Rheum* 2008; 58: 567-570.
- 44 Pivot-Pajot C, Varoqueaux F, de Saint Basile G, Bourgoin SG. Munc13-4 regulates granule secretion in human neutrophils. J Immunol 2008; **180**: 6786-6797.
- 45 Ren Q, Wimmer C, Chicka MC, Ye S, Ren Y, Hughson FM *et al.* Munc13-4 is a limiting factor in the pathway required for platelet granule release and hemostasis. *Blood* 2010; **116**: 869-877.
- 46 Zhang K, Johnson JA, Biroschak J, Villanueva J, Lee SM, Bleesing JJ et al. Familial haemophagocytic lymphohistiocytosis in patients who are heterozygous for the A91V perforin variation is often associated with other genetic defects. Int J Immunogenet 2007; 34: 231-233.
- 47 Yanagimachi M, Naruto T, Miyamae T, Hara T, Kikuchi M, Hara R et al. Association of IRF5 polymorphisms with susceptibility to macrophage activation syndrome in patients with juvenile idiopathic arthritis. J Rheumatol 2011; 38: 769-774.
- 48 Stinchcombe JC, Barral DC, Mules EH, Booth S, Hume AN, Machesky LM et al. Rab27a is required for regulated secretion in cytotoxic T lymphocytes. J Cell Biol 2001; 152: 825-834.
- 49 Crozat K, Hoebe K, Ugolini S, Hong NA, Janssen E, Rutschmann S et al. Jinx, an MCMV susceptibility phenotype caused by disruption of Unc13d: a mouse model of type 3 familial hemophagocytic lymphohistiocytosis. J Exp Med 2007; 204: 853-863.
- 50 Pachlopnik Schmid J, Ho CH, Chretien F, Lefebvre JM, Pivert G, Kosco-Vilbois M *et al.* Neutralization of IFNgamma defeats haemophagocytosis in LCMV-infected perforin- and Rab27a-deficient mice. *EMBO Mol Med* 2009; **1**: 112-124.
- 51 Pachlopnik Schmid J, Ho CH, Diana J, Pivert G, Lehuen A, Geissmann F et al. A Griscelli syndrome type 2 murine model of hemophagocytic lymphohistiocytosis (HLH). Eur J Immunol 2008; 38: 3219-3225.
- 52 Behrens EM, Canna SW, Slade K, Rao S, Kreiger PA, Paessler M *et al.* Repeated TLR9 stimulation results in macrophage activation syndrome-like disease in mice. *J Clin Invest* 2011; **121**: 2264-2277.
- 53 Fall N, Barnes M, Thornton S, Luyrink L, Olson J, llowite NT et al. Gene expression profiling of peripheral blood from patients with untreated new-onset systemic juvenile idiopathic arthritis reveals molecular heterogeneity that may predict macrophage activation syndrome. Arthritis Rheum 2007; 56: 3793-3804.
- 54 Ewald SE, Barton GM. Nucleic acid sensing Toll-like receptors in autoimmunity. *Curr Opin Immunol* 2011; 23: 3-9.
- 55 Guggemoos S, Hangel D, Hamm S, Heit A, Bauer S, Adler H. TLR9 contributes to antiviral immunity during gammaherpesvirus infection. *J Immunol* 2008; 180: 438-443.
- 56 Teramura T, Tabata Y, Yagi T, Morimoto A, Hibi S, Imashuku S. Quantitative analysis of cell-free Epstein-Barr virus genome copy number in patients with EBV-associated hemophagocytic lymphohistiocytosis. *Leuk Lymphoma* 2002; **43**: 173 179.
- 57 Sumegi J, Barnes MG, Nestheide SV, Molleran-Lee S, Villanueva J, Zhang K et al. Gene expression profiling of peripheral blood mononuclear cells from children with active hemophagocytic lymphohistiocytosis. Blood 2011; 117: e151-e160.
- 58 Fife MS, Gutierrez A, Ogilvie EM, Stock CJ, Samuel JM, Thomson W *et al.* Novel IL10 gene family associations with systemic juvenile idiopathic arthritis. *Arthritis Res Ther* 2006; **8**: R148.
- 59 Moller JC, Paul D, Ganser G, Range U, Gahr M, Kelsch R et al. IL10 promoter polymorphisms are associated with systemic onset juvenile idiopathic arthritis (SoJIA). Clin Exp Rheumatol 2010; 28: 912-918.
- 60 Krebs P, Crozat K, Popkin D, Oldstone MB, Beutler B. Disruption of MyD88 signaling suppresses hemophagocytic lymphohistiocytosis in mice. *Blood* 2011; 117: 6582-6588.
- 61 Hayashi K, Jin Z, Onoda S, Joko H, Teramoto N, Ohara N et al. Rabbit model for human EBV-associated hemophagocytic syndrome (HPS): sequential autopsy analysis and characterization of IL-2-dependent cell lines established from herpesvirus papio-induced fatal rabbit lymphoproliferative diseases with HPS. *Am J Pathol* 2003; **162**: 1721 - 1736.
- 62 Hsieh WC, Chang Y, Hsu MC, Lan BS, Hsiao GC, Chuang HC *et al.* Emergence of anti-red blood cell antibodies triggers red cell phagocytosis by activated macrophages in a rabbit model of Epstein-Barr virus-associated hemophagocytic syndrome. *Am J Pathol* 2007; **170**: 1629 1639.

- 63 Chan CB, Abe M, Hashimoto N, Hao C, Williams IR, Liu X *et al.* Mice lacking asparaginyl endopeptidase develop disorders resembling hemophagocytic syndrome. *Proc Natl Acad Sci USA* 2009; **106**: 468-473.
- 64 Yin L, Al-Alem U, Liang J, Tong WM, Li C, Badiali M et al. Mice deficient in the X-linked lymphoproliferative disease gene sap exhibit increased susceptibility to murine gammaherpesvirus-68 and hypo-gammaglobulinemia. J Med Virol 2003; 71: 446-455.
- 65 Parolini S, Bottino C, Falco M, Augugliaro R, Giliani S, Franceschini R *et al.* X-linked lymphoproliferative disease. 2B4 molecules displaying inhibitory rather than activating function are responsible for the inability of natural killer cells to kill Epstein-Barr virus-infected cells. *J Exp Med* 2000; **192**: 337-346.
- 66 Brown DE, McCoy MW, Pilonieta MC, Nix RN, Detweiler CS. Chronic murine typhoid fever is a natural model of secondary hemophagocytic lymphohistiocytosis. *PLoS One* 2010; **5**: e9441.
- 67 Yanaba K, Bouaziz JD, Matsushita T, Tsubata T, Tedder TF. The development and function of regulatory B cells expressing IL-10 (B10 cells) requires antigen receptor diversity and TLR signals. *J Immunol* 2009; **182**: 7459-7472.
- 68 Zoller EE, Lykens JE, Terrell CE, Aliberti J, Filipovich AH, Henson PM *et al.* Hemophagocytosis causes a consumptive anemia of inflammation. *J Exp Med* 2011; **208**: 1203-1214.
- 69 Schaer DJ, Schaer CA, Schoedon G, Imhof A, Kurrer MO. Hemophagocytic macrophages constitute a major compartment of heme oxygenase expression in sepsis. *Eur J Haematol* 2006; **77**: 432-436.
- 70 Hadchouel M, Prieur AM, Griscelli C. Acute hemorrhagic, hepatic, and neurologic manifestations in juvenile rheumatoid arthritis: possible relationship to drugs or infection. J Pediatr 1985; 106: 561-566.
- 71 Zeng HS, Xiong XY, Wei YD, Wang HW, Luo XP. Macrophage activation syndrome in 13 children with systemic-onset juvenile idiopathic arthritis. *World J Pediatr* 2008; **4**: 97 - 101.
- 72 Singh S, Chandrakasan S, Ahluwalia J, Suri D, Rawat A, Ahmed N *et al.* Macrophage activation syndrome in children with systemic onset juvenile idiopathic arthritis: clinical experience from northwest India. *Rheumatol Int* (in press).
- 73 Mouy R, Stephan JL, Pillet P, Haddad E, Hubert P, Prieur AM. Efficacy of cyclosporine A in the treatment of macrophage activation syndrome in juvenile arthritis: report of five cases. *J Pediatr* 1996; **129**: 750-754.
- 74 Quesnel B, Catteau B, Aznar V, Bauters F, Fenaux P. Successful treatment of juvenile rheumatoid arthritis associated haemophagocytic syndrome by cyclosporin A with transient exacerbation by conventional-dose G-CSF. Br J Haematol 1997; **97**: 508-510.
- 75 Ravelli A, De Benedetti F, Viola S, Martini A. Macrophage activation syndrome in systemic juvenile rheumatoid arthritis successfully treated with cyclosporine. *J Pediatr* 1996; **128**: 275-278.
- 76 Stephan JL, Kone-Paut I, Galambrun C, Mouy R, Bader-Meunier B, Prieur AM. Reactive haemophagocytic syndrome in children with inflammatory disorders. A retrospective study of 24 patients. *Rheumatology (Oxford)* 2001; 40: 1285-1292.
- 77 Rao A, Luo C, Hogan PG. Transcription factors of the NFAT family: regulation and function. *Annu Rev Immunol* 1997; **15**: 707 747.
- 78 Wallace CA, Sherry DD. Trial of intravenous pulse cyclophosphamide and methylprednisolone in the treatment of severe systemic-onset juvenile rheumatoid arthritis. Arthritis Rheum 1997; 40: 1852 - 1855.
- 79 Henter JI, Samuelsson-Horne A, Arico M, Egeler RM, Elinder G, Filipovich AH et al. Treatment of hemophagocytic lymphohistiocytosis with HLH-94 immunochemotherapy and bone marrow transplantation. Blood 2002; 100: 2367 - 2373.
- 80 Kishimoto T, Hamazaki T, Yasui M, Sasabe M, Okamura T, Sakata N et al. Autologous hematopoietic stem cell transplantation for 3 patients with severe juvenile rheumatoid arthritis. Int J Hematol 2003; 78: 453-456.
- 81 Brinkman DM, de Kleer IM, ten Cate R, van Rossum MA, Bekkering WP, Fasth A *et al.* Autologous stem cell transplantation in children with severe progressive systemic or polyarticular juvenile idiopathic arthritis: long-term follow-up of a prospective clinical trial. *Arthritis Rheum* 2007; **56**: 2410-2421.
- 82 Emmenegger U, Frey U, Reimers A, Fux C, Semela D, Cottagnoud P *et al.* Hyperferritinemia as indicator for intravenous immunoglobulin treatment in reactive macrophage activation syndromes. *Am J Hematol* 2001; **68**: 4-10.
- 83 Chen RL, Lin KH, Lin DT, Su JJ, Huang LM, Lee PI et al. Immunomodulation treatment for childhood virus-associated haemophagocytic lymphohistiocytosis. Br J Haematol 1995; 89: 282-290.
- 84 Larroche C, Bruneel F, Andre MH, Bader-Meunier B, Baruchel A, Tribout B et al. [Intravenously administered gamma-globulins in reactive hemaphagocytic syndrome. Multicenter study to assess their importance, by the immunoglobulins group of experts of CEDIT of the AP-HP]. Ann Med Interne (Paris) 2000; 151: 533-539.
- 85 Tristano AG, Casanova-Escalona L, Torres A, Rodriguez MA. Macrophage activation syndrome in a patient with systemic onset rheumatoid arthritis:

rescue with intravenous immunoglobulin therapy. J Clin Rheumatol 2003; 9: 253-258.

- 86 Miettunen PM, Narendran A, Jayanthan A, Behrens EM, Cron RQ. Successful treatment of severe paediatric rheumatic disease-associated macrophage activation syndrome with interleukin-1 inhibition following conventional immunosuppressive therapy: case series with 12 patients. *Rheumatology (Oxford)* 2011; **50**: 417-419.
- 87 Mahlaoui N, Ouachee-Chardin M, de Saint Basile G, Neven B, Picard C, Blanche S et al. Immunotherapy of familial hemophagocytic lymphohistiocytosis with antithymocyte globulins: a single-center retrospective report of 38 patients. *Pediatrics* 2007; **120**: e622 - e628.
- 88 Coca A, Bundy KW, Marston B, Huggins J, Looney RJ. Macrophage activation syndrome: serological markers and treatment with anti-thymocyte globulin. *Clin Immunol* 2009; **132**: 10-18.
- 89 Gurcan HM, Keskin DB, Stern JN, Nitzberg MA, Shekhani H, Ahmed AR. A review of the current use of rituximab in autoimmune diseases. *Int Immunopharmacol* 2009; **9**: 10–25.
- 90 Alexeeva El, Valieva SI, Bzarova TM, Semikina EL, Isaeva KB, Lisitsyn AO *et al.* Efficacy and safety of repeat courses of rituximab treatment in patients with severe refractory juvenile idiopathic arthritis. *Clin Rheumatol* 2011; **30**: 1163-1172.
- 91 Balamuth NJ, Nichols KE, Paessler M, Teachey DT. Use of rituximab in conjunction with immunosuppressive chemotherapy as a novel therapy for Epstein Barr virus-associated hemophagocytic lymphohistiocytosis. J Pediatr Hematol Oncol 2007; 29: 569-573.
- 92 Bosman G, Langemeijer SM, Hebeda KM, Raemaekers JM, Pickkers P, van der Velden WJ. The role of rituximab in a case of EBV-related lymphoproliferative disease presenting with haemophagocytosis. *Neth J Med* 2009; **67**: 364-365.
- 93 Aeberli D, Oertle S, Mauron H, Reichenbach S, Jordi B, Villiger PM. Inhibition of the TNF-pathway: use of infliximab and etanercept as remission-inducing agents in cases of therapy-resistant chronic inflammatory disorders. *Swiss Med Wkly* 2002; **132**: 414-422.
- 94 Emmenegger U, Reimers A, Frey U, Fux C, Bihl F, Semela D et al. Reactive macrophage activation syndrome: a simple screening strategy and its potential in early treatment initiation. *Swiss Med Wkly* 2002; **132**: 230-236.
- 95 Henzan T, Nagafuji K, Tsukamoto H, Miyamoto T, Gondo H, Imashuku S et al. Success with infliximab in treating refractory hemophagocytic lymphohistiocytosis. Am J Hematol 2006; 81: 59-61.
- 96 Maeshima K, Ishii K, Iwakura M, Akamine M, Hamasaki H, Abe I et al. Adult-onset Still's disease with macrophage activation syndrome successfully treated with a combination of methotrexate and etanercept. *Mod Rheumatol* 2012; 22: 137 - 141.
- 97 Makay B, Yilmaz S, Turkyilmaz Z, Unal N, Oren H, Unsal E. Etanercept for therapyresistant macrophage activation syndrome. *Pediatr Blood Cancer* 2008; 50: 419-421.
- 98 Prahalad S, Bove KE, Dickens D, Lovell DJ, Grom AA. Etanercept in the treatment of macrophage activation syndrome. J Rheumatol 2001; 28: 2120-2124.
- 99 Sellmer A, Stausbol-Gron B, Krag-Olsen B, Herlin T. Successful use of infliximab in macrophage activation syndrome with severe CNS involvement. *Scand J Rheumatol* 2011; **40**: 156-157.
- 100 Takahashi N, Naniwa T, Banno S. Successful use of etanercept in the treatment of acute lupus hemophagocytic syndrome. *Mod Rheumatol* 2008; 18: 72-75.
- 101 Verbsky JW, Grossman WJ. Hemophagocytic lymphohisticocytosis: diagnosis, pathophysiology, treatment, and future perspectives. Ann Med 2006; 38: 20-31.
- 102 Chauveau E, Terrier F, Casassus-Buihle D, Moncoucy X, Oddes B. [Macrophage activation syndrome after treatment with infliximab for fistulated Crohn's disease]. Presse Med 2005; 34: 583-584.
- 103 Kaneko K, Kaburaki M, Muraoka S, Tanaka N, Yamamoto T, Kusunoki Y *et al.* Exacerbation of adult-onset Still's disease, possibly related to elevation of serum tumor necrosis factor-alpha after etanercept administration. *Int J Rheum Dis* 2010; **13**: e67 - e69.
- 104 Kimura Y, Pinho P, Walco G, Higgins G, Hummell D, Szer I *et al.* Etanercept treatment in patients with refractory systemic onset juvenile rheumatoid arthritis. *J Rheumatol* 2005; **32**: 935-942.
- 105 Nadia EA, Carvalho JF, Bonfa E, Lotito AP, Silva CA. Macrophage activation syndrome associated with etanercept in a child with systemic onset juvenile idiopathic arthritis. *Isr Med Assoc J* 2009; **11**: 635-636.
- 106 Ramanan AV, Schneider R. Macrophage activation syndrome following initiation of etanercept in a child with systemic onset juvenile rheumatoid arthritis. *J Rheumatol* 2003; **30**: 401-403.
- 107 Sandhu C, Chesney A, Piliotis E, Buckstein R, Koren S. Macrophage activation syndrome after etanercept treatment. *J Rheumatol* 2007; **34**: 241-242.

- 108 Sterba G, Sterba Y, Stempel C, Blank J, Azor E, Gomez L. Macrophage activation syndrome induced by etanercept in a patient with systemic sclerosis. *Isr Med Assoc J* 2010; **12**: 443-445.
- 109 Stern A, Riley R, Buckley L. Worsening of macrophage activation syndrome in a patient with adult onset Still's disease after initiation of etanercept therapy. *J Clin Rheumatol* 2001; **7**: 252-256.
- 110 Agarwal S, Moodley J, Ajani Goel G, Theil KS, Mahmood SS, Lang RS. A rare trigger for macrophage activation syndrome. *Rheumatol Int* 2011; 31: 405-407.
- 111 Molto A, Mateo L, Lloveras N, Olive A, Minguez S. Visceral leishmaniasis and macrophagic activation syndrome in a patient with rheumatoid arthritis under treatment with adalimumab. *Joint Bone Spine* 2010; **77**: 271-273.
- 112 Behrens EM, Kreiger PA, Cherian S, Cron RQ. Interleukin 1 receptor antagonist to treat cytophagic histiocytic panniculitis with secondary hemophagocytic lymphohistiocytosis. J Rheumatol 2006; 33: 2081 - 2084.
- 113 Verbsky JW, White AJ. Effective use of the recombinant interleukin 1 receptor antagonist anakinra in therapy resistant systemic onset juvenile rheumatoid arthritis. J Rheumatol 2004; 31: 2071 - 2075.
- 114 Pascual V, Allantaz F, Arce E, Punaro M, Banchereau J. Role of interleukin-1 (IL-1) in the pathogenesis of systemic onset juvenile idiopathic arthritis and clinical response to IL-1 blockade. J Exp Med 2005; 201: 1479-1486.
- 115 Nigrovic PA, Mannion M, Prince FH, Zeft A, Rabinovich CE, van Rossum MA *et al.* Anakinra as first-line disease-modifying therapy in systemic juvenile idiopathic arthritis: report of forty-six patients from an international multicenter series. *Arthritis Rheum* 2011; **63**: 545-555.
- 116 Bruck N, Suttorp M, Kabus M, Heubner G, Gahr M, Pessler F. Rapid and sustained remission of systemic juvenile idiopathic arthritis-associated macrophage activation syndrome through treatment with anakinra and corticosteroids. *J Clin Rheumatol* 2011; **17**: 23-27.
- 117 Kelly A, Ramanan AV. A case of macrophage activation syndrome successfully treated with anakinra. *Nat Clin Pract Rheumatol* 2008; **4**: 615-620.
- 118 Record JL, Beukelman T, Cron RQ. Combination therapy of abatacept and anakinra in children with refractory systemic juvenile idiopathic arthritis: a retrospective case series. *J Rheumatol* 2011; **38**: 180-181.
- 119 Burgi U, Mendez A, Hasler P, Hullstrung HD. Hemophagocytic syndrome in adultonset Still's disease (AOSD): a must for biologics?-Case report and brief review of the literature. *Rheumatol Int* (in press).
- 120 Chou RC, Dinarello CA, Ferry JA, Dal Cin P. A 36-year-old woman with recurrent high-grade fevers, hypotension, and hypertriglyceridemia. *Arthritis Care Res* (Hoboken) 2010; 62: 128-136.
- 121 Debiais S, Maillot F, Luca L, Buret J, Fautrel B, Renard JP. Efficacy of anakinra in a case of refractory Still disease. J Clin Rheumatol 2008; **14**: 357-358.
- 122 Durand M, Troyanov Y, Laflamme P, Gregoire G. Macrophage activation syndrome treated with anakinra. *J Rheumatol* 2010; **37**: 879-880.

- 123 Fitzgerald AA, Leclercq SA, Yan A, Homik JE, Dinarello CA. Rapid responses to anakinra in patients with refractory adult-onset Still's disease. *Arthritis Rheum* 2005; 52: 1794-1803.
- 124 Kalliolias GD, Georgiou PE, Antonopoulos IA, Andonopoulos AP, Liossis SN. Anakinra treatment in patients with adult-onset Still's disease is fast, effective, safe and steroid sparing: experience from an uncontrolled trial. Ann Rheum Dis 2007; 66: 842-843.
- 125 Maier J, Birkenfeld G, Pfirstinger J, Scholmerich J, Fleck M, Bruhl H. Effective treatment of steroid refractory adult-onset Still's disease with anakinra. *J Rheumatol* 2008; **35**: 939-941.
- 126 Mehta BM, Hashkes PJ, Avery R, Deal CL. A 21-year-old man with Still's disease with fever, rash, and pancytopenia. Arthritis Care Res (Hoboken) 2010; 62: 575-579.
- 127 Naumann L, Feist E, Natusch A, Langen S, Krause A, Buttgereit F et al. IL1receptor antagonist anakinra provides long-lasting efficacy in the treatment of refractory adult-onset Still's disease. Ann Rheum Dis 2010; 69: 466-467.
- 128 Youssef J, Lazaro E, Blanco P, Viallard JF. Blockade of interleukin 1 receptor in Still's disease affects activation of peripheral T-lymphocytes. J Rheumatol 2008; 35: 2453 - 2456.
- 129 Clark SR, McMahon CJ, Gueorguieva I, Rowland M, Scarth S, Georgiou R et al. Interleukin-1 receptor antagonist penetrates human brain at experimentally therapeutic concentrations. J Cereb Blood Flow Metab 2008; 28: 387-394.
- 130 Fisher Jr CJ, Dhainaut JF, Opal SM, Pribble JP, Balk RA, Slotman GJ et al. Recombinant human interleukin 1 receptor antagonist in the treatment of patients with sepsis syndrome. Results from a randomized, double-blind, placebo-controlled trial. Phase III rhIL-1ra Sepsis Syndrome Study Group. JAMA 1994; 271: 1836-1843.
- 131 Canna S, Frankovich J, Higgins G, Narkewicz MR, Nash SR, Hollister JR et al. Acute hepatitis in three patients with systemic juvenile idiopathic arthritis taking interleukin-1 receptor antagonist. *Pediatr Rheumatol Online J* 2009; 7: 21.
- 132 Lurati A, Teruzzi B, Salmaso A, Demarco G, Pontikaki I, Gattinara M et al. Macrophage activation syndrome (MAS) during anti-IL1 therapy (anakinra) in a patient affected by systemic juvenile arthritis (soJIA): a report and review of the literature. Pediatr Rheumatol Online J 2005; 3: 79-85.
- 133 Zeft A, Hollister R, LaFleur B, Sampath P, Soep J, McNally B *et al.* Anakinra for systemic juvenile arthritis: the Rocky Mountain experience. *J Clin Rheumatol* 2009; **15**: 161-164.
- 134 Yokota S, Imagawa T, Mori M, Miyamae T, Aihara Y, Takei S et al. Efficacy and safety of tocilizumab in patients with systemic-onset juvenile idiopathic arthritis: a randomised, double-blind, placebo-controlled, withdrawal phase III trial. *Lancet* 2008; **371**: 998-1006.
- 135 Kobayashi M, Takahashi Y, Yamashita H, Kaneko H, Mimori A. Benefit and a possible risk of tocilizumab therapy for adult-onset Still's disease accompanied by macrophage-activation syndrome. *Mod Rheumatol* 2011; **21**: 92-96.

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