



# **Magnetite Nanoparticles in Magnetic Hyperthermia and Cancer Therapies: Challenges and Perspectives**

Agnieszka Włodarczyk<sup>1</sup>, Szymon Gorgoń<sup>2</sup>, Adrian Radoń<sup>3</sup> and Karolina Bajdak-Rusinek<sup>1,\*</sup>

- <sup>1</sup> Department of Medical Genetics, Faculty of Medical Sciences in Katowice, Medical University of Silesia, Medyków 18, 40-752 Katowice, Poland; awlodarczyk@sum.edu.pl
- <sup>2</sup> Department of Surgical and Perioperative Sciences, Surgery, Umeå University, 901 87 Umeå, Sweden; szymon.gorgon@umu.se
- <sup>3</sup> Łukasiewicz Research Network—Institute of Non-Ferrous Metals, Sowinskiego 5 St., 44-100 Gliwice, Poland; adrian.radon@imn.lukasiewicz.gov.pl
- \* Correspondence: kbajdak-rusinek@sum.edu.pl; Tel.: +48-32-208-8382

**Abstract:** Until now, strategies used to treat cancer are imperfect, and this generates the need to search for better and safer solutions. The biggest issue is the lack of selective interaction with neoplastic cells, which is associated with occurrence of side effects and significantly reduces the effectiveness of therapies. The use of nanoparticles in cancer can counteract these problems. One of the most promising nanoparticles is magnetite. Implementation of this nanoparticle can improve various treatment methods such as hyperthermia, targeted drug delivery, cancer genotherapy, and protein therapy. In the first case, its feature makes magnetite useful in magnetic hyperthermia. Interaction of magnetite with the altered magnetic field generates heat. This process results in raised temperature only in a desired part of a patient body. In other therapies, magnetite-based nanoparticles could serve as a carrier for various types of therapeutic load. The magnetic field would direct the drug-related magnetite nanoparticles to the pathological site. Therefore, this material can be used in protein and gene therapy or drug delivery. Since the magnetite nanoparticle can be used in various types of cancer treatment, they are extensively studied. Herein, we summarize the latest finding on the applicability of the magnetite nanoparticles, also addressing the most critical problems faced by smart nanomedicine in oncological therapies.

Keywords: magnetite nanoparticles; hyperthermia; smart nanomedicine

# 1. Introduction

Cancer is one of the most common causes of human death. In 2020 there were approximately 19.3 million cancer cases, and it is estimated that this number could increase to 28.4 million in 2040 [1]. Therefore, new cancer-treatment methods are tested and have been recently developed. These methods include improving well-established clinical methods such as chemotherapy, radiotherapy, or the development of less invasive surgery methods [2]. However, traditional methods still have a nonselective effect on neoplastic cells, thus interacting with healthy cells. This may result in limited effectiveness or adverse effect of treatment on a patient, even though these methods have the potential to kill cancer cells. Therefore, the development of new cancer treatments has focused on tumor-selective methods [3]. In recent years, selective inhibitors have been introduced into clinical use. They are used in targeted therapies against specific mutant signaling pathways [4]. Although inhibitors' effectiveness has been proven, they are connected the with development of drug resistance by cancer cells [5]. Due to limitations and problems encountered in approved cancer therapies, there is a great need for new and better cancer treatment strategies.

One of the proposed alternative approaches in cancer treatment is magnetic hyperthermia [6]. Hyperthermia is already used to support the treatment of many cancers, e.g., head and neck, breast, cervical, sarcomas, and melanomas [2]. Although selective hyperthermia



Citation: Włodarczyk, A.; Gorgoń, S.; Radoń, A.; Bajdak-Rusinek, K. Magnetite Nanoparticles in Magnetic Hyperthermia and Cancer Therapies: Challenges and Perspectives. *Nanomaterials* **2022**, *12*, 1807. https:// doi.org/10.3390/nano12111807

Academic Editors: Dimitri Stanicki and Sophie Laurent

Received: 4 May 2022 Accepted: 23 May 2022 Published: 25 May 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). is not expected to be a standalone method, it could put selective pressure on cancer cells, increasing the therapeutic effect of primary cancer treatment. In the method, thermal energy is generated when magnetic nanoparticles are influenced by an alternating magnetic field [7]. Limiting the magnetic field gradient to a pathogenic site only allows increasing body temperature in the desired region of the body selectively.

Magnetic hyperthermia is based on the use of magnetic nanoparticles. Among the dozen or so magnetic nanoparticles, the most promising is magnetite. Magnetite nanoparticles are already in clinical use: they have found application as a diagnostic tool for contrasting [8]. In addition, there are plans to use magnetite in other ways in medicine. It has been proposed to use these nanoparticles in the separation of proteins. However, the wide range of magnetite properties makes them attractive nanoparticles that could be used in oncological therapies. Their potential in targeted drug delivery, gene, or protein therapy has been explored [8]. Increased interest in the topic is reflected in the number of published scientifical papers. An NCBI database search for the combined key words "magnetite" and "cancer" gives 1390 publications, including 7 clinical trials, published in the last 5 years [9].

In the presented work, we summarize the research and achievements to date in the field of medicine, biology, and chemistry that concentrate on therapeutic use of magnetite nanoparticles in oncology. In the presented context, magnetite research faces problems typical for translational medicine, i.e., in laboratory (both chemical and preclinical) and in clinics as well. Here, we describe the problems related to the medical use of magnetite nanoparticles and future research directions.

## 2. Nanoparticles

Nanoparticles (NPs) are defined as solid particles in the size range 10–1000 nm [10]; however, the definition introduced by the European Commission states that nanoparticles include materials where at least half of the particles are equal to or smaller than 100 nm [11]. In addition, they often exhibit new and distinct electrical, optical, magnetic, biological, and chemical properties [3,12]. Furthermore, nanoparticles can be synthesized from various materials, e.g., composite polymers, semiconductors, metals, or even lipids and proteins [13], and are characteristic of different shapes, including spheres, rods, or tubes [13]. Due to their various properties, there is growing interest in the use of nanoparticles to develop new materials of unusual and novel features [14,15]. Currently nanoparticles are tested in various fields such as catalysis [16,17], energy storage [18], energy conversion [19,20], and optoelectronic, for example, as light-emitting diodes [21]. In the case of the biomedical applications, NPs are tested not only as anticancer agents, which will be discussed later in this paper but also as antibacterial agents (for example in form of the layers on medical implants) [22,23], and sensitive tests to detect various diseases [24,25]. Generally, this wide application range is related to their unique properties and highly reactive surface. Unfortunately, their high surface energy can also result in poor stability [26].

Medicine is one of the branches of science where nanoparticles are expected to have the most significant impact. First and foremost, nanoparticles are planned to be used in so-called smart nanomedicine (Table 1). Here, the nanoparticle is coated using a polymer or other nanostructures such as metallic ones are deposited on their surface. It enables to attachment of bioactive substances such as cytotoxic or therapeutic drugs [27]. Targeting and activating nanoparticles occurs under various factors, including magnetic field, light waves, ultrasounds, and internal, including pH, temperature, redox potential, or enzymes [28]. This procedure allows for the obtaining of the desired therapeutic effect at the pathogenic site only. Another promising use of nanoscale objects is a theranostic nanoparticle, i.e., a particle that can be used in diagnostics and simultaneously therapeutic agents [29–31].

Currently, they are used in cancer medicine, mainly in diagnostics. In addition, nanoparticles serve as a stabilizing factor for therapeutics when used in therapies. According to the authors' knowledge, about 10 types of nanoparticle-based therapies have been approved by the U.S. Food and Drug Administration (FDA) and European Medicines Agency(EMA) [32].

Group	Nanoparticle	Drugs, Functionalizing Agents, or Bioactive Molecules	Size (nm)	Shape	Nanoparticle Role	Application Type	Reference
	Au	hesperidin	10–50 <sup>1</sup>	spherical	Drug-delivery system	antioxidant agent; protective agent against DNA damage from H <sub>2</sub> O <sub>2</sub> , anticancer drug-delivery system for breast cancer therapy	[33]
	AgCu	sodium citrate, mercapto-propionic acid	4–32 <sup>2</sup>	spherical	anticancer agent	selective toxicity against MCF-7 breast cancer cells	[34]
Metallic	Pt	doxorubicin, fucoidan	$33 \pm 3.4$ <sup>3</sup>	coral-like	photothermal agent, drug-delivery system	nanoplatforms for synergetic biological-thermo-chemo trimodal treatment of MCF-7 breast cancer cells	[35]
	Au@Pt	methoxy-PEG-thiol	20–30	dendritic	photothermal agent	photothermal therapy under 808 nm light irradiation	[36]
	Pd	polyvinylpyrrolidone	9–15	spherical	anticancer agent	ROS generation and cleaving of the mitochondrial membrane	[37]
	MnAl layered double hydroxide	Fluorouracil	34.9 and 24.7 <sup>4</sup>	nanosheets	Drug-delivery system, MRI contrast	anticancer drug-delivery system combined with MRI	[38]
	ZnO	quercetin	21–39	hexagonal	Drug-delivery system	antioxidant agent; protective agent against DNA damage from H <sub>2</sub> O <sub>2</sub> , anticancer drug-delivery system for breast cancer therapy selective toxicity against MCF-7 breast cancer cells nanoplatforms for synergetic biological-thermo-chemo trimodal treatment of MCF-7 breast cancer cells photothermal therapy under 808 nm light irradiation ROS generation and cleaving of the mitochondrial membrane anticancer drug-delivery system	[39]
Nonmetallic	Fe <sub>3</sub> O <sub>4</sub>	BSA protein, glutaric acid	$4.5\pm0.1$ $^5;98$ $^6$	spherical	magnetic hyperthermia agent	hyperthermia therapy	[40]
	CuS	Doxorubicin, DGM <sup>7</sup> , peptides <sup>8</sup>	$22\ {}^9; 10\ {}^{10}; \\151.5\pm 2.2\ {}^{11}$	spherical	cross-linker, photothermal agent		[41]
	graphene quantum dots	TAT peptides, FA-PEG-NH $_2$ <sup>12</sup>	5	spherical	anticancer agent		[42]

Table 1. The potential	applications	of nanoparticles i	n smart nanomedicine.

Group	Nanoparticle	Drugs, Functionalizing Agents, or Bioactive Molecules	Size (nm)	Shape	Nanoparticle Role	Application Type	Reference
	Au@SiO <sub>2</sub>	poly (N-isopropylacrylamide- co-acrylic acid); indocyanine green	$59 \pm 3.6  {}^{13}; \\ 9.3 \pm 1.2  {}^{14}$	nanorods	Drug-delivery system, photothermal agent	photodynamic/photothermal tumor therapy by combining the effect of Au NPs with ICG under irradiation of 808 nm laser	[43]
	Fe <sub>3</sub> O <sub>4</sub> @SiO <sub>2</sub>	lactoferrin, doxorubicin	119 <sup>15</sup>	spherical	Drug-delivery system, magnetic hyperthermia, and photothermal agent	chemo-magnetic field-photothermal breast cancer therapy	[44]
Composite	silica-carbon nano-onion	fucoidan, doxorubicin, HM30181A	~50	nano-onions	Drug-delivery system, photothermal agent	anticancer drug releasing combined with the binding of P-glycoprotein pumps and photothermal effect	[45]
	AlMg layered double hydroxide-Fe <sub>3</sub> O <sub>4</sub>	hyaluronic acid, doxorubicin	$\begin{array}{c} 3.2 \pm 0.2 \ {}^{16} \\ 68.8 \pm 10.7 \ {}^{17} \end{array}$	disk-shaped	Drug-delivery system, MRI contrast,	T1-weighted MR imaging and chemotherapy by targeting cancer cells overexpressing CD44 receptors	[46]
	Ag@SiO <sub>2</sub> Ag@SiO <sub>2</sub> @Ag	n.a.	$77 \pm 13 \ {}^{18}; \\ 83 \pm 18 \ {}^{19}$	core-shell spherical	bioimaging agent, photothermal agents	bioimaging and photothermal therapy using 400 nm laser	[47]

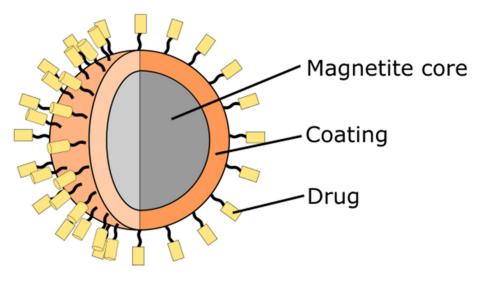
<sup>1</sup> for hesperidin-loaded Au NPs, <sup>2</sup> depending on Ag: Cu ratio, <sup>3</sup> for fucoidan coated Pt NPs, <sup>4</sup> measured at two different dimensions, <sup>5</sup> for pure Fe<sub>3</sub>O<sub>4</sub> NPs. <sup>6</sup> hydrodynamic diameter for Pro-Glu-Fe<sub>3</sub>O<sub>4</sub> NPs (Pro—BSA protein, Glu—glutaric acid), <sup>7</sup> poly(ε-caprolactone)-ss-poly(2-(diisopropylamino)ethyl methacrylate/glycidyl methacrylate/2-methylacrylloxyethyl phosphorylcholine, <sup>8</sup> cyclo(AGA-d-PC) and (YGRKKKRRQRRRC) peptides, <sup>9</sup> CuS size determined based on dynamic light scattering, <sup>10</sup> CuS size determined based on the TEM measurements, <sup>11</sup> sizes for cross-linked micelles, <sup>12</sup> folic acid-linked polyethylene glycol, <sup>13</sup> Au nanorod length, <sup>14</sup> Au nanorod width, <sup>15</sup> for lactoferrin loaded composite <sup>16</sup> for pure Fe<sub>3</sub>O<sub>4</sub> NPs, <sup>17</sup> for composite, <sup>18</sup> for nanoparticles with 5 nm Ag seeds on the surface, <sup>19</sup> for nanoparticles with 12 nm Ag NPs on the surface.

### Iron-Oxide Nanoparticles

Nanoparticles that have been used in medicine can be divided into three groups: metal, nonmetallic, and composite nanoparticles. The most commonly used are metal and metal-oxide nanoparticles [3]. Because of biocompatibility and unique magnetic properties, the most widely studied are ferrites, especially superparamagnetic iron-oxide nanoparticles (SPIONs). Recently, iron-oxide nanoparticles have attracted much consideration due to their unique properties, such as superparamagnetism, surface-to-volume ratio, greater surface area, and easy separation methodology. Superparamagnetic nanoparticles are susceptible to the effects of a magnetic field; however, when the external magnetic field is lost, they become demagnetized. This feature, together with good colloidal stability, is clinically important. It minimizes the risk of nanoparticles' aggregation in the blood [48]. Additionally, the biodegradability of SPIONs seems to be one of the most important features for clinical use [49–51].

Within the group can be found such iron oxides as maghemite ( $\gamma$ -Fe<sub>2</sub>O<sub>3</sub>), hematite ( $\alpha$ -Fe<sub>2</sub>O<sub>3</sub>), and magnetite (Fe<sub>3</sub>O<sub>4</sub>) [52]. From a medical point of view, magnetite seems to be the most promising SPION. It is due to its unique magnetic, catalytic, and biochemical properties [53]. Fe<sub>3</sub>O<sub>4</sub> nanoparticles have a large surface area needed for adsorption and immobilization of molecules or drugs. They can be easily modified to improve absorption [54]. Moreover, their size and shape can be easily controlled by using different synthesis methods [55]. It is possible to synthesize multifunctional platforms using Fe<sub>3</sub>O<sub>4</sub> nanoparticles, in which other compounds cover magnetite nanoparticles, such as SiO<sub>2</sub>, or even by the biodegradable polymers [56–58].

Therapeutic magnetite nanoparticles usually consist of three essential components: magnetite core, which serves as a carrier for therapeutic agents; a coating on the ironoxide nanoparticle, which promotes beneficial interactions in the body; and a therapeutic load that performs a designated function in vivo (Figure 1). In addition, a particle may consist of a ligand that recognizes specific receptors on the outer surface of cancer cells [59]. Interestingly, these structures can also be modified, and magnetite nanoparticles with or without coating can be deposited on the other nanostructures such as layered double hydroxides (LDHs). Interestingly,  $Fe_3O_4$  nanoparticles can also be covered by LDHs to form nanostructures with different morphology and properties [60]. This flexibility of synthesis of nanostructured multifunctional platforms results in the high interest in applying magnetite in nanomedicine. Fedorenko et al. [61] have shown that the ultrafine iron-oxide nanoparticles can be covered by the silica layer and this composite can be doped by the Gd(III) complexes to improve their MRI dual-contrast ability. Formation of the core-shell structure results in much higher stability in dispersion, which is connected with the high electrokinetic potential (about -78 mW). Moreover, not only the formation of this type of nanostructures can change the properties of iron oxides. As was recently presented, the modification of the surface by various surfactants can highly modify the magnetic properties and also the transverse relaxation rate of water molecules in the aqueous colloids of hydrophilic magnetic nanoparticles [62,63].



**Figure 1.** Schematic representation of a therapeutic magnetite-based nanoparticle. In typical magnetite-based nanoparticle, core of the nanoparticle is made of magnetite. The core is covered with a coating substance. On the surface, therapeutic load is attached.

### 3. Hyperthermia

Hyperthermia is defined as an increase in body temperature by  $\geq 1$  °C [64] or as an increase in temperature to 39–45 °C [65]. There are four types of hypothermia: fever, movement-related hyperthermia, inadequate temperature drainage hyperthermia, and pathological or medication-induced hyperthermia [66]. Healthy human somatic cells, except neurones, can survive at a temperature of 44 °C for at least 1 h [67,68]. Thus, hyperthermia does not damage healthy tissues unless the temperature exceeds the above value. On the other hand, hyperthermia in the temperature range 45–50 °C causes thermal ablation, i.e., necrosis through dehydration, protein denaturation, and cell-membrane damage [65,69]. Although hyperthermia can be dangerous to the organism, it can be of clinical use, especially when directed against specific cells or tissue. Medical hyperthermia is classified according to the size of the area where the temperature is raised. Thus, hyperthermia is divided into local-regional and whole-body hyperthermia [2].

Increasing temperature in a limited-body region allows maximizing therapeutic effect in the appropriate parts of the body while minimizing undesirable effects of heating in other body regions [70]. There are several possible therapeutically methods of increasing temperature in a specific body fragment. One of them is magnetic hyperthermia [71]. In the method, the alternating magnetic field generates electromagnetic radiation. This energy is absorbed by SPION and converted into thermal energy [72,73]. As a result, the surrounding tissue heats up. A similar method of increasing body temperature is photothermic. Here, instead of the alternating magnetic field, a laser is used as a source of electromagnetic radiation [13,73,74]. Therefore, laser-induced temperature elevation may be a more effective method of reaching the desired temperature than magnetically induced hyperthermia [74]. First of all, light with partially transparent tissues would reduce the main drawback of magnetic field-induced hyperthermia, i.e., low tissue penetration [73]. Both photothermia and magnetic hyperthermia have been intensively studied as potential cancer treatments [75–77].

#### 3.1. Hyperthermia in Oncology

In the 18th century, it was noticed that tumors shrank in patients with fever. Since then, hyperthermia has been intensively studied as a potential cancer treatment [78]. However, the first attempts to use hyperthermia in the clinic were made in the 20th century [79]. At the beginning of the century, in vitro and in vivo studies showed that neoplastic cells do not tolerate increased temperature as well as healthy cells [80,81]. Furthermore, toxicity

tests in dogs, sheep, and pigs showed that progressive necrosis of normal and neoplastic tissues occurred at temperatures above 45 °C. However, internal heat dissipation occurred as the normal tissues approached this temperature. This phenomenon resulted, in healthy tissues maintaining the temperature below 45 °C, with practically no damage to normal organs, tissue, or skin. However, most solid tumors did not have this adaptability and heated up to 50 °C [82].

So far, many studies have been carried out to confirm the effects of hyperthermia [2]. Initially, hyperthermia was planned to be used as a standalone therapy [79]. However, hyperthermia was studied mainly as a support to other therapies. This approach includes a combination of hyperthermia and chemotherapy [73,83], radiotherapy [71,84,85] or with both [86,87]. Results showed that increasing body temperature increases the effectiveness of therapies [71,73,83,88,89], which was also estimated and confirmed by many clinical studies [2,90].

Currently, oncological hyperthermia is used primarily as an adjunctive method in treating many cancer types, along with chemotherapy or radiotherapy [2,71,78]. In this case, the patient's body temperature is usually raised to around 40–43 °C [88,89]. In addition, hyperthermia is also used as an independent therapy for ablation of individual neoplastic lesions, where a temperature above 50 °C is used [71].

Although many studies show the positive effect of hyperthermia in cancer treatment, this method is still controversial, and the results of clinical trials have been criticized [2,91]. In addition, some problems make it difficult to assess the effect of hyperthermia on tumors in a clinical setting. The main problem is the lack of a precise oncological definition of hyperthermia [2]. In addition, it makes it difficult to standardize the methodology. Moreover, the unresolved technical problems with hyperthermia are limiting factors in the wide use of hyperthermia as a cancer-treatment strategy (see Section 5). Due to the above problems, hyperthermia is a rarely used strategy in oncology even when it is clinically approved for use in some cancer therapies [2,92].

#### 3.1.1. How Hyperthermia Kills Neoplastic Cells?

Although many studies confirmed the effectiveness of hyperthermia, the exact mechanism of its action is not fully understood [2,91]. The main problem in pinpointing the exact mechanism is fact that many changes occur in a cell at high temperatures [91]. So far, several explanations have been given. First of all, the effect of hyperthermia on tumors may occur at the cellular level. In vitro studies have shown that temperatures above 41 °C block the DNA double-break repair mechanism [2]. During the S-phase of cell division, cells tend to be less susceptible to the effects of therapeutics. This changes when the temperature increases and the cells show increased sensitivity to the therapies. Some cytostatics interact with cells during the S-phase of division so that hyperthermia could enhance the effect of these drugs [2]. The fast rate of cell divisions and hypoxia characteristic of cancer makes cancer cells more susceptible to hyperthermy [93]. It has also been shown that the combination of gold nanowires with hyperthermia can damage the cytoskeleton and affect the migration of cancer cells [94] or synergistically support the inactivation of the complex of oncogenic proteins PML/RAR $\alpha$  [95,96]. In the case of tumors with a high level of well-known oncogenic proteins p53 and Ki-67, the addition of hyperthermia to chemical treatment improves the effectiveness of the therapy [97]. Other studies have shown that hyperthermia can induce apoptosis through p53-dependent and -independent mechanisms [98].

Moreover, hyperthermia can help overcome the problem of drug resistance by modulation of different pathways [88]. However, more studies are needed. For example, results show that hyperthermia can act positively and negatively in developing drug resistance and can induce or help overcome resistance [88].

However, hyperthermia can affect tumors by modulation of blood flow [91]. In healthy tissue, blood vessels are hierarchically connected into efficient networks of arteries, capillaries, and veins. On the other hand, cancerous blood vessels are chaotic, leaky, and

inefficient [99]. Moderate hyperthermia (T < 42  $^{\circ}$ C) increases blood flow in the neoplastic tissue, increasing the number of cytostatic drugs in the neoplastic tissue, and thus has an adjunctive effect on chemotherapy. In addition, blood flow increases the availability of oxygen to cancer cells. Neoplastic-cell hypoxia is related to radioresistance. Thus, increased blood flow also makes the tumor more susceptible to radiotherapy [78].

Further temperature increase (above 42 °C) is connected with additional effects. High temperature damages chaotically connected blood vessels and reduces blood perfusion in the cancerous tissue. This causes local hypoxia and acidification of the tumor, and eventually necrosis. However, the increase does not strongly affect healthy tissue [91]. In addition, hyperthermia can stimulate the immune system in the cancer environment by programming the immune system to recognize cancer cells [93,100,101].

#### 3.1.2. Magnetite Nanoparticles in Magnetic Hyperthermia

There are several problems associated with hyperthermia nowadays. The main problems are related to nonselective heating up of tissues and not reaching a high enough temperature [102,103]. Therefore, attempts have been made to overcome these problems. One of the most promising solutions is based on magnetic nanoparticles. They can strongly absorb microwave radiation [104] and transform it into heat. With precise delivery of microwave radiation to the desired place, localization of hyperthermia to the pathological site is possible. Among the magnetic nanoparticles, due to the magnetic properties and efficiency in energy dissipation (in this case, the conversion of magnetic energy into thermal energy), the most promising seems to be magnetite-based (Fe<sub>3</sub>O<sub>4</sub>) [105]. There are also other significant features of Fe<sub>3</sub>O<sub>4</sub> nanoparticles that can be used in treatment. These include targeting them to neoplastic cells through increased permeability and retention [106] and enabling highly specific active and passive targeting [107]. In addition, magnetite is easy to manipulate to obtain the desired effect. The heating of the magnetite depends on the frequency and amplitude of the applied magnetic field and the size, morphology, and composition of the magnetic nanoparticles [103,108]. In addition to microwave radiation (magnetic hyperthermia), magnetite can be induced with the use of a laser (photothermal) [109].

The use of ferrites, such as magnetite, in cancer treatment has been under investigation since the 1950s [7,110]. Since then, much laboratory research has been conducted on ferrites in cancer treatment [31,111]. The magnetite nanoparticles can be synthesized in different shapes; their size and surface functionalization can be easily controlled and modified. Accordingly, the possibility of their application in the magnetic hyperthermia field is widely tested. Recent research on the applicability of magnetite nanoparticles with controlled size and shape as hyperthermia agents is summarized in Table 2. As can be seen, according to the lack of a standardized measurement procedure, it is difficult to compare results obtained for the same material:  $Fe_3O_4$  nanoparticles. The specific absorption rate (SAR), defined as the power transformed into heat per unit of mass of nanoparticles, strongly depends on the alternative current (AC) fields parameters (frequency and field strength) and the magnetic fluid concentrations. However, the size and shape of magnetite also play a crucial role in the efficiency of magnetic hyperthermia. For example, magnetite nanorings are characterized by higher SAR values than nanotubes, while spherical-shaped magnetite nanoparticles can achieve SAR values of about 11 W/g for two times lower concentrations than magnetitetruncated octahedrons in similar AC magnetic fields parameters (see Table 2).

Shape	Nanoparticle Size (nm)	C (mg/mL)	f (kHz)	H (Oe)	SAR (Wg)	Referenc
truncated octahedron	$10\pm3$	10	112	250	8.4	[112]
nanorods	50–70 <sup>1</sup> , 8–10 <sup>2</sup>	5	184	261 395 522	69 154 246	[113]
nanorings	$\begin{array}{c} 67\pm21\ {}^3,41\pm18\ {}^4,\\ 93\pm27\ {}^5,26\pm12\ {}^6\end{array}$	0.6	300	560	~1000	[114]
nanotubes	$\begin{array}{c} 470\pm45{}^{1},110\pm20{}^{4},\\ 170\pm20{}^{5},55\pm5{}^{6}\end{array}$	1	300	400	80	[115]
nanorings	$55 \pm 5^{1}, 55 \pm 5^{4}, 110 \pm 15^{5}, 55 \pm 5^{6}$	1	300	400	100	- [115]
porous	<100	5	220	117	480	— [116]
porous/hollow	350	5	220	117	140	- [116]
	26 <sup>7</sup> , 225 <sup>2</sup>	0.1.8	400	400	400         420           600.6         5000	- [117]
nanodiscs	267,2252	0.1 <sup>8</sup>	488	600.6	5000	- [11/]
ellipsoidal	250 <sup>1</sup> , 50 <sup>2</sup>	5	765	300	759 <sup>9</sup>	[118]
	5.1	- 10		300 -	10.1	
polyhedral	14.3		100		42.2	- - [119]
polyneural	53.1		100		71.6	- [119]
	225.6	-			$ \begin{array}{r} 100\\ 480\\ 140\\ 420\\ 5000\\ 759^{9}\\ 10.1\\ 42.2\\ 71.6\\ 19.1\\ 11\\ 432.1^{9}\\ 330.8^{9}\\ 189.3^{9}\\ 900 \pm 22^{9}\\ 658 \pm 53^{9}\\ 160 \pm 1.5\\ \end{array} $	_
spherical	$11.4\pm3.2$	5	109.8	240	11	[120]
	$10\pm 2$	0.5	1001.1	125.4	432.1 <sup>9</sup>	
spherical			751.5	137.6	330.8 <sup>9</sup>	[121]
1			522.2	193.4	189.3 <sup>9</sup>	_
elongated	$12\pm3$	- 9	105	100 -	$900\pm22^{\:9}$	[100]
cuboidal	$15\pm2$	- 2°	$\begin{array}{cccc} 10 & 100 & 300 & \hline 42.2 \\ \hline 71.6 \\ \hline 19.1 \\ \hline 5 & 109.8 & 240 & 11 \\ \hline 5 & 109.8 & 240 & 11 \\ \hline 0.5 & 1001.1 & 125.4 & 432.1 \\ \hline 751.5 & 137.6 & 330.8 \\ \hline 522.2 & 193.4 & 189.3 \\ \hline 522.2 & 193.4 & 189.3 \\ \hline 2 \\ 2 \\ 8 \\ \hline 2 \\ 8 \\ \hline 435 & 193.5 \\ \hline 900 \pm 22 \\ \hline 658 \pm 53 \\ 9 \\ \hline 658 \pm 53 \\ 9 \\ \hline \end{array}$	$658\pm53~^9$	- [122]	
cuboidal	80.78 ± 3.37	5	315	300 400 500 600 700 800	$\begin{array}{c} 160 \pm 1.5 \\ 166 \pm 1.4 \\ 209 \pm 1.6 \\ 317 \pm 1.2 \\ 430 \pm 1.4 \\ 1036 \pm 1.5 \end{array}$	[123]

**Table 2.** Comparison of specific absorption rate (SAR) of magnetite nanoparticles with various shapes (C—magnetic fluid concentration, f—AC magnetic field frequency, H—magnetic field strength).

 $^1$  length,  $^2$  diameter,  $^3$  height,  $^4$  inner diameter,  $^5$  external diameter,  $^6$  wall thickness,  $^7$  thickness,  $^8$  Fe concentration in dispersion,  $^9$  W/g<sub>Fe</sub>.

The effect of ferrites has been studied in in vitro cell cultures, as well as in in vivo models, both in the case of magnetically induced hyperthermia [8,124] and photothermia [74,125]. Moreover, the hypothermic effect generated by ferrites was tested on organoids [126]. Several clinical trials have been performed using magnetite-based hyperthermia [127]. The first clinical use of interstitial hyperthermia using magnetite nanoparticles was reported in 2005 [128]. The procedure was performed on the prostate gland of a 67-year-old patient. In a technique also known as magnetic fluid hyperthermia, magnetite nanoparticles were injected into the target area and selectively heated by an externally applied alternating magnetic field [128]. According to the authors' knowledge, so far, magnetite nanoparticles have been used therapeutically in procedures performed on patients who have prostate cancer [128,129], melanoma [130], and glioblastoma [131]. Although many studies have proven the positive effect of ferrites (especially magnetites in cancer

treatment), it has not been possible to develop an appropriate methodology to allow for the broader use of magnetic hyperthermia. Many preclinical studies have been conducted, but few reach the third and final phase of clinical trials [8]. The year 2021 may be the turning point. NanoTherm<sup>®</sup> (treatment of glioblastoma) completed clinical phase 2a, and the NoCanTher project has started final clinical trials (treatment of advanced pancreatic ductal adenocarcinoma) [132].

## 4. Use of Magnetite Nanoparticles in Cancer Therapies

Besides the application of magnetite in magnetic hyperthermia, there are other proposed applications of these nanoparticles in cancer therapies. The nanoparticle's properties allow use for targeting cancer. Interaction between the magnetite particle and magnetic field is the backbone of targeting. Magnetite-carrying drugs can be pulled to the desired location in the body by the magnetic field. Thus, the magnetite nanoparticles described here are carriers for therapeutics. Several studies have been made to test the possible use of magnetite in the controlled delivery of drugs or gene and protein therapy [8].

## 4.1. Targeted Delivery of Medicinal Substances

Nowadays, the main problem of cancer treatments is nonselectivity toward cancer cells; interactions with healthy tissue cause numerous side effects [133]. In chemotherapy, cytostatic drugs block cell proliferation. Cancer cells are characteristic of rapid cell division and are strongly affected by cytostatic drugs. However, healthy cells also divide, and the therapeutics interact with them. The side effects of chemotherapy include hair loss, nausea and vomiting, associated loss of appetite, or bone-marrow disorders.

Additionally, new disease entities may develop even long after the end of chemotherapy, e.g., pulmonary tissue fibrosis, infertility, cardiac arrhythmias, and peripheral neuropathies [134,135]. Precise delivery of medicinal substances to the pathological site would reduce side effects. In addition, targeted drug delivery allows the use of a higher concentration of drugs than in the case of other therapies [136]. In traditional treatment strategies, high drug concentrations could be toxic and cause severe damage to healthy tissue [137]. Injecting therapeutic close to its target could help reach a high concentration; however, it may be dangerous for a patient [136].

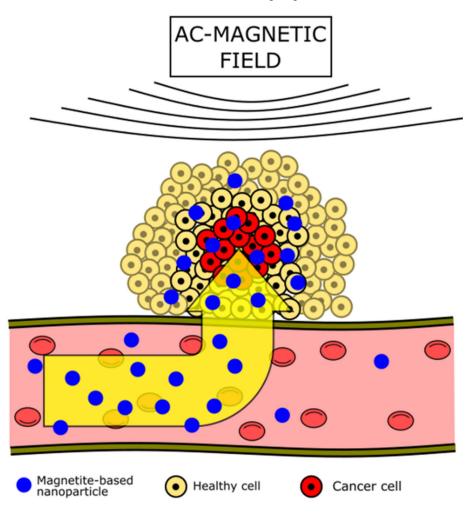
## 4.1.1. Nanopharmaceuticals

The magnetic particle can serve as a carrier in controlled drug delivery [8,133,138]. The implementation of nanoscale pharmaceutical products, i.e., nanopharmaceuticals, allows for the improvement of the physicochemical properties of not only existing drugs, but also new molecular units with biological activity [139]. In nanopharmaceuticals, active ingredients (APIs) are loaded into a nanocarrier to improve their solubility, extend the half-life, improve pharmacokinetic properties, obtain a modified release profile, and reduce their toxicity [139].

### 4.1.2. Nanopharmaceuticals in Magnetic Field for Targeted Drug Delivery

Therapeutics can then be directed to the desired location through a magnetic field [137]. The magnetic nanoparticle–drug complex is delivered by manipulating an external magnetic field using electromagnetic coils [137] to various types of permanent magnets [140] (Figure 2). In addition, nanoparticles can be functionalized, e.g., with folic-acid groups that interact with receptors specific for some tumors to increase specificity [141]. The potential of these delivery systems for anticancer drugs has been shown in several studies. For example, magnetite nanoparticles with attached carboplatin particles showed a high rate of inhibition of ovarian tumors without significant toxicity to healthy organs [142]. Similarly, promising research has been carried out on a multifunctional nanoplatform delivering cisplatin to cancer cells [143]. The platform consisted of biodegradable PLGA surface-functionalized by chitosan and polyvinyl alcohol with additional SPIONs. Tests showed increased release time of cisplatin to more than 10 days and confirmed the possibility of

using nanoparticles also as an imaging agent. Another study has shown that drug-coated polymer nanospheres and nanocapsules enhance intracellular anticancer effects [144]. The method of drug delivery into the body was also developed using multilayer microcapsules composed of poly (allylamine) and poly (sodium 4-styrene sulfonate). This method has been tested in in vitro and in vivo conditions [145].



**Figure 2.** Magnetite-based drug delivery occurs under the influence of an external AC magnetic field. Nanoparticles are pulled from a blood vessel to a pathological site and migrate toward the source of altering magnetic field. The pulled-out nanoparticles are concentrated in a tumorous site with minimal interaction with healthy cells. Then, release of the therapeutics load occurs.

However, the magnetic field could be used in drug delivery another way [146]. It was proposed to enclose drugs in a polymer–magnetic beads microparticle complex. In case of the absence of a magnetic field, the drug is slowly realized from a carrier. The drug realizes rate increases when the magnetic field is applied because pores are compressed [146].

#### 4.2. Cancer Genotherapy

Gene therapy aims to modify genetic material or correct genetic errors that cause diseases. It can be performed by injecting foreign genetic material into the body (in vivo gene therapy) [147,148] and if the modified cell is outside the patient's body, then reintroduced into the body (ex vivo gene therapy) [147–149]. Gene therapy can be used in therapies of many diseases; however, here we focus only on its use in oncology.

Several proposed oncological therapies are based on the modification of nucleic acids [148]. These include the suicide gene, immunization gene, oncogene silencing, tumor-suppressor-gene replacement, angiogenesis-targeting therapy, and others [148]. In

addition, the cancer vaccine, which is considered one of the most promising oncological therapies, is also based on DNA modifications [150]. However, despite numerous studies on the described techniques, the effectiveness of the proposed strategies still does not provide the full potential of gene therapy in oncology [148]. The main problem is the lack of an effective system to deliver genetic material to desired cells and tissue.

#### 4.2.1. Nanoparticles in Genotherapy

The introduction of therapeutic nucleic acids into the target cell can be done only using appropriate vectors. So far, several methods of delivering genetic material to the patient's body have been proposed. Among them are distinguished methods based on viral vectors and physical and chemical methods [147,151]. The use of the vectors proposed so far in cancer gene therapy is associated with immunogenicity, limited genetic burden, and cancer risk due to the insertion of a therapeutic load in the vicinity of genes responsible for controlling cell growth [152]. Consequently, there has been engagement in potentially using nanoparticles as vectors in cancer-gene therapy. Due to the simple synthesis and low cytotoxicity, nanoparticles seem to be ideal candidates.

Additionally, easy surface modification of the nanoparticles enables immune-systemresponse modulation [153]. Nanoparticles provide a new approach to gene vectors because they can wrap or adsorb nucleic acids on their surface. Specific targeting molecules, such as a monoclonal antibody, are then coupled to the particle surface, and the nanoparticles may be endocytosed or phagocytosed by cells, and the encapsulated therapeutic agents may be effective in a cell [49]. Some unique nanomaterials have magnetic, optical, and thermal properties that enable the delivery and controlled release of target genes [154]. By coating nanoparticles with hydrophilic polymers, it is possible to obtain a low level of adsorption to blood proteins, which helps to avoid phagocytosis [155]. Due to their low immunogenicity and easy cell-membrane penetration, nanoparticles are probably one of the most promising vectorization strategies for delivery, as reflected in the number of clinical trials proposed using nanovectors for cancer treatment [148]. Nanoparticles can be used with viral vectors, but as nonviral vectors [156].

## 4.2.2. Magnetite in Genotherapy

Nanovectors based on magnetite nanoparticles are also considered a potential vector in gene therapies. Initially, magnetite was tested with an adenoviral vector [157]. One of the limitations of adenoviral vectors is the low cell penetration due to the short time of exposure of the cell to the virus. Studies have shown that binding a vector to a magnetite nanoparticle can extend the exposure of the cell to the vector and thus also increase cell penetration [158]. However, magnetite has been studied as the primary vector. Magnetic transfection, or magnetofection, delivers genetic material to the cell using an external magnetic field [8,159]. Magnetite nanoparticles (with nucleic acids attached) are pulled by magnetic force to the tissue. In addition, magnetic force helps penetrate the cell membrane [160]. The presence of nanoparticles near the tissue also increases the absorption of the genetic material through endocytosis [161]. It was found that such use of the magnetic field can significantly increase the efficiency of transfection of foreign genetic material into the cell [162]. Naked magnetite particles or particles coated with, e.g., polyethyleneimine (PEI), to which plasmids, short hairpin RNA (shRNA), or antisense oligonucleotides are attached [8]. Numerous in vitro studies confirm the effectiveness of magnetofection based on SPIONs, both in animals and plants [162–168]. However, there are no clinically approved methods of delivering genetic material based on nanoparticles, and their use is limited to laboratory tests [148,156].

#### 4.3. Protein Therapy

In cancer therapies, the goal of protein therapy is to deliver a sufficient dose of the therapeutic protein to cancer cells [59]. It is considered one of the most direct and safe cancer therapies described so far [59]. Protein therapy may include blocking a cell-surface receptor involved in carcinogenesis, intracellular delivery of proteins involved in cell

signaling, inhibiting growth and invasion of cancer cells, or inducing apoptosis [59,169]. Capsule methods have been developed to maintain the function of the therapeutic protein introduced into the body [169]. However, the targeted delivery and tracking of encapsulated therapeutic proteins to the desired cells remain challenging. It turns out that the use of nanoparticles can result in the effective delivery of the therapeutic protein to cancer cells while maintaining their enzymatic activity [169,170]. Nanoparticles can protect the proteins carried against the activity of proteases or facilitate their penetration into the cell [59].

According to the authors' knowledge, magnetite nanoparticles were not clinically used in protein therapy, and magnetite was tested in in vivo conditions. In the mouse model, the therapy based on magnetite nanoparticles and the combination with chlorotoxin inhibited the invasiveness of glioblastoma [171]. In addition, enhancement of the drug effect in target cells using nanoparticles was also described [171]. This is because the therapeutic protein can be distributed explicitly on the surface of the nanoparticles. Another example is the use of iron oxide to deliver an antibody. The antibody effectively inhibited the mutant EGFR receptor [172]. In addition, the encapsulation of cytochrome c in the nanoparticle cavities induced apoptosis after delivery to tumor cells [169].

### 5. Challenges in the Development of Magnetite-Based Therapies

The development of new cancer therapies, especially selective strategies, is connected with several well-known problems that make it challenging to develop a uniform methodology. These include heterogeneity of tumors, differences between the human and mouse models used in in vivo studies and differences between neoplastic lesions [15]. In addition, there are several problems connected with the use of magnetite nanoparticles in magnetic hyperthermia that require careful further studies. These unsolved problems are related to hyperthermia, general problems with nanoparticles, and technological and methodological aspects of the synthesis and use of magnetite.

In recent years, hyperthermia has been used to support radiotherapy, chemotherapy, and immunotherapy, increasing their effectiveness [78,173]. However, hyperthermia still lacks a uniform definition in oncology. This makes the standardization of methodology impossible. It results in variation between studies, which gives contradictory results [2]. It also makes it impossible to calculate and optimize the appropriate dose of energy [2]. The exact mechanism of hyperthermia-mediated cancer treatment has to be further studied. Especially one aspect of hyperthermia in oncology needs to look deeper. Studies on the effects of hyperthermia on drug resistance show opposite results and suggest that, in some cases, hyperthermia may stimulate the development of resistance [88].

Although the properties of nanoparticles show promise from a clinical point of view, they require further research. Unfortunately, nanotechnology develops faster than legislature and it creates problem of lack of standardization of nomenclature and test methodology. There is great need for guidelines for safety assessment of nanomaterials [174]. Detailed understanding of how nanoparticles interact with cells, proteins, hormones, or immune factors is fundamental to their commercial and clinical application [104]. First of all, it is unknown what happens with nanoparticles when they are metabolized in the body. Do nanoparticles, or their metabolites, accumulate in cells or organs, thereby triggering intracellular changes, inflammatory responses, and the development of oxidative stress? It has been suggested that nanoparticles can be transported in the body by the neuronal route [175], which may contribute to the degeneration of the nervous system. Increased ROS production, induced by nanoparticles, may damage organelles and contribute to the increased aggregation of proteins involved in the pathogenesis of Alzheimer's disease and Parkinson's disease [176–178]. Some studies suggest that nanoparticles may affect the patient's endothelium and contribute to the development of cardiovascular diseases [179].

Moreover, they can enter the blood cells, which can damage morphotic elements of the blood. There are also reports of an allergic reaction following the use of nanoparticles [180,181]. Nanoparticles can also absorb biological agents such as bacterial endotoxins on their surface, which may also have an unknown risk [182].

Cytotoxicity of nanomaterials depends on the concentration, proportion, porosity, aggregation tendency, a chemical affinity for biological structures, chemical surface reactivity and even the shape, size of nanoparticles, and environmental conditions [183,184]. So far, many studies have been conducted on the cytotoxicity of magnetite [31,111,138,185–188]. First of all, it has been noticed that the previously mentioned particle size influences the cytotoxicity of magnetite. When using chicken macrophages, it was found that smaller magnetite particles (60, 120, and 250 nm were used) were more toxic to these cells [188]. No significant toxicity of particles with a size of 30 µm was detected on HeLa cells and postnatal human fibroblasts [138]. Interestingly, it has been shown that magnetite can be toxic to fibroblasts; however, the same concentration of nanoparticles does not affect leukocytes in the tested parameters [187]. Farcas et al. [111] found that small magnetite particles (up to 30 nm) are not toxic to human keranocytes and primary endodermal melanocytes but may be toxic to melanoma cells, both in the human and murine cell lines [111]. However, modifications of magnetite nanoparticles can eliminate problem of cytotoxicity [189]. An example is the study in which the cytotoxicity of nanoparticles based on the magnetite core was reduced when used with an oleic-acid shell [189].

The synthesis of magnetite is a leading technical problem and requires improvement [190]. It is challenging to obtain magnetite particles of appropriate and the same size with known synthesis methods. For extensive use of magnetite, it is also needed to reduce the cost of synthesis. Large-scale synthesis or the laboriousness of coating nanoparticles are also limiting factors [132,190]. Methods of magnetite synthesis based on the co-precipitation method allow nanoparticles with high efficiency; however, the obtained structures are strongly agglomerated [191].

On the other hand, methods based on the high-temperature decomposition of organic precursors allow for the obtaining of monodisperse nanoparticles, albeit with low efficiency compared to the reaction time [191,192]. The most frequently used method of synthesizing nanoparticles with high application potential (e.g., magnetic hyperthermia and targeted therapies) is the polyol method [193]. However, a much lower efficiency characterizes it than in the case of the coprecipitation method. It is also much more challenging to control the nucleation and growth of nanoparticles than reactions in high-boiling liquids and using, e.g., oleic acid or stearic acid [194,195]. The major problem in targeted drug delivery based on magnetite is the limited packaging capacity of the nanoparticles. Here, using magnetite together with other materials could be an option. The most promising seems to be a combination of nanoparticles and hydrogels as a drug carrier [196].

Another noteworthy obstacle connected with a large-scale synthesis of therapeutic is fact that this is clinically useful only when the quality of obtained therapeutic is unchanged between batches of manufactured product. Therefore, a reproducible, scalable-production-method nanoparticle-based therapeutic must be developed and validated [139]. Here, good manufacturing practices of nanotherapeutics should ensure that guidelines recommended by agencies that control the authorization and licensing of the pharmaceutics [139].

The clinical methodology also needs improvement. The main problem limiting the effectiveness of magnetic hyperthermia is low penetration of the magnetic field, i.e., up to 5 cm into the body due to the loss of the magnetic field gradient [133]. As a result, magnetite can only be activated in subcutaneous tissues. There are attempts to use magnetic implants that generate sufficient magnetic field strength to attract magnetic nanoparticles carrying drugs [133]. However, even with implants, an external magnetic field is required to obtain the desired effect [197]. In addition, if a patient has metal implants, e.g., of the hip joint, or a pacemaker or implantable heart defibrillator, the procedure cannot be performed [198].

Additionally, in the case of prostate cancer, the temperatures typically reached inside the tumor were generally in the range of 40–41 °C. Only a few of these studies have reached maximum intraprostate temperatures of 42.5 °C or exceeded the critical temperature of 43.0 °C. The temperature distribution in the prostate gland is heterogeneous, so reliable temperature calculations during prostate-cancer hyperthermia are challenging to obtain. Due to local differences in tissue properties, excessive temperature rise can occur, resulting in burns, blisters, pain, thrombosis, bleeding, and other undesirable effects [89]. Therefore, the region affected by the temperature rise must be strictly limited to the desired tumor volume. In the case of local-regional hyperthermia, it is necessary to diagnose all tumors. If not all lesions are recognized, unrecognized lesions are left untreated [2]. This phenomenon is fundamental when the magnetic field is intended to activate drug release from nanoparticle carriers.

### 6. Conclusions

Tumor-selective therapies should be the main oncological strategies in the future. Nanoparticles' properties could be helpful in the development of such methods. Magnetite seems to be one of the most promising nanoparticles in oncology. This is mainly because the nanoparticle can be used in different ways, and all of the proposed magnetitebased strategies selectively interact with tumors. The therapeutic application of magnetite nanoparticles has been under investigation for many years with no success. Most of the tests ended at the preclinical phase [8]. There are still several obstacles to the wide use of magnetite in medicine. However, nanoparticles are expected to revolutionize medicine and pharmacy, resulting in nanomedicine being an intensively developing branch of medicine. Many problems connected with safe use and nanoparticles synthesis will probably be solved in the near future. These problems can be connected with a few aspects. First of all, the hyperthermia measurements should be standardized. The presented research in this field currently cannot be easily compared to decide which way (such as surface, shape, or size modifications) should be tested in the future to develop highly biocompatible particles characterized by high magnetically induced hyperthermia effect. Secondly, new synthesis methods of iron oxides should be developed. These methods should be characterized by repeatability, relatively high yield, and most critically, scalability. The currently tested methods based on the coprecipitation and high-temperature degradation processes result in the synthesis of agglomerated nanoparticles or synthesis of nanoparticles with low yield but characterized by uniform size and shape. Above that, further research should also concentrate on biocompatibility, elimination from the body, and the long-term effects of nanoparticles on cells. There are many contradicting literature reports on these aspects, for example, in the case of the potential use of cobalt ferrite nanoparticles.

**Author Contributions:** Conceptualization, S.G. and A.W.; formal analysis, K.B.-R.; writing—original draft preparation, A.W., S.G., and A.R.; writing—review and editing, A.W., S.G., A.R., and K.B.-R.; supervision, A.R. and K.B.-R.; funding acquisition, K.B-R. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

## References

- Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA A Cancer J. Clin. 2021, 71, 209–249. [CrossRef] [PubMed]
- Lee, S.-Y.; Fiorentini, G.; Szasz, A.M.; Szigeti, G.; Szasz, A.; Minnaar, C.A. Quo Vadis Oncological Hyperthermia (2020)? Front. Oncol. 2020, 10, 1690. [CrossRef] [PubMed]
- Yu, Z.; Gao, L.; Chen, K.; Zhang, W.; Zhang, Q.; Li, Q.; Hu, K. Nanoparticles: A New Approach to Upgrade Cancer Diagnosis and Treatment. *Nanoscale Res. Lett.* 2021, 16, 88. [CrossRef]
- Baudino, T.A. Targeted Cancer Therapy: The Next Generation of Cancer Treatment. Curr. Drug. Discov. Technol. 2015, 12, 3–20. [CrossRef] [PubMed]
- Groenendijk, F.H.; Bernards, R. Drug Resistance to Targeted Therapies: Déjà vu All over Again. Mol. Oncol. 2014, 8, 1067–1083. [CrossRef] [PubMed]
- 6. Racca, L.; Cauda, V. Remotely Activated Nanoparticles for Anticancer Therapy. Nanomicro. Lett. 2020, 13, 11. [CrossRef]
- Liu, X.; Zhang, Y.; Wang, Y.; Zhu, W.; Li, G.; Ma, X.; Zhang, Y.; Chen, S.; Tiwari, S.; Shi, K.; et al. Comprehensive Understanding of Magnetic Hyperthermia for Improving Antitumor Therapeutic Efficacy. *Theranostics* 2020, 10, 3793–3815. [CrossRef]

- 8. Nuzhina, J.V.; Shtil, A.A.; Prilepskii, A.Y.; Vinogradov, V.V. Preclinical Evaluation and Clinical Translation of Magnetite-Based Nanomedicines. *J. Drug Delivery Sci. Technol.* **2019**, *54*, 101282. [CrossRef]
- National Center for Biotechnology Information (NCBI). [Internet]. National Library of Medicine (US), National Center for Biotechnology Information: Bethesda, MD, USA; [1988] – [Cited 2022]. Available online: https://www.ncbi.nlm.nih.gov/ (accessed on 18 May 2022).
- 10. Mohanraj, V.J.; Chen, Y. Nanoparticles—A Review. Trop. J. Pharm. Res. 2006, 5, 561–573. [CrossRef]
- 11. Linsinger, T.; Roebben, G.; Gilliland, D.; Calzolai, L.; Rossi, F.; Gibson, P.; Klein, C. *Requirements on Measurements for the Implementation of the European Commission Definition of the Term "Nanomaterial"*.; Publications Office of the European Union: Luxembourg, 2012.
- 12. Zhu, K.; Ju, Y.; Xu, J.; Yang, Z.; Gao, S.; Hou, Y. Magnetic Nanomaterials: Chemical Design, Synthesis, and Potential Applications. *Acc. Chem. Res.* 2018, *51*, 404–413. [CrossRef]
- 13. Cheng, Y.; Morshed, R.A.; Auffinger, B.; Tobias, A.L.; Lesniak, M.S. Multifunctional Nanoparticles for Brain Tumor Imaging and Therapy. *Adv. Drug. Deliv. Rev.* 2014, *66*, 42–57. [CrossRef] [PubMed]
- Navya, P.N.; Kaphle, A.; Srinivas, S.P.; Bhargava, S.K.; Rotello, V.M.; Daima, H.K. Current Trends and Challenges in Cancer Management and Therapy Using Designer Nanomaterials. *Nano Convergence*. 2019, 6, 23. [CrossRef] [PubMed]
- 15. Zhang, W.; Kohane, D.S. Keeping Nanomedicine on Target. Nano. Lett. 2021, 21, 3–5. [CrossRef] [PubMed]
- Chandrashekhar, V.G.; Senthamarai, T.; Kadam, R.G.; Malina, O.; Kašlík, J.; Zbořil, R.; Gawande, M.B.; Jagadeesh, R.V.; Beller, M. Silica-Supported Fe/Fe–O Nanoparticles for the Catalytic Hydrogenation of Nitriles to Amines in the Presence of Aluminium Additives. *Nat. Catal.* 2022, *5*, 20–29. [CrossRef]
- 17. da Silva, A.G.M.; Rodrigues, T.S.; Wang, J.; Camargo, P.H.C. Plasmonic Catalysis with Designer Nanoparticles. *Chem. Commun.* **2022**, *58*, 2055–2074. [CrossRef]
- Gogotsi, Y.; Penner, R.M. Energy Storage in Nanomaterials—Capacitive, Pseudocapacitive, or Battery-Like? ACS Nano 2018, 12, 2081–2083. [CrossRef]
- 19. Łukowiec, D.; Wasiak, T.; Janas, D.; Drzymała, E.; Depciuch, J.; Tarnawski, T.; Kubacki, J.; Wacławek, S.; Radoń, A. Pd Decorated Co–Ni Nanowires as a Highly Efficient Catalyst for Direct Ethanol Fuel Cells. *Int. J. Hydrogen Energy* **2021**. [CrossRef]
- Wu, Z.-P.; Caracciolo, D.T.; Maswadeh, Y.; Wen, J.; Kong, Z.; Shan, S.; Vargas, J.A.; Yan, S.; Hopkins, E.; Park, K.; et al. Alloying– Realloying Enabled High Durability for Pt–Pd-3d-Transition Metal Nanoparticle Fuel Cell Catalysts. *Nat. Commun.* 2021, 12, 859. [CrossRef]
- 21. Lozano, G.; Rodriguez, S.R.; Verschuuren, M.A.; Gómez Rivas, J. Metallic Nanostructures for Efficient LED Lighting. *Light Sci. Appl.* **2016**, *5*, e16080. [CrossRef]
- 22. Jiang, Y.; Zheng, W.; Tran, K.; Kamilar, E.; Bariwal, J.; Ma, H.; Liang, H. Hydrophilic Nanoparticles That Kill Bacteria While Sparing Mammalian Cells Reveal the Antibiotic Role of Nanostructures. *Nat. Commun.* **2022**, *13*, 197. [CrossRef]
- Escobar, A.; Muzzio, N.; Moya, S.E. Antibacterial Layer-by-Layer Coatings for Medical Implants. *Pharmaceutics* 2021, 13, 16. [CrossRef] [PubMed]
- McClements, J.; Bar, L.; Singla, P.; Canfarotta, F.; Thomson, A.; Czulak, J.; Johnson, R.E.; Crapnell, R.D.; Banks, C.E.; Payne, B.; et al. Molecularly Imprinted Polymer Nanoparticles Enable Rapid, Reliable, and Robust Point-of-Care Thermal Detection of SARS-CoV-2. ACS Sens. 2022, 7, 1122–1131. [CrossRef] [PubMed]
- Gomez-Marquez, J.; Hamad-Schifferli, K. Local Development of Nanotechnology-Based Diagnostics. Nat. Nanotechnol. 2021, 16, 484–486. [CrossRef] [PubMed]
- 26. Xu, L.; Liang, H.-W.; Yang, Y.; Yu, S.-H. Stability and Reactivity: Positive and Negative Aspects for Nanoparticle Processing. *Chem. Rev.* **2018**, *118*, 3209–3250. [CrossRef]
- Patra, J.K.; Das, G.; Fraceto, L.F.; Campos, E.V.R.; del P. Rodriguez-Torres, M.; Acosta-Torres, L.S.; Diaz-Torres, L.A.; Grillo, R.; Swamy, M.K.; Sharma, S.; et al. Nano Based Drug Delivery Systems: Recent Developments and Future Prospects. *J. Nanobiotechnol.* 2018, 16, 71. [CrossRef] [PubMed]
- 28. Zhou, Q.; Zhang, L.; Yang, T.; Wu, H. Stimuli-Responsive Polymeric Micelles for Drug Delivery and Cancer Therapy. *Int. J. Nanomed.* **2018**, *13*, 2921–2942. [CrossRef] [PubMed]
- Kesavan, M.P.; Kotla, N.G.; Ayyanaar, S.; Kumar, G.G.V.; Rajagopal, G.; Sivaraman, G.; Webster, T.J.; Rajesh, J. A Theranostic Nanocomposite System Based on Iron Oxide-Drug Nanocages for Targeted Magnetic Field Responsive Chemotherapy. *Nanomedicine* 2018, 14, 1643–1654. [CrossRef]
- Ryu, J.H.; Koo, H.; Sun, I.-C.; Yuk, S.H.; Choi, K.; Kim, K.; Kwon, I.C. Tumor-Targeting Multi-Functional Nanoparticles for Theragnosis: New Paradigm for Cancer Therapy. *Adv. Drug Deliv. Rev.* 2012, 64, 1447–1458. [CrossRef]
- Soleymani, M.; Khalighfard, S.; Khodayari, S.; Khodayari, H.; Kalhori, M.R.; Hadjighassem, M.R.; Shaterabadi, Z.; Alizadeh, A.M. Effects of Multiple Injections on the Efficacy and Cytotoxicity of Folate-Targeted Magnetite Nanoparticles as Theranostic Agents for MRI Detection and Magnetic Hyperthermia Therapy of Tumor Cells. *Sci. Rep.* 2020, *10*, 1695. [CrossRef]
- 32. Chiang, C.-L.; Cheng, M.-H.; Lin, C.-H. From Nanoparticles to Cancer Nanomedicine: Old Problems with New Solutions. *Nanomaterials* **2021**, *11*, 1727. [CrossRef]
- Sulaiman, G.M.; Waheeb, H.M.; Jabir, M.S.; Khazaal, S.H.; Dewir, Y.H.; Naidoo, Y. Hesperidin Loaded on Gold Nanoparticles as a Drug Delivery System for a Successful Biocompatible, Anti-Cancer, Anti-Inflammatory and Phagocytosis Inducer Model. *Sci. Rep.* 2020, 10, 9362. [CrossRef] [PubMed]

- Al Tamimi, S.; Ashraf, S.; Abdulrehman, T.; Parray, A.; Mansour, S.A.; Haik, Y.; Qadri, S. Synthesis and Analysis of Silver–Copper Alloy Nanoparticles of Different Ratios Manifest Anticancer Activity in Breast Cancer Cells. *Cancer Nanotechnol.* 2020, 11, 13. [CrossRef]
- Kang, S.; Kang, K.; Chae, A.; Kim, Y.-K.; Jang, H.; Min, D.-H. Fucoidan-Coated Coral-like Pt Nanoparticles for Computed Tomography-Guided Highly Enhanced Synergistic Anticancer Effect against Drug-Resistant Breast Cancer Cells. *Nanoscale* 2019, 11, 15173–15183. [CrossRef] [PubMed]
- Yang, Y.; Chen, M.; Wu, Y.; Wang, P.; Zhao, Y.; Zhu, W.; Song, Z.; Zhang, X.-B. Ultrasound Assisted One-Step Synthesis of Au@Pt Dendritic Nanoparticles with Enhanced NIR Absorption for Photothermal Cancer Therapy. *RSC Adv.* 2019, *9*, 28541–28547. [CrossRef] [PubMed]
- Ramalingam, V.; Raja, S.; Harshavardhan, M. In Situ One-Step Synthesis of Polymer-Functionalized Palladium Nanoparticles: An Efficient Anticancer Agent against Breast Cancer. *Dalton Trans.* 2020, 49, 3510–3518. [CrossRef]
- Dragoi, B.; Uritu, C.M.; Agrigoroaie, L.; Lutic, D.; Hulea, V.; Postole, G.; Coroaba, A.; Carasevici, E. MnAl-Layered Double Hydroxide Nanosheets Infused with Fluorouracil for Cancer Diagnosis and Therapy. ACS Appl. Nano Mater. 2021, 4, 2061–2075. [CrossRef]
- Sathishkumar, P.; Li, Z.; Govindan, R.; Jayakumar, R.; Wang, C.; Long Gu, F. Zinc Oxide-Quercetin Nanocomposite as a Smart Nano-Drug Delivery System: Molecular-Level Interaction Studies. *Appl. Surface Sci.* 2021, 536, 147741. [CrossRef]
- Gawali, S.L.; Shelar, S.B.; Gupta, J.; Barick, K.C.; Hassan, P.A. Immobilization of Protein on Fe3O4 Nanoparticles for Magnetic Hyperthermia Application. Int. J. Biol. Macromol. 2021, 166, 851–860. [CrossRef]
- 41. Wu, Z.; Zhang, P.; Wang, P.; Wang, Z.; Luo, X. Using Copper Sulfide Nanoparticles as Cross-Linkers of Tumor Microenvironment Responsive Polymer Micelles for Cancer Synergistic Photo-Chemotherapy. *Nanoscale* **2021**, *13*, 3723–3736. [CrossRef]
- 42. Qi, L.; Pan, T.; Ou, L.; Ye, Z.; Yu, C.; Bao, B.; Wu, Z.; Cao, D.; Dai, L. Biocompatible Nucleus-Targeted Graphene Quantum Dots for Selective Killing of Cancer Cells via DNA Damage. *Commun. Biol.* **2021**, *4*, 214. [CrossRef]
- Gong, B.; Shen, Y.; Li, H.; Li, X.; Huan, X.; Zhou, J.; Chen, Y.; Wu, J.; Li, W. Thermo-Responsive Polymer Encapsulated Gold Nanorods for Single Continuous Wave Laser-Induced Photodynamic/Photothermal Tumour Therapy. *J. Nanobiotechnol.* 2021, 19, 41. [CrossRef]
- Sharifi, M.; Hasan, A.; Nanakali, N.M.Q.; Salihi, A.; Qadir, F.A.; Muhammad, H.A.; Shekha, M.S.; Aziz, F.M.; Amen, K.M.; Najafi, F.; et al. Combined Chemo-Magnetic Field-Photothermal Breast Cancer Therapy Based on Porous Magnetite Nanospheres. *Sci. Rep.* 2020, *10*, 5925. [CrossRef] [PubMed]
- Wang, H.; Liang, Y.; Yin, Y.; Zhang, J.; Su, W.; White, A.M.; Jiang, B.; Xu, J.; Zhang, Y.; Stewart, S.; et al. Carbon Nano-Onion-Mediated Dual Targeting of P-Selectin and P-Glycoprotein to Overcome Cancer Drug Resistance. *Nat. Commun.* 2021, 12, 312. [CrossRef] [PubMed]
- Zhang, N.; Wang, Y.; Zhang, C.; Fan, Y.; Li, D.; Cao, X.; Xia, J.; Shi, X.; Guo, R. LDH-Stabilized Ultrasmall Iron Oxide Nanoparticles as a Platform for Hyaluronidase-Promoted MR Imaging and Chemotherapy of Tumors. *Theranostics* 2020, 10, 2791–2802. [CrossRef] [PubMed]
- 47. Manivannan, K.; Cheng, C.-C.; Anbazhagan, R.; Tsai, H.-C.; Chen, J.-K. Fabrication of Silver Seeds and Nanoparticle on Core-Shell Ag@SiO<sub>2</sub> Nanohybrids for Combined Photothermal Therapy and Bioimaging. *J. Colloid Interface Sci.* **2019**, 537, 604–614. [CrossRef]
- 48. Akbarzadeh, A.; Samiei, M.; Davaran, S. Magnetic Nanoparticles: Preparation, Physical Properties, and Applications in Biomedicine. *Nanoscale Res. Lett.* **2012**, *7*, 144. [CrossRef]
- 49. Brigger, I.; Dubernet, C.; Couvreur, P. Nanoparticles in Cancer Therapy and Diagnosis. *Adv. Drug Deliv. Rev.* 2002, 54, 631–651. [CrossRef]
- Pistone, A.; Iannazzo, D.; Panseri, S.; Montesi, M.; Tampieri, A.; Galvagno, S. Hydroxyapatite-Magnetite-MWCNT Nanocomposite as a Biocompatible Multifunctional Drug Delivery System for Bone Tissue Engineering. *Nanotechnology* 2014, 25, 425701. [CrossRef]
- 51. Weissleder, R.; Stark, D.D.; Engelstad, B.L.; Bacon, B.R.; Compton, C.C.; White, D.L.; Jacobs, P.; Lewis, J. Superparamagnetic Iron Oxide: Pharmacokinetics and Toxicity. *AJR Am. J. Roentgenol.* **1989**, *152*, 167–173. [CrossRef]
- 52. Mahmoudi, M.; Sant, S.; Wang, B.; Laurent, S.; Sen, T. Superparamagnetic Iron Oxide Nanoparticles (SPIONs): Development, Surface Modification and Applications in Chemotherapy. *Adv. Drug Deliv. Rev.* **2011**, *63*, 24–46. [CrossRef]
- Natarajan, S.; Harini, K.; Gajula, G.P.; Sarmento, B.; Neves-Petersen, M.T.; Thiagarajan, V. Multifunctional Magnetic Iron Oxide Nanoparticles: Diverse Synthetic Approaches, Surface Modifications, Cytotoxicity towards Biomedical and Industrial Applications. *BMC Mater.* 2019, 1, 2. [CrossRef]
- Radoń, A.; Hawełek, Ł.; Łukowiec, D.; Kubacki, J.; Włodarczyk, P. Dielectric and Electromagnetic Interference Shielding Properties of High Entropy (Zn, Fe, Ni, Mg, Cd)Fe<sub>2</sub>O<sub>4</sub> Ferrite. *Sci. Rep.* 2019, *9*, 20078. [CrossRef]
- Roca, A.G.; Gutiérrez, L.; Gavilán, H.; Fortes Brollo, M.E.; Veintemillas-Verdaguer, S.; del P. Morales, M. Design Strategies for Shape-Controlled Magnetic Iron Oxide Nanoparticles. *Adva. Drug Deliv. Rev.* 2019, 138, 68–104. [CrossRef] [PubMed]
- 56. Farazi, R.; Vaezi, M.R.; Molaei, M.J.; Saeidifar, M.; Behnam-Ghader, A.A. Effect of PH and Temperature on Doxorubicin Hydrochloride Release from Magnetite/Graphene Oxide Nanocomposites. *Mater. Today Proc.* **2018**, *5*, 15726–15732. [CrossRef]
- 57. Keshavarz, H.; Khavandi, A.; Alamolhoda, S.; Naimi-Jamal, M.R. PH-Sensitive Magnetite Mesoporous Silica Nanocomposites for Controlled Drug Delivery and Hyperthermia. *RSC Adv.* 2020, *10*, 39008–39016. [CrossRef]

- Zhang, J.L.; Srivastava, R.S.; Misra, R.D.K. Core–Shell Magnetite Nanoparticles Surface Encapsulated with Smart Stimuli-Responsive Polymer: Synthesis, Characterization, and LCST of Viable Drug-Targeting Delivery System. *Langmuir* 2007, 23, 6342–6351. [CrossRef]
- 59. Kievit, F.M.; Zhang, M. Surface Engineering of Iron Oxide Nanoparticles for Targeted Cancer Therapy. Acc. Chem. Res. 2011, 44, 853–862. [CrossRef]
- 60. Prasad, C.; Tang, H.; Liu, W. Magnetic Fe<sub>3</sub>O<sub>4</sub> Based Layered Double Hydroxides (LDHs) Nanocomposites (Fe<sub>3</sub>O<sub>4</sub>/LDHs): Recent Review of Progress in Synthesis, Properties and Applications. *J. Nanostruct. Chem.* **2018**, *8*, 393–412. [CrossRef]
- Fedorenko, S.; Stepanov, A.; Zairov, R.; Kaman, O.; Amirov, R.; Nizameev, I.; Kholin, K.; Ismaev, I.; Voloshina, A.; Sapunova, A.; et al. One-Pot Embedding of Iron Oxides and Gd(III) Complexes into Silica Nanoparticles—Morphology and Aggregation Effects on MRI Dual Contrasting Ability. *Colloids Surf. A Physicochem. Eng. Aspects* 2018, 559, 60–67. [CrossRef]
- Stepanov, A.; Burilov, V.; Pinus, M.; Mustafina, A.; Rümmeli, M.H.; Mendez, R.G.; Amirov, R.; Lukashenko, S.; Zvereva, E.; Katsuba, S.; et al. Water Transverse Relaxation Rates in Aqueous Dispersions of Superparamagnetic Iron Oxide Nanoclusters with Diverse Hydrophilic Coating. *Colloids Surf. A Physicochem. Eng. Aspects* 2014, 443, 450–458. [CrossRef]
- Daou, T.J.; Grenèche, J.M.; Pourroy, G.; Buathong, S.; Derory, A.; Ulhaq-Bouillet, C.; Donnio, B.; Guillon, D.; Begin-Colin, S. Coupling Agent Effect on Magnetic Properties of Functionalized Magnetite-Based Nanoparticles. *Chem. Mater.* 2008, 20, 5869–5875. [CrossRef]
- 64. Fischer, M.; Tamariz-Ellemann, A.; Gliemann, L. Hyperthermia: The Hotter the Body, the Hungrier the Brain? J. Physiol. 2020, 598, 2053–2054. [CrossRef] [PubMed]
- Datta, N.R.; Ordóñez, S.G.; Gaipl, U.S.; Paulides, M.M.; Crezee, H.; Gellermann, J.; Marder, D.; Puric, E.; Bodis, S. Local Hyperthermia Combined with Radiotherapy And-/or Chemotherapy: Recent Advances and Promises for the Future. *Cancer Treat. Rev.* 2015, 41, 742–753. [CrossRef] [PubMed]
- 66. Stitt, J.T. Fever versus Hyperthermia. Fed. Proc. 1979, 38, 39–43.
- 67. Dutz, S.; Hergt, R. Magnetic Nanoparticle Heating and Heat Transfer on a Microscale: Basic Principles, Realities and Physical Limitations of Hyperthermia for Tumour Therapy. *Int. J. Hyperthermia.* **2013**, *29*, 790–800. [CrossRef]
- 68. van der Zee, J. Heating the Patient: A Promising Approach? Ann. Oncol. 2002, 13, 1173–1184. [CrossRef]
- 69. Karwacki, J.H.; Nawrot, I.; Skalski, A.; Woźniewicz, B. Termoablacja Prądem Wysokiej Częstotliwości—Zastosowanie Kliniczne. *Adv. Clin. Exp. Med.* **2006**, 153–156.
- Kok, H.P.; Cressman, E.N.K.; Ceelen, W.; Brace, C.L.; Ivkov, R.; Grüll, H.; ter Haar, G.; Wust, P.; Crezee, J. Heating Technology for Malignant Tumors: A Review. Int. J. Hyperthermia 2020, 37, 711–741. [CrossRef]
- Peeken, J.C.; Vaupel, P.; Combs, S.E. Integrating Hyperthermia into Modern Radiation Oncology: What Evidence Is Necessary? Front. Oncol. 2017, 7, 132. [CrossRef]
- Baldi, G.; Lorenzi, G.; Ravagli, C. Hyperthermic Effect of Magnetic Nanoparticles under Electromagnetic Field. Proc. Appl. Ceramics 2009, 3. [CrossRef]
- Jabalera, Y.; Sola-Leyva, A.; Carrasco-Jiménez, M.P.; Iglesias, G.R.; Jimenez-Lopez, C. Synergistic Photothermal-Chemotherapy Based on the Use of Biomimetic Magnetic Nanoparticles. *Pharmaceutics* 2021, 13, 625. [CrossRef] [PubMed]
- Plan Sangnier, A.; Preveral, S.; Curcio, A.; K A Silva, A.; Lefèvre, C.T.; Pignol, D.; Lalatonne, Y.; Wilhelm, C. Targeted Thermal Therapy with Genetically Engineered Magnetite Magnetosomes@RGD: Photothermia Is Far More Efficient than Magnetic Hyperthermia. J. Control Release 2018, 279, 271–281. [CrossRef] [PubMed]
- Bertuit, E.; Benassai, E.; Mériguet, G.; Greneche, J.-M.; Baptiste, B.; Neveu, S.; Wilhelm, C.; Abou-Hassan, A. Structure–Property– Function Relationships of Iron Oxide Multicore Nanoflowers in Magnetic Hyperthermia and Photothermia. ACS Nano 2022, 16, 271–284. [CrossRef]
- 76. Espinosa, A.; Kolosnjaj-Tabi, J.; Abou-Hassan, A.; Plan Sangnier, A.; Curcio, A.; Silva, A.K.A.; Di Corato, R.; Neveu, S.; Pellegrino, T.; Liz-Marzán, L.M.; et al. Magnetic (Hyper)Thermia or Photothermia? Progressive Comparison of Iron Oxide and Gold Nanoparticles Heating in Water, in Cells, and In Vivo. *Adv. Funct. Mater.* 2018, 28, 1803660. [CrossRef]
- 77. Nemec, S.; Kralj, S.; Wilhelm, C.; Abou-Hassan, A.; Rols, M.-P.; Kolosnjaj-Tabi, J. Comparison of Iron Oxide Nanoparticles in Photothermia and Magnetic Hyperthermia: Effects of Clustering and Silica Encapsulation on Nanoparticles' Heating Yield. *Appl. Sci.* 2020, *10*, 7322. [CrossRef]
- Cheng, Y.; Weng, S.; Yu, L.; Zhu, N.; Yang, M.; Yuan, Y. The Role of Hyperthermia in the Multidisciplinary Treatment of Malignant Tumors. *Integr. Cancer Ther.* 2019, 18, 1534735419876345. [CrossRef]
- Seegenschmiedt, M.H.; Vernon, C.C. A Historical Perspective on Hyperthermia in Oncology. In *Thermoradiotherapy and Ther*mochemotherapy: Biology, Physiology, Physics; Seegenschmiedt, M.H., Fessenden, P., Vernon, C.C., Eds.; Medical Radiology; Springer: Berlin/Heidelberg, Germany, 1995; pp. 3–44, ISBN 978-3-642-57858-8.
- 80. Loeb, L. Über Transplantation von Tumoren. Virchows Arch. Path Anat. 1903, 172, 345–368. [CrossRef]
- 81. Rohdenburg, G.L.; Prime, F. The Effect of Combined Radiation and Heat on Neoplasms. Arch. Surg. 1921, 2, 116–129. [CrossRef]
- 82. Storm, F.K.; Harrison, W.H.; Elliott, R.S.; Morton, D.L. Normal Tissue and Solid Tumor Effects of Hyperthermia in Animal Models and Clinical Trials. *Cancer Res.* 1979, 39, 2245–2251.
- Yang, S.-J.; Huang, C.-H.; Wang, C.-H.; Shieh, M.-J.; Chen, K.-C. The Synergistic Effect of Hyperthermia and Chemotherapy in Magnetite Nanomedicine-Based Lung Cancer Treatment. *Int. J. Nanomed.* 2020, 15, 10331–10347. [CrossRef]

- Kalapurakal, J.A.; Pierce, M.; Chen, A.; Sathiaseelan, V. Efficacy of Irradiation and External Hyperthermia in Locally Advanced, Hormone-Refractory or Radiation Recurrent Prostate Cancer: A Preliminary Report. Int. J. Radiat. Oncol. Biol. Phys. 2003, 57, 654–664. [CrossRef]
- Van Vulpen, M.; De Leeuw, A.A.C.; Raaymakers, B.W.; Van Moorselaar, R.J.A.; Hofman, P.; Lagendijk, J.J.W.; Battermann, J.J. Radiotherapy and Hyperthermia in the Treatment of Patients with Locally Advanced Prostate Cancer: Preliminary Results. *BJU Int.* 2004, 93, 36–41. [CrossRef] [PubMed]
- Anscher, M.S.; Lee, C.; Hurwitz, H.; Tyler, D.; Prosnitz, L.R.; Jowell, P.; Rosner, G.; Samulski, T.; Dewhirst, M.W. A Pilot Study of Preoperative Continuous Infusion 5-Fluorouracil, External Microwave Hyperthermia, and External Beam Radiotherapy for Treatment of Locally Advanced, Unresectable, or Recurrent Rectal Cancer. *Int. J. Radiat. Oncol. Biol. Phys.* 2000, 47, 719–724. [CrossRef]
- Feyerabend, T.; Wiedemann, G.J.; Jäger, B.; Vesely, H.; Mahlmann, B.; Richter, E. Local Hyperthermia, Radiation, and Chemotherapy in Recurrent Breast Cancer Is Feasible and Effective except for Inflammatory Disease. *Int. J. Radiat. Oncol. Biol. Phys.* 2001, 49, 1317–1325. [CrossRef]
- Hildebrandt, B.; Wust, P.; Ahlers, O.; Dieing, A.; Sreenivasa, G.; Kerner, T.; Felix, R.; Riess, H. The Cellular and Molecular Basis of Hyperthermia. *Crit. Rev. Oncol. Hematol.* 2002, 43, 33–56. [CrossRef]
- 89. Wust, P.; Hildebrandt, B.; Sreenivasa, G.; Rau, B.; Gellermann, J.; Riess, H.; Felix, R.; Schlag, P.M. Hyperthermia in Combined Treatment of Cancer. *Lancet Oncol.* 2002, *3*, 487–497. [CrossRef]
- 90. Jha, S.; Sharma, P.K.; Malviya, R. Hyperthermia: Role and Risk Factor for Cancer Treatment. *Achievements Life Sci.* 2016, 10, 161–167. [CrossRef]
- 91. Hegyi, G.; Szigeti, G.P.; Szász, A. Hyperthermia versus Oncothermia: Cellular Effects in Complementary Cancer Therapy. *Evid. Based Complement. Alternat. Med.* **2013**, 2013, 672873. [CrossRef]
- Chang, D.; Lim, M.; Goos, J.A.C.M.; Qiao, R.; Ng, Y.Y.; Mansfeld, F.M.; Jackson, M.; Davis, T.P.; Kavallaris, M. Biologically Targeted Magnetic Hyperthermia: Potential and Limitations. *Front. Pharmacol.* 2018, 9, 831. [CrossRef]
- Carter, T.J.; Agliardi, G.; Lin, F.-Y.; Ellis, M.; Jones, C.; Robson, M.; Richard-Londt, A.; Southern, P.; Lythgoe, M.; Zaw Thin, M.; et al. Potential of Magnetic Hyperthermia to Stimulate Localized Immune Activation. *Small* 2021, 17, e2005241. [CrossRef]
- Liu, J.; Kang, L.; Ratnayake, I.; Ahrenkiel, P.; Smith, S.; Wang, C. Targeting Cancer Cell Adhesion Molecule, CD146, with Low-Dose Gold Nanorods and Mild Hyperthermia Disrupts Actin Cytoskeleton and Cancer Cell Migration. J. Colloid Interface Sci. 2021, 601, 556–569. [CrossRef] [PubMed]
- 95. Maimaitiyiming, Y.; Wang, Q.Q.; Yang, C.; Ogra, Y.; Lou, Y.; Smith, C.A.; Hussain, L.; Shao, Y.M.; Lin, J.; Liu, J.; et al. Hyperthermia Selectively Destabilizes Oncogenic Fusion Proteins. *Blood Cancer Discov.* **2021**, *2*, 388–401. [CrossRef] [PubMed]
- Wu, H.C.; Rérolle, D.; Thé, H. de PML/RARa Destabilization by Hyperthermia: A New Model for Oncogenic Fusion Protein Degradation? *Blood Cancer Discov.* 2021. [CrossRef] [PubMed]
- Kitamura, K.; Saeki, H.; Kawaguchi, H.; Araki, K.; Ohno, S.; Kuwano, H.; Maehara, Y.; Sugimachi, K. Immunohistochemical Status of the P53 Protein and Ki-67 Antigen Using Biopsied Specimens Can Predict a Sensitivity to Neoadjuvant Therapy in Patients with Esophageal Cancer. *Hepatogastroenterology* 2000, 47, 419–423.
- 98. Takahashi, I.; Emi, Y.; Hasuda, S.; Kakeji, Y.; Maehara, Y.; Sugimachi, K. Clinical Application of Hyperthermia Combined with Anticancer Drugs for the Treatment of Solid Tumors. *Surgery* **2002**, *131*, S78–S84. [CrossRef]
- 99. Shchors, K.; Evan, G. Tumor Angiogenesis: Cause or Consequence of Cancer? Cancer Res. 2007, 67, 7059–7061. [CrossRef]
- 100. Adnan, A.; Muñoz, N.M.; Prakash, P.; Habibollahi, P.; Cressman, E.N.K.; Sheth, R.A. Hyperthermia and Tumor Immunity. *Cancers* **2021**, *13*, 2507. [CrossRef]
- 101. Skitzki, J.J.; Repasky, E.A.; Evans, S.S. Hyperthermia as an Immunotherapy Strategy for Cancer. *Curr. Opin. Investig. Drugs* **2009**, *10*, 550–558.
- Mallory, M.; Gogineni, E.; Jones, G.C.; Greer, L.; Simone, C.B. Therapeutic Hyperthermia: The Old, the New, and the Upcoming. *Crit. Rev. Oncol. Hematol.* 2016, 97, 56–64. [CrossRef]
- Martinez-Boubeta, C.; Simeonidis, K.; Oró, J.; Makridis, A.; Serantes, D.; Balcells, L. Finding the Limits of Magnetic Hyperthermia on Core-Shell Nanoparticles Fabricated by Physical Vapor Methods. *Magnetochemistry* 2021, 7, 49. [CrossRef]
- 104. Laurent, S.; Dutz, S.; Häfeli, U.O.; Mahmoudi, M. Magnetic Fluid Hyperthermia: Focus on Superparamagnetic Iron Oxide Nanoparticles. *Adv. Colloid Interface Sci.* 2011, 166, 8–23. [CrossRef]
- 105. Caizer, C. Theoretical Study on Specific Loss Power and Heating Temperature in CoFe<sub>2</sub>O<sub>4</sub> Nanoparticles as Possible Candidate for Alternative Cancer Therapy by Superparamagnetic Hyperthemia. *Appl. Sci.* **2021**, *11*, 5505. [CrossRef]
- Maeda, H. Toward a Full Understanding of the EPR Effect in Primary and Metastatic Tumors as Well as Issues Related to Its Heterogeneity. *Adv. Drug Deliv. Rev.* 2015, *91*, 3–6. [CrossRef] [PubMed]
- Byrne, J.D.; Betancourt, T.; Brannon-Peppas, L. Active Targeting Schemes for Nanoparticle Systems in Cancer Therapeutics. *Adv. Drug Deliv. Rev.* 2008, 60, 1615–1626. [CrossRef] [PubMed]
- 108. Conde-Leboran, I.; Baldomir, D.; Martinez-Boubeta, C.; Chubykalo-Fesenko, O.; del Puerto Morales, M.; Salas, G.; Cabrera, D.; Camarero, J.; Teran, F.J.; Serantes, D. A Single Picture Explains Diversity of Hyperthermia Response of Magnetic Nanoparticles. J. Phys. Chem. C 2015, 119, 15698–15706. [CrossRef]

- 109. Jędrzak, A.; Grześkowiak, B.F.; Golba, K.; Coy, E.; Synoradzki, K.; Jurga, S.; Jesionowski, T.; Mrówczyński, R. Magnetite Nanoparticles and Spheres for Chemo- and Photothermal Therapy of Hepatocellular Carcinoma in Vitro. *IJN* 2020, *15*, 7923–7936. [CrossRef]
- 110. Vilas-Boas, V.; Carvalho, F.; Espiña, B. Magnetic Hyperthermia for Cancer Treatment: Main Parameters Affecting the Outcome of In Vitro and In Vivo Studies. *Molecules* 2020, 25, E2874. [CrossRef]
- 111. Farcas, C.G.; Macasoi, I.; Pinzaru, I.; Chirita, M.; Chirita Mihaila, M.C.; Dehelean, C.; Avram, S.; Loghin, F.; Mocanu, L.; Rotaru, V.; et al. Controlled Synthesis and Characterization of Micrometric Single Crystalline Magnetite With Superparamagnetic Behavior and Cytocompatibility/Cytotoxicity Assessments. *Front. Pharmacol.* 2020, *11*, 410. [CrossRef]
- 112. de Mello, L.B.; Varanda, L.C.; Sigoli, F.A.; Mazali, I.O. Co-Precipitation Synthesis of (Zn-Mn)-Co-Doped Magnetite Nanoparticles and Their Application in Magnetic Hyperthermia. J. Alloys Compounds 2019, 779, 698–705. [CrossRef]
- Zhao, S.; Hao, N.; Zhang, J.X.J.; Hoopes, P.J.; Shubitidze, F.; Chen, Z. Fabrication of Monodisperse Magnetic Nanorods for Improving Hyperthermia Efficacy. J. Nanobiotechnol. 2021, 19, 63. [CrossRef]
- Bao, J.; Guo, S.; Zu, X.; Zhuang, Y.; Fan, D.; Zhang, Y.; Shi, Y.; Ji, Z.; Cheng, J.; Pang, X. Polypyrrole-Coated Magnetite Vortex Nanoring for Hyperthermia-Boosted Photothermal/Magnetothermal Tumor Ablation Under Photoacoustic/Magnetic Resonance Guidance. Front. Bioeng. Biotechnol. 2021, 9. [CrossRef] [PubMed]
- 115. Das, R.; Masa, J.A.; Kalappattil, V.; Nemati, Z.; Rodrigo, I.; Garaio, E.; García, J.Á.; Phan, M.-H.; Srikanth, H. Iron Oxide Nanorings and Nanotubes for Magnetic Hyperthermia: The Problem of Intraparticle Interactions. *Nanomaterials* 2021, 11, 1380. [CrossRef] [PubMed]
- Nguyen, D.T.; Kim, K. Controlled Synthesis of Monodisperse Magnetite Nanoparticles for Hyperthermia-Based Treatments. *Powder Technol.* 2016, 301, 1112–1118. [CrossRef]
- 117. Yang, Y.; Liu, X.; Lv, Y.; Herng, T.S.; Xu, X.; Xia, W.; Zhang, T.; Fang, J.; Xiao, W.; Ding, J. Orientation Mediated Enhancement on Magnetic Hyperthermia of Fe3O4 Nanodisc. *Adv. Functional Mater.* **2015**, *25*, 812–820. [CrossRef]
- 118. Simeonidis, K.; Morales, M.P.; Marciello, M.; Angelakeris, M.; de la Presa, P.; Lazaro-Carrillo, A.; Tabero, A.; Villanueva, A.; Chubykalo-Fesenko, O.; Serantes, D. In-Situ Particles Reorientation during Magnetic Hyperthermia Application: Shape Matters Twice. Sci. Rep. 2016, 6, 38382. [CrossRef]
- 119. Zuo, X.; Ding, H.; Zhang, J.; Fang, T.; Zhang, D. Carbothermal Treated Iron Oxide Nanoparticles with Improving Magnetic Heating Efficiency for Hyperthermia. *Results Phys.* **2022**, *32*, 105095. [CrossRef]
- Perecin, C.J.; Tirich, B.M.; Nagamine, L.C.C.M.; Porto, G.; Rocha, F.V.; Cerize, N.N.P.; Varanda, L.C. Aqueous Synthesis of Magnetite Nanoparticles for Magnetic Hyperthermia: Formation Mechanism Approach, High Water-Dispersity and Stability. *Colloids Surf. A Physicochem. Eng. Aspects* 2021, 627, 127169. [CrossRef]
- 121. Kandasamy, G.; Sudame, A.; Luthra, T.; Saini, K.; Maity, D. Functionalized Hydrophilic Superparamagnetic Iron Oxide Nanoparticles for Magnetic Fluid Hyperthermia Application in Liver Cancer Treatment. ACS Omega 2018, 3, 3991–4005. [CrossRef]
- 122. Kossatz, S.; Ludwig, R.; Dähring, H.; Ettelt, V.; Rimkus, G.; Marciello, M.; Salas, G.; Patel, V.; Teran, F.J.; Hilger, I. High Therapeutic Efficiency of Magnetic Hyperthermia in Xenograft Models Achieved with Moderate Temperature Dosages in the Tumor Area. *Pharm. Res.* **2014**, *31*, 3274–3288. [CrossRef]
- Rajan, A.; Kaczmarek-Szczepańskac, B.; Sahu, N.K. Magneto-Thermal Response of Fe3O4@CTAB Nanoparticles for Cancer Hyperthermia Applications. *Mater. Today Commun.* 2021, 28, 102583. [CrossRef]
- 124. Fatima, H.; Charinpanitkul, T.; Kim, K.-S. Fundamentals to Apply Magnetic Nanoparticles for Hyperthermia Therapy. *Nanomaterials* **2021**, *11*, 1203. [CrossRef] [PubMed]
- 125. Wu, M.; Wang, Q.; Zhang, D.; Liao, N.; Wu, L.; Huang, A.; Liu, X. Magnetite Nanocluster@poly(Dopamine)-PEG@ Indocyanine Green Nanobead with Magnetic Field-Targeting Enhanced MR Imaging and Photothermal Therapy in Vivo. *Colloids Surf B Biointer.* 2016, 141, 467–475. [CrossRef] [PubMed]
- 126. Palzer, J.; Mues, B.; Goerg, R.; Aberle, M.; Rensen, S.S.; Olde Damink, S.W.M.; Vaes, R.D.W.; Cramer, T.; Schmitz-Rode, T.; Neumann, U.P.; et al. Magnetic Fluid Hyperthermia as Treatment Option for Pancreatic Cancer Cells and Pancreatic Cancer Organoids. *Int. J. Nanomed.* 2021, 16, 2965–2981. [CrossRef] [PubMed]
- Singh, D.; McMillan, J.M.; Kabanov, A.V.; Sokolsky-Papkov, M.; Gendelman, H.E. Bench-to-Bedside Translation of Magnetic Nanoparticles. *Nanomedicine* 2014, 9, 501–516. [CrossRef]
- 128. Johannsen, M.; Gneveckow, U.; Eckelt, L.; Feussner, A.; Waldöfner, N.; Scholz, R.; Deger, S.; Wust, P.; Loening, S.A.; Jordan, A. Clinical Hyperthermia of Prostate Cancer Using Magnetic Nanoparticles: Presentation of a New Interstitial Technique. *Int. J. Hyperthermia* 2005, 21, 637–647. [CrossRef]
- Johannsen, M.; Thiesen, B.; Wust, P.; Jordan, A. Magnetic Nanoparticle Hyperthermia for Prostate Cancer. Int. J. Hyperthermia 2010, 26, 790–795. [CrossRef]
- Ito, A.; Kobayashi, T. Intracellular Hyperthermia Using Magnetic Nanoparticles: A Novel Method for Hyperthermia Clinical Applications. *Thermal. Med.* 2008, 24, 113–129. [CrossRef]
- Maier-Hauff, K.; Ulrich, F.; Nestler, D.; Niehoff, H.; Wust, P.; Thiesen, B.; Orawa, H.; Budach, V.; Jordan, A. Efficacy and Safety of Intratumoral Thermotherapy Using Magnetic Iron-Oxide Nanoparticles Combined with External Beam Radiotherapy on Patients with Recurrent Glioblastoma Multiforme. J. Neurooncol. 2011, 103, 317–324. [CrossRef]

- Rubia-Rodríguez, I.; Santana-Otero, A.; Spassov, S.; Tombácz, E.; Johansson, C.; De La Presa, P.; Teran, F.J.; Del Puerto Morales, M.; Veintemillas-Verdaguer, S.; Thanh, N.T.K.; et al. Whither Magnetic Hyperthermia? A Tentative Roadmap. *Materials* 2021, 14, 706. [CrossRef]
- 133. Price, P.M.; Mahmoud, W.E.; Al-Ghamdi, A.A.; Bronstein, L.M. Magnetic Drug Delivery: Where the Field Is Going. *Front. Chem.* **2018**, *6*. [CrossRef]
- 134. Corrie, P.G. Cytotoxic Chemotherapy: Clinical Aspects. Medicine 2008, 36, 24–28. [CrossRef]
- 135. Nurgali, K.; Jagoe, R.T.; Abalo, R. Editorial: Adverse Effects of Cancer Chemotherapy: Anything New to Improve Tolerance and Reduce Sequelae? *Front. Pharmacol.* **2018**, *9*, 245. [CrossRef] [PubMed]
- 136. Zhao, Z.; Huang, D.; Yin, Z.; Chi, X.; Wang, X.; Gao, J. Magnetite Nanoparticles as Smart Carriers to Manipulate the Cytotoxicity of Anticancer Drugs: Magnetic Control and PH-Responsive Release. *J. Mater. Chem.* **2012**, *22*, 15717–15725. [CrossRef]
- 137. Hoshiar, A.K.; Le, T.-A.; Amin, F.U.; Kim, M.O.; Yoon, J. Studies of Aggregated Nanoparticles Steering during Magnetic-Guided Drug Delivery in the Blood Vessels. *J. Magnetism Magnetic Mater.* **2017**, 427, 181–187. [CrossRef]
- Anastasova, E.I.; Prilepskii, A.Y.; Fakhardo, A.F.; Drozdov, A.S.; Vinogradov, V.V. Magnetite Nanocontainers: Toward Injectable Highly Magnetic Materials for Targeted Drug Delivery. ACS Appl. Mater. Interfaces 2018, 10, 30040–30044. [CrossRef] [PubMed]
- Souto, E.B.; Silva, G.F.; Dias-Ferreira, J.; Zielinska, A.; Ventura, F.; Durazzo, A.; Lucarini, M.; Novellino, E.; Santini, A. Nanopharmaceutics: Part I-Clinical Trials Legislation and Good Manufacturing Practices (GMP) of Nanotherapeutics in the EU. *Pharmaceutics* 2020, 12, E146. [CrossRef]
- 140. Ganapathe, L.S.; Mohamed, M.A.; Mohamad Yunus, R.; Berhanuddin, D.D. Magnetite (Fe<sub>3</sub>O<sub>4</sub>) Nanoparticles in Biomedical Application: From Synthesis to Surface Functionalisation. *Magnetochemistry* **2020**, *6*, 68. [CrossRef]
- 141. Fernández, M.; Javaid, F.; Chudasama, V. Advances in Targeting the Folate Receptor in the Treatment/Imaging of Cancers. *Chem. Sci.* **2018**, *9*, 790–810. [CrossRef]
- 142. Song, H.; Quan, F.; Yu, Z.; Zheng, M.; Ma, Y.; Xiao, H.; Ding, F. Carboplatin Prodrug Conjugated Fe3O4 Nanoparticles for Magnetically Targeted Drug Delivery in Ovarian Cancer Cells. J. Mater. Chem. B 2019, 7, 433–442. [CrossRef]
- Ibarra, J.; Encinas, D.; Blanco, M.; Barbosa, S.; Taboada, P.; Juárez, J.; Valdez, M.A. Co-Encapsulation of Magnetic Nanoparticles and Cisplatin within Biocompatible Polymers as Multifunctional Nanoplatforms: Synthesis, Characterization, Andin Vitroassays. *Mater. Res. Express* 2018, 5, 015023. [CrossRef]
- Qiu, H.; Rieger, J.; Gilbert, B.; Jérôme, R.; Jérôme, C. PLA-Coated Gold Nanoparticles for the Labeling of PLA Biocarriers. *Chem. Mater.* 2004, 16, 850–856. [CrossRef]
- 145. Voronin, D.V.; Sindeeva, O.A.; Kurochkin, M.A.; Mayorova, O.; Fedosov, I.V.; Semyachkina-Glushkovskaya, O.; Gorin, D.A.; Tuchin, V.V.; Sukhorukov, G.B. In Vitro and in Vivo Visualization and Trapping of Fluorescent Magnetic Microcapsules in a Bloodstream. ACS Appl. Mater. Interfaces 2017, 9, 6885–6893. [CrossRef] [PubMed]
- 146. Langer, R. Biomaterials in Drug Delivery and Tissue Engineering: One Laboratory's Experience. Acc. Chem. Res. 2000, 33, 94–101. [CrossRef] [PubMed]
- 147. Belete, T.M. The Current Status of Gene Therapy for the Treatment of Cancer. Biologics 2021, 15, 67–77. [CrossRef] [PubMed]
- 148. Roma-Rodrigues, C.; Rivas-García, L.; Baptista, P.V.; Fernandes, A.R. Gene Therapy in Cancer Treatment: Why Go Nano? *Pharmaceutics* **2020**, *12*, 233. [CrossRef]
- Gonçalves, G.A.R.; de Melo Alves Paiva, R. Gene Therapy: Advances, Challenges and Perspectives. *Einstein* 2017, 15, 369–375.
   [CrossRef]
- 150. Lopes, A.; Vandermeulen, G.; Préat, V. Cancer DNA Vaccines: Current Preclinical and Clinical Developments and Future Perspectives. J. Exp. Clin. Cancer Res. 2019, 38, 146. [CrossRef]
- 151. Fus-Kujawa, A.; Prus, P.; Bajdak-Rusinek, K.; Teper, P.; Gawron, K.; Kowalczuk, A.; Sieron, A.L. An Overview of Methods and Tools for Transfection of Eukaryotic Cells in Vitro. *Front. Bioeng. Biotechnol.* **2021**, *9*, 701031. [CrossRef]
- 152. Sun, W.; Shi, Q.; Zhang, H.; Yang, K.; Ke, Y.; Wang, Y.; Qiao, L. Advances in the Techniques and Methodologies of Cancer Gene Therapy. *Discov. Med.* **2019**, *27*, 45–55.
- 153. Muhammad, Q.; Jang, Y.; Kang, S.H.; Moon, J.; Kim, W.J.; Park, H. Modulation of Immune Responses with Nanoparticles and Reduction of Their Immunotoxicity. *Biomater. Sci.* 2020, *8*, 1490–1501. [CrossRef] [PubMed]
- 154. Park, T.G.; Jeong, J.H.; Kim, S.W. Current Status of Polymeric Gene Delivery Systems. *Adv. Drug Deliv. Rev.* **2006**, *58*, 467–486. [CrossRef] [PubMed]
- 155. Zhou, Z.; Liu, X.; Zhu, D.; Wang, Y.; Zhang, Z.; Zhou, X.; Qiu, N.; Chen, X.; Shen, Y. Nonviral Cancer Gene Therapy: Delivery Cascade and Vector Nanoproperty Integration. *Adv. Drug Deliv. Rev.* **2017**, *115*, 115–154. [CrossRef] [PubMed]
- 156. Huang, R.-Y.; Liu, Z.-H.; Weng, W.-H.; Chang, C.-W. Magnetic Nanocomplexes for Gene Delivery Applications. J. Mater. Chem. B 2021, 9, 4267–4286. [CrossRef]
- 157. McBain, S.C.; Yiu, H.H.P.; Dobson, J. Magnetic Nanoparticles for Gene and Drug Delivery. *Int. J. Nanomed.* 2008, *3*, 169–180. [CrossRef] [PubMed]
- 158. Mah, C.; Fraites, T.J.; Zolotukhin, I.; Song, S.; Flotte, T.R.; Dobson, J.; Batich, C.; Byrne, B.J. Improved Method of Recombinant AAV2 Delivery for Systemic Targeted Gene Therapy. *Mol. Ther.* **2002**, *6*, 106–112. [CrossRef]
- 159. Estelrich, J.; Escribano, E.; Queralt, J.; Busquets, M.A. Iron Oxide Nanoparticles for Magnetically-Guided and Magnetically-Responsive Drug Delivery. *Int. J. Mol. Sci.* 2015, *16*, 8070–8101. [CrossRef]

- 160. Goodwin, T.; Huang, L. Chapter One—Nonviral Vectors: We Have Come a Long Way. In *Advances in Genetics*; Huang, L., Liu, D., Wagner, E., Eds.; Nonviral Vectors for Gene Therapy; Academic Press: Cambridge, MA, USA, 2014; Volume 88, pp. 1–12.
- Do Minh, A.; Sharon, D.; Chahal, P.; Kamen, A.A. 1.26—Cell Transfection☆. In *Comprehensive Biotechnology*, 3rd ed.; Moo-Young, M., Ed.; Pergamon: Oxford, UK, 2019; pp. 383–390, ISBN 978-0-444-64047-5.
- 162. Kim, Y.-K.; Zhang, M.; Lu, J.-J.; Xu, F.; Chen, B.-A.; Xing, L.; Jiang, H.-L. PK11195-Chitosan-Graft-Polyethylenimine-Modified SPION as a Mitochondria-Targeting Gene Carrier. *J. Drug Target* 2016, 24, 457–467. [CrossRef]
- Jin, L.; Wang, Q.; Chen, J.; Wang, Z.; Xin, H.; Zhang, D. Efficient Delivery of Therapeutic SiRNA by Fe3O4 Magnetic Nanoparticles into Oral Cancer Cells. *Pharmaceutics* 2019, 11, E615. [CrossRef]
- 164. Kievit, F.M.; Wang, K.; Ozawa, T.; Tarudji, A.W.; Silber, J.R.; Holland, E.C.; Ellenbogen, R.G.; Zhang, M. Nanoparticle-Mediated Knockdown of DNA Repair Sensitizes Cells to Radiotherapy and Extends Survival in a Genetic Mouse Model of Glioblastoma. *Nanomed. Nanotechnol. Biol. Med.* 2017, 13, 2131–2139. [CrossRef]
- Vainauska, D.; Kozireva, S.; Karpovs, A.; Čistjakovs, M.; Bariševs, M. A Novel Approach for Nucleic Acid Delivery into Cancer Cells. *Medicina* 2012, 48, 324–329. [CrossRef]
- 166. Wang, K.; Kievit, F.M.; Sham, J.G.; Jeon, M.; Stephen, Z.R.; Bakthavatsalam, A.; Park, J.O.; Zhang, M. Iron-Oxide-Based Nanovector for Tumor Targeted SiRNA Delivery in an Orthotopic Hepatocellular Carcinoma Xenograft Mouse Model. *Small* 2016, 12, 477–487. [CrossRef]
- 167. Zhao, X.; Meng, Z.; Wang, Y.; Chen, W.; Sun, C.; Cui, B.; Cui, J.; Yu, M.; Zeng, Z.; Guo, S.; et al. Pollen Magnetofection for Genetic Modification with Magnetic Nanoparticles as Gene Carriers. *Nat. Plants* 2017, *3*, 956–964. [CrossRef] [PubMed]
- Zuvin, M.; Kuruoglu, E.; Kaya, V.O.; Unal, O.; Kutlu, O.; Yagci Acar, H.; Gozuacik, D.; Koşar, A. Magnetofection of Green Fluorescent Protein Encoding DNA-Bearing Polyethyleneimine-Coated Superparamagnetic Iron Oxide Nanoparticles to Human Breast Cancer Cells. ACS Omega 2019, 4, 12366–12374. [CrossRef] [PubMed]
- Santra, S.; Kaittanis, C.; Perez, J.M. Cytochrome c Encapsulating Theranostic Nanoparticles: A Novel Bifunctional System for Targeted Delivery of Therapeutic Membrane-Impermeable Proteins to Tumors and Imaging of Cancer Therapy. *Mol. Pharm.* 2010, 7, 1209–1222. [CrossRef] [PubMed]
- 170. Revia, R.A.; Zhang, M. Magnetite Nanoparticles for Cancer Diagnosis, Treatment, and Treatment Monitoring: Recent Advances. *Mater. Today* **2016**, *19*, 157–168. [CrossRef]
- 171. Veiseh, O.; Sun, C.; Fang, C.; Bhattarai, N.; Gunn, J.; Kievit, F.; Du, K.; Pullar, B.; Lee, D.; Ellenbogen, R.G.; et al. Specific Targeting of Brain Tumors with an Optical/Magnetic Resonance Imaging Nanoprobe across the Blood-Brain Barrier. *Cancer Res.* 2009, *69*, 6200–6207. [CrossRef]
- 172. Hadjipanayis, C.G.; Machaidze, R.; Kaluzova, M.; Wang, L.; Schuette, A.J.; Chen, H.; Wu, X.; Mao, H. EGFRvIII Antibody-Conjugated Iron Oxide Nanoparticles for Magnetic Resonance Imaging-Guided Convection-Enhanced Delivery and Targeted Therapy of Glioblastoma. *Cancer Res.* **2010**, *70*, 6303–6312. [CrossRef]
- 173. Crezee, J.; Franken, N.A.P.; Oei, A.L. Hyperthermia-Based Anti-Cancer Treatments. Cancers 2021, 13, 1240. [CrossRef]
- 174. Allan, J.; Belz, S.; Hoeveler, A.; Hugas, M.; Okuda, H.; Patri, A.; Rauscher, H.; Silva, P.; Slikker, W.; Sokull-Kluettgen, B.; et al. Regulatory Landscape of Nanotechnology and Nanoplastics from a Global Perspective. *Regul. Toxicol. Pharmacol.* 2021, 122, 104885. [CrossRef]
- 175. Campbell, A.; Oldham, M.; Becaria, A.; Bondy, S.C.; Meacher, D.; Sioutas, C.; Misra, C.; Mendez, L.B.; Kleinman, M. Particulate Matter in Polluted Air May Increase Biomarkers of Inflammation in Mouse Brain. *Neurotoxicology* **2005**, *26*, 133–140. [CrossRef]
- 176. Costa, C.; Brandão, F.; Bessa, M.J.; Costa, S.; Valdiglesias, V.; Kiliç, G.; Fernández-Bertólez, N.; Quaresma, P.; Pereira, E.; Pásaro, E.; et al. In Vitro Cytotoxicity of Superparamagnetic Iron Oxide Nanoparticles on Neuronal and Glial Cells. Evaluation of Nanoparticle Interference with Viability Tests. J. Appl. Toxicol. 2016, 36, 361–372. [CrossRef] [PubMed]
- 177. Liu, Y.; Li, J.; Xu, K.; Gu, J.; Huang, L.; Zhang, L.; Liu, N.; Kong, J.; Xing, M.; Zhang, L.; et al. Characterization of Superparamagnetic Iron Oxide Nanoparticle-Induced Apoptosis in PC12 Cells and Mouse Hippocampus and Striatum. *Toxicol. Lett.* 2018, 292, 151–161. [CrossRef] [PubMed]
- 178. Murros, K.; Wasiljeff, J.; Macías-Sánchez, E.; Faivre, D.; Soinne, L.; Valtonen, J.; Pohja, M.; Saari, P.; Pesonen, L.J.; Salminen, J.M. Magnetic Nanoparticles in Human Cervical Skin. *Front. Med.* **2019**, *6*, 123. [CrossRef] [PubMed]
- Zhu, M.-T.; Wang, Y.; Feng, W.-Y.; Wang, B.; Wang, M.; Ouyang, H.; Chai, Z.-F. Oxidative Stress and Apoptosis Induced by Iron Oxide Nanoparticles in Cultured Human Umbilical Endothelial Cells. *J. Nanosci. Nanotechnol.* 2010, 10, 8584–8590. [CrossRef] [PubMed]
- Donaldson, K.; Stone, V.; Clouter, A.; Renwick, L.; MacNee, W. Ultrafine Particles. Occup. Environ. Med. 2001, 58, 211–216, 199.
   [CrossRef]
- Oberdörster, G.; Oberdörster, E.; Oberdörster, J. Nanotoxicology: An Emerging Discipline Evolving from Studies of Ultrafine Particles. *Environ. Health Perspect.* 2005, 113, 823–839. [CrossRef]
- Schins, R.P.F.; Lightbody, J.H.; Borm, P.J.A.; Shi, T.; Donaldson, K.; Stone, V. Inflammatory Effects of Coarse and Fine Particulate Matter in Relation to Chemical and Biological Constituents. *Toxicol. Appl. Pharmacol.* 2004, 195, 1–11. [CrossRef]
- Rizwan, M.; Ali, S.; Ali, B.; Adrees, M.; Arshad, M.; Hussain, A.; Zia ur Rehman, M.; Waris, A.A. Zinc and Iron Oxide Nanoparticles Improved the Plant Growth and Reduced the Oxidative Stress and Cadmium Concentration in Wheat. *Chemosphere* 2019, 214, 269–277. [CrossRef]

- 184. Shahbazi, M.-A.; Hamidi, M.; Mäkilä, E.M.; Zhang, H.; Almeida, P.V.; Kaasalainen, M.; Salonen, J.J.; Hirvonen, J.T.; Santos, H.A. The Mechanisms of Surface Chemistry Effects of Mesoporous Silicon Nanoparticles on Immunotoxicity and Biocompatibility. *Biomaterials* 2013, 34, 7776–7789. [CrossRef]
- De Simone, U.; Spinillo, A.; Caloni, F.; Avanzini, M.A.; Coccini, T. In Vitro Evaluation of Magnetite Nanoparticles in Human Mesenchymal Stem Cells: Comparison of Different Cytotoxicity Assays. *Toxicol. Mechanisms Methods* 2020, 30, 48–59. [CrossRef]
- Like, C.; Changsheng, G.; Miao, L.; Zhuan, L.; Jian, X. The Cytotoxic Effects of Magnetite Nanoparticle-Pb<sup>2+</sup> Complex on Rat Kidney Cells. *Asian J. Ecotoxicol.* 2020, 87–95. [CrossRef]
- 187. Minaeva, O.V.; Brodovskaya, E.P.; Pyataev, M.A.; Gerasimov, M.V.; Zharkov, M.N.; Yurlov, I.A.; Kulikov, O.A.; Kotlyarov, A.A.; Balykova, L.A.; Kokorev, A.V.; et al. Comparative Study of Cytotoxicity of Ferromagnetic Nanoparticles and Magnetitecontaining Polyelectrolyte Microcapsules. J. Phys. Conf. Ser. 2017, 784, 012038. [CrossRef]
- Zhang, S.; Wu, S.; Shen, Y.; Xiao, Y.; Gao, L.; Shi, S. Cytotoxicity Studies of Fe3O4 Nanoparticles in Chicken Macrophage Cells. *R. Soc. Open Sci.* 2020, 7, 191561. [CrossRef] [PubMed]
- Mohammadi, H.; Nekobahr, E.; Akhtari, J.; Saeedi, M.; Akbari, J.; Fathi, F. Synthesis and Characterization of Magnetite Nanoparticles by Co-Precipitation Method Coated with Biocompatible Compounds and Evaluation of in-Vitro Cytotoxicity. *Toxicol. Rep.* 2021, *8*, 331–336. [CrossRef] [PubMed]
- Yusefi, M.; Shameli, K.; Yee, O.S.; Teow, S.-Y.; Hedayatnasab, Z.; Jahangirian, H.; Webster, T.J.; Kuča, K. Green Synthesis of Fe<sub>3</sub>O<sub>4</sub> Nanoparticles Stabilized by a Garcinia Mangostana Fruit Peel Extract for Hyperthermia and Anticancer Activities. *IJN* 2021, 16, 2515–2532. [CrossRef]
- 191. Majetich, S.A.; Wen, T.; Mefford, O.T. Magnetic Nanoparticles. MRS Bull. 2013, 38, 899–903. [CrossRef]
- 192. Lassenberger, A.; Grünewald, T.A.; van Oostrum, P.D.J.; Rennhofer, H.; Amenitsch, H.; Zirbs, R.; Lichtenegger, H.C.; Reimhult, E. Monodisperse Iron Oxide Nanoparticles by Thermal Decomposition: Elucidating Particle Formation by Second-Resolved in Situ Small-Angle X-Ray Scattering. *Chem. Mater.* 2017, 29, 4511–4522. [CrossRef]
- Wan, J.; Cai, W.; Meng, X.; Liu, E. Monodisperse Water-Soluble Magnetite Nanoparticles Prepared by Polyol Process for High-Performance Magnetic Resonance Imaging. *Chem. Commun.* 2007, 5004–5006. [CrossRef]
- 194. Wang, H.; Shrestha, T.B.; Basel, M.T.; Pyle, M.; Toledo, Y.; Konecny, A.; Thapa, P.; Ikenberry, M.; Hohn, K.L.; Chikan, V.; et al. Hexagonal Magnetite Nanoprisms: Preparation, Characterization and Cellular Uptake. J. Mater. Chem. B 2015, 3, 4647–4653. [CrossRef]
- 195. Wetterskog, E.; Agthe, M.; Mayence, A.; Grins, J.; Wang, D.; Rana, S.; Ahniyaz, A.; Salazar-Alvarez, G.; Bergström, L. Precise Control over Shape and Size of Iron Oxide Nanocrystals Suitable for Assembly into Ordered Particle Arrays. *Sci. Technol. Adv. Mater.* 2014, 15, 055010. [CrossRef]
- Merino, S.; Martín, C.; Kostarelos, K.; Prato, M.; Vázquez, E. Nanocomposite Hydrogels: 3D Polymer-Nanoparticle Synergies for On-Demand Drug Delivery. ACS Nano 2015, 9, 4686–4697. [CrossRef] [PubMed]
- 197. Sensenig, R.; Sapir, Y.; MacDonald, C.; Cohen, S.; Polyak, B. Magnetic Nanoparticle-Based Approaches to Locally Target Therapy and Enhance Tissue Regeneration in Vivo. *Nanomedicine* 2012, 7, 1425–1442. [CrossRef] [PubMed]
- Thiesen, B.; Jordan, A. Clinical Applications of Magnetic Nanoparticles for Hyperthermia. *Int. J. Hyperthermia* 2008, 24, 467–474. [CrossRef] [PubMed]