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# Marine invasion genetics: from spatio-temporal patterns to evolutionary outcomes

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## Abstract

1  
2 Over the last 15 years studies on invasion genetics have provided important insights to  
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4 unravel cryptic diversity, track the origin of colonizers and reveal pathways of introductions.  
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6 Despite all these advances, to date little is known about how evolutionary processes influence  
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8 the observed genetic patterns in marine biological invasions. Here, we firstly review the  
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10 literature on invasion genetics that include samples from European seas. These seas constitute  
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12 a wide array of unique water masses with diverse degrees of connectivity, and have a long  
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14 history of species introductions. We found that only a small fraction of the recorded  
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16 introduced species has been genetically analysed. Furthermore, most studies restrict their  
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18 approach to describe patterns of cryptic diversity and genetic structure, with the underlying  
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20 mechanisms involved in the invasion process being largely understudied. Secondly, we  
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22 analyse how genetic, reproductive and anthropogenic traits shape genetic patterns of marine  
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24 introduced species. We found that most studies reveal similar genetic diversity values in both  
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26 native and introduced ranges, report evidence of multiple introductions, and show that genetic  
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28 patterns in the introduced range are not explained by taxonomic group or reproductive  
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30 strategy. Finally, we discuss the evolutionary implications derived from genetic patterns  
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32 observed in non-indigenous species. We identify different scenarios that are determined by  
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34 propagule pressure, phenotypic plasticity and pre-adaptation, and the effects of selection and  
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36 genetic admixture. We conclude that there is a need for further investigations of evolutionary  
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38 mechanisms that affect individual fitness and adaptation to rapid environmental change.  
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51 **Keywords:** Europe, hybridisation, introduced species, invasion routes, cryptic  
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## Introduction

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2 Evolutionary genetics is a well-established and fertile field of research that has been  
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4 central for understanding biological invasions (Lee 2002). It has provided the opportunity to  
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6 unravel concealed patterns and processes involved with species introductions (e.g. Balanyà et  
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8 al. 1994; Taylor and Keller 2007; Cadotte et al. 2009; Lawson Handley et al. 2011) and has  
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10 boosted contentious scientific debates (e.g. genetic paradox in invasion biology, Tsutsui et al.  
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12 2000; Frankham 2005; Roman and Darling 2007). Population genetics, molecular barcoding  
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14 and phylogenetics are increasingly being used to understand aspects of biological invasions  
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16 such as cryptic diversity and cryptic introductions, tracking the origin of introductions or  
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18 characterising colonisation pathways (Estoup and Guillemaud 2010; Geller et al. 2010). The  
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20 term ‘invasion genetics’ was coined in the late 1990s (Villablanca et al. 1998; Davies et al.  
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22 1999) and since then invasion genetics studies have proliferated, fuelled by both  
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24 methodological and analytical advances in genetic techniques (Cock et al. 2010; Reitzel et al.  
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26 2013).

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29 As is the case in the field of ecology (Menge et al. 2009), terrestrial and aquatic  
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31 invasion biology studies have historically progressed in relative isolation (Ruiz et al. 2000;  
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33 Kolar and Lodge 2001; Grosholz 2002; Callaway and Maron 2006). This may be due to the  
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35 traditional design of academic pathways or the idiosyncratic and/or antagonistic  
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37 characteristics of these ecosystems. However, there is no reason to maintain this segregation,  
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39 as most concepts in invasion biology are applicable across ecosystems (e.g. Roman and  
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41 Darling 2007; Wilson et al. 2009; Blackburn et al. 2011). Marine ecosystems have a broad  
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43 range of characteristics that make them ideal model systems for invasion genetics studies.  
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45 This includes the open nature of the seascape, the diversity of life-histories and dispersal  
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47 capabilities of marine biota, and the type and frequency of introduction vectors (see details  
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49 below). Holland (2000) produced the first study that specifically reviewed knowledge on  
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1 marine invasion genetics and detailed the usefulness of genetic techniques for understanding  
2 ecological processes involved in biological invasions. More recently, a study reviewing the  
3 latest findings in marine invasion genetics emphasized the practicality of genetic tools for the  
4 identification of cryptic invasions and sources of introduced populations (Geller et al. 2010).  
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9 However, to date no review has addressed the link between observed genetic patterns and  
10 evolutionary processes in marine biological invasions.  
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14 In this review, we begin by examining all available information from invasion genetic  
15 studies conducted using samples collected from European seas to assess the state of  
16 knowledge and the importance of a number of genetic and life-history traits relevant for  
17 colonisation and spread. We chose the marine waters surrounding Europe (NE Atlantic Ocean  
18 and the Baltic, Mediterranean and Black seas) as this region provides an excellent setting to  
19 investigate genetic patterns related to marine biological invasions. European shores comprise  
20 a wide array of unique water masses with diverse degrees of connectivity. Also, they have a  
21 long history of species introductions - both as a donor and as a receiver (e.g. Patti and Gambi  
22 2001; Roman and Palumbi 2004; Rius et al. 2014). We then analyse how genetic diversity,  
23 reproductive strategies and anthropogenic vectors shape invasion genetic patterns. From there  
24 we obtain and discuss insights into the evolutionary processes that may underlie biological  
25 invasions and how the latest developments in genetic techniques can help to unravel these.  
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#### 44 **History, regional specificities and genetic patterns of European seas**

45 Inhabitants of the European region were amongst the first sailors to establish  
46 transoceanic routes for commercial and colonisation purposes (Scammell 1981). This  
47 represented the dawn of long-distance artificial translocation of marine species. European  
48 shipping traffic to African, American and Asian shores commenced in the 15<sup>th</sup> century  
49 (Scammell 1991), which established global commercial routes. Europe has been the cradle of  
50 taxonomy, and as a result descriptions of marine species from both Europe and abroad  
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1 emerged in the eighteenth / nineteenth centuries, when transoceanic shipping had already been  
2 in place for centuries. Many species that arrived in Europe in historical times were  
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4 mislabelled as native simply because they were already there when taxonomists came on to  
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6 the stage (Carlton 2000; Carlton 2009). Many purportedly ‘European’ species are simply early  
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8 introductions, or should be labelled at best as cryptogenic species (Carlton 1996; Haydar  
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10 2012). Consequently, Europe is plagued with historical taxonomic problems and instances of  
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12 ‘pseudoinigenous’ species (sensu Carlton 2009). Genetic tools have greatly helped to resolve  
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14 this issue (Geller et al. 2010), although in some cases the genetic signal has been blurred by  
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16 centuries of global transportation. Thus, it is nowadays extremely challenging to track the  
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18 origins and introduction pathways of cosmopolitan species or species complexes (e.g. Zhan et  
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20 al. 2010; Pineda et al. 2011; Pérez-Portela et al. 2013).

26 Concerning the geographic particularities of the European region, the NE Atlantic  
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28 Ocean shores include some of the most frequented shipping harbours worldwide (Kaluza et al.  
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30 2010; Keller et al. 2011). These harbours have provided an extensive and long-term man-  
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32 made interchange of marine biota among temperate regions, and such exposure to  
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34 introductions has led to high propagule pressure within the N Atlantic region (Briski et al.  
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36 2012). Such large contingents of introduced individuals have resulted in a long list of reported  
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38 non-indigenous species, often complemented with accurate species range descriptions, but  
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40 rarely studied in detail retrospectively (how did the introduction and expansion happen?) or  
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42 prospectively (what are the chances for future introductions?) (but see for example Bolte et al.  
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44 2013). Another peculiarity of the European seas is the presence of important semi-enclosed  
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46 seas, namely the Baltic, Mediterranean and Black seas. These basins are connected to ‘outside  
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48 basins’ by extremely narrow links (Kattegat / Skagerrak, Gibraltar, and Sea of Marmara). This  
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50 particular geographic setting has led some authors to consider these seas as giant mesocosms,  
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whereby the outcomes of ‘natural experiments’ occurring there have the potential to enhance our understanding of major processes affecting the world’s oceans (Lejeusne et al. 2010).

The Mediterranean Sea is the most altered and invaded of all European seas, with recent estimates of the number of introduced species ranging between 500 to 1000 species (Streftaris et al. 2005; Streftaris and Zenetos 2006; Galil 2009; Coll et al. 2010; Zenetos et al. 2012). The Gibraltar strait represents a strong biogeographic break (Patarnello et al. 2007) and is a hub of human activity, making it difficult to disentangle natural range expansions from human-related species introductions (Rius et al. 2012; Zenetos et al. 2012). The opening of the Suez Canal in 1869 initiated a species migration from the Red Sea to the Mediterranean Sea (the so-called Lessepsian migration) (Streftaris et al. 2005; Galil 2009; Coll et al. 2010). To date more than 330 alien species are considered to have been introduced through the Suez Canal, resulting in a drastic biodiversity change in the Mediterranean Sea (Galil 2007; Galil 2012; Belmaker et al. 2013). However, the perception of the role of the Suez Canal and the Gibraltar Strait as entrance gates may be biased by the failure to recognize early introductions as such (as outlined above), while modern introductions are more easily identifiable. The Lessepsian invasions represent a unique scientific model, as the date of the opening of the Canal is known and the route of invasion is generally unidirectional. Thus, the Lessepsian system allows, within a relatively simple setting, for the testing of specific hypotheses related to the genetics of biological invasions. For example, Lessepsian introductions support the idea that reduction in genetic diversity is not inherent to the invasion process, because since the earliest genetic studies of Lessepsian species, little evidence for genetic bottlenecks has been found (Golani and Ritte 1999; Hassan et al. 2003; Hassan and Bonhomme 2005; Azzurro et al. 2006; Bernardi et al. 2010). In only one instance, namely that of the highly successful introduced bluespotted cornetfish *Fistularia commersonii*, is there evidence of lowered genetic diversity in the Mediterranean Sea (Golani et al. 2007; Sanna et al. 2011).

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Contrary to what happens with the Lessepsian connection, the Black Sea and adjacent Azov Sea are naturally connected to the Mediterranean Sea, albeit with extreme restrictions at the Bosphorus and Dardanelles, which are very shallow straits (approximately 50 m in depth) (Yaltırak et al. 2000). Additionally, the influx of freshwater into the Black Sea results in lower salinity compared to the Mediterranean Sea (Sorokin 2002). This translates into the Black Sea waters naturally flowing out towards the Mediterranean, through an upper layer of lower salinity water, while higher salinity Mediterranean water tends to flow in towards the Black Sea as a bottom layer, (similarly to the situation in the Gibraltar strait, where the heavier Mediterranean water flows out towards the Atlantic Ocean) (Murray et al. 1991; Sorokin 2002). Due to these particularities, the Mediterranean and Black seas' biota have been isolated with strong limitations to gene flow in place. However, in recent years, changes in the hydrology and abiotic characteristics of the Black sea, together with an increase in ship traffic, have made this natural boundary more permeable (Gregg and Özsoy 2002). Contrary to the Lessepsian situation, the source and route of biological introductions to and from the Black Sea need to be studied carefully, as they may follow different modes. For example, the ctenophore *Mnemiopsis leidyi*, which displays a native range along the eastern seaboard of the United States of America (Reusch et al. 2010), was first recorded in the Black Sea and only later recorded in the Mediterranean Sea (Ghabooli et al. 2011). Genetic studies, based on mitochondrial and nuclear markers confirmed that this ctenophore species was first introduced in the Black Sea (supposedly via ballast water from the Gulf of Mexico Reusch et al. 2010), and from there invaded the Mediterranean Sea (Ghabooli et al. 2013). These results are consistent with a similar study based on microsatellite markers (Bolte et al. 2013) and, importantly, with the overall hydrography of the region, where surface water (where dispersive propagules are more concentrated) tends to flow out of the Black Sea into the Mediterranean Sea.

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Finally, the Baltic Sea includes coastal ecosystems with brackish waters that are considered important centres of xenodiversity (Leppäkoski et al. 2002). The oceanographic similarity between the Black and Baltic seas has led to large scale colonisation pressure of Ponto-Caspian taxa of which representatives of crustaceans (e.g. Cristescu et al. 2004; Cristescu and Hebert 2005; Audzijonyte et al. 2007; Audzijonyte et al. 2013) and gobiids (e.g. Brown and Stepien 2008; Feldheim et al. 2009; Neilson and Stepien 2011) have been traced genetically. In some cases, such as with the introduction of the polychaete species *Marenzelleria neglecta*, *M. viridis* and *M. arctia* (Blank and Bastrop 2009), the introduction of non-indigenous species has resulted in significant biogeochemical impacts on coastal ecosystems of the Baltic Sea (Norkko et al. 2012).

### **Literature review**

Table S1 lists 87 studies on invasion genetics (as of 2013, the last full year available) that include samples from European seas, including information on taxonomic affiliations, natural dispersal capabilities, genetic markers used and some major genetic traits. Genetic studies of alien species in European seas started in the 1990s with allozyme analyses, and were subsequently followed by studies using DNA sequence and microsatellite data. Mitochondrial DNA, particularly the cytochrome c oxidase subunit I gene, remains the most widely used genetic marker (44% of the studies). One surprising result from this review is that after an initial period (1996-2003) of low publication rate (average of 2.3 works per year), the rate increased in 2004 but has since remained fairly constant (average of 7.1 works per year during the period 2004-2013) (Fig. 1A). This is despite a boom in the use of genetic tools in marine research in recent years (e.g. Uriz and Turon 2012).

The geographic areas covered by these studies are depicted in Fig. 1B, showing a predominance of studies covering the NE Atlantic Ocean and the Mediterranean Sea, while smaller areas such as the Baltic and Black seas were the subjects of fewer studies. When we



1 analysed the taxonomic affiliation of studied taxa (Fig. 1C), we found a clear dominance of  
2 chordates. Taking as a reference point the taxonomic distribution of alien species reported by  
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4 Zenetos et al. (2012) in the Mediterranean Sea, the major introduced groups are Mollusca,  
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6 Crustacea, Polychaeta, Macrophyta, and Fish. Of these, only Macrophyta and Fish have  
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8 received attention to match their importance in terms of genetic studies (Fig. 1C). In turn,  
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10 ctenophores, and particularly ascidians, are overrepresented in genetic studies.  
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14 The results strongly favour the notion that genetic diversity is not generally impaired  
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16 when species are introduced. Of the ca. 54% of studies that did compare genetic diversity in  
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18 both native and introduced populations, the majority of them (ca. 74%) found similar diversity  
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20 in some or all introduced populations. Only ca. 23% of the studies reported lower genetic  
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22 diversity in introduced populations, with one study (ca. 2%) reporting higher diversity in the  
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24 introduced range (Fig. 1D). Interestingly, when we categorized the dispersal capabilities of  
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26 the species from studies that compare genetic diversity between native and introduced  
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28 populations into short dispersal (pelagic lifetime less than one week, usually corresponding to  
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30 lecithotrophic larvae or direct developers) and long dispersal (pelagic lifetime more than one  
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32 week, corresponding to long-lived propagules or planktotrophic larvae), we found that both  
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34 strategies are represented (Fig. 1D). However, we found a slightly higher frequency of forms  
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36 with long dispersal abilities in species with similar genetic diversity in native and introduced  
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38 populations (ca. 63% vs ca. 55% of species featuring higher diversity in the native  
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40 populations). Almost half of the studies in Table S1 (ca. 47%) reported evidence of multiple  
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42 introductions. This percentage reaches ca. 57% in studies reporting similar diversity in native  
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44 and introduced populations, but drops to ca. 45% in studies that found higher diversity in  
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46 native populations. Overall, the evidence from published studies points towards a low  
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48 frequency of founder effects (i.e. severe genetic drift responsible for a loss of genetic  
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50 variation after the colonisation of a new site by a limited number of individuals) in marine  
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1 introductions to European waters, with multiple introductions playing an important role in  
2 generating genetic patterns.  
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4 As a whole, we found a dearth of genetic studies of European introduced species. Out  
5 of an estimated 500 - 1000 introduced species (Zenetos et al. 2012), the reviewed genetic  
6 studies focussed on only 59 species (or species-complexes). In addition, most studies  
7 reviewed here have focussed on resolving cryptic diversity and describing patterns of genetic  
8 structure across species ranges, with little work done on adaptive or evolutionary processes.  
9

### 10 **Genetic diversity, reproductive strategies and anthropogenic vectors**

11 Marine genetic studies routinely encounter a realm full of cryptic diversity (Appeltans  
12 et al. 2012). This requires uncovering and understanding a wealth of ecological and  
13 evolutionary mechanisms ruling speciation. Studies on marine biological invasions face an  
14 additional challenge as a result of human transport, which can determine the distribution of  
15 genotypes of species or species complexes (e.g. Daguin and Borsa 2000; Pérez-Portela et al.  
16 2013; Rius and Teske 2013). Even within well-defined genetic lineages, the relationship  
17 between genetic diversity and colonisation success has generated an intense debate in the  
18 invasion biology literature. Genetic studies consistently show that marine biological invasions  
19 are often characterized by the presence of high propagule pressure from multiple and highly  
20 divergent sources from the native range (e.g. Simon-Bouhet et al. 2006; Rius et al. 2012).  
21 Another possibility is that the high genetic diversity of localized introduced populations is  
22 explained by introductions from a few but genetically highly polymorphic sources (Riquet et  
23 al. 2013), a feature that has also been documented in terrestrial populations (Meimberg et al.  
24 2010). Based on these two processes (i.e. introduction from genetically disparate sources or  
25 genetically diversified sources), the introduced range generally has similar or higher mean  
26 values of genetic diversity than the native range (e.g. *Crepidula fornicata*, Fig. 2). However,  
27 there are exceptions to this (Koskinen et al. 2002; Gaither et al. 2012) and important  
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1 population processes such as genetic bottleneck, genetic drift, lag period and / or Allee effects  
2 (Tsutsui et al. 2000; Leung et al. 2004; Crooks 2005; Dawson & Hamner 2005; Taylor and  
3 Hastings 2005) can be responsible for reducing levels of genetic diversity. Some studies on  
4 marine introduced species have indeed reported low levels of genetic diversity (e.g. Provan et  
5 al. 2005; Dupont et al. 2007b; Golani et al. 2007; Tepolt et al. 2009; Pérez-Portela et al. 2012;  
6 Rius and Shenkar 2012). In some species, strong founder effects during expansion may result  
7 in a decrease in genetic diversity over time (e.g. *Perophora japonica*, Pérez-Portela et al.  
8 2012) or create an expansion wave (see Ciosi et al. 2011 for a terrestrial example). It has been  
9 shown that such low levels of genetic diversity do not generally compromise population  
10 establishment and the spread of non-indigenous species (Roman and Darling 2007). Some  
11 studies report a complex interplay of primary and secondary introductions (also called pre-  
12 border and post-border stages, see Forrest et al. 2009), with the former transfer happening  
13 unidirectionally (from the native to the introduced range), while the latter occurs within the  
14 introduced range. In such cases, the native range is normally characterized by a well-defined  
15 genetic structure while the introduced range is diverse but highly homogenous (Voisin et al.  
16 2005; Simon-Bouhet et al. 2006; Rius et al. 2012). When human-mediated transport results in  
17 bi-directional relocation of propagules between the native and introduced ranges, a random  
18 reshuffling of genotypes occurs, which may lead to global genetic homogenisation (Zhan et  
19 al. 2010; Pineda et al. 2011).

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46 When genetic attributes are compared among species with different reproductive  
47 strategies, common outcomes can be found. For instance, both poor dispersers, such as  
48 direct-developers (e.g. the gastropod *Cylope neritea*, Simon-Bouhet et al. 2006) or species  
49 with lecitotrophic larvae (e.g. the ascidian *Microcosmus squamiger*, Ordóñez et al. 2013), and  
50 highly dispersive species, like benthic-pelagic species with long-lived larvae (e.g. the  
51 gastropod *Crepidula fornicata*, Riquet et al. 2013 and references therein), are all successful  
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1 invaders that show no reduction in genetic diversity compared to the native range. This is  
2 counterintuitive as in the absence of human-mediated transport of species, short and long  
3 planktonic larval duration display different dispersal kernel distributions and thus dissimilar  
4 genetic dispersal estimates (Siegel et al. 2003). Differences may be expected when  
5 considering social organisation or gregarious behaviour. For instance, colonial vs. solitary  
6 ascidians were shown to display contrasted genetic patterns in their native range (Ayre et al.  
7 2009). And yet, both colonial and solitary non-indigenous species show similar genetic  
8 patterns (Bock et al. 2011; Goldstien et al. 2011; Bock et al. 2012; Rius et al. 2014). Taken  
9 together, genetic studies analysing the role of reproductive strategies of introduced species  
10 suggest that these different strategies have little effect on the composition and distribution of  
11 genetic signatures.

12 The open nature of most seascapes prevents the efficient containment of introduced  
13 marine species and facilitates the spread of initial and successive waves of founders in the  
14 wild. Such spread has been demonstrated using bio-physical modelling of the larval dispersal  
15 of the *C. fornicata* in its European introduced range (Viard et al. 2006; Dupont et al. 2007a).  
16 Furthermore, the long-lived larval stages of many marine organisms allow them to typically  
17 remain in the water column for a few weeks (Shanks 2009), which might be an advantageous  
18 trait in artificial transportation (e.g. ballast waters). In addition, organisms can survive long  
19 periods of transport time (e.g. several months for the benthic amphipod, *Corophium*  
20 *acherusicum*, in ballast water tanks, Gollasch 1996). In yet another instance, Gollasch et al.  
21 (2000) showed that the abundance of the copepod *Tisbe graciloides* grows exponentially in  
22 ballast waters irrespective of transport time and species diversity. Anthropogenic vectors (e.g.  
23 ballast waters, aquaculture, and fouling of ship hulls and sea-chests) represent highly efficient  
24 ways to transport large numbers of individuals per species and an array of species (Carlton  
25 and Geller 1993; Cohen and Carlton 1998; Carlton 2001). Similarly, marine infrastructures

1 (i.e. man-made mobile or immobile structures) have been identified as major vectors for both  
2 primary introductions and subsequent expansions (i.e. stepping stones, corridors or  
3 bridgehead effects; Bulleri and Airoidi 2005; Dafforn et al. 2009; Mineur et al. 2012).  
4 However, studies comparing different vectors are needed. For example, ballast water is  
5 known to be very effective at transporting invertebrate larvae but most likely plays a minor  
6 role in the transport of fish species (with the notable exception of the round goby, *Neogobius*  
7 *melanostomus*, introduced from the Black Sea to the Great Lakes, Fuller et al. 2014). Ship  
8 fouling is a non-selective vector for non-indigenous benthic species, particularly those that  
9 take refuge in sea chests and other structures less exposed to drag during navigation (Coutts  
10 and Dodgshun 2007). In another notable instance, recreational boating is an important vector  
11 for the spread of non-indigenous species (Acosta and Forrest 2009), specially during the  
12 secondary stage of introduction (Mineur et al. 2008). However, this vector remains the largest  
13 unregulated vector for the introduction and spread of marine non-indigenous species (Clarke  
14 Murray et al. 2011).

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34 Population genetic studies have revealed interesting patterns of introduction and  
35 spread via aquaculture. This vector is believed to account for as many introductions as  
36 shipping (Savini et al. 2000; Wolff and Reise 2002; Streftaris et al. 2005; Mineur et al. 2007).  
37 Contrary to the somewhat haphazard role of shipping, aquaculture is particularly prone to  
38 repeated introductions due to the continued interchange of species among facilities. For  
39 instance, the rapid spread of the gastropod *Cyclope neritea* along the French Atlantic coasts  
40 was shown to be associated with an admixture event between divergent genetic lineages,  
41 which increased genetic diversity in the newly recorded French populations (Fig. 3). Such a  
42 pattern is best explained by recurrent introductions from several areas in the Mediterranean  
43 Sea, including important shellfish culture sites in Spain, France and Italy (Simon-Bouhet et al.  
44 2006). Another instance is the Japanese kelp *Undaria pinnatifida*, which is native to Asia and

1 was first accidentally introduced in the Thau lagoon (French Mediterranean Sea) in the 1970s  
2 and subsequently intentionally transported to NE Atlantic shores to be cultivated. Genetic  
3 studies have shown that sustainable populations can be established after their escape from  
4 aquaculture facilities (e.g. in the English Channel, Grulois et al. 2011). The cultivated origin  
5 of wild populations of *U. pinnatifida* is supported by evidence that cultivated and wild  
6 populations belong to the same genetic lineage (Voisin et al. 2005). The escape and  
7 sustainable establishment of cultivated stocks has also been clearly demonstrated for the  
8 Pacific oyster *Crassostrea gigas* in Ireland (Kochmann et al. 2012). For both the Japanese  
9 kelp and the Pacific oyster, genetic data strongly suggest that wild populations have become  
10 self-sustainable. Another type of introduction derived from aquaculture activities concerns  
11 escapes of domesticated fish and their associated fauna, which in some cases has fully  
12 replaced native populations or have become introgressed with native populations (e.g. Glover  
13 et al. 2013). Such processes may, in addition, facilitate the translocation of non-indigenous  
14 pathogens to native species. This is exemplified by the large-scale infestation of the European  
15 eel *Anguilla anguilla* by the Asian parasite *Anguillicola crassus*. This is despite the fact that  
16 its original host (*Anguilla japonica*) has never successfully established in European waters  
17 (Wielgoss et al. 2008).

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New regulations and international conventions aimed at controlling the transport of marine species (e.g. European Community No 708/2007 of 11<sup>th</sup> June 2007 concerning the use of alien and locally absent species in aquaculture, or the new European Union 2020 Biodiversity Strategy that proposes a firm commitment to protect biodiversity and mitigate introduced species that threaten important economic activities) will contribute to the constraint of future introductions. However, it is uncertain how these measures will affect genetic patterns of non-indigenous species.

### **Evolutionary outcomes**

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As seen above, studies of successful marine non-indigenous species routinely report the presence of populations with high genetic diversity in the introduced range. This situation may be linked to evolutionary neutral or selective processes. In the simplest case, all introduced genotypes establish and spread in the host environment, which can be similar to the native range. If the environment in the native and introduced range differs, phenotypic plasticity may be responsible for counteracting any effect of environmental filtering (Emerson and Gillespie 2008; Davidson et al. 2011). Phenotypic plasticity is a heritable trait that can accelerate adaptive processes (e.g. genetic assimilation Lande 2009; Fierst 2011). As introduced populations face important selective constraints and pressures over contemporary time scales (Huey et al. 2005), selection acting on the genetic background of introduced individuals is likely to be important (Novak 2007). In such cases, a central question arises: what are the evolutionary mechanisms based on selective processes that promote the sustainable and successful establishment of populations of non-indigenous species? Three possible explanations are proposed for selective processes occurring during the introduction of multiple divergent genotypes into a new range (Fig. 4).

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The first scenario considers a situation whereby the introduced range receives introductions from divergent genetic backgrounds but only one of them establishes (p.eg. exclusion as a result of environmental filtering). The successful genotype may be better adapted to the new environment (often called ‘pre-adapted’ genotype, see Dlugosch and Parker 2007). This scenario requires some sort of match between conditions in the host environment and the evolutionary processes that occurred over long periods of time in the native range (Facon et al. 2006). This situation has been reported in some studies of marine systems. For instance, cryptic invasions by an introduced lineage were the most likely explanation for a new rapid expansion of *Carcinus maenas* along the NE Pacific shoreline (Roman 2006; Tepolt et al. 2009). This ‘failure-success’ scenario is difficult to prove without

1 regular temporal surveys and specimen collections, including sampling from early or  
2 unsuccessful introductions. Museum collections and / or herbarium specimens (e.g. Lees et al.  
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4 2011) are of primary interest to resolve this point. Studies of failed introductions and  
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6 monitoring invaders over time (e.g. survey of ‘boom and bust’ dynamics) are crucial for  
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8 understanding long-term invasion dynamics (Suarez et al. 2005; Strayer et al. 2006; Pérez-  
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10 Portela et al. 2012) although opportunities to study such events are rare.  
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14 The second possible scenario (Fig. 4) involves selection forces that affect the standing  
15 genetic variation (i.e. pre-existing polymorphisms) introduced from the native range. This  
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17 scenario mimics a situation often encountered in marine invertebrates with: 1) high genetic  
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19 diversity and little genetic structure in the native range, and 2) high propagule pressure  
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21 following the introduction process. Interestingly, scenario 2 differs from scenario 1 in that  
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23 introductions from isolated regions of the native range are not required. Introduction may  
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25 simply occur from a genetically polymorphic source, and subsequently selection (e.g. through  
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27 competition and predation) would follow. Selection on pre-existing polymorphisms may  
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29 operate over short time scales compared to selection on new mutations (Barrett and Schluter  
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31 2008; Vandepitte et al. 2014). The genetic variants on which selection operates in the  
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33 introduced range may be neutral or even slightly deleterious in the native range but confer  
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35 some advantage to the introduced individuals in their new environment. Genome scans are  
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37 appropriate methods to detect loci influenced by selection processes (i.e. outliers) in the  
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39 absence of knowledge about the genes and traits that may be under selection (Bierne et al.  
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41 2011). Using such an approach, Riquet et al. (2013) did not find any outlier loci when  
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43 comparing native American and introduced European populations of the slipper limpet *C.*  
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45 *fornicata*, while their study revealed outliers between populations in the native range. This  
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47 outcome suggests that either no selection on standing genetic variation occurred following the  
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49 introduction process or, simply, that genome scans, particularly when using a low number of  
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1 markers, may not be efficient for identifying selection during colonisation because of the  
2 short time elapsed since the introduction. In contrast, Rohfritsch et al. (2013) showed the  
3 existence of outlier loci in introduced European populations of the Pacific oyster *C. gigas*.  
4 Two groups of populations were identified based on some of the outliers, but such genetic  
5 structure was uncorrelated with the one observed from neutral loci. While the authors did not  
6 fully exclude the effect of secondary multiple introductions, they suggested that parallel  
7 adaptations occurred in similar environments (in this case fjord-like environments). The two  
8 examples detailed above advocate for further investigation of the adaptive genomic  
9 architecture of native and introduced populations.  
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21 The last selective evolutionary scenario (Fig. 4) is based on the idea that new adaptive  
22 genetic combinations could arise from intraspecific crosses among individuals characterized  
23 by different genetic backgrounds (Facon et al. 2008; Lawson Handley et al. 2011). Similarly  
24 to interspecific hybridisation (Ellstrand and Schierenbeck 2000; Seehausen 2004; de Carvalho  
25 et al. 2013), genetic admixture may create novel allelic combinations that enhance the fitness  
26 of introduced individuals. The underlying rationale is that new heritable traits arise because of  
27 novel genetic combinations and that selection operates on these new traits (Rius and Darling  
28 2014). In the same way as heterosis (or hybrid vigour) among interspecific lineages operates  
29 (e.g. Drake 2006), strong evidence of correlation between genetic admixture and fitness of  
30 introduced individuals has been shown (e.g. Kolbe et al. 2007; Turgeon et al. 2011). Although  
31 the consequences of genetic admixture of divergent genotypes have been rarely studied in  
32 marine systems, studies reporting genetic admixture in introduced populations are  
33 proliferating. Pineda et al. (2011) found an excess of admixed genotypes of two nuclear  
34 lineages in introduced populations of the ascidian *Styela plicata*, which suggests that selection  
35 favours novel genetic combinations. Similarly, introduced populations of other widespread  
36 ascidian species show genetic admixture of divergent lineages (Rius et al. 2012; Zhan et al.  
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2012). However, this does not necessarily mean that admixed genotypes are always favoured in the introduced range, as the predominance of unadmixed genotypes has also been reported (Ordóñez et al. 2013).

Taken together, pre-adaption, selection on standing genetic variation and genetic novelties arising from post-immigration genetic admixture can be important drivers influencing evolutionary outcomes. Further research should yield important insights about these mechanisms in marine introductions.

### **Concluding remarks and future directions**

Studies on marine invasion genetics have generally shown that introduced populations rarely suffer from founder effects and are characterized by high genetic diversity, most likely as a result of repeated introductions and high propagule pressure. Despite almost 20 years of research, our study suggest that marine invasion genetics is still in its infancy. Most studies in European seas have focussed on cryptic diversity and descriptive comparisons of genetic composition and structure, from which insights into introduction patterns (pathways, sources, connectivity, etc.) have been drawn. However, little information is available on the underlying evolutionary mechanisms involved in marine biological invasions.

The currently limited genetic information of marine non-indigenous species makes the link between genetic information and rapid adaptation, phenotypic plasticity or fitness, elusive. The lack of comprehensive associations between genetic diversity and the success of marine biological invasions also brings into question the plausibility of using neutral genetic diversity as a proxy for adaptive genetic diversity and colonisation success (Hansen et al. 2012). The patterns observed to date advocate for further investigation of genetic and genomic architecture of marine non-indigenous species as well as in-depth analyses of the link between genotypes and phenotypes, considering in particular the fitness and adaptive potential of admixed genotypes.

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Key concerns such as the introduction history and the traits that determine invasiveness of non-indigenous species may benefit immensely from major advances in molecular biology and bioinformatics. Issues related to phylogeny and identification, historical and contemporaneous sources of non-indigenous species, population characteristics, and potential or realised adaptation can be tackled through an increasingly sophisticated analytical toolbox (reviews by Allendorf et al. 2010; Cock et al. 2010; Angeloni et al. 2012). The advent of next generation sequencing techniques (NGS) has revolutionized the potential for genetic and phenotypic analyses of the invasion process in non-model organisms. New markers such as single nucleotide polymorphisms (SNPs) can be easily developed from sequence data of full or partial genomes or transcriptomes (Everett et al. 2011; Helyar et al. 2011; Seeb et al. 2011; Reitzel et al. 2013). They allow for precise population genetic analyses, while at the same time allowing for the detection of adaptive changes through the study of outlier loci with non-neutral behaviour (Stapley et al. 2010; Rohfritsch et al 2013). NGS methods allow the study of hybridisation patterns of sibling species in secondary contact zones (e.g. Blank et al. 2012). They also enable the study of global gene expression patterns through so-called RNA-seq techniques (Wang et al. 2009; Wolf 2013), which can highlight differential expression levels of key genes as a response to new conditions in the introduced range. NGS techniques are also being increasingly used in the study of environmental DNA for several applications (Bohmann et al. 2014). Among these, environmental DNA can be a powerful tool for the early detection of introduced species, even before they become conspicuous in new localities (Bott et al. 2010; Bucklin et al. 2011). Another promising technique involves the sequencing of ancient DNA and historical (often low quality) DNA, which is highly applicable to museum collections that harbour alien taxa (e.g. Roman 2006; Lees et al. 2011). Finally, it is increasingly recognized that epigenetics, the study of heritable changes in gene expression and function that cannot be explained by changes in DNA

1 sequence, affects the phenotype and hence the potential to adapt. This happens through  
2 changes in the DNA methylation status, remodelling of chromatin structure and the regulation  
3 of expression mediated by microRNA (Bossdorf et al. 2008 and references therein). Such  
4 routes have yet to be explored in marine non-indigenous species, but may provide the key to  
5 understanding illusive factors affecting invasiveness.  
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11 Overall, the combination of genome-, transcriptome- and proteome-based methods  
12 will increasingly facilitate the investigation of a large set of issues in biological invasion  
13 studies; from the identification of non-indigenous species to the disentangling of the roles of  
14 phenotypic adaptation versus evolutionary change in the success of introductions. Much  
15 remains to be done in order for molecular techniques to become fully implemented in  
16 invasion genetics, especially with regards to describing and understanding the evolutionary  
17 mechanisms involved in the invasion process in marine ecosystems.  
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## Figures captions

**Fig. 1.** Genetic studies of marine biological invasions that include samples from European seas (see Table S1 for details). A) Number of studies over time, B) Number of studies per region, C) Number of species analysed genetically (white bars and left Y-axis, note that estimates are conservative: species complexes are considered as a single species, and multispecies works with no clear indication of the non-indigenous status of the species have been excluded) and number of introduced species reported [black bars and right Y-axis, using Zenetos et al. (2012) as a reference], D) Outcome of studies that compare patterns of genetic diversity between native and introduced populations, categorized by offspring dispersal capabilities (short or long, above) and the presence / absence of evidence for multiple introductions (below).

**Fig. 2.** Genetic diversity using: A) 17 microsatellites and B) 327 AFLP loci from native (black bars; NW Atlantic) and introduced (white bars; Mediterranean Sea, NE Atlantic and NE Pacific) populations of *Crepidula fornicata*. Different styles of dashed lines represent mean diversity across all populations of the native and introduced ranges respectively. Comparisons of genetic diversity among native and introduced populations yield non-significant differences ( $P = 0.99$  and  $P = 0.85$  for microsatellites and AFLP, respectively, based on a permutation test, 5000 permutations of individuals between groups). Sources of the introduction are presumably located in the northern part of the native range. Data from Riquet et al. (2013).

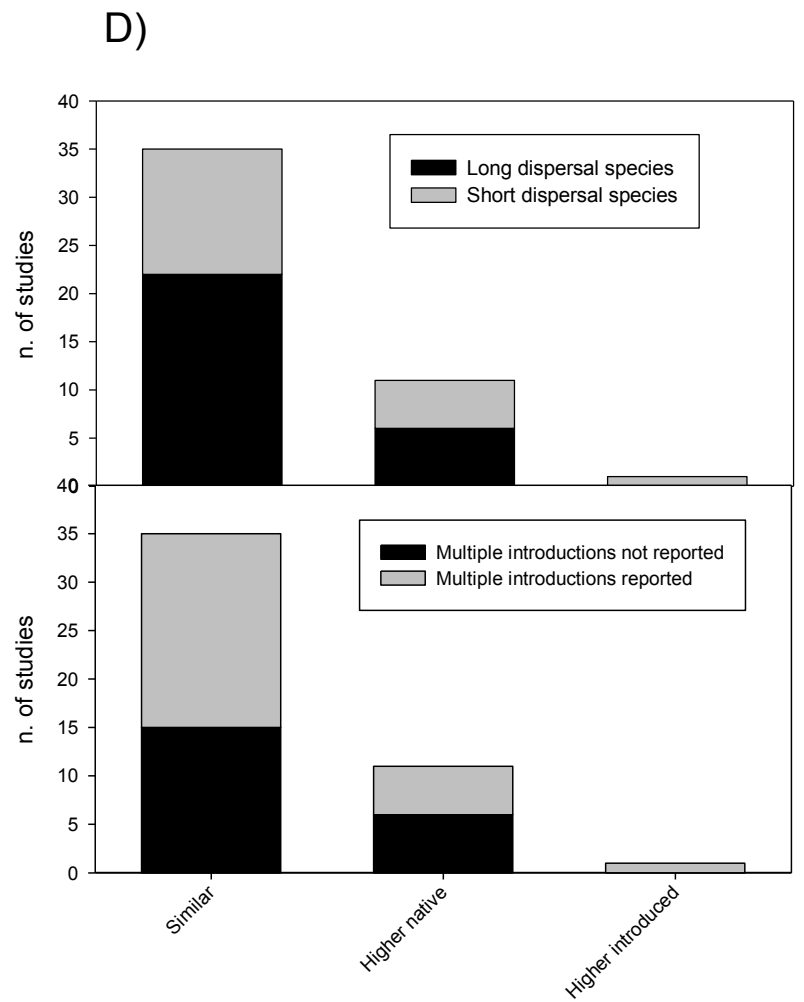
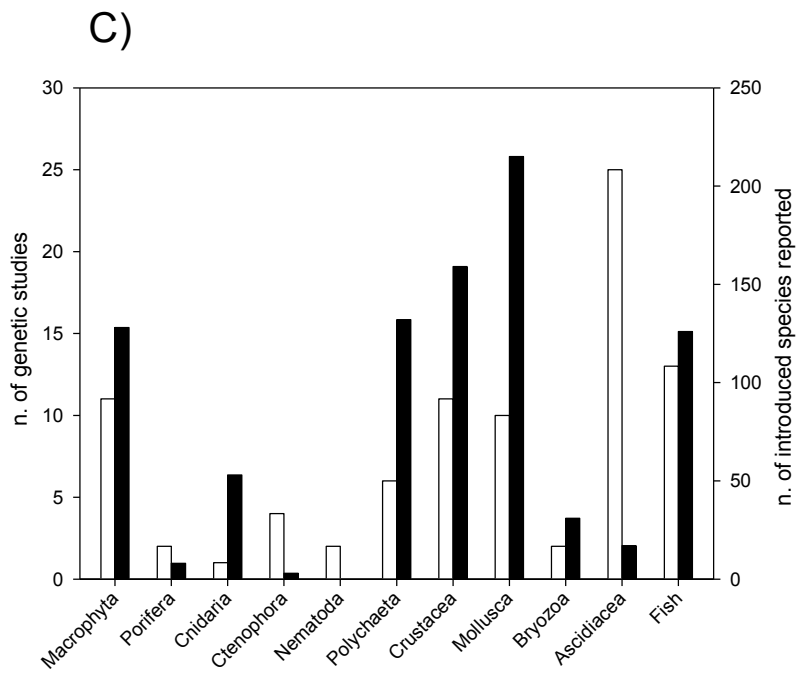
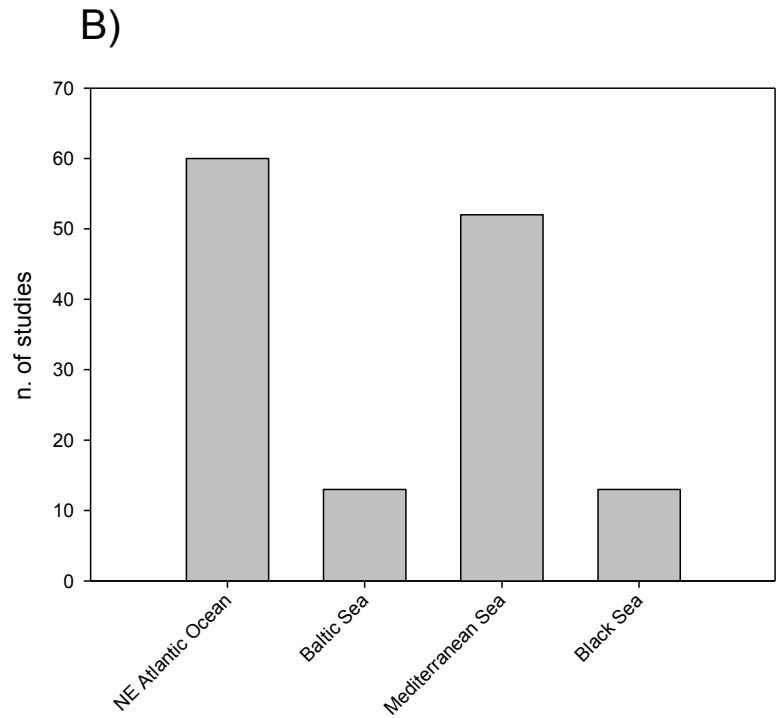
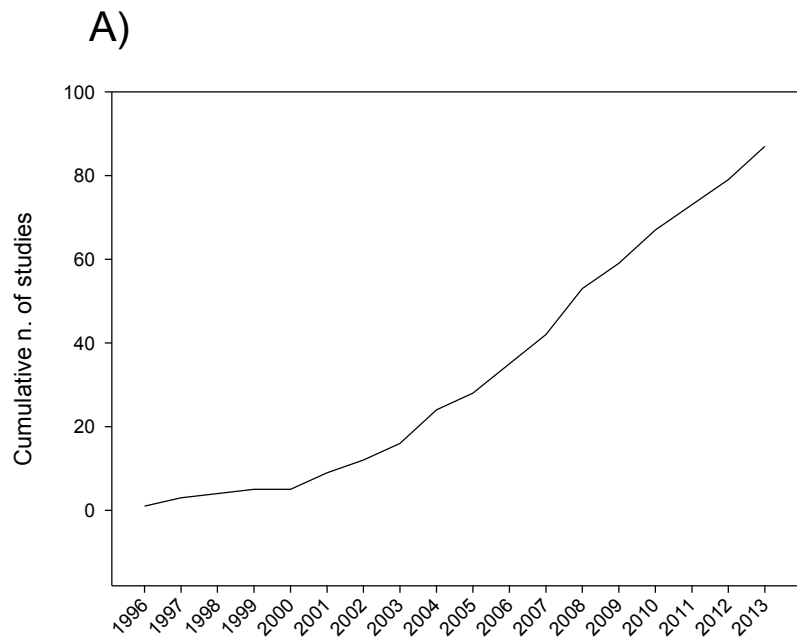
**Fig. 3.** Increased genetic variance and admixture between genetically divergent lineages of *Cyclope neritea* in the NE Atlantic introduced range (red line in left map; population samples a-e) as compared to its native range in the Mediterranean and Black seas (yellow line; samples 1-13). Ephemeral populations are also documented along the Iberian Peninsula (blue line, samples f-h). Circles show the frequency distribution of mitochondrial haplotypes at

1 population level. Colours indicate different groups of divergent haplotypes. Details are  
2 provided in Simon-Bouhet et al. (2006), but for two populations (Adriatic and Black seas) that  
3 are included here but were not analysed in the paper.  
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7 **Fig. 4.** Potential selective processes acting during the introduction of multiple genetically  
8 polymorphic and / or divergent sources. Each colour represents individuals with a specific  
9 genetic background from the native range that may be neutral, adaptive or deleterious in the  
10 introduced range. Squares in the native range feature genetically divergent groups of  
11 individuals along the species range. Arrows in the introduced range represent temporal  
12 changes. Note that the first scenario includes temporally sequential introductions but that the  
13 same outcome may occur when the three genotypes are introduced at the same time.  
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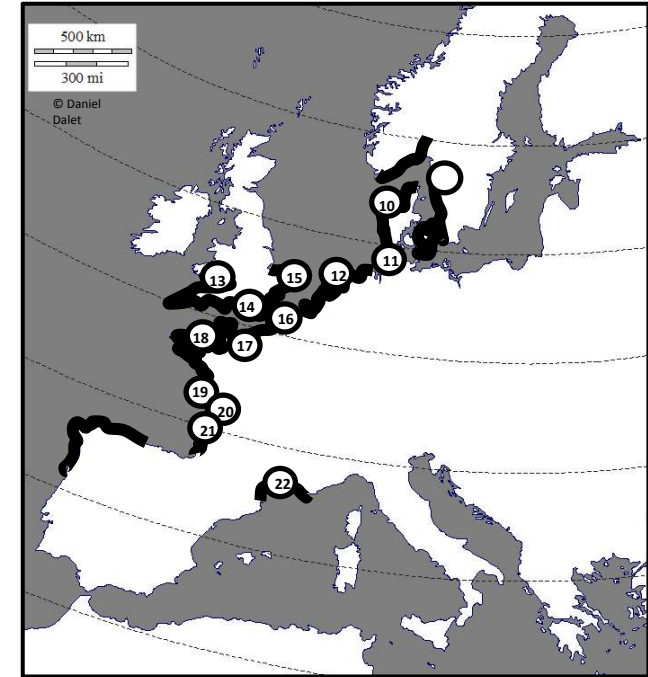
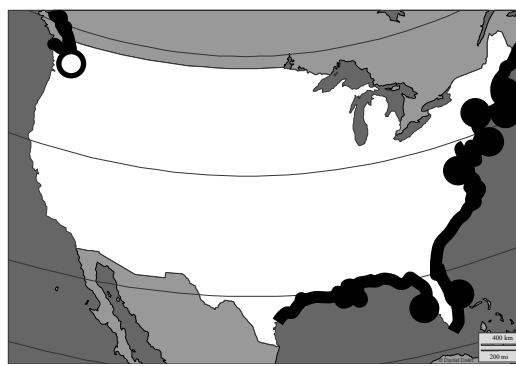


Figure 1

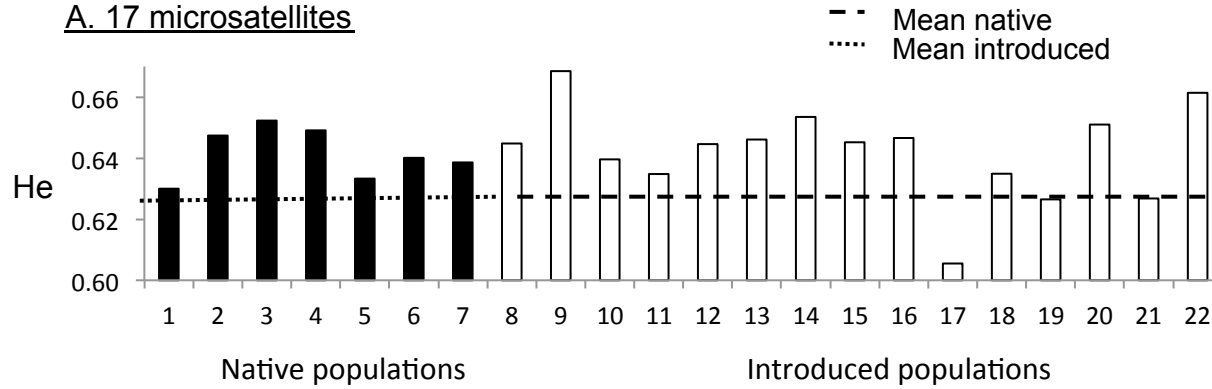


Genetic diversity of native vs introduced populations

Figure 2



**A. 17 microsatellites**



**B. 327 AFLPs**

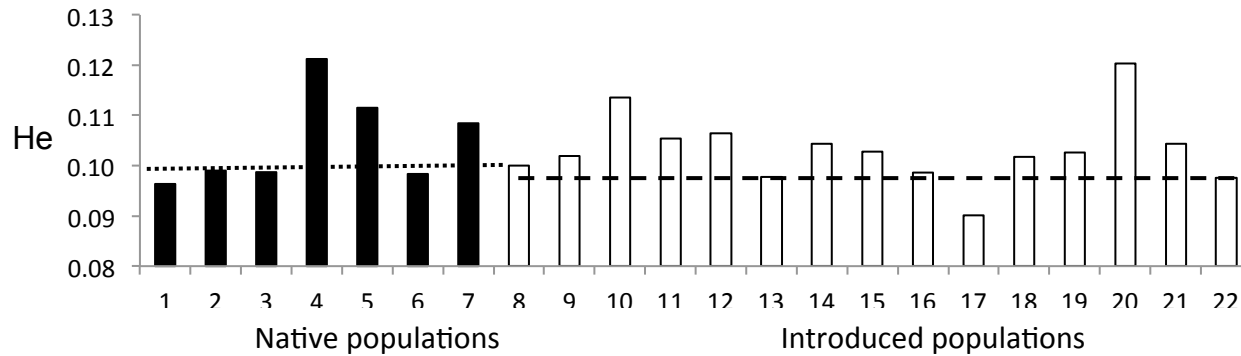


Figure 3

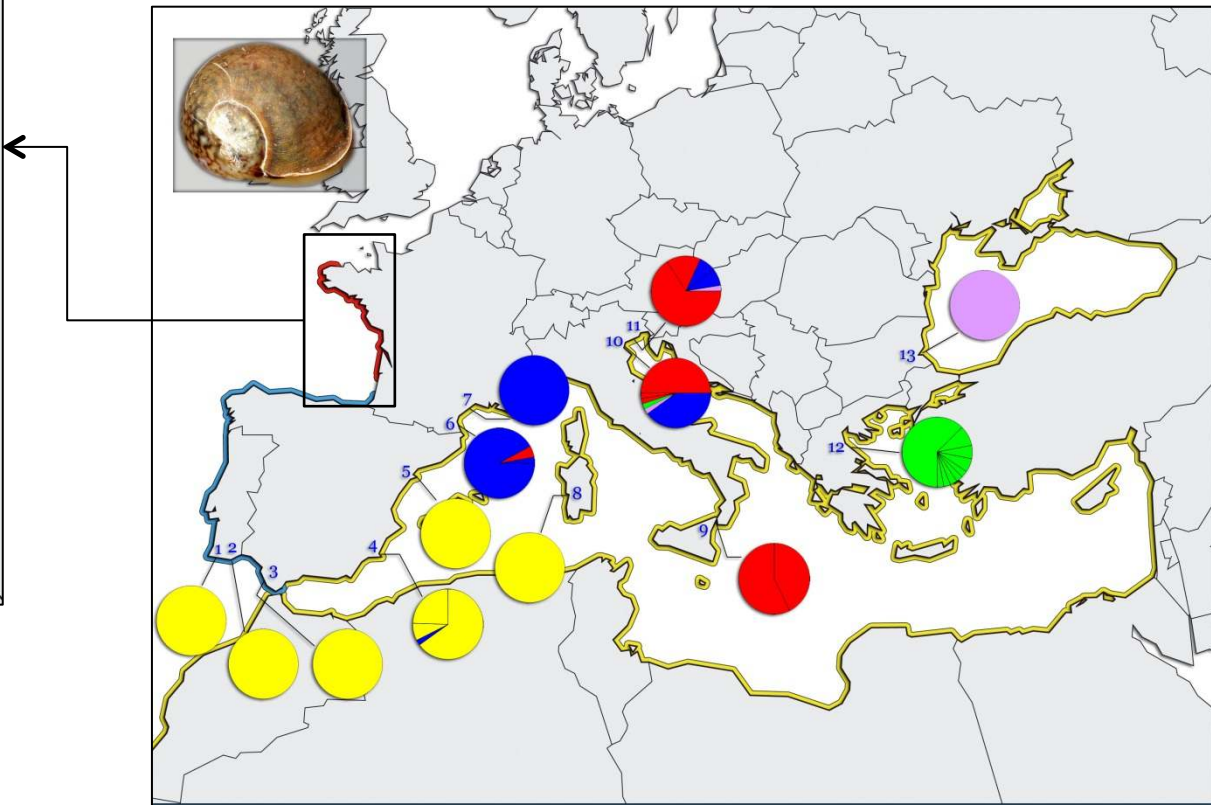
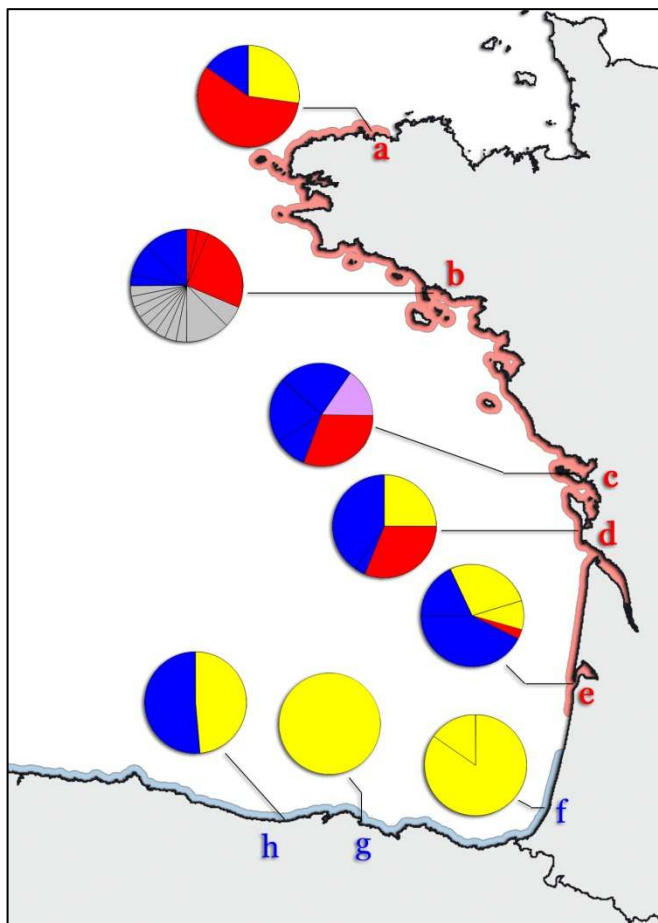
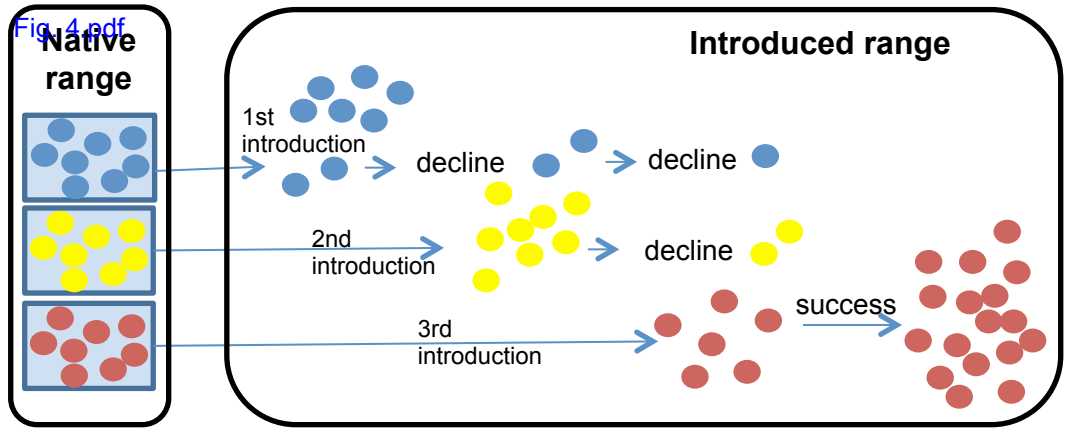
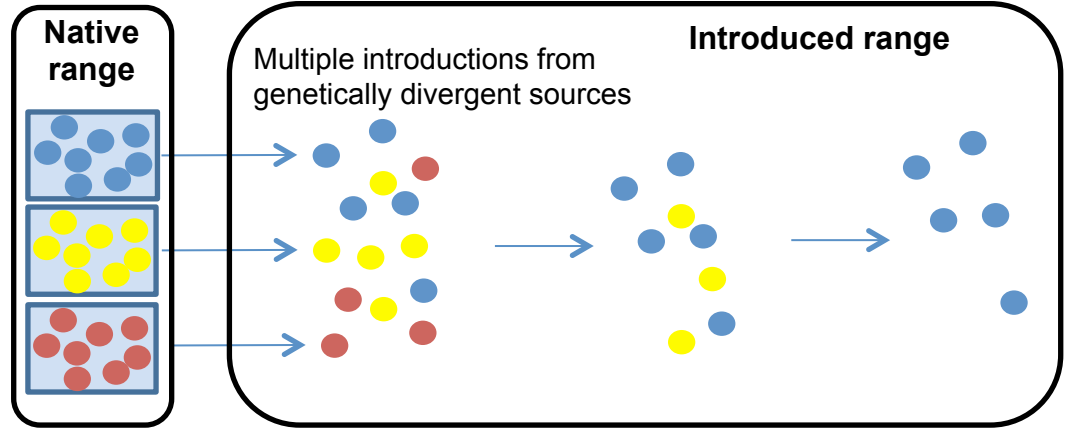


Figure 4

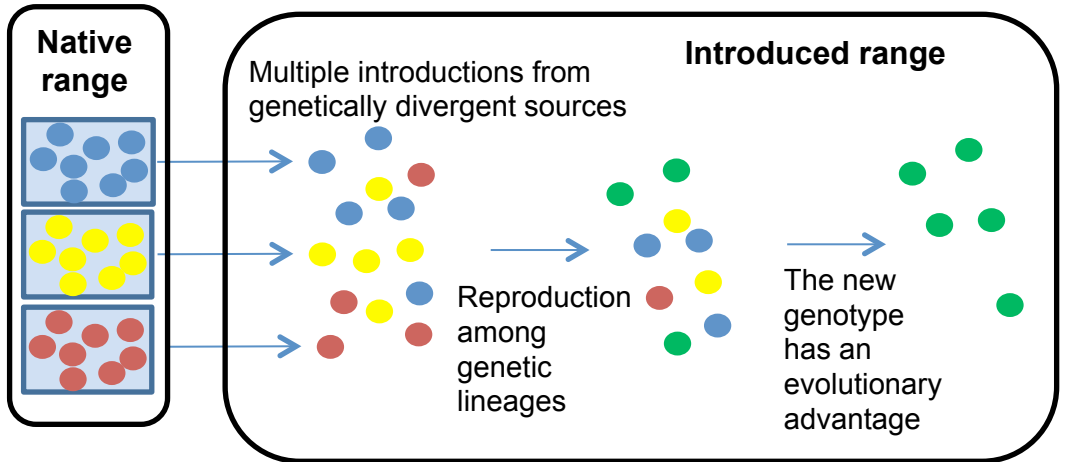
**Scenario 1**  
 Repeated introductions but only one introduction (likely a pre-adapted genotype) becomes successful



**Scenario 2**  
 Selective forces act on the overall introduced genetic variation



**Scenario 3**  
 Evolutionary novelties arise following genetic admixture



Time →

Lead author	Year	Taxon	Phylum	Dispersal	Genetic marker	Region in European waters				Genetic Diversity Native Range	Genetic Diversity Introduced Range	Evidence of multiple introductions
						NI Atlantic Ocean	Baltic Sea	Mediterranean Sea	Black Sea			
Bauer	1996	<i>Marenzelleria</i> sp.	Annelida	Long	Allozymes	X				Similar native and introduced	Yes	
Bastrop	1997	<i>Marenzelleria</i> spp.	Annelida	Long	Allozymes and mitochondrial 16S rDNA	X	X			Data in Rohrer study	Yes	
Andrew	1999	<i>Sabella spallanzanii</i>	Annelida	Long	Allozymes	X		X		Not provided	Not provided	Not provided
Ivansson	1998	<i>Caulerpa taxifolia</i>	Chlorophyta	Long	ITS-DNA 18S rDNA, 28S rDNA	X		X		Not provided	Not provided	Not provided
Golani	1999	7 <i>Mullididae</i> species	Chordata	Long	Electrophoretic examination of presumptive loci	X		X		Similar native and introduced	Not provided	Not provided
Wiedenmann	2001	<i>Caulerpa taxifolia</i>	Chlorophyta	Long	DNA fingerprinting	X		X		Not provided	Not provided	Not provided
Patti	2001	<i>Sabella spallanzanii</i>	Annelida	Long	ITS2	X		X		High	Not provided	Not provided
Tarjuelo	2001	<i>Cleavelina lepadiformis</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		Not provided	Not provided	Not provided
Mchvor	2001	<i>Polysiphonia horveyi</i>	Rhodophyta	Long	rbcL sequence	X	X	X		Not provided	Not provided	Yes
Bucchiarelli	2002	<i>Aethanammus lucanosus</i>	Chordata	Long	Mitochondrial control region	X		X		Similar native and introduced	Not provided	Not provided
Durand	2002	<i>Caulerpa racemosa</i>	Chlorophyta	Long	ITS1-ITS2 and 18S	X		X		Not provided	Not provided	Yes
Messner	2002	<i>Caulerpa taxifolia</i>	Chlorophyta	Long	Nuclear rDNA ITS, cp 16S rDNA intron-2	X		X		Not provided	Not provided	Yes
Turon	2003	<i>Cleavelina lepadiformis</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		Not provided	Not provided	Not provided
Dupont	2003	<i>Crepidula fornicata</i>	Mollusca	Long	Polyomorphic enzyme loci	X		X		Similar native and introduced	Not provided	Yes
Bonhomme	2003	<i>Siganus rivulatus</i>	Chordata	Long	mtDNA cytochrome b gene	X		X		Not provided	Not provided	Yes
Hassan	2003	<i>Siganus rivulatus</i> and <i>Siganus luridus</i>	Chordata	Long	mtDNA cytochrome b gene, Intron length polymorphisms and inter-Simple Sequence Repeats	X		X		Similar native and introduced	Not provided	Yes
Andreakis	2004	<i>Asparagopsis tomentosus</i> and <i>Asparagopsis armata</i>	Rhodophyta	Long	Nuclear LSU rDNA gene, plastid <i>Rubisco</i> spacer and mitochondrial <i>cox2-3</i> spacer	X		X		Not provided	Not provided	Not provided
Shefer	2004	<i>Brochodontes phurazonis</i>	Mollusca	Long	cytochrome c oxidase subunit I	X		X		Similar native and introduced	Not provided	Not provided
Roman	2004	<i>Carcinus maenas</i>	Arthropoda	Long	cytochrome c oxidase subunit I	X		X		High	Not provided	Not provided
Duran	2004	<i>Cambar crabs</i>	Porifera	Short	Nuclear ribosomal internal transcribed spacers (ITS-1 and ITS-2)	X		X		High	Not provided	Not provided
Theriault	2004	<i>Mytilus leucophaea</i>	Mollusca	Long	mtDNA (16S rRNA and cytochrome c oxidase subunit I)	X		X	X	Not provided	Not provided	Not provided
Martel	2004	<i>Ocinetrella inornatus</i>	Mollusca	Short	Allozymes and mitochondrial DNA	X		X		High	Not provided	Not provided
Martel	2004	<i>Ocinetrella inornatus</i>	Mollusca	Short	Polyomorphic enzyme loci	X		X		Similar native and introduced	Not provided	Not provided
Cristescu	2004	<i>Echinogammarus ischnus</i>	Arthropoda	Short	cytochrome c oxidase subunit I	X		X	X	High	Low	Not provided
Provan	2005	<i>Codium fragile tomentosoides</i>	Chlorophyta	Long	Microsatellites and DNA sequence data from three regions of the plastid genome	X		X		Similar native and introduced	Not provided	Not provided
Vosjan	2005	<i>Undaria pinnatifida</i>	Ochrophyta	Long	Mitochondrial intergenic spacer regions	X		X		High	Low and high	Yes
Cristescu	2005	14 Ponto-Caspian crustacean species	Arthropoda	Long	cytochrome c oxidase subunit I, 16S and 28S	X		X	X	Not provided	Yes	Not provided
Hassan	2005	<i>Upeneus moluccensis</i>	Chordata	Long	Nuclear (intron) and mitochondrial markers	X		X		Not provided	Not provided	Not provided
López-Legentil	2006	<i>Batryllus schlosseri</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		Unresolved origin	High	Not provided
Ben-Shlomo	2006	<i>Batryllus schlosseri</i>	Chordata	Short	Microsatellites	X		X		Unresolved origin	High	Not provided
Dupont	2006	<i>Crepidula fornicata</i>	Mollusca	Long	Microsatellites	X		X		Not provided	High	Yes
Simon-Bouhet	2006	<i>Cyrtolpe neritea</i>	Mollusca	Short	cytochrome c oxidase subunit I	X		X		Low	High	Yes
Kelly	2006	<i>Gommarus tigrinus</i>	Arthropoda	Long	cytochrome c oxidase subunit I	X		X		High	Low and high	Yes
Bastrop	2006	<i>Marenzelleria neglecta</i>	Annelida	Long	mtDNA (16S rRNA and cytochrome c oxidase subunit I, cytochrome b)	X		X		Not provided	Not provided	Not provided
Azzurro	2006	<i>Siganus luridus</i>	Chordata	Long	Mitochondrial control region	X		X		Similar native and introduced	Not provided	Yes
Andreakis	2007	<i>Asparagopsis tomentosus</i>	Rhodophyta	Long	Hypervariable domains D1-D8 of the nuclear rDNA LSU, the plastid spacer between the large and small subunits of <i>Rubisco</i> and the mitochondrial <i>cox 3</i> intergenic spacer	X		X		High	Not provided	Not provided
Nydam	2007	<i>Ciona intestinalis</i> species complex	Chordata	Short	cytochrome c oxidase subunit I	X		X		Not provided	Not provided	Not provided
Herborg	2007	<i>Erachter sinensis</i>	Arthropoda	Long	Microsatellites	X		X	X	Not provided	Low and high	Not provided
Laessle	2007	<i>Ciona intestinalis</i> species complex	Chordata	Short	Mitochondrial genome	X		X		Not provided	Not provided	Not provided
Caputi	2007	<i>Ciona intestinalis</i> species complex	Chordata	Short	cytochrome c oxidase subunit I, nuclear (ITS-2, EPIC Hox4 and Gata introns, Hox13 exon)	X		X		Not provided	Not provided	Not provided
Dupont	2007	<i>Carella eumytila</i>	Chordata	Short	Microsatellites	X		X		Similar native and introduced	Not provided	Not provided
Dupont	2007	<i>Fistulohyla commersonii</i>	Chordata	Long	Mitochondrial control region	X		X		High	Low and high	Not provided
Ashton	2008	<i>Caprella mutica</i>	Arthropoda	Short	cytochrome c oxidase subunit I	X		X		High	Low	Yes
Audzinyte	2008	<i>Hemimysis anomala</i>	Arthropoda	Short	cytochrome c oxidase subunit I	X		X	X	Similar native and introduced	Yes	Not provided
Blaževic	2008	<i>Littorina littorea</i>	Mollusca	Long	cytochrome b and cytochrome oxidase I	X		X		High	Low	Yes
Blank	2008	<i>Marenzelleria</i> spp.	Annelida	Long	16S rDNA, cytochrome b, cytochrome oxidase subunit I	X		X		Not provided	Not provided	Not provided
Wiegoss	2008	<i>Anguillicola crassus</i>	Nematoda	Short	Microsatellite loci and one mitochondrial marker	X		X	X	Similar native and introduced	Not provided	Not provided
Darling	2008	<i>Carcinus maenas</i> and <i>C. aestuarii</i>	Arthropoda	Long	cytochrome c oxidase subunit I	X		X		High	Low	Yes
Ashton	2008	<i>Caprella mutica</i>	Arthropoda	Long	cytochrome c oxidase subunit I	X		X		High	Low and high	Yes
Hughes	2008	<i>Celleporella</i> spp.	Bryozoa	Short	cytochrome c oxidase subunit I	X		X		Not provided	Not provided	Yes
Rius	2008	<i>Microcosmus squamiger</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		Similar native and introduced	Not provided	Yes
Wiegoss	2008	<i>Anguillicola crassus</i>	Nematoda	Short	Microsatellites	X		X		High	Low	Not provided
Brown	2008	<i>Apolonia melanostoma</i>	Chordata	Long	cytochrome b and microsatellites	X		X	X	High	Low and high	Not provided
Folino-Roreem	2009	<i>Cordyliphora</i> spp.	Cnidaria	Long	2 mitochondrial loci (16S rRNA and cytochrome c oxidase subunit I) and one nuclear locus (28S rRNA)	X		X		Not provided	Not provided	Not provided
Neilson	2009	13 Ponto-Caspian "neogobelin" species	Chordata	Long	Mitochondrial genes (cyt b and cytochrome c oxidase subunit I) and nuclear genes (RAG1 and S7)	X		X	X	Not provided	Not provided	Not provided
Stefaniak	2009	<i>Didemnum vesillum</i>	Chordata	Short	cytochrome c oxidase subunit I and thox2 gene	X		X		Not provided	Not provided	Not provided
Feldheim	2009	Different species of gobies	Chordata	Long	Microsatellites	X		X		High	Low and high	Not provided
Banche	2009	<i>Sorus ghibbanus</i>	Chordata	Long	Mitochondrial control region and ATPase	X		X		Similar native and introduced	Not provided	Not provided
Dupont	2009	<i>Styela clava</i>	Chordata	Short	Microsatellites	X		X		Not provided	High	Yes
Zhan	2010	<i>Ciona intestinalis</i> species complex	Chordata	Short	Mitochondrial (cytochrome c oxidase subunit 3-NADH dehydrogenase subunit 1 region and NADH dehydrogenase subunit 4 gene) and nuclear (internal transcribed spacer 1)	X		X		Unknown origin	Low and high	Not provided
Reusch	2010	<i>Mnemopsis leidy</i>	Ctenophora	Long	Microsatellites	X		X		Similar native and introduced	Not provided	Not provided
Dupont	2010	<i>Styela clava</i>	Chordata	Short	Microsatellites	X		X		Similar native and introduced	Yes	Not provided
Reuschel	2010	<i>Palaeomon elegans</i>	Arthropoda	Long	16S and cytochrome c oxidase subunit I	X		X	X	High	Low	Not provided
Haydar	2010	<i>Malgula montanensis</i>	Chordata	Short	Nuclear 18S rDNA and mtDNA cytochrome c oxidase subunit I sequences	X		X		High	Low and high	Yes
Blaževic	2010	<i>Carcinus maenas</i>	Arthropoda	Long	cytochrome c oxidase subunit I	X		X		High	Low	Yes
Cheang	2010	<i>Sargassum muticum</i>	Ochrophyta	Short	RUBISCO spacer, TrnW <sub>1</sub> spacer and ITS2	X		X		Similar native and introduced	Not provided	Not provided
Dupont	2010	<i>Styela clava</i>	Chordata	Short	Microsatellites	X		X		Similar native and introduced	Yes	Not provided
Lepasseur	2011	<i>Batryllus schlosseri</i> and <i>Batryllus violaceus</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		Unknown origin and genetic diversity variable	Yes	Not provided
Grubis	2011	<i>Undaria pinnatifida</i>	Ochrophyta	Short	Microsatellites	X		X		Not provided	High	Not provided
Ghahabi	2011	<i>Mnemopsis leidy</i>	Ctenophora	Long	ITS-1 and ITS-2	X		X		High	Low and high	Yes
Neilson	2011	<i>Neogobius fluviatilis</i> and <i>N. pallasi</i>	Chordata	Long	mtDNA cytochrome b, nuclear RAG-1 and microsatellites	X		X	X	High	Low	Not provided
Goldstein	2011	<i>Styela clava</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		High	Low and high	Yes
Pineda	2011	<i>Styela plicata</i>	Chordata	Short	cytochrome c oxidase subunit I and ANT	X		X		Unknown origin	Low and high	Yes
Bock	2012	<i>Batryllus schlosseri</i>	Chordata	Short	cytochrome c oxidase subunit I, 18S, microsatellites	X		X		Unknown origin and genetic diversity variable	Not provided	Not provided
Golani	2012	<i>Callinectes filamentosus</i>	Chordata	Long	2 mitochondrial loci (control region and cytochrome oxidase I) and two nuclear loci (2nd intron of the ribosomal protein S7 and rhodopsin)	X		X		Similar native and introduced	Yes	Not provided
Rius	2012	<i>Herdmania homus</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		High	Low and high	Yes
Rius	2012	<i>Microcosmus squamiger</i>	Chordata	Short	Microsatellites	X		X		Similar native and introduced	Not provided	Not provided
Guardiola	2012	<i>Paraleucilla magna</i>	Porifera	Short	Microsatellites	X		X		Not provided	High	Not provided
Pérez-Portela	2012	<i>Paraphora japonica</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		Not provided	Low and high	Yes
Riquet	2013	<i>Crepidula fornicata</i>	Mollusca	Long	Microsatellite and AFLP-based genome scans	X		X		Similar native and introduced	Not provided	Not provided
Pérez-Portela	2013	<i>Diplomeia listerianum</i>	Chordata	Short	cytochrome c oxidase subunit I	X		X		Unknown origin and genetic diversity variable	Not provided	Not provided
Ordóñez	2013	<i>Microcosmus squamiger</i>	Chordata	Short	Microsatellites	X		X		Similar native and introduced	Yes	Not provided
Bioke	2013	<i>Mnemopsis leidy</i>	Ctenophora	Long	Microsatellites	X		X	X	High	Low	Yes
Rohlfritsch	2013	<i>Crossostrea gigas</i>	Mollusca	Long	Microsatellites, AFLPs and SNPs	X		X		Similar native and introduced	Not provided	Not provided
Bishop	2013	<i>Asterocarpa humilis</i>	Chordata	Short	18S and cytochrome c oxidase subunit I	X		X		Not provided	Not provided	Not provided
Ghahabi	2013	<i>Mnemopsis leidy</i>	Ctenophora	Long	cytochrome c oxidase subunit I and ITS-1 & ITS-2	X		X	X	Similar native and introduced	Not provided	Not provided
Mackie	2013	<i>Watersipora subaerata</i>	Bryozoa	Short	cytochrome c oxidase subunit I	X		X		Not provided	Low and high	Yes