# Matrix Remodeling in Pulmonary Fibrosis and Emphysema

Tejaswini Kulkarni<sup>1,2</sup>, Philip O'Reilly<sup>1,2</sup>, Veena B. Antony<sup>1,2</sup>, Amit Gaggar<sup>1,2,3</sup>, and Victor J. Thannickal<sup>1,2,3</sup>

<sup>1</sup>Department of Medicine, Division of Pulmonary, Allergy and Critical Care Medicine, University of Alabama at Birmingham, Birmingham, Alabama; <sup>2</sup>Program in Protease and Matrix Biology Center, Birmingham, Alabama; and <sup>3</sup>Birmingham VA Medical Center, Birmingham, Alabama

ORCID ID: 0000-0002-4251-4988 (T.K.).

### Abstract

Pulmonary fibrosis and emphysema are chronic lung diseases characterized by a progressive decline in lung function, resulting in significant morbidity and mortality. A hallmark of these diseases is recurrent or persistent alveolar epithelial injury, typically caused by common environmental exposures such as cigarette smoke. We propose that critical determinants of the outcome of the injury-repair processes that result in fibrosis versus emphysema are mesenchymal cell fate and associated extracellular matrix dynamics. In this review, we explore the concept that regulation of mesenchymal cells under the influence of soluble factors, in particular transforming growth factor- $\beta_1$ , and the extracellular matrix determine the divergent tissue remodeling responses seen in pulmonary fibrosis and emphysema.

# **Keywords:** extracellular matrix; myofibroblasts; transforming growth factor-β; pulmonary fibrosis; emphysema

### **Clinical Relevance**

Pulmonary fibrosis and emphysema are chronic lung diseases that result from dysregulated injury-repair responses to chronic lung injury. This review explores the concept that the distinct clinical-radiological-pathological presentation of these diseases is determined by a central role of the extracellular matrix and the associated fate/function of extracellular matrix-producing mesenchymal cells.

Pulmonary fibrosis and emphysema represent distinct clinical-radiologicalpathological syndromes that result from injury-repair responses to chronic lung injury, most notably cigarette smoke. The pathobiological mechanisms that result in these distinct remodeling responses in the lung are not well understood. Idiopathic pulmonary fibrosis (IPF) is a chronic, progressive, and fatal form of interstitial lung disease that occurs primarily in older individuals; it is associated with disordered deposition of extracellular matrix (ECM) (1, 2). IPF is defined by the histopathological pattern of usual interstitial pneumonia characterized by spatial-temporal heterogeneity and the accumulation of myofibroblasts in

fibroblastic foci (3, 4). Emphysema is a chronic, progressive, clinical phenotype of chronic obstructive pulmonary disease (COPD), with increasing morbidity, mortality, and economic burden worldwide. COPD is characterized by airway remodeling with increased ECM deposition in the bronchi and bronchioles; ECM degradation appears to be enhanced in the alveolar regions (5, 6). IPF and COPD are distinctive in their clinical presentations and physiology, yet they carry common risk factors and presumably distinct pathobiological mechanisms.

Cigarette smoke is the most common risk factor for both IPF and COPD. In addition to cigarette smoke, a number of potential environmental exposures have been linked to IPF; these include metal dusts (brass, lead, and steel), agricultural dust, and wood dust (7, 8). In addition, a substantial proportion of nonsmokers with emphysema from occupational exposure to mineral and biological dusts, gases, and fumes, in particular biomass fuel, in enclosed environments are being identified (9). These abnormalities in IPF and emphysema are typically associated with chronic inflammation from recurrent environmental insults and persistent tissue injury. However, emerging data support the concept that dysregulated lung repair after alveolar epithelial injury may give rise to these distinct clinical entities. Several common host response pathways after lung injury have been identified in

(Received in original form May 16, 2015; accepted in final form December 26, 2015)

This work was supported by National Institutes of Health (NIH) grants P01 HL114470 and R01 AG046210 (V.J.T.), and NIH grant HL102371 and Veteren's Administration grant I01BX001756 (A.G.).

Author Contributions: Conception and design—T.K. and V.J.T.; drafting manuscript for intellectual content—T.K., P.O'R., V.B.A., A.G., and V.J.T.

Am J Respir Cell Mol Biol Vol 54, Iss 6, pp 751-760, Jun 2016

Copyright © 2016 by the American Thoracic Society

Originally Published in Press as DOI: 10.1165/rcmb.2015-0166PS on January 7, 2016

Internet address: www.atsjournals.org

Correspondence and requests for reprints should be addressed to Victor J. Thannickal, M.D., 1900 University Boulevard, 422 THT, Birmingham, AL 35294. E-mail: vjthan@uab.edu

both these chronic lung diseases, yet there are uncertainty and debate regarding theories for the pathogenesis of their phenotypes (10). In this review, we explore the concept that the distinct pathobiology of these clinical syndromes is determined by a central role for the ECM and the associated fate/function of ECM-producing mesenchymal cells.

# Mechanisms of Lung Injury and Repair

Host responses to persistent tissue injury induce inflammatory and repair responses that involve myriad interactions among various cells and soluble factors in an attempt to restore normal organ structure and function. Disruption of this tightly orchestrated host response can lead to distinct pathological consequences: pulmonary fibrosis and emphysema.

Current concepts of the pathogenesis of pulmonary fibrosis center primarily on dysregulated wound repair and altered epithelial-mesenchymal communication after repetitive epithelial injury. A number of factors that contribute to this dysregulation have been identified and include impaired fibrinolysis, imbalance of tissue inhibitor of metalloproteinases (TIMPs)-matrix metalloproteinases (MMPs), oxidative stress, elaboration of profibrotic cytokines/chemokines, and eicosanoid imbalance (11). Epithelial and inflammatory cells release several profibrotic mediators, including transforming growth factor (TGF)- $\beta$ 1, platelet-derived growth factor (PDGF), connective tissue growth factor, IL-1β, and tumor necrosis factor- $\alpha$  (12–16). Various MMPs expressed by different cell types in the alveolar microenvironment mediate diverse pro- and antifibrotic functions (17). A consistent feature of the pathobiology of IPF is the accumulation of myofibroblasts that acquire an apoptosisresistant and invasive phenotype, thus perpetuating alveolar collapse and ECM remodeling (18, 19) (Figure 1).

In emphysema, inflammatory cells accumulate in the airways and alveolar spaces in response to chronic exposure to cigarette smoke or environmental toxins. They release proteolytic enzymes including elastases (such as neutrophil elastase and macrophage metalloelastase [MMP-12]) and collagenases/gelatinases (such as

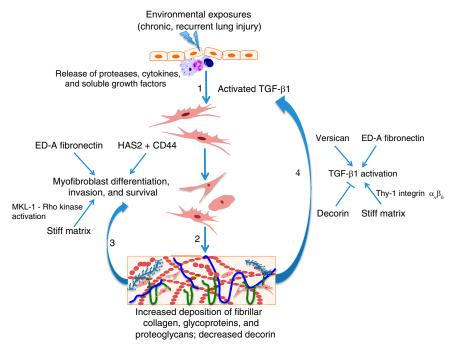
MMP-8 and MMP-9) (20-22). Proteolytic cleavage of elastin and other ECM components by these proteases results in degradation of the ECM, leading to loss of elastic recoil and the generation of matrikines, which may perpetuate inflammatory responses. Cleaved elastin fragments and collagen-derived peptide Pro-Gly-Pro (PGP) from ECM degradation mediate chemotactic activity and promote airway inflammation, thus precipitating a self-propagating cycle of inflammation and ECM proteolysis (23, 24). In addition, proteases may inactivate TIMPs, thus reducing the antiproteolytic activity within the matrix and leading to a protease-antiprotease imbalance (22, 25). Furthermore, the normal repair functions of fibroblasts may be suppressed, in part because of their reduced responsiveness to TGF-β1 signals (26) (Figure 2). Thus, dysregulated inflammation and repair may result in distinct fibroblast-ECM remodeling responses that account for the variability in the clinical-pathological presentations seen in pulmonary fibrosis and emphysema.

## **Fibroblast Biology**

Fibroblasts are versatile cells that undergo differentiation to myofibroblasts and secrete ECM proteins during physiological tissue repair in almost all organ systems studied (27). Myofibroblasts are defined by their capacity to contract and by the expression of contractile proteins, particularly,  $\alpha$ -smooth muscle actin  $(\alpha$ -SMA) stress fibers (28). The highly controlled synthetic capacity of myofibroblasts is an important feature of tissue repair and ECM remodeling in the lung. The deletion of PDGF-A-expressing myofibroblasts results in developmental emphysema caused by a failure of alveolar septation (29). Aberrant fibroblast responses in the injured alveolar microenvironment may be a key mechanism driving the distinct clinical phenotypes in pulmonary fibrosis and emphysema. Pulmonary fibrosis is characterized by persistent and heightened myofibroblast activity, whereas emphysema appears to be associated with reduced fibroblastic activity.

Fibroblastic foci (FF), the histopathological hallmark of usual interstitial pneumonia, comprise aggregates of fibroblasts and myofibroblasts. Previous studies suggested that myofibroblasts originate from multiple different sources (30-32); however, more recent studies indicate that almost all myofibroblasts originate from the differentiation of resident fibroblast progenitors (33). The regulation of myofibroblast differentiation in pulmonary fibrosis is complex and involves the action of growth factors/cytokines such as TGF- $\beta$ 1 (34, 35) in concert with ECM-derived biochemical and biomechanical signals (36-42). The acquisition of an apoptosis-resistant myofibroblast phenotype by both soluble mediators and the ECM may drive the nonresolving nature of IPF (43, 44). In addition, the acquisition of invasive properties by myofibroblasts (45) may lead to disruption of the basement membrane, which is a distinctive feature of IPF. In addition to enhanced synthetic and invasive capacity, activated myofibroblasts exert isometric contractile activity. This activity is maintained by a single-cell lock-step model via the combined actions of intracellular calcium and Rho/Rho kinase (Rho/ROCK)mediated calcium-independent contraction (46). The contractile activity of myofibroblasts results in macroscopic tissue contracture (28), which likely contributes to alveolar collapse and the observed reduction in total lung capacity in IPF.

In contrast to pulmonary fibrosis, there appears to be an impaired fibroblast response to epithelial injury in emphysema. Fibroblasts obtained from patients with COPD demonstrate decreased proliferation, migration, and contractility (26). This effect occurs potentially through increased expression of prostaglandin E2, a known inhibitor of fibroblast repair function, with a resultant decrease in synthesis and deposition of ECM components (47-49). Importantly, fibroblasts in emphysema have reduced responsiveness to transcriptional regulation by TGF-β1 and myofibroblast differentiation (26). Neutrophil elastase, which is highly expressed in emphysema, releases epidermal growth factor from the cell surface and initiates epidermal growth factor receptor/mitogen-activated protein kinase kinase/extracellular signal-regulated kinase (EGFR/MEK/ERK) signaling to down-regulate elastin synthesis in rat lung fibroblasts (50, 51); this is mediated at least in part by stabilization of the Smad



**Figure 1.** Loss of cellular and extracellular matrix (ECM) homeostasis in the pathogenesis of pulmonary fibrosis. Exposure to inhaled toxins such as cigarette smoke leads to epithelial cell injury, inflammation, and release of proteases and soluble growth factors. Latent transforming growth factor-β1 (TGF-β1) may be activated by multiple mechanisms such as integrin  $\alpha_v\beta_6$  (1). TGF-β1 signaling in fibroblasts mediates myofibroblast differentiation and survival with enhanced ECM synthesis (2). ECM deposition and remodeling results in matrix stiffening, which activates GTPase RhoA and its effector Rho kinase via induction of MKL-1, perpetuating myofibroblast activation (3). Extradomain A (ED-A) fibronectin is essential for myofibroblastic differentiation, whereas HAS2 and CD44 are essential for fibroblast migration/invasion. Positive feedback loops maintain TGF-β1 activation via ECM-derived signals that include Thy-1 integrin  $\alpha_v\beta_5$ , ED-A fibronectin, and versican; in contrast, decorin may inhibit TGF-β1 activity by direct binding (4). HAS2, hyaluronan synthase 2; MKL-1, megakaryoblastic leukemia factor-1.

corepressor (TG-interacting factor), which inhibits TGF- $\beta$  signaling (52). In addition to reduced synthetic capacity, fibroblasts exposed to cigarette smoke may induce higher proteolytic activity in the extracellular milieu by production/release of proteases. Increased expression of MMP-1 and MMP-2 in fibroblasts exposed to cigarette smoke has been reported (53, 54). Furthermore, low concentrations of cigarette smoke have also been shown to induce fibroblast apoptosis (55), which may favor an emphysema phenotype. Collectively, these studies highlight a divergence of tissue repair responses after lung injury, leading to myofibroblast activation in pulmonary fibrosis, whereas this activity is inhibited in emphysema. These abnormal mesenchymal responses further contribute to the alterations in the biochemical and biomechanical properties of the ECM that are emerging as newly appreciated

contributors to the pathogenesis of these two chronic pulmonary diseases.

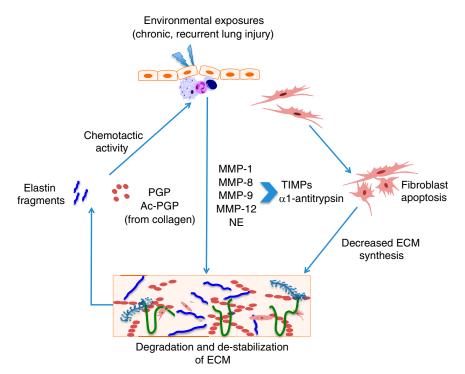
# ECM Composition and Dynamics

The ECM is an interconnected lattice of secreted proteins that forms a scaffold and functions as bidirectional signals for cells within tissues of all adult organs. The ECM is composed of almost 300 proteins forming the core "matrisome," together with glycosoaminoglycans (GAGs), polysaccharides, ECM-binding growth factors, and ECM-modifying enzymes (56, 57). ECM is a highly dynamic structure that is constantly remodeled either enzymatically or nonenzymatically. It confers unique biochemical and mechanical properties to each organ through its interactions with the cellular components such as fibroblasts and epithelial and endothelial cells.

ECM remodeling is a highly regulated process in tissue homeostasis and injury repair (58). In this broad context, ECM remodeling in lung injury repair is defined as dynamic changes in the quantity and quality of ECM components that result in restoration of normal tissue structure/ function or pathological changes such as fibrosis or emphysema. Thus, dysregulated ECM remodeling may result from an imbalance in ECM secretion, degradation, organization, and/or post-translational modifications. The extent to which each of these contribute to fibrosis versus emphysema is not well understood. In addition, whether altered lung mechanics resulting from changes in the quantity or quality of ECM proteins contribute to distinct clinical phenotypes has not been established. In this section, we briefly review the role of fibrous proteins (collagen, elastin), glycoproteins (fibronectin [FN], tenascin), proteoglycans (decorin, versican [VCAN]), and glycosaminoglycans (hyaluronan [HA]) in the development of lung fibrosis and emphysema.

#### Collagens

Collagens are proteins composed of different types of chain-forming helical regions via repeating sequences of three amino acids (Gly-x-y). Fibrillar collagens (types I, II, III, V, and XI) are the most commonly found collagen in the lungs and are critical in maintaining tensile strength and distensibility of the lungs. Type IV collagen, a nonfibrillar collagen, is present in the basement membrane of the lungs and plays an important role at the blood-gas barrier (57). Ultrastructural studies have shown an increased deposition of collagen in ECM surrounding the myofibroblasts in FF in patients with IPF (3). In addition, the expression of lysyl oxidase-like 2, an enzyme that catalyzes cross-linking of fibrillar collagens, is increased in IPF (59). Further investigation is needed to evaluate whether alterations in the mechanical properties of the lung are caused by the increased deposition and/or abnormal structural modifications of collagen. In addition to its structural properties, fibrillar type I collagen limits fibroblast proliferation in normal lung repair by interacting with the cell surface receptor  $\alpha_2\beta_1$  integrin (60). This interaction leads to suppression of the phosphoinositide 3-kinase signal



**Figure 2.** Loss of cellular and ECM homeostasis in the pathogenesis of emphysema. Exposure to inhaled toxins such as cigarette smoke and environmental dust leads to epithelial cell injury and recruitment of inflammatory cells. Inflammatory cells release proteolytic enzymes that include elastases such as NE, macrophage metalloelastase (MMP-12), and collagenases/gelatinases (e.g., MMP-8 and MMP-9). Elastase and MMP-12 cleave elastin to release elastin peptide fragments. The coordinated action of MMP-8 and MMP-9 in the presence of PE results in specific cleavage of collagen to release Pro-Gly-Pro (PGP) and its acetylated form, Ac-PGP. Cleaved elastin fragments and the collagen-derived peptides mediate chemotactic activity for monocytes and neutrophils, respectively, and perpetuate a self-propagating cycle of inflammation and ECM proteolysis. Impaired fibroblast proliferation and survival, with reduced responses to transforming growth factor- $\beta$ 1, down-regulate ECM production. Ac-PGP, N-acetyl Pro-Gly-Pro; MMP, matrix metalloproteinases; NE, neutrophil elastase; PE, propyl endopeptidase; TIMP, tissue inhibitor of metalloproteinase.

pathway caused by maintenance of high phosphatase activity of the tumor suppressor phosphatase and tensin homolog. Loss of this restraint because of a pathologically altered fibroblast-integrin interaction may contribute to their abnormal proliferation in pulmonary fibrosis (60, 61).

In contrast, there is an increase in collagenase activity in emphysema, with weakening of the collagen fiber network and loss of the mechanical forces that appear to be a prerequisite for the development of emphysema (62). In addition to their structural roles, proteolytic products of collagen may have profound effects on repair responses and the perpetuation of inflammation. Neutrophilderived MMP-8 and MMP-9 cleave whole collagen in fragments of 30 to 100 amino acids followed by specific cleavage to release tripeptides, PGP, and its acetylated form, Ac-PGP by the action of propyl endopeptidase (23, 63). The multistep pathway in the degradation of collagen, in the presence of MMPs and propyl endopeptidase, was shown in murine models of cigarette smoke (CS)-induced emphysema, and further *in vitro* studies reported the breakdown of whole collagen to PGP and Ac-PGP (23, 63, 64). These collagen fragments induce neutrophil chemotaxis through CXC-receptors 1 and 2 on neutrophils, thus precipitating an inflammatory cycle (65).

#### Elastin

Elastin, which is the primary component of elastic fibers, is an insoluble polymer of the monomeric soluble precursor, tropoelastin. Elastic fibers are found interspersed together with collagen in the ECM, and they contribute to the elastic recoil properties of the lung (66). Lysyloxidase-catalyzed cross-linking of lysine residues on tropoelastin monomers is essential for elastic fiber assembly. There is a paucity of studies investigating the role of elastin in lung fibrosis. Increased deposition of elastin was demonstrated in a mouse model of bleomycin-induced lung fibrosis (39). Elastin was also shown to enhance TGF-B1-induced myofibroblast differentiation with increased gene expression of  $\alpha$ -SMA and type I collagen in human lung fibroblasts (39). In contrast, the role of elastin in the pathogenesis of emphysema is well established. Elastin and collagen degradation by neutrophil and macrophage proteases results in ECM proteolysis in vitro and emphysema in vivo. There is a protease-antiprotease imbalance, as shown by early studies in patients with  $\alpha$ -1 antitrypsin deficiency, a major inhibitor of elastases (67). In addition, intratracheal administration of elastase induces emphysema, and both macrophage-elastase (MMP-12) and neutrophil-elastase-null mice were protected against CS-induced emphysema (22, 68). However, this protection against CS in MMP-12 and neutrophilelastase-null mice may not be solely caused by a lack of elastase production. Similar to collagen fragments, elastin fragments generated from the elastin polymer by proteolysis have increased chemotactic activity, resulting in macrophage accumulation and further ECM degradation by MMP-12 (24). Elastin fragment antagonism with a monoclonal antibody abrogated this feedback loop of macrophage accumulation and ECM elastin degradation in a mouse emphysema model (24), suggesting a potential target for therapeutic intervention.

#### **Glycoproteins**

FN is a dimeric glycoprotein that mediates cell-matrix adhesions through the binding of ECM proteins such as collagen and cell-surface integrins (69). Partly through its effects on reorganization of the cytoskeleton, FN facilitates cell migration and differentiation during the process of wound healing. Alternative splicing of the type III domains results in polymorphism of FN, particularly extradomain-A (ED-A)-FN, which is

essential for wound healing (70) and fibrogenesis (41, 42). The role of alternative splicing of multiple genes in the regulation of the immune response is better appreciated (71) than its role in regulating wound healing and fibrosis. Studies on the role of alternative splicing involving other ECM genes and their receptors (e.g., CD44) are likely to provide novel insights into the diversity of the wound-healing responses.

Tenascin-C (TNC) is another large hexameric glycoprotein that is up-regulated during tissue injury. ED-A-FN and TNC are present in abnormally larger quantities in fibrotic lung with increased synthesis of these glycoproteins by fibroblasts from patients with IPF compared with fibroblasts from normal subjects (41, 72). They have been shown to colocalize with collagen and VCAN in fibroblastic foci, which are active sites of fibrogenesis (72). Expression of these glycoproteins is stimulated by several soluble factors such as TGF-B1 and PDGF. TGF-B1 preferentially promotes the accumulation of ED-A-FN (73, 74), the predominant isoform in pulmonary fibrosis. ED-A-FN and TNC, in turn, have been shown to be essential for myofibroblast differentiation, cellular motility, and collagen production (41, 42, 75, 76). The mechanism of ED-A-FN-mediated fibroblast differentiation is incompletely understood; the involvement of  $\alpha_4\beta_7$  integrin and the activation of focal adhesion kinase and mitogen-activated protein kinase-associated signaling pathways have been reported (77). ED-A-FN-null mice exposed to bleomycin demonstrate diminished pulmonary fibrosis with fewer myofibroblasts compared with wild-type mice; this correlated with diminished activation of TGF- $\beta$ 1 (41). Similarly, TNC-null mice exposed to bleomycin demonstrate reduced fibrosis from impaired TGF-β1 responsiveness (78). These data suggest that glycoproteins, released from fibroblasts, may play an important role in TGF-B1-mediated myofibroblast differentiation, resulting in a feed-forward loop. Increased peribronchial deposition of collagen,  $\alpha$ -SMA, TNC, and FN in patients with emphysema suggests a similar mechanism of aberrant repair after epithelial injury, with resultant airway remodeling and airflow obstruction in COPD (79, 80). However, our current understanding of the role of specific

glycoproteins in the matrix remodeling associated with emphysema is limited.

#### Proteoglycans

Proteoglycans consist of core proteins to which highly charged GAGs are covalently linked. They are critical constituents of the hydrated gel or "ground substance" in which collagen and elastin fibers are embedded (81). The lung tissue comprises sulfated (chondroitin sulfate, heparan sulfate, and keratin sulfate) and nonsulfated (hyaluronic acid) GAGs (82). The PGs normally contribute to the mechanical stability of the collagen-elastin network and prevent collapse of alveolar structures caused by electrostatic interactions from the negatively charged GAGs (81). Structural remodeling of the lung resulting from abnormal expression of PG-GAG complexes and structural changes to the GAG side chains can contribute to the development of pulmonary diseases (45, 83-88).

VCAN is a chondroitin sulfate/ dermatan sulfate attached PG that is important for matrix assembly and regulation of collagen fibrillation. Increased deposition of VCAN has been demonstrated around active areas of fibrogenesis in the fibrotic lungs of patients with IPF (72). Increased expression of VCAN and enzymes essential for the synthesis and sulfation of associated GAGs was demonstrated in bleomycin-instilled rats and in fibrotic lung fibroblasts (88). Furthermore, this study showed that the up-regulation of VCAN in rat lung fibroblasts was mediated by the TGF-B type I receptor/activin receptor-like kinase 5/p38 mitogen-activated protein kinase signaling pathway. Interestingly, recombinant VCAN-expressing mouse fibroblasts were shown to display a TGF-β1-mediated myofibroblast-like phenotype with increased proliferation and synthetic activity in association with enhanced collagen gel contractility (40). Data exploring the role of VCAN in the pathogenesis of emphysema are limited. Increased deposition of VCAN from fibroblasts in the distal airways and alveolar walls of patients with COPD has been demonstrated previously (89, 90). However, this is negatively correlated with the amount of elastin and elastin-binding protein in COPD, suggesting that modulation of VCAN could influence the

resynthesis of elastin fibers through its activity on elastin-binding protein.

Decorin, biglycan, and fibromodulin are small, leucine-rich PGs with complex roles in the structural and signaling functions of the ECM. In rat models of pulmonary fibrosis, increased levels of biglycan and fibromodulin with reduced levels of decorin have been reported (86). Interestingly, intratracheal delivery of recombinant decorin by adenovirus gene transfer was shown to decrease subpleural fibroproliferation in a bleomycin mouse model (91). Similarly, intranasal administration of a decorin-encoding adenoviral vector resulted in transient increase in decorin that ameliorated bleomycin-induced lung fibrosis in mice (92). The antifibrotic effects of decorin have been attributed to inhibition of TGF-B1 activity through binding of the patent peptide to leucine-rich repeats in the core protein (92, 93). There is no difference in peribronchial or parenchymal deposition of decorin in mild/moderate COPD compared with normal lungs; however, decreased expression of peribronchiolar decorin has been shown in patients with severe emphysema (79, 94). In addition, TGF-B1 has been shown to down-regulate decorin expression in fibroblasts from patients with severe emphysema compared with fibroblasts from patients with mild emphysema (95). Interestingly, it was shown that fluticasone induced decorin production in airway fibroblasts from patients with severe COPD (96). From these studies, it appears that restoration of decorin may result in an improvement in airway obstruction by ameliorating the negative effects of TGF-B1 on airway remodeling.

#### Glycosaminoglycans

HA synthesized by membrane-bound HA synthases is a large, nonsulfated GAG that has been shown to accumulate at sites of tissue injury and play a role in normal wound healing (97, 98). In a mice model of bleomycin-induced lung injury, mice with myofibroblast-specific overexpression of membrane-bound HA synthases (HAS2) were shown to develop severe fibrosis; silencing of the *HAS2* gene in fibroblasts inhibited matrix invasion and myofibroblast accumulation (45). The absence of CD44, an HA cell-surface receptor, or treatment with anti-CD44 antibodies, also reduced lung fibrosis, suggesting that the HAS2-CD44 axis is critical for tissue invasion and relentless fibrosis (45). In addition, HA-Toll-like receptor 2 and HA-Toll-like receptor 4 interactions have been shown to provide signals that initiate inflammatory responses, maintain epithelial cell integrity, and promote recovery from lung injury (99), supporting the concept that HA may function as one of a growing number of damage-associated molecular patterns. Interestingly, the chain length of HA is reduced significantly by cigarette smoke-induced free radical damage, reducing the viscosity of aggregated PGs in emphysema (100). Other studies have shown that low molecular weight HA fragments create a proinflammatory state by stimulating macrophages to release chemokines and MMPs (specifically, MMP-12), resulting in chronic inflammation, destruction of the lung parenchyma, and airway wall remodeling in emphysema (101, 102). In a recent study using a mouse model of porcine pancreatic elastase-induced emphysema, the loss of the structural role of PGs related to the change in charge density on the GAGs reduced alveolar stability and led to a more progressive disease phenotype (85). In that study, the changes in mouse lung tissue stiffness and the alveolar shape distortion under the influence of varying tonicity conditions were increased in experimental emphysema, thus suggesting that the loss of PGs affects lung tissue stability and the progression of emphysema.

# Mechanical Properties of ECM

There is emerging evidence that sustained matrix stiffness from abnormal ECM deposition and contractile activity of myofibroblasts is critically involved in influencing fibroblast function. The interactions between the ECM and fibroblasts were examined recently by culturing fibroblasts from IPF or normal lung on decellularized ECM from IPF or normal lung (37). Interestingly, diseased ECM had a greater impact on pathological gene expression at the level of translation compared with the cell autonomous changes themselves; in addition, IPF ECM promoted the translational activation of genes encoding ECM proteins in fibroblasts, suggesting a positive feedback loop.

Several pathways have been implicated in myofibroblast activation by stiff matrices. A stiff matrix may regulate myofibroblast differentiation through an extrinsic mechanotransduction pathway, leading to latent TGF- $\beta$ 1 activation by the induction of Thy-1 integrin  $\alpha_v\beta_5$  (38). A stiff matrix also activates GTPase RhoA and its effector Rho kinase via induction of megakaryoblastic leukemia factor-1, which has been shown to regulate the expression of fibrotic genes (36). This intrinsic mechanotransduction pathway involving RhoA/ROCK-dependent actin cytoskeletal remodeling and megakaryoblastic leukemia factor-1 activation mediates myofibroblast differentiation in vitro and experimental lung fibrosis in vivo (103). It is not known whether alterations in ECM stiffness and mechanotransduction influence the function and behavior of fibroblasts in emphysema or, reciprocally, whether cell autonomous (epigenetic) changes in fibroblasts influence ECM production and remodeling. Ex vivo cell/tissue culture models that recapitulate the biochemical and biomechanical properties of the ECM are still evolving. Polyacrylamide hydrogel systems with tunable substrate stiffness are well established; however, they fail to reconstruct all the properties of diseased ECM. Development of new tools to simulate three-dimensional models of acellular, normal, and diseased human lung matrices may be vital to understanding the intricate effects of ECM in regulating cellular phenotypes/fates relevant to pulmonary fibrosis and emphysema.

# TGF- $\beta$ 1 in Fibrosis and Emphysema

Although the biochemical and biomechanical properties of ECM contribute to myofibroblast differentiation and survival, the best recognized soluble mediator in myofibroblast activation and tissue fibrosis is TGF-B1. However, the role of TGF- $\beta$ 1 in emphysema development/progression is limited. TGF-β1 is a 75-kD polypeptide synthesized as a large latent precursor molecule bound to the latency-associated peptide by various cells and is incorporated into the ECM (104). Latent TGF-β1 may be activated by both protease-dependent and protease-independent mechanisms (104). Integrin  $\alpha_{v}\beta_{6}$ -mediated activation of

latent TGF-B1 is the most recognized and studied mechanism in pulmonary fibrosis (105). Other mechanisms of TGF-B1 activation involving plasmin, MMPs, thrombospondin, and extracellular acidification have also been described (104), although their relative roles in different modes of lung injury repair are not well defined. In addition, ECM proteins, VCAN, and ED-A-FN can potentially activate latent TGF-B1 (41, 88); whether these are related to mechanosensing in specific cell types warrant further investigation (106). Activated TGF-B1 binds to the TGF- $\beta$  receptor (TGF- $\beta$ R) complex composed of dimers of the transmembrane serine/threonine kinase receptors- type I and II TGF-BRs (107, 108). This leads to transphosphorylation of the kinase domain of TGF-BRI by TGF-βRII, inducing the phosphorylation and activation of the intracellular receptor-regulated R-Smad (Smad2 and Smad3) proteins in fibroblasts (107). The R-Smads then interact with Smad4 and translocate to the nucleus to activate transcription of the genes involved in fibroblast differentiation and matrix synthesis. In addition to its fundamental role in myofibroblast differentiation, TGF-B1 promotes myofibroblast survival through the activation and coordination of two major prosurvival pathways, focal adhesion kinase and protein kinase B (109). These pathways may contribute to the prolonged survival of myofibroblasts in fibrotic tissues.

Some studies in human subjects suggest that, in contrast to the profibrotic effects of TGF-B1 signaling in pulmonary fibrosis, TGF-B1 may play a protective role in emphysema. Among several polymorphisms for TGF-β1, SNP T+869C (leucine to proline exchange caused by a transition of nucleotide T to C at position 869) and T allele of SNP rs1800469, both of which are associated with higher TGF-B1 production, are more prevalent in control smokers than in patients with emphysema (110, 111), suggesting that TGF- $\beta$ 1 may be protective. TGF-B1 normally suppresses the expression of MMPs and increases TIMP expression in regulated repair processes and is important for fibroblastmediated ECM synthesis and contractility (112, 113). In addition, despite adequate levels, it is possible that reduced fibroblast responsiveness to TGF-B1 may influence fibroblast function and ECM maintenance (26, 47, 49). Animal models in which

TGF-B1 signaling has been disrupted support this apparent dichotomy in TGF-B signaling. Mice deficient in the  $\beta 6$ subunit of integrin  $\alpha_v \beta_6$  are unable to activate latent TGF-B1; they are protected from bleomycin-induced lung fibrosis and develop age-dependent emphysema (105, 114). Similarly, mice deficient in Smad3, a TGF-BR-regulated effector protein, are protected from bleomycin and TGF- $\beta$ 1-mediated fibrosis (115), but these mice also develop abnormal alveolarization followed by progressive emphysema and alveolar wall destruction (116). These studies support a role for the TGF-B1/Smad pathway in susceptibility to, and potentially progression of, lung fibrosis and emphysema.

### Conclusions

In summary, despite common risk factors (e.g., cigarette smoke, aging) and

similar fates of susceptible cell populations (e.g., alveolar epithelial cell injury/ apoptosis), the distinct tissue remodeling responses seen in lung fibrosis and emphysema are not well understood. In this review, we posited the idea that differences in mesenchymal cell fate/ function and ECM dynamics may account for phenotypic expression of fibrosis versus emphysema. Given the multitude of functions ascribed to the ECM and their proteolytic products, dysregulation of the inflammatory and/or repair after diverse forms of lung injury may result in distinct clinical syndromes. Alterations to any of the critical steps in the injury-repair response with aberrant cross-talk between reparative cells and the ECM could result in nonresolving, perpetuating cycles of inflammation and repair. Critical pathways involving TGF-B1/Smad, integrins, PGs and their receptors, glycoproteins, and the biomechanical properties of the ECM have been identified as regulating fibroblast function and fate, and new pathways undoubtedly remain to be discovered.

Halting the molecular events propagating feed-forward loops that contribute to disease progression in pulmonary fibrosis and emphysema may be clinically as important as the development of strategies to reverse the disease process. However, the potential off-target effects associated with the disruption of key injury-repair response pathways need to be taken into consideration. As we gain an improved understanding of the role of the lesser-known components of the ECM and decipher the complexities of the cell-matrix interactions in these diseases, the future development of novel matrix-directed therapies are anticipated.

Author disclosures are available with the text of this article at www.atsjournals.org.

#### References

- 1. Thannickal VJ, Henke CA, Horowitz JC, Noble PW, Roman J, Sime PJ, Zhou Y, Wells RG, White ES, Tschumperlin DJ. Matrix biology of idiopathic pulmonary fibrosis: a workshop report of the National Heart, Lung, and Blood Institute. Am J Pathol 2014;184:1643-1651.
- 2. Gross TJ, Hunninghake GW. Idiopathic pulmonary fibrosis. N Engl J Med 2001;345:517-525.
- 3. Kuhn III C, Boldt J, King TE Jr, Crouch E, Vartio T, McDonald JA. An immunohistochemical study of architectural remodeling and connective tissue synthesis in pulmonary fibrosis. Am Rev Respir Dis 1989;140:1693-1703.
- 4. Raghu G, Collard HR, Egan JJ, Martinez FJ, Behr J, Brown KK, Colby TV, Cordier JF, Flaherty KR, Lasky JA, et al.; ATS/ERS/JRS/ALAT Committee on Idiopathic Pulmonary Fibrosis. An official ATS/ERS/ JRS/ALAT statement: idiopathic pulmonary fibrosis: evidence-based guidelines for diagnosis and management. Am J Respir Crit Care Med 2011;183:788-824.
- 5. McDonough JE, Yuan R, Suzuki M, Seyednejad N, Elliott WM, Sanchez PG, Wright AC, Gefter WB, Litzky L, Coxson HO, et al. Small-airway obstruction and emphysema in chronic obstructive pulmonary disease. N Engl J Med 2011;365:1567-1575.
- 6. Taraseviciene-Stewart L, Voelkel NF. Molecular pathogenesis of emphysema. J Clin Invest 2008;118:394-402.
- 7. Awadalla NJ, Hegazy A, Elmetwally RA, Wahby I. Occupational and environmental risk factors for idiopathic pulmonary fibrosis in Egypt: a multicenter case-control study. Int J Occup Environ Med 2012;3: 107-116.
- 8. Taskar V, Coultas D. Exposures and idiopathic lung disease. Semin Respir Crit Care Med 2008;29:670-679.
- 9. Salvi SS, Barnes PJ. Chronic obstructive pulmonary disease in nonsmokers. Lancet 2009;374:733-743.
- 10. Chilosi M, Poletti V, Rossi A. The pathogenesis of COPD and IPF: distinct horns of the same devil? Respir Res 2012;13:3.
- 11. Thannickal VJ, Toews GB, White ES, Lynch III JP, Martinez FJ. Mechanisms of pulmonary fibrosis. Annu Rev Med 2004;55:395-417.
- 12. Antoniades HN, Bravo MA, Avila RE, Galanopoulos T, Neville-Golden J, Maxwell M, Selman M. Platelet-derived growth factor in idiopathic pulmonary fibrosis. J Clin Invest 1990;86:1055-1064.

- 13. Khalil N, O'Connor RN, Unruh HW, Warren PW, Flanders KC, Kemp A, Bereznay OH, Greenberg AH. Increased production and immunohistochemical localization of transforming growth factor-  $\beta$  in idiopathic pulmonary fibrosis. Am J Respir Cell Mol Biol 1991;5: 155-162.
- 14. Pan LH, Yamauchi K, Uzuki M, Nakanishi T, Takigawa M, Inoue H, Sawai T. Type II alveolar epithelial cells and interstitial fibroblasts express connective tissue growth factor in IPF. Eur Respir J 2001;17: 1220-1227.
- 15. Piquet PF, Ribaux C, Karpuz V, Grau GE, Kapanci Y. Expression and localization of tumor necrosis factor- $\!\alpha$  and its mRNA in idiopathic pulmonary fibrosis. Am J Pathol 1993;143:651-655.
- 16. Kolb M, Margetts PJ, Anthony DC, Pitossi F, Gauldie J. Transient expression of IL-1β induces acute lung injury and chronic repair leading to pulmonary fibrosis. J Clin Invest 2001;107: 1529-1536.
- 17. Giannandrea M, Parks WC. Diverse functions of matrix metalloproteinases during fibrosis. Dis Model Mech 2014;7: 193-203.
- 18. Noble PW, Barkauskas CE, Jiang D. Pulmonary fibrosis: patterns and perpetrators. J Clin Invest 2012;122:2756-2762.
- 19. Thannickal VJ, Zhou Y, Gaggar A, Duncan SR. Fibrosis: ultimate and proximate causes. J Clin Invest 2014;124:4673-4677.
- 20. Molet S, Belleguic C, Lena H, Germain N, Bertrand CP, Shapiro SD, Planquois JM, Delaval P, Lagente V. Increase in macrophage elastase (MMP-12) in lungs from patients with chronic obstructive pulmonary disease. Inflamm Res 2005;54:31-36.
- 21. Russell RE, Culpitt SV, DeMatos C, Donnelly L, Smith M, Wiggins J, Barnes PJ. Release and activity of matrix metalloproteinase-9 and tissue inhibitor of metalloproteinase-1 by alveolar macrophages from patients with chronic obstructive pulmonary disease. Am J Respir Cell Mol Biol 2002;26:602-609.
- 22. Shapiro SD, Goldstein NM, Houghton AM, Kobayashi DK, Kelley D, Belaaouaj A. Neutrophil elastase contributes to cigarette smokeinduced emphysema in mice. Am J Pathol 2003;163:2329-2335.
- 23. Gaggar A, Jackson PL, Noerager BD, O'Reilly PJ, McQuaid DB, Rowe SM, Clancy JP, Blalock JE. A novel proteolytic cascade generates an extracellular matrix-derived chemoattractant in chronic neutrophilic inflammation. J Immunol 2008;180:5662-5669.

- 24. Houghton AM, Quintero PA, Perkins DL, Kobayashi DK, Kelley DG, Marconcini LA, Mecham RP, Senior RM, Shapiro SD. Elastin fragments drive disease progression in a murine model of emphysema. J Clin Invest 2006;116:753-759.
- 25. Liu Z, Zhou X, Shapiro SD, Shipley JM, Twining SS, Diaz LA, Senior RM, Werb Z. The serpin  $\alpha$ 1-proteinase inhibitor is a critical substrate for gelatinase B/MMP-9 in vivo. Cell 2000;102:647-655.
- 26. Togo S, Holz O, Liu X, Sugiura H, Kamio K, Wang X, Kawasaki S, Ahn Y, Fredriksson K, Skold CM, et al. Lung fibroblast repair functions in patients with chronic obstructive pulmonary disease are altered by multiple mechanisms. Am J Respir Crit Care Med 2008;178:248-260.
- 27. Hinz B, Phan SH, Thannickal VJ, Prunotto M, Desmoulière A, Varga J, De Wever O, Mareel M, Gabbiani G. Recent developments in myofibroblast biology: paradigms for connective tissue remodeling. Am J Pathol 2012;180:1340-1355.
- 28. Tomasek JJ, Gabbiani G, Hinz B, Chaponnier C, Brown RA. Myofibroblasts and mechano-regulation of connective tissue remodelling. Nat Rev Mol Cell Biol 2002;3:349-363.
- 29. Boström H, Willetts K, Pekny M, Levéen P, Lindahl P, Hedstrand H, Pekna M, Hellström M, Gebre-Medhin S, Schalling M, et al. PDGF-A signaling is a critical event in lung alveolar myofibroblast development and alveogenesis. Cell 1996;85:863-873.

- 30. Hashimoto N, Jin H, Liu T, Chensue SW, Phan SH. Bone marrowderived progenitor cells in pulmonary fibrosis. J Clin Invest 2004;113: 243-252.
- 31. Quan TE, Cowper S, Wu SP, Bockenstedt LK, Bucala R. Circulating fibrocytes: collagen-secreting cells of the peripheral blood. Int J Biochem Cell Biol 2004;36:598-606.
- 32. Willis BC, Liebler JM, Luby-Phelps K, Nicholson AG, Crandall ED, du Bois RM, Borok Z. Induction of epithelial-mesenchymal transition in alveolar epithelial cells by transforming growth factor- $\beta$ 1: potential role in idiopathic pulmonary fibrosis. Am J Pathol 2005;166: 1321-1332.
- 33. Rock JR, Barkauskas CE, Cronce MJ, Xue Y, Harris JR, Liang J, Noble PW, Hogan BL. Multiple stromal populations contribute to pulmonary fibrosis without evidence for epithelial to mesenchymal transition. Proc Natl Acad Sci USA 2011;108: E1475-E1483.
- 34. Desmoulière A, Geinoz A, Gabbiani F, Gabbiani G. Transforming growth factor- $\beta$  1 induces  $\alpha$ -smooth muscle actin expression in granulation tissue myofibroblasts and in quiescent and growing cultured fibroblasts. J Cell Biol 1993;122:103-111.
- 35. Sime PJ, Xing Z, Graham FL, Csaky KG, Gauldie J. Adenovectormediated gene transfer of active transforming growth factor-B1 induces prolonged severe fibrosis in rat lung. J Clin Invest 1997;100: 768-776.
- 36. Huang X, Yang N, Fiore VF, Barker TH, Sun Y, Morris SW, Ding Q, Thannickal VJ, Zhou Y. Matrix stiffness-induced myofibroblast differentiation is mediated by intrinsic mechanotransduction. Am J Respir Cell Mol Biol 2012;47:340-348.
- 37. Parker MW, Rossi D, Peterson M, Smith K, Sikström K, White ES, Connett JE, Henke CA, Larsson O, Bitterman PB. Fibrotic extracellular matrix activates a profibrotic positive feedback loop. J Clin Invest 2014;124:1622-1635.
- 38. Zhou Y, Hagood JS, Lu B, Merryman WD, Murphy-Ullrich JE. Thy-1-integrin  $\alpha v\beta 5$  interactions inhibit lung fibroblast contraction-induced latent transforming growth factor-B1 activation and myofibroblast differentiation. J Biol Chem 2010;285:22382-22393.
- 39. Blaauboer ME, Boeijen FR, Emson CL, Turner SM, Zandieh-Doulabi B, Hanemaaijer R, Smit TH, Stoop R, Everts V. Extracellular matrix proteins: a positive feedback loop in lung fibrosis? Matrix Biol 2014:34:170-178.
- 40. Carthy JM, Meredith AJ, Boroomand S, Abraham T, Luo Z, Knight D, McManus BM, Versican V1 overexpression induces a myofibroblast-like phenotype in cultured fibroblasts. PLoS One 2015;10:e0133056.
- 41. Muro AF, Moretti FA, Moore BB, Yan M, Atrasz RG, Wilke CA, Flaherty KR, Martinez FJ, Tsui JL, Sheppard D, et al. An essential role for fibronectin extra type III domain A in pulmonary fibrosis. Am J Respir Crit Care Med 2008;177:638-645.

- 42. Serini G, Bochaton-Piallat ML, Ropraz P, Geinoz A, Borsi L, Zardi L, Gabbiani G. The fibronectin domain ED-A is crucial for myofibroblastic phenotype induction by transforming growth factorβ1. J Cell Biol 1998;142:873-881.
- 43. Hecker L, Logsdon NJ, Kurundkar D, Kurundkar A, Bernard K, Hock T, Meldrum E, Sanders YY, Thannickal VJ. Reversal of persistent fibrosis in aging by targeting Nox4-Nrf2 redox imbalance. Sci Transl Med 2014;6:231ra47.
- 44. Thannickal VJ, Horowitz JC. Evolving concepts of apoptosis in idiopathic pulmonary fibrosis. Proc Am Thorac Soc 2006;3:350-356.
- 45. Lovgren AK, Kovacs JJ, Xie T, Potts EN, Li Y, Foster WM, Liang J, Meltzer EB, Jiang D, Lefkowitz RJ, et al. β-arrestin deficiency protects against pulmonary fibrosis in mice and prevents fibroblast invasion of extracellular matrix. Sci Transl Med 2011;3:74ra23.
- 46. Castella LF, Buscemi L, Godbout C, Meister JJ, Hinz B. A new lockstep mechanism of matrix remodelling based on subcellular contractile events. J Cell Sci 2010;123:1751-1760.
- 47. Huang S, Wettlaufer SH, Hogaboam C, Aronoff DM, Peters-Golden M. Prostaglandin E(2) inhibits collagen expression and proliferation in patient-derived normal lung fibroblasts via E prostanoid 2 receptor and cAMP signaling. Am J Physiol Lung Cell Mol Physiol 2007;292: 1405-1413.
- 48. Kohyama T, Ertl RF, Valenti V, Spurzem J, Kawamoto M, Nakamura Y, Veys T, Allegra L, Romberger D, Rennard SI. Prostaglandin E(2) inhibits fibroblast chemotaxis. Am J Physiol Lung Cell Mol Physiol 2001:281:L1257-L1263.
- 49. Li YJ, Wang XQ, Sato T, Kanaji N, Nakanishi M, Kim M, Michalski J, Nelson AJ, Sun JH, Farid M, et al. Prostaglandin E<sub>2</sub> inhibits human lung fibroblast chemotaxis through disparate actions on different E-prostanoid receptors. Am J Respir Cell Mol Biol 2011;44:99-107.
- 50. DiCamillo SJ, Carreras I, Panchenko MV, Stone PJ, Nugent MA, Foster JA, Panchenko MP. Elastase-released epidermal growth factor recruits epidermal growth factor receptor and extracellular signalregulated kinases to down-regulate tropoelastin mRNA in lung fibroblasts. J Biol Chem 2002;277:18938-18946.
- 51. DiCamillo SJ, Yang S, Panchenko MV, Toselli PA, Naggar EF, Rich CB, Stone PJ, Nugent MA, Panchenko MP. Neutrophil elastase-initiated EGFR/MEK/ERK signaling counteracts stabilizing effect of autocrine TGF-β on tropoelastin mRNA in lung fibroblasts. Am J Physiol Lung Cell Mol Physiol 2006;291:L232-L243.
- 52. Yang S, Nugent MA, Panchenko MP. EGF antagonizes TGF-β-induced tropoelastin expression in lung fibroblasts via stabilization of Smad corepressor TGIF. Am J Physiol Lung Cell Mol Physiol 2008;295: L143-L151.
- 53. Ning W, Dong Y, Sun J, Li C, Matthay MA, Feghali-Bostwick CA, Choi AM. Cigarette smoke stimulates matrix metalloproteinase-2 activity via EGR-1 in human lung fibroblasts. Am J Respir Cell Mol Biol 2007;36:480-490.
- 54. Kim H, Liu X, Kohyama T, Kobayashi T, Conner H, Abe S, Fang Q, Wen FQ, Rennard SI. Cigarette smoke stimulates MMP-1 production by human lung fibroblasts through the ERK1/2 pathway. COPD 2004;1: 13-23.
- 55. Carnevali S, Petruzzelli S, Longoni B, Vanacore R, Barale R, Cipollini M, Scatena F, Paggiaro P, Celi A, Giuntini C. Cigarette smoke extract induces oxidative stress and apoptosis in human lung fibroblasts. Am J Physiol Lung Cell Mol Physiol 2003;284:L955–L963.
- 56. Hynes RO, Naba A. Overview of the matrisome-an inventory of extracellular matrix constituents and functions. Cold Spring Harb Perspect Biol 2012:4:a004903.
- 57. Dunsmore SE, Rannels DE. Extracellular matrix biology in the lung. Am J Physiol 1996;270:L3-L27.
- 58. Bonnans C, Chou J, Werb Z. Remodelling the extracellular matrix in development and disease. Nat Rev Mol Cell Biol 2014;15:786-801.
- 59. Chien JW, Richards TJ, Gibson KF, Zhang Y, Lindell KO, Shao L, Lyman SK, Adamkewicz JI, Smith V, Kaminski N, et al. Serum lysyl oxidase-like 2 levels and idiopathic pulmonary fibrosis disease progression. Eur Respir J 2014;43:1430-1438.
- 60. Xia H, Seeman J, Hong J, Hergert P, Bodem V, Jessurun J, Smith K, Nho R, Kahm J, Gaillard P, et al. Low  $\alpha(2)\beta(1)$  integrin function enhances the proliferation of fibroblasts from patients with idiopathic pulmonary fibrosis by activation of the β-catenin pathway. Am J Pathol 2012;181:222-233.

- 61. Xia H, Diebold D, Nho R, Perlman D, Kleidon J, Kahm J, Avdulov S, Peterson M, Nerva J, Bitterman P, *et al*. Pathological integrin signaling enhances proliferation of primary lung fibroblasts from patients with idiopathic pulmonary fibrosis. *J Exp Med* 2008;205: 1659–1672.
- 62. Kononov S, Brewer K, Sakai H, Cavalcante FS, Sabayanagam CR, Ingenito EP, Suki B. Roles of mechanical forces and collagen failure in the development of elastase-induced emphysema. *Am J Respir Crit Care Med* 2001;164:1920–1926.
- 63. Braber S, Koelink PJ, Henricks PA, Jackson PL, Nijkamp FP, Garssen J, Kraneveld AD, Blalock JE, Folkerts G. Cigarette smoke-induced lung emphysema in mice is associated with prolyl endopeptidase, an enzyme involved in collagen breakdown. *Am J Physiol Lung Cell Mol Physiol* 2011;300:L255–L265.
- 64. O'Reilly PJ, Hardison MT, Jackson PL, Xu X, Snelgrove RJ, Gaggar A, Galin FS, Blalock JE. Neutrophils contain prolyl endopeptidase and generate the chemotactic peptide, PGP, from collagen. *J Neuroimmunol* 2009;217:51–54.
- 65. Weathington NM, van Houwelingen AH, Noerager BD, Jackson PL, Kraneveld AD, Galin FS, Folkerts G, Nijkamp FP, Blalock JE. A novel peptide CXCR ligand derived from extracellular matrix degradation during airway inflammation. *Nat Med* 2006;12:317–323.
- Shifren A, Mecham RP. The stumbling block in lung repair of emphysema: elastic fiber assembly. *Proc Am Thorac Soc* 2006;3: 428–433.
- Owen CA. Proteinases and oxidants as targets in the treatment of chronic obstructive pulmonary disease. *Proc Am Thorac Soc* 2005; 2: 373–385.
- Hautamaki RD, Kobayashi DK, Senior RM, Shapiro SD. Requirement for macrophage elastase for cigarette smoke-induced emphysema in mice. *Science* 1997;277:2002–2004.
- 69. Pankov R, Yamada KM. Fibronectin at a glance. *J Cell Sci* 2002;115: 3861–3863.
- Muro AF, Chauhan AK, Gajovic S, Iaconcig A, Porro F, Stanta G, Baralle FE. Regulated splicing of the fibronectin EDA exon is essential for proper skin wound healing and normal lifespan. *J Cell Biol* 2003;162: 149–160.
- 71. Martinez NM, Lynch KW. Control of alternative splicing in immune responses: many regulators, many predictions, much still to learn. *Immunol Rev* 2013;253:216–236.
- 72. Estany S, Vicens-Zygmunt V, Llatjós R, Montes A, Penín R, Escobar I, Xaubet A, Santos S, Manresa F, Dorca J, et al. Lung fibrotic tenascin-C upregulation is associated with other extracellular matrix proteins and induced by TGFβ1. BMC Pulm Med 2014;14:120.
- 73. Borsi L, Castellani P, Risso AM, Leprini A, Zardi L. Transforming growth factor-β regulates the splicing pattern of fibronectin messenger RNA precursor. *FEBS Lett* 1990;261:175–178.
- 74. Balza E, Borsi L, Allemanni G, Zardi L. Transforming growth factor β regulates the levels of different fibronectin isoforms in normal human cultured fibroblasts. *FEBS Lett* 1988;228:42–44.
- Olsen AL, Sackey BK, Marcinkiewicz C, Boettiger D, Wells RG. Fibronectin extra domain-A promotes hepatic stellate cell motility but not differentiation into myofibroblasts. *Gastroenterology* 2012; 142:928–937.
- Trebaul A, Chan EK, Midwood KS. Regulation of fibroblast migration by tenascin-C. *Biochem Soc Trans* 2007;35:695–697.
- 77. Kohan M, Muro AF, White ES, Berkman N. EDA-containing cellular fibronectin induces fibroblast differentiation through binding to α4β7 integrin receptor and MAPK/Erk 1/2-dependent signaling. *FASEB J* 2010;24:4503–4512.
- Carey WA, Taylor GD, Dean WB, Bristow JD. Tenascin-C deficiency attenuates TGF-B-mediated fibrosis following murine lung injury. *Am J Physiol Lung Cell Mol Physiol* 2010;299:L785–L793.
- 79. Annoni R, Lanças T, Yukimatsu Tanigawa R, de Medeiros Matsushita M, de Morais Fernezlian S, Bruno A, Fernando Ferraz da Silva L, Roughley PJ, Battaglia S, Dolhnikoff M, *et al*. Extracellular matrix composition in COPD. *Eur Respir J* 2012;40:1362–1373.
- Karvonen HM, Lehtonen ST, Harju T, Sormunen RT, Lappi-Blanco E, Mäkinen JM, Laitakari K, Johnson S, Kaarteenaho RL. Myofibroblast expression in airways and alveoli is affected by smoking and COPD. *Respir Res* 2013;14:84.

- Schaefer L, Schaefer RM. Proteoglycans: from structural compounds to signaling molecules. *Cell Tissue Res* 2010;339:237–246.
- Bensadoun ES, Burke AK, Hogg JC, Roberts CR. Proteoglycan deposition in pulmonary fibrosis. *Am J Respir Crit Care Med* 1996;154:1819–1828.
- 84. Bracke KR, Dentener MA, Papakonstantinou E, Vernooy JH, Demoor T, Pauwels NS, Cleutjens J, van Suylen RJ, Joos GF, Brusselle GG, *et al.* Enhanced deposition of low-molecular-weight hyaluronan in lungs of cigarette smoke-exposed mice. *Am J Respir Cell Mol Biol* 2010;42:753–761.
- Takahashi A, Majumdar A, Parameswaran H, Bartolák-Suki E, Suki B. Proteoglycans maintain lung stability in an elastase-treated mouse model of emphysema. *Am J Respir Cell Mol Biol* 2014;51:26–33.
- Venkatesan N, Ebihara T, Roughley PJ, Ludwig MS. Alterations in large and small proteoglycans in bleomycin-induced pulmonary fibrosis in rats. *Am J Respir Crit Care Med* 2000;161:2066–2073.
- Venkatesan N, Ouzzine M, Kolb M, Netter P, Ludwig MS. Increased deposition of chondroitin/dermatan sulfate glycosaminoglycan and upregulation of β1,3-glucuronosyltransferase I in pulmonary fibrosis. *Am J Physiol Lung Cell Mol Physiol* 2011;300:L191–L203.
- 88. Venkatesan N, Tsuchiya K, Kolb M, Farkas L, Bourhim M, Ouzzine M, Ludwig MS. Glycosyltransferases and glycosaminoglycans in bleomycin and transforming growth factor-β1-induced pulmonary fibrosis. *Am J Respir Cell Mol Biol* 2014;50:583–594.
- Hallgren O, Nihlberg K, Dahlbäck M, Bjermer L, Eriksson LT, Erjefält JS, Löfdahl CG, Westergren-Thorsson G. Altered fibroblast proteoglycan production in COPD. *Respir Res* 2010;11:55.
- Merrilees MJ, Ching PS, Beaumont B, Hinek A, Wight TN, Black PN. Changes in elastin, elastin binding protein and versican in alveoli in chronic obstructive pulmonary disease. *Respir Res* 2008;9:41.
- Shimizukawa M, Ebina M, Narumi K, Kikuchi T, Munakata H, Nukiwa T. Intratracheal gene transfer of decorin reduces subpleural fibroproliferation induced by bleomycin. *Am J Physiol Lung Cell Mol Physiol* 2003;284:L526–L532.
- 92. Kolb M, Margetts PJ, Sime PJ, Gauldie J. Proteoglycans decorin and biglycan differentially modulate TGF-β-mediated fibrotic responses in the lung. *Am J Physiol Lung Cell Mol Physiol* 2001;280: L1327–L1334.
- Yamaguchi Y, Mann DM, Ruoslahti E. Negative regulation of transforming growth factor-β by the proteoglycan decorin. *Nature* 1990;346:281–284.
- 94. van Straaten JF, Coers W, Noordhoek JA, Huitema S, Flipsen JT, Kauffman HF, Timens W, Postma DS. Proteoglycan changes in the extracellular matrix of lung tissue from patients with pulmonary emphysema. *Mod Pathol* 1999;12:697–705.
- Noordhoek JA, Postma DS, Chong LL, Menkema L, Kauffman HF, Timens W, van Straaten JF, van der Geld YM. Different modulation of decorin production by lung fibroblasts from patients with mild and severe emphysema. *COPD* 2005;2:17–25.
- 96. Brandsma CA, Timens W, Jonker MR, Rutgers B, Noordhoek JA, Postma DS. Differential effects of fluticasone on extracellular matrix production by airway and parenchymal fibroblasts in severe COPD. Am J Physiol Lung Cell Mol Physiol 2013;305: L582–L589.
- 97. Turino GM, Cantor JO. Hyaluronan in respiratory injury and repair. *Am J Respir Crit Care Med* 2003;167:1169–1175.
- 98. Jiang D, Liang J, Noble PW. Hyaluronan in tissue injury and repair. Annu Rev Cell Dev Biol 2007;23:435–461.
- 99. Jiang D, Liang J, Fan J, Yu S, Chen S, Luo Y, Prestwich GD, Mascarenhas MM, Garg HG, Quinn DA, et al. Regulation of lung injury and repair by Toll-like receptors and hyaluronan. Nat Med 2005;11:1173–1179.
- 100. McDevitt CA, Beck GJ, Ciunga MJ, O'Brien J. Cigarette smoke degrades hyaluronic acid. *Lung* 1989;167:237–245.
- Horton MR, Shapiro S, Bao C, Lowenstein CJ, Noble PW. Induction and regulation of macrophage metalloelastase by hyaluronan fragments in mouse macrophages. *J Immunol* 1999;162:4171–4176.

- 102. McKee CM, Penno MB, Cowman M, Burdick MD, Strieter RM, Bao C, Noble PW. Hyaluronan (HA) fragments induce chemokine gene expression in alveolar macrophages. The role of HA size and CD44. *J Clin Invest* 1996;98:2403–2413.
- 103. Zhou Y, Huang X, Hecker L, Kurundkar D, Kurundkar A, Liu H, Jin TH, Desai L, Bernard K, Thannickal VJ. Inhibition of mechanosensitive signaling in myofibroblasts ameliorates experimental pulmonary fibrosis. J Clin Invest 2013;123:1096–1108.
- 104. Annes JP, Munger JS, Rifkin DB. Making sense of latent TGFβ activation. *J Cell Sci* 2003;116:217–224.
- 105. Munger JS, Huang X, Kawakatsu H, Griffiths MJ, Dalton SL, Wu J, Pittet JF, Kaminski N, Garat C, Matthay MA, *et al.* The integrin  $\alpha_{v}\beta_{6}$  binds and activates latent TGF- $\beta$ 1: a mechanism for regulating pulmonary inflammation and fibrosis. *Cell* 1999;96: 319–328.
- Wipff PJ, Rifkin DB, Meister JJ, Hinz B. Myofibroblast contraction activates latent TGF-β1 from the extracellular matrix. *J Cell Biol* 2007;179:1311–1323.
- 107. Massagué J. TGFβ signalling in context. Nat Rev Mol Cell Biol 2012;13:616–630.
- 108. Shi Y, Massagué J. Mechanisms of TGF-β signaling from cell membrane to the nucleus. *Cell* 2003;113:685–700.
- 109. Horowitz JC, Rogers DS, Sharma V, Vittal R, White ES, Cui Z, Thannickal VJ. Combinatorial activation of FAK and AKT by transforming growth factor-β1 confers an anoikis-resistant phenotype to myofibroblasts. *Cell Signal* 2007;19:761–771.

- 110. Wu L, Chau J, Young RP, Pokorny V, Mills GD, Hopkins R, McLean L, Black PN. Transforming growth factor-β1 genotype and susceptibility to chronic obstructive pulmonary disease. *Thorax* 2004;59:126–129.
- 111. Zhang L, Chang WW, Ding H, Su H, Wang HY. Transforming growth factor-β1 polymorphisms and chronic obstructive pulmonary disease: a meta-analysis. *Int J Tuberc Lung Dis* 2011;15: 1301–1307.
- Kessenbrock K, Plaks V, Werb Z. Matrix metalloproteinases: regulators of the tumor microenvironment. Cell 2010;141:52–67.
- 113. Kuang PP, Zhang XH, Rich CB, Foster JA, Subramanian M, Goldstein RH. Activation of elastin transcription by transforming growth factor-β in human lung fibroblasts. *Am J Physiol Lung Cell Mol Physiol* 2007;292:L944–L952.
- 114. Morris DG, Huang X, Kaminski N, Wang Y, Shapiro SD, Dolganov G, Glick A, Sheppard D. Loss of integrin  $\alpha_{\nu}\beta_{6}$ -mediated TGF- $\beta$  activation causes Mmp12-dependent emphysema. *Nature* 2003;422:169–173.
- 115. Zhao J, Shi W, Wang YL, Chen H, Bringas P Jr, Datto MB, Frederick JP, Wang XF, Warburton D. Smad3 deficiency attenuates bleomycin-induced pulmonary fibrosis in mice. *Am J Physiol Lung Cell Mol Physiol* 2002;282:L585–L593.
- 116. Bonniaud P, Kolb M, Galt T, Robertson J, Robbins C, Stampfli M, Lavery C, Margetts PJ, Roberts AB, Gauldie J. Smad3 null mice develop airspace enlargement and are resistant to TGF-β-mediated pulmonary fibrosis. *J Immunol* 2004;173:2099–2108.