




OPEN

Mechanism and consequences for avoidance of superparasitism in the solitary parasitoid *Cotesia vestalis*


Wen-bin Chen^{1,2}, Liette Vasseur^{1,3}, Shuai-qi Zhang^{1,2}, Han-fang Zhang^{1,2}, Jun Mao^{1,2}, Tian-sheng Liu^{1,2}, Xian-yong Zhou^{1,2}, Xin Wang^{1,2}, Jing Zhang^{1,2}, Min-sheng You^{1,2}  & Geoff M. Gurr^{1,4} 

A parasitoid's decision to reject or accept a potential host is fundamental to its fitness. Superparasitism, in which more than one egg of a given parasitoid species can deposit in a single host, is usually considered sub-optimal in systems where the host is able to support the development of only a single parasitoid. It follows that selection pressure may drive the capacity for parasitoids to recognize parasitized hosts, especially if there is a fitness cost of superparasitism. Here, we used microsatellite studies of two distinct populations of *Cotesia vestalis* to demonstrate that an egg laid into a diamondback moth (*Plutella xylostella*) larva that was parasitized by a conspecific parasitoid 10 min, 2 or 6 h previously was as likely to develop and emerge successfully as was the first-laid egg. Consistent with this, a naive parasitoid encountering its first host was equally likely to accept a healthy larva as one parasitized 10 min prior, though handling time of parasitized hosts was extended. For second and third host encounters, parasitized hosts were less readily accepted than healthy larvae. If 12 h elapsed between parasitism events, the second-laid egg was much less likely to develop. Discrimination between parasitized and healthy hosts was evident when females were allowed physical contact with hosts, and healthy hosts were rendered less acceptable by manual injection of parasitoid venom into their hemolymph. Collectively, these results show a limited capacity to discriminate parasitized from healthy larvae despite a viability cost associated with failing to avoid superparasitism.

Host selection and discrimination are essential behaviors of parasitoids¹. Encountering a parasitized host may evoke different responses between gregarious and solitary parasitoid species². In gregarious parasitoids, multiple eggs from one or more females can be laid in the host and complete development. In contrast, solitary parasitoids usually lay only one egg per host that reaches adulthood³. These parasitoids are generally predicted to avoid already parasitized (superparasitized) hosts because the earliest laid parasitoid larva may have to compete with or directly kill a subsequently laid newest competitor^{4,5}.

Superparasitism is a common phenomenon in solitary parasitoids⁶. For a long time, most researchers believed that laying more than one egg in a host was sub-optimal and regarded as a mistake in the case of solitary parasitoids^{6,7}. Superparasitism can, however, be adaptive under some conditions^{6,7}. In parasitoid *Microplitis rufiventris* (Hymenoptera: Braconidae), superparasitism increases the success rate of emergence of wasp larvae when using a suboptimal host (advanced host)⁸. In the parasitoid *Spalangia cameroni* (Hymenoptera: Pteromalidae), when hosts are scarce, superparasitism is observed and offspring production per parasitized host increases⁹. Superparasitism can help larvae of the parasitoid *Pseudapanteles dignus* evade host encapsulation¹⁰. Parasitoids are thought to use encounters with unparasitized hosts as a source of information to evaluate host density in the habitat¹¹. Clearly, such responses can only happen when the parasitoid has the capacity to discriminate parasitized from unparasitized hosts, a phenomenon described as host discrimination¹².

In parasitoid wasps, external or internal chemical cues are thought to play key role in host discrimination^{13–16}. These cues, also referred as host marking pheromones (HMP), can be detected by conspecific's antennae in

¹State Key Laboratory of Ecological Pest Control for Fujian and Taiwan Crops, Institute of Applied Ecology, Fujian Agriculture and Forestry University, Fuzhou, China. ²Joint International Research Laboratory of Ecological Pest Control, Ministry of Education, Fuzhou, China. ³Department of Biological Sciences, Brock University, St. Catharines, Ontario, Canada. ⁴Graham Centre, Charles Sturt University, Orange, New South Wales, Australia.  e-mail: msyou@fafu.edu.cn; GGurr@csu.edu.au

the case of external cues¹⁷ or by ovipositors to detect internal cues^{13,18}. External cues have been described in multiple parasitoid species^{19–21} such as *Anaphes victus* (Hymenoptera: Mymaridae), allowing other females to reject previously-parasitized hosts without antennal probing²². In contrast, the parasitoid *Leptopilina heterotoma* (Hymenoptera: Eucoilidae) larva employs internal cues to discriminate hosts after ovipositor insertion²³. Even though the cues can be used for host discrimination with some variation, it is believed that hemolymph composition, may be one of the most important cues for parasitized host discrimination²⁴.

Cotesia vestalis (Hymenoptera: Braconidae) is an internal and solitary parasitoid that can successfully parasitize 2nd to 4th larval instars of its main host, the globally significant brassica pest, diamondback moth, *Plutella xylostella* (Lepidoptera: Plutellidae) although it prefers 2nd and 3rd instars^{25–27}. *Cotesia vestalis* is a typical hymenopteran parasitoid in that unfertilized eggs develop into males and fertilized eggs develop into females²⁸. The eggs of *C. vestalis* hatch about 36 h to 48 h after oviposition^{29,30}, and the duration of egg and larval stage is about 6.8 days²⁹. The longevity of *C. vestalis* adult averages 10–15 days with food supplied, and the expected maximum parasitization rate of females is about 30 hosts per day³¹. *C. vestalis* is known to use chemical cues such as the plant volatiles to locate the host habits^{32,33}, and the antenna and ovipositor play important roles in host recognition³².

Superparasitism is common in *C. vestalis*. When this occurs, the larvae use their mandibles to attack other parasitoid larvae present in the host and the first-hatched generally considered to have the best opportunity for success³⁰. There is little evidence showing that a larva emerging from a second egg can successfully develop when superparasitism occurs in *C. vestalis*, suggesting a strong selective advantage of avoiding superparasitism. This parasitoid is used worldwide as a biological control agent of *P. xylostella*, so it is of practical as well as scientific interests to establish the significance of superparasitism. Accordingly, we investigated the effects of superparasitism and the capacity of parasitoid females to discriminate between healthy and parasitized hosts. We hypothesized that there would be a measurable fitness cost to *C. vestalis* of superparasitism and this would drive selection for at least some capacity to recognize superparasitized host larvae based on female HMPs.

Materials and Methods

Insects. Both *P. xylostella* and *C. vestalis* were maintained and tested in environmental chambers at 25 °C ± 2 °C, 60% ± 10% RH, and a photoperiod of L14:D10. Founders of *P. xylostella* and a Fuzhou strain (FZ strain) of *C. vestalis* cultures were sourced from pupae/cocoons collected from a cabbage field in Nantong Town, Fuzhou, China (25.95°N, 119.27°E), in May 2014. A second strain of *C. vestalis* was obtained from Zhejiang University in Hangzhou (Zhejiang population, ZJ strain) (30.27°N, 120.15°E) that originated from that more northerly part of Eastern China. Both strains were reared in laboratory for more than two years; the earlier observations showed that they had similar development, parasitism rate, and body size. Both strains were used in experiment 1.1. For all other experiments only the FZ strain was used. The *P. xylostella* population was reared on radish sprouts and the *C. vestalis* population was reared on larvae of *P. xylostella*, both for more than 20 generations before being used for experiments.

Mating and parasitism procedures. Newly emerged *C. vestalis* females were placed individually in plastic tubes (diameter: 8 cm, length: 11 cm) each containing a newly emerged male. A drop of honey and cotton ball soaked with water was placed on the top of the tube for parasitoid feeding and left for 24 h. Preliminary studies had established that after 24 h under these conditions, over 95% of females produced female progeny, indicating they had effectively mated. All the females used for parasitism and superparasitism experiments were previously mated. After 24 h, the female was transferred into a glass tube (diameter: 13 mm, length: 55 mm) with a drop of honey on the side. The *P. xylostella* larvae were individually introduced into the glass tube for parasitism. Earlier work showed that >93% of ovipositor insertions lead to oviposition by *C. vestalis* (refer to Chen *et al.*³⁴).

Survival of superparasitized eggs. To test whether a second-laid egg could develop into an adult, a study was conducted using both the Zhejiang (ZJ) and the Fuzhou (FZ) strains. A population of 18 ZJ and another population of 22 FZ mated females were used as maternal generation. A third instar *P. xylostella* larva was introduced in a glass tube with one female *C. vestalis* ZJ-strain (selected randomly from ZJ population) for oviposition. After 10 minutes, the ZJ-strain female was removed and replaced with a female *C. vestalis* of FZ-strain (selected randomly from ZJ population) to obtain conspecific-superparasitism. The same procedure was repeated with intervals between parasitism and superparasitism of 2, 6 and 12 h (each interval time was used 40 larvae). All superparasitized larvae from the same interval treatment were reared together in a plastic chamber (with a mesh roof, diameter: 15 cm, height: 8 cm) until pupation or death. The sex of emerged *C. vestalis* offspring was identified and recorded, the offspring were then individually placed in Eppendorf tube and stored at -80 °C to be used for DNA extraction. All the mothers and their offspring from the different time intervals were genetically assessed using seven microsatellites³⁵. GENALEX v6.5^{36,37} was used to determine whether the offspring belonged to the ZJ or FZ strain (see supplementary material for detail). Since only one adult can emerge from parasitism from a solitary parasitoid, it was predicted that ≤40 offspring (ZJ + FZ) would emerge. The proportions of emergence of ZJ or FZ strain offspring in the same time interval treatment were compared using Chi-Square test.

Effect of superparasitism on progeny production and bionomics. In order to test whether superparasitism affected progeny when the interval between parasitism and superparasitism was short, *P. xylostella* larvae were exposed to three types of parasitism (parasitism, self-superparasitism, conspecific-superparasitism), using FZ strain females. For the parasitism treatment, 3rd-instar *P. xylostella* larvae were individually exposed to only one parasitoid with one oviposition event allowed then reared individually until the emergence of the parasitoid.

For the self-superparasitism, a 3rd-instar *P. xylostella* larva was first exposed to a female parasitoid for oviposition, and then kept with the same parasitoid for a second oviposition event (interval within 10 min). The larva was then transferred into a Petri dish to be reared until emergence of the parasitoid. For the

conspicuous-superparasitism, a 3rd-instar *P. xylostella* larva was first exposed to a female parasitoid that oviposited. This parasitoid was then removed and replaced with another naive female parasitoid to oviposit (interval within 10 min). For each treatment, the procedure was repeated with 60 larvae. Once completed, larvae were transferred into a Petri dish until emergence. The duration of each host larval stage until the emergence of the parasitoid cocoon was recorded, as well as the weight of the cocoon (within 24 h of pupation). The cocoons were then left to emerge. The emerged parasitoids were reared individually in a Petri dish with 20% honey solution. After parasitoid death, the lengths of the left hind tibia and the left wing were measured under a calibrated stereomicroscope (Olympus-SZX2) with the software “cellSens dimension” (Olympus Ltd, Tokyo, Japan) to quantify body size. Life history traits of the progeny from three treatments were compared using a General Linear Model (GLM) with treatment being the main factor.

Discrimination between parasitized and unparasitized larvae. To obtain parasitized *P. xylostella* larvae, FZ strain *C. vestalis* mated females (20 replicates) were used for parasitism, and the larvae were used for the following two experiments (1.3.1 and 1.3.2).

Comparing rate of oviposition of *C. vestalis* on unparasitized and parasitized *P. xylostella* larvae Oviposition rate of *C. vestalis* was examined when the parasitoid was exposed to 1) larvae that were not parasitized (n = 86), 2) larvae parasitized 10 min prior to the start of the experiment (n = 67), and 3) larvae parasitized 12 h prior to the start of the experiment (n = 52). To better understand whether repeated encounters of such larvae could influence the rate of oviposition, the same female parasitoid was presented to three different larvae over a period of three minutes. In the first minute, a *P. xylostella* larva from one of the three treatments was introduced into a tube (diameter: 13 mm, length: 55 mm). Subsequent acceptance (i.e., insertion of the ovipositor) or rejection of the larva within the following 20 sec was recorded computer-based video recording system installed in an animal behavior observing chamber, as described by Chen *et al.*³⁴. After 30 sec, the larva was removed. In the second minute, a new *P. xylostella* larva from the same treatment was introduced to the same parasitoid. The oviposition or rejection was again recorded, and the second larva was removed following the same procedure than in the first minute. In the third minute, a third larva (same treatment) was introduced, following the same procedure. Data were analyzed using a Generalized Linear Mixed Model (GLMM) and binary logistic regression as target distribution and relationship (link) with the linear model, with “acceptance (0)” as the reference value. In this, treatment served as main factor and the sequence was the second linked variable. Differences in acceptance rate in each encounter point were compared by Chi-square test. The Bonferroni modification was applied with the correction of *P* values and alpha level. Analyses were conducted using SPSS version 24.

Discrimination by *C. vestalis* females between self-parasitized and conspecific-parasitized *P. xylostella* larvae To determine whether *C. vestalis* females could discriminate between self-parasitized and conspecific-parasitized hosts, the following procedure was followed. Fifty-six female parasitoids were used for self-superparasitism test and 58 for conspecific-superparasitism test. Each female parasitoid was allowed to parasitize three unparasitized 3rd-instar *P. xylostella* larvae in a tube, after which the larvae parasitized by this specific wasp were placed in a glass tube (the tube was marked with the same number as the parasitoid) for the following treatments after a period of rest of 10 min. For the self-superparasitism treatment, the parasitoid that was previously exposed to the three larvae (tube with the same number) was again exposed to these larvae following the same procedure as described in section 1.3.1 i.e., the three larvae were sequentially exposed to the parasitoid over a period of three minutes. For conspecific-superparasitism treatment, the parasitoid was presented to three larvae that were parasitized by a different parasitoid in sequence, using the same procedure as self-superparasitism. The superparasitism decision (acceptance or rejection) was recorded as previously described. Statistical analyses were as described in 1.3.1.

Effect of topical application of hemolymph on parasitism acceptance by naive parasitoids. In order to test whether topical application of hemolymph influenced host acceptance by *C. vestalis*, the hemolymph from parasitized *P. xylostella* larvae was isolated and applied onto the cuticle of unparasitized larvae. Third instar *P. xylostella* larvae that had been parasitized 10–30 minutes previously were pricked with a drawn-glass needle and the hemolymph collected by pressing the larva. Then the hemolymph was immediately placed onto unparasitized *P. xylostella* larvae using an insect dissecting needle. Larvae were then exposed to *C. vestalis* within 10 min of being treated prior to any visual sign of melanisation of the hemolymph. The control consisted of the hemolymph from unparasitized larvae, collected and applied with the same method as parasitized larvae. The acceptance and rejection rates were recorded for 53 inexperienced (mated but not prior oviposition) parasitoid females when exposed to unparasitized larvae spread with unparasitized larval hemolymph, and for 51 inexperienced parasitoid females exposed to unparasitized larvae but spread with parasitized larval hemolymph. The acceptance and rejection rates were recorded using the criteria previously described. A GLMM was used to compare the difference of parasitoid's parasitism decision between larvae applied with unparasitized larval hemolymph or with parasitized larval hemolymph.

Effect of topical application of hemolymph on parasitism acceptance by experienced parasitoids. To determine whether having oviposition experience may influence acceptance or rejection rates when encountering unparasitized larvae with larval hemolymph from unparasitized or parasitized larvae, a similar experiment as in section 1.4 was conducted with parasitoids that oviposited 10 minutes before. Fifty-four experienced parasitoid females were presented to unparasitized larvae covered with unparasitized larval hemolymph, and 52 experienced parasitoid females were exposed to unparasitized larvae covered with parasitized larval hemolymph. The acceptance and rejection rates were recorded using the criteria previously described. A GLMM was used to compare the difference of parasitoid's parasitism decision between larvae applied with unparasitized larval hemolymph or with parasitized larval hemolymph. Differences in acceptance rate in each encounter point were compared by Chi-square test.

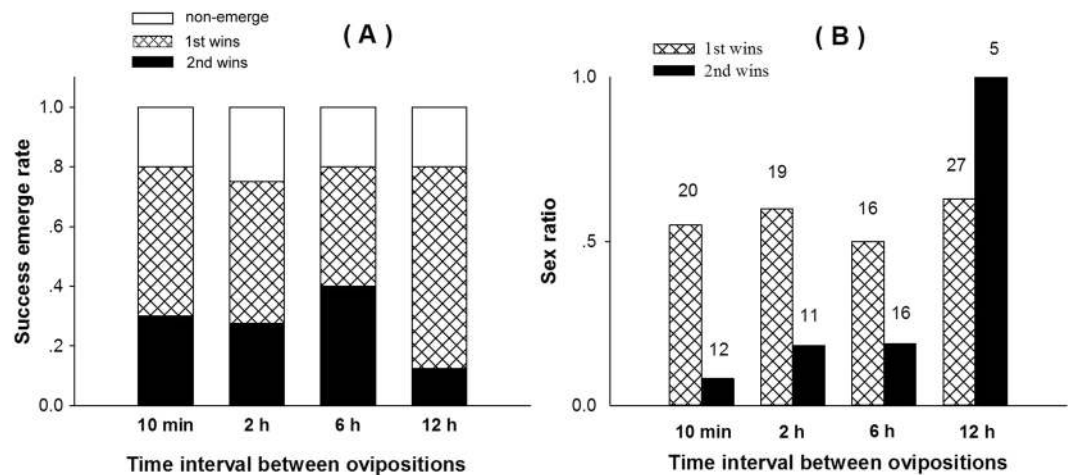


Figure 1. Effect of time intervals between ovipositions on emergence success of the superparasitism eggs. **(A)** The emergence success rate of first and second eggs from 40 oviposition events. **(B)** The sex ratio (proportion of male progeny emerging from contests won by the first and second females) of offspring developed from parasitized (first) eggs or superparasitized (second) eggs. Numbers above bars indicate numbers of individuals in each sample.

Influence of hemolymph injection from parasitized larvae on parasitism acceptance or rejection rate of inexperienced parasitoid. In order to test whether discrimination of the parasitoid between parasitized and unparasitized larvae was affected by non-topical hemolymph-mediated effects, unparasitized host larvae were injected with the hemolymph of either parasitized or unparasitized larvae. In this experiment, we used early stage 4th instar *P. xylostella* larvae for injection and hemolymph collection. For the parasitized treatment, donor larvae were observed to have been stung three times. The hemolymph was then collected as described in the previous experiment but placed in an Eppendorf tube in an ice-bath, immediately centrifuged at 12000 g for 1 min, and the resulting upper-liquid was extracted for the treatment. The extract was injected into unparasitized larvae by a nanoliter injector (NANOLITER 2010 (World Precision Instruments Inc, Florida, USA)) at 70 nl/larva. The hemolymph from unparasitized larvae was used for injection as control treatment. The procedure of three encounters of larvae by *C. vestalis* was used to determine acceptance or rejection as described in 1.3.1. Fifty-two female parasitoids were exposed to larvae injected with parasitized host's hemolymph, and 53 were exposed to larvae injected with unparasitized host's hemolymph. The data of parasitism decisions of the two treatments were compared by GLMMs. Differences in acceptance rate in each encounter point were compared by Chi-square test.

Effect of parasitoid venom on parasitism acceptance/rejection rate of inexperienced parasitoids. As far as we know, *C. vestalis* injects venom and polydnavirus (PDV) into the host during oviposition and, because *C. vestalis* was observed to reject hosts parasitized only minutes previously, we selected venom as the likely marking component in this step of experiment. We tested whether venom was the factor for oviposition decision of a new parasitoid female, venom was collected from 10 mated *C. vestalis* females that were younger than 5 days from hatching. Parasitoids were frozen at -50 °C for 3 min then surface-sterilized with 75% ethanol and sterilized-deionized water. The parasitoids were then dissected to collect the intact venom reservoir, which was then ruptured using dissecting forceps in 20 µl ddH₂O on a Petri dish. Venom was collected in an Eppendorf tube and centrifuged at 12000 g for 1 min, the venom reservoir tissue discarded, and the supernatant used for injection at 70 nl/larva. Early stage 4th instar *P. xylostella* larvae were used for injection and be exposed to inexperienced parasitoids (n = 51). For the control treatment (parasitoids n = 52), the larvae were injected with ddH₂O. Parasitism decision procedure and data analysis were as described above.

Results

Effect of time interval on development success of superparasitism eggs. Studies using the two different strains of parasitoids showed that eggs of both strains could develop from superparasitism events, but their survival rates varied depending on the time interval between initial oviposition and superparasitism oviposition. There were no significant differences in the rate of successful parasitoid development when parasitoids were exposed at 10 min ($\chi^2 = 3.333$, df = 1, p = 0.068), 2 h ($\chi^2 = 3.413$, df = 1, p = 0.065) and 6 h ($\chi^2 = 0$, df = 1, p = 1) intervals between first and second events (Fig. 1A, Supplementary Figs. 2–5). However, when parasitoid females (FZ strain) attacked *P. xylostella* larvae parasitized by a primary parasitoid (ZJ strain) 12 h before, emergence rate was significantly lower for the superparasitized strain than the primary strain ($\chi^2 = 25.208$, df = 1, p < 0.001) (Fig. 1A).

The sex ratio of the first egg that developed to adulthood was not influenced by short time intervals of superparasitism. For the interval of 12 h, if the first eggs survives, the sex ratio of parasitized eggs was 0.63 but, if the second eggs survives and develops, it produced only males (superparasitized eggs), although in very low number (n = 5) (Fig. 1B).

Sex of insect Type of treatment	Number of successfully developed individual ^a	Time to cocoon formation (d)	Time to adult emergence from cocoon (d)	Adult duration (d)	Cocoon weight (mg)	Hind tibia length (mm)	Forewing length (mm)
Females							
Parasitism	23	6.1 ± 0.8	4.1 ± 0.7	10.3 ± 5.1	1.78 ± 0.246	0.756 ± 0.050	2.174 ± 0.106
Self-superparasitism	20	6.3 ± 0.7	4.0 ± 0.5	11.4 ± 5.4	1.72 ± 0.202	0.749 ± 0.052	2.150 ± 0.103
Conspecific-superparasitism	21	6.2 ± 0.8	4.1 ± 0.7	9.9 ± 5.1	1.72 ± 0.333	0.755 ± 0.069	2.186 ± 0.110
Males							
Parasitism	14	5.8 ± 0.6	4.1 ± 0.9	9.4 ± 5.3	1.72 ± 0.205	0.744 ± 0.034	2.207 ± 0.086
Self-superparasitism	15	6.1 ± 0.6	3.4 ± 0.7	10.7 ± 4.5	1.68 ± 0.190	0.738 ± 0.040	2.152 ± 0.095
Conspecific-superparasitism	15	5.9 ± 0.7	3.8 ± 0.9	9.1 ± 5.1	1.58 ± 0.296	0.715 ± 0.051	2.101 ± 0.143

Table 1. Life history traits of *C. vestalis* exposed to parasitism, self-superparasitism and conspecific-superparasitism treatments (mean ± SD). ^aIndividuals that successfully reached adulthood from the initial 60 replicates (including males and females).

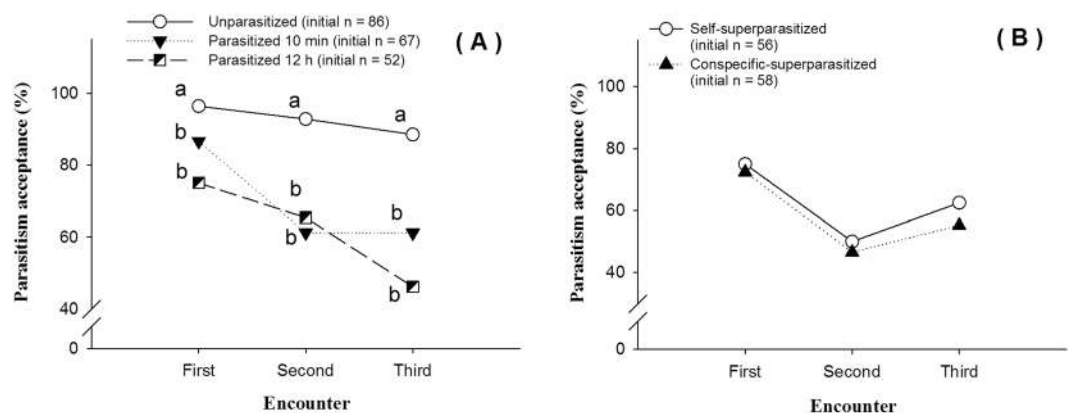


Figure 2. Parasitized host discrimination ability detection of *C. vestalis*. (A) Parasitism acceptance rates of *C. vestalis* to unparasitized or parasitized *P. xylostella* larvae, which had been parasitized 10 minutes or parasitized 12 h prior (GLMMs, Treatment main effect: $F = 34.448$, $df = 2$, $P < 0.001$; Encounter main effect: $F = 11.658$, $df = 2$, $P < 0.001$; Treatment × Encounter: $F = 0.838$, $df = 4$, $P = 0.501$); Different letter in same encounter indicate significant difference ($P < 0.017$), Bonferroni test. (B) Parasitism acceptance rates of *C. vestalis* to self-parasitized and conspecific-parasitized *P. xylostella* larvae (GLMMs, Treatment main effect: $F = 0.035$, $df = 1$, $P = 0.851$, Encounter main effect: $F = 8.925$, $df = 2$, $P < 0.001$; Treatment × Encounter: $F = 0.039$, $df = 2$, $P = 0.961$).

Effect of superparasitism on progeny production and bionomics of *C. vestalis* offspring.

Superparasitism had no adverse effect on progeny production and bionomics in *C. vestalis* (Table 1). Parasitized, self-superparasitized and conspecific-superparasitized offspring showed similar survival rates (Supplementary Table 2). With single parasitism, 62% of emerged individuals were females, and self-superparasitism treatment and conspecific-superparasitism produced 57% and 58%, respectively.

Discrimination between parasitized and unparasitized larvae. Comparing rate of oviposition of *C. vestalis* on unparasitized and parasitized *P. xylostella* larvae. When a *C. vestalis* female encountered a host in the glass tube, she became more active, moving very rapidly. This behavior was similar when encountering a unparasitized or parasitized host (Supplementary Videos 2 and 3). She used her antenna to detect the host and decided to accept or reject the host before ovipositing (Supplementary Video 4). On some occasions, the larva could escape when feeling the antennae, suggesting its capacity to detect the parasitoid (Supplementary Video 1).

Acceptance rate was the highest at the first encounter event when exposed to three larvae sequentially in all treatments (Fig. 2A). The acceptance rate, however, declined significantly over time with the third exposure having the lowest value (Fig. 2A). Acceptance rates were significantly different between treatments ($P < 0.001$), and when the parasitoids were exposed to unparasitized larvae, they exhibited the highest acceptance rates.

Discrimination of *C. vestalis* female between self-parasitized and conspecific-parasitized *P. xylostella* larvae. Female *C. vestalis* did not discriminate between larvae that were self-parasitized and conspecific-parasitized (Fig. 2B).

Effect of topical hemolymph application on parasitism acceptance or rejection rate of inexperienced parasitoid females. Hemolymph volatiles did not affect parasitism acceptance rate of *C. vestalis*. Females showed similar acceptance rates when exposed to unparasitized larvae topically treated with hemolymph

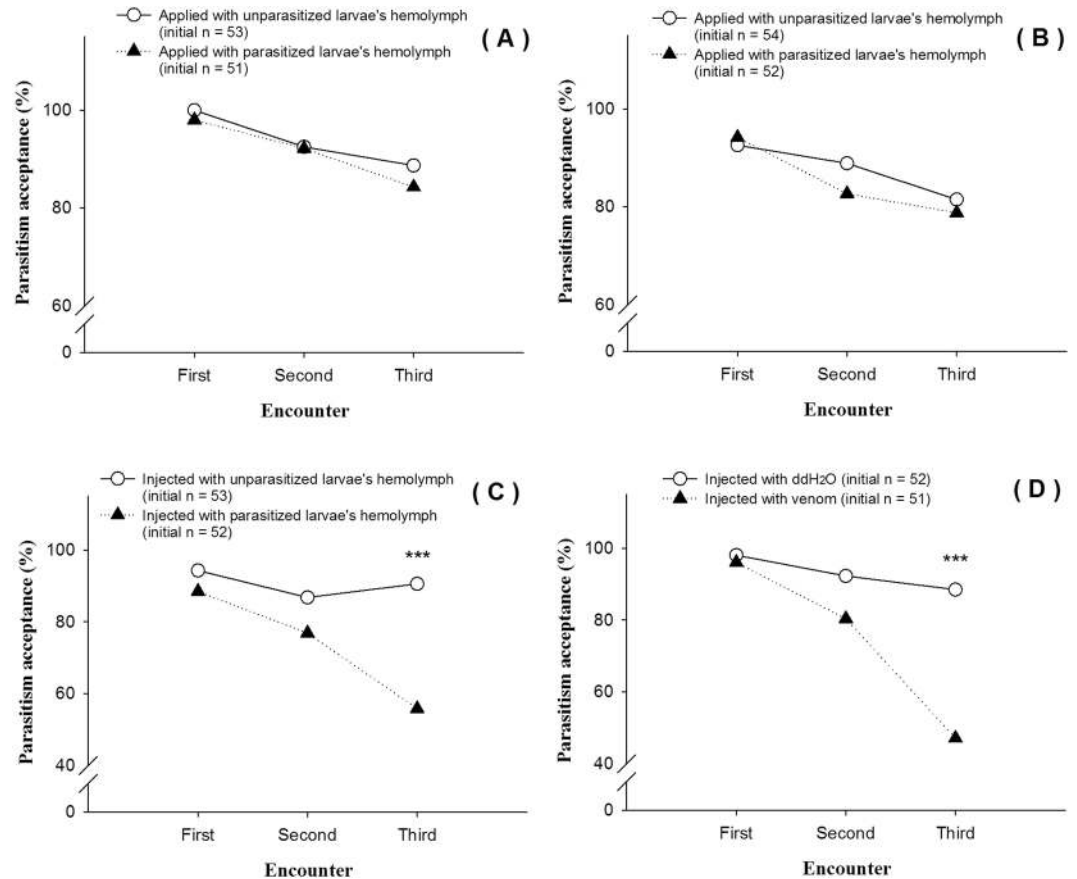


Figure 3. Results of the host discrimination experiments (sections 1.4, 1.5, 1.6 and 1.7). (A) Parasitism acceptance rates of inexperienced *C. vestalis* to unparasitized *P. xylostella* larvae with odor of parasitized larvae's hemolymph (GLMMs, Treatment main effect: $F = 0.258$, $df = 1$, $P = 0.612$; Encounter main effect: $F = 3.612$, $df = 2$, $P = 0.028$; Treatment \times Encounter: $F = 0.095$, $df = 2$, $P = 0.909$). (B) Parasitism acceptance rates of experienced *C. vestalis* to unparasitized *P. xylostella* larvae with odor of parasitized larvae's hemolymph (GLMMs, Treatment main effect: $F = 0.172$, $df = 1$, $P = 0.678$; Encounter main effect: $F = 4.117$, $df = 2$, $P = 0.017$; Treatment \times Encounter: $F = 0.348$, $df = 2$, $P = 0.706$). (C) Parasitism acceptance rates of inexperienced *C. vestalis* to unparasitized *P. xylostella* larvae which injected with parasitized larvae's hemolymph (GLMMs, Treatment main effect: $F = 8.898$, $df = 1$, $P = 0.003$, Encounter main effect: $F = 3.508$, $df = 2$, $P = 0.031$; Treatment \times Encounter: $F = 1.902$, $df = 2$, $P = 0.151$); The Chi-square test for each encounter: First encounter: $\chi^2 = 1.157$, $df = 1$, $P = 0.282$; Second encounter: $\chi^2 = 1.725$, $df = 1$, $P = 0.189$; Third encounter: $\chi^2 = 16.252$, $df = 1$, $P < 0.001$. (D) Parasitism acceptance rates of inexperienced *C. vestalis* to unparasitized *P. xylostella* larvae which injected with parasitoid's venom solution (GLMMs, Treatment main effect: $F = 6.378$, $df = 1$, $P = 0.012$, Encounter main effect: $F = 8.937$, $df = 2$, $P < 0.001$; Treatment \times Encounter: $F = 1.252$, $df = 2$, $P = 0.287$); The Chi-square test for each encounter: First encounter: $\chi^2 = 0.364$, $df = 1$, $P = 0.546$; Second encounter: $\chi^2 = 3.113$, $df = 1$, $P = 0.078$; Third encounter: $\chi^2 = 20.270$, $df = 1$, $P < 0.001$.

of parasitized of *P. xylostella* larvae ($P = 0.612$). Of the initial 53 inexperienced parasitoids exposed to larvae with unparasitized hemolymph, 53 were accepted in the first minute, 49 during the second minute and, 47 in the third minute. Numbers were similar when exposed to parasitized larvae (initial parasitoids = 51, first minute, 50 accepted, second minute, 47 and third minutes 43) (Fig. 3A).

Effect of topical hemolymph application on parasitism acceptance or rejection rate of experienced parasitoid females. Like in the case of inexperienced parasitoids, experienced females showed similar responses to unparasitized larvae applied with hemolymph of parasitized of *P. xylostella* larvae ($P = 0.678$). Hemolymph volatiles did not affect parasitism acceptance rate of experienced *C. vestalis* (Fig. 3B).

Influence of injected hemolymph extract from parasitized larvae on parasitism acceptance or rejection rate of inexperienced parasitoids. The injection of hemolymph from parasitized larvae or unparasitized larvae into unparasitized larvae significantly influenced inexperienced parasitoid host acceptance. Larvae injected with the hemolymph extract from unparasitized larvae had a significantly higher acceptance rate by *C. vestalis* females ($n = 53$) than those treated with hemolymph from parasitized hosts ($P = 0.003$) (Fig. 3C).

While the rate of parasitism acceptance remained relatively constant for the three minutes for unparasitized larvae, there was a significant decline over time for the larvae injected with parasitized larva's hemolymph.

Effect of parasitoid venom on parasitism acceptance/rejection rate of inexperienced parasitoids. When a parasitoid encountered the first larva, the acceptance rate did not significantly differ whether the larva was injected with parasitoid venom or water (Fig. 3D). While acceptance rates remained relatively constant over time for larvae injected with water, parasitoids sequentially exposed to larvae injected with venom showed a decline in acceptance rate over the three minutes (Fig. 3D).

Discussion

In this study, we showed that superparasitism did not necessarily have adverse effect on the parasitoid. Superparasitized hosts could support parasitoid development to adulthood at a similar rate to that of unparasitized hosts. Importantly, however, this was the case only when superparasitism occurred 6 h or less after the initial parasitism event. At 12 h there was a marked cost of superparasitism in terms of reduced emergence and male dominated progeny. Parasitoid foraging behavior and egg production strategies greatly vary among parasitoid species^{38,39}. In solitary parasitoids, superparasitism may be disadvantageous when larvae from eggs laid later must compete with the older conspecifics, and then are less likely to develop to adulthood⁴⁰. Superparasitism may only be considered adaptive if the second eggs maintain similar rates of survival and growth than the first eggs⁴¹ and as shown in our study.

Microsatellites as molecular markers are increasingly used to study genetic diversity in natural pest enemy populations⁴². Based on seven microsatellites, we used two laboratory *C. vestalis* strains (ZJ-strain and FZ-strain) coming from different populations to determine the success rates of first and second eggs to emerge and found that eggs laid by FZ parasitoids could successfully develop to adulthood at a similar emerge rate to primary eggs (laid by ZJ strain) for the first three time intervals (10 min, 2 and 6 h). Superparasitism may have adverse effects on biological traits of parasitoids. For instance, in *Anaphes nitens*, it leads to more male offspring⁴³. In the present study, the sex ratio of the first egg was not influenced by superparasitism, but the sex ratio of the superparasitized egg was obviously changed depending on the time interval between oviposition events. We assume that, when the time interval between first and second oviposition extends to 12 h, few second eggs survive over the first egg. Moreover, due to males developing faster than females, when superparasitism occurs (interval time >12 h), female eggs from second oviposition may have no chance to survive.

Superparasitism may also cause other adverse effects on biological traits of parasitoids. For example, it can cause smaller body size and longer development time in *Cotesia flavipes*⁵. High level of superparasitism (superparasitism more than twice) in *C. vestalis* can lead to longer immature duration and more male offspring²⁹. However, when low level of superparasitism was investigated, we did not detect such adverse effects in life history traits of the progeny including immature duration, adult duration and body size. We hypothesized that the few adverse effects in low level of superparasitism might be an adaptive trait of *C. vestalis* and suggest low level superparasitism in rearing this parasitoid could be acceptable.

Superparasitism was clearly not advantageous for *C. vestalis* when the time interval between oviposition events was 12 h. The emergence success of the second egg was reduced after 12 h and likely due to the first egg having enough time to develop to a sufficiently advanced stage such that this individual was able to compete successfully against the second-laid individual. Our finding is consistent with a study by Zhang *et al.*⁴⁴ that report, through dissection of the superparasitized *P. xylostella* larvae, rates of first egg success of 86.7% after 24 h and 95.0% after 48 h⁴⁴. In general, the probability of a second egg competing and emerging depends on time elapsed since the first parasitism in solitary parasitoids⁴⁵. This similar phenomenon has been observed in other gregarious and solitary parasitoid species, such as *Bracon hebetor*⁴⁶, *Metaphycus flavus*⁴⁷, *Echthrodelphax fairchildii*⁴⁸, and *Trissolcus basalis*⁴⁹.

In the present study, the highest rate of host acceptance was during the first encounter between a parasitoid and a potential host. Inexperienced *C. vestalis* were highly likely to parasitize the first encountered host whether already parasitized or not. This phenomenon may be adaptive, because the naïve parasitoid's encounter with the initial host potentially represents the only host, she will ever encounter given that hosts may be scarce, and she may die before encountering another one. Encounters with subsequent hosts signal the likelihood that she will find others before death so can afford to be progressively more selective and less accepting of previously parasitized host. Host selection is an important factor for parasitoid survival¹. The quality and availability of the host (i.e. resources for egg development) is a key issue for the fitness of parasitoids⁹. In the solitary parasitoid *Diachasmimorpha longicaudata* (Hymenoptera: Braconidae), for example, when the host density fluctuates between high and low per day, high superparasitism rate is associated with the low host density⁵⁰. In *Cotesia flavipes*, the naïve females treat the parasitized host similar to unparasitized host, but the experienced females reject the parasitized host⁵.

Our results showed that when *C. vestalis* was provided successively with more hosts, acceptance rates of parasitized hosts declined. Thus, *C. vestalis* showed some capacity for discrimination between parasitized and unparasitized hosts, as it has been seen for other species⁵¹⁻⁵⁴. For solitary parasitoids, previously parasitized hosts are referred to as low quality, and foraging theory predicts that the parasitoid should reject superparasitism⁴.

Our results strongly suggest that *C. vestalis* can discriminate parasitized *P. xylostella* from unparasitized one and studies with venom and hemolymph treated hosts illustrate the mechanism of host discrimination. Various parasitoids can use pheromones for host discrimination, such as heptanone and alkadienes to avoid already parasitized larvae^{14,21,55} and these are detected by the parasitoid antennae¹⁷. Our videos showed *C. vestalis* using their antennae on host larvae, there was no discrimination between self-parasitized and conspecific-parasitized hosts, which contrasts with *Muscidifurax zaraptor*⁴⁰, *Echthrodelphax fairchildii*⁵⁶, and *Aphelinus asychis*⁵⁷. We propose that *C. vestalis* use internal cues for parasitized host discrimination, because injecting parasitoid host

hemolymph or parasitoid venom into unparasitized hosts altered *C. vestalis* acceptance. This did not occur in response to topically applied hemolymph. This contrasts with other species such as *Anaphes victus* (Hymenoptera: Mymaridae) that can determine host status based on antennal contact²², and similar to *Dolichogenidea tasmanica* (Hymenoptera: Braconidae), which discriminated host status by ovipositor injection⁵⁸.

Irrespective of the potential for *C. vestalis* to determine the parasitism status of a host larvae, this may be overridden by the need to make a very rapid decision to oviposit. *P. xylostella* larvae were able to escape from parasitoids during some encounters even in the enclosed conditions of the present tests. In the field, larvae have the additional option of dropping from the plant to escape danger⁵⁹. This risk to the parasitoid of losing contact with vigorous host larvae that have effective behavioral defenses may have favored selection for rapid acceptance of hosts immediately after encounter, albeit with the tradeoff of a fitness cost should that host already be parasitized.

In conclusion, *C. vestalis* exhibits limited capacity for avoidance of superparasitism, even though it can discriminate between unparasitized and already parasitized hosts, because discrimination requires the insertion of the ovipositor into a vigorously mobile host that is likely to escape unless rapidly utilised. Recognition that a host is already parasitized seems to require first the immediate insertion of the ovipositor and the female parasitoid needs to decide very rapidly whether to deposit an egg. As a new experienced female, she will use her sensory apparatus to apply some level of host discrimination in subsequently-encountered hosts. The cost of this limited discrimination capacity appears to be low if the time interval between parasitism events is low (minutes) but, under longer and biologically realistic time intervals (hours, and presumably days) there is a clear cost of superparasitism. The extent to which this behavior may be adaptive or not for parasitoids under field conditions where hosts have the chance of escaping if not rapidly parasitized, remains to be investigated.

Received: 20 February 2019; Accepted: 12 May 2020;

Published online: 10 July 2020

References

- Vinson, S. B. Host selection by insect parasitoids. *Annual Review of Entomology* **21**, 109–133 (1976).
- Tormos, J. *et al.* Superparasitism in laboratory rearing of *Spalangia cameroni* (Hymenoptera: Pteromalidae), a parasitoid of medfly (Diptera: Tephritidae). *Bulletin of Entomological Research* **102**, 51–61 (2012).
- Reynolds, K. T. & Hardy, I. C. W. Superparasitism: a non-adaptive strategy? *Trends in Ecology & Evolution* **19**, 347–348 (2004).
- Parker, G. A. & Courtney, S. P. Models of clutch size in insect oviposition. *Theoretical Population Biology* **26**, 27–48 (1984).
- Potting, R. P. J., Snellen, H. M. & Vet, L. E. M. Fitness consequences of superparasitism and mechanism of host discrimination in the stemborer parasitoid *Cotesia flavipes*. *Entomologia Experimentalis et Applicata* **82**, 341–348 (1997).
- Van Alphen, J. J. & Visser, M. E. Superparasitism as an adaptive strategy for insect parasitoids. *Annual Review of Entomology* **35**, 59–79 (1990).
- Hubbard, S. F., Harvey, I. F. & Fletcher, J. P. Avoidance of superparasitism: a matter of learning? *Animal Behaviour* **57**, 1193–1197 (1999).
- Khafagi, W. E. & Hegazi, E. M. Does superparasitism improve host suitability for parasitoid development? A case study in the *Microplitis rufiventris* - *Spodoptera littoralis* system. *Biocontrol* **53**, 427–438 (2008).
- Böckmann, E. A., Tormos, J., Beitia, F. & Fischer, K. Offspring production and self-superparasitism in the solitary ectoparasitoid *Spalangia cameroni* (Hymenoptera: Pteromalidae) in relation to host abundance. *Bulletin of Entomological Research* **102**, 131–137 (2012).
- Luna, M. G., Desneux, N. & Schneider, M. I. Encapsulation and self-superparasitism of *Pseudapanteles dignus* (Muesebeck) (Hymenoptera: Braconidae), a parasitoid of *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiidae). *Plos One* **11** (2016).
- Varaldi, J., Fouillet, P., Bouletreau, M. & Fleury, F. Superparasitism acceptance and patch-leaving mechanisms in parasitoids: a comparison between two sympatric wasps. *Animal Behaviour* **69**, 1227–1234 (2005).
- Van Lenteren, J. C. The development of host discrimination and the prevention of superparasitism in the parasite *Pseudeucoila bochei* Weld (Hym.: Cynipidae). *Netherlands Journal of Zoology* **26**, 1–83 (1975).
- Agboka, K. *et al.* Self-, intra-, and interspecific host discrimination in *Telenomus busseolae* Gahan and *Tisis Polaszek* (Hymenoptera: Scelionidae), sympatric egg parasitoids of the African cereal stem borer *Sesamia calamistis* Hampson (Lepidoptera: Noctuidae). *Journal of Insect Behavior* **15**, 1–12 (2002).
- Gauthier, N. & Bénédet, F. Y., T., Monge, J. P. & Huignard, J. Marking behavior and discrimination of concealed hosts by the ectoparasitoid, *Dinarmus basalis* Rond. (Hym. Pteromalidae). *Journal of Insect Behavior* **15**, 589–606 (2002).
- Jaloux, B., Errard, C., Mondy, N., Vannier, F. & Monge, J. P. Sources of chemical signals which enhance multiparasitism preference by a cleptoparasitoid. *Journal of Chemical Ecology* **31**, 1325–1337 (2005).
- Stelinski, L. L., Oakleaf, R. & Rodriguez, C. Oviposition-detering pheromone deposited on blueberry fruit by the parasitic wasp, *Diachasma alloeum*. *Behaviour* **144**, 429–445 (2007).
- Wu, Z. X. & Nordlund, D. A. Superparasitism of *Lygus hesperus* Knight eggs by *Anaphes iole* Girault in the Laboratory. *Biological Control* **23**, 121–126 (2002).
- Ganesalingam, V. K. Mechanism of discrimination between parasitized and unparasitized hosts by *Venturia canescens* (Hymenoptera: Ichneumonidae). *Entomologia Experimentalis et Applicata* **17**, 36–44 (1974).
- Micha, S. G., Stammel, J. & Höller, C. 6-methyl-5-heptene-2-one, a putative sex and spacing pheromone of the aphid hyperparasitoid, *Alloxysta victrix* (Hymenoptera: Alloxystidae). *European Journal of Entomology* **90**, 439–442 (1993).
- Sheehan, W., Wäckers, F. L. & Lewis, W. J. Discrimination of previously searched, host-free sites by *Microplitis croceipes* (Hymenoptera: Braconidae). *Journal of Insect Behavior* **6**, 323–331 (1993).
- Syvrtsen, T. C., Jackson, L. L., Blomquist, G. J. & Vinson, S. B. Alkadienes mediating courtship in the parasitoid *Cardiochiles nigricipes* (Hymenoptera: Braconidae). *Journal of Chemical Ecology* **21**, 1971–1989 (1995).
- Van Baaren, J. & Boivin, G. Learning affects host discrimination behavior in a parasitoid wasp. *Behavioral Ecology & Sociobiology* **42**, 9–16 (1998).
- Ruschioni, S., van Loon, J. J. A., Smid, H. M. & van Lenteren, J. C. Insects can count: Sensory basis of host discrimination in parasitoid wasps revealed. *Plos One* **10** (2015).
- Nufio, C. R. & Papaj, D. R. Host marking behavior in phytophagous insects and parasitoids. *Entomologia Experimentalis et Applicata* **99**, 273–293 (2001).
- Velasco, L. R. I. The life history of *Apanteles plutellae* Kurdj. (Braconidae), a parasitoids of the diamondback moth. *Philippines Entomology* **5**, 385–399 (1982).
- Talekar, N. S. & Yang, J. C. Characteristic of parasitism of diamondback moth by two larval parasites. *Entomophaga* **36**, 95–104 (1991).

27. Shi, Z. H., Liu, S. S. & Li, Y. X. *Cotesia plutellae* parasitizing *Plutella xylostella*: Host-age dependent parasitism and its effect on host development and food consumption. *Biocontrol* **47**, 499–511 (2002).
28. De Boer, J. G., Ode, P. J., Vet, L. E. M., Whitfield, J. & Heimpel, G. E. Complementary sex determination in the parasitoid wasp *Cotesia vestalis* (C. plutellae). *Journal of Evolutionary Biology* **20**, 340–348 (2010).
29. Li, Y., Liu, Y. & Liu, S. Effect of superparasitism on bionomics of *Cotesia plutellae*. *Chinese Journal of Biological Control* **17**, 151–154 (2001).
30. Yu, R.-X., Shi, M., Huang, F. & Chen, X.-X. Immature development of *Cotesia vestalis* (Hymenoptera: Braconidae), an endoparasitoid of *Plutella xylostella* (Lepidoptera: Plutellidae). *Annals of the Entomological Society of America* **101**, 189–196 (2008).
31. Mitsunaga, T., Shimoda, T. & Yano, E. Influence of food supply on longevity and parasitization ability of a larval endoparasitoid, *Cotesia plutellae* (Hymenoptera: Braconidae). *Appl. Entomol. Zool* **39**, 691–697 (2004).
32. Roux, O., Van Baaren, J., Gers, C., Arvanitakis, L. & Legal, L. Antennal structure and oviposition behavior of the *Plutella xylostella* specialist parasitoid: *Cotesia plutellae*. *Microscopy Research & Technique* **68**, 36–44 (2005).
33. Potting, R. P. J., Poppy, G. M. & Schuler, T. H. The role of volatiles from cruciferous plants and pre-flight experience in the foraging behaviour of the specialist parasitoid *Cotesia plutellae*. *Entomologia Experimentalis et Applicata* **93**, 87–95 (1999).
34. Chen, W. B. *et al.* Parasitised caterpillars suffer reduced predation: potential implications for intra-guild predation. *Scientific Reports* **7** (2017).
35. Liu, T. S. *et al.* Isolation and characterization of microsatellite loci for *Cotesia plutellae* (Hymenoptera: Braconidae). *Insects* **8** (2017).
36. Peakall, R. & Smouse, P. E. Genalex 6: genetic analysis in Excel. Population genetic software for teaching and research. *Molecular Ecology Notes* **6**, 288–295 (2006).
37. Peakall, R. & Smouse, P. E. GenALEX 6.5: genetic analysis in Excel. Population genetic software for teaching and research—an update. *Bioinformatics* **28**, 2537–2539 (2012).
38. Jervis, M. A., Ellers, J. & Harvey, J. A. Resource acquisition, allocation, and utilization in parasitoid reproductive strategies. *Annual Review of Entomology* **53**, 361–385 (2008).
39. Couchoux, C. & van Nouhuys, S. Effects of intraspecific competition and host-parasitoid developmental timing on foraging behaviour of a parasitoid wasp. *Journal of Insect Behavior* **27**, 283–301 (2014).
40. McKay, T. & Broce, A. B. Discrimination of self-parasitized hosts by the pupal parasitoid *Muscidifurax zaraptor* (Hymenoptera: Pteromalidae). *Annals of the Entomological Society of America* **97**, 592–599 (2004).
41. Yamada, Y. Y. & Sugaura, K. Evidence for adaptive self-superparasitism in the dryinid parasitoid *Haplogonatopus atratus* when conspecifics are present. *Oikos* **103**, 175–181 (2003).
42. Gariepy, T. D., Kuhlmann, U., Gillott, C. & Erlandson, M. Parasitoids, predators and PCR: the use of diagnostic molecular markers in biological control of Arthropods. *Journal of Applied Entomology* **131**, 225–240 (2007).
43. Santolamazza-Carbone, S. & Rivera, A. C. Superparasitism and sex ratio adjustment in a wasp parasitoid: results at variance with local mate competition? *Oecologia* **136**, 365–373 (2003).
44. Zhang, L., Bai, S., Yu, H. & Li, X. Intraspecific competition in *Cotesia vestalis*, a solitary endoparasitoid of *Plutella xylostella* larvae. *Chinese Journal of Biological Control* **30**, 128–133 (2014).
45. Lebreton, S., Christidès, J. P., Bagnères, A. G., Chevrièr, C. & Darrouzet, E. Modifications of the chemical profile of hosts after parasitism allow parasitoid females to assess the time elapsed since the first attack. *Journal of Chemical Ecology* **36**, 513–521 (2010).
46. Strand, M. R. & Godfray, H. C. J. Superparasitism and ovidic in parasitic Hymenoptera: theory and a case study of the ectoparasitoid *Bracon hebetor*. *Behavioral Ecology & Sociobiology* **24**, 421–432 (1989).
47. Tena, A., Kapranas, A., Garcia-Marí, F. & Luck, R. F. Host discrimination, superparasitism and infanticide by a gregarious endoparasitoid. *Animal Behaviour* **76**, 789–799 (2008).
48. Yamada, Y. Y. & Ikawa, K. Superparasitism strategy in a semisolitary parasitoid with imperfect self/non-self recognition, *Echthrodelpfax fairchildii*. *Entomologia Experimentalis et Applicata* **114**, 143–152 (2005).
49. Field, S., Keller, M. & Calbert, G. The pay-off from superparasitism in the egg parasitoid *Trissolcus basalis*, in relation to patch defence. *Ecological Entomology* **22**, 142–149 (1997).
50. Diazfleischer, F., Galvez, C. & Montoya, P. Oviposition, superparasitism, and egg load in the solitary parasitoid *Diachasmimorpha longicaudata* (Hymenoptera: Braconidae): response to host availability. *Annals of the Entomological Society of America* **108**, 235–241 (2015).
51. Gauthier, N., Monge, J. P. & Huignard, J. Superparasitism and host discrimination in the solitary ectoparasitoid *Dinarmus basalis*. *Entomologia Experimentalis et Applicata* **79**, 91–99 (1996).
52. Weber, C. A., Smilanick, J. M., Ehler, L. E. & Zalom, F. G. Ovipositional behavior and host discrimination in three scelionid egg parasitoids of stink bugs. *Biological Control* **6**, 245–252 (1996).
53. Darrouzet, E., Bignon, L. & Chevrièr, C. Impact of mating status on egg-laying and superparasitism behaviour in a parasitoid wasp. *Entomologia Experimentalis et Applicata* **123**, 279–285 (2007).
54. Lebreton, S., Labarussias, M., Chevrièr, C. & Darrouzet, E. Discrimination of the age of conspecific eggs by an ovipositing ectoparasitic wasp. *Entomologia Experimentalis et Applicata* **130**, 28–34 (2009).
55. Hoffmeister, T. S. & Roitberg, B. D. Evolutionary ecology of oviposition marking pheromones. In: *Hilker, M., Meiners, T., (eds.), Chemoecology of insect eggs and egg deposition.* (Germany: Blackwell Publishing, 2008).
56. Ito, E. & Yamada, Y. Y. Self-/conspecific discrimination and superparasitism strategy in the ovidical parasitoid *Echthrodelpfax fairchildii* (Hymenoptera: Dryinidae). *Insect Science* **21**, 741–749 (2014).
57. Liang, Q. F., Jia, Y. J. & Liu, T. X. Self- and conspecific discrimination between unparasitized and parasitized green peach aphid (Hemiptera: Aphididae), by *Aphelinus asychis* (Hymenoptera: Aphelinidae). *Journal of Economic Entomology* **110**, 430–437 (2017).
58. Yazdani, M., Glatz, R. & Keller, M. Host discrimination by the solitary endoparasitoid *Dolichogenidea tasmanica* (Hymenoptera: Braconidae). *Biocontrol Sci. Technol.* **25**, 155–162 (2015).
59. Wang, X. G. & Keller, M. A. A comparison of the host-searching efficiency of two larval parasitoids of *Plutella xylostella*. *Ecological Entomology* **27**, 105–114 (2002).

Acknowledgements

The work was supported by National Natural Science Foundation of China (No. 31230061 and No. 31320103922), Science and Technology Major Project of Fujian Province (No. 2018NZ01010013). LV is supported by the Minjiang Scholar Program in Fujian Province (PRC) and the Advanced Talents of SAFEA, and GMG by the National Thousand Talents Program in China and the Advanced Talents of SAFEA. We thank Mrs A. Johnson of Charles Sturt University for assistance with manuscript preparation. Associate Professor Feng-luan Yao from Fujian Academy of Agricultural Sciences for Figure drawing assistance. Professor Xue-xin Chen from Zhejiang University for supplying ZJ-strain parasitoids.

Author contributions

W.-B.C., G.M.G., M.-S.Y. and L.V. designed the research; W.-B.C., S.-Q.Z., H.-F.Z., J.M., T.-S.L., X.-Y.Z., X.W. and J.Z. performed the research; W.-B.C., G.M.G., M.-S.Y. and L.V. wrote and revised the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at <https://doi.org/10.1038/s41598-020-67050-1>.

Correspondence and requests for materials should be addressed to M.-s.Y. or G.M.G.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2020