Advanced Access publication on October 16, 2013 doi:10.1093/humupd/dmt054

human reproduction update

Melatonin and stable circadian rhythms optimize maternal, placental and fetal physiology

Russel J. Reiter^{1,*}, Dun Xian Tan¹, Ahmet Korkmaz², and Sergio A. Rosales-Corral³

¹Department of Cellular and Structural Biology, University of Texas Health Science Center, San Antonio, TX, USA ²Present address: Department of Physiology, Gulhane Medical School, Ankara, Turkey ³Present address: Centro de Investigacion Biomedica de Occidente del Instituto Mexicano del Seguno Social, Guadalajara, Mexico

Correspondence address. Tel: +210-567-3859; Fax: +210-567-3803; E-mail: reiter@uthscsa.edu

Submitted on July 17, 2013; resubmitted on September 6, 2013; accepted on September 11, 2013

TABLE OF CONTENTS

- Introduction
- Methods
- Melatonin as a free radical scavenger and as an antioxidant
- Melatonin production and function in the placenta
- Programming fetal circadian rhythmicity
- Pre-eclampsia and placental damage: protection by melatonin
- Melatonin in relation to parturition
- Conclusion
- References

BACKGROUND: Research within the last decade has shown melatonin to have previously-unsuspected beneficial actions on the peripheral reproductive organs. Likewise, numerous investigations have documented that stable circadian rhythms are also helpful in maintaining reproductive health. The relationship of melatonin and circadian rhythmicity to maternal and fetal health is summarized in this review.

METHODS: Databases were searched for the related published English literature up to 15 May 2013. The search terms used in various combinations included melatonin, circadian rhythms, biological clock, suprachiasmatic nucleus, ovary, pregnancy, uterus, placenta, fetus, pre-eclampsia, intrauterine growth restriction, ischemia-reperfusion, chronodisruption, antioxidants, oxidative stress and free radicals. The results of the studies uncovered are summarized herein.

RESULTS: Both melatonin and circadian rhythms impact reproduction, especially during pregnancy. Melatonin is a multifaceted molecule with direct free radical scavenging and indirect antioxidant activities. Melatonin is produced in both the ovary and in the placenta where it protects against molecular mutilation and cellular dysfunction arising from oxidative/nitrosative stress. The placenta, in particular, is often a site of excessive free radical generation due to less than optimal adhesion to the uterine wall, which leads to either persistent hypoxia or intermittent hypoxia and reoxygenation, processes that cause massive free radical generation and organ dysfunction. This may contribute to pre-eclampsia and other disorders which often complicate pregnancy. Melatonin has ameliorated free radical damage to the placenta and to the fetus in experiments using non-human mammals. Likewise, the maintenance of a regular maternal light/dark and sleep/wake cycle is important to stabilize circadian rhythms generated by the maternal central circadian pacemaker, the suprachiasmatic nuclei. Optimal circadian rhythmicity in the mother is important since her circadian clock, either directly or indirectly via the melatonin rhythm, programs the developing master oscillator of the fetus. Experimental studies have shown that disturbed maternal circadian rhythms, referred to as chronodisruption, and perturbed melatonin cycles have negative consequences for the maturing fetal oscillators, which may lead to psychological and behavioral problems in the newborn. To optimize regular circadian rhythms and prevent disturbances of the melatonin cycle during pregnancy, shift work and bright light exposure at night should be avoided, especially during the last trimester of pregnancy. Finally, melatonin synergizes with oxytocin to promote delivery of the

fetus. Since blood melatonin levels are normally highest during the dark period, the propensity of childbirth to occur at night may relate to the high levels of melatonin at this time which work in concert with oxytocin to enhance the strength of uterine contractions.

CONCLUSIONS: A number of conclusions naturally evolve from the data summarized in this review: (i) melatonin, of both pineal and placental origin, has essential functions in fetal maturation and placenta/uterine homeostasis; (ii) circadian clock genes, which are components of all cells including those in the peripheral reproductive organs, have important roles in reproductive and organismal (fetal and maternal) physiology; (iii) due to the potent antioxidant actions of melatonin, coupled with its virtual absence of toxicity, this indoleamine may have utility in the treatment of pre-eclampsia, intrauterine growth restriction, placental and fetal ischemia/reperfusion, etc. (iv) the propensity for parturition to occur at night may relate to the synergism between the nocturnal increase in melatonin and oxytocin.

Key words: circadian rhythms / fetus / melatonin / placenta / pre-eclampsia

Introduction

Even before the discovery of melatonin in the pineal gland (Lerner et al., 1958), the organ was experimentally linked to reproduction, but the findings were unpersuasive (Quay, 1956; Mogler, 1958). The decade of the 1960s, however, was accompanied by a series of seminal studies which showed that the gland functioned as a photoneuroendocrine transducer (Quay, 1963; Axelrod *et al.*, 1964, 1965) and it was definitively proved to have a physiological association with the hypothalamo-pituitary-gonadal axis (Hoffman and Reiter, 1965, 1966; Reiter and Hester, 1966). In particular, the daily changes in the duration of darkness under natural photoperiod conditions were found, via the pineal gland, to drive seasonal variations in reproductive capability in photosensitive species (Reiter, 1973, 1974). That melatonin was the mediator of pineal origin which determined the waxing and waning of reproductive competence under these conditions was uncovered shortly thereafter (Reiter *et al.*, 1976; Tamarkin *et al.*, 1976; Carter and Goldman, 1983).

In addition to its importance in intervening between the changing light:dark cycle and seasonal reproduction in photoperiodic species (Revel *et al.*, 2009; Lincoln and Hazlerigg, 2010), melatonin has numerous other actions on the gonads and adnexae of non-seasonally breeding mammals, including the human, that are beneficial to optimal peripheral reproductive organ health (Reiter *et al.*, 2009c, 2013a). The new data relating melatonin to pregnancy and delivery are summarized in this survey.

Research on circadian rhythms progressed concurrent with that of melatonin. Given that a normal light:dark cycle is of 24 h duration and has been for eons, it was advantageous for animals to modulate their metabolism, locomotor activity, etc. over a 24-h period to preserve energy, reduce predation pressure and avoid the likelihood of antagonistic cellular processes, e.g. the maximal activities of lipolytic and lipogenic enzymes in hepatocytes, from occurring simultaneously (Li et al., 2012; Thut et al., 2012). Thus, vertebrate species developed a light-sensitive biological clock whose function is governed by the prevailing light:dark cycle, the most regular recurring variable in the environment (Brainard et al., 1983, 1984; Meng et al., 2011; Ruger et al., 2013). This master clock is located in the suprachiasmatic nucleus (SCN) of the hypothalamus (Swanson and Cowan, 1975; Lydic et al., 1980). The SCN synchronizes many circadian rhythms in humans as in other vertebrates (Gillette and Tischkau, 1999; Hofman, 2000). Since the circadian production of melatonin in the pineal gland is also under the influence of the SCN, it is often difficult to separate the biological effects of general circadian disruption from those following from disturbances in the circadian melatonin cycle (Reiter et al., 2009b, 2012b).

In the following survey, the importance of the circadian clock to optimal reproductive physiology is often discussed separately from that due to alterations in melatonin (and vice versa). In reality, however, there is often no way of knowing whether a particular abnormal process relates to a derangement of the biological clock or to a perturbation or total suppression of the cyclic production of melatonin (Dumont et al., 2012).

Methods

The published literature utilized to compile this review was retrieved from a number of sources. Because of the relative diversity of subjects reviewed (i.e. mechanisms of melatonin synthesis in the pineal and in the placenta, peripheral melatonin receptors, free radical biology and the role of melatonin and its derivatives as free radical scavengers and as antioxidants, functional aspects of the central and peripheral circadian clocks, maturation and development of fetal physiology, diseases of pregnancy and the control of parturition), search terms were broad and numerous. These included, in various combinations, melatonin, circadian rhythms, biological clock, SCN, ovary, pregnancy, uterus, placenta, fetus, pre-eclampsia, intrauterine growth restriction, ischemia/reperfusion, chronodisruption, antioxidants oxidative stress and free radicals. All germane literature uncovered is summarized in this report.

Melatonin as a free radical scavenger and as an antioxidant

Whereas an extensive discussion of the actions of melatonin as a direct free radical scavenger (Tan *et al.*, 1993; Galano *et al.*, 2011) and as an indirect antioxidant (Barlow-Walden *et al.*, 1995; Rodriguez *et al.*, 2004) is beyond the scope of this review, the data overwhelmingly show that this indoleamine is highly effective in reducing oxidative stress throughout the body (Maldonado *et al.*, 2007; de Matos *et al.*, 2012; Tamura *et al.*, 2013) including in the ovary, placenta, fetus and mother (see below). In studies where melatonin was compared with other better-known antioxidants (e.g. vitamins C, E, etc.) in terms of their protective efficiency against the damaging actions of toxic oxygen and nitrogen-based reactants, melatonin has proved more effective (Martin *et al.*, 2000a; Reiter *et al.*, 2009a; Milczarek *et al.*, 2010). Recently, investigators also found that when melatonin was compared with chemically designed mitochondriatargeted antioxidants, again melatonin was superior in reducing molecular damage resulting from free radicals (Lowes *et al.*, 2013).

There are several features that make melatonin highly efficient in protecting beleaguered macromolecules from oxidative/nitrosative stress (Fig. 1). First, there are no morphophysiological barriers to melatonin, e.g. it readily crosses the blood–brain barrier and the placenta (Schenker et *al.*, 1998; Okatani *et al.*, 1999), and so in addition to all maternal organs it also protects the placenta and the fetus. Melatonin readily gains access to the major sites of free radical generation, e.g. mitochondria. Besides being located in several subcellular compartments (Menendez-Pelaez and Reiter, 1993; Venegas *et al.*, 2012), melatonin actually may be produced in the mitochondria (Tan *et al.*, 2013) where free radical generation is especially high. Not only melatonin but also several of its metabolites that are formed when it functions as a directfree radical scavenger, i.e. cyclic 3-hydroxymelatonin, N^1 -acetyl- N^2 -formyl-5-methoxy-kynuramine (AFMK), N^1 -acetyl-5-methoxykynuramine (AMK), etc. are

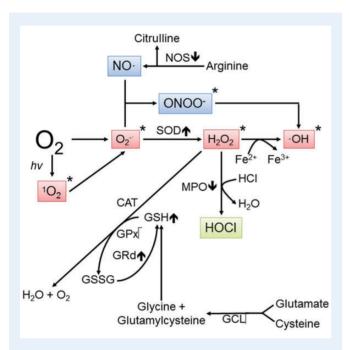


Figure | Pathways showing the conversion of molecular oxygen to reactive oxygen and nitrogen species. An estimated 1-4% of the O2 inhaled is eventually converted to reactive products. When produced in excess, free radicals and other derivatives of oxygen are highly destructive to peripheral reproductive tissues. A single electron reduction of melatonin generates the superoxide anion radical $(O_2^{\bullet-})$ which is either quickly metabolized by superoxide dismutase (SOD) to hydrogen peroxide (H_2O_2) or it couples with nitric oxide (NO^{\bullet}) to produce the peroxynitrite anion (ONOO⁻). H₂O₂ is transformed to the hydroxyl radical (*OH) in the presence of transition metals, represented here by iron. O_2 can also be converted to singlet oxygen (1O_2), a less common but toxic species. $O_2^{\bullet-}$, 1O_2 , H_2O_2 and ${}^{\bullet}OH$ are often classified as reactive oxygen species (ROS), whereas NO[•] and ONOO⁻ are classified as reactive nitrogen species (RNS). Of these products, the [•]OH and ONOO⁻ are considered the most reactive and damaging. The items marked with an asterisk are scavenged by melatonin and/or its metabolites. The up and down arrows associated with the enzymes indicate whether the action of melatonin is either stimulatory or inhibitory to its activity, respectively. H_2O_2 is metabolized via several routes which eventually leads to the production of non-toxic species. CAT, catalase; GCL, glutamyl cysteine ligase; GPx, glutathione peroxidase; GRd, glutathione reductase; GSSG, glutathione disulfide; HOCI, hypochlorous acid; MPO, myeloperoxidase; NOS, nitric oxide synthase.

also radical scavengers (Tan et al., 2007; Hardeland et al., 2009; Hardeland, 2012) and some may be better at doing so than melatonin itself (Galano et al., 2013). Thus, melatonin is the progenitor of a variety of free radical scavengers which function in an antioxidant cascade to prevent radical-mediated damage (Fig. 2) (Tan et al., 2002). Finally, in terms of its indirect functions in limiting oxidative stress, melatonin stimulates several antioxidative enzymes (Barlow-Walden et al., 1995; Pablos et al., 1998; Rodriguez et al., 2004), likely via receptormediated processes (Tomas-Zapico and Coto-Montes, 2005). Its ubiquitous distribution, ease of transfer between subcellular compartments and broad spectrum capabilities make melatonin highly effective in combatting free radical damage in the reproductive system and elsewhere.

In addition to removing toxic oxygen and nitrogen species from cells by either directly quenching them or by metabolizing them to non-reactive products, melatonin also exhibits significant anti-inflammatory activity (Carrillo-Vico *et al.*, 2005; Mauriz *et al.*, 2013). Melatonin's efficacy as an anti-inflammatory agent stems from its ability to reduce gene expression and activities of inducible nitric oxide synthase and cyclo-oxygenase, and by limiting the production of a variety of pro-inflammatory molecules including prostanoids, leukotrienes, cytokines and adhesion molecules. Melatonin modulates the production of these factors by regulating several transcription factors including nuclear factor - κ B, nuclear factor

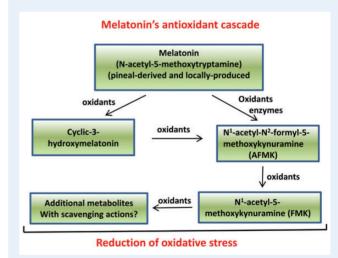


Figure 2 In what is referred to as the antioxidant cascade, melatonin and its metabolites function in the reduction of oxidative stress. Melatonin is a powerful antioxidant due to its ability to neutralize toxic-free radicals in both the mother and the fetus. The metabolism of melatonin is a continuum when it functions as a radical scavenger such that each metabolite, like melatonin, also detoxifies radical species. This pathway is referred to as melatonin's antioxidant cascade. In this process, not only melatonin-but also cyclic 3-hydroxymelatonin (cyclic-3OH-melatonin), N^1 -acetyl- N^2 -formyl-5-methoxykynuramine (known as AFMK) and N^{1} -acetyl-5-methoxykynuramine (known as AMK)—all function as radical scavengers. Finally, preliminary findings indicate that the molecule(s) produced when AMK neutralizes radicals may also function as a scavenger(s). Among the metabolic products listed, cyclic-3OH-melatonin and AMK may actually be more effective radical scavengers than melatonin itself. Thus, melatonin could be referred to as a pro-antioxidant or pro-drug.

erythroid-2 related factor 2 and hypoxia-inducible factor (Mauriz et al., 2013). As a component of this process, melatonin reduces the recruitment of leukocytes to areas of injury; this also limits oxidative damage since leukocytes can be a major source of free radicals and, therefore, of the associated molecular damage.

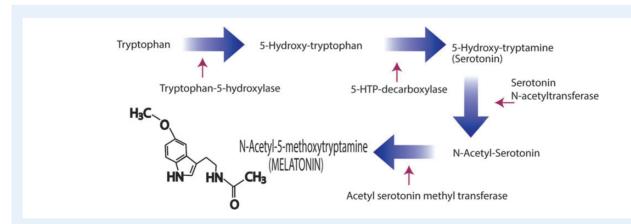
While melatonin's multi-faceted antioxidant actions are of fundamental importance and were the presumed original functions of melatonin during evolution (Tan et al., 2013), it is not the exclusive role of melatonin to guench and/or neutralize free radicals (Reiter et al., 2010a; 2012a). Rather, this ubiquitously-acting molecule has a variety of other functions that have been well documented in mammals including man; some of these may involve not only changes in the absolute levels of melatonin but also perturbations of its circadian rhythm. Thus, changes in melatonin have been linked to sleep disturbances (Santhi et al., 2012), psychological depression (Cardinali et al., 2012), metabolism leading to body weight regulation (Fenn et al., 2011), attention deficit/hyperactivity disorder (Chaste et al., 2011) and many others. Research on melatonin and circadian rhythms is progressing at a rapid pace and it seems likely that the discoveries to date are only a fraction of the real functions of melatonin and circadian rhythmicity. How all these facets of melatonin and circadian rhythms impact reproductive physiology are only beginning to be uncovered.

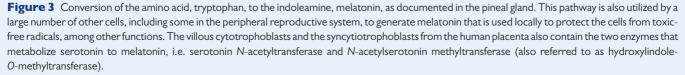
Melatonin production and function in the placenta

Melatonin, *N*-acetyl-5-methoxytryptamine, is synthesized from tryptophan (Fig. 3) and was initially thought to be solely of pineal origin (Lerner et al., 1958). This idea was dispelled, however, when melatonin showed up in other organs along with the enzymes that produce it (Reiter et al., 2013b). While the pineal gland is the only organ in which melatonin has been found to be generated in a light:dark-dependent circadian manner (Axelrod et al., 1965; Panke et al., 1979), as reflected in the blood levels of this indole (Vaughan et al., 1976; Wehr, 1991; Kennaway et al., 1992), it is now clear that many, perhaps all, organs have acquired the ability to generate this important molecule. If melatonin is synthesized in mitochondria, as recently proposed (Tan et al., 2013), then its ubiquitous production in organisms would be assured since all eukaryotic cells are endowed with these energy-generating organelles. The advantage of mitochondria being a source of this antioxidant would certainly be fortuitous since these organelles are a major site of origin of damaging oxygen derivatives. Thus in this location, melatonin, due to its potent radical scavenging activity (Tan et al., 1993; Allegra et al., 2003; Reiter et al., 2009a; Hardeland, 2012), would be ideally situated to prevent these destructive agents from mutilating neighboring molecules. In essence, melatonin would provide on-site protection. Certainly, this is consistent with the high levels of melatonin in mitochondria (Venegas et al., 2012) and the ability of melatonin to protect this organelle from damage and preserve its physiology (Martin et al., 2000b; Acuna-Castroviejo et al., 2002, 2011; Lowes et al., 2013). The vastly different levels of melatonin in fluids and subcellular organelles have led to a debate about what constitutes a physiological concentration of melatonin (Reiter and Tan, 2003).

Specifically in the reproductive system, melatonin is reportedly produced in the ovary (Itoh et al., 1999), in the oocyte per se (Sakaguchi et al., 2013), in the cumulus cells surrounding the oocyte (El-Raey et al., 2011) and in the placenta (Lanoix et al., 2008). Moreover, melatonin levels are higher in the ovarian follicular fluid than they are in the general circulation (Brzezinski et al., 1987; Röunberg et al., 1990; Nakamura et al., 2003). In the follicular fluid, melatonin has access to the surrounding tissues where it may have the important function of protecting the oocyte from oxidative damage during both maturation and ovulation (Tamura et al., 2013), a time when free radical formation is elevated (Brannstrom and Norman, 1993; Behrman et al., 2001; Sugino, 2005). This idea is consistent with the observation that melatonin was effective in reducing molecular damage to human ova (Tamura et al., 2008a, b) that were used for IVF-embryo transfer. Just as melatonin prevents free radical damage to ova, it also does so for sperm (Ortiz et al., 2011; Reiter et al., 2013a).

The placental villous trophoblasts are not only a source of melatonin but also they contain the classic transmembrane receptors for the





indole, i.e. MTI and MT2 (Lanoix et al., 2008). To document this, villous cytotrophoblasts were isolated from human term placentas (37-41 weeks of gestation) after vaginal delivery. The two enzymes that convert the precursor, serotonin, to melatonin, i.e. arylalkylamine N-acetyltransferase (AANAT) and acetylserotonin methyltransferase (ASMT) (formerly known as hydroxyindole-O-methyltransferase) (Fig. 3) are expressed and are active in both cytotrophoblast and syncytiotrophoblast cells as well as in IEG-3 and BeWo choriocarcinoma cells, commonly used as in vitro models for human trophoblasts. Given that villous trophoblasts apparently have the capability of generating melatonin and they also possess membrane receptors, Lanoix et al. (2008) speculated that locally produced melatonin likely has paracrine, autocrine and/or intracrine actions in the placenta. In addition to working via MT1 and MT2 receptors, melatonin also could directly scavenge radicals and reduce oxidative damage to placental tissues, which would be important given the large number of radicals that are often produced in this tissue when its function becomes compromised (Pringle et al., 2010).

In the placenta, the primary villous trophoblasts include two distinct categories of cells, i.e. the mononuclear villous cytotrophoblasts (vCTB) and the multinucleated syncytiotrophoblast (STB). The fusion of the vCTB to form the STB is a highly regulated process. To ensure the transition of vCTB to the STB, the vCTB cells are generally not lost due to apoptosis. Based on the recent *in vitro* findings of Lanoix *et al.* (2012b), melatonin reduces the loss of human vCTB cells by seemingly increasing their resistance to apoptosis via the intrinsic apoptotic pathway. The suppression of the apoptotic response by melatonin in vCTB cells was shown to be a membrane receptor-mediated process, given that the addition of luzindole, an MT1/MT2 receptor antagonist, to the culture medium blocked the anti-apoptotic actions of melatonin in vCTB cells.

The STB is a non-proliferative tissue with a pro-apoptotic phenotype (Vaillancourt *et al.*, 2009). Thus, throughout pregnancy, lost STB must be continually regenerated by fusion of vCTB cells. In the absence of the persistent input of vCTB cells into the STB, the latter would quickly be reduced in size since its life span is short (Huppertz and Kingdom, 2004; Vaillancourt *et al.*, 2009). Because melatonin preserves the vCTB and thereby makes them available for formation of the STB, melatonin may have an important role in maintaining homeostasis in the placenta (Fig. 4). STB degeneration is a consequence of syncytial loss due to both the intrinsic and extrinsic pathways (Heazell *et al.*, 2008). The specific actions of melatonin on the apoptotic processes of STB has yet to be defined; this action could be the same or different than melatonin's function at the level of the vCTB.

Interestingly, trophoblast-derived cancer cells respond differently to melatonin than do the vCTB cells. Several choriocarcinoma cell lines have been established. One of the best studied is the BeWo line (Lewis *et al.*, 1994). BeWo cells also have the capability of fusing to form a syncytium and they synthesize melatonin and possess its receptors (Lanoix *et al.*, 2006, 2008). As with numerous other cancer cells (Sainz *et al.*, 2003; Uguz *et al.*, 2012; Wang *et al.*, 2012), BeWo cells respond to melatonin by undergoing apoptosis, a response that is opposite that of normal vCTB. Thus, the actions of melatonin in the placenta are clearly context specific. As already mentioned, it is common that melatonin preserves normal cells while promoting processes that cause degeneration and death of cancer cells (Choi *et al.*, 2008; Proietti *et al.*, 2013).

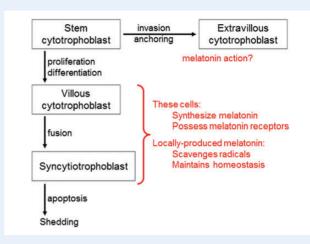


Figure 4 The functions of locally produced melotonin in the villous cytotrophoblasts of the placenta. The enzymatic machinery required for the synthesis of melatonin has been identified in the human villous cytotrophoblasts and in the syncytiotrophoblasts. Although clock genes have been identified in the placenta, whether there is a 24-h rhythm in melatonin production in this tissue is yet to be examined. The classic membrane receptors, MTI and MT2, for this indoleamine are also present in the placenta. Locally generated melatonin likely functions in the protection of the placenta from oxidative stress employing both receptor-dependent and receptor-independent processes. Additionally, melatonin probably reduces the loss of villous cytotrophoblasts by preventing apoptosis of these cells; melatonin's anti-apoptotic actions are well known in normal cells. Via its influence on cell survival, melatonin presumably maintains the stable turnover of the syncytiotrophoblast. Whether melatonin has any actions at the level of the extravillous trophoblasts, especially during invasion of the uterine wall should be examined.

Germane to a discussion of melatonin's action in the placenta is a report which claimed that night-time circulating melatonin levels are depressed in women suffering with severe pre-eclampsia when compared with those in women with a normal pregnancy or with mild pre-eclampsia (Nakamura *et al.*, 2001). Since especially severe pre-eclampsia is believed to involve the excessive production of free radicals, the lower melatonin levels could be a consequence of a more rapid utilization of this free radical scavenger or a consequence of a reduced production. Also, given that the night-time level of melatonin was measured at a single time point in these three groups of women, the apparently lower values in those with severe pre-eclampsia could have been a result of a shifted nocturnal melatonin peak rather than an actual depression of its absolute concentration.

Consistent with their continuing interest in the relevance of melatonin to placental physiology, Lanoix *et al.* (2012b) compared the levels of melatonin, its precursor, its synthesizing enzymes and its membrane receptors in pre-eclamptic placentas with those from gestation-matched normotensive control placentas. They observed that relative to control tissues, pre-eclamptic placentas had reduced AANAT gene expression and enzyme activity along with lower ASMT activity. Correlating with these observations were dramatically depressed concentrations of melatonin and elevated levels of its precursor, serotonin. Finally, preeclampsia was associated with a significant reduction in both the MTI and MT2 receptor in the placenta. In view of these outcomes, it seems clear that the markedly depressed level of melatonin in pre-eclamptic placentas is a consequence of impaired production of the indoleamine due to a major deficiency in the rate-limiting enzyme AANAT. The drop in melatonin in the pre-eclamptic placenta may help to explain the lower levels of blood melatonin in women with pre-eclampsia (Nakamura et al., 2001); however, this correlation implies that placenta-derived melatonin is normally released into the blood, perhaps especially in nearterm pregnancy, when blood levels of the indoleamine reach their maximal values (Tamura et al., 2008a). Lanoix et al. (2012b) also suggested that depressed circulating melatonin levels during pregnancy may be a biomarker for the diagnosis of pre-eclampsia. They also surmise that melatonin may be useful as a treatment for pre-eclampsia.

Programming fetal circadian rhythmicity

The maternal circadian system is organized by the master clock, a small group of neurons located bilaterally at the base of the diencephalon/telencephalon junction in the anterior hypothalamus (Mason and Lincoln, 1976). These neurons, referred to as the SCN, regulate circadian rhythms of most if not all organs in the body. At the molecular level, the rhythms in the SCN are driven by a transcriptional/translational feedback loop that involves genes *Per 1, Cry 1, Cry 2, Clock* and *Bmal 1* (Bass and Takahashi, 2010); these rhythms in turn regulate circadian oscillations in peripheral organs (Menaker *et al.*, 2013).

Clock genes have been described in two functionally distinct zones (junctional and labyrinth) of the rat placenta near the time of delivery (Wharfe et al., 2011). Clock gene expression was compared in placentas collected at four time points throughout a light:dark cycle (collected at 0800, 1400, 2000 and 0200 h). The findings revealed that although the canonical circadian genes were expressed in both placenta zones, the circadian changes were not robust nor were they well co-ordinated. Despite these data, Waddell et al. (2012) caution against discounting a potential role of circadian rhythms as a key component of the normal placental phenotype.

A primary entrainer of the cyclic function of the SCN is the light:dark cycle, as detected by the retinas. The eyes are connected to the SCN via a specialized group of axons from a select population of retinal ganglion cells (Sand *et al.*, 2011; Lucas, 2013); the axons of these cells are referred to as the retinohypothalamic tract (Mason and Lincoln, 1976) and they course with the optic nerve to the SCN.

Axons of SCN cells project to adjacent hypothalamic neurons where they synchronize overt circadian rhythms, such as body temperature, sleeping/waking, feeding and adrenocorticotrophic hormone/corticosteroid. Additionally, via a more complex neural route involving central and peripheral sympathetic neurons, the SCN regulates the production and release of melatonin from the pineal gland (Stehle *et al.*, 2011). Blood melatonin levels are typically 10-15 times higher at night than during the day as a result of the nocturnal production and release of the indole from the pineal gland. The maternal melatonin rhythm may have a major role in influencing the development of the fetus since unaltered melatonin readily crosses the placenta (Okatani *et al.*, 1999; Schenker *et al.*, 1998). Melatonin produced in organs other than the pineal gland is minimally released into the blood, but rather is used in the organ where it is synthesized as an autocoid or paracoid (Tan *et al.*, 2003). The circadian clocks in many peripheral organs, for example the kidney, adrenal gland, lung, are entrained by SCN-derived information supplied to them by their autonomic innervation (Bass and Takahashi, 2010). Also, daily fluctuations in body temperature and cortisol may aid in synchronizing the clocks of peripheral organs. Finally, the day/ night changes in circulating melatonin cue peripheral organs of the prevailing light:dark environment (Pevet and Challet, 2011). Thus, the metabolism of cells throughout the body is exposed to a variety of circadian messages that serve to regulate gene expression. The expression of an estimated 5–15% of the genes in peripheral organs is circadian in nature (Richards and Gumz, 2012).

The maturation and synchronization of the fetal circadian system has eluded detailed examination to date. Evidence from studies conducted in humans and non-human primates has revealed entrained 24-h rhythms in fetal heart rate and respiratory movements during the latter half of pregnancy (Seron-Ferre *et al.*, 2007). Whether the circadian system of the fetus, particularly in late pregnancy, has anything reminiscent of the pervasive influence of the maternal SCN is unknown. It has been demonstrated that the post-natal development of the peripheral circadian system in offspring of mothers lacking a functional master clock (due to lesions of the SCN or to gene knockouts) is unexpectedly relatively normal (Jud and Albrecht, 2006).

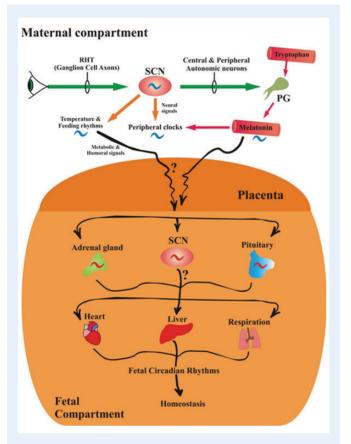
In those mammals (including the human) where studies have been done, the SCN is already morphologically distinguishable in the basal hypothalamus of the fetus by mid-gestation. Before birth it exhibits rhythms in mRNA for vasopressin (an important transmitter in the SCN) and cfos mRNA and protein (Reppert and Schwartz, 1983; Novakova et al., 2010). Moreover, the innervating fibers from the retinohypothalamic tract to the SCN are apparent before term delivery (Weinert, 2005). The oscillations that occur in the fetal SCN are typically of lower amplitude than those in the adult master clock (Watanabe et al., 2006). While the retinohypothalamic input to the SCN is apparent in the late-gestation fetus, the output of the fetal clock to the pineal gland seems to be incomplete until after birth (Reiter, 1991). Thus, there is no evidence that a fetal melatonin rhythm influences any aspect of fetal development since the pineal gland requires an innervation from the SCN via the peripheral sympathetic nervous system to produce melatonin.

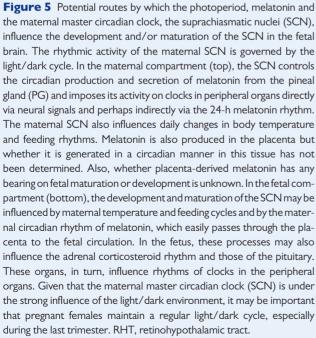
Melatonin of maternal origin, however, does influence the fetus by virtue of its rhythm and the ease with which it crosses the placenta. Studies in the human have repeatedly confirmed that the cycle of melatonin in the maternal blood also occurs in the fetal circulation (Kennaway et al., 1996; Okatani et al., 1999). This rhythm, when abolished by exposure of the mother to constant light, changes the rhythmic expression in fetal clock genes; these changes are reversed when daily melatonin injections are given to the mother (Torres-Farfan et al., 2006). This documents that the fetal clock is imprinted by melatonin, which under normal circumstances is of maternal origin. The human fetus displays 24-h rhythms in temperature and oxygen consumption by 32–33 weeks of gestation (Bauer et al., 2009).

In addition to the maternal melatonin rhythm, judging from the cyclic fluctuation of cortisol levels in the umbilical artery and vein, the term fetus is also exposed to this rhythm (Seron-Ferre et al., 2001). The so-called fetal zone of the fetal adrenal gland also secretes large amounts of dehy-droepiandrosterone sulfate (DHEAS), which causes a 24-h rhythm in the fetal circulation; DHEAS is a necessary precursor for placental estrogen production during pregnancy (Seron-Ferre et al., 2007). Whether there

is any functional interaction of the fetal melatonin and DHEAS rhythms at the level of the fetal SCN or elsewhere is unknown.

Evidence accumulated to date suggests that the circadian architecture in the fetus, particularly in the SCN, is entrained by the maternal melatonin cycle (Fig. 5) and possibly to a lesser extent by manipulation of





the feeding times (Novakova et al., 2010). As mentioned above, exposure of pregnant mothers to constant light disrupts the maternal activity cycle and suppresses the circadian melatonin rhythm; these perturbations impact gene expression in the fetal SCN. Studies in rodents have shown that maternal-generated rhythms, when disrupted during pregnancy, also alter the post-natal behavioral circadian rhythms in the offspring. For example, maternal pinealectomy, which abolishes the circadian melatonin cycle, significantly interferes with the drinking rhythm in the offspring while this cycle is restored when the pinealectomized mothers are given regular melatonin injections in late pregnancy (Bellavia et al., 2006). These observations emphasize the potential danger of constant light exposure, unusual light/dark cycles or night shift work for pregnant humans especially during the last trimester. This is certainly not trivial given that throughout the world an estimated one-fourth to one-third of female employees work at night and, moreover, considering the high degree of light pollution in urban areas, avoiding light at night sufficient to alter the function of the biological clock and circulating melatonin levels is becoming progressively more difficult.

There is now general agreement that maternal circadian rhythms are influential in the entrainment and programming of fetal and newborn circadian rhythms (Fig. 5), but what rhythms are affected may be species specific. While information in this field is still rudimentary, evidence has shown that disturbances of the fetal circadian system, regardless of the cause of those perturbations, have long-term consequences in the offspring. As an example, women who engage in shift work during pregnancy have an increased incidence of spontaneous abortions, premature deliveries and low birthweight infants (Zhu et al., 2004). Shift work greatly alters the melatonin cycle, the sleep/wake rhythm and feeding times which also could be instrumental in contributing to these complications. Exposure of rats to simulated shift work caused increased hyperleptinemia and adiposity of the offspring at 3 months after birth and altered glucose tolerance and insulin resistance, reminiscent of that seen in metabolic syndrome, when the offspring were I year of age (Varcoe et al., 2011). Clearly, the impact of a disturbed light: dark cycle in late pregnancy and during the perinatal period may have major effects on subsequent behavioral and metabolic functions (Ferreira et al., 2012). Given the remarkable rise in the diagnosis of metabolic syndrome, obesity, attention deficit-hyperactivity disorder, autism spectrum disorders, etc., it may be worthwhile to be more attentive to the light:dark environment during pregnancy (Hardeland et al., 2012). This is especially true since the frequency of these conditions has run in parallel with excessive use of light at night, a change that is difficult to avoid in current societies.

Offspring that are delivered prematurely are not exposed to a normal maternal melatonin rhythm that they would be if they were still *in utero* and they do not generate a melatonin rhythm on their own (Kennaway et al., 1992). Thus, preterm infants are deprived of exposure to the melatonin cycle during a critical interval of their development. How or whether the lack of exposure to this rhythm in these premature infants has consequences on any aspect of development remains unexamined. This problem could be potentially partially rectified if premature infants would be breast fed since a melatonin rhythm normally exists in human breast milk, with higher levels at night than during the day (Illner-ova et al., 1993). This still would require, however, that the mother be in darkness for several hours before and at the time of night-time breast feeding and also the much lower levels of melatonin in the breast milk (relative to those in the blood) may render these concentrations insignificant in terms of programming the SCN of the newborn.

Pre-eclampsia and placental damage: protection by melatonin

Pre-eclampsia is a dangerous complication of pregnancy which causes significant maternal and fetal morbidity and death (Hung and Burton, 2006; Nelissen *et al.*, 2011; Redman, 2011). It is commonly diagnosed in the latter half of pregnancy but it may involve poor placentation during the earliest stage of pregnancy (Fig. 6). It can progress to an even more serious condition, eclampsia, which is accompanied by life-threatening seizures.

Pre-eclampsia has multiple facets with a commonly observed sign being hypertension. As the condition progresses, the damaged placenta releases molecules into the circulation which cause endothelial dysfunction leading to an elevation in blood pressure (Roberts *et al.*, 1989; Roberts and Hubel, 2009) and eventually more systemic damage, for example at the level of the kidney: these latter changes initiate proteinuria as a result of glomerular endotheliosis (Gaber *et al.*, 1994). Several features which characterize pre-eclampsia contribute to the pathophysiology of the disease including hypertension, inflammation and oxidative/nitrosative stress. The only effective treatment for preeclampsia is the delivery of the fetus and the placenta, although the condition can occur during the first 2 weeks after child birth.

The role of oxidative and/or nitrosative stress in pre-eclampsia is well accepted (Myatt *et al.*, 1996; Hung and Burton, 2006), although it is debated as to what causes the generation of the toxic oxygen derivatives that contribute to molecular damage. One theory states that free radicals and other reactive species are a consequence of a compromised

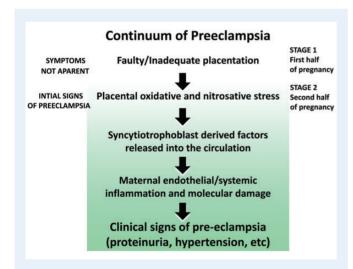


Figure 6 The potential sequence of events that contribute to preeclampsia. The oxidative/nitrosative stress that occurs may be a result of permanent hypoxia or intermittent hypoxia/reoxygenation resulting from placental maldevelopment because of inadequate attachment of the placenta to the uterine wall. Melatonin, due to its potent antioxidative actions, reduces ischemia/reperfusion injury to the placenta under experimental conditions and could potentially alleviate some of the signs of pre-eclampsia for the same reason. Melatonin also has antihypertensive actions which may aid in lowering the blood pressure in pre-eclamptic women. Not illustrated in this figure are the negative effects that chronodisruption has during pregnancy (see text). uteroplacental arterial flow because of the improper remodeling of the spiral arteries during placentation (Kaufmann et al., 2003). This leads to a low oxygen tension and a relative hypoxic state in the placenta. Another view is that it is not a matter of a chronically diminished blood flow to the placenta, but rather the problem resides with the intermittency of blood flow to this tissue. This results in intervals of ample arterial blood flow followed by intervals when blood flow is inadequate, which leads to hypoxia/reoxygenation injury (Hung and Burton, 2006). These presumptive explanations differ in that in the first case the placenta is in a chronic hypoxic state while in the second situation there are repeated intervals of hypoxia followed by reoxygenation. Under both conditions, massive numbers of free radicals would be generated thereby ensuring elevated molecular damage to the uterine trophoblasts and the release of a wide range of factors that are known to be altered in pre-eclampsia that then promote inflammation. A review by Redman and Sargent (2009) describes these inflammatory agents in detail and an extensive discussion of these data is considered beyond the scope of this report.

Considering the proposed involvement of free radicals and oxidative stress as contributors to the placental damage that occurs early in preeclampsia, it was anticipated that antioxidants may be beneficial in ameliorating the severity or progression of this disease. When tested, however, the data do not generally support the use of either vitamins C or E as having benefit in the treatment of pre-eclampsia (Conde-Agudelo *et al.*, 2011; Rossi and Mullin, 2011). This is also the case with some other antioxidants, for example co-enzyme Q10, that have been examined (Briceno-Perez *et al.*, 2009).

As noted above, melatonin as an antioxidant has a much greater capacity to scavenge radicals and reduce oxidative damage than do either vitamins C or E, including in the placenta (Martin et al., 2000a; Milczarek et al., 2010). Melatonin and its secondary, tertiary and quaternary metabolites function as radical scavengers and so melatonin neutralizes many more toxic reactants than do the vitamin antioxidants (Tan et al., 1993, 2002; Galano et al., 2011, 2013). Moreover, melatonin acts as an indirect antioxidant by stimulating antioxidative enzymes (Barlow-Walden et al., 1995; Pablos et al., 1998; Rodriguez et al., 2004) and glutathione production (Urata et al, 1999), a tripeptide with significant antioxidative activity.

Since the initial assault that leads to pre-eclampsia may be oxidative damage to the placenta arising from chronic hypoxia (Kaufmann et al., 2003) or repeated hypoxia and reoxygenation (Hung and Burton, 2006) of the placental tissue, Okatani et al. (2001) examined whether melatonin would protect against placental ischemia and reperfusion. To achieve this, the authors occluded the utero-ovarian arteries bilaterally for 20 min on Day 19 of pregnancy in rats, half of which were treated with melatonin (10 mg/kg) 60 min in advance of arterial occlusion. The hypoxic/reoxygenation episode was followed by a reduction in the placental mitochondrial respiratory control index and drop in the ratio of adenosine-5-diphosphate (ADP) concentration to oxygen consumption (ADP/O) during state 3 respiration, while the level of oxidized lipids in the placenta rose. These changes were prevented when melatonin was given documenting that the indole, likely due to its antioxidant activity, is capable of protecting the placenta from ischemia/reperfusion injury. Moreover, the authors surmised that melatonin may be a beneficial treatment in other situations where free radicals contribute to fetal problems, e.g. fetal growth restriction and fetal hypoxia associated with a difficult delivery. Interestingly, subsequent reports have proved this speculation valid; melatonin reduces fetal growth restriction

in rats due to either placental ischemia/reperfusion (Nagai et al., 2008) or undernourishment during pregnancy (Richter et al., 2009) and in sheep resulting from a period of ischemia followed by reperfusion (Lemley et al., 2012). In humans as well, melatonin reduced oxidative damage in newborns that experienced a period of asphyxia due to difficult delivery (Fulia et al., 2001).

Lanoix et al. (2012a, 2013) recently completed a series of *in vitro* studies in which they used melatonin to protect primary human STB cells from apoptosis due to 4-h hypoxia followed by 18 h of reoxygenation. The hypoxia/reoxygenation procedure clearly caused oxidative stress in the STB cells, which led to an activation of the Bax/Bcl-2 mitochondrial apoptosis pathway and DNA fragmentation. In this *in vitro* model, melatonin markedly reduced both apoptosis and the associated DNA damage. These findings support the possibility that melatonin may be useful *in vivo* to limit complications of pregnancy that involve damage to STB cells and their premature loss. For example, STB apoptosis is elevated in pre-eclampsia and intrauterine growth restriction (Heazell et al., 2008; Tomas et al., 2011).

In addition to oxidative damage to the placenta, etc., circadian misalignment may be one of many factors that contribute to pre-eclampsia (Ditisheim et al., 2013). Chronodisruption has been tentatively experimentally linked to other reproductive malfunctions including interruption of pregnancy, spontaneous abortion and low birthweight infants (Mahoney, 2010; Summa et al., 2012). The epidemiological data, however, have not uniformly supported an association between preeclampsia and shift work, a known circadian disrupter (Wergeland and Strand, 1997; Haelterman et al., 2007; Chang et al., 2010).

Gestational hypertension is one aspect of pre-eclampsia that may be linked to circadian disruption given that these rhythmic perturbations are also often associated with melatonin suppression. Normally, blood pressure in humans varies over a 24-h period with lowest pressure occurring at night (de la Sierra et al., 2009; Reiter et al., 2010b). This nocturnal reduction is mediated by the concurrent rise in circulating melatonin (Obayashi et al., 2013). Moreover, exogenously administered melatonin reduces hypertension in humans (Arangino et al., 1999). Thus, with the elimination of the night-time elevation of melatonin due to shift work, night-time light exposure, etc., some degree of hypertension would be expected, such as that which occurs in pre-eclampsia. Finally, placental diseases including pre-eclampsia may have an impact on the cardiovascular health of the mother later in life (Mosca et al., 2011). The cause of this association presumably relates to the endothelial damage sustained during a difficult pregnancy (Siddiqui and Hladunewich, 2011). Thus, if melatonin would have utility in reducing the severity of pre-eclampsia one downstream effect may also be an improvement of the cardiovascular health of the mother in the long term.

Pre-eclampsia is often accompanied by what is referred to as the HELLP syndrome; this condition includes hemolysis, elevated liver enzymes and low platelet count (Weinstein, 1982). Since it occurs coincident with pre-eclampsia, it is speculated to have a similar causative basis in terms of abnormal placental development or physiology (Abildgaard and Heimdal, 2013). Whether the signs of HELLP have any relation to melatonin has never been investigated. Platelets do, however, normally contain melatonin and receptors for this indole (Vacas *et al.*, 1991; Morera and Abreu, 2005). Also, hemolysis and elevated liver enzymes are frequently a result of oxidative damage to the respective cells (Pacini *et al.*, 2011). This being the case, melatonin, due to its antioxidant properties, may be beneficial in this condition. Finally, melatonin was observed to have anti-epileptic actions in humans (Molino-Carballo *et al.*, 1997; Sanchez-Barcelo *et al.*, 2011). This suggests that melatonin may reduce the likelihood of pre-eclampsia progressing to eclampsia.

Lipopolysaccharide (LPS) from the cell membranes of Gram-negative bacteria is a commonly used toxin in experimental situations to promote free radical-mediated oxidative damage. In a comprehensive molecular study designed to test whether exogenously administered melatonin would combat oxidative destruction at the level of the placenta, Wang et al. (2011) treated mice at 15 days of pregnancy with 300 μ g/kg LPS without or with concurrent melatonin (5 mg/kg) treatment. Virtually every placental change induced by LPS, including depressed glutathione, elevated inducible nitric oxide synthase (iNOS), enhanced levels of 3-nitrotyrosine residues, reduced GRP78 expression, elevated elF2 α and JNK phosphorylation and increased CHOP expression, was negated in mice treated with melatonin. The results of this thorough study strongly emphasize the potentially important role that endogenously produced placental melatonin could play in preserving homeostasis in this pregnancy-critical organ.

Melatonin not only protects the placenta from LPS-mediated oxidative toxicity but also, likewise, reduces the damage inflicted by LPS on the fetus. When mice were treated (on gravid Days 15-17) with LPS, the toxin increased intrauterine fetal death, caused intrauterine growth retardation (IUGR) and induced biochemical changes consistent with elevated oxidative stress in both the mothers and the fetuses (Chen *et al.*, 2006). As in other experiments of this type, melatonin was effective in eliminating almost all the changes resulting from LPS toxicity, although it did not totally restore the reduced fetal weights of the recovered fetuses.

Other actions of melatonin at the placental level also document that the indole enhances the total antioxidative capacity of this organ. When rats that were undernourished (35% food reduction) from gestation days 15–20 were given melatonin in their drinking water, the relative expression of the antioxidative enzymes manganese superoxide dismutase and catalase were highly significantly elevated over those in the underfed animals not treated with melatonin (Richter et al., 2009). While a similar stimulation of glutathione peroxidase expression was not observed, the findings do show the likely value of melatonin in protecting the placenta from molecular mutilation by free radicals. The enzyme increases were accompanied by an improvement in the growth of the placenta (reflected in the fetal weight to placental weight ratio), which benefitted the fetuses since mean fetal bodyweights were also elevated. Thus in this case, melatonin overcame IUGR which was a result of undernourishment, a procedure believed to involve excessive free radical generation (Franco et al., 2006).

Melatonin in relation to parturition

The processes of labor and delivery of a child involve increasingly frequent and robust contractions of the myometrium accompanied by cervical effacement and expulsion of the fetus. While these events obviously occur at any time during the day or night, there are reports noting that the onset of human term labor more commonly takes place in the late night and early morning hours (Glattre and Bjerkedal, 1983; Cagnacci and Soldani, 1998; Lindow et al., 2000). Similar claims have been made relative to preterm labor (lams *et al.*, 2002), at least after 28 weeks of gestation (Vatish *et al.*, 2010). During evolution there were likely selective survival advantages for both the mother and the newborn to enter parturition at night since predation pressure was lowest at that time. As in the human, similar 24-h circadian variations of birth are seen in other species (Lincoln and Porter, 1976; Zahn and Hattensperger, 1993; Farber *et al.*, 1997).

The circadian signals that drive the nocturnal changes that culminate in the delivery of offspring in late term human pregnancy are only beginning to be unraveled (Olcese, 2012; Olcese et al., 2013). Moreover, the circadian mechanisms related to night-time parturition are probably breaking down because of the widespread misuse of light at night. Throughout evolution where the regularly recurring light:dark cycles were undisturbed, internal rhythms were more strongly coupled to the photoperiodic environment (Golombek et al., 2013). With the advent of the developments in artificial lighting, the natural environmental light/dark cycles are being markedly subverted (witness, for example, the light environments in a hospital setting) and normally entrained circadian rhythms are likewise under increasingly greater pressure. The dysynchronization of circadian rhythms, also referred to as chronodisruption (Erren and Reiter, 2009; 2013), due to perturbed or unusual light/dark cycles generally negatively influence physiological and metabolic processes and sometimes lead to disease (Blask et al., 2011; Bass, 2012; Münch and Bromundt, 2012; Golombek et al., 2013).

Within the last decade, melatonin, which exhibits a pronounced circadian rhythm in both the pineal gland and the circulation of all mammals (Vaughan et al., 1976; Panke et al., 1979; Reiter, 1986; Stehle et al., 2011), has been shown to modulate uterine physiology. With the aid of receptor autoradiography and radioreceptor assays, Schlabritz-Loutsevitch et al. (2003) identified specific high-affinity, G-protein coupled, melatonin-binding sites on the membranes of uterine myometrial cells obtained from both non-pregnant and pregnant humans. These findings are consistent with the presence of the well-defined 7-transmembrane melatonin receptors typical of many cells (Stankov and Reiter, 1990; Dubocovich and Markowska, 2005; Slominski et al., 2012). Moreover, this same group (Schlabritz-Loutsevitch et al., 2003) reported that the accumulation of cyclic adenosine monophosphate (cAMP) in cultured primary myocytes obtained from the human uterus was not suppressed by melatonin after treating the cells with forskolin. This showed that the common G_i protein coupling of the uterine melatonin receptors to adenylyl cyclase was not essential in these cells (Dubocovich and Markowska, 2005). Interestingly, however, it was found that melatonin activates the same intracellular signaling pathway as does oxytocin in uterine myometrial cells. These intracellular events include stimulation of phospholipase C, protein kinase C and myosin light chain kinase (Sharkey et al., 2009, 2010). This being the case, oxytocin and melatonin may act synergistically to promote and sustain strong uterine contractility which aids in the delivery of the fetus. This interaction between oxytocin and melatonin, which is normally elevated during darkness, could explain the higher night-time delivery of offspring (Lincoln and Porter, 1976; Zahn and Hattensperger, 1993; Farber et al., 1997). That melatonin actually sensitizes the human uterus to oxytocin is supported by the finding that cultured myometrial cells treated with a concentration of oxytocin that by itself did not induce contractions undergo powerful contractions when melatonin is added with the low oxytocin concentration (Sharkey et al., 2009, 2010).

An examination of the potential mechanisms involved in the oxytocin/ melatonin synergy showed that melatonin also enhances gap junction activity and mRNA as well as protein expression for connexin 43, a gap junction protein common to many cells, including uterine myocytes. Thus, melatonin seems to assist in the co-ordination of uterine muscle cells during labor to intensify contractile strength (Olcese *et al.*, 2013). Further examination of myometrial cells collected during labor revealed elevated 2-1¹²⁵-melatonin binding compared with smooth muscle cells harvested from the non-laboring uterus. Since 2-1¹²⁵-melatonin binds to both the MT I and MT2 membrane receptors, the obvious implication is that these receptors are more abundant in the uterus during labor than during a more quiescent phase.

Preliminary studies recently performed in humans support a significant role for melatonin in aiding in the delivery of the fetus by enhancing the strength of uterine contractions (Olcese et al., 2013). Pregnant volunteers (>38 weeks gestation) were monitored continuously from 19:00-07:00 h under dim white light as to the frequency and strength of their uterine contractions. When these females were exposed to a 10 000 lux full spectrum light at 23:00 h for 1 h, the strength of their uterine contractions was diminished during the remainder of the night. This light intensity is known to suppress endogenous circulating melatonin levels (Lewy et al., 1980; Revell and Skene, 2007), although in this study melatonin was not measured. The regularity and the strength of the uterine contractions were monitored by an experienced obstetrical nurse but the study was not blinded. The results, however, generally support the idea that the nocturnal rise in melatonin contributes to the increased night-time labor onset and delivery of the offspring due to its synergistic interaction with oxytocin.

The results summarized above are intriguing in view of what is known about the melatonin rhythm during pregnancy. The amplitude of the nocturnal melatonin peak becomes steadily higher in late pregnancy such that, near term, the night-time melatonin levels are higher than those measured at any other time (Fig. 7) (Nakamura *et al.*, 2001; Tamura *et al.*, 2008a) thereby enhancing the likelihood of its synergy with oxytocin. This rise may be an evolutionary adaptation to enhance uterine contractions near the time when delivery should occur. Immediately after delivery, nocturnal melatonin levels in the mother return to values typical of those in non-pregnant women (Tamura *et al.*, 2008a).

Several conclusions and/or recommendations follow from the data summarized: (i) considering the increasingly widespread use of manufactured light, to the point where darkness in urban areas is disappearing and humans are becoming progressively more melatonin deficient, it seems likely that chronodisruption and the loss of melatonin will remain a continuing problem; (ii) since melatonin seems to synergize with oxytocin to strengthen uterine contractions, when spontaneous labor begins melatonin could be given as a supplement to augment uterine contractions, to reduce the duration of labor and make delivery easier. Parenthetically, melatonin also may be useful during delivery since it has been shown to alleviate pain (Gitto et al., 2012; Srinivasan et al., 2012). (iii) When labor, especially during the day, is induced using oxytocin, it could be given with melatonin to intensify the strength of uterine contractions; (iv) women who have a history of early delivery (e.g. before 32 weeks of gestation) should probably avoid melatonin use during pregnancy and (v) throughout pregnancy, but especially during the last trimester, women should maintain a regular light/dark cycle, without interrupting the period of darkness with even brief periods of light, for the purpose of stabilizing

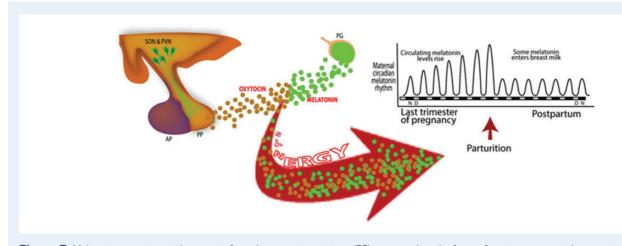


Figure 7 Melatonin synergizes with oxytocin from the posterior pituitary (PP) to strengthen the force of uterine myometrial contractions during labor. Since maximal melatonin levels occur at night in darkness, its synergistic actions with oxytocin may explain the elevated frequency of labor onset at night. In developed countries, the widespread use of manufactured light at night alters circadian rhythms and suppresses the nocturnal production of melatonin thereby likely reducing nighttime labor onset and spreading this process throughout the 24-h period. AP, anterior pituitary; PG, pineal gland; PVN, paraventricular nucleus; SON, supraoptic nucleus.

their circadian rhythms and maximizing the nocturnal melatonin rise, both of which influence the maturation of the fetal circadian system.

Conclusion

Both melatonin and circadian rhythms are influential in determining optimal reproductive physiology during pregnancy. The discoveries related to melatonin production in several peripheral reproductive organs suggest that this indole has several important functions in these tissues, one of which very likely relates to its ability to obviate oxidative/nitrosative stress and placental dysfunction. The production of this highly effective antioxidant in the placenta may be of special significance since it would likely protect against conditions that involve less than optimal placentation, a situation that leads to either prolonged or intermittent hypoxia/reoxygenation and elevated free radical generation. Moreover, pre-eclampsia, which many researchers agree involves exaggerated free radical generation that leads to the manifestation of the signs of this condition, also may be ameliorated by this indoleamine. Given that melatonin readily crosses the placenta it also protects the fetus from oxidative/nitrosative stress. Considering the uncommonly low, or lack of, toxicity of melatonin, including during pregnancy (Jahnke et al., 1999), melatonin may prove to be highly valuable in optimizing the physiology of the reproductive system not only during pregnancy but also at other times (Reiter et al., 2013a). Certainly, it is the hope that research in this area will flourish during the next decade.

Circadian rhythms of the mother are seemingly highly important in programming the fetal clock, either via the melatonin signal and/or by other means. This being the case, it is imperative that during pregnancy, and perhaps especially during the last trimester, pending mothers maintain a regular sleep/wake and light:dark cycle. The light:dark cycle is especially important because the photic environment is an important impeller of the maternal master biological clock, the SCN, which in turn directly or indirectly provides circadian information to the fetus. Thus, shift work and unusual exposure to bright light at night should be avoided since they cause chronodisruption and melatonin suppression. There is a large amount of experimental and clinical data showing that disturbances caused by these processes have untoward consequences for the fetus.

Authors' roles

R.J.R. performed literature searches, had critical discussion of the data with each of the co-workers and wrote the preliminary and final drafts of the paper. D.X.T. performed literature searches, analyzed and discussed data and read drafts of the paper and made suggestions for changes/additions. A.K. and S.A.R.-C. prepared figures, discussed data and read all drafts of the paper.

Funding

No funding was obtained from any source to support this work.

Conflict of interest

The authors declare no conflict of interest.

References

- Abildgaard U, Heimdal K. Pathogenesis of the syndrome of hemolysis, elevated liver enzymes, and low platelet county (HELLP): a review. *Eur J Obstet Gynecol Reprod Biol* 2013;**166**:117–123.
- Acuna-Castroviejo D, Escames G, Carazo A, Leon J, Khaldy H, Reiter RJ. Melatonin, mitochondrial homeostasis and mitochondrial-related diseases. *Curr Top Med Chem* 2002;2:133–151.
- Acuna-Castroviejo D, Lopez LC, Escames G, Lopez A, Garcia JA, Reiter RJ. Melatonin-mitochondria interplay in health and disease. *Curr Top Med Chem* 2011; 11:221–240.
- Allegra M, Reiter RJ, Tan DX, Gentile C, Tesoriere L, Livrea MA. The chemistry of melatonin's interaction with reactive species. *J Pineal Res* 2003;**34**:1–10.

- Arangino S, Cagnacci A, Angiolucci M, Vacca AM, Longu G, Volpe A, Melis GB. Effects of melatonin on vascular reactivity, catecholamine levels, and blood pressure in healthy men. Am J Cardiol 1999;83:1417–1419.
- Axelrod J, Wurtman RJ, Winget CM. Melatonin synthesis in the hen pineal gland and its control by light. *Nature* 1964;**201**:1134.
- Axelrod J, Wurtman RJ, Snyder SH. Control of hydroxyindole-O-methyltransferase activity in the rat pineal gland by environmental lighting. J Biol Chem 1965; 240:949–954.
- Barlow-Walden LR, Reiter RJ, Abe M, Cablos M, Menendez-Pelaez A, Chen LD, Poeggeler B. Melatonin stimulates brain glutathione peroxidase activity. *Neurochem Int* 1995;**26**:497–502.
- Bass J. Circadian topology of metabolism. Nature 2012;491:348-356.
- Bass J, Takahashi JS. Circadian integration of metabolism and energetics. *Science* 2010; **37**:1349–1354.
- Bauer J, Janecke A, Gerss J, Masjosthusmann K, Werner C, Hoffmann G. Circadian variation in oxygen consumption in preterm infants. J Perinat Med 2009;37:413–417.
- Behrman HR, Kodaman PH, Preston SL, Gao S. Oxidative stress and the ovary. J Soc Gynecol Investig 2001;**8**:S40–S42.
- Bellavia SL, Carpentieri AR, Vaque AM, Macchione AF, Vermouth NT. Pup circadian rhythm entrainment-effect of maternal ganglionectomy or pinealectomy. *Physiol Behav* 2006;**89**:342–349.
- Blask DE, Hill SM, Dauchy RT, Xiang S, Yuan L, Duplessis T, Mao L, Dauchy E, Saver LA. Circadian regulation of molecular, dietary and metabolic signaling mechanisms of human breast cancer growth by the nocturnal melatonin signal and the consequences of its disruption by light at night. J Pineal Res 2011;**51**:259–269.
- Brainard GC, Richardson BA, King TS, Matthews SA, Reiter RJ. The suppression of pineal melatonin content and N-acetyltransferase activity by different light irradiances in the syrian hamster: a dose–response relationship. *Endocrinology* 1983;113:293–298.
- Brainard GC, Richardson BA, King TS, Reiter RJ. Influence of different light spectra on the suppression of pineal melatonin content in the Syrian hamster. *Brain Res* 1984; 294:333–339.
- Brannstrom M, Norman RJ. Involvement of leukocytes and cytokines in the ovulatory process and corpus luteum function. *Hum Reprod* 1993;8:1762–1775.
- Briceno-Perez C, Briceno-Sanabria L, Vigil-De Garcia P. Prediction and preeclampsia. Hyperten Pregnancy 2009;28:138–155.
- Brzezinski A, Seibel MM, Lynch HJ, Deng MH, Wurtman RJ. Melatonin in human preovulatory follicular fluid. *J Clin Endocrinol Metab* 1987;**64**:864–867.
- Cagnacci A, Soldani R, Melis GB, Volpe A. Diurnal rhythms of labor and delivery in women: modulation by parity and reasons. *Am J Obst Gynecol* 1998;**178**(1 Pt 1):140–145.
- Cardinali DP, Srinivasan V, Brzezinski A, Brown GM. Melatonin and its analogs in insomnia and depression. J Pineal Res 2012;52:365–375.
- Carrillo-Vico A, Guerrero JM, Lardone PJ, Reiter RJ. A review of the multiple actions of melatonin on the immune system. *Endocrine* 2005;27:189–200.
- Carter DS, Goldman BD. Antigonadal effect of timed melatonin infusion in pinealectomized male Djungarian hamsters (*Phodopus sungorus sungorus*): duration is the critical parameter. *Endocrinology* 1983;113:1261–1267.
- Chang PJ, Chu LC, Hsieh WS, Chuang YL, Lin SJ, Chen PC. Working hours and risk of gestational hypertension and pre-eclampsia. Occup Med 2010;60:66–71.
- Chaste P, Clement N, Botros HG, Guillaume JL, Konyukh M, Pagan C, Scheid I, Nygren G, Anckarsater H, Rastam M *et al.* Genetic variations of the melatonin pathway in patients with attention-deficit and hyperactivity disorders. *J Pineal Res* 2011;**51**:394–399.
- Chen YH, Xu DX, Wang JP, Wang H, Wei LZ, Sun MF, Wei W. Melatonin protects against lipopolysaccharide-induced intrauterine fetal death and growth retardation in mice. *J Pineal Res* 2006;**40**:40–47.
- Choi J, Park SM, Lee E, Kim JH, Jeong YI, Lee JY, Park SW, Kim HS, Hossein MS, Jeong YW et al. Anti-apoptotic effect of melatonin on preimplantation development of porcine parthenogenetic embryos. *Mol Reprod Dev* 2008; 75:1127–1135.
- Conde-Agudelo A, Romero R, Kusanovic JP, Hassan SS. Supplementation with vitamins C and E during pregnancy for the prevention of preeclampsia and other adverse maternal and perinatal outcomes: a systematic review and meta analysis. *Am J Obstet Gynecol* 2011;**204**:503.e1–503.e12.
- de la Sierra A, Redon J, Banegas JR, Segura J, Parati G, Gorostidi M, de la Cruz JJ, Sobrino J, Llisterri JL, Alonso J *et al.* Prevalence and factors associated with circadian blood pressure patterns in hypertensive patients. *Hypertension* 2009; **53**:466–472.

- de Matos Cavalcante AG, Carvalhedo de Bruin PF, Sales de Bruin VM, Nunes DM, Pereira ED, Cavalcante MM, Andrade GM. Melatonin reduces lung oxidative stress in patients with chronic obstructive pulmonary disease: a randomized, double-blind, placebo-controlled study. J Pineal Res 2012;53:238–244.
- Ditisheim AJ, Dibner C, Phillippe J, Pechere-Bertschi A. Biological rhythms and preeclampsia. *Front Endocrinol* 2013; doi: 10.3389/fendo.2013.00047.
- Dubocovich ML, Markowska M. Functional MTI and MT2 melatonin receptors in mammals. *Endocrine* 2005;**27**:101–110.
- Dumont M, Lanctot V, Cadieux-Viau R, Paquet J. Melatonin production and light exposure of rotating night workers. *Chronobiol Int* 2012;**29**:203–210.
- El-Raey M, Geshi M, Somfai T, Kaneda M, Hirako M, Abdel-Ghaffar AE, Sosa GA, El-Roos ME, Nagai T. Evidence of melatonin synthesis in the cumulus oocyte complexes and it role in embracing oocyte maturation in vitro in cattle. *Mol Reprod Dev* 2011;**78**:250–262.
- Erren TC, Reiter RJ. Defining chronodisruption. J Pineal Res 2009;46:245-247.
- Erren TC, Reiter RJ. Revisiting chronodisruption: when the physiological nexus between internal and external time splits in humans. *Naturwissenschaften* 2013; **100**:291–298.
- Farber DM, Giussani DA, Jenkins SC, Mecenas CA, Winter SA, Wentworth RA, Nathanielsz PW. Timing of the switch from myometrial contractures to contractions in late-gestation pregnant Rhesus monkeys as recorded by myometrial electromyogram during spontaneous term and androstenedioneinduced labor. *Biol Reprod* 1997;56:557–562.
- Fenn AM, Fonken LK, Nelson RJ. Sustained melatonin treatment blocks body mass, pelage, reproductive and fever responses to short day lengths in female Siberian hamsters. J Pineal Res 2011;51:180–186.
- Ferreira DS, Amaral FG, Mesquita CC, Barbosa AP, Lellis-Santos C, Turati AO, Santos LR, Sollon CS, Gomes PR, Faria JA. Maternal melatonin programs the daily pattern of energy metabolism in adult offspring. *PLoS One* 2012;**7**:e38795.
- Franco MC, Akamine EH, Reboucas N, Carvalho MH, Tostes RC, Nigro D, Fortes ZB. Long-term effects of intrauterine malnutrition on vascular function in female offspring: implications of oxidative stress. *Life Sci* 2006;**80**:709–715.
- Fulia F, Gitto E, Cuzzocrea S, Reiter RJ, Dugo L, Gittoi P, Barberi S, Cordaro S, Barberi I. Increased levels of malondialdehyde and nitrite/nitrate in the blood of asphyxiated newborns: reduction by melatonin. J Pineal Res 2001;31:343–349.
- Gaber LW, Spargo BH, Lindheimer MD. Renal pathology in pre-eclampsia. *Baillieres Clin Obstet Gynoecol* 1994;**8**:443–468.
- Galano A, Tan DX, Reiter RJ. Melatonin as a natural ally against oxidative stress: a physiochemical examination. *J Pineal Res* 2011;**51**:1–16.
- Galano A, Tan DX, Reiter RJ. On the free radical scavenging activities of melatonin's metabolites, AFMK and AMK. *J Pineal Res* 2013;**54**:245–257.
- Gillette MU, Tischkau SA. Suprachiasmatic nucleus: the brain's circadian clock. *Recent Prog Horm* Res 1999;**54**:33–58.
- Gitto E, Aversa S, Salpietro CD, Barberi I, Arrigo T, Trimarchi G, Reiter RJ, Pelligrino S. Pain in neonatal intensive care: role of melatonin as an analgesic antioxidant. *J Pineal Res* 2012;**52**:291–295.
- Glattre E, Bjerkedal T. The 24-hour rhythmicity of birth: a population study. Acta Obstet Gynecol Scand 1983;**62**:31–36.
- Golombek DA, Casiraghi LP, Agostino PV, Paladino N, Duhart JM, Plano SA, Chiesa JJ. The times they're a-changing: effects of circadian desynchronization on physiology and disease. J Physiol Paris 2013; 107:310–322.
- Haelterman E, Marcoux S, Croteau A, Dramaix M. Population-based study on occupational risk factors for preeclampsia and gestational hypertension. J Work Environ Health 2007;33:304–317.
- Hardeland R. Melatonin in aging and disease—multiple consequences of reduced secretion, options and limits of treatment. *Aging Dis* 2012;**3**:194–225.
- Hardeland R, Tan DX, Reiter RJ. Kynuramines, metabolites of melatonin and other indoles: the resurrection of an almost forgotten class of biogenic amines. *J Pineal Res* 2009;**47**:109–126.
- Hardeland R, Madrid JA, Tan DX, Reiter RJ. Melatonin, the circadian multioscillator system and health: the need for detailed analyses of peripheral melatonin signaling. *J Pineal Res* 2012;**52**:139–166.
- Heazell AE, Buttle HR, Baker PN, Crocker IP. Altered expression of regulators of caspase activity within trophoblast of normal pregnancies and pregnancies complicated by preeclampsia. *Reprod Sci* 2008; **15**:1034–1043.

Hofman MA. The human circadian clock and aging. Chronobiol Int 2000; 17:245–259.

Hoffman RA, Reiter RJ. Pineal gland: influence on gonads of male hamsters. *Science* 1965;**148**:1609–1611.

- Hoffman RA, Reiter RJ. Influence of compensatory mechanisms and the pineal gland on dark-induced gonadal atrophy in male hamsters. *Nature* 1966;**207**:656–657.
- Hung TH, Burton G. Hypoxia and reoxygenation: a possible mechanism for placental oxidative stress in pre-eclampsia. *Taiwan J Obstet Gynec* 2006;**45**:189–200.
- Huppertz B, Kingdom JCP. Apoptosis in the trophoblast—role of apoptosis in placental morphogenesis. J Soc Gynecol Investig 2004; 11:353–362.
- Iams JD, Newman RB, Thom EA, Goldenberg RL, Mueller-Heubach E, Maowad A, Sibai BM, Caritis SN, Miodovnik M, Paul RH et al. Frequency of uterine contractions and the risk of spontaneous preterm delivery. N Engl | Med 2002;346:250-255.
- Illnerova H, Buresova M, Presl J. Melatonin rhythm in human milk. J Clin Endocrinol Metab 1993;**77**:838–841.
- Itoh MT, Ishizuka B, Kuribayashi Y, Amemiya A, Sumi Y. Melatonin, its precursors, and synthesizing enzyme activities in the human ovary. *Mol Human Reprod* 1999; 5:402–408.
- Jahnke G, Marr M, Myers C, Wilson R, Travlos G, Price C. Maternal and developmental toxicity evaluation of melatonin administered orally to pregnant Sprague–Dawley rats. *Toxicol Sci* 1999;**50**:271–279.
- Jud C, Albrecht U. Circadian rhythms in murine pups develop in the absence of a functional maternal circadian clock. J Biol Rhythms 2006;21:149–154.
- Kaufmann P, Black S, Huppertz B. Endovascular trophoblast invasion: implications for the pathogenesis of intrauterine growth retardation and preeclampsia. *Biol Reprod* 2003;69:1–7.
- Kennaway DJ, Stamp GE, Goble FC. Development of melatonin production in infants and impact of prematurity. J Clin Endocrinol Metab 1992;75:397–399.
- Kennaway DJ, Goble FC, Stamp GE. Factors influencing the development of melatonin rhythmicity in humans. J Clin Endocrinol Metab 1996;81:1525–1532.
- Lanoix D, Ouellette R, Vaillancourt C. Expression of melatoninergic receptors in human placenta choriocarcinoma cell lines. *Hum Reprod* 2006;**21**:1981–1988.
- Lanoix D, Beghdadi H, Lafond J, Vaillancourt C. Human placental trophoblasts synthesize melatonin and express its receptors. J Pineal Res 2008;45:50–60.
- Lanoix D, Lacasse AA, Reiter RJ, Vaillancourt C. Melatonin: the smart killer: the human trophoblast as a model. *Mol Cell Endrocrinol* 2012a;**348**:1–11.
- Lanoix D, Guerin P, Vaillancourt C. Placental melatonin production and melatonin receptor expression are altered in preeclampsia: new insights into the role of this hormone in pregnancy. J Pineal Res 2012b;54:417–425.
- Lanoix D, Lacasse AA, Reiter RJ, Vaillancourt C. Melatonin: the watchdog of villous trophoblast homeostasis against hypoxia/reoxygenation-induced oxidative stress and apoptosis. *Mol Cell Endocrinol* 2013;**38**:35–45.
- Lemley CO, Meyer AM, Camacho LE, Neville TL, Newman DJ, Caton JS, Vonnahme KA. Melatonin supplementation alters uteroplacental hemodynamic and fetal development in an ovine model of intrauterine growth restriction. Am J Physiol Regul Integr Comp Physiol 2012;302:R454–R467.
- Lerner AB, Case JD, Takahashi Y, Mori W. Isolation of melatonin, the pineal gland factor that lightens melanocytes. *J Am Chem Soc* 1958;**80**:2587.
- Lewis MP, Sullivan MH, Elder MG. Regulation of interlookin-1 beta of growth and collagenase production by choriocarcinoma cells. *Placenta* 1994;15:13–20.
- Lewy AJ, Wehr TA, Goodwin FK, Newsome DA, Markey SP. Light suppresses melatonin secretion in humans. *Science* 1980;**210**:1267–1269.
- Li MD, Li CM, Wang Z. The role of circadian clocks in metabolic disease. Yale J Biol Med 2012;85:387–401.
- Lincoln GA, Hazlerigg DG. Mammalian circannual pacemakers. *Soc Reprod Fertil* 2010; **67**(Suppl.):171–186.
- Lincoln DW, Porter DG. Timing of the photoperiod and the hour of delivery in rats. *Nature* 1976;**260**:780–781.
- Lindow SW, Jha RR, Thompson JW. 24-hour rhythm to the onset of preterm labor. Br J Obstet Gynecol 2000; **107**:1145–1148.
- Lowes DA, Webster NR, Murphy MP, Galley HF. Antioxidants that protect mitochondria reduce interleukin-6 and oxidative stress, improve mitochondrial function, and reduce biochemical markers of organ dysfunction in a rat model of acute sepsis. *Br J Anaesth* 2013;**110**:472–480.
- Lucas RJ. Mammalian inner retinal photoreception. Curr Biol 2013;23:R125-R133.
- Lydic R, Schoene WC, Czeisler CA, Moore-Ede MC. Suprachiasmatic region of the human hypothalamus: homolog to the primate circadian pacemaker? *Sleep* 1980; 2:355–361.
- Mahoney MM. Shift work, jet lag, and female reproduction. Int J Endocrinol 2010;813764.
- Maldonado MD, Murrillo-Cabezas F, Terron MP, Flores LJ, Tan DX, Manchester LC, Reiter RJ. The potential of melatonin in reducing morbidity-mortality after craniocerebral trauma. J Pineal Res 2007;42:1–11.

- Martin M, Macias M, Escames G, Leon J, Acuna-Castroviejo D. Melatonin but not vitamins C and E maintains glutathione homeostasis in t-butyl hydroperoxideinduced mitochondrial oxidative stress. FASEB J 2000a; 14:1677–1679.
- Martin M, Macias M, Escames G, Reiter RJ, Agapito MT, Ortiz GG, Acuna-Castroviejo D. Melatonin-induced increased activity of the respiratory chain complexes I and IV can prevent mitochondrial damage induced by ruthenium red in vivo. *J Pineal Res* 2000b;**28**:242–248.
- Mason CA, Lincoln DW. Visualization of the retino-hypothalamic projection in the rat by cobalt precipitation. *Cell Tiss Res* 1976;**168**:117–131.
- Mauriz JL, Collado PS, Veneroso C, Reiter RJ, Gonzalez-Gallego J. A review of the molecular aspects of melatonin's anti-inflammatory actions: recent insights and new perspectives. J Pineal Res 2013;54:1–14.
- Menaker M, Murphy ZC, Sellix MT. Central control of peripheral oscillators. *Curr Opin Neurobiol* 2013;23:741–746.
- Menendez-Pelaez A, Reiter RJ. Distribution of melatonin in mammalian tissue: the relative importance of nuclear verses cytosolic localization. *J Pineal Res* 1993; **15**:59–69.
- Meng Y, He Z, Yin J, Zhang Y, Zhang T. Quantitative calculation of human melatonin suppression induced by inappropriate light at night. *Med Biol Eng Comput* 2011; **49**:1083–1088.
- Milczarek R, Hallmann A, Sokolowska E, Kaletha K, Klimek J. Melatonin enhances antioxidant action of alpha-tocopherol and ascorbate against NADPH- and iron-dependent lipid peroxidation in human placental mitochondria. *J Pineal Res* 2010;49:149–155.
- Mogler RKH. Das Endokrine System des Syrischen Goldhamsters unter natürlichen Winterschlaf. Z Morphol Oekal Tiere 1958;**47**:267–308.
- Molino-Carballo A, Munoz-Hoyos A, Reiter RJ, Sanchez-Forte M, Moreno-Madrid F, Rufo-Campos M, Molino-Font JA, Acuna-Castroviejo D. Utility of high doses of melatonin as adjunctive anticonvulsant therapy in a child with severe myoclonic epilepsy: two years' experience. J Pineal Res 1997;23:97–105.
- Morera AL, Abreu P. Existence of melatonin in human platelets. *J Pineal Res* 2005; **39**:432–433.
- Mosca L, Benjamin EJ, Berra K, Bezanson JL, Dolor RJ, Lloyd-Jones DM, Newby LK, Pina IL, Roger VL, Shawl J et al. Effectiveness-based guidelines for the prevention of cardiovascular disease in women—2011 update: a guideline from the American Heart Association. J Am Coll Cardiol 2011;57:1404–1423.
- Münch M, Bromundt V. Light and chronobiology: implications for health and disease. Dialogues Clin Neurosci 2012; 14:448–453.
- Myatt L, Rosenfield RB, Eis AL, Brockman DE, Greer I, Lyall F. Nitrotyrosine residues in placenta. Evidence of peroxynitrite formation and action. *Hypertension* 1996; 28:488–493.
- Nagai R, Watanabe K, Wakatsuki A, Hamada F, Shinohara K, Hayashi Y, Imamura R, Fukaya T. Melatonin preserves fetal growth in rats by protecting against ischemia/ reperfusion-induced oxidative/nitrosative mitochondrial damage in the placenta. *J Pineal Res* 2008;**45**:271–276.
- Nakamura Y, Tamura H, Kashida S, Takayama H, Yamagata Y, Karube A, Sugino N, Kato H. Changes of serum melatonin level and its relationship to feto-placental unit. J Pineal Res 2001;**30**:29–33.
- Nakamura Y, Tamura H, Takayama H, Kato H. Increased endogenous level of melatonin in preovulatory human follicles does not directly influence progesterone production. *Fertil Steril* 2003;**80**:1012–1016.
- Nelissen EC, van Montfoort AP, Dumoulin JC, Evers JL. Epigenetics and the placenta. Hum Reprod Update 2011;17:397–417.
- Novakova M, Sladek M, Sumova A. Exposure of pregnant rats to restricted feeding schedule synchronizes the SCN clocks of their fetuses under constant light but not under a light:dark regime. *J Biol Rhythms* 2010;**25**:350–360.
- Obayashi K, Saeki K, Iwamoto J, Okamoto N, Tomioka K, Nezu S, Ikada Y, Kurumatani N. Nocturnal urinary melatonin excretion is associated with non-dipper pattern in elderly hypertensives. *Hyperten Res* 2013;**36**:736–740.
- Okatani Y, Okamoto K, Hayashi K, Wakatsuki A, Tamura S, Sagara Y. Matemal-fetal transfer of melatonin in pregnant women near term. J Pineal Res 1999;25:129–134.
- Okatani Y, Wakatsuki A, Schinohara K, Taniguchi K, Fukaya T. Melatonin protects against oxidative mitochondrial damage induced by rat placenta by ischemia and reperfusion. *J Pineal Res* 2001;**31**:173–178.
- Olcese J. Circadian aspects of mammalian parturition: a review. *Mol Cell Endocrinol* 2012;**349**:62–67.
- Olcese J, Lazier S, Paradise C. Melatonin and the circadian timing of human parturition. Reprod Sci 2013;**20**:168–174.

- Ortiz A, Espino J, Bejarano I, Lozano GM, Monllor F, Garcia JF, Pariente JA, Rodriguez A. High endogenous melatonin concentrations enhance sperm quality and short-term in vitro exposure to melatonin improves aspects of sperm motility. *J Pineal Res* 2011; **50**:132–139.
- Pablos MI, Reiter RJ, Ortiz GG, Guerro JM, Agapito MT, Chuang JI, Sewerynek E. Rhythms of glutathione peroxidase and glutathione reductase in brain of chick and their inhibition by light. *Neurochem Int* 1998;**32**:69–75.
- Pacini N, Ferrari A, Menozzi M, Borziani F. Melatonin enhances the in vitro action of cytochalasin B on globular resistance and osmotic fragility of erythrocytes. *Neuro Endocrinol Lett* 2011;**32**:292–300.
- Panke E, Rollag MD, Reiter RJ. Pineal melatonin concentrations in the Syrian hamster. Endocrinology 1979;104:194–197.
- Pevet P, Challet E. Melatonin: both master clock output and internal time-giver in the circadian clocks network. J Physiol (Paris) 2011;105:170–182.
- Pringle KG, Kind KL, Sferruzzi-Perri AN, Thompson JG, Roberts CT. Beyond oxygen: complex regulation and activity of hypoxia inducible factors in pregnancy. *Human Reprod Update* 2010;16:415–431.
- Proietti S, Cucina A, Reiter RJ, Bizzarri M. Molecular mechanisms of melatonin's inhibitory actions on breast cancer. *Cell Mol Life Sci* 2013;**70**:2139–2157.
- Quay WB. Volumetric and cytologic variation in the pineal body of *Peromyscus leucopus* (Rodentia) with respect to sex, captivity and daylength. *J Morphol* 1956;**98**:471–495.
- Quay WB. Circadian rhythm in rat pineal serotonin and its modification by estrous cycle and photoperiod. *Gen Comp Endocrinol* 1963;**3**:473–479.
- Redman CWG. Pre-eclampsia: a multi-stress disorder. *Rev Med Int* 2011;**32**(Suppl. 1):S41–S44.
- Redman CWG, Sargent IL. Placental stress and pre-eclampsia: a revised review. *Placenta* 2009;**30**(Suppl. A):S38–S42.
- Reiter RJ. Pineal control of a seasonal reproductive rhythm in male golden hamsters exposed to natural daylight and temperature. *Endocrinology* 1973;**92**:423–430.
- Reiter RJ. Circannual reproductive rhythms in mammals related to photoperiod and pineal function. *Chronobiologia* 1974;1:365–395.
- Reiter RJ. Normal patterns of melatonin levels in the pineal gland and body fluids of humans and experimental animals. *J Neural Transm* 1986;**35**(Suppl. 21):35–54.
- Reiter RJ. Pineal melatonin: cell biology of its synthesis and of its physiological interactions. *Endocrine Rev* 1991;12:151-180.
- Reiter RJ, Hester RJ. Interrelationships of the pineal gland, the superior cervical ganglia and the photoperiod in the regulation of the endocrine system of hamsters. *Endocrinology* 1966;**79**:1168–1170.
- Reiter RJ, Tan DX. What constitutes a physiological concentration of melatonin? *J Pineal* Res 2003;**34**:79–80.
- Reiter RJ, Blask DE, Johnson LY, Rudeen PK, Vaughan MK, Waring PJ. Melatonin inhibition of reproduction in the male hamsters: its dependency on time of day of administration and on an intact and sympathetically innervated pineal gland. *Neuroendocrinology* 1976;22:107–116.
- Reiter RJ, Paredes SD, Manchester LC, Tan DX. Reducing oxidative/nitrosative stress: a newly-discovered genre for melatonin. *Crit Rev Biochem Mol Biol* 2009a; 44:175–200.
- Reiter RJ, Tan DX, Erren TC, Fuentes-Broto L, Paredes SD. Light-mediated perturbations of circadian timing and cancer risk: a mechanistic analysis. *Integr Cancer Ther* 2009b;8:354–360.
- Reiter RJ, Tan DX, Manchester LC, Paredes SD, Mayo JC, Sainz RM. Melatonin and reproduction revisited. *Biol Reprod* 2009c;81:445–456.
- Reiter RJ, Tan DX, Fuentes-Broto L. Melatonin: a multitasking molecule. *Prog Brain Rec* 2010a; **181**:127–151.
- Reiter RJ, Tan DX, Paredes SD, Fuentes-Broto L. Beneficial effects of melatonin in cardiovascular disease. Ann Med 2010b;42:276–285.
- Reiter RJ, Tan DX, Korkmaz A, Ma S. Obesity and metabolic syndrome: association with chronodisruption, sleep deprivation and melatonin suppression. *Ann Med* 2012a;**44**:564–577.
- Reiter RJ, Tan DX, Madrid JA, Erren TC. When the circadian clock becomes a ticking time bomb. *Chronobiol Int* 2012b;29:1286–1287.
- Reiter RJ, Rosales-Corral SA, Manchester LC, Tan DX. Peripheral reproductive organ health and melatonin: ready for prime time. *Int J Mol Sci* 2013a; **14**:7231–7272.
- Reiter RJ, Tan DX, Rosales-Corral SA, Manchester LC. The universal nature, unequal distribution and antioxidant functions of melatonin and its derivatives. *Mini-Rev Med Chem* 2013b; 13:373–384.
- Reppert SM, Schwartz WJ. Maternal coordination of the fetal biological clock in utero. Science 1983;**220**:969–971.

- Revel FG, Masson-Pevet M, Pevet P, Mikkelsen JD, Simonneaux V. Melatonin controls seasonal breeding by a network of hypothalamic targets. *Neuroendocrinology* 2009; **90**:1–4.
- Revell VL, Skene DJ. Light-induced melatonin suppression in humans with polychromatic and monochromatic light. *Chronobiol Int* 2007;**24**:1125–1137.
- Richards J, Gumz M. Advances in understanding the peripheral circadian clocks. *FASEBJ* 2012;**26**:3602–3613.
- Richter HG, Hansell JA, Raut S, Giussani DA. Melatonin improves placenta efficiency and birth weight and increases the placental expression of antioxidant enzymes in undernourished rats. *J Pineal Res* 2009;**46**:357–364.
- Roberts JM, Hubel CA. The two stage model of preeclampsia: variations on the theme. *Placenta* 2009;**23**(Suppl. A):S43–S48.
- Roberts JM, Taylor RN, Musci TJ, Rodgers GM, Hubel CA, McLaughlin MK. Preeclampsia: an endothelial cell disorder. *AmJ Obst Gynecol* 1989; **161**:1200–1204.
- Rodriguez C, Mayo JC, Sainz RM, Antolini I, Herrera F, Martin V, Reiter RJ. Regulation of antioxidant enzymes: a significant role for melatonin. J Pineal Res 2004;36:1–9.
- Rodriguez C, Martin V, Herrera F, Garcia-Santos G, Rodriguez-Blanco J, Casado-Zapico S, Sanchez-Sanchez AM, Suarez S, Puente-Moncada N, Anitua MJ et al. Mechanisms involved in the pro-apoptotic effect of melatonin in cancer cells. Int J Mol Sci 2013;14:6597–6613.
- Rossi AC, Mullin PM. Prevention of pre-eclampsia with low dose aspirin or vitamins C and E in women at high and low risk: a systemic review with meta-analysis. *EurJ Obstet Gynecol Reprod Biol* 2011; **158**:9–16.
- Röunberg I, Kauppila A, Leppaluoto J, Martikainen H, Vakkuri O. Circadian and seasonal variation in human preovulatory follicular fluid melatonin concentration. *J Clin Endocrinol Metab* 1990;**71**:492–496.
- Ruger M, St Hilaire MA, Brainard GC, Khalsa SB, Kronauer RE, Czeisler CA, Lockley SW. Human phase response curve to a single 6.5 h pulse of short-wavelength light. J Physiol 2013;591:353–363.
- Sainz RM, Mayo JC, Rodriguez C, Tan DX, Lopez-Burillo S, Reiter RJ. Melatonin and cell death: differential actions on apoptosis in normal and cancer cells. *Cell Mol Life Sci* 2003;60:1407–1426.
- Sakaguchi K, Itoh MT, Takahashi N, Tarumi W, Ishizuka B. The rat oocyte synthesizes melatonin. Reprod Fertil Rev 2013;25:674–682.
- Sanchez-Barcelo EJ, Mediavilla MD, Reiter RJ. Clinical uses of melatonin in pediatrics. Int J Pediatr 2011;892624.
- Sand A, Schmidt TM, Kofuji P. Diverse types of ganglion cell photoreceptors in the mammalian retina. *Prog Retin Eye Res* 2011;**51**:17–43.
- Santhi N, Thorne HC, van der Veen DR, Johnsen S, Mills SL, Hommes V, Schlangen LJ, Archer SN, Dijk DJ. The spectral composition of evening light and individual differences in the suppression of melatonin and delay of sleep in humans. *J Pineal Res* 2012;**53**:47–59.
- Schenker S, Yang Y, Perez A, Acuff RV, Papas AM, Henderson G, Lee MP. Antioxidant transport by the human placenta. *Clin Nutr* 1998;**17**:159–167.
- Schlabritz-Loutsevitch N, Hellner N, Middendorf R, Müller D, Olcese J. The human myometrium as a target for melatonin. J Clin Endocrinol Metab 2003;88:908–913.
- Seron-Ferre M, Riffo R, Valenzuela GJ, Germain AM. Twenty-four-hour pattern of cortisol in the human fetus at term. Am J Obst Gynecol 2001;184:1278–1283.
- Seron-Ferre M, Valenzuela GJ, Torres-Farfan C. Circadian clocks during embryonic and fetal development. *Birth Defects Res C Embryo Today* 2007;**81**:204–214.
- Sharkey JT, Puttaramu R, Word RA, Olcese J. Melatonin synergizes with oxytocin to enhance contractility of human myometrial smooth muscle cells. J Clin Endocrinol Metab 2009;94:421–427.
- Sharkey JT, Cable C, Olcese J. Melatonin sensitizes human myometrial cells to oxytocin in a PKCα/ERK-dependent manner. *J Clin Endocrinol Metab* 2010;**95**:2902–2908.
- Siddiqui N, Hladunewich M. Understanding the link between the placenta and future cardiovascular disease. Trends Cardiovasc Med 2011;21:188–193.
- Slominski RM, Reiter RJ, Schlabritz-Loutsevitch N, Ostrom RS, Slominski AT. Melatonin membrane receptors in peripheral tissues: distribution and functions. *Mol Cell Endocrinol* 2012;**351**:152–166.
- Srinivasan V, Lauterbach EC, Ho KY, Acuna-Castroviejo D, Zakaria R, Brzezinski A. Melatonin in antinociception: its therapeutic applications. *Curr Neuropharmacol* 2012;10:167–178.
- Stankov B, Reiter RJ. Melatonin receptors: current status, facts and hypotheses. *Life Sci* 1990;**46**:971–982.
- Stehle JH, Saade A, Rawashdeh O, Ackermann K, Jilg A, Sebesteny T, Maronde E. A survey of molecular details in the human pineal gland in the light of phylogeny, structure, function and chronobiological diseases. J Pineal Res 2011;51:17–43.

Sugino N. Reactive oxygen species in ovarian physiology. *Reprod Med Biol* 2005; **4**:31–44.

- Summa KC, Vitaterna MH, Turek FW. Environment perturbation of the circadian clock disrupts pregnancy in the mouse. *PLoS* 2012;**7**:e637668.
- Swanson LW, Cowan WM. The efferent connections of the suprachiasmatic nucleus of the hypothalamus. *J Comp Neurol* 1975; **160**:1–12.
- Tamarkin L, Westrom WK, Hamill AI, Goldman BD. Effect of melatonin on the reproductive system of male and female Syrian hamsters: diurnal rhythm in sensitivity to melatonin. *Endocrinology* 1976;**99**:1534–1541.
- Tamura H, Nakamura Y, Terron MP, Flores LJ, Manchester LC, Tan DX, Sugino N, Reiter RJ. Melatonin and pregnancy in the human. *Reprod Toxicol* 2008a;25:291–303.
- Tamura H, Takasaki A, Miwa I, Taniguchi K, Maekawa R, Asada H, Taketani T, Matsuoka A, Yamagata Y, Shimamura K *et al.* Oxidative stress impairs oocyte quality and melatonin protects oocytes from free radical damage and improves fertilization rates. *J Pineal Res* 2008b;**44**:280–287.
- Tamura H, Takasaki A, Taketani T, Tanabe M, Kuzuka F, Lee L, Tamura I, Maekawa R, Asada H, Yamagata Y *et al.* Melatonin as a free radical scavenger in the ovarian follicle. *Endocr* J 2013;**60**:1–13.
- Tan DX, Chen LD, Poeggeler B, Manchester LC, Reiter RJ. Melatonin: a potent, endogenous hydroxyl radical scavenger. *EndocrJ* 1993;1:57–60.
- Tan DX, Reiter RJ, Manchester LC, Yan MT, El-Sawi M, Sainz RM, Mayo JC, Kohen R, Allegra M, Hardeland R. Chemical and physical properties and potential mechanisms: melatonin as a broad spectrum antioxidant and free radical scavenger. *Curr Top Med Chem* 2002;2:181–197.
- Tan DX, Manchester LC, Hardeland R, Lopez-Burillo S, Mayo JC, Sainz RM, Reiter RJ. Melatonin: a hormone, a tissue factor, an autocoid, a paracoid, and an antioxidant vitamin. J Pineal Res 2003;34:75–78.
- Tan DX, Manchester LC, Terron MP, Flores LJ, Reiter RJ. One molecule, many derivatives: a never-ending interaction of melatonin with reactive oxygen and nitrogen species? *J Pineal Res* 2007;**42**:28–42.
- Tan DX, Manchester LC, Liu X, Rosales-Corral SA, Acuna-Castroviejo D, Reiter RJ. Mitochondria and chloroplasts as the original sites of melatonin synthesis: a hypothesis related to melatonin's primary function and evolution in eukaryotes. *J Pineal Res* 2013;**54**:127–138.
- Thut G, Miniussi C, Gross J. The functional importance of rhythmic activity in the brain. *Curr Biol* 2012;**22**:R658–R663.
- Tomas SZ, Prusac IK, Roje D, Tadin I. Trophoblast apoptosis in placentas from pregnancies complicated with pre-eclampsia. *Gynecol Obstet Invest* 2011; **71**:250–255.
- Tomas-Zapico C, Coto-Montes A. A proposed mechanism to explain the stimulatory effect of melatonin on antioxidative enzymes. *J Pineal Res* 2005;**39**:99–104.
- Torres-Farfan C, Rocco V, Monso C, Valenzuela FJ, Campino C, Germain A, Torrealba F, Valenzuela GJ, Seron-Ferre M. Maternal melatonin effects on clock gene expression in a nonhuman primate fetus. *Endocrinology* 2006; 147:4618–4626.
- Uguz AC, Cig B, Espino J, Bejarano I, Naziroglu M, Rodriguez AB, Pariente JA. Melatonin potentiates chemotherapy-induced cytotoxicity and apoptosis in pancreatic tumor cells. J Pineal Res 2012;**53**:91–98.

- Urata Y, Honma S, Goto S, Todoroki S, Iida T, Cho S, Honma K, Kondo T. Melatonin induces gamma-glutamylcysteine synthetase mediated by activator protein-1 in human vascular endothelial cells. *Free Radic Bio Med* 1999;**27**:838–847.
- Vacas MI, Del Zar MM, Martinuzzo M, Cardinali DP. Binding sites for [3H]-melatonin in humans platelets. J Pineal Res 1991;11:135–139.
- Vaillancourt C, Lanoix D, Le Bellego F, Daoud G, Lafond J. Involvement of MAPK signaling in human villous trophoblast differentiation. *Mini-Rev Med Chem* 2009; 9:962–973.
- Varcoe TJ, Wight N, Voultsios A, Salkeld MD, Kennaway DJ. Chronic phase shifts of the photoperiod throughout pregnancy programs glucose intolerance and insulin resistance in the rat. *PLoS One* 2011;**6**:e18504.
- Vatish M, Steer PJ, Blanks AM, Horn M, Thorton S. Diurnal variation is lost in preterm deliveries before 28 weeks of gestation. Br J Obstet Gynecol 2010;107:1145–1148.
- Vaughan GM, Pelham RW, Pang SF, Loughlin LL, Wilson KM, Sandock KL, Vaughan MK, Koslow SH, Reiter RJ. Nocturnal elevation of plasma melatonin and urinary 5-hydroxyindoleacetic acid in young men: attempts at modification by brief changes in environmental lighting and sleep and by autonomic drugs. J Clin Endocrinol Metab 1976;42:752–764.
- Venegas C, Garcia JA, Escames G, Ortiz F, Lopez A, Doerrier C, Garcia-Corzo L, Lopez LC, Reiter RJ, Acuna-Castroviejo D. Extrapineal melatonin: analysis of its subcellular distribution and daily fluctuations. *J Pineal Res* 2012;**52**:217–227.
- Waddell BJ, Wharfe MD, Crew RC, Mark PJ. A rhythmic placenta? Circadian variation, clock genes and placental function. *Placenta* 2012;**33**:533–539.
- Wang H, Li L, Zhao M, Zhang ZH, Zhang C, Ji YL, Meng XH, Xu DX. Melatonin alleviates lipopolysaccharide-induced placental cellular stress response in mice. *J Pineal Res* 2011;**50**:418–426.
- Wang J, Xiao X, Zhang Y, Shi D, Chen W, Fu L, Liu L, Xie F, Kang T, Huang W et al. Simultaneous modulation of COX-2, p300, Akt, and Apaf-1 signaling by melatonin to inhibit proliferation and induce apoptosis in breast cancer cells. J Pineal Res 2012;53:77–90.
- Watanabe T, Kojima M, Tomida S, Nakamura TJ, Yamamura T, Nakao N, Yasuo S, Yoshimura T, Ebihara S. Peripheral clock gene expression in CS mice with bimodal locomotor rhythms. *Neurosci Res* 2006;**54**:295–301.
- Wehr TA. The durations of human melatonin secretion and sleep respond to changes in daylength. J Clin Endocrinol Metab 1991;**73**:1276–1280.
- Weinert D. Ontogenetic development of the mammalian circadian system. *Chronobiol* Int 2005;**22**:179–205.
- Weinstein L. Syndrome of hemolysis, elevated liver enzymes, and low platelet count: a severe consequence of hypertension in pregnancy. Am J Obstet Gynecol 1982; 142:159–167.
- Wergeland E, Strand K. Working conditions and prevalence of pre-eclampsia, Norway 1989. Int J Gynecol Obstet 1997;**58**:189–196.
- Wharfe MD, Mark PJ, Waddell BJ. Circadian variation in placental and hepatic clock genes in rat pregnancy. *Endocrinology* 2011;152:3552–3560.
- Zahn V, Hattensperger W. Circadian rhythm of pregnancy contractions. Z Geburtshilfe Perinatol 1993;197:1–10.
- Zhu JL, Hjollund NH, Andersen AM, Olsen J. Shift work, job stress, and late fetal loss: the National Birth Cohort in Denmark. *J Occup Environ Med* 2004;**46**:1144–1149.