

Review Article

Mesenchymal Stem Cells Migration Homing and Tracking

Abhishek Sohni and Catherine M. Verfaillie

Stem Cell Biology and Embryology, Department of Development and Regeneration, KU Leuven, 3000 Leuven, Belgium

Correspondence should be addressed to Abhishek Sohni; abhishek.sohni@med.kuleuven.be and Catherine M. Verfaillie; catherine.verfaillie@med.kuleuven.be

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In this review, we discuss the migration and homing ability of mesenchymal stem cells (MSCs) and MSC-like cells and factors influencing this. We also discuss studies related to the mechanism of migration and homing and the approaches undertaken to enhance it. Finally, we describe the different methods available and frequently used to track and identify the injected cells *in vivo*.

1. Potential of MSC and MSC-Like Cells for Cell-Based Therapies

For stem cells to be used in the clinical setting, they should be safe, that is, do not form tumors, and be readily harvested and/or expanded. Although embryonic stem cells (ESCs) are pluripotent and could be used to replace any tissue, they can form teratomas. Hence, their potential use in cell-based therapies will require that no undifferentiated ESCs persist in the graft. In addition, culture methods for human ESCs are still quite demanding; hence, scaleup is not yet straightforward.

The best-studied transplanted stem cell is the hematopoietic stem cell (HSC) that can be harvested from different sources (bone marrow, blood, and umbilical cord blood) in sufficient numbers for transplantation. HSCs have then also been used for cell-based therapies especially in an allogeneic setting for more than a quarter of a century.

Other adult stem cell populations that are being evaluated clinically are MSCs and multipotent adult progenitor cells (MAPCs; trade name MultiStem) both derived from human postnatal tissue.

MSCs were first described in the 1970s by Friedenstein et al. who described a population of cells derived from bone marrow that had the appearance of fibroblasts and could generate aside from fibroblasts, also adipocytes, chondrocytes, and osteocytes [1, 2]. These cells grew out as colonies and were therefore termed “colony forming units” or CFUs. Later on, Caplan and others termed these cells “mesenchymal

stem cells” (MSCs) [3, 4]. MSCs are classically isolated from bone marrow however; they can be found in multiple tissues, such as adipose tissue, fetal lung, placenta, Wharton’s jelly, and UCB, among others [5–8]. MSCs are characterized as adherent cells with the ability to differentiate into fibroblasts, adipocytes, chondrocytes, osteocytes, and smooth muscle cells apart from supportive hematopoietic “stromal” cells [1, 3, 4, 9] and with a characteristic cell surface antigen profile.

Many MSCs or MSC-like cells with varying differentiation potential have been described and reviewed elsewhere [10]. Although the cell surface repertoire and the gene expression pattern vary among these cells, this is likely a reflection of the tissue of origin or the culture conditions used for maintenance of these cells [11, 12]. A standardized phenotype was proposed for MSCs by the International Society for Cellular Therapies. A typical human (h)MSC should express CD105, CD90, and CD73 but not CD79a, CD45, CD34, CD19, CD14, CD11b, and HLA-DR on its surface [3, 4, 13]. Most hMSCs or hMSC-like adult progenitors give rise to mesoderm derivatives such as fat, bone, and cartilage [9]. Apart from the mesenchymal lineages, MSCs and MSC-like cells such as hMAPC have been reported to also be able to give rise to skeletal myocytes, cardiomyocytes smooth muscle cells, and endothelial cells [11, 14–16] (also reviewed in [10]). Although some studies have suggested that MSCs can give rise to neurons and endodermal progeny [17–19], it remains unclear whether such progeny has all properties of primary neuroectodermal and endodermal cells.

Apart from being able to differentiate to multiple cell types *in vitro* (and *in vivo*), MSCs and MSC-like adult stem cells have extensive immunomodulatory and immunological tolerance inducing characteristics [20–26]. hMSCs that characteristically lack expression of MHC-II, CD40, CD80 and CD86 but express MHC-I present themselves as nonimmunogenic. Although the presence of MHC-I may activate T-cells, due to lack of costimulatory molecules, MSCs fail to elicit an immune response [27]. MSCs also efficiently suppress an immune response by modulating T-cell activation and proliferation [28, 29], either by a direct cell-cell interaction [30] or mediated via soluble factors [28, 31] and this is independent of MHC matching. This immunomodulating effect of MSCs is being explored as adjuvants during allogeneic transplantation to prevent graft-versus-host disease (GVHD) [32, 33] and during organ transplantation to prevent immune rejection [29, 34–36]. In addition, the immunomodulatory characteristics of MSCs are being evaluated in the setting of autoimmune diseases, such as Crohn's Disease, among others [37, 38].

MSCs also produce innumerable growth factors and cytokines, which make them suitable for inducing endogenous repair. For instance, MSCs express bone morphogenic protein(s) (BMPs) which is effective in enhancing cartilage, bone, and tendon repair [39]. Likewise, MSCs produce factors that enhance revascularisation, even if their nature is not understood, and are therefore being evaluated in therapies for ischemic disorders, such as stroke, myocardial infarct, or peripheral arterial disorders. Yet another field of therapeutic applications is grafting MSCs that have been genetically modified to overexpress a protein in diseased tissues due to the genetic mutation of the given factor.

2. Homing of MSCs

The lingering problem in the field of cell-based therapies is the delivery of the cells to the site of injury, a process termed “homing.” As discussed above, the therapeutic efficacy of MSCs is greatly dependent on their ability to produce juxtacrine or paracrine factors that enhance regeneration from endogenous (stem) cells. For juxtacrine effects to be possible, migration of MSCs to the diseased organ/tissue is required. Migration and homing to the tissue of injury is influenced by multiple factors including age and passage number of the cells, culture conditions, and the delivery method, among others. We here provide a review of the literature demonstrating the effect of various factors on migration and homing of MSCs.

2.1. Age, Passage Number, and Dosage of MSCs. It has been shown that with higher passage number, the engraftment efficiency of MSCs decreased. Rombouts et al. had performed a time course experiment, where they showed that freshly isolated MSCs had a better efficiency of homing compared to cultured cells [40]. Moreover, they showed that culture of MSCs for 24 hr decreased the homing efficiency to 10% from 55–65% and to near 0% when cultured for 48 hr. It is well documented that with age, the ability of an organism to repair

and heal goes down which is in part due to decreased potency of resident stem/progenitor cell. Thus, it is possible that *in vitro* multiplication also causes “aging” and hence decreases potency. However, another possibility is that for other stem cells like HSCs, culture alters the expression and function of cell surface ligands required for homing; it will be discussed below.

2.2. Source and Culture Conditions of MSCs. As alluded earlier, MSCs can and have been isolated from multiple different tissues [41] with differences in the phenotype of the cells isolated [42]. These differences are likely in part due to differences in the native microenvironment from where they are isolated [43]. This presents a challenge for the use of MSCs for therapeutic purposes. In order to define an MSC, the Mesenchymal and Tissue Stem Cell Committee of the International Society for Cellular Therapy (ISCT) proposed certain standards to be considered while using human MSCs therapeutically [44]. Apart from the source of the MSCs, culture methods greatly influence MSC characteristics, including their homing potential. As mentioned earlier, freshly isolated MSCs home better than their cultured counterparts [40]. The CXCR4 chemokine receptor that recognizes CXCL12 (also termed SDF-1 α) is highly expressed on bone marrow MSCs, but is lost upon culturing [45, 46]. However, when MSCs are cultured with cytokines (such as HGF, SCF, IL-3, and IL-6) [47], and under hypoxic conditions, CXCR4 expression can be reestablished [48]. Similarly, matrix metalloproteases (MMPs), known to be important in migration of cells, have been demonstrated to play a role in MSC migration [49–51]. Expression of MMPs in MSCs is influenced by factors such as hypoxia [50] and increased culture confluence [49]. Moreover, inflammatory cytokines TGF- β 1, IL-1 β , and TNF- α also enhance migration by upregulation of MMPs (MMPs) [51] affecting homing of MSCs. Hence, culture conditions to which MSCs are exposed play a vital role in their homing ability.

2.3. Delivery Method. The efficacy, bioavailability, and functionality of a pharmacological drug are dependent on the method via which it is being administered. In order to enhance efficacy and availability, the method of administration of MSCs should hence facilitate homing of MSCs to the desired tissue. Intravenous infusion is one of the major routes of administration of MSC [52–55]. When MSCs are infused systemically, they are trapped into capillary beds of various tissues, especially the lungs [52, 56–58]. Therefore, intra-arterial injection of MSCs has been assessed. Delivery of MSCs via the internal carotid artery significantly improved their migration and homing in the injured brain compared with injection via the femoral vein [59]. Similarly, in humans with subacute spinal cord injury (SCI), delivery of MSCs via the vertebral artery leads to a greater functional improvement than when cells were administered via the intravenous route [60]. However, delivery of cells in an artery may lead to “microvascular occlusions” [59]. While to treat myocardial infarction (MI), delivery of bone marrow cells or MSCs directly in the heart or close to the site of injury

enhances the number of cells found in the peri-infarct region [61]. Similarly, direct injection of adipose-derived MSC in damaged skeletal muscle leads to an increase in mass and functional capacity [62].

2.4. Host Receptability-Injury versus Noninjured. MSCs have the luxury of being tolerated by the host immune system due to low immunogenicity as discussed earlier. Their bioavailability and efficacy are dependent on the host pathological condition. During an injury, host cells release different chemo-attractants that have a positive influence on homing of MSCs. This possibly explains the observation that MSCs home better when injected 24 hrs after injury than after 14 days in a myocardial injury model [63]. Many such chemoattractants and the associated receptors on MSCs have been identified. Moreover, MSCs are being genetically engineered to overexpress such receptors to enhance their homing to the damaged tissue [61, 63–66]. Moreover, strategies to precondition the host for better distribution and to prevent injected cells from being entrapped in small vessels especially of the lungs have proven beneficial. One such approach was the pretreatment of host with vasodilator such as sodium nitroprusside (SNP) which resulted in increased MSC passage through the lung microvasculature compared to untreated hosts [58].

3. Mechanism of Homing

Most insights in the mechanisms underlying migration and homing are from studies that evaluated leukocyte migration [67] into inflamed tissues, HSCs [68] the and metastatic cancer cells [69]. A significant body of the literature also exists related to mechanism of MSCs migration towards the target tissue and the role of cell surface receptors and molecules in aiding this migration. The role of activated endothelial cells in migration of MSCs is also being extensively studied. We here describe the factors that aid MSCs in migration and homing to tissue of interest.

3.1. Expression of Receptors and Adhesion Molecules. Similar to leucocytes, MSCs express many receptors and cell adhesion molecules that aid in migration and homing to target tissues. However, the precise mechanisms by which MSCs are recruited are not yet fully understood.

Homing is in a significant part dependent on the chemokine receptor, CXCR4, and its binding partner that was previously characterized in HSC homing, that is, stromal-derived factor-1 CXCL12 [61, 64, 70–72]. Wynn et al. demonstrated that CXCR4 is present on a subpopulation of MSCs, which aid in CXCL12-dependent migration and homing [45]. Aside from CXCR4, freshly isolated BM MSCs and cultured MSCs also express CCR1, CCR4, CCR7, CCR10, CCR9, CXCR5, and CXCR6 [72, 73] which are also involved in MSC migration

Integrins are another family of cell surface molecules involved in migration of variety of cells and are expressed on adipose-derived MSC-like cells [74]. Neutralizing antibodies against integrins, more specifically the integrin-beta1

integrin, but not integrin-alpha4, inhibit, MSC homing to infarcted myocardium [75]. However, other studies have shown that integrin-alpha4 plays a role in MSC migration [76]. Interestingly, integrin ligands such as VCAM and ICAM are also expressed on MSCs [77].

3.2. Interaction with Endothelial Cells. Migration and homing requires that cells can attach to and migrate between endothelial cells (ECs) to enter the target tissue. While it is well established that leukocytes attach to ECs, roll over the ECs, and then transmigrate between ECs, how MSCs interact with ECs is not well understood. MSCs express molecules as a number of adhesion molecules, including selectins and integrins, involved in these steps. Ruster et al. using a parallel plate flow chamber, demonstrated that MSCs like HSCs bind to ECs derived from human umbilical cord vein (HUVECs) [76]. The binding was enhanced when ECs were activated by TNF- α [76]. The cells migrate by extending podia followed by rolling and adhesion on the EC. They further demonstrated that the binding and rolling of MSCs were mediated by the P-selectin adhesion molecule, whereas migration involved the binding of VLA-4 (or integrin-beta1 & integrin-alpha4 dimer) on MSC with VCAM-1 found on ECs [76]. Steingen et al. found a similar mechanism by which VLA-4/VCAM-1 is required for transendothelial migration. In addition, migration was dependent on the phenotype of the vascular bed [78] and also involved proteolytic enzymes [78]. This is consistent with the studies from De Becker et al. and others demonstrating a role of the MMP class of proteolytic enzymes in MSC homing and migration [49, 51]. MMP-2 belongs to the gelatinase class of proteolytic enzymes that cleave gelatin and collagen-IV, the two major constituents of basement membrane.

4. Approaches to Improve Homing

For MSCs to home and target a specific tissue, they require the right combination of signaling molecules from the injured tissue and the corresponding receptors on MSC. The expression of chemokine receptors on MSCs is influenced by many factors. Although freshly isolated MSCs home better, only limited numbers of cells can be isolated. Therefore, approaches to expand MSCs while retaining expression of receptors needed for efficient homing are being developed. For instance, pretreatment of cultured MSCs with cytokines (such as IL-6, HGF, etc.) increased expression of chemokine receptors (CXCR4) and improved their migration both *in vivo* [47] and *in vitro* [79]. Likewise, IL-1 β pre-treatment enhanced the efficacy by MSCs homing in a colitis model [80].

Other approaches include changing culture conditions and coculture of MSC. Hung et al. demonstrated that short-term exposure of MSCs to hypoxia leads to increased expression of chemokine receptors (CX3CR1 and CXCR4) that may aid engraftment *in vivo* [81]. A similar increase in chemokine receptor (CXCR4) apart from cell proliferation-associated cyclin (cyclin D1, D3) expression was observed when human umbilical cord MSCs were cocultured with

Sertoli cells [82]. Yet another approach is the use of genetically engineered MSCs that overexpress chemokine receptors such as CXCR4 and integrin- α 4 to influence their homing ability. Kumar et al. transduced MSCs with an adenovirus encoding integrin- α 4, which enhanced their ability to home to bone [83]. A similar approach was taken to overexpress CXCR4 in MSCs to enhance their homing ability and to improve recovery after myocardial infarction [84]. Compared with untransduced MSCs, CXCR4 overexpressing MSCs resulted in a decrease in anterior wall thinning, and left ventricular chamber dimensions were better maintained and remodeling was observed [84]. Although these genetically modified MSCs may not yet be available for therapeutic use in humans, pre-treatment approach may well be applicable.

5. Tracking Mesenchymal Stem Cells *In Vivo*

As homing of MSCs is inefficient and many MSCs are trapped in the lung following systemic administration, it is imperative that we can trace the fate of the injected cells. One classical method to label cells is with retroviral vectors to express fluorescent proteins, which has been helpful in gaining insights in MSC homing and engraftment [85]. However, the visualization of cells that homed in different organs requires sacrificing of the animal, as the tissue penetrability of fluorescence is limited. Hence, more advanced techniques to track the injected cells *in vivo*, such as bioluminescence imaging (BLI), single-photon emission CT (SPECT), positron emission tomography (PET), multiple photon microscopy, and magnetic resonance imaging (MRI), are being employed. The noninvasive cellular imaging allows for tracking the injected cells in multiple tissues and over time.

To trace cells by MRI requires labeling of the cells with contrast reagents in order for the cells to be visualized. Cells can be labeled with contrast agents: either positive contrast agents used in T_1 -weighted MRI such as lanthanide chelates [86] or Mn-containing compounds [87, 88]; or negative contrast agents, such as superparamagnetic iron oxide (SPIO) [89–92], ultra-small superparamagnetic iron oxide (USPIO) particles [90, 92, 93], or micron-sized iron oxide particles [94, 95], that are highly sensitive and have a dominant effect on the T_2/T_2^* relaxation times, causing negative contrast enhancement in the regions of interest. For use with stem cells, the role of these agents on cell potency and function needs to be evaluated. Crabbe et al. evaluated the effect of different MRI contrast reagents on the cellular function of embryonic and postnatal stem cells including MSC in an ischemic stroke model [96]. Differences were observed in terms of size, densities, and number of inclusions among the different reagents tested. Moreover, the labeling did not interfere with the migratory capacities of these cells *in vivo* [96]. One drawback of MRI-based cell tracking is that whole body scans are difficult to achieve, and, hence, determination of where the labeled cells traffic to other than the tissue that is damaged, is difficult. A second drawback is that the iron oxide particles are retained in a tissue even if the grafted (stem) cell dies, hence, leading to false positive signals. Yet another problem as shown by Vandeputte et al. is that a pronounced

hypointense signal intensity on 3D T_2^* MR images can be seen spontaneously, corresponding to damaged blood vessels and inflammatory cells, thus, warranting caution [97].

PET imaging relies on the activation of a tracer dye by a protein such as herpes simplex virus type 1 thymidine kinase (HSV1-tk) or varicella zoster virus thymidine kinase (VZV-tk), expressed by genetically engineering into the injected cells [98, 99]. Two main tracer classes used as probes for HSV1-tk substrate are pyrimidine nucleoside derivatives and acycloguanosine derivatives [100]. PET/SPECT visualizes the emission from a tracer dye that can be toxic and interferes with cellular function. Alternatively, cells can be directly labeled with tracer dyes such as ^{18}F -FDG. Wolfs et al. demonstrated that ^{18}F -FDG, a positron emitting glucose analogue, can be easily taken up by MSC and MSC-like cells without interfering with cellular functions [101]. Further, the presence of this analogue did not significantly affect the viability, proliferation, differentiation, and migratory capacities of these cells [101]. However, this only allows following the cells for a short period of time, as the tracer decays over 109 minutes [102].

BLI requires the expression of a bioluminescence protein that can be visualized and is hindered less by tissue mass unlike fluorescent proteins. With the availability of instrumentation for visualization, the BLI method has been quite useful in tracing cells in mouse models. Bioluminescence can be obtained using proteins from the luciferase family (firefly or *Renilla*), which again needs to be genetically engineered in the injected MSCs [103, 104]. In a study evaluating the immunogenicity of such xenoproteins in genetically engineered MSCs, Bergwerf et al. found no immunological response when such MSCs were injected in the brain. In contrast, they found reporter gene-specific immune-reactive T-cell responses when they were injected in the muscle [105].

Kraitchman et al., using dual labeled cells (tracer and contrast reagent), were able to follow the cells for up to a week in an MI mouse model [106]. Their tracing showed that the cells initially home to the lungs followed by redistribution to nontarget organs within 48 hrs. MSCs were also found at the site of infarct up to one week after injection [106]. However, the cells were unable to be located using MRI, rather, they were traceable using high sensitive SPECT [106].

Many such reagents and cell tracing methodologies have been developed and evaluated for stem and progenitor cells. The efficacy, toxicity and resolution are the main factors that determine the choice of imaging technique.

6. Conclusion

Postnatal MSC or MSC-like cells are currently the primary source of stem cells that have found clinical relevance. Embryo-derived stem cells such as ESC although have a greater differentiation potential, they suffer from their ability to induce teratomas *in vivo*; hence, it has been difficult to translate their clinical use. As alluded here, a great deal of work has been done to harness the potential of these adult stem cells for the treatment of patients. MSC and the likes are already undergoing clinical trials for use in patients

especially for their immunomodulatory features. However, their heterogeneity and off-target homing especially lodging in the lungs impede the clinical use of MSC and MSC-like cells. Owing to this, a large number of cells are required to obtain desired effect at the target organ(s). Different methods as discussed above, such as targeted delivery, cytokine pre-treatment, and assisted homing, are being used to circumvent such impedances.

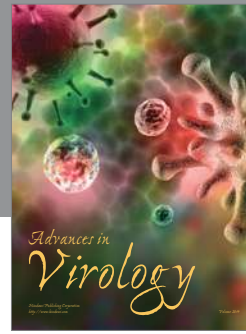
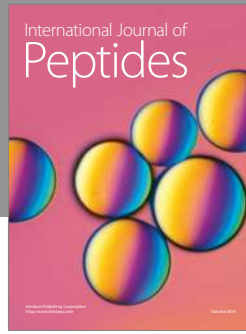
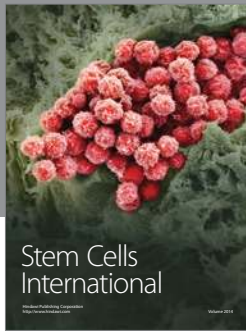
References

- [1] A. J. Friedenstein, R. K. Chailakhyan, and N. V. Latsinik, "Stromal cells responsible for transferring the microenvironment of the hemopoietic tissues. Cloning in vitro and retransplantation in vivo," *Transplantation*, vol. 17, no. 4, pp. 331–340, 1974.
- [2] A. J. Friedenstein, U. F. Deriglasova, and N. N. Kulagina, "Precursors for fibroblasts in different populations of hematopoietic cells as detected by the in vitro colony assay method," *Experimental Hematology*, vol. 2, no. 2, pp. 83–92, 1974.
- [3] A. I. Caplan, "Mesenchymal stem cells," *Journal of Orthopaedic Research*, vol. 9, no. 5, pp. 641–650, 1991.
- [4] D. J. Prockop, "Marrow stromal cells as stem cells for non-hematopoietic tissues," *Science*, vol. 276, no. 5309, pp. 71–74, 1997.
- [5] A. Erices, P. Conget, and J. J. Minguell, "Mesenchymal progenitor cells in human umbilical cord blood," *British Journal of Haematology*, vol. 109, no. 1, pp. 235–242, 2000.
- [6] D. A. De Ugarte, K. Morizono, A. Elbarbary et al., "Comparison of multi-lineage cells from human adipose tissue and bone marrow," *Cells Tissues Organs*, vol. 174, no. 3, pp. 101–109, 2003.
- [7] R. Anzalone, M. L. Iacono, S. Corrao et al., "New emerging potentials for human wharton's jelly mesenchymal stem cells: immunological features and hepatocyte-like differentiative capacity," *Stem Cells and Development*, vol. 19, no. 4, pp. 423–438, 2010.
- [8] S. Kern, H. Eichler, J. Stoeve, H. Klüter, and K. Bieback, "Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue," *Stem Cells*, vol. 24, no. 5, pp. 1294–1301, 2006.
- [9] M. F. Pittenger, "Multilineage potential of adult human mesenchymal stem cells," *Science*, vol. 284, no. 5411, pp. 143–147, 1999.
- [10] A. Sohni and C. M. Verfaillie, "Multipotent adult progenitor cells," *Best Practice and Research*, vol. 24, no. 1, pp. 3–11, 2011.
- [11] V. D. Roobrouck, C. C. Carlos Clavel, S. A. Jacobs et al., "Differentiation potential of human postnatal mesenchymal stem cells, mesoangioblasts, and multipotent adult progenitor cells reflected in their transcriptome and partially influenced by the culture conditions," *Stem Cells*, vol. 29, no. 5, pp. 871–882, 2011.
- [12] V. D. Roobrouck, K. Vanuytsel, and C. M. Verfaillie, "Concise review: culture mediated changes in fate and/or potency of stem cells," *Stem Cells*, vol. 29, no. 4, pp. 583–589, 2011.
- [13] B. Sacchetti, A. Funari, S. Michienzi et al., "Self-renewing osteoprogenitors in bone marrow sinusoids can organize a hematopoietic microenvironment," *Cell*, vol. 131, no. 2, pp. 324–336, 2007.
- [14] X. L. Aranguren, A. Luttun, C. Clavel et al., "In vitro and in vivo arterial differentiation of human multipotent adult progenitor cells," *Blood*, vol. 109, no. 6, pp. 2634–2642, 2007.
- [15] M. Kucia, M. Halasa, M. Wysoczynski et al., "Morphological and molecular characterization of novel population of CXCR4⁺ SSEA-4⁺ Oct-4⁺ very small embryonic-like cells purified from human cord blood—preliminary report," *Leukemia*, vol. 21, no. 2, pp. 297–303, 2007.
- [16] G. Kogler, S. Sensken, J. A. Airey et al., "A new human somatic stem cell from placental cord blood with intrinsic pluripotent differentiation potential," *The Journal of Experimental Medicine*, vol. 200, no. 2, pp. 123–135, 2004.
- [17] D. Woodbury, E. J. Schwarz, D. J. Prockop et al., "Adult rat and human bone marrow stromal cells differentiate into neurons," *Journal of Neuroscience Research*, vol. 61, no. 4, pp. 364–370, 2000.
- [18] Y. Sato, H. Araki, J. Kato et al., "Human mesenchymal stem cells xenografted directly to rat liver are differentiated into human hepatocytes without fusion," *Blood*, vol. 106, no. 2, pp. 756–763, 2005.
- [19] K. Timper, D. Seboek, M. Eberhardt et al., "Human adipose tissue-derived mesenchymal stem cells differentiate into insulin, somatostatin, and glucagon expressing cells," *Biochemical and Biophysical Research Communications*, vol. 341, no. 4, pp. 1135–1140, 2006.
- [20] M. Kovacovics-Bankowski, P. R. Streeter, K. A. Mauch et al., "Clinical scale expanded adult pluripotent stem cells prevent graft-versus-host disease," *Cellular Immunology*, vol. 255, no. 1–2, pp. 55–60, 2009.
- [21] W. Van't Hof, N. Mal, Y. Huang et al., "Direct delivery of syngeneic and allogeneic large-scale expanded multipotent adult progenitor cells improves cardiac function after myocardial infarct," *Cytotherapy*, vol. 9, no. 5, pp. 477–487, 2007.
- [22] S. Beyth, Z. Borovsky, D. Mevorach et al., "Human mesenchymal stem cells alter antigen-presenting cell maturation and induce T-cell unresponsiveness," *Blood*, vol. 105, no. 5, pp. 2214–2219, 2005.
- [23] R. Ramasamy, C. K. Tong, H. F. Seow, S. Vidyadaran, and F. Dazzi, "The immunosuppressive effects of human bone marrow-derived mesenchymal stem cells target T cell proliferation but not its effector function," *Cellular Immunology*, vol. 251, no. 2, pp. 131–136, 2008.
- [24] F. Dazzi and F. M. Marelli-Berg, "Mesenchymal stem cells for graft-versus-host disease: close encounters with T cells," *European Journal of Immunology*, vol. 38, no. 6, pp. 1479–1482, 2008.
- [25] X. Jiang, Y. Zhang, B. Liu et al., "Human mesenchymal stem cells inhibit differentiation and function of monocyte-derived dendritic cells," *Blood*, vol. 105, no. 10, pp. 4120–4126, 2005.
- [26] A. Corcione, F. Benvenuto, E. Ferretti et al., "Human mesenchymal stem cells modulate B-cell functions," *Blood*, vol. 107, no. 1, pp. 367–372, 2006.
- [27] W. T. Tse, J. D. Pendleton, W. M. Beyer, M. C. Egalka, and E. C. Guinan, "Suppression of allogeneic T-cell proliferation by human marrow stromal cells: implications in transplantation," *Transplantation*, vol. 75, no. 3, pp. 389–397, 2003.
- [28] M. D. Nicola, C. Carlo-Stella, M. Magni et al., "Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli," *Blood*, vol. 99, no. 10, pp. 3838–3843, 2002.
- [29] A. Bartholomew, C. Sturgeon, M. Siatskas et al., "Mesenchymal stem cells suppress lymphocyte proliferation in vitro and prolong skin graft survival in vivo," *Experimental Hematology*, vol. 30, no. 1, pp. 42–48, 2002.

- [30] M. Krampera, S. Glennie, J. Dyson et al., "Bone marrow mesenchymal stem cells inhibit the response of naive and memory antigen-specific T cells to their cognate peptide," *Blood*, vol. 101, no. 9, pp. 3722–3729, 2003.
- [31] K. Le Blanc, L. Tammik, B. Sundberg, S. E. Haynesworth, and O. Ringdén, "Mesenchymal stem cells inhibit and stimulate mixed lymphocyte cultures and mitogenic responses independently of the major histocompatibility complex," *Scandinavian Journal of Immunology*, vol. 57, no. 1, pp. 11–20, 2003.
- [32] H. M. Lazarus, O. N. Koc, S. M. Devine et al., "Cotransplantation of HLA-identical sibling culture-expanded mesenchymal stem cells and hematopoietic stem cells in hematologic malignancy patients," *Biology of Blood and Marrow Transplantation*, vol. 11, no. 5, pp. 389–398, 2005.
- [33] K. Le Blanc, I. Rasmusson, B. Sundberg et al., "Treatment of severe acute graft-versus-host disease with third party haploidentical mesenchymal stem cells," *The Lancet*, vol. 363, no. 9419, pp. 1439–1441, 2004.
- [34] C. Wan, R. Cheng, H. Wang, and T. Liu, "Immunomodulatory effects of mesenchymal stem cells derived from adipose tissues in a rat orthotopic liver transplantation model," *Hepatobiliary and Pancreatic Diseases International*, vol. 7, no. 1, pp. 29–33, 2008.
- [35] M. J. Crop, C. C. Baan, S. S. Korevaar et al., "Donor-derived mesenchymal stem cells suppress alloreactivity of kidney transplant patients," *Transplantation*, vol. 87, no. 6, pp. 896–906, 2009.
- [36] S. Inoue, F. C. Popp, G. E. Koehl et al., "Immunomodulatory effects of mesenchymal stem cells in a rat organ transplant model," *Transplantation*, vol. 81, no. 11, pp. 1589–1595, 2006.
- [37] K. Le Blanc, F. Frassoni, L. Ball et al., "Mesenchymal stem cells for treatment of steroid-resistant, severe, acute graft-versus-host disease: a phase II study," *The Lancet*, vol. 371, no. 9624, pp. 1579–1586, 2008.
- [38] R. E. Newman, D. Yoo, M. A. LeRoux, and A. Danilkovitch-Miagkova, "Treatment of inflammatory diseases with mesenchymal stem cells," *Inflammation and Allergy*, vol. 8, no. 2, pp. 110–123, 2009.
- [39] A. J. Nixon, L. R. Goodrich, M. S. Scimeca et al., "Gene therapy in musculoskeletal repair," *Annals of the New York Academy of Sciences*, vol. 1117, pp. 310–327, 2007.
- [40] W. J. C. Rombouts and R. E. Ploemacher, "Primary murine MSC show highly efficient homing to the bone marrow but lose homing ability following culture," *Leukemia*, vol. 17, no. 1, pp. 160–170, 2003.
- [41] M. Crisan, S. Yap, L. Casteilla et al., "A perivascular origin for mesenchymal stem cells in multiple human organs," *Cell Stem Cell*, vol. 3, no. 3, pp. 301–313, 2008.
- [42] P. Bianco, P. G. Robey, and P. J. Simmons, "Mesenchymal stem cells: revisiting history, concepts, and assays," *Cell Stem Cell*, vol. 2, no. 4, pp. 313–319, 2008.
- [43] L. D. Meirelles, P. C. Chagastelles, and N. B. Nardi, "Mesenchymal stem cells reside in virtually all post-natal organs and tissues," *Journal of Cell Science*, vol. 119, part 11, pp. 2204–2213, 2006.
- [44] M. Dominici, K. le Blanc, I. Mueller et al., "Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement," *Cytotherapy*, vol. 8, no. 4, pp. 315–317, 2006.
- [45] R. F. Wynn, C. A. Hart, C. Corradi-Perini et al., "A small proportion of mesenchymal stem cells strongly expresses functionally active CXCR4 receptor capable of promoting migration to bone marrow," *Blood*, vol. 104, no. 9, pp. 2643–2645, 2004.
- [46] D. G. Phinney and D. J. Prockop, "Concise review: mesenchymal stem/multipotent stromal cells: the state of transdifferentiation and modes of tissue repair—current views," *Stem Cells*, vol. 25, no. 11, pp. 2896–2902, 2007.
- [47] M. Shi, J. Li, L. Liao et al., "Regulation of CXCR4 expression in human mesenchymal stem cells by cytokine treatment: role in homing efficiency in NOD/SCID mice," *Haematologica*, vol. 92, no. 7, pp. 897–904, 2007.
- [48] T. Schioppa, B. Uranchimeg, A. Saccani et al., "Regulation of the chemokine receptor CXCR4 by hypoxia," *Journal of Experimental Medicine*, vol. 198, no. 9, pp. 1391–1402, 2003.
- [49] A. De Becker, P. Van Hummelen, M. Bakkus et al., "Migration of culture-expanded human mesenchymal stem cells through bone marrow endothelium is regulated by matrix metalloproteinase-2 and tissue inhibitor of metalloproteinase-3," *Haematologica*, vol. 92, no. 4, pp. 440–449, 2007.
- [50] B. Annabi, Y. Lee, S. Turcotte et al., "Hypoxia promotes murine bone-marrow-derived stromal cell migration and tube formation," *Stem Cells*, vol. 21, no. 3, pp. 337–347, 2003.
- [51] C. Ries, V. Egea, M. Karow, H. Kolb, M. Jochum, and P. Neth, "MMP-2, MT1-MMP, and TIMP-2 are essential for the invasive capacity of human mesenchymal stem cells: differential regulation by inflammatory cytokines," *Blood*, vol. 109, no. 9, pp. 4055–4063, 2007.
- [52] R. F. Pereira, M. D. O'Hara, A. V. Laptev et al., "Marrow stromal cells as a source of progenitor cells for nonhematopoietic tissues in transgenic mice with a phenotype of osteogenesis imperfecta," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 95, no. 3, pp. 1142–1147, 1998.
- [53] E. M. Horwitz, D. J. Prockop, L. A. Fitzpatrick et al., "Transplantability and therapeutic effects of bone marrow-derived mesenchymal cells in children with osteogenesis imperfecta," *Nature Medicine*, vol. 5, no. 3, pp. 309–313, 1999.
- [54] Y. Akiyama, C. Radtke, O. Honmou, and J. D. Kocsis, "Remyelination of the spinal cord following intravenous delivery of bone marrow cells," *Glia*, vol. 39, no. 3, pp. 229–236, 2002.
- [55] T. Nomura, O. Honmou, K. Harada, K. Houkin, H. Hamada, and J. D. Kocsis, "I.v. infusion of brain-derived neurotrophic factor gene-modified human mesenchymal stem cells protects against injury in a cerebral ischemia model in adult rat," *Neuroscience*, vol. 136, no. 1, pp. 161–169, 2005.
- [56] R. H. Lee, A. A. Pulin, M. J. Seo et al., "Intravenous hMSCs improve myocardial infarction in mice because cells embolized in lung are activated to secrete the anti-inflammatory protein TSG-6," *Cell Stem Cell*, vol. 5, no. 1, pp. 54–63, 2009.
- [57] J. Gao, J. E. Dennis, R. F. Muzic, M. Lundberg, and A. I. Caplan, "The dynamic in vivo distribution of bone marrow-derived mesenchymal stem cells after infusion," *Cells Tissues Organs*, vol. 169, no. 1, pp. 12–20, 2001.
- [58] S. Schrepfer, T. Deuse, H. Reichenspurner, M. P. Fischbein, R. C. Robbins, and M. P. Pelletier, "Stem cell transplantation: the Lung barrier," *Transplantation Proceedings*, vol. 39, no. 2, pp. 573–576, 2007.
- [59] P. Walczak, J. Zhang, A. A. Gilad et al., "Dual-modality monitoring of targeted intraarterial delivery of mesenchymal stem cells after transient ischemia," *Stroke*, vol. 39, no. 5, pp. 1569–1574, 2008.
- [60] E. Syková, P. Jendelová, L. Urdzíkova, P. Lesný, and A. Hejčl, "Bone marrow stem cells and polymer hydrogels—two strategies for spinal cord injury repair," *Cellular and Molecular Neurobiology*, vol. 26, no. 7–8, pp. 1113–1129, 2006.

- [61] D. Zhang, G. Fan, X. Zhou et al., "Over-expression of CXCR4 on mesenchymal stem cells augments myoangiogenesis in the infarcted myocardium," *Journal of Molecular and Cellular Cardiology*, vol. 44, no. 2, pp. 281–292, 2008.
- [62] F. Bacou, R. Boubaker El Andaloussi, P. Daussin et al., "Transplantation of adipose tissue-derived stromal cells increases mass and functional capacity of damaged skeletal muscle," *Cell Transplantation*, vol. 13, no. 2, pp. 103–111, 2004.
- [63] S. Schenk, N. Mal, A. Finan et al., "Monocyte chemotactic protein-3 is a myocardial mesenchymal stem cell homing factor," *Stem Cells*, vol. 25, no. 1, pp. 245–251, 2007.
- [64] Y. Zhuang, X. Chen, M. Xu, L. Zhang, and F. Xiang, "Chemokine stromal cell-derived factor 1/CXCL12 increases homing of mesenchymal stem cells to injured myocardium and neovascularization following myocardial infarction," *Chinese Medical Journal*, vol. 122, no. 2, pp. 183–187, 2009.
- [65] J. Tang, J. Wang, J. Yang et al., "Mesenchymal stem cells over-expressing SDF-1 promote angiogenesis and improve heart function in experimental myocardial infarction in rats," *European Journal of Cardio-Thoracic Surgery*, vol. 36, no. 4, pp. 644–650, 2009.
- [66] S. K. Ghadge, S. Mühlstedt, C. Özcelik, and M. Bader, "SDF-1 α as a therapeutic stem cell homing factor in myocardial infarction," *Pharmacology and Therapeutics*, vol. 129, no. 1, pp. 97–108, 2011.
- [67] B. Moser and P. Loetscher, "Lymphocyte traffic control by chemokines," *Nature Immunology*, vol. 2, no. 2, pp. 123–128, 2001.
- [68] A. Peled, I. Petit, O. Kollet et al., "Dependence of human stem cell engraftment and repopulation of NOD/SCID mice on CXCR4," *Science*, vol. 283, no. 5403, pp. 845–848, 1999.
- [69] A. Müller, B. Homey, H. Soto et al., "Involvement of chemokine receptors in breast cancer metastasis," *Nature*, vol. 410, no. 6824, pp. 50–56, 2001.
- [70] C. H. Ryu, S. A. Park, S. M. Kim et al., "Migration of human umbilical cord blood mesenchymal stem cells mediated by stromal cell-derived factor-1/CXCR4 axis via Akt, ERK, and p38 signal transduction pathways," *Biochemical and Biophysical Research Communications*, vol. 398, no. 1, pp. 105–110, 2010.
- [71] A. Kortessidis, A. Zannettino, S. Isenmann, S. Shi, T. Lapidot, and S. Gronthos, "Stromal-derived factor-1 promotes the growth, survival, and development of human bone marrow stromal stem cells," *Blood*, vol. 105, no. 10, pp. 3793–3801, 2005.
- [72] M. Honczarenko, Y. Le, M. Swierkowski, I. Ghiran, A. M. Glodek, and L. E. Silberstein, "Human bone marrow stromal cells express a distinct set of biologically functional chemokine receptors," *Stem Cells*, vol. 24, no. 4, pp. 1030–1041, 2006.
- [73] I. Von Lüttichau, M. Notohamiprodjo, A. Wechselberger et al., "Human adult CD34⁺ progenitor cells functionally express the chemokine receptors CCR1, CCR4, CCR7, CXCR5, and CCR10 but not CXCR4," *Stem Cells and Development*, vol. 14, no. 3, pp. 329–336, 2005.
- [74] D. A. De Ugarte, Z. Alfonso, P. A. Zuk et al., "Differential expression of stem cell mobilization-associated molecules on multi-lineage cells from adipose tissue and bone marrow," *Immunology Letters*, vol. 89, no. 2–3, pp. 267–270, 2003.
- [75] J. E. Ip, Y. Wu, J. Huang, L. Zhang, R. E. Pratt, and V. J. Dzau, "Mesenchymal stem cells use integrin β 1 not CXC chemokine receptor 4 for myocardial migration and engraftment," *Molecular Biology of the Cell*, vol. 18, no. 8, pp. 2873–2882, 2007.
- [76] B. Rüster, S. Göttig, R. J. Ludwig et al., "Mesenchymal stem cells display coordinated rolling and adhesion behavior on endothelial cells," *Blood*, vol. 108, no. 12, pp. 3938–3944, 2006.
- [77] M. Krampera, G. Pizzolo, G. Aprili, and M. Franchini, "Mesenchymal stem cells for bone, cartilage, tendon and skeletal muscle repair," *Bone*, vol. 39, no. 4, pp. 678–683, 2006.
- [78] C. Steingen, F. Brenig, L. Baumgartner, J. Schmidt, A. Schmidt, and W. Bloch, "Characterization of key mechanisms in transmigration and invasion of mesenchymal stem cells," *Journal of Molecular and Cellular Cardiology*, vol. 44, no. 6, pp. 1072–1084, 2008.
- [79] A. L. Ponte, E. Marais, N. Gallay et al., "The in vitro migration capacity of human bone marrow mesenchymal stem cells: comparison of chemokine and growth factor chemotactic activities," *Stem Cells*, vol. 25, no. 7, pp. 1737–1745, 2007.
- [80] H. Fan, G. Zhao, L. Liu et al., "Pre-treatment with IL-1 β enhances the efficacy of MSC transplantation in DSS-induced colitis," *Cellular & Molecular Immunology*, vol. 9, no. 6, pp. 473–481, 2012.
- [81] S. Hung, R. R. Pochampally, S. Hsu et al., "Short-term exposure of multipotent stromal cells to low oxygen increases their expression of CX3CR1 and CXCR4 and their engraftment in vivo," *PLoS ONE*, vol. 2, no. 5, article e416, 2007.
- [82] F. Zhang, Y. Hong, W. Liang et al., "Co-culture with Sertoli cells promotes proliferation and migration of umbilical cord mesenchymal stem cells," *Biochemical and Biophysical Research Communications*, vol. 427, no. 1, pp. 86–90, 2012.
- [83] S. Kumar and S. Ponnazhagan, "Bone homing of mesenchymal stem cells by ectopic α 4 integrin expression," *The FASEB Journal*, vol. 21, no. 14, pp. 3917–3927, 2007.
- [84] Z. Cheng, L. Ou, X. Zhou et al., "Targeted migration of mesenchymal stem cells modified with CXCR4 gene to infarcted myocardium improves cardiac performance," *Molecular Therapy*, vol. 16, no. 3, pp. 571–579, 2008.
- [85] F. Belema-Bedada, S. Uchida, A. Martire, S. Kostin, and T. Braun, "Efficient homing of multipotent adult mesenchymal stem cells depends on FROUNT-mediated clustering of CCR2," *Cell Stem Cell*, vol. 2, no. 6, pp. 566–575, 2008.
- [86] S. G. Crich, L. Biancone, V. Cantaluppi et al., "Improved route for the visualization of stem cells labeled with a Gd-/Eu-chelate as dual (MRI and fluorescence) agent," *Magnetic Resonance in Medicine*, vol. 51, no. 5, pp. 938–944, 2004.
- [87] M. Widerøe, Ø. Olsen, T. B. Pedersen et al., "Manganese-enhanced magnetic resonance imaging of hypoxic-ischemic brain injury in the neonatal rat," *NeuroImage*, vol. 45, no. 3, pp. 880–890, 2009.
- [88] M. Yamada and P. Yang, "In vitro labeling of human embryonic stem cells for magnetic resonance imaging," *Journal of Visualized Experiments*, no. 18, article e827, 2008.
- [89] C. Vandeputte, D. Thomas, T. Dresselaers et al., "Characterization of the inflammatory response in a photothrombotic stroke model by MRI: implications for stem cell transplantation," *Molecular Imaging and Biology*, vol. 13, no. 4, pp. 663–671, 2011.
- [90] C. Berger, M. Rausch, P. Schmidt, and M. Rudin, "Feasibility and limits of magnetically labeling primary cultured rat T cells with ferumoxides coupled with commonly used transfection agents," *Molecular Imaging*, vol. 5, no. 2, pp. 93–104, 2006.
- [91] J. W. M. Bulte and D. L. Kraitchman, "Iron oxide MR contrast agents for molecular and cellular imaging," *NMR in Biomedicine*, vol. 17, no. 7, pp. 484–499, 2004.

- [92] E. Küstermann, U. Himmelreich, K. Kandal et al., "Efficient stem cell labeling for MRI studies," *Contrast Media and Molecular Imaging*, vol. 3, no. 1, pp. 27–37, 2008.
- [93] M. Hoehn, E. Küstermann, J. Blunk et al., "Monitoring of implanted stem cell migration in vivo: a highly resolved in vivo magnetic resonance imaging investigation of experimental stroke in rat," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 25, pp. 16267–16272, 2002.
- [94] E. M. Shapiro, O. Gonzalez-Perez, J. Manuel García-Verdugo, A. Alvarez-Buylla, and A. P. Koretsky, "Magnetic resonance imaging of the migration of neuronal precursors generated in the adult rodent brain," *NeuroImage*, vol. 32, no. 3, pp. 1150–1157, 2006.
- [95] J. P. Sumner, E. M. Shapiro, D. Maric, R. Conroy, and A. P. Koretsky, "In vivo labeling of adult neural progenitors for MRI with micron sized particles of iron oxide: quantification of labeled cell phenotype," *NeuroImage*, vol. 44, no. 3, pp. 671–678, 2009.
- [96] A. Crabbe, C. Vandeputte, T. Dresselaers et al., "Effects of MRI contrast agents on the stem cell phenotype," *Cell Transplantation*, vol. 19, no. 8, pp. 919–936, 2010.
- [97] C. Vandeputte, J. Taymans, C. Casteels et al., "Automated quantitative gait analysis in animal models of movement disorders," *BMC Neuroscience*, vol. 11, article 92, 2010.
- [98] C. M. Deroose, S. K. Chitneni, R. Gijssbers et al., "Preliminary validation of varicella zoster virus thymidine kinase as a novel reporter gene for PET," *Nuclear Medicine and Biology*, vol. 39, no. 8, pp. 1266–1274, 2012.
- [99] S. S. Gambhir, J. R. Barrio, M. E. Phelps et al., "Imaging adenoviral-directed reporter gene expression in living animals with positron emission tomography," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 96, no. 5, pp. 2333–2338, 1999.
- [100] H. Hong, Y. Yang, and W. Cai, "Imaging gene expression in live cells and tissues," *Cold Spring Harbor Protocols*, vol. 6, no. 4, 2011.
- [101] E. Wolfs, T. Struys, T. Notelaers et al., "¹⁸F-FDG labeling of mesenchymal stem cells and multipotent adult progenitor cells for PET imaging: effects on ultrastructure and differentiation capacity," *Journal of Nuclear Medicine*, vol. 54, no. 3, pp. 447–454, 2013.
- [102] C. Caracó, L. Aloj, L. Chen, J. Y. Chou, and W. C. Eckelman, "Cellular release of [¹⁸F]2-fluoro-2-deoxyglucose as a function of the glucose-6-phosphatase enzyme system," *The Journal of Biological Chemistry*, vol. 275, no. 24, pp. 18489–18494, 2000.
- [103] J. Tolar, A. J. Nauta, M. J. Osborn et al., "Sarcoma derived from cultured mesenchymal stem cells," *Stem Cells*, vol. 25, no. 2, pp. 371–379, 2007.
- [104] N. de Vocht, D. Lin, J. Praet et al., "Quantitative and phenotypic analysis of mesenchymal stromal cell graft survival and recognition by microglia and astrocytes in mouse brain," *Immunobiology*, vol. 218, no. 5, pp. 696–705, 2013.
- [105] I. Bergwerf, N. De Vocht, B. Tambuyzer et al., "Reporter gene-expressing bone marrow-derived stromal cells are immunotolerated following implantation in the central nervous system of syngeneic immunocompetent mice," *BMC Biotechnology*, vol. 9, article 1, 2009.
- [106] D. L. Kraitchman, M. Tatsumi, W. D. Gilson et al., "Dynamic imaging of allogeneic mesenchymal stem cells trafficking to myocardial infarction," *Circulation*, vol. 112, no. 10, pp. 1451–1461, 2005.



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