

 Open access • Journal Article • DOI:10.1038/NG.685

Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution — [Source link](#)

Iris M. Heid, Anne U. Jackson, Joshua C. Randall, Thomas W. Winkler ...+352 more authors

Institutions: [University of Regensburg](#), [University of Michigan](#), [University of Oxford](#), [Harvard University](#) ...+86 more institutions

Published on: 01 Nov 2010 - [Nature Genetics](#) (Nature Publishing Group)

Topics: [Body mass index](#)

Related papers:

- [Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index](#)
- [Genome-wide association scan meta-analysis identifies three Loci influencing adiposity and fat distribution](#)
- [Six new loci associated with body mass index highlight a neuronal influence on body weight regulation](#)
- [New genetic loci link adipose and insulin biology to body fat distribution](#)
- [A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity](#)

Share this paper:    

View more about this paper here: <https://typeset.io/papers/meta-analysis-identifies-13-new-loci-associated-with-waist-4b398tx0c0>

Published in final edited form as:

Nat Genet. 2010 November ; 42(11): 949–960. doi:10.1038/ng.685.

Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution

Iris M Heid^{1,2,214}, Anne U Jackson^{3,214}, Joshua C Randall^{4,214}, Thomas W Winkler^{1,214}, Lu Qi^{5,6,214}, Valgerdur Steinthorsdottir^{7,214}, Gudmar Thorleifsson^{7,214}, M Carola Zillikens^{8,9}, Elizabeth K Speliotes^{10,11}, Reedik Mägi⁴, Tsegaselassie Workalemahu⁵, Charles C White¹², Nabila Bouatia-Naji^{13,14}, Tamara B Harris¹⁵, Sonja I Berndt¹⁶, Erik Ingelsson¹⁷, Cristen J Willer³, Michael N Weedon¹⁸, Jian'An Luan¹⁹, Sailaja Vedantam^{10,20}, Tõnu Esko^{21,23}, Tuomas O Kilpeläinen¹⁹, Zoltán Kutalik^{24,25}, Shengxu Li¹⁹, Keri L Monda²⁶, Anna L Dixon²⁷, Christopher C Holmes^{28,29}, Lee M Kaplan^{11,30,31}, Liming Liang^{32,33}, Josine L Min³⁴, Miriam F Moffatt³⁵, Cliona Molony³⁶, George Nicholson²⁹, Eric E Schadt^{37,38}, Krina T Zondervan³⁹, Mary F Feitosa⁴⁰, Teresa Ferreira⁴, Hana Lango Allen¹⁸, Robert J Weyant³, Eleanor Wheeler⁴¹, Andrew R Wood¹⁸, MAGIC⁴², Karol Estrada^{8,9,43}, Michael E Goddard^{44,45}, Guillaume Lettre^{46,47}, Massimo Mangino⁴⁸, Dale R Nyholt⁴⁹, Shaun Purcell^{50,52}, Albert Vernon Smith^{53,54}, Peter M Visscher⁵⁵, Jian Yang⁵⁵, Steven A McCarroll^{50,56,57}, James Nemesh⁵⁶, Benjamin F Voight^{50,56,57}, Devin Absher⁵⁸, Najaf Amin⁴³, Thor Aspelund^{53,54}, Lachlan Coin⁵⁹, Nicole L Glazer^{60,61}, Caroline Hayward⁶², Nancy L Heard-costa⁶³, Jouke-Jan Hottenga⁶⁴, Åsa Johansson^{65,66}, Toby Johnson^{24,25,67,68}, Marika Kaakinen^{69,70}, Karen Kapur^{24,25}, Shamika Ketkar⁴⁰, Joshua W Knowles⁷¹, Peter Kraft^{32,33}, Aldi T Kraja⁴⁰, Claudia Lamina^{2,72}, Michael F Leitzmann¹, Barbara McKnight⁷³, Andrew P Morris⁴, Ken K Ong¹⁹, John R B Perry¹⁸, Marjolein J Peters^{8,9}, Ozren Polasek^{74,75}, Inga Prokopenko^{4,76}, Nigel W Rayner^{4,76}, Samuli Ripatti^{77,78}, Fernando Rivadeneira^{8,9,43}, Neil R Robertson^{4,76}, Serena Sanna⁷⁹, Ulla Sovio⁵⁹, Ida Surakka^{77,78}, Alexander Teumer⁸⁰, Sophie van Wingerden⁴³, Veronique Vitart⁶², Jing Hua Zhao¹⁹, Christine Cavalcanti-Proença^{13,14}, Peter S Chines⁸¹, Eva Fisher⁸², Jennifer R Kulzer⁸³, Cecile Lecoeur^{13,14}, Narisu Narisu⁸¹, Camilla Sandholt⁸⁴, Laura J Scott³, Kaisa Silander^{77,78}, Klaus Stark⁸⁵, Mari-Liis Tammesoo²¹, Tanya M Teslovich³, Nicholas John Timpson⁸⁶, Richard M Watanabe^{87,88}, Ryan Welch³, Daniel I Chasman^{30,89}, Matthew N Cooper⁹⁰, John-Olov Jansson⁹¹, Johannes Kettunen^{77,78}, Robert W Lawrence⁹⁰, Niina Pellikka^{77,78}, Markus Perola^{77,78}, Liesbeth Vandenput⁹², Helene Alavere²¹, Peter Almgren⁹³, Larry D Atwood⁶³, Amanda J Bennett⁷⁶, Reiner Biffar⁹⁴, Lori L Bonnycastle⁸¹, Stefan R Bornstein⁹⁵, Thomas A Buchanan^{87,96}, Harry Campbell⁹⁷, Ian N M Day⁸⁶, Mariano Del⁷⁹, Marcus Dörr⁹⁸, Paul Elliott^{59,99}, Michael R Erdos⁸¹, Johan G Eriksson^{100,104}, Nelson B Freimer¹⁰⁵, Mao Fu¹⁰⁶, Stefan Gaget^{13,14}, Eco J C Geus⁶⁴, Anette P Gjesing⁸⁴, Harald Grallert², Jürgen Gräßler¹⁰⁷, Christopher J Groves⁷⁶, Candace Guiducci¹⁰, Anna-Liisa Hartikainen¹⁰⁸, Neelam Hassanali⁷⁶, Aki S Havulinna¹⁰⁹, Karl-Heinz Herzig^{70,110,111}, Andrew A Hicks¹¹², Jennie Hui^{90,113,114}, Wilmar Igl⁶⁵, Pekka Jousilahti¹⁰⁹, Antti Jula¹¹⁵, Eero Kajantie^{101,116}, Leena Kinnunen¹¹⁷, Ivana Kolcic⁷⁴, Seppo Koskinen¹⁰⁹, Peter Kovacs¹¹⁸, Heyo K Kroemer¹¹⁹, Vjekoslav Krzelj¹²⁰, Johanna Kuusisto¹²¹, Kirsti Kvaloy¹²², Jaana Laitinen¹²³, Olivier Lantieri¹²⁴, G Mark Lathrop¹²⁵, Marja-Liisa Lokki¹²⁶, Robert N Luben¹²⁷, Barbara Ludwig⁹⁵, Wendy L McArdle¹²⁸, Anne McCarthy¹²⁹, Mario A Morken⁸¹, Mari Nelis^{21,23}, Matt J Neville⁷⁶, Guillaume Paré¹³⁰, Alex N Parker¹³¹, John F Peden^{4,132}, Irene Pichler¹¹², Kirsi H Pietiläinen^{133,134}, Carl G P Platou^{122,135}, Anneli Pouta^{108,136}, Martin Ridderstråle¹³⁷, Nilesh J Samani^{138,139}, Jouko Saramies¹⁴⁰, Juha Sinisalo¹⁴¹, Jan H Smit¹⁴², Rona J Strawbridge¹⁴³, Heather M Stringham³, Amy J Swift⁸¹, Maris Teder-Laving^{22,23}, Brian Thomson¹⁰, Gianluca Usala⁷⁹, Joyce B J van Meurs^{8,9,43}, Gert-Jan van Ommen^{144,145}, Vincent Vatin^{13,14}, Claudia B Volpato¹¹², Henri Wallaschofski¹⁴⁶, G Bragi

Walters⁷, Elisabeth Widen⁷⁷, Sarah H Wild⁹⁷, Gonneke Willemssen⁶⁴, Daniel R Witte¹⁴⁷, Lina Zgaga⁷⁴, Paavo Zitting¹⁴⁸, John P Beilby^{113,114,149}, Alan L James^{114,150}, Mika Kähönen¹⁵¹, Terho Lehtimäki¹⁵², Markku S Nieminen¹⁴¹, Claes Ohlsson⁹², Lyle J Palmer^{90,114}, Olli Raitakari^{153,154}, Paul M Ridker^{30,89}, Michael Stumvoll^{155,156}, Anke Tönjes^{155,157}, Jorma Viikari¹⁵⁸, Beverley Balkau^{159,160}, Yoav Ben-Shlomo¹⁶¹, Richard N Bergman⁸⁷, Heiner Boeing⁸², George Davey Smith⁸⁶, Shah Ebrahim^{162,163}, Philippe Froguel^{13,14,164}, Torben Hansen^{84,165}, Christian Hengstenberg^{166,167}, Kristian Hveem¹²², Bo Isomaa^{103,168}, Torben Jørgensen^{169,170}, Fredrik Karpe^{76,171}, Kay-Tee Khaw¹²⁷, Markku Laakso¹²¹, Debbie A Lawlor⁸⁶, Michel Marre^{172,173}, Thomas Meitinger^{174,175}, Andres Metspalu^{21,23}, Kristian Midthjell¹²², Oluf Pedersen^{84,176,177}, Veikko Salomaa¹⁰⁹, Peter E H Schwarz¹⁷⁸, Tiinamaija Tuomi^{103,179,180}, Jaakko Tuomilehto^{117,181,182}, Timo T Valle¹¹⁷, Nicholas J Wareham¹⁹, Alice M Arnold^{73,183}, Jacques S Beckmann^{24,184}, Sven Bergmann^{24,25}, Eric Boerwinkle¹⁸⁵, Dorret I Boomsma⁶⁴, Mark J Caulfield⁶⁸, Francis S Collins⁸¹, Gudny Eiriksdottir⁵³, Vilmundur Gudnason^{53,54}, Ulf Gyllenstein⁶⁵, Anders Hamsten¹⁴³, Andrew T Hattersley¹⁸, Albert Hofman^{9,43}, Frank B Hu^{5,6,32}, Thomas Illig², Carlos Iribarren^{186,187}, Marjo-Riitta Jarvelin^{59,69,70,136}, W H Linda Kao¹⁸⁸, Jaakko Kaprio^{77,133,189}, Lenore J Launer¹⁵, Patricia B Munroe⁶⁸, Ben Oostra¹⁹⁰, Brenda W Penninx^{142,191,192}, Peter P Pramstaller^{112,193,194}, Bruce M Psaty^{195,196}, Thomas Quertermous⁷¹, Aila Rissanen¹³⁴, Igor Rudan^{97,120}, Alan R Shuldiner^{106,197}, Nicole Soranzo^{41,48}, Timothy D Spector⁴⁸, Ann-Christine Syvanen¹⁹⁸, Manuela Uda⁷⁹, André Uitterlinden^{8,9,43}, Henry Völzke¹⁹⁹, Peter Vollenweider²⁰⁰, James F Wilson⁹⁷, Jacqueline C Witteman^{9,43}, Alan F Wright⁶², Gonçalo R Abecasis³, Michael Boehnke³, Ingrid B Borecki^{40,201}, Panos Deloukas⁴¹, Timothy M Frayling¹⁸, Leif C Groop⁹³, Talin Haritunians²⁰², David J Hunter^{5,6,32}, Robert C Kaplan²⁰³, Kari E North^{26,204}, Jeffrey R O'connell¹⁰⁶, Leena Peltonen^{41,51,77,101,205}, David Schlessinger²⁰⁶, David P Strachan²⁰⁷, Joel N Hirschhorn^{10,20,208}, Themistocles L Assimes⁷¹, H-Erich Wichmann^{2,209,210}, Unnur Thorsteinsdottir^{7,211}, Cornelia M van Duijn^{9,43}, Kari Stefansson^{7,211,215}, L Adrienne Cupples^{12,215}, Ruth J F Loos^{19,215}, Inês Barroso^{41,212,215}, Mark I McCarthy^{4,76,171,215}, Caroline S Fox^{213,215}, Karen L Mohlke^{83,215}, and Cecilia M Lindgren^{4,76,215}

¹Regensburg University Medical Center, Department of Epidemiology and Preventive Medicine, Regensburg, Germany. ²Institute of Epidemiology, Helmholtz Zentrum München-German Research Center for Environmental Health, Neuherberg, Germany. ³Department of Biostatistics, Center for Statistical Genetics, University of Michigan, Ann Arbor, Michigan, USA. ⁴Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, UK. ⁵Department of Nutrition, Harvard School of Public Health, Boston, Massachusetts, USA. ⁶Channing Laboratory, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts, USA. ⁷deCODE Genetics, Reykjavik, Iceland. ⁸Department of Internal Medicine, Erasmus Medical Center (MC), Rotterdam, The Netherlands. ⁹Netherlands Genomics Initiative (NGI)-sponsored Netherlands Consortium for Healthy Aging (NCHA), Rotterdam, The Netherlands. ¹⁰Metabolism Initiative and Program in Medical and Population Genetics, Broad Institute, Cambridge, Massachusetts, USA. ¹¹Division of Gastroenterology, Massachusetts General Hospital, Boston, Massachusetts, USA. ¹²Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts, USA. ¹³Centre National de la Recherche Scientifique (CNRS), UMR8199-IBL-Institut Pasteur de Lille, Lille, France. ¹⁴University Lille Nord de France, Lille, France. ¹⁵Laboratory of Epidemiology, Demography, Biometry, National Institute on Aging, National Institutes of Health, Bethesda, Maryland, USA. ¹⁶Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Department of Health and Human Services, Bethesda, Maryland, USA. ¹⁷Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden. ¹⁸Genetics of Complex Traits, Peninsula College of Medicine and Dentistry, University of Exeter, Exeter, UK. ¹⁹Medical Research Council (MRC) Epidemiology Unit, Institute of Metabolic Science, Addenbrooke's Hospital, Cambridge, UK. ²⁰Divisions of Genetics and Endocrinology and Program in Genomics,

Children's Hospital, Boston, Massachusetts, USA. ²¹Estonian Genome Center, University of Tartu, Tartu, Estonia. ²²Estonian Biocenter, Tartu, Estonia. ²³Institute of Molecular and Cell Biology, University of Tartu, Tartu, Estonia. ²⁴Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland. ²⁵Swiss Institute of Bioinformatics, Lausanne, Switzerland. ²⁶Department of Epidemiology, School of Public Health, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, USA. ²⁷Department of Pharmacy and Pharmacology, University of Bath, Bath, UK. ²⁸MRC Harwell, Harwell Science and Innovation Campus, Oxfordshire, UK. ²⁹Department of Statistics, University of Oxford, Oxford, UK. ³⁰Harvard Medical School, Boston, Massachusetts, USA. ³¹Massachusetts General Hospital (MGH) Weight Center, Massachusetts General Hospital, Boston, Massachusetts, USA. ³²Department of Epidemiology, Harvard School of Public Health, Boston, Massachusetts, USA. ³³Department of Biostatistics, Harvard School of Public Health, Boston, Massachusetts, USA. ³⁴Human Genetics, Leiden University Medical Center, Leiden, The Netherlands. ³⁵National Heart and Lung Institute, Imperial College London, London, UK. ³⁶Merck Research Laboratories, Merck & Co., Inc., Boston, Massachusetts, USA. ³⁷Pacific Biosciences, Menlo Park, California, USA. ³⁸Sage Bionetworks, Seattle, Washington, USA. ³⁹Genetic and Genomic Epidemiology Unit, Wellcome Trust Centre for Human Genetics, Oxford, UK. ⁴⁰Department of Genetics, Washington University School of Medicine, St. Louis, Missouri, USA. ⁴¹Wellcome Trust Sanger Institute, Hinxton, Cambridge, UK. ⁴²On behalf of the MAGIC (Meta-Analyses of Glucose and Insulin-related traits Consortium) investigators. ⁴³Department of Epidemiology, Erasmus MC, Rotterdam, The Netherlands. ⁴⁴University of Melbourne, Parkville, Australia. ⁴⁵Department of Primary Industries, Melbourne, Victoria, Australia. ⁴⁶Montreal Heart Institute, Montreal, Quebec, Canada. ⁴⁷Department of Medicine, Université de Montréal, Montreal, Quebec, Canada. ⁴⁸Department of Twin Research and Genetic Epidemiology, King's College London, London, UK. ⁴⁹Neurogenetics Laboratory, Queensland Institute of Medical Research, Queensland, Australia. ⁵⁰Center for Human Genetic Research, Massachusetts General Hospital, Boston, Massachusetts, USA. ⁵¹The Broad Institute of Harvard and Massachusetts Institute of Technology (MIT), Cambridge, Massachusetts, USA. ⁵²Department of Psychiatry, Harvard Medical School, Boston, Massachusetts, USA. ⁵³Icelandic Heart Association, Kopavogur, Iceland. ⁵⁴University of Iceland, Reykjavik, Iceland. ⁵⁵Queensland Statistical Genetics Laboratory, Queensland Institute of Medical Research, Queensland, Australia. ⁵⁶Program in Medical and Population Genetics, Broad Institute of Harvard and MIT, Cambridge, Massachusetts, USA. ⁵⁷Department of Molecular Biology, Massachusetts General Hospital, Boston, Massachusetts, USA. ⁵⁸Hudson Alpha Institute for Biotechnology, Huntsville, Alabama, USA. ⁵⁹Department of Epidemiology and Biostatistics, School of Public Health, Faculty of Medicine, Imperial College London, London, UK. ⁶⁰Department of Medicine, University of Washington, Seattle, Washington, USA. ⁶¹Cardiovascular Health Research Unit, University of Washington, Seattle, Washington, USA. ⁶²MRC Human Genetics Unit, Institute for Genetics and Molecular Medicine, Western General Hospital, Edinburgh, Scotland, UK. ⁶³Department of Neurology, Boston University School of Medicine, Boston, Massachusetts, USA. ⁶⁴Department of Biological Psychology, Vrije Universiteit (VU) University Amsterdam, Amsterdam, The Netherlands. ⁶⁵Department of Genetics and Pathology, Rudbeck Laboratory, University of Uppsala, Uppsala, Sweden. ⁶⁶Department of Cancer Research and Molecular Medicine, Faculty of Medicine, Norwegian University of Science and Technology (NTNU), Trondheim, Norway. ⁶⁷Clinical Pharmacology, William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, Queen Mary, University of London, London, UK. ⁶⁸Clinical Pharmacology and Barts and The London Genome Centre, William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London, UK. ⁶⁹Institute of Health Sciences, University of Oulu, Oulu, Finland. ⁷⁰Biocenter Oulu, University of Oulu, Oulu, Finland. ⁷¹Department of Medicine, Stanford University School of Medicine, Stanford, California, USA. ⁷²Division of Genetic Epidemiology, Department of Medical Genetics, Molecular and Clinical Pharmacology, Innsbruck Medical University, Innsbruck, Austria. ⁷³Department of

Biostatistics, University of Washington, Seattle, Washington, USA. ⁷⁴Andrija Stampar School of Public Health, Medical School, University of Zagreb, Zagreb, Croatia. ⁷⁵Gen-Info Ltd, Zagreb, Croatia. ⁷⁶Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, Oxford, UK. ⁷⁷Institute for Molecular Medicine Finland (FIMM), University of Helsinki, Helsinki, Finland. ⁷⁸National Institute for Health and Welfare, Department of Chronic Disease Prevention, Unit of Public Health Genomics, Helsinki, Finland. ⁷⁹Istituto di Neurogenetica e Neurofarmacologia del CNR, Monserrato, Cagliari, Italy. ⁸⁰Interfaculty Institute for Genetics and Functional Genomics, Ernst-Moritz-Arndt-University Greifswald, Greifswald, Germany. ⁸¹National Human Genome Research Institute, National Institutes of Health, Bethesda, Maryland, USA. ⁸²Department of Epidemiology, German Institute of Human Nutrition Potsdam-Rehbruecke, Nuthetal, Germany. ⁸³Department of Genetics, University of North Carolina, Chapel Hill, North Carolina, USA. ⁸⁴Hagedorn Research Institute, Gentofte, Denmark. ⁸⁵Regensburg University Medical Center, Clinic and Policlinic for Internal Medicine II, Regensburg, Germany. ⁸⁶MRC Centre for Causal Analyses in Translational Epidemiology, Department of Social Medicine, Oakfield House, Bristol, UK. ⁸⁷Department of Physiology and Biophysics, Keck School of Medicine, University of Southern California, Los Angeles, California, USA. ⁸⁸Department of Preventive Medicine, Keck School of Medicine, University of Southern California, Los Angeles, California, USA. ⁸⁹Division of Preventive Medicine, Brigham and Women's Hospital, Boston, Massachusetts, USA. ⁹⁰Centre for Genetic Epidemiology and Biostatistics, University of Western Australia, Crawley, Western Australia, Australia. ⁹¹Department of Physiology, Institute of Neuroscience and Physiology, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden. ⁹²Department of Internal Medicine, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden. ⁹³Lund University Diabetes Centre, Department of Clinical Sciences, Lund University, Malmö, Sweden. ⁹⁴Zentrum für Zahn-, Mund- und Kieferheilkunde, Greifswald, Germany. ⁹⁵Department of Medicine III, University of Dresden, Dresden, Germany. ⁹⁶Division of Endocrinology, Keck School of Medicine, University of Southern California, Los Angeles, California, USA. ⁹⁷Centre for Population Health Sciences, University of Edinburgh, Teviot Place, Edinburgh, Scotland, UK. ⁹⁸Department of Internal Medicine B, Ernst-Moritz-Arndt University, Greifswald, Germany. ⁹⁹MRC-Health Protection Agency (HPA) Centre for Environment and Health, London, UK. ¹⁰⁰Department of General Practice and Primary Health Care, University of Helsinki, Helsinki, Finland. ¹⁰¹National Institute for Health and Welfare, Helsinki, Finland. ¹⁰²Helsinki University Central Hospital, Unit of General Practice, Helsinki, Finland. ¹⁰³Folkhalsan Research Centre, Helsinki, Finland. ¹⁰⁴Vasa Central Hospital, Vasa, Finland. ¹⁰⁵Center for Neurobehavioral Genetics, University of California, Los Angeles, California, USA. ¹⁰⁶Department of Medicine, University of Maryland School of Medicine, Baltimore, Maryland, USA. ¹⁰⁷Department of Medicine III, Pathobiochemistry, University of Dresden, Dresden, Germany. ¹⁰⁸Department of Clinical Sciences/Obstetrics and Gynecology, University of Oulu, Oulu, Finland. ¹⁰⁹National Institute for Health and Welfare, Department of Chronic Disease Prevention, Chronic Disease Epidemiology and Prevention Unit, Helsinki, Finland. ¹¹⁰Institute of Biomedicine, Department of Physiology, University of Oulu, Oulu, Finland. ¹¹¹Department of Psychiatry, Kuopio University Hospital and University of Kuopio, Kuopio, Finland. ¹¹²Institute of Genetic Medicine, European Academy Bozen-Bolzano (EURAC), Bolzano-Bozen, Italy (affiliated Institute of the University of Lübeck, Lübeck, Germany). ¹¹³PathWest Laboratory of Western Australia, Department of Molecular Genetics, J Block, QEII Medical Centre, Nedlands, Western Australia, Australia. ¹¹⁴Busselton Population Medical Research Foundation Inc., Sir Charles Gairdner Hospital, Nedlands, Western Australia, Australia. ¹¹⁵National Institute for Health and Welfare, Department of Chronic Disease Prevention, Population Studies Unit, Turku, Finland. ¹¹⁶Hospital for Children and Adolescents, Helsinki University Central Hospital and University of Helsinki, Helsinki, Finland. ¹¹⁷National Institute for Health and Welfare, Diabetes Prevention Unit, Helsinki, Finland. ¹¹⁸Interdisciplinary Centre for Clinical Research, University of Leipzig, Leipzig, Germany. ¹¹⁹Institut für Pharmakologie, Universität Greifswald, Greifswald, Germany. ¹²⁰Croatian

Centre for Global Health, School of Medicine, University of Split, Split, Croatia. ¹²¹Department of Medicine, University of Kuopio and Kuopio University Hospital, Kuopio, Finland. ¹²²Nord-Trøndelag Health Study (HUNT) Research Centre, Department of Public Health and General Practice, Norwegian University of Science and Technology, Levanger, Norway. ¹²³Finnish Institute of Occupational Health, Oulu, Finland. ¹²⁴Institut inter-regional pour la sante (IRSA), La Riche, France. ¹²⁵Centre National de Genotypage, Evry, Paris, France. ¹²⁶Transplantation Laboratory, Haartman Institute, University of Helsinki, Helsinki, Finland. ¹²⁷Department of Public Health and Primary Care, Institute of Public Health, University of Cambridge, Cambridge, UK. ¹²⁸Avon Longitudinal Study of Parents and Children (ALSPAC) Laboratory, Department of Social Medicine, University of Bristol, Bristol, UK. ¹²⁹Division of Health, Research Board, An Bord Taighde Sláinte, Dublin, Ireland. ¹³⁰Department of Pathology and Molecular Medicine, McMaster University, Hamilton, Ontario, Canada. ¹³¹Amgen, Cambridge, Massachusetts, USA. ¹³²Department of Cardiovascular Medicine, University of Oxford, John Radcliffe Hospital, Headington, Oxford, UK. ¹³³Finnish Twin Cohort Study, Department of Public Health, University of Helsinki, Helsinki, Finland. ¹³⁴Obesity Research Unit, Department of Psychiatry, Helsinki University Central Hospital, Helsinki, Finland. ¹³⁵Department of Medicine, Levanger Hospital, The Nord-Trøndelag Health Trust, Levanger, Norway. ¹³⁶National Institute for Health and Welfare, Oulu, Finland. ¹³⁷Department of Clinical Sciences, Lund University, Malmö, Sweden. ¹³⁸Department of Cardiovascular Sciences, University of Leicester, Glenfield Hospital, Leicester, UK. ¹³⁹Leicester National Institute for Health Research (NIHR) Biomedical Research Unit in Cardiovascular Disease, Glenfield Hospital, Leicester, UK. ¹⁴⁰South Karelia Central Hospital, Lappeenranta, Finland. ¹⁴¹Division of Cardiology, Cardiovascular Laboratory, Helsinki University Central Hospital, Helsinki, Finland. ¹⁴²Department of Psychiatry/Instituut voor Extramuraal Geneeskundig Onderzoek (EMGO) Institute, VU University Medical Center, Amsterdam, The Netherlands. ¹⁴³Atherosclerosis Research Unit, Department of Medicine, Solna, Karolinska Institutet, Karolinska University Hospital, Stockholm, Sweden. ¹⁴⁴Department of Human Genetics, Leiden University Medical Center, Leiden, The Netherlands. ¹⁴⁵Center of Medical Systems Biology, Leiden University Medical Center, Leiden, The Netherlands. ¹⁴⁶Institut für Klinische Chemie und Laboratoriumsmedizin, Universität Greifswald, Greifswald, Germany. ¹⁴⁷Steno Diabetes Center, Gentofte, Denmark. ¹⁴⁸Department of Physiatrics, Lapland Central Hospital, Rovaniemi, Finland. ¹⁴⁹School of Pathology and Laboratory Medicine, University of Western Australia, Nedlands, Western Australia, Australia. ¹⁵⁰School of Medicine and Pharmacology, University of Western Australia, Perth, Western Australia, Australia. ¹⁵¹Department of Clinical Physiology, University of Tampere and Tampere University Hospital, Tampere, Finland. ¹⁵²Department of Clinical Chemistry, University of Tampere and Tampere University Hospital, Tampere, Finland. ¹⁵³Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku, Turku, Finland. ¹⁵⁴The Department of Clinical Physiology, Turku University Hospital, Turku, Finland. ¹⁵⁵Department of Medicine, University of Leipzig, Leipzig, Germany. ¹⁵⁶Leipziger Interdisziplinärer Forschungskomplex zu molekularen Ursachen umwelt- und lebensstilassoziierter Erkrankungen (LIFE) Study Centre, University of Leipzig, Leipzig, Germany. ¹⁵⁷Coordination Centre for Clinical Trials, University of Leipzig, Leipzig, Germany. ¹⁵⁸Department of Medicine, University of Turku and Turku University Hospital, Turku, Finland. ¹⁵⁹INSERM Centre de Recherche en Epidémiologie et Santé des Populations (CESP) U1018, Villejuif, France. ¹⁶⁰University Paris Sud 11, Unité Mixte de Recherche en Santé (UMRS) 1018, Villejuif, France. ¹⁶¹Department of Social Medicine, University of Bristol, Bristol, UK. ¹⁶²The London School of Hygiene and Tropical Medicine, London, UK. ¹⁶³South Asia Network for Chronic Disease, New Delhi, India. ¹⁶⁴Department of Genomics of Common Disease, School of Public Health, Imperial College London, London, UK. ¹⁶⁵Faculty of Health Science, University of Southern Denmark, Odense, Denmark. ¹⁶⁶Klinik und Poliklinik für Innere Medizin II, Universität Regensburg, Regensburg, Germany. ¹⁶⁷Regensburg University Medical Center, Innere Medizin II, Regensburg, Germany. ¹⁶⁸Department of Social Services and Health Care, Jakobstad, Finland. ¹⁶⁹Research

Centre for Prevention and Health, Glostrup University Hospital, Glostrup, Denmark. ¹⁷⁰Faculty of Health Science, University of Copenhagen, Copenhagen, Denmark. ¹⁷¹NIHR Oxford Biomedical Research Centre, Churchill Hospital, Oxford, UK. ¹⁷²Department of Endocrinology, Diabetology and Nutrition, Bichat-Claude Bernard University Hospital, Assistance Publique des Hôpitaux de Paris, Paris, France. ¹⁷³Cardiovascular Genetics Research Unit, Université Henri Poincaré-Nancy 1, Nancy, France. ¹⁷⁴Institute of Human Genetics, Klinikum rechts der Isar der Technischen Universität München, Munich, Germany. ¹⁷⁵Institute of Human Genetics, Helmholtz Zentrum München-German Research Center for Environmental Health, Neuherberg, Germany. ¹⁷⁶Institute of Biomedical Sciences, University of Copenhagen, Copenhagen, Denmark. ¹⁷⁷Faculty of Health Science, University of Aarhus, Aarhus, Denmark. ¹⁷⁸Department of Medicine III, Prevention and Care of Diabetes, University of Dresden, Dresden, Germany. ¹⁷⁹Department of Medicine, Helsinki University Central Hospital, Helsinki, Finland. ¹⁸⁰Research Program of Molecular Medicine, University of Helsinki, Helsinki, Finland. ¹⁸¹Hjelt Institute, Department of Public Health, University of Helsinki, Helsinki, Finland. ¹⁸²South Ostrobothnia Central Hospital, Seinäjoki, Finland. ¹⁸³Collaborative Health Studies Coordinating Center, Seattle, Washington, USA. ¹⁸⁴Service of Medical Genetics, Centre Hospitalier Universitaire Vaudois (CHUV) University Hospital, Lausanne, Switzerland. ¹⁸⁵Human Genetics Center and Institute of Molecular Medicine, University of Texas Health Science Center, Houston, Texas, USA. ¹⁸⁶Division of Research, Kaiser Permanente Northern California, Oakland, California, USA. ¹⁸⁷Department of Epidemiology and Biostatistics, University of California, San Francisco, San Francisco, California, USA. ¹⁸⁸Department of Epidemiology and Medicine, Johns Hopkins Bloomberg School of Public Health, Baltimore, Maryland, USA. ¹⁸⁹National Institute for Health and Welfare, Department of Mental Health and Substance Abuse Services, Unit for Child and Adolescent Mental Health, Helsinki, Finland. ¹⁹⁰Department of Clinical Genetics, Erasmus MC, Rotterdam, The Netherlands. ¹⁹¹Department of Psychiatry, Leiden University Medical Centre, Leiden, The Netherlands. ¹⁹²Department of Psychiatry, University Medical Centre Groningen, Groningen, The Netherlands. ¹⁹³Department of Neurology, General Central Hospital, Bolzano, Italy. ¹⁹⁴Department of Neurology, University of Lübeck, Lübeck, Germany. ¹⁹⁵Departments of Epidemiology, Medicine and Health Services, University of Washington, Seattle, Washington, USA. ¹⁹⁶Group Health Research Institute, Group Health, Seattle, Washington, USA. ¹⁹⁷Geriatrics Research and Education Clinical Center, Baltimore Veterans Administration Medical Center, Baltimore, Maryland, USA. ¹⁹⁸Uppsala University, Department of Medical Sciences, Molecular Medicine, Uppsala, Sweden. ¹⁹⁹Institut für Community Medicine, Greifswald, Germany. ²⁰⁰Department of Internal Medicine, Centre Hospitalier Universitaire Vaudois (CHUV) University Hospital, Lausanne, Switzerland. ²⁰¹Division of Biostatistics, Washington University School of Medicine, St. Louis, Missouri, USA. ²⁰²Medical Genetics Institute, Cedars-Sinai Medical Center, Los Angeles, California, USA. ²⁰³Department of Epidemiology and Population Health, Albert Einstein College of Medicine, Bronx, New York, USA. ²⁰⁴Carolina Center for Genome Sciences, School of Public Health, University of North Carolina Chapel Hill, Chapel Hill, North Carolina, USA. ²⁰⁵Department of Medical Genetics, University of Helsinki, Helsinki, Finland. ²⁰⁶Laboratory of Genetics, National Institute on Aging, Baltimore, Maryland, USA. ²⁰⁷Division of Community Health Sciences, St George's, University of London, London, UK. ²⁰⁸Department of Genetics, Harvard Medical School, Boston, Massachusetts, USA. ²⁰⁹Klinikum Grosshadern, Munich, Germany. ²¹⁰Ludwig-Maximilians-Universität, Institute of Medical Informatics, Biometry and Epidemiology, Munich, Germany. ²¹¹Faculty of Medicine, University of Iceland, Reykjavík, Iceland. ²¹²University of Cambridge Metabolic Research Labs, Institute of Metabolic Science Addenbrooke's Hospital, Cambridge, UK. ²¹³Division of Intramural Research, National Heart, Lung, and Blood Institute, Framingham Heart Study, Framingham, Massachusetts, USA.

Abstract

Waist-hip ratio (WHR) is a measure of body fat distribution and a predictor of metabolic consequences independent of overall adiposity. WHR is heritable, but few genetic variants influencing this trait have been identified. We conducted a meta-analysis of 32 genome-wide association studies for WHR adjusted for body mass index (comprising up to 77,167 participants), following up 16 loci in an additional 29 studies (comprising up to 113,636 subjects). We identified 13 new loci in or near *RSPO3*, *VEGFA*, *TBX15-WARS2*, *NFE2L3*, *GRB14*, *DNM3-PIGC*, *ITPR2-SSPN*, *LY86*, *HOXC13*, *ADAMTS9*, *ZNRF3-KREMEN1*, *NISCH-STAB1* and *CPEB4* ($P = 1.9 \times 10^{-9}$ to $P = 1.8 \times 10^{-40}$) and the known signal at *LYPLAL1*. Seven of these loci exhibited marked sexual dimorphism, all with a stronger effect on WHR in women than men (P for sex difference = 1.9×10^{-3} to $P = 1.2 \times 10^{-13}$). These findings provide evidence for multiple loci that modulate body fat distribution independent of overall adiposity and reveal strong gene-by-sex interactions.

Central obesity and body fat distribution, as measured by waist circumference and WHR, are associated with individual risk of type 2 diabetes (T2D)^{1,2} and coronary heart disease³ and with mortality from all causes⁴. These effects are independent of overall adiposity as measured by body mass index (BMI). WHR is of particular interest as a measure of body fat distribution because it integrates the adverse metabolic risk associated with increasing waist circumference with the more protective role of gluteal fat deposition with respect to diabetes, hypertension and dyslipidemia^{5,6}.

There is abundant evidence that body fat distribution is influenced by genetic loci distinct from those regulating BMI and overall adiposity. First, even after accounting for BMI, individual variation in WHR is heritable^{7,8}, with heritability estimates ranging from 22%–61%^{7–10}. Second, the striking abnormalities of regional fat deposition associated with lipodystrophic syndromes demonstrate that genetic variation can have dramatic effects on the development and maintenance of specific fat depots^{11,12}. Third, in a previous genome-wide association analysis, we identified a locus near *LYPLAL1* strongly associated with WHR independent of any effects on BMI¹³, providing proof of principle for the genetic control of body fat distribution distinct from that of overall adiposity.

Within the Genetic Investigation of Anthropometric Traits (GIANT) consortium, we performed a large-scale meta-analysis of genome-wide association studies (GWAS) informative for WHR using adjustment for BMI to focus discovery toward genetic loci associated with body fat distribution rather than overall adiposity^{14–16}.

RESULTS

Genome-wide significant association of WHR with 14 SNPs

We conducted a two-stage study among individuals of European descent (Supplementary Table 1 and Online Methods). In the discovery stage, up to 2,850,269 imputed and genotyped SNPs were examined in 32 GWAS comprising up to 77,167 participants informative for anthropometric measures of body fat distribution. We performed a fixed-effects meta-analysis of WHR, employing study-specific linear regression adjusted for BMI and age, stratified by gender, and using an additive genetic model. After genomic control adjustment per each individual study and in the meta-analysis, these analyses revealed a substantial excess of low P values (Fig. 1a,b).

We selected SNPs representing the top 16 independent (defined as being located >1 Mb apart) regions of association (discovery $P < 1.4 \times 10^{-6}$; Table 1) and evaluated them in 29 additional, independent studies (comprising up to 113,636 individuals) using a mixture of *in silico* data and *de novo* genotyping. In these follow-up studies, 14 of the 16 SNPs analyzed

showed strong directionally consistent evidence for replication ($P < 1.0 \times 10^{-3}$) and ten SNPs reached genome-wide significance ($P < 5.0 \times 10^{-8}$). Joint analysis of the discovery and follow-up results revealed genome-wide significant associations for 14 signals (with P values between 1.9×10^{-9} and 1.8×10^{-40} ; Table 1). Between-study heterogeneity was low ($I^2 < 30\%$) for all but two signals (*GRB14* and *LYPLAL1*; Supplementary Note), and all 14 associations remained genome-wide significant in a random-effects meta-analysis (Supplementary Table 2).

One of these SNPs, rs4846567, is in linkage disequilibrium (LD) ($r^2 = 0.64$, $D' = 0.84$; HapMap European CEU population) with the previously reported WHR-associated variant near *LYPLAL1* (rs2605100)¹³. The remaining 13 loci were in or near genes not previously associated with WHR or other measures of adiposity: *RSPO3*, *VEGFA*, *TBX15-WARS2*, *NFE2L3*, *GRB14*, *DNM3-PIGC*, *ITPR2-SSPN*, *LY86*, *HOXC13*, *ADAMTS9*, *ZNRF3-KREMEN1*, *NISCH-STAB1* and *CPEB4* (Fig. 2). These 14 loci explain 1.03% of the variance in WHR (after adjustment for BMI, age and sex), with each locus contributing from 0.02% (*ZNRF3-KREMEN1*) to 0.14% (*RSPO3*) of the variance based on effect estimates in the follow-up stage.

Sexual dimorphism at several of the WHR loci

Given the known sexual dimorphism of WHR and the evidence from variance decomposition studies that this reflects sex-specific genetic effects¹⁷, we performed sex-specific meta-analyses for the 14 WHR-associated SNPs. These analyses included up to 108,979 women (42,735 in the discovery stage and 66,244 in the follow up) and 82,483 men (34,601 in the discovery and 47,882 in the follow up). In a joint analysis of discovery and follow-up data, 12 of the 14 SNPs reached genome-wide significance in women, but only three SNPs reached genome-wide significance in men (Table 2). At all but one locus (*TBX15-WARS2*), effect-size estimates were numerically greater in women. At seven of the loci (those near *RSPO3*, *VEGFA*, *GRB14*, *LYPLAL1*, *HOXC13*, *ITPR2-SSPN* and *ADAMTS9*), there were marked differences in sex-specific β coefficients (with P values ranging from 1.9×10^{-3} to 1.2×10^{-13}). All loci displayed consistent patterns of sex-specific differences in both the discovery and follow-up studies (Table 2). These 14 loci explain 1.34% of the variance in WHR (after adjustment for BMI and age) in women but only 0.46% of the variance in WHR in men.

Association with other anthropometric measures

By focusing on WHR after adjustment for BMI, our goal was to detect effects on body fat distribution independent of those influencing overall adiposity. As expected, we found very little evidence that known BMI-associated variants were detected in our WHR analysis. Of the ten loci shown to be associated with BMI in previous GWAS^{14,15,18}, only two showed nominally significant ($P < 0.05$) associations for BMI-adjusted WHR in the discovery analysis (*FTO*, rs8050136, $P = 0.03$, $n = 77,074$; *TMEM18*, rs6548238, $P = 3.0 \times 10^{-3}$, $n = 77,016$).

We also tested the 14 WHR-associated SNPs for their effect on BMI using data from up to 242,530 participants available from the GIANT consortium (including most of the studies available for WHR association). Of the 14 WHR loci, four (near *TBX15-WARS2*, *CPEB4*, *LYPLAL1* and *GRB14*) also showed evidence of association with BMI ($4.1 \times 10^{-3} \leq P \leq 3.2 \times 10^{-6}$), with the WHR-increasing allele associated with decreased BMI (Supplementary Table 3). After adding an interaction term of SNP with BMI into the model, we observed that BMI modified the WHR association at the *LY86* locus (P for interaction = 9.5×10^{-5}), with a larger WHR effect among obese individuals compared to non-obese individuals (Supplementary Note).

To determine whether the WHR-associated signals exert their effects primarily through an effect on waist or hip circumference, we performed meta-analyses for these specific phenotypes in the discovery and follow-up studies (**Supplementary Tables 1 and 3**). Overall, we observed stronger associations for hip circumference than for waist circumference. Effect-size estimates were numerically greater for hip circumference than for waist circumference at 11 of the 14 loci, and there were nominal associations ($P < 0.05$) with hip circumference for 12 of the WHR-associated loci but there were only four associations with waist circumference. In both sexes, the WHR-associated loci displaying nominal association with hip circumference always featured the WHR-increasing allele associated with reduced hip circumference. In contrast, we observed sexual dimorphism in the pattern of waist circumference associations. In women, the WHR-increasing allele at all 14 loci was associated with increased waist circumference, whereas this was only true for six of these loci in men (Fig. 3). At *GRB14*, for example, the WHR-increasing allele was associated with increased waist circumference in women ($P = 3.6 \times 10^{-4}$) but with decreased waist circumference in men ($P = 6.8 \times 10^{-3}$). These differences in the relationships between waist circumference, hip circumference and WHR underlie some of the sexual dimorphism in the patterns of WHR association.

Enrichment of association with metabolic traits

We evaluated the 14 WHR-associated loci for their relationships with related metabolic traits using GWAS data provided by trait-specific consortia^{19,21} as well as our *de novo* genotyped follow-up studies. As expected given the sample overlap between this GWAS data and our WHR GWAS data as well as information on known trait correlations (Supplementary Table 4), we observed directionally consistent enrichment of associations ($P < 0.05$) between the 14 WHR-associated alleles and increased triglycerides, low-density lipoprotein (LDL) cholesterol, fasting insulin and homeostasis model assessment (HOMA)-derived measures of insulin resistance (binomial P from 3.2×10^{-4} to 1.8×10^{-8} ; Table 3 and Supplementary Table 5). For example, the WHR-increasing allele at *GRB14* showed strong associations with increased triglycerides ($P = 7.4 \times 10^{-9}$), fasting insulin levels ($P = 5.0 \times 10^{-6}$) and insulin resistance ($P = 1.9 \times 10^{-6}$). Eleven of the 14 WHR-associated loci showed directionally consistent associations with T2D, with three of these loci (at *ADAMTS9*, *NISCH-STAB1* and *ITPR2-SSPN*) reaching nominal significance ($P < 0.05$) (Table 3 and Supplementary Table 5). Because the association signals for correlated traits in this analysis were vulnerable to overestimation given the overlap in the GWAS samples examined, we repeated these analyses and restricted the samples included to those from our *de novo* genotyped follow-up studies. Although this also resulted in a lower sample size, similar patterns of enrichment were still observed (Supplementary Table 5).

Pathway analysis and potential biological roles

To identify potential functional connections and pathway relationships between genes mapping at the WHR-associated loci, we focused on the 95 genes located in a 2-Mb interval centered around each of the 48 independent SNPs that attained $P < 1.0 \times 10^{-5}$ in the WHR discovery studies.

First, we performed a survey of the published literature using GRAIL²² to search for connectivity between the genes and specific keywords that describe these functional connections (Online Methods). Although there was no evidence after correcting for multiple testing that the connectivity between these genes was greater than chance, we identified eight genes with nominal significance ($P < 0.05$) for potential functional connectivity (*PLXND*, *HOXC10*, *TBX15*, *RSPO3*, *HOXC4*, *HOXC6*, *KREMEN1* and *HOXC11*). The keywords associated with these connections included ‘vegf’, ‘homeobox’, ‘patterning’, ‘mesenchyme’, ‘embryonic’, ‘development’ and ‘angiogenesis’.

Additionally, we performed pathway analyses using the PANTHER database²³ based on the same set of 95 genes (Online Methods and Supplementary Note). This analysis generated some evidence for over-representation of ‘developmental processes’ ($P = 5.8 \times 10^{-8}$) and ‘mRNA transcription regulation’ ($P = 2.7 \times 10^{-6}$) but neither of these factors retained nominal significance after adjustment for bias (for example, due to non-random SNP coverage in relation to genes) and the number of biological processes tested (Supplementary Note and Supplementary Table 6).

Finally, we examined the described functional roles of some of the most compelling candidates based on either proximity to the signal or the other analyses described in this paper. These analyses uncovered possible genetic roles in adipocyte development (*TBX15*), pattern formation during embryonic development (*HOXC13*), angiogenesis (*VEGFA*, *RSPO3* and *STAB1*), Wnt and β -catenin signaling (*RSPO3* and *KREMEN1*), insulin signaling (*ADAMTS9*, *GRB14* and *NISCH*), lipase activity (*LYPLAL1*), lipid biosynthesis (*PIGC*) and intracellular calcium signaling (*ITPR2*) (Supplementary Note).

Evaluation of copy number variants and non-synonymous changes

Both common and rare copy number variants (CNVs) have been reported to be associated with overall adiposity^{14, 15, 24, 25}, but the impact of CNVs on fat distribution has not been evaluated previously. To examine the potential contribution of common CNVs to variation in WHR, we looked for evidence of association in our genome-wide association discovery meta-analysis using a set of 6,018 CNV-tagging SNPs which collectively capture >40% of common CNVs that are greater than 1 kb in length^{26, 27} (Online Methods and Supplementary Note).

One CNV-tagging SNP (rs1294421 in *LY86*) was observed among our 14 WHR-associated loci. This SNP is in strong LD ($r^2 = 0.98$) with a 2,832-bp duplication variant (CNVR2760.1)²⁷ located 12 kb from an expressed sequence tag (BC039678) and 87 kb from *LY86* such that the duplication allele is associated with reduced WHR. The duplicated region consists entirely of noncoding sequence but includes part of a predicted enhancer sequence (E.5552.1)²⁸.

To identify other putatively causal variants in our associated regions, we searched for non-synonymous coding SNPs in strong LD (defined as $r^2 > 0.7$) with the most strongly associated SNPs at each locus using data from the HapMap (Build 21) and 1000 Genomes Project (April and August 2009 releases). In this search, one lead SNP (rs6784615, at the *NISCH-STAB1* locus) was correlated with non-synonymous changes in two nearby genes, *DNAH1* (p.Val441Leu, p.Arg1285Trp and p.Arg3809Cys) and *GLYCTK* (p.Leu170Val). Fine-mapping and functional studies will be required to determine whether the *DNAH1* or *GLYCTK* SNPs or the *LY86* CNV are causal for the WHR associations at these loci.

Effect of WHR associations on expression in relevant tissues

Expression quantitative trait locus (eQTL) data can implicate regional transcripts that mediate trait associations, and we therefore examined the 14 WHR-associated loci using eQTL data from human subcutaneous adipose tissue (SAT)²⁹ (two separate sample sets, $n = 610$ and $n = 603$), omental fat³⁰ ($n = 740$), liver³⁰ ($n = 518$), blood²⁹ ($n = 745$) and lymphocytes³¹ ($n = 830$) (Online Methods and Supplementary Note).

At six of the loci, the WHR-associated SNP was either the strongest SNP associated with significant ($P < 1.0 \times 10^{-5}$) expression of a local (within 1 Mb) gene transcript or explained the majority of the association between the most significant eQTL SNP and the gene transcript in conditional analyses (adjusted $P > 0.05$; Table 4). For example, the WHR-associated SNP rs1011731 (near *DNM3-PIGC*) was strongly associated with expression of

PIGC in lymphocytes ($P = 5.9 \times 10^{-10}$); furthermore, rs1011731 is in high LD ($r^2 = 1.00$, $D' = 1.00$ from the HapMap CEU population) with the SNP with the strongest effect on *PIGC* expression (rs991790), and this *cis* eQTL association was abolished by conditioning on rs1011731. These analyses therefore indicate that these two signals are coincident and that *PIGC* is a strong candidate for mediating the WHR association at rs1011731. We found similar evidence for coincidence of the WHR signal with expression for rs984222 (*TBX15* in omental fat), rs1055144 (expressed sequence tag AA553656 in SAT), rs10195252 (*GRB14* in SAT), rs4823006 (*ZNRF3* in SAT and omental fat) and rs6784615 (*STAB1* in blood) (Table 4). Taken together, the overlap between trait association and gene expression at these loci suggests that the WHR associations may be driven through altered expression of *PIGC*, *TBX15*, AA553656, *GRB1*, *ZNRF3* and *STAB1*.

RNA expression of gluteal and abdominal fat tissue

To determine whether genes within the WHR-associated loci showed evidence of differential transcription in distinct fat depots, we compared expression levels in gluteal or abdominal SAT in 49 individuals. We focused on the 15 genes with the strongest credentials for causal involvement (on the basis of proximity to the lead SNP and/or other biological or functional data; Table 1) for which expression data were available. Five of these genes (*RSPO3*, *TBX15*, *ITPR2*, *WARS2* and *STAB1*) were differentially expressed between the two tissues (using an *F* test, corrected for false discovery rate across the 15 expressed genes, $P < 0.05$; Supplementary Table 7). This supports the hypothesis that, at some loci at least, the association with WHR reflects depot-specific differences in expression patterns.

DISCUSSION

Overall, our findings demonstrate that the genetic regulation of body fat distribution involves loci and processes that are largely distinct from those that influence BMI and risk of obesity. This finding is consistent with the evidence that WHR displays substantial heritability even after adjustment for BMI. The loci that emerged from this study display no overlap with those shown to be associated with BMI either in previous reports^{14,16} or in the expanded meta-analysis recently completed by the GIANT consortium³².

Another point of distinction between our findings and those for BMI relates to the evidence for sexual dimorphism that we observed at several of the WHR-associated loci. Sex differences in the regulation of body fat distribution have long been acknowledged without a clear understanding of the underlying molecular mechanisms. These differences become apparent during puberty and are generally attributed to the influence of sex hormones³³. Consistent with our findings, variance decomposition studies have shown that the genetic contribution to the overall variance in WHR, waist and hip circumference is greater in women¹⁷. Although there is some evidence for loci with differential sex effects influencing lipids³⁴, uric acid levels³⁵ and risk of schizophrenia³⁶, we are unaware of prior reports indicating such strong enrichment of female-specific associations for any other phenotype, including BMI³².

The primary objective of genetic discovery efforts is to characterize the specific mechanisms involved in regulating the trait of interest. Despite the considerable challenges associated with moving from common variant association signals to defining causal alleles and pathways, we have identified strong candidates at several of the loci. For example, the *cis* eQTL data implicate *GRB14* as a compelling candidate for the WHR association on chromosome 2, and we were able to show that the same *GRB14* variants are also associated with triglyceride and insulin levels, consistent with previous association of this locus with high-density lipoprotein (HDL) cholesterol³⁷. These inferences about the role of *GRB14* are supported by evidence that *Grb14*-deficient mice exhibit improved glucose homeostasis

despite lower circulating insulin levels, as well as enhanced insulin signaling in liver and skeletal muscle³⁸. The signal near *ADAMTS9* overlaps a previously-reported T2D locus³⁹, and the lead SNP for WHR in our study is identical to the SNP displaying the strongest T2D association in a previous expanded T2D meta-analysis⁴⁰. Given evidence that *ADAMTS9* T2D risk alleles are associated with insulin resistance in peripheral tissues⁴¹, these findings are consistent with a primary effect of *ADAMTS9* variants on body fat distribution. At the chromosome 6 locus, *VEGFA* is the most apparent biological candidate given the presumed role of *VEGFA* as a mediator of adipogenesis⁴² and evidence that serum levels of VEGFA are correlated with obesity^{43, 44}. Finally, at the *TBX15-WARS2* locus, *TBX15* emerges as the strongest candidate based on the *cis* eQTL data in omental fat, marked depot-specific differences in adipose tissue expression in mice and humans and associations between *TBX15* expression in visceral fat and WHR^{45, 46}.

Our efforts to use pathway- and literature-mining approaches to look for functional enrichment of the genes mapping to associated regions met with only limited success but did provide some support for over-representation of developmental processes. Developmental genes have been implicated in fat accumulation and distribution^{45, 46}, and recent evidence supports a link between developmental genes, including *HOXC13* (ref. ⁴⁷) and *TBX15* (refs. ^{45, 48}), and body fat distribution. Developmental genes may in part determine the adipocyte-specific expression patterns that have been observed in different fat depots⁴⁵. Taken together, our findings point to a set of genes influencing body fat distribution that have their principal effects in adipose tissue. This is in contrast to the predominantly central (hypothalamic) processes that are involved in the regulation of BMI and overall adiposity⁴⁹.

By providing new insights into the regulation of body fat distribution, the present study raises a number of issues for future investigation. From the genetic perspective, re-sequencing, dense-array genotyping and fine-mapping approaches will be required to characterize causal variants at the loci we have identified and to support further discoveries that may account for the substantial proportion of genetic variance unexplained by our findings. From the clinical perspective, it will be important to explore the relationship of these variants to more refined measures of body fat distribution derived from detailed imaging studies, to use the variants identified to characterize the causal relationships between body fat distribution and related metabolic and cardiovascular traits and to explore population differences in patterns of body fat distribution. Efforts to tackle overall obesity through therapeutic or lifestyle-based modulation of overall energy balance have proved extremely challenging to implement, and the manipulation of processes associated with more beneficial patterns of fat distribution offers an alternative perspective for future drug discovery.

METHODS

Methods and any associated references are available in the online version of the paper at <http://www.nature.com/naturegenetics/>.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Footnotes

Correspondence should be addressed to C.M.L. (celi@well.ox.ac.uk), K.L.M. (mohlke@med.unc.edu), C.S.F. (foxca@nhlbi.nih.gov), M.I.M. (mark.mccarthy@dr1.ox.ac.uk) or I.M.H. (iris.heid@klinik.uni-regensburg.de).

²¹⁴These authors contributed equally to this work.

²¹⁵These authors jointly directed this work.

AUTHOR CONTRIBUTIONS Writing group: I.B., C.S.F., I.M.H. (lead), C.M.L. (lead), M.I.M., K.L. Mohlke, L.Q., V. Steinthorsdottir, G.T., M.C.Z.

Waist phenotype working group: T.L.A., N.B., I.B., L.A.C., C.M.D., C.S.F., T.B.H., I.M.H., A.U.J., C.M.L. (lead), R.J.F.L., R.M., M.I.M., K.L. Mohlke, L.Q., J.C.R., E.K.S., V. Steinthorsdottir, K. Stefansson, G.T., U.T., C.C.W., T.W., T.W.W., H.E.W., M.C.Z.

Data cleaning and analysis: S.I.B., I.M.H. (lead), E.I., A.U.J., H.L., C.M.L. (lead), R.J.F.L. (lead), J.L., R.M., L.Q., J.C.R., E.K.S., G.T., S.V., M.N.W., E.W., C.J.W., T.W.W., T.W.

Sex-specific analyses: S.I.B., T.E., I.M.H., A.U.J., T.O.K., Z.K., S.L., C.M.L., R.J.F.L., R.M., K.L. Monda, K.E.N., L.Q., J.C.R. (lead), V. Steinthorsdottir, G.T., T.W.W. (lead).

eQTL and expression analyses: S.I.B., A.L.D., C.C.H., J.N.H., F.K., L.M.K., C.M.L., L.L., R.J.F.L., J.L., M.F.M., J.L.M., C.M., G.N., E.E.S., E.K.S., V. Steinthorsdottir, G.T., K.T.Z.

Pathway and CNV analyses: C.M.L., S.A.M., M.I.M., J.N., V. Steinthorsdottir, G.T., B.F.V.

Secondary analyses: S.I.B., I.B.B., N.C., K.E., T.M.F., M.F.F., T.F., M.E.G., J.N.H., E.I., G.L., C.M.L., H.L., R.M., M. Mangino, M.I.M., K.L. Mohlke, D.R.N., J.R.O., S.P., J.R.B.P., J.C.R., A.V.S., E.K.S., P.M.V., M.N.W., C.J.W., R.J.W., E.W., A.R.W., J.Y.

Study-specific analyses: G.R.A., D.A., N.A., T.A., T.L.A., N.B., C.C., P.S.C., L.C., L.A.C., D.I.C., M.N.C., C.M.D., T.E., K.E., E.F., M.F.F., T.F., A.P.G., N.L.G., M.E.G., C. Hayward, N.L.H., I.M.H., J.J.H., A.U.J., Å.J., T. Johnson, J.O.J., J.R.K., M. Kaakinen, K. Kapur, S. Ketkar, J.W.K., P. Kraft, A.T.K., Z.K., J. Kettunen, C. Lamina, R.J.F.L., C. Lecocour, H.L., M.F.L., C.M.L., J.L., R.W.L., R.M., M. Mangino, B.M., K.L. Monda, A.P.M., N.N., K.E.N., D.R.N., J.R.O., K.K.O., C.O., M.J.P., O. Polasek, I. Prokopenko, N.P., M.P., L.Q., J.C.R., N.W.R., S.R., F.R., N.R.R., C.S., L.J.S., K. Silander, E.K.S., K. Stark, S.S., A.V.S., N.S., U.S., V. Steinthorsdottir, D.P.S., I.S., M.L.T., T.M.T., N.J.T., A.T., G.T., A.U., S.V., V. Vitart, L.V., P.M.V., R.M.W., R.W., R.J.W., S.W., M.N.W., C.C.W., C.J.W., T.W.W., A.R.W., J.Y., J.H.Z., M.C.Z.

Study-specific genotyping: D.A., T.L.A., L.D.A., N.B., I.B., A.J.B., E.B., L.L.B., I.B.B., H.C., D.I.C., I.N.M.D., M. Dei, M.R.E., P.E., K.E., N.B.F., M.F., A.P.G., H.G., C.G., E.J.C.G., C.J.G., T. Hansen, A.L.H., N.H., C. Hayward, A.A.H., J.J.H., F.B.H., D.J.H., J.H., W.I., M.R.J., Å.J., J.O.J., J.W.K., P. Kovacs, A.T.K., H.K.K., J. Kettunen, P. Kraft, R.N.L., C.M.L., R.J.F.L., J.L., M.L.L., M.A.M., M. Mangino, W.L.M., M.I.M., J.B.J.M., M.J.N., M.N., D.R.N., K.K.O., C.O., O. Pedersen, L.P., M.J.P., G.P., A.N.P., N.P., L.Q., N.W.R., F.R., N.R.R., C.S., A.J.S., N.S., A.C.S., M.T., B.T., A.U., G.U., V. Vatin, P.M.V., H.W., P.Z.

Study-specific phenotyping: H.A., P.A., D.A., A.M.A., T.L.A., B.B., S.R.B., R.B., E.B., I.B.B., J.P.B., M. Dörr, C.M.D., P.E., M.F.F., C.S.F., T.M.F., M.F., S.G., J.G., L.C.G., T. Hansen, A.S.H., C. Hengstenberg, A.L.H., A.T.H., K.H.H., A. Hofman, F.B.H., D.J.H., B.I., T.I., T. Jørgensen, P.J., M.R.J., Å.J., A.J., A.L.J., J.O.J., F.K., L.K., J. Kuusisto, K. Kvaloy, R.K., S. Ketkar, J.W.K., I.K., S. Koskinen, V.K., M. Kähönen, P. Kovacs, O.L., R.N.L., B.L., J.L., G.M.L., R.J.F.L., T.L., M. Mangino, M.I.M., C.O., B.M.P., O. Pedersen, C.G.P.P., J.F.P., I. Pichler, K.P., O. Polasek, A.P., L.Q., M.R., I.R., O.R., V. Salomaa, J. Saramies, P.E.H.S., K. Silander, N.J.S., J.H.S., T.D.S., D.P.S., R.S., H.M.S., J. Sinisalo, T.T., A.T., M.U., P.V., C.B.V., L.V., J.V., D.R.W., G.B.W., S.H.W., G.W., J.C.W., A.F.W., L.Z., P.Z.

Study-specific management: G.R.A., A.M.A., B.B., Y.B.S., R.N.B., H.B., J.S.B., S.B., M.B., E.B., D.I.B., I.B.B., J.P.B., M.J.C., F.S.C., L.A.C., G.D., C.M.D., S.E., G.E., P.F., C.S.F., T.M.F., L.C.G., V.G., U.G., M.E.G., T. Hansen, C. Hengstenberg, K.H., A. Hamsten, T.B.H., A.T.H., A. Hofman, F.B.H., D.J.H., B.I., T.I., C.I., T. Jørgensen, M.R.J., A.L.J., F.K., K.T.K., W.H.L.K., R.K., J. Kaprio, M. Kähönen, M.L., D.A.L., L.J.L., C.M.L., R.J.F.L., T.L., M. Marre, T.M., A.M.E.T., K.M., M.I.M., K.L. Mohlke, P.B.M., K.E.N., M.S.N., D.R.N., B.O., C.O., O. Pedersen, L.P., B.W.P., P.P.P., B.M.P., L.J.P., T.Q., A.R., I.R., O.R., P.M.R., V. Salomaa, P.S., D.S., A.R.S., N.S., T.D.S., K. Stefansson, D.P.S., A.C.S., M.S., T.T., J.T., U.T., A.T., M.U., A.U., T.T.V., P.V., H.V., J.V., P.M.V., N.J.W., H.E.W., J.F.W., J.C.W., A.F.W.

Steering committee: G.R.A., T.L.A., I.B., S.I.B., M.B., I.B.B., P.D., C.M.D., C.S.F., T.M.F., L.C.G., T. Haritunians, J.N.H. (chair), D.J.H., E.I., R.K., R.J.F.L., M.I.M., K.L. Mohlke, K.E.N., J.R.O., L.P., D.S., D.P.S., U.T., H.E.W.

Note: Supplementary information is available on the Nature Genetics website.

URLs. LocusZoom, <http://csg.sph.umich.edu/locuszoom/>.

COMPETING FINANCIAL INTERESTS The authors declare competing financial interests: details accompany the full-text HTML version of the paper at <http://www.nature.com/naturegenetics/>.

Reprints and permissions information is available online at <http://npg.nature.com/reprintsandpermissions/>.

Acknowledgments

Funding for this study was provided by the Academy of Finland (grants 104781, 120315, 129269, 117797, 121584, 126925, 129418, 129568, 77299, 124243, 213506, 129680, 129494, 10404, 213506, 129680, 114382, 126775, 127437, 129255, 129306, 130326, 209072, 210595, 213225 and 216374); an ADA Mentor-Based Postdoctoral Fellowship grant; Affymetrix, Inc., for genotyping services (N02-HL-6-4278); ALF/LUA Gothenburg; Althingi (the Icelandic Parliament); Amgen; AstraZeneca AB; Augustinus Foundation; Becket Foundation; Biocentrum

Helsinki; Biomedicum Helsinki Foundation; Boston Obesity Nutrition Research Center (DK46200); British Diabetes Association (1192); British Diabetic Association Research; British Heart Foundation (97020, PG/02/128); Busselton Population Medical Research Foundation; Cambridge NIHR Comprehensive Biomedical Research Centre; CamStrad; Chief Scientist Office of the Scottish Government; Contrat Plan Etat Région de France; Danish Centre for Health Technology Assessment; Danish Diabetes Association; Danish Ministry of Internal Affairs and Health; Danish Heart Foundation; Danish Pharmaceutical Association; Danish Research Council; DIAB Core (German Network of Diabetes); Diabetes UK; Donald W. Reynolds Foundation; Dresden University of Technology Funding Grant, Med Drive; EMGO+ institute; Emil and Vera Cornell Foundation; Erasmus Medical Center and Erasmus University, Rotterdam, The Netherlands; Estonian Government SF0180142s08; European Commission (2004310, 212111, 205419, 245536, DG XII, HEALTH-F4-2007-201413, FP7/2007-2013, QLG1-CT-2000-01643, QLG2-CT-2002-01254, LSHG-CT-2006-018947, LSHG-CT-2006-01947, LSHG-CT-2004-512066, LSHM-CT-2007-037273, EU/WLRT-2001-01254, LSHG-CT-2004-518153, SOC 95201408 05F02, Marie Curie Intra-European Fellowship); Federal Ministry of Education and Research, Germany (01ZZ9603, 01ZZ0103, 01ZZ0403, 03ZIK012, 01 EA 9401); Federal State of Mecklenburg-West Pomerania; Finnish Diabetes Research Foundation; Finnish Diabetes Research Society; Finnish Foundation for Pediatric Research; Finnish Foundation of Cardiovascular Research; Finnish Medical Society; Finska Läkaresällskapet; Finnish Ministry of Education; Folkhälsan Research Foundation; Fond Européen pour le Développement Régional; Fondation LeDucq; Foundation for Life and Health in Finland; GEN-AU 'GOLD' from Austria; German Bundesministerium fuer Forschung und Technology (# 01 AK 803 A-H, # 01 IG 07015 G); German National Genome Research Net NGFN2 and NGFNplus (01GS0823, FKZ 01GS0823); German Research Council (KFO-152); GlaxoSmithKline; Göteborg Medical Society; Gyllenberg Foundation; Health Care Centers in Vasa, Närpes and Korsholm; Healthway, Western Australia; Helmholtz Center Munich; Helsinki University Central Hospital; Hjartavernd (the Icelandic Heart Association); Ib Henriksen Foundation; IZKF (B27); Jalmari and Rauha Ahokas Foundation; Juho Vainio Foundation; Juvenile Diabetes Research Foundation International (JDRF); Karolinska Institute and the Stockholm County Council (560183); Knut and Alice Wallenberg Foundation; Lundbeck Foundation Centre of Applied Medical Genomics for Personalized Disease Prediction, Prevention and Care; Knut Krohn, Microarray Core Facility of the Interdisciplinary Centre for Clinical Research (IZKF), University of Leipzig, Germany; Lundberg Foundation; MC Health; Ministry of Cultural Affairs of the Federal State of Mecklenburg-West Pomerania, Germany; South Tyrol Ministry of Health; Ministry of Science, Education and Sport of the Republic of Croatia (216-1080315-0302); Medical Research Council UK (G0000649, G0601261, G9521010D, G0000934, G0500539, G0600331, PrevMetSyn); Montreal Heart Institute Foundation; MRC Centre for Obesity-Related Metabolic Disease; Municipal Health Care Center and Hospital in Jakobstad; Municipality of Rotterdam; Närpes Health Care Foundation; National Health and Medical Research Council of Australia and the Great Wine Estates Auctions; Netherlands Centre for Medical Systems Biology (SPI 56-464-1419); Netherlands Ministry for Health, Welfare and Sports; Netherlands Ministry of Education, Culture and Science; Netherlands Genomics Initiative; Netherlands Consortium for Healthy Aging (050-060-810); Netherlands Organisation of Scientific Research Nederlandse Organisatie voor Wetenschappelijk Onderzoek (NWO) Investments (175.010.2005.011, 911-03-012, 904-61-090, 904-61-193, 480-04-004, 400-05-717); National Institute on Aging Intramural Research Program; US National Institutes of Health (CA047988, CA65725, CA87969, CA49449, CA67262, CA50385, DK075787, DK062370, DK58845, DK072193, K23-DK080145, K99HL094535, N01-HC85079 through N01-HC85086, N01-HG-65403, N01-AG-12100, N01-HC-25195, N01-HC35129, N01-HC15103, N01-HC55222, N01-HC75150, N01-HC45133, N01-HC55015, N01-HC55016, N01-HC-55018, N01-HC-55019, N01-HC-55020, N01-HC-55021, N01-HC-55022, N01-AG-1-2109, HL71981, HG005581, HG002651, HL084729, HL043851, HHSN268200625226C, K23-DK080145, MH084698, P30-DK072488, R01-DK075787, R01 HL087652, R01-HL087641, R01-HL59367, R01-HL086694, R01-HL087647, R01-HL087679, R01-HL087700, R01-AG031890, R01-HL088119, R01-DK068336, R01-DK075681, R01-DK-073490, R01-DK075787, R01-MH63706, U01-HL72515, U01-GM074518, U01-HL084756, U01-HG004399, U01-CA098233, UL1-RR025005, UL1-RR025005, U01-HG004402, U01-DK062418, U01 HL080295, T32-HG00040, 263-MA-410953, 1RL1-MH083268-01, intramural project 1Z01-HG000024); National Institute for Health Research (NIHR); Neuroscience Campus Amsterdam; Novo Nordisk Foundation; Novo Nordisk Inc., Research Foundation of Copenhagen County; Ollqvist Foundation; Paavo Nurmi Foundation; Päivikki and Sakari Sohlberg Foundation; Pew Scholarship for the Biomedical Sciences; Perklén Foundation; Petrus and Augusta Hedlunds Foundation; Research Institute for Diseases in the Elderly (014-93-015, RIDE, RIDE2); Sahlgrenska Center for Cardiovascular and Metabolic Research (CMR, A305:188); Siemens Healthcare, Erlangen, Germany; Signe and Ane Gyllenberg Foundation; Sigrid Juselius Foundation; Social Insurance Institution of Finland; Social Ministry of the Federal State of Mecklenburg-West Pomerania, Germany; South Tyrolean Sparkasse Foundation; State of Bavaria, Germany; Support for Science Funding programme; Swedish Cultural Foundation in Finland; Swedish Foundation for Strategic Research (SSF); Swedish Heart-Lung Foundation; Swedish Medical Research Council (8691, K2007-66X-20270-01-3, K2010-54X-09894-19-3); Swedish Society of Medicine; Swiss National Science Foundation (33CSO-122661); the Royal Society; the Royal Swedish Academy of Science; Torsten and Ragnar Söderberg's Foundation; Turku University Hospitals; UK Department of Health Policy Research Programme; University and Research of the Autonomous Province of Bolzano; University Hospital Medical funds to Tampere; University Hospital Oulu, Biocenter, University of Oulu, Finland (75617); Västra Götaland Foundation; Wellcome Trust (077016/Z/05/Z, 068545/Z/02, 072960, 076113, 083270, 085301, 079557, 081682, 075491, 076113/B/04/Z, 091746/Z/10/Z, 079895, WT086596/Z/08/Z, WT Research Career Development Fellowship; WT Career Development Award); Western Australian Genetic

Epidemiology Resource and the Western Australian DNA Bank (both National Health and Medical Research Council of Australia Enabling Facilities); Yrjö Jahnsson Foundation.

References

1. Carey VJ, et al. Body fat distribution and risk of non-insulin-dependent diabetes mellitus in women. The Nurses' Health Study. *Am. J. Epidemiol.* 1997; 145:614–619. [PubMed: 9098178]
2. Wang Y, Rimm EB, Stampfer MJ, Willett WC, Hu FB. Comparison of abdominal adiposity and overall obesity in predicting risk of type 2 diabetes among men. *Am. J. Clin. Nutr.* 2005; 81:555–563. [PubMed: 15755822]
3. Canoy D. Distribution of body fat and risk of coronary heart disease in men and women. *Curr. Opin. Cardiol.* 2008; 23:591–598. [PubMed: 18830075]
4. Pischon T, et al. General and abdominal adiposity and risk of death in Europe. *N. Engl. J. Med.* 2008; 359:2105–2120. [PubMed: 19005195]
5. Snijder MB, et al. Associations of hip and thigh circumferences independent of waist circumference with the incidence of type 2 diabetes: the Hoorn Study. *Am. J. Clin. Nutr.* 2003; 77:1192–1197. [PubMed: 12716671]
6. Snijder MB, et al. Trunk fat and leg fat have independent and opposite associations with fasting and postload glucose levels: the Hoorn study. *Diabetes Care.* 2004; 27:372–377. [PubMed: 14747216]
7. Mills GW, et al. Heritability estimates for beta cell function and features of the insulin resistance syndrome in UK families with an increased susceptibility to type 2 diabetes. *Diabetologia.* 2004; 47:732–738. [PubMed: 15298351]
8. Souren NY, et al. Anthropometry, carbohydrate and lipid metabolism in the East Flanders Prospective Twin Survey: heritabilities. *Diabetologia.* 2007; 50:2107–2116. [PubMed: 17694296]
9. Rose KM, Newman B, Mayer-Davis EJ, Selby JV. Genetic and behavioral determinants of waist-hip ratio and waist circumference in women twins. *Obes. Res.* 1998; 6:383–392. [PubMed: 9845227]
10. Selby JV, et al. Genetic and behavioral influences on body fat distribution. *Int. J. Obes.* 1990; 14:593–602. [PubMed: 2228394]
11. Agarwal AK, Garg A. Genetic disorders of adipose tissue development, differentiation, and death. *Annu. Rev. Genomics Hum. Genet.* 2006; 7:175–199. [PubMed: 16722806]
12. Garg A. Acquired and inherited lipodystrophies. *N. Engl. J. Med.* 2004; 350:1220–1234. [PubMed: 15028826]
13. Lindgren CM, et al. Genome-wide association scan meta-analysis identifies three loci influencing adiposity and fat distribution. *PLoS Genet.* 2009; 5:e1000508. [PubMed: 19557161]
14. Thorleifsson G, et al. Genome-wide association yields new sequence variants at seven loci that associate with measures of obesity. *Nat. Genet.* 2009; 41:18–24. [PubMed: 19079260]
15. Willer CJ, et al. Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. *Nat. Genet.* 2009; 41:25–34. [PubMed: 19079261]
16. Loos RJ, et al. Common variants near MC4R are associated with fat mass, weight and risk of obesity. *Nat. Genet.* 2008; 40:768–775. [PubMed: 18454148]
17. Zillikens MC, et al. Sex-specific genetic effects influence variation in body composition. *Diabetologia.* 2008; 51:2233–2241. [PubMed: 18839131]
18. Frayling TM, et al. A common variant in the *FTO* gene is associated with body mass index and predisposes to childhood and adult obesity. *Science.* 2007; 316:889–894. [PubMed: 17434869]
19. Kathiresan S, et al. Common variants at 30 loci contribute to polygenic dyslipidemia. *Nat. Genet.* 2009; 41:56–65. [PubMed: 19060906]
20. Dupuis J, et al. New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. *Nat. Genet.* 2010; 42:105–116. [PubMed: 20081858]
21. Saxena R, et al. Genetic variation in *GIPR* influences the glucose and insulin responses to an oral glucose challenge. *Nat. Genet.* 2010; 42:142–148. [PubMed: 20081857]
22. Raychaudhuri S, et al. Identifying relationships among genomic disease regions: predicting genes at pathogenic SNP associations and rare deletions. *PLoS Genet.* 2009; 5:e1000534. [PubMed: 19557189]

23. Thomas PD, et al. PANTHER: a library of protein families and subfamilies indexed by function. *Genome Res.* 2003; 13:2129–2141. [PubMed: 12952881]
24. Bochukova EG, et al. Large, rare chromosomal deletions associated with severe early-onset obesity. *Nature.* 2010; 463:666–670. [PubMed: 19966786]
25. Walters RG, et al. A new highly penetrant form of obesity due to deletions on chromosome 16p11.2. *Nature.* 2010; 463:671–675. [PubMed: 20130649]
26. Conrad DF, et al. Origins and functional impact of copy number variation in the human genome. *Nature.* 2010; 464:704–712. [PubMed: 19812545]
27. Wellcome Trust Case Control Consortium, et al. Genome-wide association study of CNVs in 16,000 cases of eight common diseases and 3,000 shared controls. *Nature.* 2010; 464:713–720. [PubMed: 20360734]
28. Pennacchio LA, Loots GG, Nobrega MA, Ovcharenko I. Predicting tissue-specific enhancers in the human genome. *Genome Res.* 2007; 17:201–211. [PubMed: 17210927]
29. Emilsson V, et al. Genetics of gene expression and its effect on disease. *Nature.* 2008; 452:423–428. [PubMed: 18344981]
30. Schadt EE, et al. Mapping the genetic architecture of gene expression in human liver. *PLoS Biol.* 2008; 6:e107. [PubMed: 18462017]
31. Dixon AL, et al. A genome-wide association study of global gene expression. *Nat. Genet.* 2007; 39:1202–1207. [PubMed: 17873877]
32. Speliotes EK, et al. Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat. Genet.* Oct 10.2010 advance online publication, doi:10.1038/ng.686. [PubMed: 20935630]
33. Wells JC. Sexual dimorphism of body composition. *Best Pract. Res. Clin. Endocrinol. Metab.* 2007; 21:415–430. [PubMed: 17875489]
34. Aulchenko YS, et al. Loci influencing lipid levels and coronary heart disease risk in 16 European population cohorts. *Nat. Genet.* 2009; 41:47–55. [PubMed: 19060911]
35. Döring A, et al. *SLC2A9* influences uric acid concentrations with pronounced sex-specific effects. *Nat. Genet.* 2008; 40:430–436. [PubMed: 18327256]
36. Shifman S, et al. Genome-wide association identifies a common variant in the *reelin* gene that increases the risk of schizophrenia only in women. *PLoS Genet.* 2008; 4:e28. [PubMed: 18282107]
37. Ridker PM, et al. Polymorphism in the *CETP* gene region, HDL cholesterol, and risk of future myocardial infarction: genomewide analysis among 18,245 initially healthy women from the Women's Genome Health Study. *Circ. Cardiovasc. Genet.* 2009; 2:26–33. [PubMed: 20031564]
38. Cooney GJ, et al. Improved glucose homeostasis and enhanced insulin signalling in *Grb14*-deficient mice. *EMBO J.* 2004; 23:582–593. [PubMed: 14749734]
39. Zeggini E, et al. Meta-analysis of genome-wide association data and large-scale replication identifies additional susceptibility loci for type 2 diabetes. *Nat. Genet.* 2008; 40:638–645. [PubMed: 18372903]
40. Voight BF, et al. Twelve type 2 diabetes susceptibility loci identified through large-scale association analysis. *Nat. Genet.* 2010; 42:579–589. [PubMed: 20581827]
41. Boesgaard TW, et al. Variant near *ADAMTS9* known to associate with type 2 diabetes is related to insulin resistance in offspring of type 2 diabetes patients–EUGENE2 study. *PLoS ONE.* 2009; 4:e7236. [PubMed: 19789630]
42. Nishimura S, et al. Adipogenesis in obesity requires close interplay between differentiating adipocytes, stromal cells, and blood vessels. *Diabetes.* 2007; 56:1517–1526. [PubMed: 17389330]
43. Silha JV, Krsek M, Sucharda P, Murphy LJ. Angiogenic factors are elevated in overweight and obese individuals. *Int. J. Obes. (Lond).* 2005; 29:1308–1314. [PubMed: 15953938]
44. García de la Torre N, et al. Effects of weight loss after bariatric surgery for morbid obesity on vascular endothelial growth factor-A, adipocytokines, and insulin. *J. Clin. Endocrinol. Metab.* 2008; 93:4276–4281. [PubMed: 18713823]
45. Gesta S, et al. Evidence for a role of developmental genes in the origin of obesity and body fat distribution. *Proc. Natl. Acad. Sci. USA.* 2006; 103:6676–6681. [PubMed: 16617105]

46. Gesta S, Tseng YH, Kahn CR. Developmental origin of fat: tracking obesity to its source. *Cell*. 2007; 131:242–256. [PubMed: 17956727]
47. Lanctôt C, Kaspar C, Cremer T. Positioning of the mouse *Hox* gene clusters in the nuclei of developing embryos and differentiating embryoid bodies. *Exp. Cell Res.* 2007; 313:1449–1459. [PubMed: 17346703]
48. Candille SI, et al. Dorsoventral patterning of the mouse coat by *Tbx15*. *PLoS Biol.* 2004; 2:E3. [PubMed: 14737183]
49. O’Rahilly S. Human genetics illuminates the paths to metabolic disease. *Nature*. 2009; 462:307–314. [PubMed: 19924209]

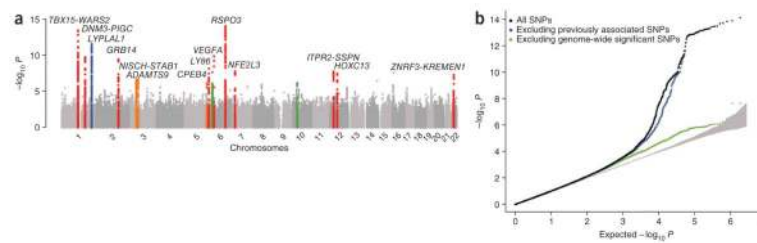


Figure 1.

Genome-wide association analyses for WHR in discovery studies. (a) Manhattan plot shows results of the WHR association meta-analysis in discovery studies (with P values on the y axis and the SNP genomic position on the x axis). Colored genomic loci indicate significant association ($P < 5 \times 10^{-8}$) detected previously (blue)¹³, in our GWAS stage (red) and after the meta-analysis combining GWAS data with that from the follow-up studies (orange). Two loci tested in the follow-up stage did not achieve genome-wide significance (green). (b) Quantile-quantile plot of SNPs for the discovery meta-analysis of WHR (black) and after removing SNPs within 1 Mb of either the recently reported *LYPLAL1* signal (blue) or the 14 significant associations (green). The gray area represents the 95% CI around the test statistic under the null distribution.

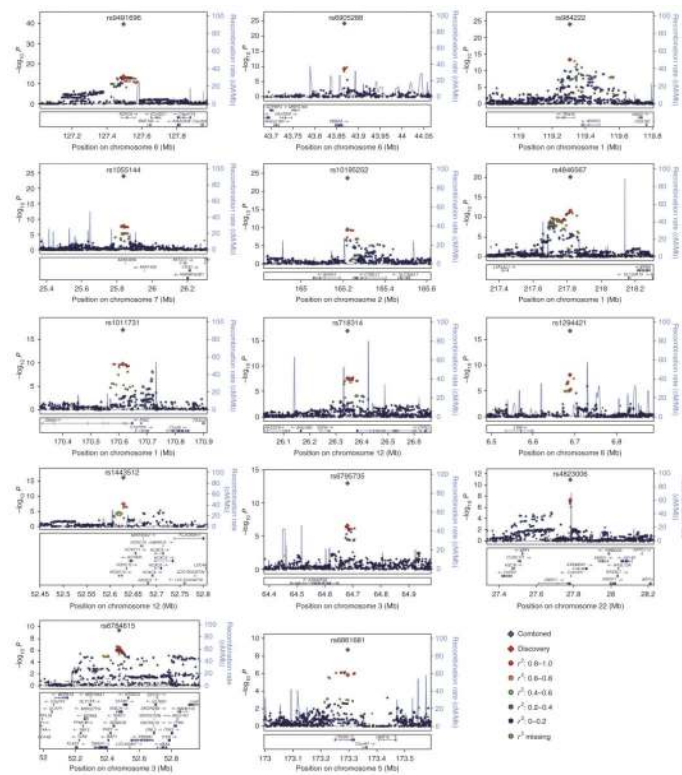


Figure 2.

Regional plots of 14 loci with genome-wide significant association. Shown is the SNP association with WHR in the meta-analysis of discovery studies for 14 loci (with $-\log_{10} P$ values on the y axis and the SNP genomic position on the x axis). In each panel, an index SNP is denoted with a purple diamond and plotted using the P attained across discovery and follow-up data (Table 1). Estimated recombination rates are plotted in blue. SNPs are colored to reflect LD with the index SNP (pairwise r^2 values from HapMap CEU). Gene and microRNA annotations are from the UCSC genome browser.

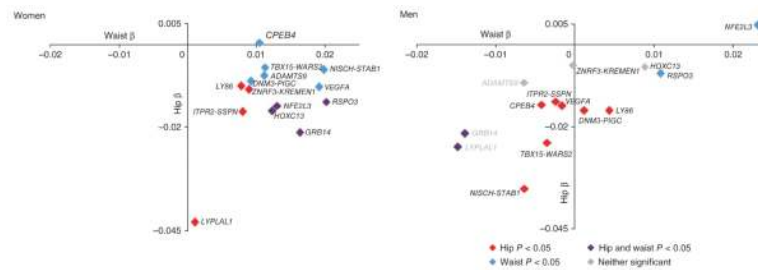


Figure 3.

Association of the 14 WHR loci with waist and hip circumference. β coefficients for waist circumference (WC, x axis) and hip circumference (HIP, y axis) in women and men derived from the joint discovery and follow-up analysis. P for WC and HIP are represented by color. In men, gray gene labels refer to those SNPs that were not significant in the male-specific WHR analysis. More details can be found in Supplementary Table 3.

Fourteen SNPs associated with WHr at genome-wide significant levels

Table 1

SNP	Chr.	Position (h36)	Nearby genes	EA ^a	EAF ^b	Discovery			Follow-up			Combined		
						P	β	n	P	β	n	P	β	n
SNPs evaluated in follow up achieving genome-wide significance														
rs9491696	6	127,494,332	RSPO3	G	0.520	2.10 × 10 ⁻¹⁴	0.037	77,164	3.27 × 10 ⁻²⁸	0.045	113,582	1.84 × 10 ⁻⁴⁰	0.042	
rs6905288	6	43,866,851	VEGFA	A	0.562	4.72 × 10 ⁻¹⁰	0.033	77,129	1.18 × 10 ⁻¹⁶	0.039	95,430	5.88 × 10 ⁻²⁵	0.036	
rs984222	1	119,305,366	TBX15-WARS2	G	0.365	3.81 × 10 ⁻¹⁴	0.037	77,167	1.56 × 10 ⁻¹²	0.031	109,623	8.69 × 10 ⁻²⁵	0.034	
rs1055144	7	25,837,634	NFE2L3	T	0.210	1.49 × 10 ⁻⁸	0.034	77,145	3.26 × 10 ⁻¹⁸	0.043	113,636	9.97 × 10 ⁻²⁵	0.040	
rs10195252	2	165,221,337	GIRB14	T	0.599	3.23 × 10 ⁻¹⁰	0.031	77,119	3.18 × 10 ⁻¹⁶	0.036	102,449	2.09 × 10 ⁻²⁴	0.033	
rs4846567	1	217,817,340	LYPLAL1	G	0.283	2.37 × 10 ⁻¹²	0.037	77,167	3.15 × 10 ⁻¹⁰	0.032	91,820	6.89 × 10 ⁻²¹	0.034	
rs1011731	1	170,613,171	DNM3-PIGC	G	0.572	1.72 × 10 ⁻¹⁰	0.031	77,094	7.47 × 10 ⁻⁹	0.026	92,018	9.51 × 10 ⁻¹⁸	0.028	
rs718314	12	26,344,550	ITPR2-SSPN	G	0.741	2.41 × 10 ⁻⁸	0.031	77,167	1.49 × 10 ⁻¹⁰	0.030	107,503	1.14 × 10 ⁻¹⁷	0.030	
rs1294421	6	6,688,148	LY86	G	0.387	6.31 × 10 ⁻⁹	0.029	77,154	2.69 × 10 ⁻¹⁰	0.028	102,189	1.75 × 10 ⁻¹⁷	0.028	
rs1443512	12	52,628,951	HOXC13	A	0.239	3.33 × 10 ⁻⁸	0.031	77,165	2.92 × 10 ⁻¹⁰	0.030	112,353	6.38 × 10 ⁻¹⁷	0.031	
rs6795735	3	64,680,405	ADAMTS9	C	0.406	2.47 × 10 ⁻⁷	0.025	77,162	6.75 × 10 ⁻⁸	0.026	84,480	9.79 × 10 ⁻¹⁴	0.025	
rs4823006	22	27,781,671	ZNRF3-KREMEN1	A	0.569	4.47 × 10 ⁻⁸	0.027	77,086	2.41 × 10 ⁻⁵	0.019	93,911	1.10 × 10 ⁻¹¹	0.023	
rs6784615	3	52,481,466	NISCH-STAB1	T	0.941	3.18 × 10 ⁻⁷	0.052	76,859	1.56 × 10 ⁻⁴	0.036	109,028	3.84 × 10 ⁻¹⁰	0.043	
rs6861681	5	173,295,064	CPEB4	A	0.340	1.40 × 10 ⁻⁶	0.026	77,164	2.13 × 10 ⁻⁴	0.019	85,722	1.91 × 10 ⁻⁹	0.022	
Further SNPs evaluated in follow up but not achieving genome-wide significance in the combined analysis														
rs2076529	6	32,471,933	BTNL2	C	0.570	2.22 × 10 ⁻⁸	0.041	34,532	0.012	0.011	92,778	3.71 × 10 ⁻⁷	0.020	
rs7081678	10	32,030,629	ZEB1	A	0.085	5.76 × 10 ⁻⁷	0.045	76,270	0.094	0.013	100,527	5.57 × 10 ⁻⁶	0.027	

P values and *β* coefficients (per change of WHr-increasing allele) for the association with WHr on the inverse normal transformed ranked scale in the meta-analyses of discovery studies (up to 77,167 subjects), follow-up studies (up to 113,636 subjects) and both combined (up to 190,781 subjects). Fourteen of the sixteen SNPs examined in the follow-up samples showed genome-wide significant results (*P* < 5 × 10⁻⁸) in the combined analysis. *P* values in the discovery stage were genomic control corrected per study and in the meta-analysis. Details on between-study heterogeneity are given in Supplementary Table 1c.

^aEA, effect allele (WHR-increasing allele on the forward strand).

^bEAF, effect allele frequency. Chr., chromosome.

Table 2

Evidence of sex-differences in the WHr association at seven of the 14 associated loci

SNP	Nearby genes	Men						Women						Sex difference					
		Discovery			Follow up			Combined			Discovery			Follow up			Combined		
		P	β	P	P	β	P	P	β	P	P	β	P	β	P	P	β	P	
rs9491696	<i>RSPO3</i>	1.68 × 10 ⁻⁴	0.026	6.97 × 10 ⁻⁹	0.036	1.05 × 10 ⁻¹¹	0.031	1.62 × 10 ⁻¹²	0.047	8.84 × 10 ⁻²²	0.053	1.93 × 10 ⁻³²	0.050	1.94 × 10 ⁻³					
rs6905288	<i>VEGFA</i>	0.066	0.013	2.09 × 10 ⁻⁴	0.025	7.38 × 10 ⁻⁵	0.020	7.72 × 10 ⁻¹³	0.052	3.14 × 10 ⁻¹⁵	0.051	2.27 × 10 ⁻²⁶	0.052	5.20 × 10 ⁻⁶					
rs984222	<i>TBX15</i> , <i>WARS2</i>	3.32 × 10 ⁻⁹	0.041	2.43 × 10 ⁻⁵	0.029	9.41 × 10 ⁻¹³	0.035	1.21 × 10 ⁻⁷	0.036	1.33 × 10 ⁻⁸	0.033	1.02 × 10 ⁻¹⁴	0.034	0.951					
rs1055144	<i>NFE2L3</i>	6.00 × 10 ⁻⁴	0.029	5.67 × 10 ⁻⁸	0.040	2.52 × 10 ⁻¹⁰	0.035	2.34 × 10 ⁻⁶	0.040	7.13 × 10 ⁻¹²	0.046	1.41 × 10 ⁻¹⁶	0.044	0.270					
rs10195252	<i>GRB14</i>	0.201	0.009	0.114	0.011	0.043	0.010	6.33 × 10 ⁻¹⁵	0.053	4.95 × 10 ⁻²¹	0.054	3.84 × 10 ⁻³⁴	0.054	1.41 × 10 ⁻¹¹					
rs4846567	<i>LYPLAL1</i>	0.191	0.010	0.982	0.000	0.358	0.005	4.84 × 10 ⁻¹⁸	0.064	8.12 × 10 ⁻¹⁷	0.055	4.95 × 10 ⁻³³	0.059	1.18 × 10 ⁻¹³					
rs1011731	<i>DNM3</i> - <i>PIGC</i>	4.88 × 10 ⁻⁷	0.034	1.95 × 10 ⁻³	0.022	7.81 × 10 ⁻⁹	0.028	2.13 × 10 ⁻⁵	0.028	7.03 × 10 ⁻⁷	0.030	6.90 × 10 ⁻¹¹	0.029	0.855					
rs718314	<i>ITPR2</i> - <i>SSPN</i>	0.177	0.010	2.02 × 10 ⁻³	0.022	1.41 × 10 ⁻³	0.017	8.29 × 10 ⁻¹⁰	0.047	4.21 × 10 ⁻⁹	0.038	2.41 × 10 ⁻¹⁷	0.042	4.67 × 10 ⁻⁴					
rs1294421	<i>LY86</i>	4.18 × 10 ⁻³	0.020	7.00 × 10 ⁻⁶	0.030	1.63 × 10 ⁻⁷	0.025	3.44 × 10 ⁻⁸	0.038	7.32 × 10 ⁻⁶	0.026	2.40 × 10 ⁻¹²	0.031	0.357					
rs1443512	<i>HOXC13</i>	0.184	0.011	9.74 × 10 ⁻⁴	0.024	9.45 × 10 ⁻⁴	0.018	1.43 × 10 ⁻⁹	0.048	3.09 × 10 ⁻⁸	0.035	6.38 × 10 ⁻¹⁶	0.040	2.23 × 10 ⁻³					
rs6795735	<i>AD</i> , <i>AMTS9</i>	0.011	0.017	0.614	0.004	0.027	0.011	7.85 × 10 ⁻⁷	0.033	2.95 × 10 ⁻¹¹	0.042	1.92 × 10 ⁻¹⁶	0.038	8.50 × 10 ⁻⁵					
rs4823006	<i>ZNRF3</i> - <i>KRE</i> , <i>MEN1</i>	6.87 × 10 ⁻³	0.019	0.094	0.012	1.94 × 10 ⁻³	0.015	6.86 × 10 ⁻⁸	0.037	3.81 × 10 ⁻⁵	0.024	3.24 × 10 ⁻¹¹	0.030	0.032					
rs6784615	<i>NISCH</i> , <i>STAB1</i>	1.51 × 10 ⁻³	0.045	0.033	0.032	1.68 × 10 ⁻⁴	0.039	6.23 × 10 ⁻⁵	0.057	1.72 × 10 ⁻³	0.039	6.01 × 10 ⁻⁷	0.047	0.574					
rs6861681	<i>CPEB4</i>	1.88 × 10 ⁻³	0.023	0.045	0.015	3.03 × 10 ⁻⁴	0.019	2.14 × 10 ⁻⁴	0.027	1.58 × 10 ⁻³	0.021	1.55 × 10 ⁻⁶	0.024	0.555					

P values and *β* coefficients (per change of WHr-increasing allele in the sex-combined analysis as in Table 1) for the WHr association are given for the discovery (up to 34,601 men and 42,735 women), the follow-up (up to 47,882 men and 65,780 women) and the combined meta-analysis (up to 81,301 men and 107,429 women). Also given are the *P* values for testing for difference between sex-specific *β* coefficients in the combined meta-analysis; SNPs with *P* for sex difference < 3.6 × 10⁻³ (0.05/14) were considered to show a significant sex difference.

Table 3
WHR signals show enrichment of association with other traits related to metabolic disorders

Trait	Sample size ^d	SNPs in concordant direction ^b		SNPs in concordant direction with $P < 0.05^c$	
		n	P	n	P
Triglycerides	43,826	14	6.10×10^{-5}	7	1.79×10^{-8}
HDL-C	45,561	13	9.16×10^{-4}	4	3.20×10^{-4}
LDL-C	43,889	10	0.090	1	0.298
Fasting glucose	63,849	10	0.090	1	0.298
Fasting insulin	54,883	13	9.16×10^{-4}	5	1.62×10^{-5}
HOMA-IR	53,625	13	9.16×10^{-4}	6	6.17×10^{-7}
2 h glucose	27,011	7	0.605	0	1.000
Type 2 diabetes	10,128 ^d	11	0.029	3	4.62×10^{-3}

The 14 WHR SNPs were tested for association with other traits by meta-analysis of GWAS data from previous reports^{19, 21, 39} together with our non-overlapping *de novo* genotyped follow-up studies. HDL-C, high density lipoprotein cholesterol; LDL-C, low density lipoprotein cholesterol; HOMA-IR, index of insulin resistance; 2 h glucose, glucose levels 2 h after an oral glucose challenge.

^aMaximum number of subjects available for any of the 14 SNPs.

^bNumber of the 14 SNPs for which the WHR-increasing allele is associated with the trait in the concordant direction (that is, increased levels, except for HDL-C) and corresponding binomial P -value to test whether this number is greater than that expected by chance and not accounting for the correlation between the traits.

^cNumber of SNPs in concordant direction that show $P < 0.05$ for the association with the trait and the corresponding binomial P -value as in ^b.

^d4,549 cases, 5579 controls.

Expression quantitative trait locus analysis for 11 of the 14 WHR signals

WHR SNP	Tissue	Gene	Effect ^a	WHR SNP association with transcript (<i>P</i>)			Transcript peak SNP ^b	Peak SNP association with transcript (<i>P</i>)		
				Unadj.	Adj. for peak SNP	LD (<i>r</i> ²) ^c		Unadj.	Adj. for WHR SNP	
rs9491696	SAT-D	<i>RSPO3</i>	+	1.10 × 10 ⁻⁷	0.03		rs1936795	0.26	2.20 × 10 ⁻¹³	7.40 × 10 ⁻⁸
rs984222	Omental	<i>TBX15</i>	+	7.90 × 10 ⁻¹⁰	1.00		rs984222	1.00	7.90 × 10 ⁻¹⁰	1.00
	Omental	<i>WARS2</i>	+	5.11 × 10 ⁻³⁶	0.03		rs10802075	0.27	1.31 × 10 ⁻¹⁶³	1.33 × 10 ⁻⁸⁸
	Subcutaneous fat	<i>WARS2</i>	+	1.67 × 10 ⁻²⁵	0.01		rs10802075	0.22	3.88 × 10 ⁻¹¹⁰	1.01 × 10 ⁻⁶³
	Lymphocytes	<i>WARS2</i>	-	4.30 × 10 ⁻¹⁸	5.47 × 10 ⁻⁵		rs2645305	0.27	5.57 × 10 ⁻⁴⁰	6.88 × 10 ⁻²⁶
	Liver	<i>WARS2</i>	+	2.57 × 10 ⁻¹⁷	0.07		rs1057990	0.26	6.69 × 10 ⁻⁵⁹	1.97 × 10 ⁻³²
	SAT-D	<i>WARS2</i>	+	1.10 × 10 ⁻¹⁸	0.51		rs1057990	0.26	5.80 × 10 ⁻¹³⁰	5.80 × 10 ⁻¹⁰⁰
	Blood	<i>WARS2</i>	+	6.10 × 10 ⁻¹⁷	0.11		rs1057990	0.26	6.30 × 10 ⁻⁷⁵	1.10 × 10 ⁻⁵⁴
rs1055144	SAT-D	AA553656 ^d	-	1.20 × 10 ⁻¹¹	0.96		rs7798002	0.95	7.20 × 10 ⁻¹²	0.32
	SAT-M	AA553656 ^d	-	2.46 × 10 ⁻⁷	0.65		rs1451385	0.77	5.93 × 10 ⁻⁸	0.38
rs10195252	SAT-D	<i>GRB14</i>	+	4.40 × 10 ⁻¹¹	1.00		rs10195252	1.00	4.40 × 10 ⁻¹¹	1.00
	SAT-M	<i>GRB14</i>	+	5.51 × 10 ⁻⁶	1.00		rs10184004	1.00	5.51 × 10 ⁻⁶	1.00
	Omental	<i>GRB14</i>	+	1.02 × 10 ⁻¹³	1.00		rs10195252	1.00	1.02 × 10 ⁻¹³	1.00
	SAT-M	<i>SLC38A11</i>	-	3.93 × 10 ⁻⁶	0.66		rs10184126	0.18	7.76 × 10 ⁻⁴⁴	8.57 × 10 ⁻³⁴
	SAT-D	<i>SLC38A11</i>	-	3.70 × 10 ⁻⁹	0.35		rs10184126	0.18	2.40 × 10 ⁻⁹⁴	7.40 × 10 ⁻⁸²
rs1011731	Blood	<i>C1orf105</i>	+	3.80 × 10 ⁻¹⁶	0.20		rs2157451	0.28	1.30 × 10 ⁻³³	8.20 × 10 ⁻¹⁸
	Lymphocytes	<i>PIGC</i>	-	5.87 × 10 ⁻¹⁰	1.00		rs991790	1.00	5.65 × 10 ⁻¹⁰	1.00
rs718314	Lymphocytes	<i>ITPR2</i>	+	1.79 × 10 ⁻⁹	0.98		rs7976877	0.45	2.21 × 10 ⁻¹⁸	1.91 × 10 ⁻⁶
	Blood	<i>ITPR2</i>	-	2.40 × 10 ⁻⁹	0.20		rs2570	0.41	2.40 × 10 ⁻³⁷	1.80 × 10 ⁻²⁸
rs1294421	SAT-M	<i>BC039678</i>	-	2.43 × 10 ⁻⁷	0.38		rs1294404	0.64	1.89 × 10 ⁻¹⁶	3.42 × 10 ⁻⁴
	Omental	<i>BC039678</i>	-	1.09 × 10 ⁻⁶	0.33		rs912056	0.71	8.28 × 10 ⁻¹⁷	4.26 × 10 ⁻⁵
rs6795735	SAT-D	<i>ADAMTS9</i>	-	1.50 × 10 ⁻⁶	0.04		rs7372321	0.11	1.10 × 10 ⁻⁹	2.30 × 10 ⁻⁵
	Omental	<i>AK022320</i>	-	7.99 × 10 ⁻¹⁵	0.64		rs4521216	0.02	5.15 × 10 ⁻⁴²	1.49 × 10 ⁻¹⁹
	SAT-D	<i>AK022320</i>	-	2.24 × 10 ⁻¹⁰	0.98		rs4521216	0.02	9.62 × 10 ⁻³⁷	7.58 × 10 ⁻¹⁹

WHR SNP	Tissue	Gene	Effect ^a	WHR SNP association with transcript (<i>P</i>)		Transcript peak SNP ^b	Peak SNP association with transcript (<i>P</i>)	
				Unadj.	Adj. for peak SNP		Unadj.	Adj. for WHR SNP
rs4823006	SAT-D	<i>ZNRF3</i>	–	2.40 × 10 ^{–8}	0.63	rs3178915	6.70 × 10 ^{–11}	8.90 × 10 ^{–4}
	SAT-M	<i>ZNRF3</i>	–	1.08 × 10 ^{–18}	0.93	rs6005975	1.59 × 10 ^{–19}	0.50
	Omental	<i>ZNRF3</i>	–	9.13 × 10 ^{–18}	0.98	rs6005975	6.07 × 10 ^{–21}	0.27
rs6784615	Blood	<i>STAB1</i>	+	2.80 × 10 ^{–9}	0.32	rs9846089	9.40 × 10 ^{–10}	0.08
rs6861681	Lymphocytes	<i>CPEB4</i>	+	3.79 × 10 ^{–22}	0.89	rs7705502	4.95 × 10 ^{–29}	2.00 × 10 ^{–3}
	Blood	<i>HMP19</i>	+	1.60 × 10 ^{–16}	0.97	rs10516107	1.10 × 10 ^{–21}	4.30 × 10 ^{–6}

Association between the 14 WHR SNPs and expression of transcripts located within 1 Mb of the WHR SNP in two sets of abdominal subcutaneous adipose tissue (SAT-D from deCODE and SAT-M from Massachusetts General Hospital), omental fat, liver, lymphocytes and blood (Supplementary Note). Results are given if the unadjusted WHR SNP association showed $P < 1.00 \times 10^{-5}$. Findings are highlighted in bold font where the WHR SNP was the transcript peak SNP or where the WHR signal and the *cis*-eQTL signal were considered coincident (that is, the transcript peak SNP was highly correlated with the WHR SNP, $r^2 > 0.7$ and the transcript peak association disappeared by adjusting on the WHR SNP, $P > 0.05$); see also Online Methods. Unadj., unadjusted; Adj., adjusted.

^aEffect direction for the WHR-increasing allele.

^bSNP with the strongest association with the transcript in the region (transcript peak SNP).

^cCorrelation (HapMap CEU, build 36) between the WHR SNP and the transcript peak SNP.

^dThe transcript labeled AA553656 was detected as Contig27623_RC and corresponds to chromosome 7 locations 25,854,143–25,854,203 (HapMap build 36).