

RESEARCH ARTICLE

Metabolomic profiling reveals correlations between spermogram parameters and the metabolites present in human spermatozoa and seminal plasma

Kathrin M. Engel^{1,2}, Sven Baumann³, Ulrike Rolle-Kampczyk³, Jürgen Schiller², Martin von Bergen^{3,4}, Sonja Grunewald^{1*}

1 Training Center of the European Academy of Andrology (EAA), Dermatology, Venerology and Allergology Clinic, University Hospital Leipzig, Leipzig, Germany, **2** Institute of Medical Physics and Biophysics, Faculty of Medicine, University of Leipzig, Leipzig, Germany, **3** Department of Molecular Systems Biology, Helmholtz Centre for Environmental Research – UFZ, Leipzig, Germany, **4** Institute of Biochemistry, Faculty of Life Sciences, University of Leipzig, Leipzig, Germany

☞ These authors contributed equally to this work.
 ✉ Current address: Umweltbundesamt – UBA, Dessau-Roßlau, Germany
 * sonja.grunewald@medizin.uni-leipzig.de



OPEN ACCESS

Citation: Engel KM, Baumann S, Rolle-Kampczyk U, Schiller J, von Bergen M, Grunewald S (2019) Metabolomic profiling reveals correlations between spermogram parameters and the metabolites present in human spermatozoa and seminal plasma. PLoS ONE 14(2): e0211679. <https://doi.org/10.1371/journal.pone.0211679>

Editor: Roberto Amendola, ENEA Centro Ricerche Casaccia, ITALY

Received: October 25, 2018

Accepted: January 20, 2019

Published: February 20, 2019

Copyright: © 2019 Engel et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its Supporting Information files.

Funding: The study was funded by grants for SG and JS of the German Research Council (Deutsche Forschungsgemeinschaft, <http://www.dfg.de/>, grant numbers DFG GR 2075/7-1 and DFG Schi 476/12-2). The funder had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Abstract

In 50% of all infertility cases, the male is subfertile or infertile, however, the underlying mechanisms are often unknown. Even when assisted reproductive procedures such as *in vitro* fertilization and intracytoplasmic sperm injection are performed, the causes of male factor infertility frequently remain elusive. Since the overall activity of cells is closely linked to their metabolic capacity, we analyzed a panel of 180 metabolites in human sperm and seminal plasma and elucidated their associations with spermogram parameters. Therefore, metabolites from a group of 20 healthy donors were investigated using a targeted LC-MS/MS approach. The correlation analyses of the amino acids, biogenic amines, acylcarnitines, lysophosphatidylcholines, phosphatidylcholines, sphingomyelins and sugars from sperm and seminal plasma with standard spermogram parameters revealed that metabolites in sperm are closely related to sperm motility, whereas those in seminal plasma are closely related to sperm concentration and morphology. This study provides essential insights into the metabolome of human sperm and seminal plasma and its associations with sperm functions. This metabolomics technique could be a promising screening tool to detect the factors of male infertility in cases where the cause of infertility is unclear.

Introduction

Fertility problems are increasingly occurring in couples worldwide, and male factors are involved in approximately 50% of all infertility cases [1]. To predict male reproductive potential and record treatment success, clinicians currently primarily rely on semen analyses conducted according to the guidelines of the World Health Organization (WHO) [2]. This

Competing interests: The authors have declared that no competing interests exist.

Abbreviations: Ac-Orn, acetylcarnitine; ADMA, asymmetrically dimethylated arginine; alpha-AAA, α -amino adipic acid; Arg, arginine; Asp, aspartate; BMI, body mass index; CO, DL-carnitine; C10:1, decenoyl-L-carnitine; C12:1, dodecenoyl-L-carnitine; C14:2-OH, hydroxytetradecenyl-L-carnitine; C16, hexadecanoyl-L-carnitine; C16:2, hexadecadienyl-L-carnitine; C16-OH, hydroxyhexadecanoyl-L-carnitine; C18, octadecanoyl-L-carnitine; C18:1, octadecenyl-L-carnitine; C18:1-OH, hydroxyoctadecenyl-L-carnitine; C2, acetyl-L-carnitine; C3, propionyl-L-carnitine; C4, butyryl-L-carnitine; C4-OH (C3-DC), hydroxybutyryl-L-carnitine; C5, valeryl-L-carnitine; C5:1, tiglyl-L-carnitine; C5:1-DC, glutacoyl-L-carnitine; C5-DC (C6-OH), glutaryl-L-carnitine (hydroxyhexanoyl-L-carnitine); C5-M-DC, methylglutaryl-L-carnitine; C5-OH (C3-DC-M), hydroxyvaleryl-L-carnitine (methylmalonyl-L-carnitine); C6 (C4:1-DC), hexanoyl-L-carnitine (fumaryl-L-carnitine); C6:1, hexenoyl-L-carnitine; C8, octanoyl-L-carnitine; C9, nonanoyl-L-carnitine; CASA, computer-assisted sperm analysis; Cit, Citrulline; ESI, electrospray ionization; FIA, flow injection analysis; GC, gas chromatography; Gln, glutamine; Gly, glycine; GPCe, etherphosphorylcholine; His, histidine; ICSI, intracytoplasmic sperm injection; Ile, isoleucine; IVF, *in vitro* fertilization; LC, liquid chromatography; Leu, leucine; LPC, lysophosphatidylcholine; Lys, lysine; Met, methionine; Met-SO, methionine sulfoxide; MS, mass spectrometry; Nitro-Tyr, 3-nitrotyrosine; NMR, nuclear magnetic resonance; OH-Pro, L-4-hydroxyproline; Orn, ornithine; PC, phosphatidylcholine; PEA, phenethylamine; Phe, phenylalanine; PITC, phenylisothiocyanate; r_s , Spearman correlation coefficient; SD, standard deviation; SM, sphingomyelin; total DMA, total dimethylamine; Trp, tryptophan; Tyr, tyrosine; UPLC, ultra-performance liquid chromatography; Val, valine; WHO, World Health Organization.

protocol comprises the determination of rather superficial parameters, such as semen volume, semen pH, sperm cell count, sperm motility, and sperm morphology. With the exception of motility these parameters do not allow the detailed assessment of the functional status of the sperm cell. Therefore, the causes of male factor infertility frequently remain elusive even when assisted reproductive procedures such as *in vitro* fertilization (IVF) and intracytoplasmic sperm injection (ICSI) are performed.

New technologies, including genomics, proteomics and, most recently, metabolomics, have spurred the search for novel male infertility biomarkers [3]. In this context, the detection of genes, proteins [4], or metabolites unique to the infertile male might lead to an improved understanding of male sub- and infertility. This approach would also permit the identification of an accurate set of biomarkers that could be used to diagnose such patients with increased sensitivity and specificity.

At present, very limited data on the metabolome of healthy human semen are available. For human seminal plasma, some data on metabolites are available in the literature [5,6]. The correlation analysis of the spermogram parameters obtained by a computer-assisted sperm analysis (CASA) and sperm metabolites have also been reported [6]. In human sperm, 42 and 27 metabolites have been identified by untargeted approaches using nuclear magnetic resonance (NMR) spectroscopy and gas chromatography coupled to mass spectrometry (GC-MS), respectively [7]. MS-based methods were also used to characterize the lipid composition of sperm and seminal plasma from different species [8–10]. However, to the authors' best knowledge, a targeted approach to identify metabolites present in sperm and seminal plasma obtained from the same donors by liquid chromatography (LC) and the determination of the correlation of the levels of these metabolites in both types of specimens with manually assessed spermogram parameters have not yet been reported.

Sperm cell lysis is challenging [11] due to the small cell size, proteins that are almost completely bound to the cytoskeleton, high compartmentalization and limited cytoplasm. In this study, we established reliable protocols for the investigation of the metabolome and the identification of potential metabolomic candidates in the sperm and seminal plasma obtained from 20 healthy donors. The total concentrations of amino acids, biogenic amines, acylcarnitines, lysophosphatidylcholines, phosphatidylcholines, sphingomyelins and the sum of hexoses were determined, and studies correlating the levels of these metabolites with standard spermogram parameters were performed.

Materials and methods

Semen specimens

The study was approved by the Institutional Review Board of the University of Leipzig (No. 136-10-31052010) considering the Declaration of Helsinki. All patients signed a written informed consent form. Semen samples were collected from 20 healthy donors (age (mean \pm SD) 31.7 \pm 6.8 years, body mass index (BMI) 22.4 \pm 2.2, abstention time 3.6 \pm 1.1 days). The measured semen parameters of all of the samples were within the reference ranges defined by the World Health Organization for the normal fertile population [2].

Inclusion and exclusion criteria

Donors were invited to take part in the present study if they were 20 to 45 years old, normal weight (BMI 19 to 24), and did not suffer from any metabolic or cardiovascular diseases. Donors were informed that they should stay abstinent for 2 to up to 7 days. These criteria were confirmed by signing the written informed consent form.

Semen samples were excluded from the study if the sperm concentration was $< 15 \times 10^6$ cells/ml and/or if sperm total motility (progressive + local motility) was $< 40\%$.

Manual evaluation of sperm motility and sperm morphology

The motility of sperm in a sample volume of 10 μ l was evaluated microscopically at 400 \times magnification. Sperm were classified as progressively motile, locally motile or immotile as suggested by the WHO (WHO, 2010).

For the assessment of sperm morphology, smears were air-dried and stained according to the modified Papanicolaou staining method using Orange G 6, Mayer's Hemalum solution (Dr. K. Hollborn & Söhne GmbH & Co. KG, Leipzig, Germany) and EA-50 (Sigma-Aldrich Chemie GmbH, Taufkirchen, Germany). The morphology of 200 spermatozoa was determined under a light microscope at 1,000 \times magnification using an oil-immersion objective [12]. The head, neck, midpiece and tail of sperm cells were evaluated according to the WHO laboratory handbook [2].

The characteristics of the study group and the determined spermogram parameters are shown in [S1 Table](#).

Density gradient centrifugation

The liquefied semen samples were subjected to density gradient centrifugation (ISolate sperm separation medium, Irvine Scientific, Santa Ana, CA, USA). In brief, the samples were loaded onto a 50% and 90% discontinuous gradient of sperm separation medium, and centrifuged at 600 \times g for 20 min at room temperature (22°C). The supernatant, which consisted of the seminal plasma, was collected in a separate tube and stored at -80°C until further processing. The 90% pellet, which represented the mature sperm fraction, was resuspended in phosphate-buffered saline (PBS, pH 7.4). The sperm cells present in this fraction were then counted using a Neubauer hemocytometer. After an additional centrifugation of the re-suspended mature sperm fraction for 5 min at 5,300 \times g and 4°C, the supernatant was discarded.

Metabolic profiling of healthy semen donors

The following protocol was established to ensure a valid analytical procedure. All samples were treated in a standardized manner. The preanalytical period was kept as short as possible.

The metabolomic analysis of human sperm and seminal plasma was conducted using the AbsoluteIDQ p180 kit (Biocrates Life Sciences AG, Innsbruck, Austria) as previously described [13]. All samples were prepared and analyzed according to the protocol provided by Biocrates. Briefly, sperm metabolites were extracted by adding 2 ml methanol/water (1:1, v/v) to the sperm pellet and subjecting it to ultrasonic homogenization for 2 min on ice; this resulted in sperm cell lysis. The samples were centrifuged for 5 min at 5,300 \times g and 4°C, and the supernatants were treated according to the protocol provided by the manufacturer.

The amino acids and biogenic amines were analyzed by liquid chromatography (LC)-mass spectrometry (MS) (LC-MS/MS). Acylcarnitines, phosphatidylcholines (including lysophosphatidylcholines), sphingomyelins and hexoses were analyzed by flow injection analysis (FIA)-MS/MS measurements (FIA-MS/MS). Samples were pipetted into the wells of a 96-well sandwich filter plate, which included stable isotope-labeled internal standards. The filters were dried under a stream of nitrogen, the amino acids in the sample were derivatized with 5% phenylisothiocyanate reagent (PITC), and the filters were dried again. After the extraction of the metabolites and internal standards with 300 μ l 5 mM ammonium acetate in methanol and shaking at 450 rpm (Thermomixer comfort, Eppendorf, Hamburg, Germany) for 30 min at

room temperature, the solution was centrifuged through a solid-phase filter membrane and diluted for further analysis.

All analyses were performed on a QTRAP mass spectrometer under electrospray ionization (ESI) (ABI Sciex API 5500 Q-TRAP). The MS was coupled to an ultra-performance liquid chromatography (UPLC) system (Waters Acquity, Waters Corporation, Milford, CT, USA). Metabolite detection was achieved by positive and negative multiple reaction monitoring (MRM) using an injection volume of 20 μ l for FIA-MS/MS and an injection volume of 10 μ l for LC-MS/MS. For calibration, a calibration mixture consisting of 7 different concentrations was used. Quality controls obtained from lyophilized human plasma samples were included at 3 different concentration levels.

Raw data were collected in Analyst software (Sciex, Framingham, MA, USA) and processed using MetIDQ software, which is an integrated part of the p180 Kit (Biocrates). The quantification of LC-MS/MS data was performed using the IntelliQuan algorithm in Analyst software (Sciex). Subsequently, the determined metabolite concentrations of each sample were normalized to the amount per 100×10^6 sperm cells.

In general, 171 out of 180 metabolites (present as reference compounds in the kit) could be detected in sperm, and 177 out of 180 metabolites were present in seminal plasma. Only those metabolites that were present in $\geq 75\%$ of the samples were used in further analyses. In detail, the aspartate (Asp), citrulline (Cit), glycine (Gly), histidine (His), lysine (Lys), methionine (Met), ornithine (Orn), tryptophan (Trp), valine (Val) and etherphosphorylcholine (GPCe) 42:4 levels in sperm cells and the acetylornithine (Ac-Orn), phenethylamine (PEA), phosphatidylcholine (PC) 30:2 and PC 38:1 levels in seminal plasma were not included in the correlation analyses.

For a summary of the study please see [S1 Fig](#).

Statistical analysis

Statistical analyses were performed using GraphPad Prism 6 (GraphPad Software, Inc., La Jolla, CA, USA). The mean values and standard deviations of the concentrations of all metabolites were calculated. The data were analyzed for normal distribution and logarithmic normal distribution (if data were not normally distributed) using the D'Agostino & Pearson omnibus normality test. Because not all data were normally distributed, Spearman correlation coefficients (r_s) were calculated. All tests were nonparametric and two-tailed, and significance was indicated by $P < 0.05$.

Results and discussion

Despite the standardization of the experimental procedure, high deviations were observed for some analytes ([S2 Table](#)), possibly as a result of interindividual differences. The normalization to 100×10^6 sperm cells could also contribute to the deviations because the concentrations of "0" remain unaffected by this normalization. However, this normalization is less error-prone than dilution of the samples or the attempt to concentrate the samples to 100×10^6 sperm cells prior to MS analysis. The results of the seminal plasma analysis are more reliable than those of the sperm analysis.

The differences in the metabolite concentrations between sperm and seminal plasma indicate proper sample preparation and negligible mixing of the sperm and seminal plasma. Additionally, it can be assumed that sperm lysis occurred only after the separation of the sperm from the seminal plasma.

Amino acids and biogenic amines

Mature sperm are transcriptionally and translationally inert [14,15]. The proteins present in sperm are synthesized by the precursor cells during spermatogenesis.

As expected, all proteinogenic amino acids could be extracted from human sperm cells and seminal plasma. The presence of some amino acids, namely, alanine (Ala), arginine (Arg), glutamine (Gln), Gly, serine (Ser), tyrosine (Tyr), Val, leucine (Leu), isoleucine (Ile) and threonine (Thr) in human sperm has already been shown by NMR spectroscopic and GC-MS-based approaches [7].

In the present study the total amount of amino acids present in seminal plasma was higher than that in sperm, with a mean 800-fold increase. However, the total amino acid content of sperm and seminal plasma correlated positively with sperm count ($r_s = 0.535$ and 0.544 , respectively). The distribution pattern of the proteinogenic amino acids in sperm and in seminal plasma differed. As shown in Fig 1 and S2 Table, although the nonessential amino acid Gln was the most abundant amino acid in both specimens, in absolute counts, it was far more abundant in sperm than in seminal plasma (mean \pm SD $33.9 \pm 23.8\%$ vs. $11.1 \pm 0.7\%$). The high Gln content of sperm proteins is likely due to the presence of the androgen receptor, which contains polyglutamine stretches (CAG repeats) in its transactivation domain [16,17].

Previously published metabolomic analyses [18–20] reported that Gln is also present in high concentrations in human blood plasma and serum. In the study of Breier and colleagues the mean relative amounts of Gln measured in 22 subjects (approximately 21% for blood plasma and serum) were higher than the relative amounts found in seminal plasma but lower than the relative amounts found in sperm in the present study (Fig 1).

In contrast, not all of the investigated nonproteinogenic amino acids and biogenic amines could be detected in human sperm and seminal plasma. Whereas asymmetrically dimethylated arginine (ADMA), Cit, kynurenine, Orn, putrescine, sarcosine, serotonin, spermidine, spermine, taurine and total dimethylamine (total DMA) were present in both types of specimens, Ac-Orn, α -aminoadipic acid (alpha-AAA), carnosine, creatinine, L-4-hydroxyproline (OH-Pro) and PEA were detected only in seminal plasma. Histamine, methionine sulfoxide (Met-SO) and 3-nitrotyrosine (Nitro-Tyr) were found neither in sperm nor in seminal plasma (S2 Table). This is in contrast to Paiva *et al.* [7] and Zhao *et al.* [21], who also found alpha-AAA, creatinine and PEA in human sperm cells, but with different techniques, namely, NMR spectroscopy or GC.

Correlations among amino acids and biogenic amines

The quantities of most amino acids in sperm and seminal plasma were correlated in a significantly positive manner (S3 and S4 Tables). In contrast, the amounts of most individual amino acids in sperm and seminal plasma did not show cross-correlation (Fig 2). Only a few statistically significant associations were found between the levels of specific amino acids in sperm and those in seminal plasma. Among those, Gln in sperm showed a significant positive correlation ($P < 0.05$) with Arg ($r_s = 0.477$), Ile (0.454), Leu (0.512), phenylalanine (Phe, 0.568) and Tyr (0.476) in seminal plasma. A significant positive correlation was also found for Leu in sperm with Tyr (0.448) in seminal plasma (Fig 2, S5 Table).

Correlations between amino acid and biogenic amine concentrations and spermogram parameters

The concentrations of some amino acids and biogenic amines in sperm were correlated with spermogram parameters (Fig 3A and S6 Table). In detail, abstention time tended to correlate

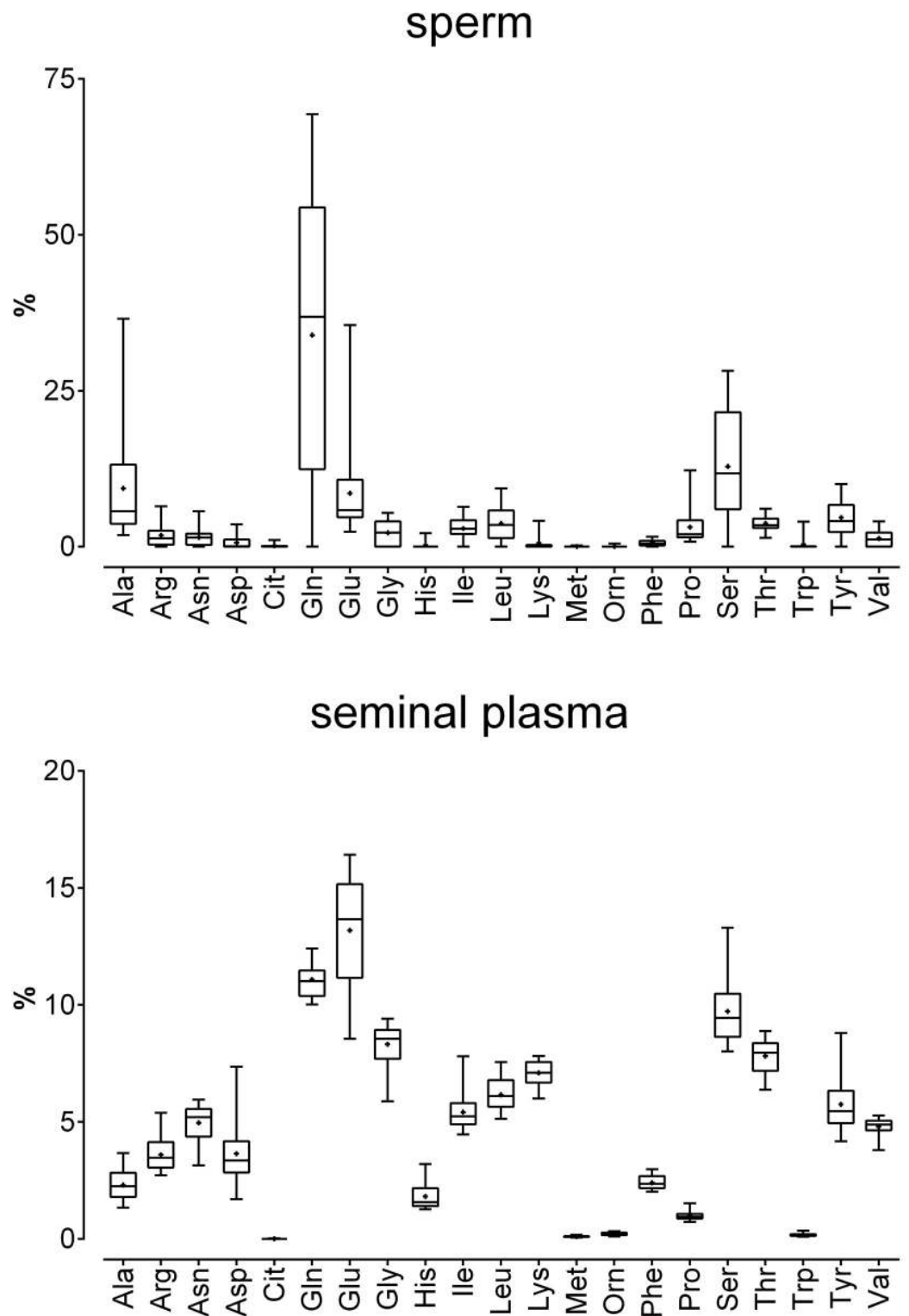


Fig 1. Relative amino acid concentrations in sperm and seminal plasma obtained from 20 healthy human donors. The box plots show the minimum, maximum and mean (black cross) of the data. Relative amounts were calculated for each sample based on the total amount of amino acids in the sample and are given in %. Total concentrations (in μmol) are given in the [S2 Table](#).

<https://doi.org/10.1371/journal.pone.0211679.g001>

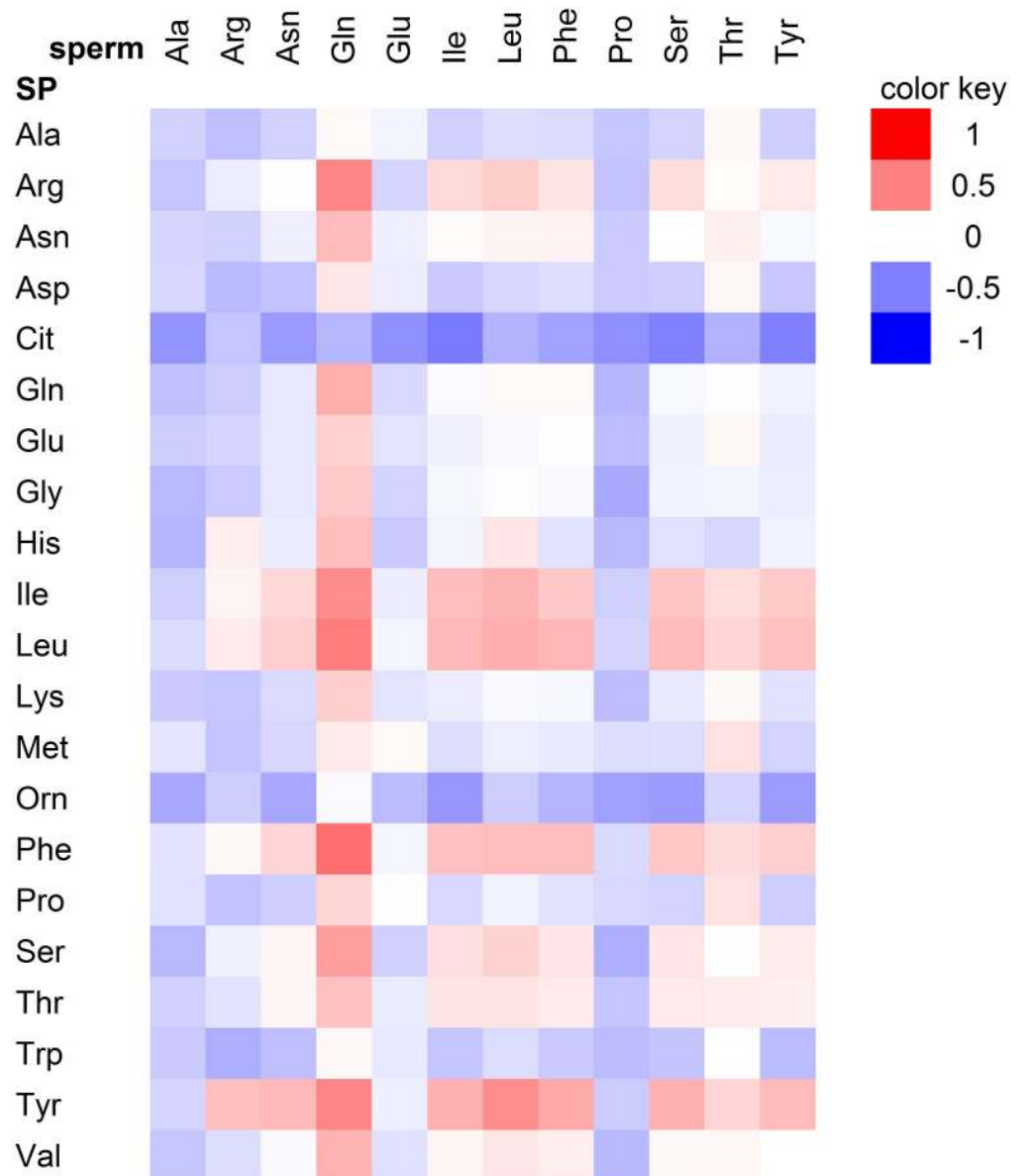


Fig 2. Heat map of the correlation coefficients between levels of amino acids from ejaculated mature human sperm and seminal plasma. Red colored cells represent positive correlations, and blue colored cells represent negative correlations. Only metabolites that were present in $\geq 75\%$ of the samples were considered for correlation analyses.

<https://doi.org/10.1371/journal.pone.0211679.g002>

negatively with amino acid content, and the correlation was significant ($P < 0.05$) for Ile ($r_s = -0.478$), Ser (-0.456), Tyr (-0.523) and putrescine (-0.598). Certain amino acids showed a positive correlation with progressive motility of sperm. This correlation was significant for Arg (0.499), Gln (0.562), Ile (0.629), Leu (0.603), Phe (0.674), Ser (0.639) and Tyr (0.622). NMR- and GC-MS-based studies found that the concentrations of some amino acids of sperm cells are changed significantly in asthenozoospermic patients where the motility of sperm is severely reduced. In accordance with our study the Leu levels were decreased in these patients [21].

Previous publications have described Arg as a crucial component in spermatogenesis. Diets deficient in Arg lead to the presence of multinuclear giant cells in the testis and to the

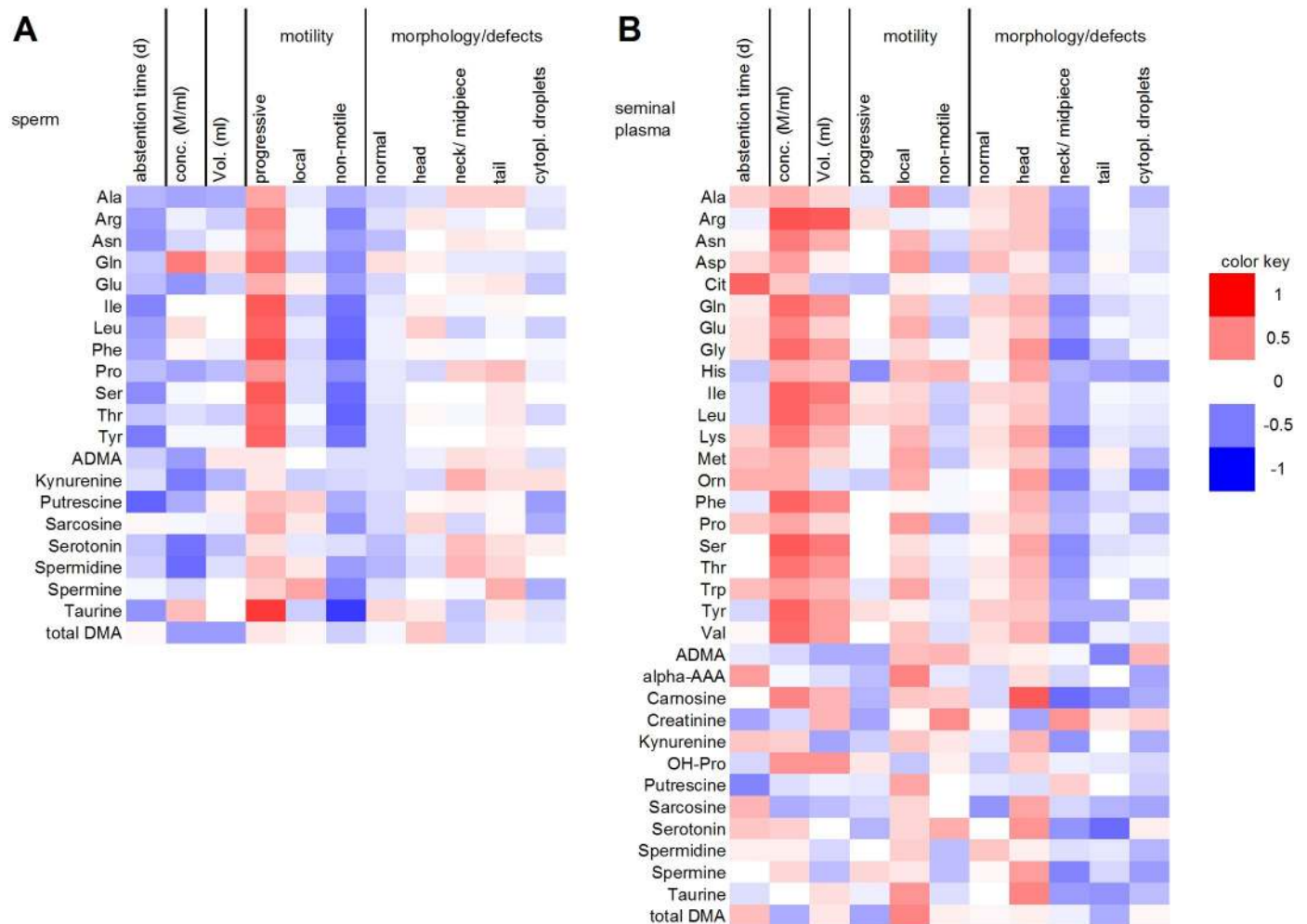


Fig 3. Heat map of the correlation coefficients between spermogram parameters and the levels of amino acids and biogenic amines from ejaculated mature human sperm (A) and seminal plasma (B). Red colored cells represent positive correlations, blue colored cells represent negative correlations. Only metabolites that were present in $\geq 75\%$ of the samples were included in the correlation analyses.

<https://doi.org/10.1371/journal.pone.0211679.g003>

impairment of spermatogenesis in men [22]. It was also shown that following the consumption of an Arg-deficient diet, the number and motility of human sperm decreased dramatically [23]. In contrast, Arg supplementation in men and boars increased the number and motility of sperm [24,25].

Gln and Arg, Gln and Ser, as well as Phe and Tyr, are connected via their synthesis pathways. The Ser and Tyr residues in proteins are potential sites for protein phosphorylation, and such sites have been shown to undergo crucial modifications during signal transduction cascades involved in sperm motility [26–29]. Additionally, several proteins involved in energy metabolism, protein folding/degradation, vesicle trafficking and cytoskeleton function have been reported to be associated with sperm motility [30–32]. The highest and most significant correlation was found between the sperm concentration of taurine and progressive motility ($r_s = 0.784, P = 0.0001$). This finding is consistent with the results of earlier studies in which it was shown that the addition of taurine to the medium improves the sperm motility in various species [33,34].

Although the sperm concentration of ejaculates tended to be negatively associated with their amino acid and biogenic amine content (significant for kynurenine (-0.526), serotonin (-0.545) and spermidine (-0.585)), a significant positive correlation was found for Gln (0.530).

No significant correlations between sperm morphology and the amino acid or biogenic amine content of the sperm were found.

The **seminal plasma** content of most proteinogenic amino acids and carnosine was significantly positively correlated with sperm concentration ($P < 0.05$). This result is in accordance with Qiao *et al.*, who also showed a positive correlation of the concentrations of some amino acids detected by GC-MS with sperm concentration [6]. Nonproteinogenic amino acids and other biogenic amines showed no association with sperm concentration (Fig 3B and S7 Table). Accordingly, the amount of all proteinogenic amino acids in seminal plasma also tended to be positively correlated with the volume of the ejaculate, reaching statistical significance for Arg, Ile, Phe and Ser ($r_s = 0.642, 0.524, 0.469$ and 0.528 , respectively). The levels of nonproteinogenic amino acids and biogenic amines showed no clear relationship to the volume of the ejaculate.

High creatinine levels in seminal plasma were significantly positively correlated with the amount of immotile sperm ($P < 0.05$). This finding is in accordance with an NMR-based study that showed increased creatinine levels in the seminal plasma of patients with asthenozoospermia [35]. Creatinine, a degradation product of creatine, is one of the most abundant biogenic amines in human blood serum [20] and is cleared via the kidneys [36]. Therefore, the creatinine in seminal plasma could result from a contamination of the ejaculate with urine. In stallions, urine contamination of ejaculates has been shown to lead to a significant reduction in total and progressive sperm motility [37]. Furthermore, there are reports of elevated creatine kinase activity in subfertile patients [38,39].

The inverse correlation of the putrescine concentration in sperm with abstention time is consistent with the observation of a similar negative correlation in seminal plasma. Putrescine is a basic, polycationic product of Arg degradation and is further converted into spermidine and spermine. In sperm and seminal plasma, the concentrations of spermine were tenfold higher than those of spermidine. A high abundance of spermine was also found in earlier studies in which seminal plasma was shown to contain the highest spermine concentration of all body fluids [40]. The concentrations of putrescine, spermidine and spermine tended to be inversely correlated with sperm immotility. This is consistent with reports in oligospermic men showing that treatment with S-adenosylmethionine, a polyamine precursor, leads to an increase in polyamine content and improved sperm motility [41]. Furthermore, compared to that of normal donors, higher concentrations of spermine and spermidine were reported in the seminal plasma of obese men (BMI > 30) with a poor spermiogram [42]. Low concentrations of spermine and ornithine (a precursor of putrescine) in seminal plasma were associated with higher frequencies of neck/midpiece defects in sperm cells. Spermine has been described as a decapacitation factor for sperm that ensures the proper timing of the acrosome reaction and capacitation [43].

Components of the serotonin (5-HT) system have been shown to be present in human ejaculate. The observed inverse correlation of the serotonin concentration in sperm with the sperm concentration is consistent with the results of former studies in which it was shown that hyperserotonemia leads to azoospermia [44] and that increased serotonin concentration in the spermatic vein is correlated with significantly lower sperm quality [45]. Impaired semen quality has also been associated with the use of serotonin reuptake inhibitors, which block serotonin receptors and thereby lead to an increase in serotonin concentration [46,47].

Normal sperm morphology was not significantly correlated with the levels of any amino acids or biogenic amines in seminal plasma. Interestingly, in-depth morphological analysis

revealed significant positive correlations of the carnosine and taurine content of seminal plasma with the occurrence of head defects and significant negative correlations of Gln, Gly, Lys, Orn, Ser, carnosine and spermine ($P < 0.05$; $r_s = -0.444, -0.538, -0.498, -0.484, -0.454, -0.576$ and -0.473 , respectively) levels with neck and midpiece defects in sperm cells. Tail defects were significantly negatively correlated ($P < 0.05$) with seminal plasma levels of ADMA ($r_s = -0.488$), carnosine (-0.446) and serotonin (-0.590). The presence of cytoplasmic droplets was inversely associated with Orn levels in seminal plasma (-0.457).

Acylcarnitines

Among acylcarnitines, DL-carnitine (C0) and acetyl-L-carnitine (C2) were mainly detected in sperm. All other acylcarnitines were found in amounts less than 1 μmol . The four most abundant acylcarnitines in seminal plasma were C0, C2, butyryl-L-carnitine (C4) and propionyl-L-carnitine (C3). The detailed results are presented in [Fig 4](#) and [S2 Table](#).

Correlations among acylcarnitines

The quantities of most acylcarnitines in sperm as well as in seminal plasma correlated with each other in a significant positive manner ($P < 0.05$). In contrast, significant cross-correlations between sperm and seminal plasma were observed only for C3 ($r_s = 0.460$) and hexadecadienyl-L-carnitine (C16:2, -0.453). The amounts of C3-DC-M/C5-OH and C4:1-DC/C6 in seminal plasma were significantly negatively correlated with those of nearly every other acylcarnitine species in sperm. The methylglutaryl-L-carnitine (C5-M-DC) content of sperm was significantly negatively correlated with the levels of C3, C6, octanoyl-L-carnitine (C8), C16:2 and all other C5 metabolites.

The C2 and C3 levels in sperm correlated positively with the levels of acylcarnitines in seminal plasma but were significantly negatively correlated with decenoyl-L-carnitine (C10:1) and dodecenoyl-L-carnitine (C12:1) levels. The correlation coefficients for the acylcarnitine concentrations in sperm with those in seminal plasma are presented in [S8 Table](#).

Correlations between acylcarnitine concentrations and spermogram parameters

The concentration of every acylcarnitine **in sperm** was significantly correlated ($P < 0.05$) with at least one spermogram parameter ([Fig 5A](#)). In detail, the concentrations of most acylcarnitines, except C0, C16, C16-OH, C18, C18:1, C3, C3-DC, C4, C5, C5-OH, C5:1 and C6, were significantly negatively correlated with sperm concentration. Interestingly, those acylcarnitines that were not significantly correlated with sperm concentration were linked to sperm motility. Among them were the most abundant acylcarnitines, namely, C0, C2 and C3. This result is accordance with the relevance of acylcarnitines in the energy production of mammalian cells. The complete mechanisms were recently summarized in [48]. With respect to sperm, it has been shown that high concentrations of free acylcarnitines are positively correlated with progressive sperm movement [49]. Furthermore, a study reported lower C2 concentrations in semen of oligozoospermic men [50]. In accordance with this finding, Busetto and colleagues reported that L-carnitine, acetyl-L-carnitine, vitamins and zinc diet supplementation has a beneficial effect on sperm concentration and sperm motility in men with abnormal spermograms that show oligo-, astheno-, or teratozoospermia [51].

In **seminal plasma**, in contrast to sperm, the acylcarnitine content tended to correlate positively with sperm concentration ([Fig 5B](#)). The correlations were significant for C14:2-OH, C16:2, C18:1-OH, C2, C3, C5, C5-DC, C5-OH, C5:1-DC, C6 and C6:1. This finding is consistent with an earlier comparison of infertile and fertile men that showed significantly reduced

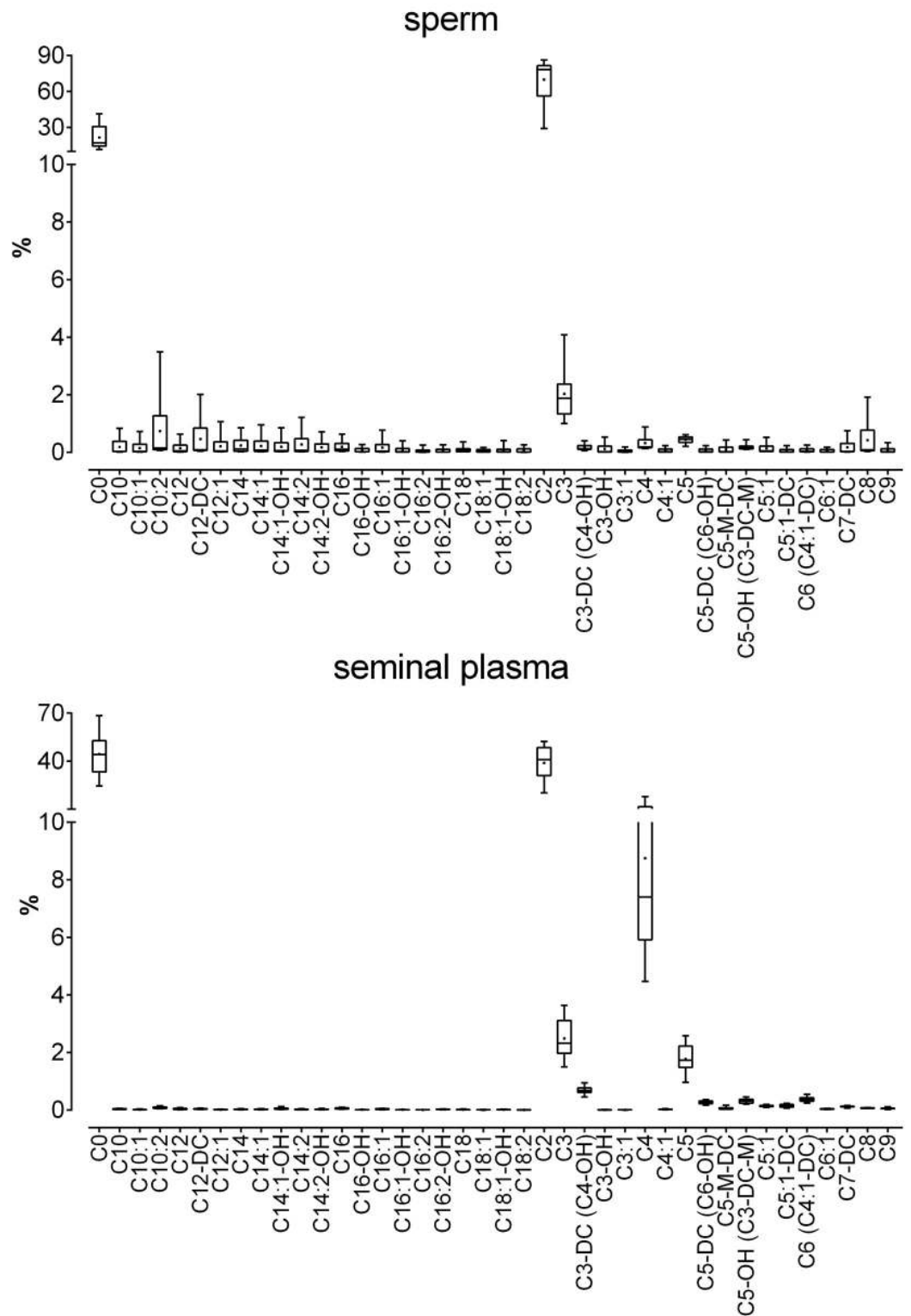


Fig 4. Relative acylcarnitine concentrations in sperm and seminal plasma obtained from 20 healthy human donors. The box plots show the minimum, maximum and mean (black cross) values of the data. The relative amounts were calculated for each sample based on the total amount of acylcarnitines in the sample and are given in %. Total concentrations (in μmol) are given in [S2 Table](#).

<https://doi.org/10.1371/journal.pone.0211679.g004>

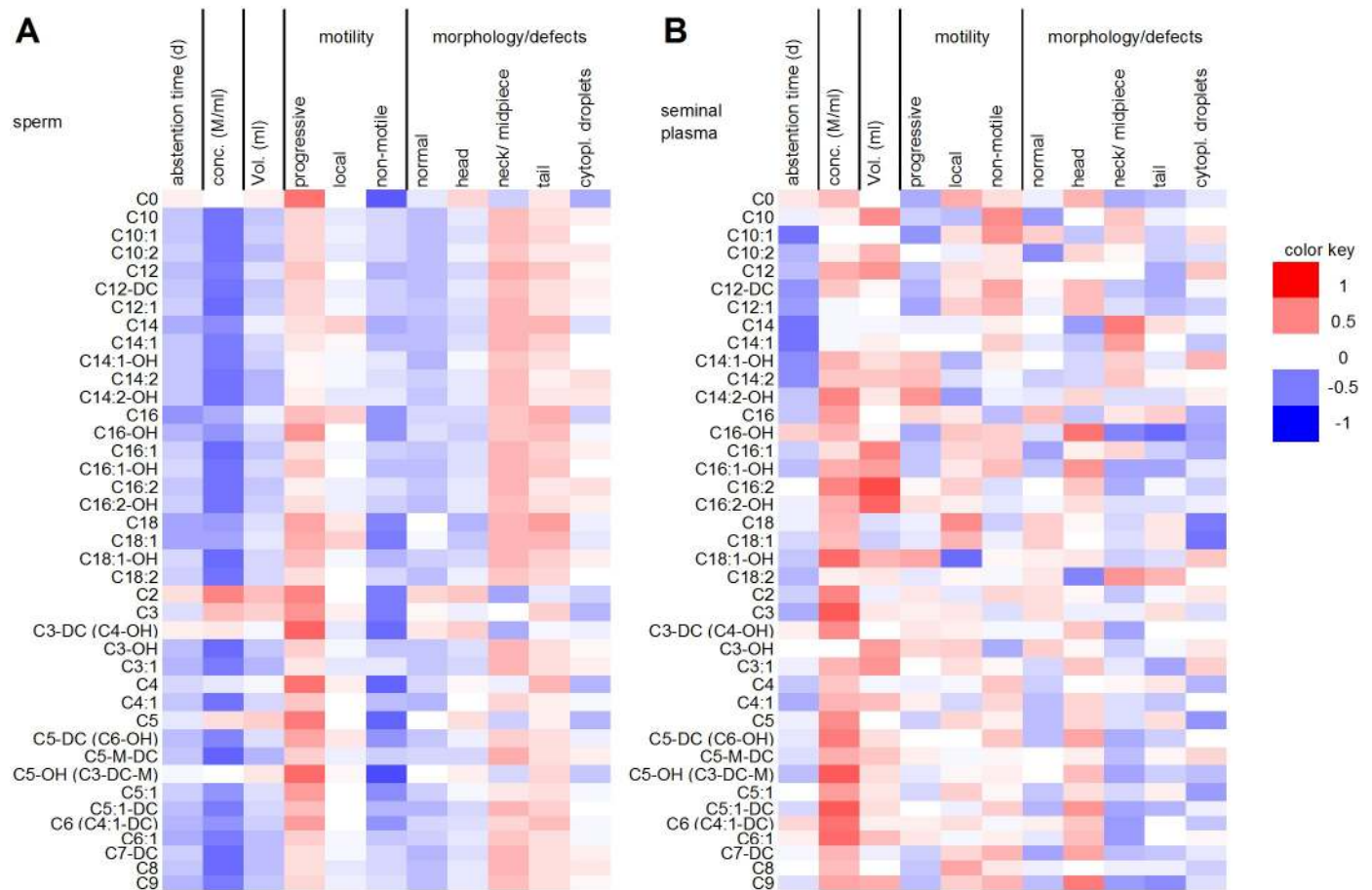


Fig 5. Heat map of the correlation coefficients between spermogram parameters and the levels of acylcarnitines from ejaculated mature human sperm (A) and seminal plasma (B). Red colored cells represent positive correlations, and blue colored cells represent negative correlations.

<https://doi.org/10.1371/journal.pone.0211679.g005>

concentrations of free and total carnitine in the infertile cohort [52]. Because there were no significant associations of acylcarnitine levels with sperm motility, it might be reasonable to assume that acylcarnitines in seminal plasma are responsible for the protection of the sperm, particularly against oxidative damage.

The amount of C16-OH in seminal plasma was associated with morphological defects; specifically, it showed a significant positive association with head defects ($r_s = 0.548$) and a significant negative association with neck (-0.477) and tail defects (-0.563). A similar trend was observed for C9.

Lipids

Because most of the currently available metabolomic data describing sperm and seminal plasma have been either performed with NMR spectroscopy or GC-MS, there are no metabolomics data of the PC compositions and intact lipids available. The three most abundant lipids in sperm were sphingomyelin (SM) 16:0, PC 38:6 and PC 28:1. In seminal plasma, SM 16:0, PC 34:1 and SM 24:0 were present at the highest concentrations of all lipids investigated. These results are in accordance with previously published studies that relied on another MS-based technique, namely, MALDI-TOF MS [53,54]. For detailed results, please see Figs 6–9 and S2 Table.

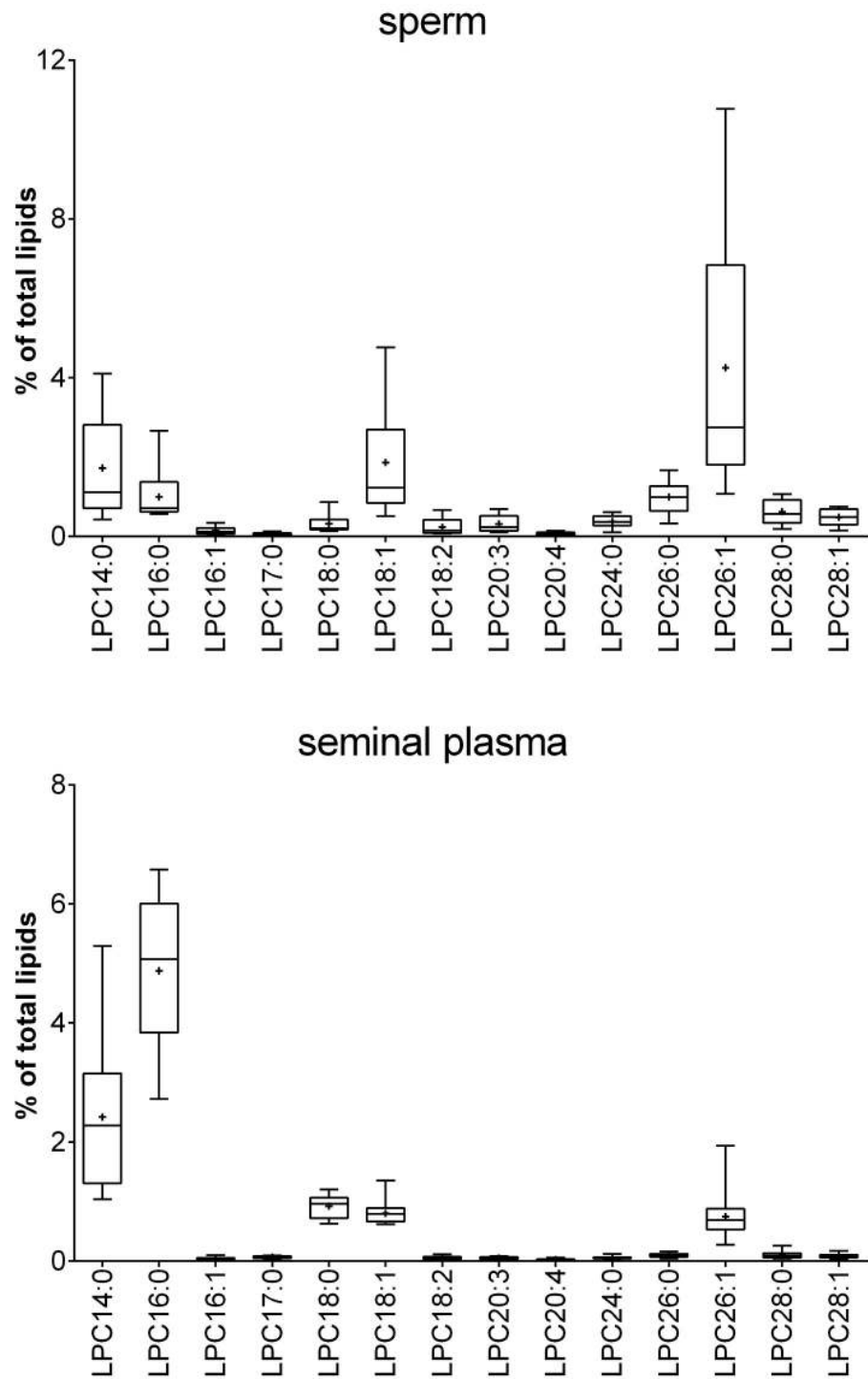


Fig 6. Relative lysophosphatidylcholine (LPC) concentrations in sperm and seminal plasma obtained from 20 healthy human donors. The box plots show the minimum, maximum and mean (black cross) values of the data. The relative amounts were calculated for each sample based on the total amount of lipids in the sample and are given in %. Total concentrations (in μmol) are given in [S2 Table](#).

<https://doi.org/10.1371/journal.pone.0211679.g006>

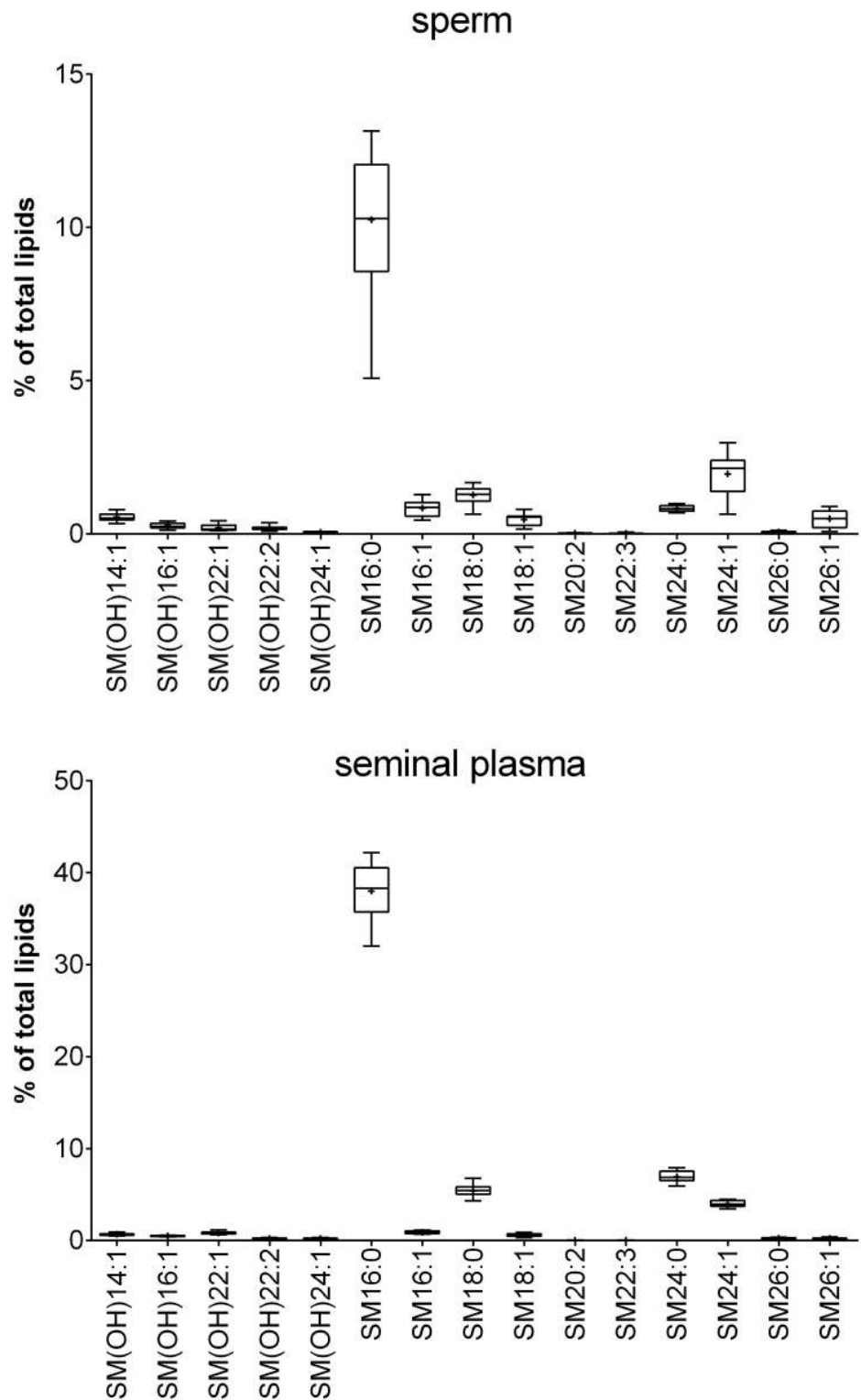


Fig 7. Relative sphingomyelin (SM) concentrations in sperm and seminal plasma obtained from 20 healthy human donors. The box plots show the minimum, maximum and mean (black cross) values of the data. The relative amounts were calculated for each sample based on the total amount of lipids in the sample and are given in %. Total concentrations (in μmol) are given in [S2 Table](#).

<https://doi.org/10.1371/journal.pone.0211679.g007>

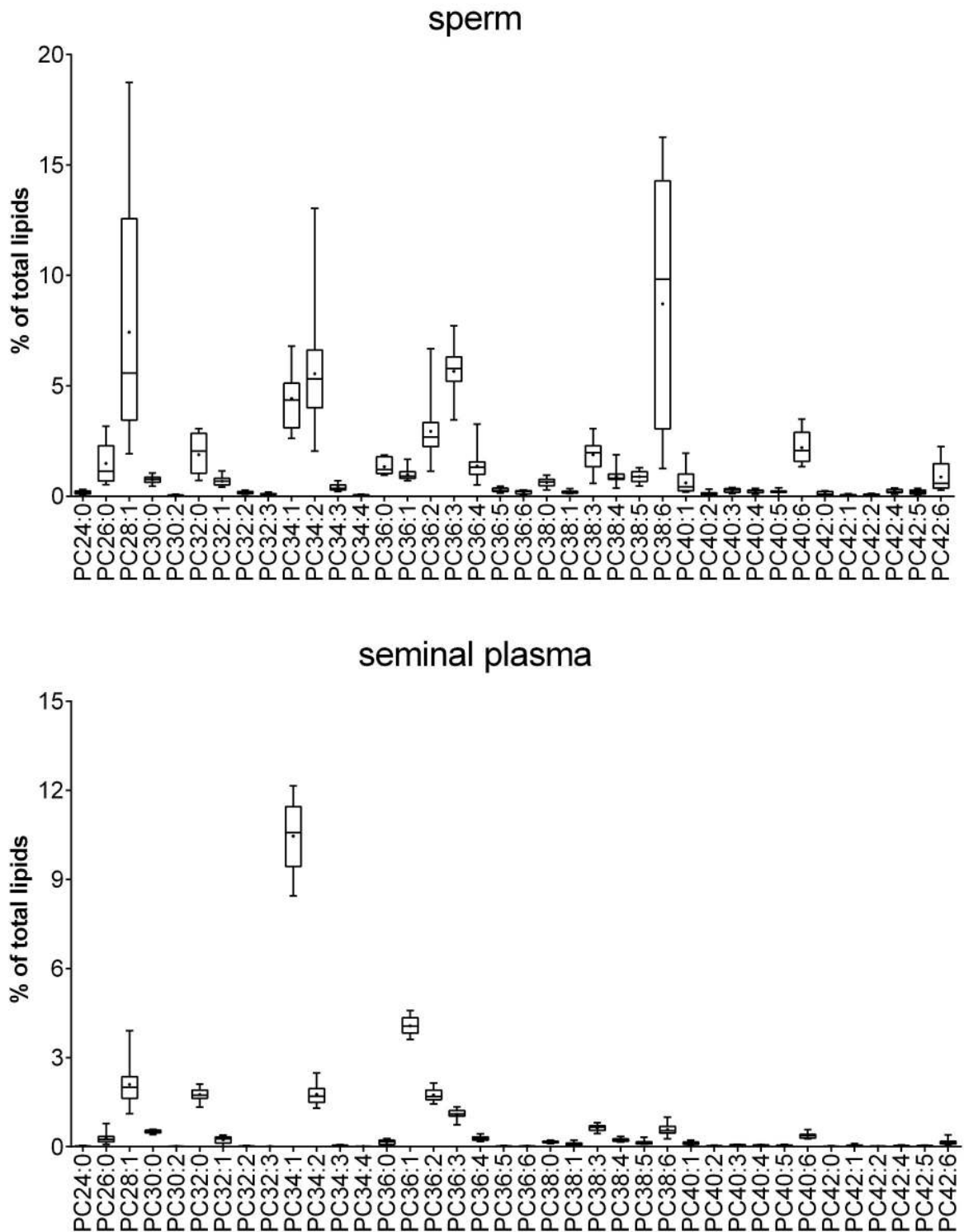


Fig 8. Relative phosphatidylcholine (PC) concentrations in sperm and seminal plasma obtained from 20 healthy human donors. The box plots show the minimum, maximum and mean (black cross) values of the data. The relative amounts were calculated for each sample based on the total amount of lipids in the sample and are given in %. Total concentrations (in μmol) are given in [S2 Table](#).

<https://doi.org/10.1371/journal.pone.0211679.g008>

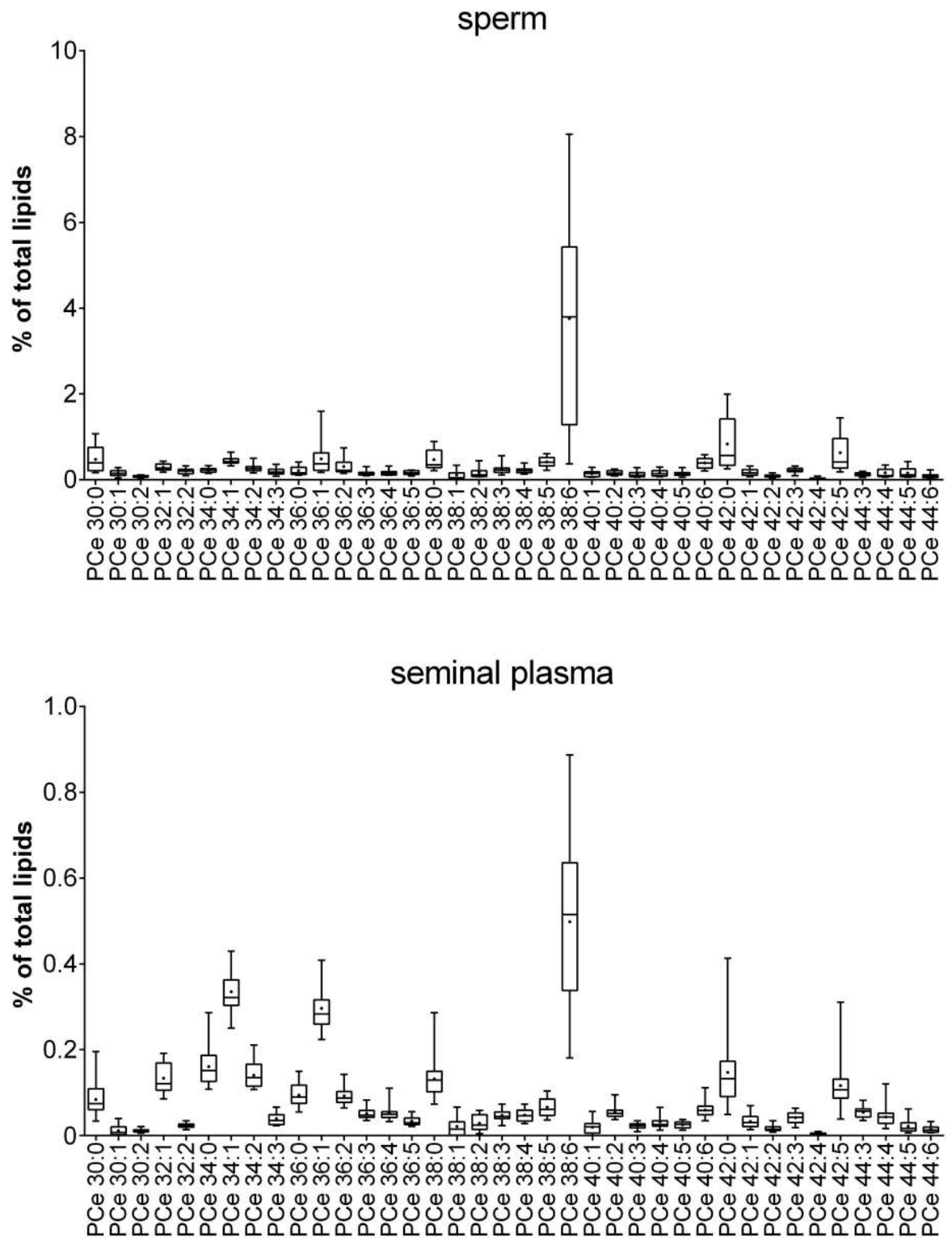


Fig 9. Relative etherphosphorylcholine (GPCe) concentrations in sperm and seminal plasma obtained from 20 healthy human donors. The box plots show the minimum, maximum and mean (black cross) values of the data. The relative amounts have been calculated for each sample based on the total amount of lipids in the sample and are given in %. Total concentrations (in μmol) are given in [S2 Table](#).

<https://doi.org/10.1371/journal.pone.0211679.g009>

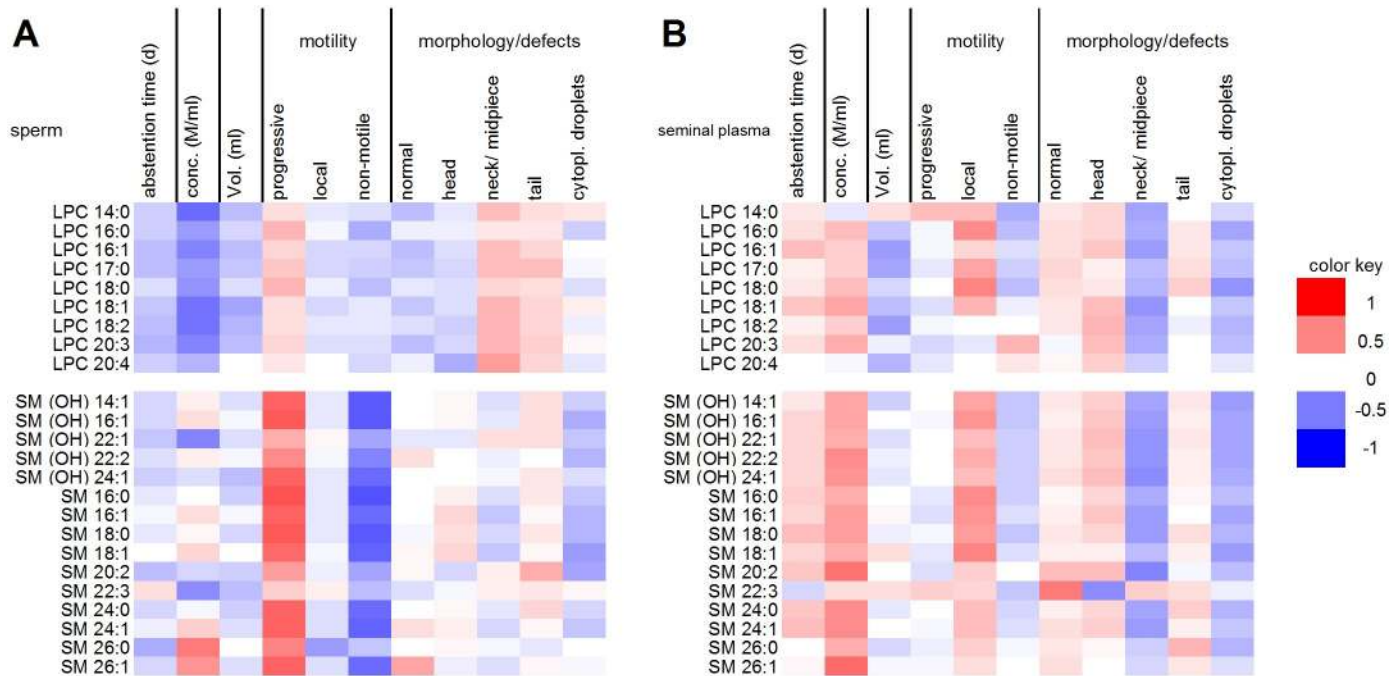


Fig 10. Heat map of the correlation coefficients between spermogram parameters and the levels of lysophosphatidylcholines (LPC) and sphingomyelins (SM) from ejaculated mature human sperm (A) and seminal plasma (B). Red colored cells represent positive correlations, and blue colored cells represent negative correlations. Only metabolites that were present in $\geq 75\%$ of the samples were considered in the correlation analyses.

<https://doi.org/10.1371/journal.pone.0211679.g010>

Correlations among lipids

In sperm cells, the levels of all LPC species were correlated in a significantly positive manner ($P < 0.05$). This correlation was also observed for most PC, ether GPC (GPCe) and SM species (Figs 10A and 11A). Only a few correlations were found for PC 28:1, PC 30:2, PC 40:1, PC 42:6, GPCe 38:6 and SM 22:3 within the respective lipid classes.

The correlation analysis of LPC with PC species resulted in significant positive associations in most cases, except for LPC 14:0 and PC 38:6. Regarding the association of LPC with GPCe and SM in human spermatozoa, no associations were found for GPCe 38:6 and SM 26:0 levels with any LPC species.

With the exception of PC 38:6, most PC species also correlated with GPCe. Considering the associations between PC and SM as well as GPCe and SM species, only SM 22:3 did not correlate with most PC and GPCe species.

The results for seminal plasma were similar, and many significant correlations were found (Figs 10B and 11B). However, no associations were found for GPCe 40:1. Among the lipids investigated, fewer than ten correlations were found for LPC 14:0 (8), PC 26:0 (7), PC 36:0 (5), PC 40:1 (9), GPCe 42:0 (5), GPCe 42:5 (7), GPCe 44:5 and SM 22:3 (2). GPCe 44:4 was correlated only with GPCe 38:1, and GPCe 30:1 was correlated only with PC 40:2.

Correlations between lipids and spermogram parameters

Significant positive correlations with progressive motility were found for more than 50% (40 of 76) of the analyzed PC species and for 80% (12 of 15) of the analyzed SM species in sperm, indicating a decisive role of lipid metabolites in sperm motility. The classification of PC and PCe into subgroups with different double bond contents shows significant correlations of highly unsaturated PC and PCe in sperm cells with sperm motility (Fig 12). This is in

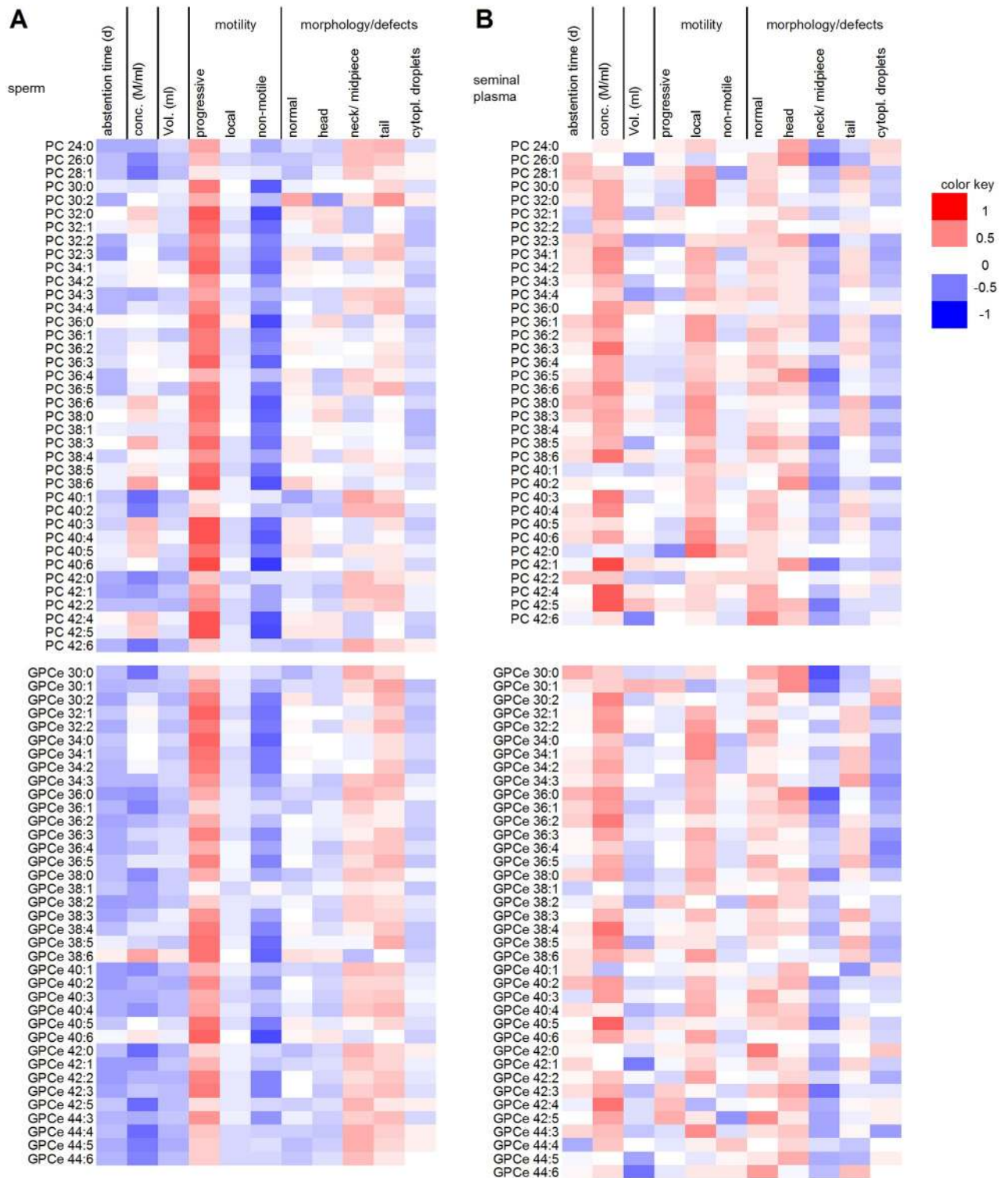


Fig 11. Heat map of the correlation coefficients between spermogram parameters and the levels of phosphatidylcholines (PC) and ether-bound glycerophosphorylcholines (GPCe) from ejaculated mature human sperm (A) and seminal plasma (B). Red colored cells represent positive correlations, and blue colored cells represent negative correlations. Only metabolites that were present in $\geq 75\%$ of the samples were included in the correlation analyses.

<https://doi.org/10.1371/journal.pone.0211679.g011>

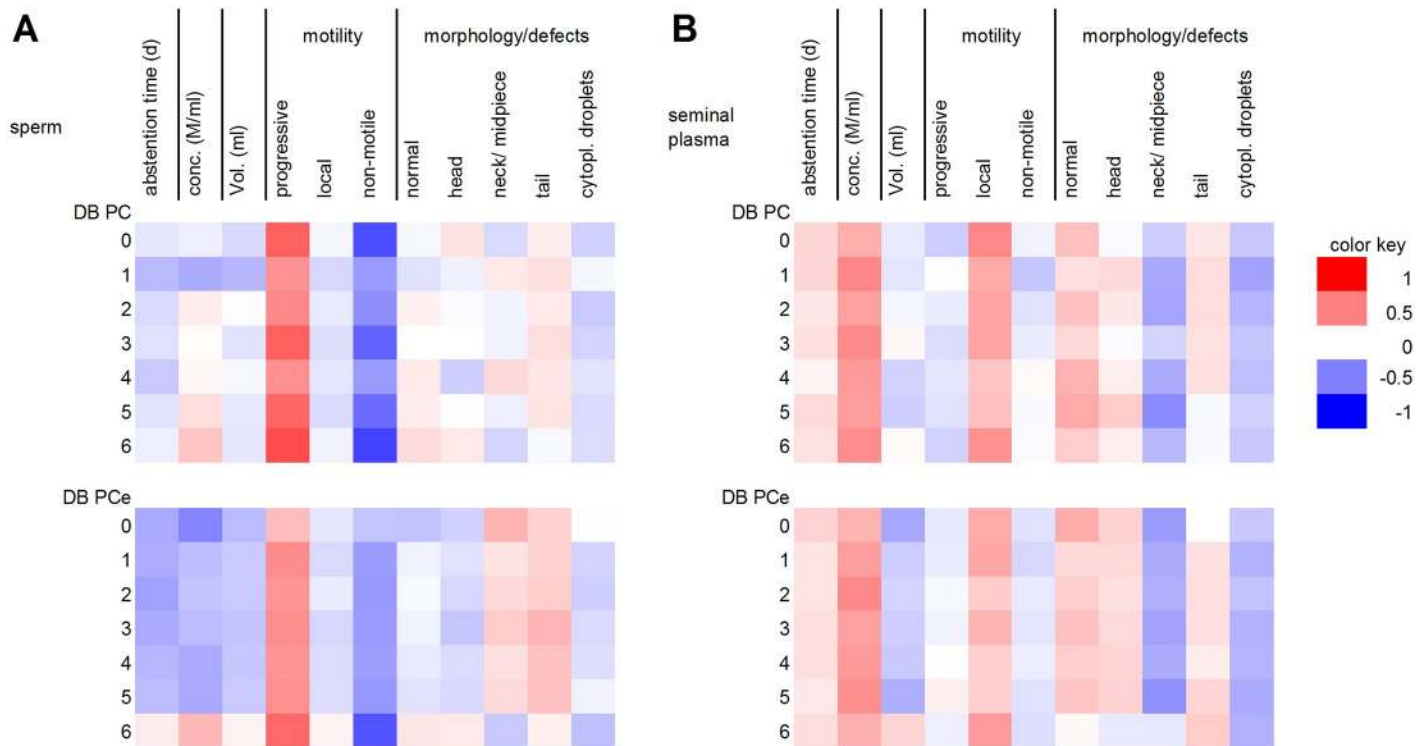


Fig 12. Heat map of the correlation coefficients between spermogram parameters and the levels of phosphatidylcholines (PC) and ether-bound glycerophosphorylcholines (GPCe) with different double bond contents from ejaculated mature human sperm (A) and seminal plasma (B). Red colored cells represent positive correlations, and blue colored cells represent negative correlations.

<https://doi.org/10.1371/journal.pone.0211679.g012>

accordance with publications showing, that the assembly of docosahexaenoic acid (C22:6) into sperm lipids during the maturation of sperm in the epididymis increases the fluidity of the membrane and, thus, promotes the motility of the sperm cells [10,55]. Overall, LPC and PC species tended to correlate negatively with sperm concentration, whereas no clear trend was observed for SM. Lysolipids (such as LPC) can act as detergents and are therefore capable of destroying lipid bilayers [56,57]. In this context, they also play a crucial role during the fertilization process [57,58]. As the sperm cell approaches the oocyte, the concentration of lysolipids in the sperm cell membrane increases, and lysolipids facilitate the fusion of the sperm and oocyte membranes [59]. However, if the lysolipid concentration in the sperm membrane increases too early, the cell can be destroyed before it reaches its final destination. Increased LPC concentrations have been shown in the sperm of very obese patients [60]. Interestingly, no correlations between lipid concentrations and morphological features were found in the present study; such associations would have been expected if the structure of the sperm membrane was damaged.

In contrast to that of sperm, the lipid concentration of seminal plasma was positively associated with sperm concentration and local motility and tended to correlate negatively with morphological abnormalities involving neck/midpiece defects. A study on Chinese patients reported that the levels of triglycerides, total cholesterol, and low- and high-density lipoproteins in seminal plasma were higher in individuals with oligo-, astheno- or teratozoospermia than in individuals with normal sperm concentration, motility and morphology [61]. These lipid species were not investigated in the present study. A significant positive correlation with

normomorphonic sperm was found for the levels of PC 42:6, GPCe 42:0, GPCe 42:5 and SM 22:3 in seminal plasma.

A complete list of the observed correlations between the sperm and seminal plasma components and the spermogram parameters is provided in [S6](#) and [S7](#) Tables. The correlation coefficients for lipid concentrations in sperm with those in seminal plasma are shown in [S9–S12](#) Tables. The results of the correlation analysis indicate that only a small exchange of metabolites occurs between sperm and seminal plasma.

Conclusion

Here, we present the first targeted approach to a metabolomic analysis of human sperm and seminal plasma using sophisticated and reliable sample preparation methods and liquid chromatography/flow injection analysis coupled with electrospray ionization mass spectrometry under high-throughput conditions. This pilot study is a first step in linking traditional spermogram parameters with sophisticated mass spectrometric tools. Although further studies will be necessary to define the normal range in the concentrations of the metabolites in the sperm and seminal plasma of healthy men, the multiple correlations with standard sperm parameters found in this study demonstrate that metabolomics is a very useful tool in the identification of biomarkers for male infertility. As a next step, metabolomic investigations of semen samples from subfertile and infertile men should be performed.

Supporting information

S1 Fig. Summary of the study design and outcome. Each ejaculate was analyzed manually and information about the abstention time, semen volume, pH, leucocyte concentration, sperm concentration, sperm motility and sperm morphology were recorded. Subsequently, the semen sample was separated by density gradient centrifugation and the mature sperm fraction and the seminal plasma were collected. Both were analyzed by a targeted metabolomics approach. Whereas in sperm 171 out of 180 metabolites were identified, 177 out of 180 metabolites were identified in seminal plasma. Concentrations of sperm metabolites were adjusted to 100×10^6 cells. Metabolite concentrations were correlated to spermogram parameters. Overall, the metabolome of sperm is more closely related to sperm count and sperm motility, whereas the metabolome of the seminal plasma is more closely related to sperm count and sperm morphology.
(TIF)

S1 Table. Age, body mass index (BMI) and spermogram parameters of 20 healthy human subjects. Data illustrate the quantitative aspects of the specimens as well as their characteristics with respect to sperm motility and morphology (left column). Data of donors are given as mean \pm SD. For comparison the reference ranges established by the WHO (5th edition, 2010) are shown in the right column.
(DOCX)

S2 Table. Concentrations of metabolites in sperm and seminal plasma of 20 healthy human subjects. Data are given in μmol as mean \pm SD. Abbreviations: Ala—alanine, Arg—arginine, Asn—asparagine, Asp—aspartate, Cit—citrulline, Gln—glutamine, Glu—glutamate, Gly—glycine, His—histidine, Ile—isoleucine, Leu—leucine, Lys—lysine, Met—methionine, Orn—ornithine, Phe—phenylalanine, Pro—proline, Ser—serine, Thr—threonine, Trp—tryptophan, Tyr—tyrosine, Val—valine, Ac-Orn—acetylornithine, ADMA—asymmetrically dimethylated arginine, alpha-AAA—alpha-amino adipic acid, Met-SO—methionine sulfoxide, Nitro-Tyr—nitrotyrosine, OH-Pro—hydroxyproline, PEA—phenylethylamine, DMA—

dimethylamine, C0—DL-carnitine, C10—decanoyl-L-carnitine, C10:1—decenoyl-L-carnitine, C10:2—decadienyl-L-carnitine, C12—dodecanoyl-L-carnitine, C12:1—dodecenoyl-L-carnitine, C12-DC—dodecanedioyl-L-carnitine, C14—tetradecanoyl-L-carnitine, C14:1—tetradecenoyl-L-carnitine, C14:1-OH—hydroxytetradecenoyl-L-carnitine, C14:2—tetradecadienyl-L-carnitine, C14:2-OH—hydroxytetradecadienyl-L-carnitine, C16—hexadecanoyl-L-carnitine, C16:1—hexadecenoyl-L-carnitine, C16:1-OH—hydroxyhexadecenoyl-L-carnitine, C16:2—hexadecadienyl-L-carnitine, C16:2-OH—hydroxyhexadecadienyl-L-carnitine, C16-OH—hydroxyhexadecanoyl-L-carnitine, C18—octadecanoyl-L-carnitine, C18:1—octadecenoyl-L-carnitine, C18:1-OH—hydroxyoctadecenoyl-L-carnitine, C18:2—octadecadienyl-L-carnitine, C2—acetyl-L-carnitine, C3—propionyl-L-carnitine, C3:1—propenyl-L-carnitine, C3-DC/C4-OH—malonyl-L-carnitine/hydroxybutyryl-L-carnitine, C3-DC-M/C5-OH—methylmalonyl-L-carnitine/hydroxyvaleryl-L-carnitine, C3-OH—hydroxypropionyl-L-carnitine, C4—butyryl-L-carnitine, C4:1—butenyl-L-carnitine, C4:1-DC/C6—fumaryl-L-carnitine/hexanoyl-L-carnitine, C5—valeryl-L-carnitine, C5:1—tiglyl-L-carnitine, C5:1-DC—glutaconyl-L-carnitine, C5-DC/C6-OH—glutaryl-L-carnitine/hydroxyhexanoyl-L-carnitine, C5-M-DC—methylglutaryl-L-carnitine, C6:1—hexenoyl-L-carnitine, C7-DC—pimetyl-L-carnitine, C8—octanoyl-L-carnitine, C8:1—octenoyl-L-carnitine, C9—nonayl-L-carnitine, LPC—lysophosphatidylcholine, PC—phosphatidylcholine, GPCe—etherphosphorylcholine, SM—sphingomyelin.

(DOCX)

S3 Table. Correlation analysis of amino acids in sperm of 20 healthy donors. Data are Spearman correlation rank coefficients. Significant correlations are highlighted in bolt red. Abbreviations: Ala—alanine, Arg—arginine, Asn—asparagine, Gln—glutamine, Glu—glutamate, Ile—isoleucine, Leu—leucine, Phe—phenylalanine, Pro—proline, Ser—serine, Thr—threonine, Tyr—tyrosine.

(DOCX)

S4 Table. Correlation analysis of amino acids in seminal plasma of 20 healthy donors. Data are Spearman correlation rank coefficients. Significant correlations are highlighted in bolt. Abbreviations: Ala—alanine, Arg—arginine, Asn—asparagine, Asp—aspartate, Cit—citrulline, Gln—glutamine, Glu—glutamate, Gly—glycine, His—histidine, Ile—isoleucine, Leu—leucine, Lys—lysine, Met—methionine, Orn—ornithine, Phe—phenylalanine, Pro—proline, Ser—serine, Thr—threonine, Trp—tryptophan, Tyr—tyrosine, Val—valine.

(DOCX)

S5 Table. Correlation analysis of amino acids in sperm and seminal plasma (SP) of 20 healthy donors. Data are Spearman correlation rank coefficients. Significant correlations are highlighted in bolt. Abbreviations: Ala—alanine, Arg—arginine, Asn—asparagine, Asp—aspartate, Cit—citrulline, Gln—glutamine, Glu—glutamate, Gly—glycine, His—histidine, Ile—isoleucine, Leu—leucine, Lys—lysine, Met—methionine, Orn—ornithine, Phe—phenylalanine, Pro—proline, Ser—serine, Thr—threonine, Trp—tryptophan, Tyr—tyrosine, Val—valine.

(DOCX)

S6 Table. Correlation analysis of metabolites in sperm of 20 healthy donors with spermogram parameters. Data are Spearman correlation rank coefficients. Significant positive correlations are highlighted in red, significant negative correlations in blue. Abbreviations: Ala—alanine, Arg—arginine, Asn—asparagine, Gln—glutamine, Glu—glutamate, Gly—glycine, His—histidine, Ile—isoleucine, Leu—leucine, Phe—phenylalanine, Pro—proline, Ser—serine, Thr—threonine, Tyr—tyrosine, ADMA—asymmetrically dimethylated arginine, DMA—

dimethylamine, C0—DL-carnitine, C10—decanoyl-L-carnitine, C10:1—decanoyl-L-carnitine, C10:2—decadienyl-L-carnitine, C12—dodecanoyl-L-carnitine, C12:1—dodecenoyl-L-carnitine, C12-DC—dodecanedioyl-L-carnitine, C14—tetradecanoyl-L-carnitine, C14:1—tetradecenoyl-L-carnitine, C14:1-OH—hydroxytetradecenoyl-L-carnitine, C14:2—tetradecadienyl-L-carnitine, C14:2-OH—hydroxytetradecadienyl-L-carnitine, C16—hexadecanoyl-L-carnitine, C16:1—hexadecenoyl-L-carnitine, C16:1-OH—hydroxyhexadecenoyl-L-carnitine, C16:2—hexadecadienyl-L-carnitine, C16:2-OH—hydroxyhexadecadienyl-L-carnitine, C16-OH—hydroxyhexadecanoyl-L-carnitine, C18—octadecanoyl-L-carnitine, C18:1—octadecenoyl-L-carnitine, C18:1-OH—hydroxyoctadecenoyl-L-carnitine, C18:2—octadecadienyl-L-carnitine, C2—acetyl-L-carnitine, C3—propionyl-L-carnitine, C3:1—propenyl-L-carnitine, C3-DC/C4-OH—malonyl-L-carnitine/hydroxybutyryl-L-carnitine, C3-DC-M/C5-OH—methylmalonyl-L-carnitine/hydroxyvaleryl-L-carnitine, C3-OH—hydroxypropionyl-L-carnitine, C4—butyryl-L-carnitine, C4:1—butenyl-L-carnitine, C4:1-DC/C6—fumaryl-L-carnitine/hexanoyl-L-carnitine, C5—valeryl-L-carnitine, C5:1—tiglyl-L-carnitine, C5:1-DC—glutaconyl-L-carnitine, C5-DC/C6-OH—glutaryl-L-carnitine/hydroxyhexanoyl-L-carnitine, C5-M-DC—methylglutaryl-L-carnitine, C6:1—hexenoyl-L-carnitine, C7-DC—pimetyl-L-carnitine, C8—octanoyl-L-carnitine, C8:1—octenoyl-L-carnitine, C9—nonayl-L-carnitine, LPC—lysophosphatidylcholine, PC—phosphatidylcholine, GPCe—etherphosphorylcholine, SM—sphingomyelin.

(DOCX)

S7 Table. Correlation analysis of metabolites in seminal plasma with spermogram parameters of 20 healthy donors. Data are Spearman correlation rank coefficients. Significant

positive correlations are highlighted in red, significant negative correlations in blue. Abbreviations: Ala—alanine, Arg—arginine, Asn—asparagine, Asp—aspartate, Cit—citrulline, Gln—glutamine, Glu—glutamate, Gly—glycine, His—histidine, Ile—ileucine, Leu—leucine, Lys—lysine, Met—methionine, Orn—ornithine, Phe—phenylalanine, Pro—proline, Ser—serine, Thr—threonine, Trp—tryptophan, Tyr—tyrosine, Val—valine, ADMA—asymmetrically dimethylated arginine, alpha-AAA— α -amino adipic acid, OH-Pro—hydroxyproline, DMA—dimethylamine, C0—DL-carnitine, C10—decanoyl-L-carnitine, C10:1—decanoyl-L-carnitine, C10:2—decadienyl-L-carnitine, C12—dodecanoyl-L-carnitine, C12:1—dodecenoyl-L-carnitine, C12-DC—dodecanedioyl-L-carnitine, C14—tetradecanoyl-L-carnitine, C14:1—tetradecenoyl-L-carnitine, C14:1-OH—hydroxytetradecenoyl-L-carnitine, C14:2—tetradecadienyl-L-carnitine, C14:2-OH—hydroxytetradecadienyl-L-carnitine, C16—hexadecanoyl-L-carnitine, C16:1—hexadecenoyl-L-carnitine, C16:1-OH—hydroxyhexadecenoyl-L-carnitine, C16:2—hexadecadienyl-L-carnitine, C16:2-OH—hydroxyhexadecadienyl-L-carnitine, C16-OH—hydroxyhexadecanoyl-L-carnitine, C18—octadecanoyl-L-carnitine, C18:1—octadecenoyl-L-carnitine, C18:1-OH—hydroxyoctadecenoyl-L-carnitine, C18:2—octadecadienyl-L-carnitine, C2—acetyl-L-carnitine, C3—propionyl-L-carnitine, C3:1—propenyl-L-carnitine, C3-DC/C4-OH—malonyl-L-carnitine/hydroxybutyryl-L-carnitine, C3-DC-M/C5-OH—methylmalonyl-L-carnitine/hydroxyvaleryl-L-carnitine, C3-OH—hydroxypropionyl-L-carnitine, C4—butyryl-L-carnitine, C4:1—butenyl-L-carnitine, C4:1-DC/C6—fumaryl-L-carnitine/hexanoyl-L-carnitine, C5—valeryl-L-carnitine, C5:1—tiglyl-L-carnitine, C5:1-DC—glutaconyl-L-carnitine, C5-DC/C6-OH—glutaryl-L-carnitine/hydroxyhexanoyl-L-carnitine, C5-M-DC—methylglutaryl-L-carnitine, C6:1—hexenoyl-L-carnitine, C7-DC—pimetyl-L-carnitine, C8—octanoyl-L-carnitine, C8:1—octenoyl-L-carnitine, C9—nonayl-L-carnitine, LPC—lysophosphatidylcholine, PC—phosphatidylcholine, GPCe—etherphosphorylcholine, SM—sphingomyelin.

(DOCX)

S8 Table. Correlation analysis of acylcarnitines in sperm with those in seminal plasma (SP) of 20 healthy donors. Data are Spearman correlation rank coefficients. Significances are highlighted in bolt. Abbreviations: C0—DL-carnitine, C10—decanoyl-L-carnitine, C10:1—decenoyl-L-carnitine, C10:2—decadienyl-L-carnitine, C12—dodecanoyl-L-carnitine, C12:1—dodecenoyl-L-carnitine, C12-DC—dodecanedioyl-L-carnitine, C14—tetradecanoyl-L-carnitine, C14:1—tetradecenoyl-L-carnitine, C14:1-OH—hydroxytetradecenoyl-L-carnitine, C14:2—tetradecadienyl-L-carnitine, C14:2-OH—hydroxytetradecadienyl-L-carnitine, C16—hexadecanoyl-L-carnitine, C16:1—hexadecenoyl-L-carnitine, C16:1-OH—hydroxyhexadecenoyl-L-carnitine, C16:2—hexadecadienyl-L-carnitine, C16:2-OH—hydroxyhexadecadienyl-L-carnitine, C16-OH—hydroxyhexadecanoyl-L-carnitine, C18—octadecanoyl-L-carnitine, C18:1—octadecenoyl-L-carnitine, C18:1-OH—hydroxyoctadecenoyl-L-carnitine, C18:2—octadecadienyl-L-carnitine, C2—acetyl-L-carnitine, C3—propionyl-L-carnitine, C3:1—propionyl-L-carnitine, C3-DC/C4-OH—malonyl-L-carnitine/hydroxybutyryl-L-carnitine, C3-DC-M/C5-OH—methylmalonyl-L-carnitine/hydroxyvaleryl-L-carnitine, C3-OH—hydroxypropionyl-L-carnitine, C4—butyryl-L-carnitine, C4:1—butenyl-L-carnitine, C4:1-DC/C6—fumaryl-L-carnitine/hexanoyl-L-carnitine, C5—valeryl-L-carnitine, C5:1—tiglyl-L-carnitine, C5:1-DC—glutaconyl-L-carnitine, C5-DC/C6-OH—glutaryl-L-carnitine/hydroxyhexanoyl-L-carnitine, C5-M-DC—methylglutaryl-L-carnitine, C6:1—hexenoyl-L-carnitine, C7-DC—pimelyl-L-carnitine, C8—octanoyl-L-carnitine, C8:1—octenoyl-L-carnitine, C9—nonayl-L-carnitine.
(DOCX)

S9 Table. Correlation analysis of lysophospholipids in sperm with lyso-phosphatidylcholines (LPC), phosphatidylcholines (PC), etherphosphorylcholines (GPCe) and sphingomyelins (SM) in seminal plasma (SP) of 20 healthy donors. Data are Spearman correlation rank coefficients. Significances are highlighted in bolt.
(DOCX)

S10 Table. Correlation analysis of phosphatidylcholines in sperm with lyso-phosphatidylcholines (LPC), phosphatidylcholines (PC), etherphosphorylcholines (GPCe) and sphingomyelins (SM) in seminal plasma (SP) of 20 healthy donors. Data are Spearman correlation rank coefficients. Significances are highlighted in bolt.
(DOCX)

S11 Table. Correlation analysis of ether-phosphatidylcholines in sperm with lyso-phosphatidylcholines (LPC), phosphatidylcholines (PC), etherphosphorylcholines (GPCe) and sphingomyelins (SM) in seminal plasma (SP) of 20 healthy donors. Data are Spearman correlation rank coefficients. Significances are highlighted in bolt.
(DOCX)

S12 Table. Correlation analysis of sphingomyelins (SM) in sperm with lyso-phosphatidylcholines (LPC), phosphatidylcholines (PC), etherphosphorylcholines (GPCe) and SM in seminal plasma (SP) of 20 healthy donors. Data are Spearman correlation rank coefficients. Significances are highlighted in bolt.
(DOCX)

Acknowledgments

The authors would like to thank Gabriele Kersten for technical assistance.

Author Contributions

Conceptualization: Sven Baumann, Martin von Bergen, Sonja Grunewald.

Data curation: Sven Baumann, Ulrike Rolle-Kampczyk.

Formal analysis: Kathrin M. Engel, Sonja Grunewald.

Funding acquisition: Jürgen Schiller, Sonja Grunewald.

Investigation: Kathrin M. Engel, Sven Baumann, Sonja Grunewald.

Methodology: Sven Baumann, Ulrike Rolle-Kampczyk, Sonja Grunewald.

Project administration: Ulrike Rolle-Kampczyk, Sonja Grunewald.

Resources: Sonja Grunewald.

Software: Kathrin M. Engel.

Supervision: Sonja Grunewald.

Validation: Kathrin M. Engel, Sven Baumann, Ulrike Rolle-Kampczyk, Jürgen Schiller, Martin von Bergen, Sonja Grunewald.

Visualization: Kathrin M. Engel.

Writing – original draft: Kathrin M. Engel, Sven Baumann, Sonja Grunewald.

Writing – review & editing: Kathrin M. Engel, Sven Baumann, Ulrike Rolle-Kampczyk, Jürgen Schiller, Martin von Bergen, Sonja Grunewald.

References

1. Esteves SC, Miyaoka R, Agarwal A. An update on the clinical assessment of the infertile male. *Clinics*. 2011; 66: 691–700. <https://doi.org/10.1590/S1807-59322011000400026> PMID: 21655766
2. World Health Organization. WHO Laboratory manual for the examination and processing of human sperm. 5th ed. Geneva: World Health Organization; 2010.
3. Courant F, Antignac J-P, Monteau F, Le Bizec B. Metabolomics as a potential new approach for investigating human reproductive disorders. *J Proteome Res*. 2013; 12: 2914–2920. <https://doi.org/10.1021/pr400204q> PMID: 23651426
4. Baker MA. The ‘omics revolution and our understanding of sperm cell biology. *Asian J Androl*. 2011; 13: 6–10. <https://doi.org/10.1038/aja.2010.62> PMID: 20972449
5. Silvestroni L, Morisi G, Malandrino F, Frajese G. Free Amino Acids in Semen. Measurement and Significance in Normal and Oligozoospermic Men. *Archives of Andrology*. 2009; 2: 257–261. <https://doi.org/10.3109/01485017908987322>
6. Qiao S, Wu W, Chen M, Tang Q, Xia Y, Jia W, et al. Seminal plasma metabolomics approach for the diagnosis of unexplained male infertility. *PLoS ONE*. 2017; 12: e0181115. <https://doi.org/10.1371/journal.pone.0181115> PMID: 28797078
7. Paiva C, Amaral A, Rodriguez M, Canyellas N, Correig X, Ballescà JL, et al. Identification of endogenous metabolites in human sperm cells using proton nuclear magnetic resonance ((1) H-NMR) spectroscopy and gas chromatography-mass spectrometry (GC-MS). *Andrology*. 2015; 3: 496–505. <https://doi.org/10.1111/andr.12027> PMID: 25854681
8. Engel KM, Schiller J, Müller K, Dannenberger D, Jakop U. The Phospholipid Composition of Kangaroo Spermatozoa Verified by Mass Spectrometric Lipid Analysis. *Lipids*. 2017; 52: 857–869. <https://doi.org/10.1007/s11745-017-4283-9> PMID: 28801719
9. Teuber K, Schiller J, Jakop U, Lüpold S, Orledge JM, Blount JD, et al. MALDI-TOF mass spectrometry as a simple tool to determine the phospholipid/glycolipid composition of sperm. Pheasant spermatozoa as one selected example. *Anim Reprod Sci*. 2011; 123: 270–278. <https://doi.org/10.1016/j.anireprosci.2011.01.009> PMID: 21295419
10. Nimptsch A, Pyttel S, Paasch U, Mohr C, Heinrich J-M, Schiller J. A MALDI MS investigation of the lysophosphatidylcholine/phosphatidylcholine ratio in human spermatozoa and erythrocytes as a useful fertility marker. *Lipids*. 2014; 49: 287–293. <https://doi.org/10.1007/s11745-013-3870-7> PMID: 24307496

11. Pearson BL, Posselt LP, Henzel KS, Ehninger D. Limited efficacy of somatic cell lysis buffer to purify laboratory mouse sperm. *Epigenomics*. 2018; 10: 689–694. <https://doi.org/10.2217/epi-2018-0015> PMID: [29979107](https://pubmed.ncbi.nlm.nih.gov/29979107/)
12. Grunewald S, Paasch U. Basic diagnostics in andrology. *Journal der Deutschen Dermatologischen Gesellschaft*. 2013; 11: 799. <https://doi.org/10.1111/ddg.12177> PMID: [23957479](https://pubmed.ncbi.nlm.nih.gov/23957479/)
13. Kenéz Á, Dánicse S, Rolle-Kampczyk U, von Bergen M, Huber K. A metabolomics approach to characterize phenotypes of metabolic transition from late pregnancy to early lactation in dairy cows. *Metabolomics*. 2016; 12: 1073. <https://doi.org/10.1007/s11306-016-1112-8>
14. Grunewald S, Paasch U, Glander H-J, Anderegg U. Mature human spermatozoa do not transcribe novel RNA. *Andrologia*. 2005; 37: 69–71. <https://doi.org/10.1111/j.1439-0272.2005.00656.x> PMID: [16026427](https://pubmed.ncbi.nlm.nih.gov/16026427/)
15. Johnson GD, Sendler E, Lalancette C, Hauser R, Diamond MP, Krawetz SA. Cleavage of rRNA ensures translational cessation in sperm at fertilization. *Mol Hum Reprod*. 2011; 17: 721–726. <https://doi.org/10.1093/molehr/gar054> PMID: [21831882](https://pubmed.ncbi.nlm.nih.gov/21831882/)
16. Tut TG, Ghadessy FJ, Trifiro MA, Pinsky L, Yong EL. Long polyglutamine tracts in the androgen receptor are associated with reduced trans-activation, impaired sperm production, and male infertility. *J Clin Endocrinol Metab*. 1997; 82: 3777–3782. <https://doi.org/10.1210/jcem.82.11.4385> PMID: [9360540](https://pubmed.ncbi.nlm.nih.gov/9360540/)
17. Zitzmann M, Nieschlag E. The CAG repeat polymorphism within the androgen receptor gene and male-ness1. *Int J Androl*. 2003; 26: 76–83. <https://doi.org/10.1046/j.1365-2605.2003.00393.x> PMID: [12641825](https://pubmed.ncbi.nlm.nih.gov/12641825/)
18. Brosnan JT. Interorgan amino acid transport and its regulation. *J Nutr*. 2003; 133: 2068S–2072S. <https://doi.org/10.1093/jn/133.6.2068S> PMID: [12771367](https://pubmed.ncbi.nlm.nih.gov/12771367/)
19. Schmidt JA, Rinaldi S, Scalbert A, Ferrari P, Achaintre D, Gunter MJ, et al. Plasma concentrations and intakes of amino acids in male meat-eaters, fish-eaters, vegetarians and vegans. A cross-sectional analysis in the EPIC-Oxford cohort. *Eur J Clin Nutr*. 2016; 70: 306–312. <https://doi.org/10.1038/ejcn.2015.144> PMID: [26395436](https://pubmed.ncbi.nlm.nih.gov/26395436/)
20. Breier M, Wahl S, Prehn C, Fugmann M, Ferrari U, Weise M, et al. Targeted metabolomics identifies reliable and stable metabolites in human serum and plasma samples. *PLoS ONE*. 2014; 9: e89728. <https://doi.org/10.1371/journal.pone.0089728> PMID: [24586991](https://pubmed.ncbi.nlm.nih.gov/24586991/)
21. Zhao K, Zhang J, Xu Z, Xu Y, Xu A, Chen W, et al. Metabolomic Profiling of Human Spermatozoa in Idiopathic Asthenozoospermia Patients Using Gas Chromatography-Mass Spectrometry. *Biomed Res Int*. 2018; 2018: 8327506. <https://doi.org/10.1155/2018/8327506> PMID: [29682560](https://pubmed.ncbi.nlm.nih.gov/29682560/)
22. Shettles LB. The Relation of Dietary Deficiencies to Male Fertility. *Fertility and Sterility*. 1960; 11: 88–99. [https://doi.org/10.1016/S0015-0282\(16\)33665-2](https://doi.org/10.1016/S0015-0282(16)33665-2) PMID: [14445928](https://pubmed.ncbi.nlm.nih.gov/14445928/)
23. Holt L, Emmett Jr., Albanese Anthony, A. Observations on amino acid deficiencies in man. *Tans Assoc Am Physicians*. 1944: 143–156.
24. Wu G, Bazer FW, Davis TA, Kim SW, Li P, Marc Rhoads J, et al. Arginine metabolism and nutrition in growth, health and disease. *Amino Acids*. 2009; 37: 153–168. <https://doi.org/10.1007/s00726-008-0210-y> PMID: [19030957](https://pubmed.ncbi.nlm.nih.gov/19030957/)
25. Tanimura J. Studies on arginine in human semen. II. The effects of medication with L-arginine-HCL on male infertility. *Bull Osaka Med Sch*. 1967; 13: 84–89. PMID: [6080242](https://pubmed.ncbi.nlm.nih.gov/6080242/)
26. Ijiri TW, Mahbub Hasan AKM, Sato K-I. Protein-tyrosine kinase signaling in the biological functions associated with sperm. *J Signal Transduct*. 2012; 2012: 181560. <https://doi.org/10.1155/2012/181560> PMID: [23209895](https://pubmed.ncbi.nlm.nih.gov/23209895/)
27. Mahony MC, Gwathmey T. Protein Tyrosine Phosphorylation during Hyperactivated Motility of Cynomolgus Monkey (*Macaca fascicularis*) Spermatozoa1. *Biology of Reproduction*. 1999; 60: 1239–1243. <https://doi.org/10.1095/biolreprod60.5.1239> PMID: [10208990](https://pubmed.ncbi.nlm.nih.gov/10208990/)
28. Bhattacharjee R, Goswami S, Dudiki T, Popkie AP, Phiel CJ, Kline D, et al. Targeted disruption of glycogen synthase kinase 3A (GSK3A) in mice affects sperm motility resulting in male infertility. *Biology of Reproduction*. 2015; 92: 65. <https://doi.org/10.1095/biolreprod.114.124495> PMID: [25568307](https://pubmed.ncbi.nlm.nih.gov/25568307/)
29. Dudiki T, Kadunganattil S, Ferrara JK, Kline DW, Vijayaraghavan S. Changes in Carboxy Methylation and Tyrosine Phosphorylation of Protein Phosphatase PP2A Are Associated with Epididymal Sperm Maturation and Motility. *PLoS ONE*. 2015; 10: e0141961. <https://doi.org/10.1371/journal.pone.0141961> PMID: [26569399](https://pubmed.ncbi.nlm.nih.gov/26569399/)
30. Pereira R, Sá R, Barros A, Sousa M. Major regulatory mechanisms involved in sperm motility. *Asian J Androl*. 2017; 19: 5–14. <https://doi.org/10.4103/1008-682X.167716> PMID: [26680031](https://pubmed.ncbi.nlm.nih.gov/26680031/)
31. Amaral A, Paiva C, Attardo Parrinello C, Estanyol JM, Ballescà JL, Ramalho-Santos J, et al. Identification of proteins involved in human sperm motility using high-throughput differential proteomics. *J Proteome Res*. 2014; 13: 5670–5684. <https://doi.org/10.1021/pr500652y> PMID: [25250979](https://pubmed.ncbi.nlm.nih.gov/25250979/)

32. Dungdung SR, Bhoumik A, Saha S, Ghosh P, Das K, Mukherjee S, et al. Sperm Motility Regulatory Proteins. A Tool to Enhance Sperm Quality. In: Payan Carreira R, editor. *Insights from Animal Reproduction*: InTech; 2016.
33. Bottrel M, Acha D, Ortiz I, Hidalgo M, Gósalvez J, Camisã J, et al. Cryoprotective effect of glutamine, taurine, and proline on post-thaw semen quality and DNA integrity of donkey spermatozoa. *Anim Reprod Sci*. 2018; 189: 128–135. <https://doi.org/10.1016/j.anireprosci.2017.12.021> PMID: 29325880
34. Partyka A, Rodak O, Bajzert J, Kochan J, Niżański W. The Effect of L-Carnitine, Hypotaurine, and Taurine Supplementation on the Quality of Cryopreserved Chicken Semen. *Biomed Res Int*. 2017; 2017: 7279341. <https://doi.org/10.1155/2017/7279341> PMID: 28523277
35. Zhang X, Diao R, Zhu X, Li Z, Cai Z. Metabolic characterization of asthenozoospermia using nontargeted seminal plasma metabolomics. *Clin Chim Acta*. 2015; 450: 254–261. <https://doi.org/10.1016/j.cca.2015.09.001> PMID: 26342261
36. Edwards KDG, Whyte HM. Plasma creatinine level and creatinine clearance as tests of renal function. *Australasian Annals of Medicine*. 1959; 8: 218–224. <https://doi.org/10.1111/imj.1959.8.3.218> PMID: 13819423
37. Ellerbrock RE, Honorato J, Curcio BR, Stewart JL, Souza JAT, Love CC, et al. Effect of urine contamination on stallion semen freezing ability. *Theriogenology*. 2018; 117: 1–6. <https://doi.org/10.1016/j.theriogenology.2018.05.010> PMID: 29800826
38. Sidhu RS, Sharma RK, Agarwal A. Relationship between creatine kinase activity and semen characteristics in subfertile men. *Int J Fertil Womens Med*. 1998; 43: 192–197. PMID: 9726847
39. Hallak J, Sharma RK, Pasqualotto FF, Ranganathan P, Thomas AJ, Agarwal A. Creatine kinase as an indicator of sperm quality and maturity in men with oligospermia. *Urology*. 2001; 58: 446–451. PMID: 11549497
40. Mann T. Secretory function of the prostate, seminal vesicle and other male accessory organs of reproduction. *J Reprod Fertil*. 1974; 37: 179–188. PMID: 4593605
41. Vanella A, Pinturo R, Vasta M, Piazza G, Rapisarda A, Savoca S, et al. Polyamine levels in human semen of infertile patients. Effect of S-adenosylmethionine. *Acta Eur Fertil*. 1978; 9: 99–103. PMID: 726797
42. Guo D, Wu W, Tang Q, Qiao S, Chen Y, Chen M, et al. The impact of BMI on sperm parameters and the metabolite changes of seminal plasma concomitantly. *Oncotarget*. 2017; 8: 48619–48634. <https://doi.org/10.18632/oncotarget.14950> PMID: 28159940
43. Rubinstein S, Breitbart H. Role of spermine in mammalian sperm capacitation and acrosome reaction. *Biochem J*. 1991; 278 (Pt 1): 25–28.
44. Gonzales GF, Garcia-Hjarles M, Velasquez G. Hyperprolactinaemia and hyperserotoninaemia. Their relationship to seminal quality. *Andrologia*. 1992; 24: 95–100. PMID: 1605404
45. Caldamone AA, Al-Juburi A, Cockett AT. The varicocele. Elevated serotonin and infertility. *J Urol*. 1980; 123: 683–685. PMID: 7420556
46. Tanrikut C, Schlegel PN. Antidepressant-associated changes in semen parameters. *Urology*. 2007; 69: 185.e5–7. <https://doi.org/10.1016/j.urology.2006.10.034> PMID: 17270655
47. Tanrikut C, Feldman AS, Altemus M, Paduch DA, Schlegel PN. Adverse effect of paroxetine on sperm. *Fertility and Sterility*. 2010; 94: 1021–1026. <https://doi.org/10.1016/j.fertnstert.2009.04.039> PMID: 19515367
48. Longo N, Frigeni M, Pasquali M. Carnitine transport and fatty acid oxidation. *Biochim Biophys Acta*. 2016; 1863: 2422–2435. <https://doi.org/10.1016/j.bbamcr.2016.01.023> PMID: 26828774
49. Aliabadi E, Soleimani Mehranjani M, Borzoei Z, Talaei-Khozani T, Mirkhani H, Tabesh H. Effects of L-carnitine and L-acetyl-carnitine on testicular sperm motility and chromatin quality. *Iran J Reprod Med*. 2012; 10: 77–82. PMID: 25242977
50. Kohengkul S, Tanphaichitr V, Muangmun V, Tanphaichitr N. Levels of L-carnitine and L-O-acetylcarnitine in normal and infertile human semen: a lower level of L-O-acetylcarnitine in infertile semen. *Fertility and Sterility*. 1977; 28: 1333–1336. PMID: 590542
51. Busetto GM, Agarwal A, Virmani A, Antonini G, Ragonesi G, Del Giudice F, et al. Effect of metabolic and antioxidant supplementation on sperm parameters in oligo-astheno-teratozoospermia, with and without varicocele. A double-blind placebo-controlled study. *Andrologia*. 2018; 50. <https://doi.org/10.1111/and.12927> PMID: 29315686
52. Zöpfigen A, Priem F, Sudhoff F, Jung K, Lenk S, Loening SA, et al. Relationship between semen quality and the seminal plasma components carnitine, alpha-glucosidase, fructose, citrate and granulocyte elastase in infertile men compared with a normal population. *Hum Reprod*. 2000; 15: 840–845. PMID: 10739829

53. Schiller J, Arnhold J, Glander H-J, Arnold K. Lipid analysis of human spermatozoa and seminal plasma by MALDI-TOF mass spectrometry and NMR spectroscopy—effects of freezing and thawing. *Chemistry and Physics of Lipids*. 2000; 106: 145–156. [https://doi.org/10.1016/S0009-3084\(00\)00148-1](https://doi.org/10.1016/S0009-3084(00)00148-1) PMID: [10930566](https://pubmed.ncbi.nlm.nih.gov/10930566/)
54. Lessig J, Gey C, Süß R, Schiller J, Glander H-J, Arnhold J. Analysis of the lipid composition of human and boar spermatozoa by MALDI-TOF mass spectrometry, thin layer chromatography and ³¹P NMR spectroscopy. *Comp Biochem Physiol B, Biochem Mol Biol*. 2004; 137: 265–277. <https://doi.org/10.1016/j.cbpc.2003.12.001> PMID: [14990223](https://pubmed.ncbi.nlm.nih.gov/14990223/)
55. Andersen JM, Rønning PO, Herning H, Bekken SD, Haugen TB, Witczak O. Fatty acid composition of spermatozoa is associated with BMI and with semen quality. *Andrology*. 2016; 4: 857–865. <https://doi.org/10.1111/andr.12227> PMID: [27371336](https://pubmed.ncbi.nlm.nih.gov/27371336/)
56. Henriksen JR, Andresen TL, Feldborg LN, Duelund L, Ipsen JH. Understanding detergent effects on lipid membranes. A model study of lysolipids. *Biophys J*. 2010; 98: 2199–2205. <https://doi.org/10.1016/j.bpj.2010.01.037> PMID: [20483328](https://pubmed.ncbi.nlm.nih.gov/20483328/)
57. Glander H-J, Schiller J, Süß R, Paasch U, Grunewald S, Arnhold J. Deterioration of spermatozoal plasma membrane is associated with an increase of sperm lyso-phosphatidylcholines. *Andrologia*. 2002; 34: 360–366. PMID: [12472619](https://pubmed.ncbi.nlm.nih.gov/12472619/)
58. Fuchs B, Muller K, Paasch U, Schiller J. Lysophospholipids. Potential markers of diseases and infertility. *Mini Rev Med Chem*. 2012; 12: 74–86. PMID: [22070693](https://pubmed.ncbi.nlm.nih.gov/22070693/)
59. Pyttel S, Nimptsch A, Böttger J, Zschörnig K, Jakob U, Wegener J, et al. Changes of murine sperm phospholipid composition during epididymal maturation determined by MALDI-TOF mass spectrometry. *Theriogenology*. 2014; 82: 396–402. <https://doi.org/10.1016/j.theriogenology.2014.04.017> PMID: [24877723](https://pubmed.ncbi.nlm.nih.gov/24877723/)
60. Pyttel S, Zschörnig K, Nimptsch A, Paasch U, Schiller J. Enhanced lysophosphatidylcholine and sphingomyelin contents are characteristic of spermatozoa from obese men—A MALDI mass spectrometric study. *Chemistry and Physics of Lipids*. 2012; 165: 861–865. <https://doi.org/10.1016/j.chemphyslip.2012.11.001> PMID: [23146833](https://pubmed.ncbi.nlm.nih.gov/23146833/)
61. Lu J-C, Jing J, Yao Q, Fan K, Wang G-H, Feng R-X, et al. Relationship between Lipids Levels of Serum and Seminal Plasma and Semen Parameters in 631 Chinese Subfertile Men. *PLoS ONE*. 2016; 11: e0146304. <https://doi.org/10.1371/journal.pone.0146304> PMID: [26726884](https://pubmed.ncbi.nlm.nih.gov/26726884/)