

RESEARCH

Open Access

MicroRNA expressions associated with progression of prostate cancer cells to antiandrogen therapy resistance

Richard Ottman, Camha Nguyen, Robert Lorch and Ratna Chakrabarti*

Abstract

Background: Development of resistance to androgen deprivation therapy (ADT) is a major obstacle for the management of advanced prostate cancer. Therapies with androgen receptor (AR) antagonists and androgen withdrawal initially regress tumors but development of compensatory mechanisms including AR bypass signaling leads to re-growth of tumors. MicroRNAs (miRNAs) are small regulatory RNAs that are involved in maintenance of cell homeostasis but are often altered in tumor cells.

Results: In this study, we determined the association of genome wide miRNA expression (1113 unique miRNAs) with development of resistance to ADT. We used androgen sensitive prostate cancer cells that progressed to ADT and AR antagonist Casodex (CDX) resistance upon androgen withdrawal and treatment with CDX. Validation of expression of a subset of 100 miRNAs led to identification of 43 miRNAs that are significantly altered during progression of cells to treatment resistance. We also show a correlation of altered expression of 10 proteins targeted by some of these miRNAs in these cells.

Conclusions: We conclude that dynamic alterations in miRNA expression occur early on during androgen deprivation therapy, and androgen receptor blockade. The cumulative effect of these altered miRNA expression profiles is the temporal modulation of multiple signaling pathways promoting survival and acquisition of resistance. These early events are driving the transition to castration resistance and cannot be studied in already developed CRPC cell lines or tissues. Furthermore our results can be used a prognostic marker of cancers with a potential to be resistant to ADT.

Keywords: MicroRNA, Prostate cancer, Antiandrogen, Androgen deprivation, Drug resistance, Expression profiling

Background

Prostate cancer is the most commonly diagnosed cancer and a leading cause of cancer related death in men in developed countries. Because androgen is required for normal growth and functioning of the prostate gland and also for development of cancer androgen deprivation therapy (ADT) has become the mainstay for advanced prostate cancer [1]. Although most patients initially respond to ADT by showing low PSA values, they eventually develop more aggressive castration resistant prostate cancer (CRPC). Androgen, working through androgen receptor (AR) triggers transcriptional

activation of a variety of genes that are essential for growth and survival of prostate epithelial cells. However, prolonged androgen blockade using steroidal or non-steroidal inhibitors such as, cyproterone acetate, Casodex or hydroxyl flutamide, leads to activation of various adaptive mechanisms after initial retardation of cell proliferation [2,3].

Development of resistance to ADT, which includes reduction in androgen synthesis and direct antagonism of the androgen receptors (AR), can occur as a result of high expression and activation of AR [4]. Activation of AR without androgen is through switching of AR to alternative mechanism of activation. Commonly noted mechanisms include AR gene amplification, increased coactivator expression, selection of AR gene mutation and sensitivity to growth factors and cytokines [5]. AR

* Correspondence: ratna.chakrabarti@ucf.edu
Burnett School of Biomedical Sciences, University of Central Florida, 12722
Research Parkway, Orlando, Florida, USA

expression and activity also increased after long-term androgen ablation to a level that was comparable to that in parental cell lines or androgen dependent (AD) tumors prior to castration [6]. It has been proposed that the increased activity of AR in androgen independent (AI) cells or in relapsed tumors in castrated xenograft mice is mediated through ligand independent mechanism [7] or through promiscuous sensitivity of AR to other steroid hormone, growth factors or cytokines [8]. Androgen blockade therapy can accumulate mutations causing AR to become sensitive to androgen antagonists, which then act as agonists [2,9]. It is now accepted that CRPC maintains functional AR signaling [10,11] but androgen refractoriness is through an AR bypass or adaptive mechanism, which is possibly, mediated through cytokines or other survival factors. Irrespective of the specific phenotypes acquired by the AD prostate cancers, the outcome is altered expression of protein-coding genes as a whole that are responsible for progression and metastasis of prostate cancer. In addition, nonsteroidal agents such as flutamide can alter gene expression in AR negative prostate tumors [12]. Gene expression profiling in androgen dependent and androgen independent prostate cancers revealed an increasingly complex profile of gene expression in prostate cancer with respect to the status of androgen sensitivity or refractoriness [13]. However, the exact mechanism of altered gene expression in CRPC is not clear.

The role of small noncoding microRNAs (miRNAs) in regulation of gene expression, which is mediated by inhibition of translation or degradation of target mRNAs is an established phenomenon [14]. MiRNAs belong to a class of 17–22 nucleotides, which contains a specific sequence at the 5' end and regulates translation through binding to 3'UTR of the mRNAs [14]. To date, there are 1921 distinct human miRNAs have been identified, each of which regulate multiple target mRNAs (www.mirbase.org Nov 2011). Genes encoding miRNAs are located in the intergenic regions or within the protein-coding genes either alone or in clusters [15]. There is now abundant evidence that aberrant expression of miRNAs occurs in diverse types of cancer including prostate cancer and during different stages of disease progression [16,17]. The role of miRNAs in regulation of post-transcriptional gene expression has been implicated in 30% of the protein-coding genes [18]. Because miRNAs can be overexpressed or down regulated in cancer cells these noncoding RNAs are designated as oncogenic miRNAs or suppressor miRNAs. Functionally, miRNAs reduce the levels of many of their target mRNAs and the amount of proteins encoded by these mRNAs [19]. Because a given miRNA may have many mRNA targets the biological effects of changes in miRNA expression is likely to be dependent on the cellular environment.

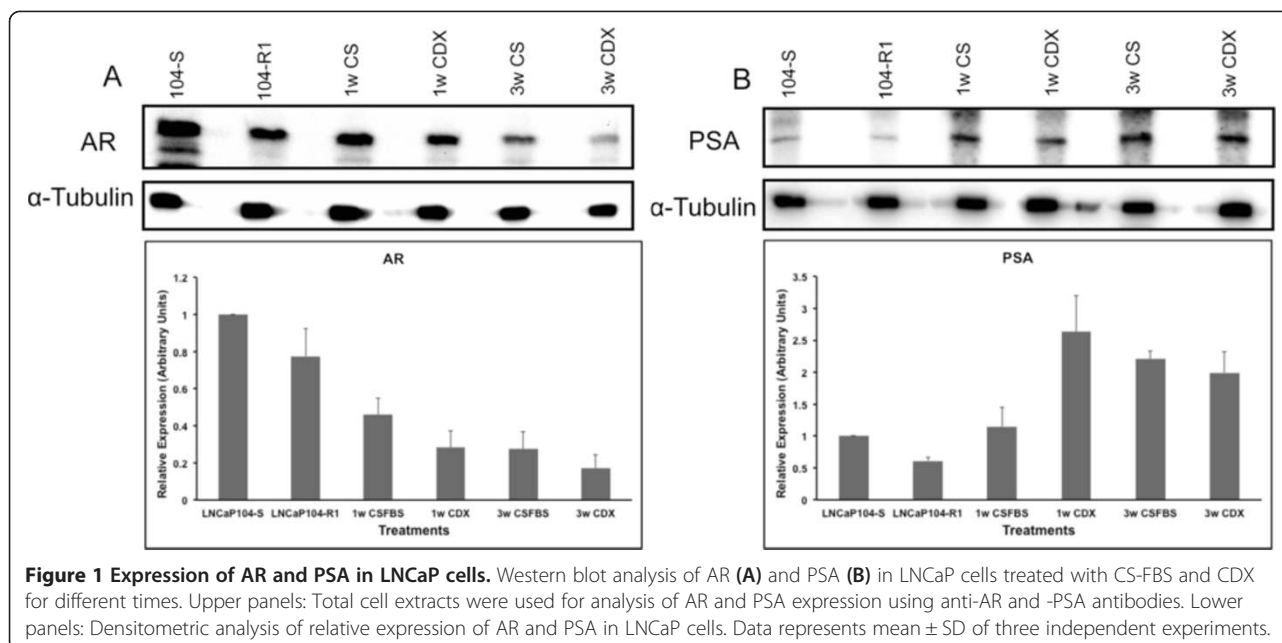
A number of studies indicated aberrant expression of miRNAs in CRPC compared to AD prostate cancer cells [20,21]. Several miRNAs, such as miR-21, miR-125 and miR-32 are directly regulated by androgens in cells and xenograft models [20,22,23]. Studies include comparative analysis between androgen sensitive (AS) and -resistant prostate cancer cells, with or without treatment of AS cells with androgens or normal vs. hormone refractory prostate cancer tissues, which only provides steady state status of miRNA and gene expression. However none of these studies provide information on the mechanism of transition of androgen-sensitive or dependent prostate cancer cells to antiandrogen resistant cells. In this study, we show, for the first time alteration in expression of miRNAs and their target proteins as the cells progress to antiandrogen resistance, some of which are not detectable in the established AI cell line.

Results

MicroRNA expression profile differentiates between untreated LNCaP cells and cells treated with Casodex or subjected to androgen withdrawal:

We used genome-wide miRNA array (1113 unique primers) profiling approach to identify specific miRNAs that are involved in development of resistance to Casodex (CDX). A clonal subline of LNCaP cells LNCaP-104S (-104S) and its androgen-independent derivative LNCaP-104R1 (-104R1) were used for monitoring differential expression of miRNAs upon treatment with CDX. LNCaP-104S cells are CDX-sensitive, whereas LNCaP-104R1 cells are not despite expressing AR at a basal level higher than LNCaP-104S cells [24]. LNCaP-104S cells require DHT for maintaining their AD status but when treated with CDX for 3 weeks in CS-FBS (charcoal-stripped FBS), CDX insensitive colonies develop that are independent of androgen (CDXR). During the first week of both CS-FBS and CDX treatments, LNCaP-104S cells grew without significant cell death, however, during the second week of CDX treatment about 60% cells died and detached. From the residual 40% cells, half of the cell population remained viable during the third week of CDX treatment. CS-FBS -treated cells did not show as much cell death during the second week of treatment and started to regrow during the third week of treatment. Cell morphology also changed as they are treated with CDX and CS-FBS (Additional file 1: Figure S1). Cells viable after third week of treatments were used for miRNA profiling experiments. Expression of AR in these cells showed gradual reduction as the treatment progresses, but the PSA expression increased, which suggests increased transcriptional activity of the residual AR in the treated cells (Figure 1).

We compared miRNA expressions in RNA extracts from untreated and CDX treated -104S cells at 0 hr, 1wk and 3wks and untreated -104R1 cells (Table 1). Hierarchical



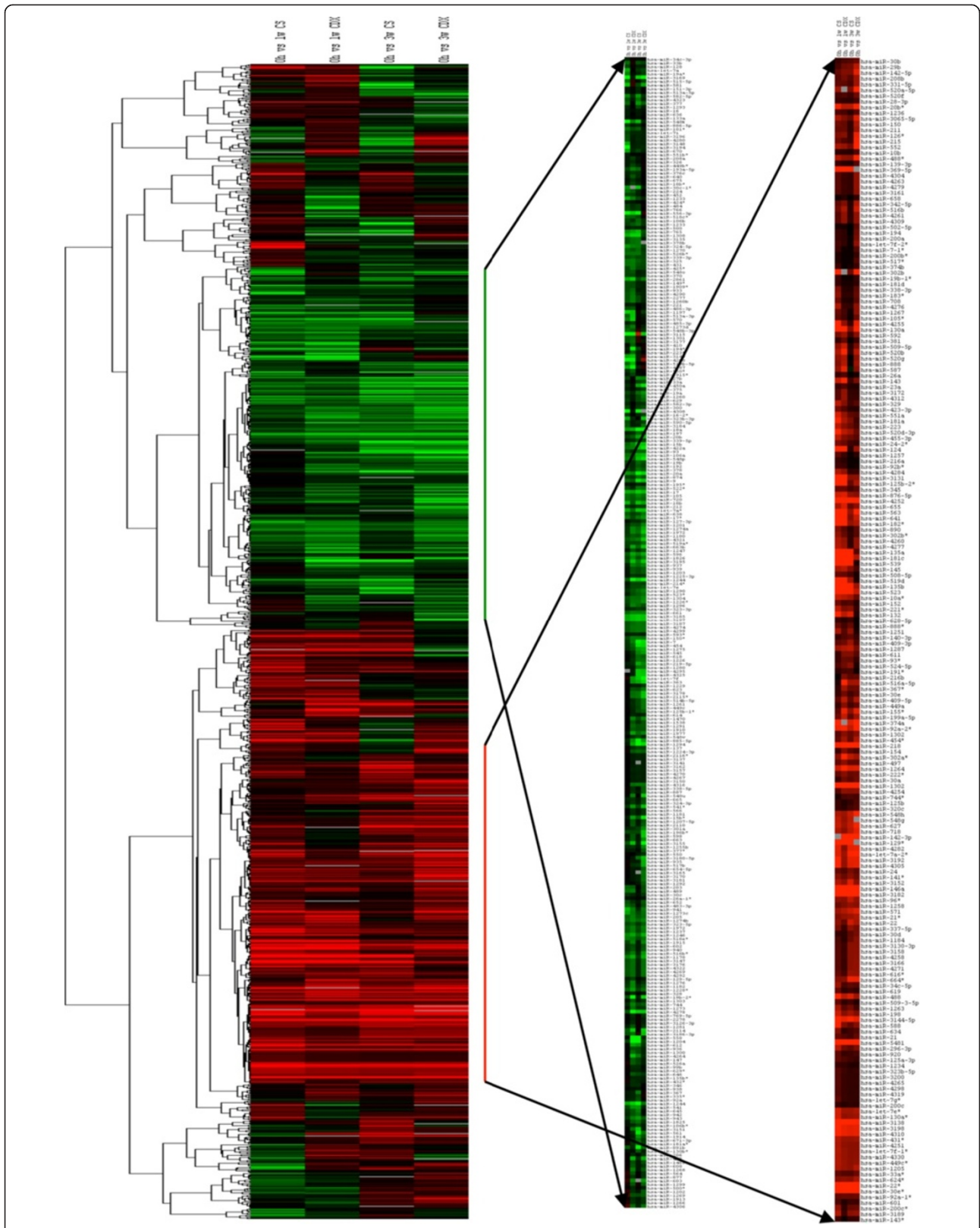
clustering of the normalized and \log_2 transformed expression data (Additional file 2: Table S1) showed 5 distinct clusters of miRNAs (Additional file 3: Figure S2). Comparative analysis between samples from different conditions using two samples Welch t-test with \log_{10} transformed data and p values of 0.05 showed significant miRNAs that are differentially expressed between conditions (Additional file 4: Table S2 and Additional file 5: Figure S3). Volcano plot (V plots) of the t-test between LNCaP-104S cells and all other samples showed 38 significant miRNAs, of which 27 miRNAs were up regulated and 11 down regulated compared to -104S (Additional file 5: Figure S3A). Comparison between untreated -104S and -104R1 cells showed 24 significant miRNAs, which includes 16 down regulated and 8 up regulated miRNAs in -104R1 (Additional file 5: Figure S3B). Differential expression of 17 significant miRNAs was observed between untreated LNCaP-104S cells and -104S cells treated with CDX, of which 13 were up regulated and 4 were down regulated CDX treated cells (Additional file 5:

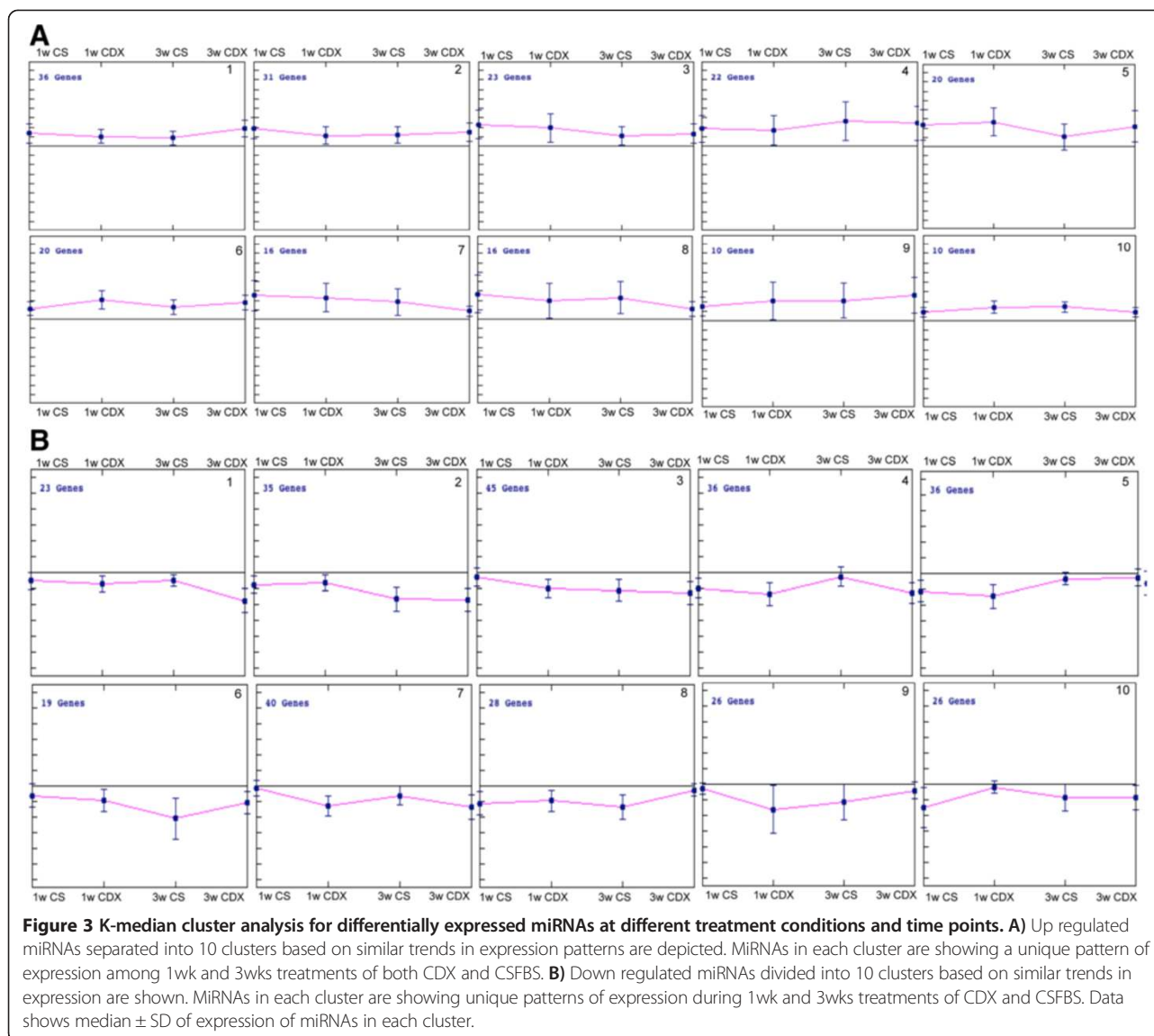
Figure S3C). LNCaP-104S and -104S cells treated with CSFBS also showed 9 up regulated and 5 down regulated microRNAs in CSFBS treated cells (Additional file 5: Figure S3D). Although -104R1 cells are CDX resistant there are differences in miRNA expression when -104S cells were treated with CDX (Additional file 5: Figure S3E). T-test analysis showed 24 significant miRNAs of which 18 miRNAs were up regulated and 6 down regulated in -104R1 cells. Difference in miRNA expressions was also noted between -104S cells maintained in androgen-depleted condition and AI -104R1 cells. Twenty-four significant miRNAs were identified of which 12 were up regulated and 12 down regulated in -104R1 cells (Additional file 5: Figure S3F). Comparison between androgen depletion and CDX treatment showed 5 significant miRNAs, 4 of which were up regulated and one down regulated in CDX treated cells (Additional file 5: Figure S3G).

Clustering analyses using \log_2 transformed fold change (FC) values of four treatment conditions compared to -104S untreated cells showed two distinct clusters of up and down regulated miRNAs, which includes 307 down regulated and 197 up regulated miRNAs (Figure 2 and Additional file 6: Table S3). K-median clustering for the up-regulated miRNAs showed a trend of gradual increase in median FC in miRNAs in some clusters (cluster 1, 4 and 9) and a gradual decrease in some clusters (clusters 3, 7 and 8) from 1 week to 3 weeks treatments (Figure 3A and Additional file 7: Table S4). In down regulated profile there is also a trend of gradual decrease in median expression of miRNAs in some clusters (clusters 1, 2, 3 and 7) (Figure 3B and Additional file 7: Table S4). Of these lists, 100 miRNAs were chosen based on fold

Table 1 Cell lines and treatments

		Samples		
		Cell line	Treatment	Time point
Reference	0 hr	LNCaP-104S	FBS-DHT 1nM	0 hr
	1wk CSFBS	LNCaP-104S	CSFBS	1 wk
	3wks CSFBS	LNCaP-104S	CSFBS	3 wks
	1wk CDX	LNCaP-104S	CSFBS/5 μ M CDX	1 wk
	3wks CDX	LNCaP-104S	CSFBS/5 μ M CDX	3 wks
Test/Reference	0 hr	LNCaP-104R1	CSFBS	0 wks





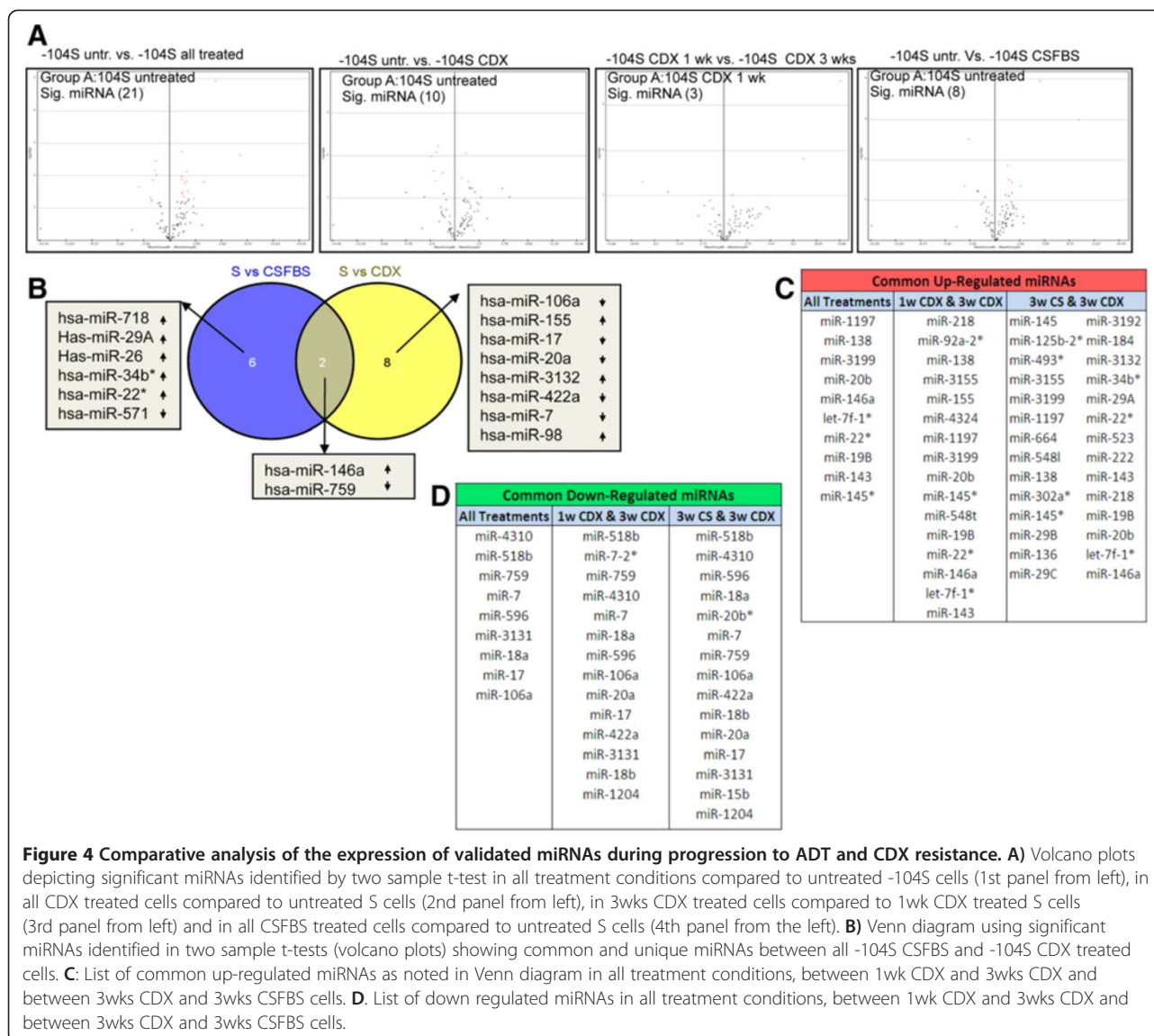
change and/or the z score ≥ 3.0 or ≤ -3.0 for validation using qPCR.

Validated expression of miRNAs revealed distinct differences in expression in different treatment conditions

Analysis of qPCR data indicated a variable expression profile of a subset of miRNAs in LNCaP cells exposed to CDX and/or androgen withdrawal (CSFBS) for different time periods. Two sample Welch t-test with a p -value < 0.05 showed a significant change in expression of 21 miRNAs in treated samples compared to the untreated -104S cells (Figure 4A and Additional file 8: Table S5). Comparison between -104S untreated and either CDX or CSFBS treated samples identified 10 and 8 significant miRNAs respectively (Figure 4A) Comparative expression analysis between CDX 1wk and CDX 3wks also revealed significant change in expression in 3 miRNAs (Figure 4A). Venn Diagram of

miRNA expression profiles following CDX treatment and androgen withdrawal indicated two common miRNAs (miR-146a and miR-759) but treatment specific differential expression was observed with 8 and 6 unique miRNAs, in CDX and CSFBS treated respectively (Figure 4B). Analysis of the up-regulated miRNAs revealed 10 miRNAs that showed 2-fold or higher expressions in all treatment conditions and time points (Figure 4C). There are also 30 miRNAs that showed 2-fold or higher expression in more than one treatment conditions. In the down regulated list, 9 miRNAs showed at least 2-fold reduction in expression in all treatment conditions and 15 miRNAs with ≥ 2.0 -fold down regulation in more than one treatment conditions (Figure 4D).

Cluster analysis of \log_2 transformed FC in expression of 100 miRNAs showed three distinct clusters displaying common expression changes in 2 time points of two

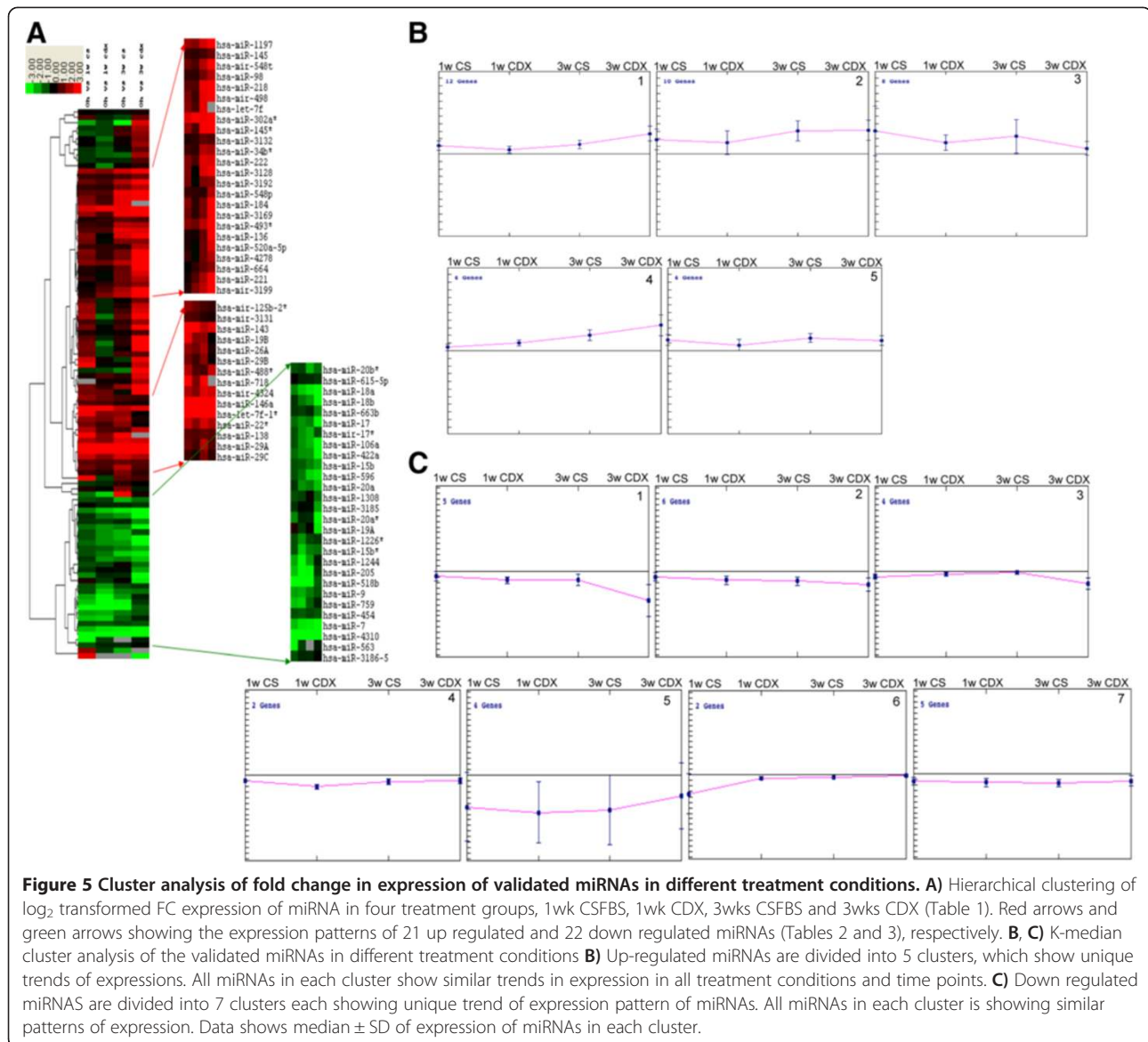


reatment conditions. Two clusters contained miRNAs with increased expression and one cluster containing down regulated miRNAs (Figure 5A and Additional file 9: Table S6). K-median clustering of the up-regulated miRNAs showed a distinct trend of gradual increase (clusters 1 [32%], 2 [26%] and 4[11%]) and gradual decrease (cluster 3 14%) in FC expressions as the treatment progressed (Figure 5B and Additional file 10: Table S7). Some of the miRNAs (21) showed across the board up regulations (Table 2). Similarly, in the list of down-regulated miRNAs a gradual decrease in expression (clusters 1[18%] and 2 [21%]) could be noted as the treatment progressed (Figure 5C) and a subset of miRNAs (22) showed down-regulation in all treatment conditions (Table 3). When compared with published expression database there are supporting reports on expression patterns of specific miRNAs in different types of cancer

(Tables 2 and 3). There are also opposing reports on expression of miRNAs, which includes, miR-106a [25], miR-15b [26], miR-17 [27], miR-18 [28], miR-205 [29], miR-20 [30] and miR-7 [31] for down regulated miRNAs; and let-7f-1 [32], miR-136a [33], miR-143 [34], miR-146a [35], miR-218 [36], miR-22 [37], miR-222 [38] miR-29a [39], miR-34b [40], and miR-493 [41] for up-regulated miRNAs.

Involvement of miRNAs in specific cellular processes which differ between treatment conditions

MicroRNAs exhibiting up regulation or down regulation in all treatment conditions compared to untreated -104S cells (Tables 2 and 3), were used for function and disease relevance to understand the possible alterations in the cellular processes as LNCaP cells progressed towards androgen withdrawal and AR antagonist resistance. We



used RT-PCR FC values of the up regulated or down regulated miRNA for the analysis using IPA software (Ingenuity Systems). Based on $-\log(p\text{-values})$ with $-values < 0.05$, a percentage of miRNAs were assigned to certain disorder or cellular processes (Additional file 11: Figure S4). When the functional profiles of the p-regulated miRNAs were compared, there is a decrease in the percentage of miRNAs involved in cancer, dermatological disease, and hematological disease between 3wks and 1wk CDX treated cells whereas an increase in the percentage of miRNAs involved in endocrine system disease, gastrointestinal disease hepatic system disease, reproductive system disease and metabolic disease in 3wks CDX treated S cells compared to 1wk CDX treated cells (Additional file 11: Figure S4A). In cells subjected to androgen withdrawal, there are also differences in percentage of miRNAs in different cellular processes between 1wk

and 3wks treatments, which show a reduction in hematological diseases, cellular development, and reproductive system disease. An increased involvement of miRNAs in a variety of cellular processes also was noted in 3wk CSFBS treated cells, which includes cell death and survival, cell movement, endocrine system disease, gastrointestinal disease, hematological disease, hepatic system disease and metabolic disease (Additional file 11: Figure S4A).

In the list of down-regulated miRNAs, there is an increase in percentage of miRNAs involved in cell morphology, cell movement, dermatological disease, gastrointestinal disease, renal urological disease, and reproductive system disease as cells progressed from 1wk to 3wks CDX treatment. A decreased percentage of down regulated miRNAs in specific cellular processes also could be noted in these cells, which includes

Table 2 Fold changes in expressions of up regulated miRNAs and its relevance with published reports

	1wk CSFBS Δ change	1wk CDX Δ change	3wks CSFBS Δ change	3wk CDX Δ change	Expression in other Cancer	References
hsa-let-7f-1	12.50	12.17	13.91	17.64	▲ ▼	[42,43]
hsa-miR-136	3.69	3.44	5.16	4.30	▲ ▼	[44,45]
hsa-miR-143	15.80	19.67	13.92	6.25	▲ ▼	[46,47]
hsa-miR-146a	31.44	20.83	56.46	50.68	▲ ▼	[48,49]
hsa-miR-218	4.64	2.33	18.44	11.49	▲ ▼	[50,51]
hsa-miR-22	2.39	2.43	4.58	3.10	▲ ▼	[52]
hsa-miR-22*	15.91	11.56	11.78	22.47	▲ ▼	NA
hsa-miR-222	3.65	1.99	7.05	4.63	▲ ▼	[53,54]
hsa-miR-29a	2.09	1.58	4.60	4.24	▲ ▼	[55,56]
hsa-miR-302a*	2.97	7.73	3.87	4.48	▲	[57,58]
hsa-miR-3138	6.53	5.94	6.66	5.96	-	NA
hsa-miR-3144b-5p	11.25	4.39	4.47	8.53	▲	[59]
hsa-miR-3192	8.70	2.11	5.52	5.71	-	NA
hsa-miR-3199	1.25	4.54	2.71	6.76	-	NA
hsa-miR-34b*	3.15	2.34	6.69	8.64	▲ ▼	[60,61]
hsa-miR-493*	1.31	1.42	2.31	15.06	▲ ▼	[62]
hsa-miR-548h	2.12	3.76	4.06	6.72	-	NA
hsa-miR-548l	3.94	3.11	8.66	7.40	-	NA
hsa-miR-548p	3.88	2.44	4.64	1.55	-	NA
hsa-miR-548t	4.89	9.65	1.26	5.46	-	NA
hsa-miR-664	3.81	4.01	5.16	5.05	▲	[63]

NA: Not available. *denotes the star strand, which is currently designated as -3p or -5p. Thickness of arrows represents number of publications available corresponding to the expression pattern.

cellular development, endocrine system disease, hepatic system disease, tumor morphology and inflammatory response. Androgen withdrawal for 1wk to 3wks also showed changes in percentage of miRNA involvement (Additional file 11: Figure S4B). An increased percentage of down regulated miRNAs involved in cancer, cell death and survival, dermatological disease, endocrine system

disease, gastrointestinal disease, metabolic disease, and tumor morphology is noted in 3wks CSFBS treated cells. 3wks androgen withdrawal also showed a decrease in miRNA percentage involved in cell growth and proliferation, cell movement, cell-cell signaling, DNA replication and repair and renal-urological disease (Additional file 11: Figure S4B).

Table 3 Fold changes in expressions of down regulated miRNAs and its relevance with published reports

	1wk CSFBS Δ change	1wk CDX Δ change	3wks CSFBS Δ change	3wk CDX Δ change	Expression in other Cancer	References
hsa-miR-106a	-2.37	-3.18	-5.29	-9.40	▲▼	[64,65]
hsa-miR-1244	-3.40	-8.55	-5.14	-3.38	-	NA
hsa-miR-15b	-1.86	-2.02	-2.85	-3.33	▲▼	[66,67]
hsa-miR-15b*	-2.42	-4.65	-3.83	-9.76	▲▼	[68,69]
hsa-miR-17	-2.91	-3.37	-2.50	-24.99	▲▼	[70,71]
hsa-miR-17*	-3.15	-3.13	-5.26	-3.94	▲▼	[68,72]
hsa-miR-18a	-2.01	-4.36	-14.70	-14.13	-	[73]
hsa-miR-18b	-1.56	-2.28	-4.92	-21.36	▲▼	[74,75]
hsa-miR-205	-6.33	-12.51	-12.06	3.89	▲▼	[76,77]
hsa-miR-205*	-0.35	-1.34	-2.85	-3.23	-	NA
hsa-miR-20a	-2.73	-3.31	-3.78	-4.12	▲▼	[71,75,78]
hsa-miR-20a*	-1.78	-1.25	-2.64	-18.17	-	[69,75]
hsa-miR-20b*	-0.64	-0.56	-7.99	-4.60	-	[74,75,79]
hsa-miR-3131	-2.35	-2.97	-3.14	-3.41	-	[80]
hsa-miR-3185	-2.95	-3.24	-4.17	-5.76	-	[80]
hsa-miR-422a	-3.15	-3.15	-4.92	-6.87	-	[81]
hsa-miR-454	-0.58	-1.89	-3.28	-3.76	▼	[75,82]
hsa-miR-518b	-2.64	-6.42	-5.61	-12.23	-	[83]
hsa-miR-596	-2.45	-3.94	-15.83	-12.71	-	[84]
hsa-miR-759	-13.05	-10.92	-6.17	-5.86	▼	[82]
hsa-miR-7	-3.00	-5.23	-6.76	-11.08	-	[85]
hsa-miR-9	-2.64	-4.35	-4.53	-3.55	▼	[86]

NA: Not available, *denotes the star strand, which is currently designated as -3p or -5p. Thickness of arrows represents number of publications available corresponding to the expression pattern.

The *in silico* function analysis of the target miRNAs in different treatment conditions indicated a complex interaction of a network of miRNAs and their target proteins in these cells which rendered them adaptive to the androgen withdrawal and treatments with AR antagonists. Next, we analyzed the network of interactions among target miRNAs. We used the \log_2 transformed FC values of the subset of validated miRNA (Tables 2 and 3) for analysis of the functional interrelationship among

miRNAs (Figure 6). The network for up-regulated miRNAs and its putative targets in 3wks CDX showed that miR-3192 directly and miR-218 indirectly through DEAD box protein DDX20 target p53. MiR-146a also directly regulates a number of Toll-like receptors (TLR1, TLR9 and TLR10), cytokine receptors and its associated proteins (IL1R1, IL12RB2, IL1RAP) and chemokine receptors (CXCR4, CCR3) (Figure 6A). In addition, miR-146a directly targets a number of growth factors and

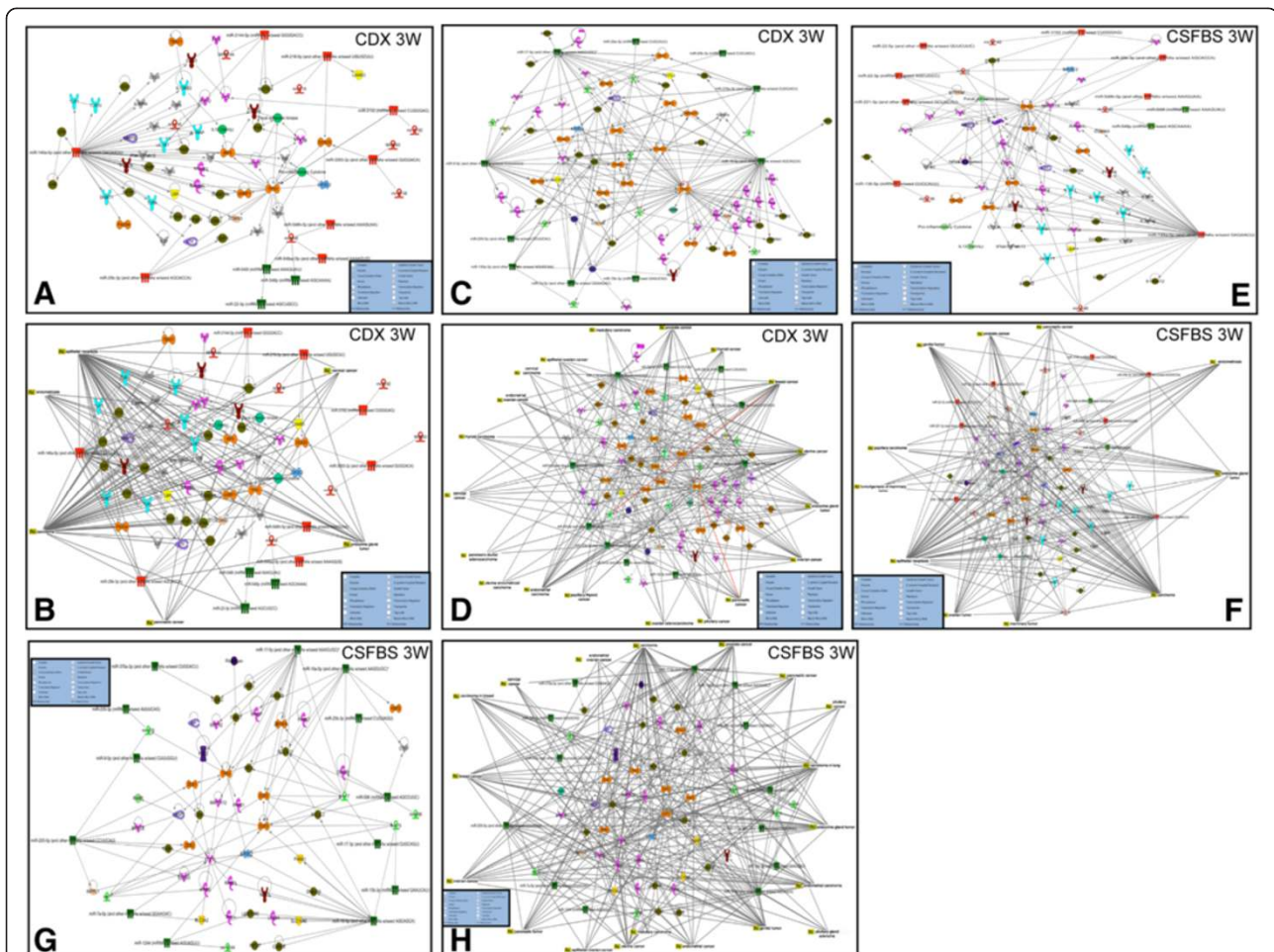


Figure 6 Analysis of miRNA-regulated network of interacting partners involved various signaling pathways. A) Log₂ transformed FC values of 21 up regulated miRNAs were imported into the Core analysis tool of IPA software and a network for interacting miRNAs and their direct target proteins and miRNAs that target any of the up regulated miRNAs has been generated for 3wks CDX treated cells. Red symbols denote miRNAs or mature miRNAs. Proteins include growth factor/cytokines, enzymes, kinases, phosphatases, peptidases, translational regulator, transcriptional regulators, and transporters. **B)** The network in A is overlaid with disease relationship showing association of miRNA and protein interactions with various types of cancer. **C and D)** Regulatory network of the log₂ transformed FC values of 22 down regulated miRNAs and their target proteins in 3wks CDX treated cells (**C**) and overlay of disease association of the miRNA-protein regulatory network (**D**). Green symbols represent down regulated miRNAs and mature miRNAs. **E and F)** Regulatory network of log₂ transformed FC values of the up regulated miRNAs and their target proteins in cells treated with CSFBS for 3wks (**E**) and an overlay of disease relationship with the regulatory network in E. Red symbols represent miRNAs and mature miRNAs. **G and H)** Regulatory network of log₂ transformed FC values of the down regulated miRNAs and their target proteins in cells treated with CSFBS for 3wks (**G**) and an overlay of disease association with the regulatory network in G (**H**). Green symbols represent down regulated miRNAs and mature miRNAs.

cytokines (Figure 6A). These miR-146a targets are also shown in the up-regulated network of 3wks CSFBS, in addition to BRCA, which is a direct target of miR-146a (Figure 6E). MiR-3192 targeting of p53 is also noted in 3wks CSFBS but not of miR-218 (Figure 6E). As noted in the networks, p53 also regulates a number of miRNAs, such as, miR-29b-3p, miR-22-3p, miR-22-5p and miR-221-3p (Figure 6A and E). When the networks are overlaid with diseases, a number of miRNAs in the up-regulated list of both 3wks CDX and 3wks CSFBS showed involvement in a variety of cancers including

pancreatic cancer, endocrine gland tumor, prostate cancer, ovarian tumor, mammary tumor, cervical cancer and epithelial neoplasia (Figure 6B and F).

In the list of down-regulated miRNAs there is direct targeting of E2F2, E2F3, MAP3K12, FXR1 and AR interacting nuclear protein kinase HIPK3 by miR-17-5p. MiR-16p also directly targets BCL2L2, cell death suppressor BNIP2, EGFR, RAB21 and single strand purine rich DNA binding protein PURA (Figure 6C and G) in both 3wks CDX and 3wks CSFBS treated cells. Also a number of miRNAs in the down regulated list down regulate a number of common

targets, such as E2F3 is targeted by miR-1244, miR-596, miR-18a-5p, miR-16-5p and miR-17-5p. EGFR is also targeted by miR-7a-5p and miR-16-5p. Overlay of diseases on networks revealed a distinct association of miR-17-5p, with various cancers including prostate cancer, breast cancer and pancreatic cancer and endocrine gland tumors. An association of miR-16-5p with prostate cancer, pancreatic cancer, pituitary cancer and endometrial cancer also could be noted (Figure 6D and H). These networks indicate a complex interplay of microRNAs and proteins during development of resistance of LNCaP cells to androgen withdrawal and treatment with CDX.

Target identification of the subset of miRNAs revealed potential activation and/or inactivation of a number of proteins involved in different signaling networks

A total of 21 up regulated and 22 down regulated miRNAs in all treatment conditions were used for identification of protein targets using IPA, miRDB and TargetScan software. Venn diagram of the putative targets of up-regulated miRNAs derived from the network generated using the IPA software showed 27 proteins that are common in all treatment conditions (Figure 7A). A number of which are known tumor suppressors such as BRCA1[87], TP53 [88], RAD54L [89], IRF5 [90], DUSP2 [91], IKK1 or CHUK [92]. Other targets of up regulated miRNAs in different treatment conditions also includes proteins that inhibit tumor progression, such as DNMT3A [93], FADD [94], pTEN [95], FOXO3[96], DDX20 [97] and PA2G4 (EBP1

[98]. There are also 41 common targets of the down-regulated miRNAs in all treatment conditions (Figure 7B). Some of the targets are known oncoproteins, which includes EGFR [99], VEGFA [100], ZBTB7 (POKEMON) [101], acid phosphatase 2(ACP2) [102] and NFKB1 [103]. Additionally, proteins that are overexpressed in cancer cells are among the targets of down-regulated miRNAs in different treatment conditions, which includes Wnt3A [104], PPARα [105], UCP2 [106], CSF1 [107], and MED1 [108].

Targets of miRNAs showed the predicted expression profiles in LNCaP cells subjected to androgen withdrawal and CDX treatment

Further identification of the targets of selected miRNA (Table 2) was conducted by miRDB and TargetScan database search and proteins that are regulated by one or multiple miRNAs from our list and received higher target scores in either or both searches are presented in Tables 4 and 5. The selected proteins that are regulated by the up-regulated miRNAs include TNF receptor associated factor 6 (TRAF6) [109], interleukin receptor associated kinase1 (IRAK1), BCL6 co-repressor-like 1(BCORL1) [110], Cbl [109], neuro oncological ventral antigen1 (NOVA1) [111], coiled coil domain containing 67 (CCDC67) [112] NET1 [113], ZFAND1(Acc.# NM_024699), RGS6 [114] CDKN1B (p27Kip1) [115], IGFBP5 [116] and RNASEL [117] (Table 4). The proteins that are potentially regulated by the down-regulated miRNAs include, ABHD3 [118], FGD4 [119], CCNJ [120], CHAMP1 [121], Myb [122],

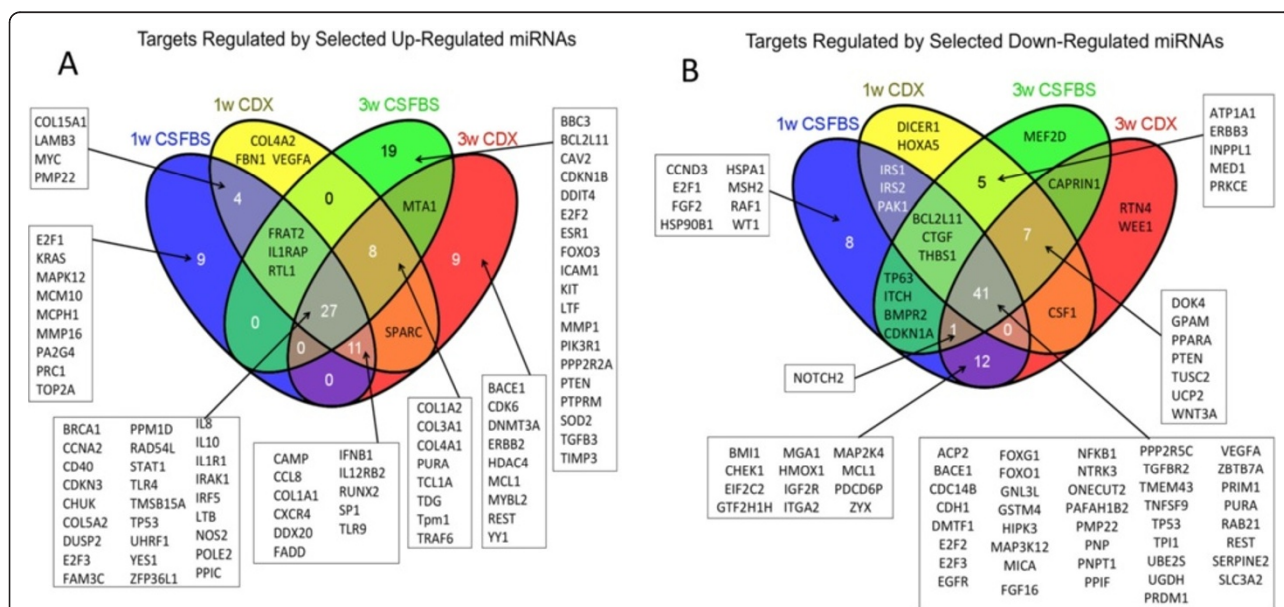


Figure 7 Analysis of treatment specific targets regulated by validated miRNAs. A) Venn diagram showing specific targets that are potentially regulated by miRNAs that are up regulated in different treatment conditions. Putative targets were identified based on the regulatory network generated by IPA software. **B)** Venn diagram depicting predicted targets regulated by the down-regulated miRNAs in different treatment conditions. Targets were identified based on the regulatory network using IPA software.

Table 4 Predicted mRNA Targets of Up-Regulated miRNAs

Gene ID	Gene description	miRNAs	miRDB (target score)	TargetScan (Context + Percentile)
TRAF6	TNF receptor-associated factor 6, E3 ubiquitin protein ligase	miR-146a	100	94,94,65,50,38,19
IRAK1	Interleukin receptor associated kinase 1	miR-146a	87	98
BCORL1	BCL6 corepressor-like 1	miR-146a, miR-143, miR-548t	77,68	98,80
NOVA-1	Neuro-oncological ventral antigen1	miR-146a, miR-143, miR-548l	97,93,86	97,89,87
CCDC67	Coiled-coil domain containing 67	miR-22	77	99,97
Cbl	Cbl proto-oncogene, E3 ubiquitin protein ligase	miR-22, miR-222,miR-136, miR-3199,miR-1197	87,83,64,55	86,84,75,61,43,33,32,31
NET1	Neuroepithelial cell transforming 1	miR-22, miR-222,miR-143	93,61	98
ZFAND1	Zinc finger, AN1-type domain 1	miR-548h, miR-548l,has-miR-548t,miR-136	96,70	93,83,99,52
RGS6	Regulator of G-protein signaling 6	miR-222	75	91,96,12
CDKN1B	Cyclin-dependent kinase inhibitor 1B (p27, Kip1)	miR-222	85	93,95
IGFBP5	Insulin-like growth factor binding protein 5	miR-143	N/A	99
RNASEL	Ribonuclease L	miR-146a, miR-548l	74	81

PIK3CD [85], VEGFA [123], SPOPL [124], RAB9B [125], EGFR [126], E2F1 [127] and DOK4 [128]. Western blot analysis confirmed the predicted expression profiles of some of the targets. Expression of Cbl, p27Kip1, TRAF6, IRAK1, and ZFAND1 were significantly and progressively down regulated in cells treated with CDX or subjected to androgen deprivation (Figure 8). Similarly, expression of FGD4, VEGFA, EGFR, DOK4 and ABHD3 were up regulated to moderate to high levels in cells treated with CDX or CSFBS (Figure 9).

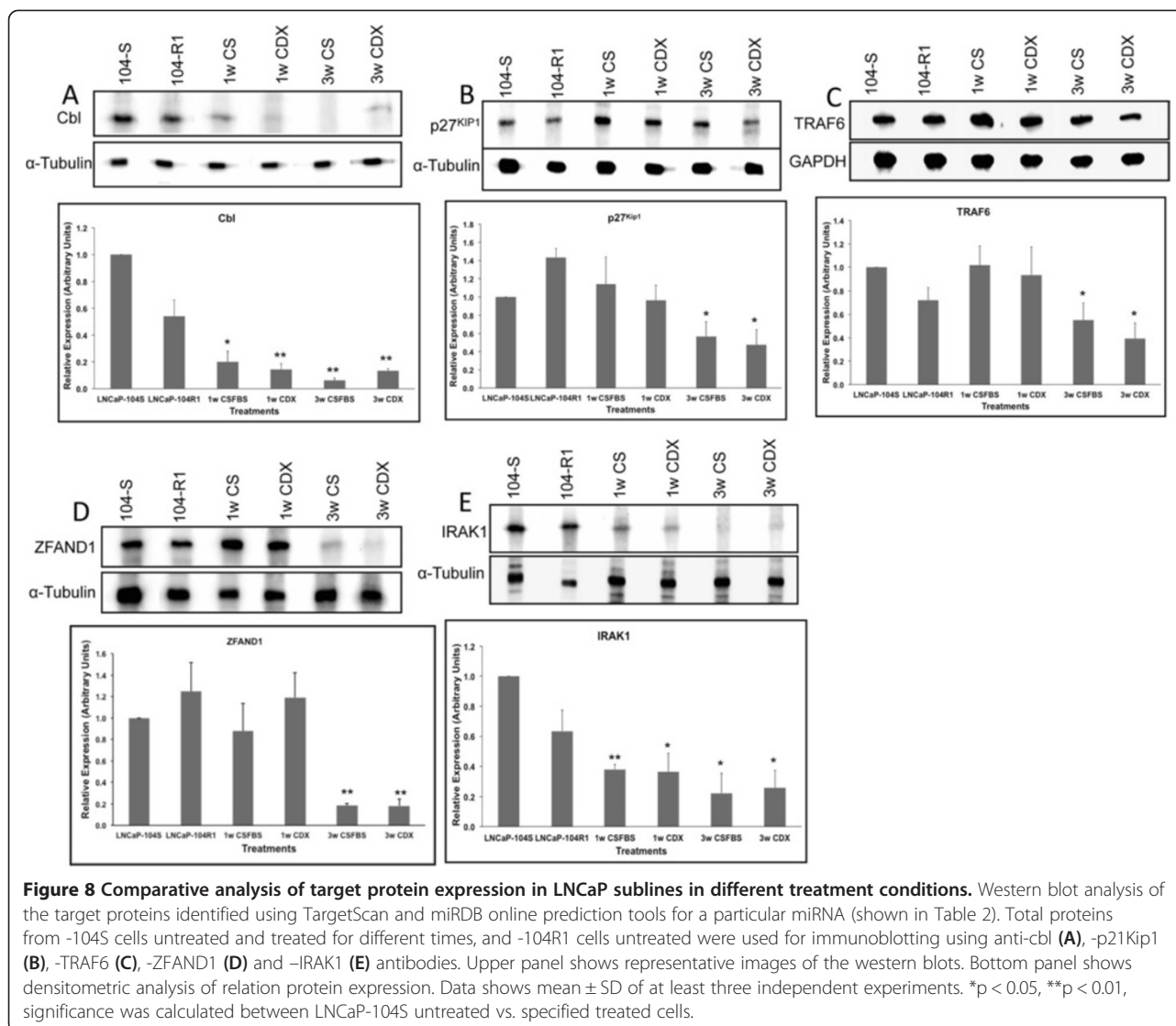
Discussion

Our studies on profiling and validation of miRNA expressions during transition of AD LNCaP-104S cells to AI and

CDX resistant cells revealed activation and inactivation of several signaling networks. We noted a difference in miRNA expressions between the AI subline LNCaP-104R1, and freshly generated CDX resistant LNCaP-104S cells, which includes some of the up regulated (miR-146a) and down-regulated miRNAs (miR-15b-3p and miR-18b). Although we noted differential expression of miRNAs between CS-FBS and CDX treated LNCaP cells we selected only miRNAs that showed either up regulation or down regulation in all treated samples for analysis of their putative targets. Despite similar expression profile of specific miRNAs in CS-FBS and CDX treated samples, some the targets such as DOK4 and VEGF showed differential expression in CSFBS and CDX treated cells. Presumably, this

Table 5 Predicted mRNA Targets of Down-Regulated miRNAs

GeneID	Gene Description	miRNAs	miRDB (target score)	TargetScan (Context + Percentile)
ABHD3	Abhydrolase domain containing 3	miR-1244, miR-130a, miR-205	74, > 85	84
CCNJ	Cyclin J	miR-205-5p	90	98
CHAMP1/ZNF828	Chromosome alignment maintaining phosphoprotein 1	miR-7-5p, miR-378a-3p	60-96	96
MYB	v-Myb myeloblastosis viral oncogene homolog (avian)	miR-15b-5p, miR-16-5p	74	96
PIK3CD	Phosphoinositide-3-kinase, catalytic, delta polypeptide	miR-7-5p	94	91
VEGFA	Vascular endothelial growth factor A	miR-205, miR-15b	79	97
FGD4	RhoGEF and PH domain containing 4	miR-17, miR-106a, miR-20a	100	98
SPOPL	Speckle-type POZ protein-like	miR-9-3p	97	97
RAB9B	RAB9B, member RAS oncogene family	miR-15b-5p, miR-16-5p, miR-130a-3P, miR-9-5p	67-86	67-86
EGFR	Epidermal growth factor receptor	miR-7	81	93
E2F1	E2F transcription factor 1	miR-205, miR-17, miR-20b	77	80
DOK4/IRS-5	Docking protein 4	miR-205	N/A	94

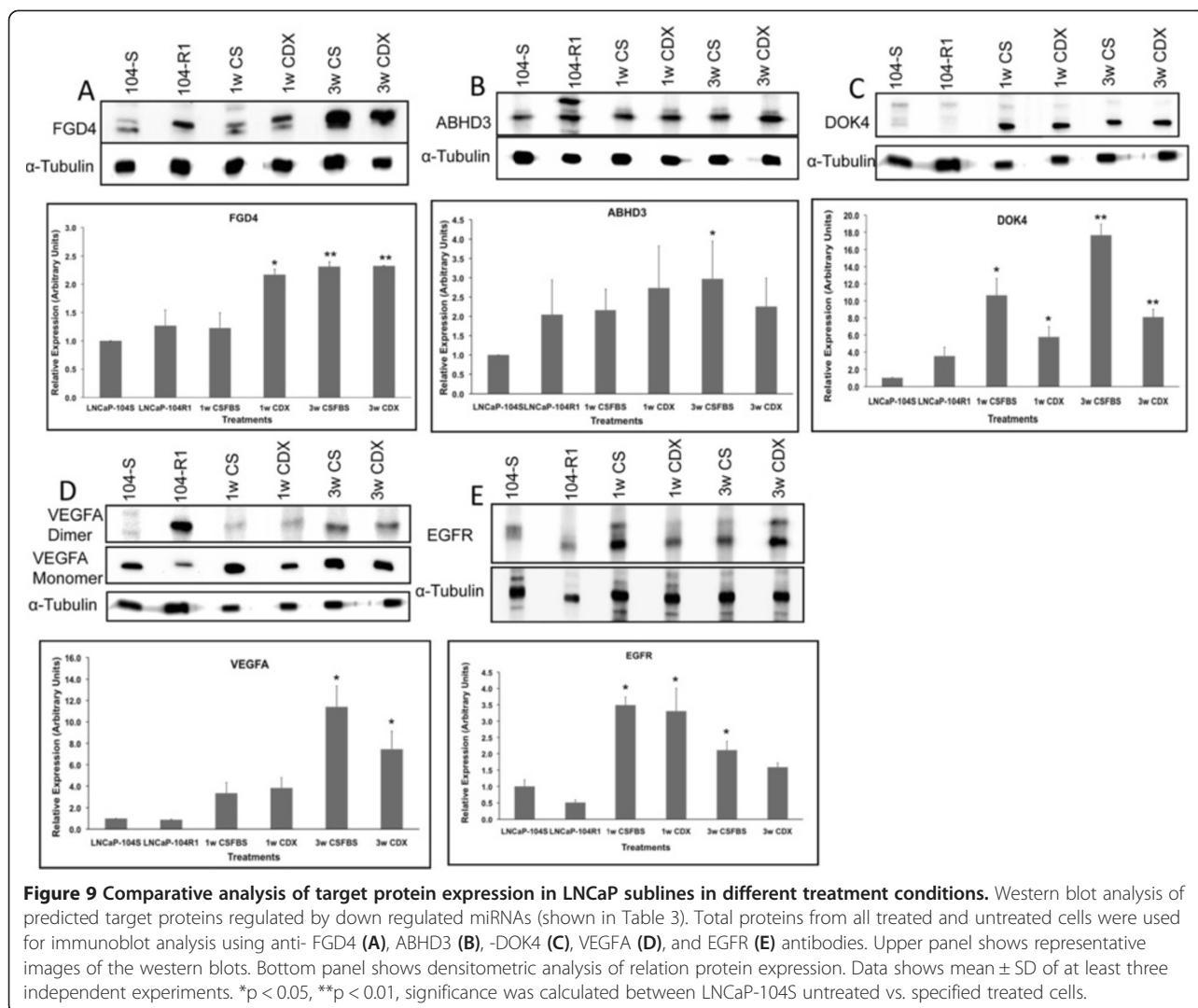


could be the effect of regulation of multiple targets by a given miRNA, which may indirectly affect the net expression of DOK4 and VEGF.

Over expression of miR-146a was noted in all treated cells contrary to the study showing loss of expression of miR-146a in CRPC [129]. Increased miR-146a expression was substantiated by down regulation of its two bona fide targets TRAF6 and IRAK1 in both -104R1 and 3wks treated -104S cells. MiR-146a expression is induced by NF- κ B [130] and acts in a negative feedback loop through degradation of TRAF6 and IRAK1 to reduce NF- κ B signaling and inflammatory response. An increase in transcription of miR-146a, as a result of elevated NF- κ B activity is noted in thyroid cancer [131] and down regulation of miR-146a is associated with hyperactivation of NF- κ B [132]. Despite this association, an increase in NF- κ B1 expression could be predicted in treated -104S cells, as NF- κ B1 is a direct target of the down-regulated miRNA

miR-9 [133]. Increased expression of the other subunit RelA, which heterodimerizes with NF- κ B1 also could be predicted as it is a direct target of the down-regulated miR-7 [134]. It appears that the NF- κ B signaling pathway is activated in the early stages of gaining resistance to CDX/ androgen blockade and the increased expression of miR-146a is a secondary effect of the activation of the NF κ B pathway. As a result, in the initial stages of anti-androgen drug resistance there is decreased inflammatory response but down regulation of tumor suppressor targets of miR-146a, BCORL1 [135] and RNASEL [136], which may not be detected in fully developed CRPC.

The EGFR signaling pathway could be activated also in the early stages of androgen blockade and CDX treatment. The evidence of EGFR pathway activation in the treated -104S cells is from our results showing an increased expression of EGFR, down regulation of miR-7 and up-regulation miR-222, which are the miRNA



regulators of EGFR. Decreased expression of p27Kip1 and Cbl, as two other targets of the up-regulated miR-222, further aid activation of EGFR signaling. C-Cbl, an E3 ubiquitin ligase, inactivates ligand-bound EGFR through EGFR-Cbl complex formation leading to its degradation [137]. C-Cbl activation mediates the tumor suppressive effects of EphB6 and inhibits cancer cell invasiveness [138]. C-Cbl is also targeted by the up regulated miRNA miR-136 in all treated cells. Down regulation of c-Cbl in treated -104S and untreated -104R1 cells possibly promotes antiandrogen resistance through EGFR stabilization. A loss of expression of AR was noted in these cells (data not shown), which supports the report showing an inverse relationship between expression of AR and EGFR in prostate cancer patients [139]. Up regulation of miR-136 in treated -104S cells was substantiated by the loss of expression of the miR-136 target ZFAND1, an uncharacterized AN1 type zinc finger domain 1 containing protein.

Activation of PI3K/AKT signaling axis also could be predicted in treated -104S cells, as a result of down regulation of miR-7, which inhibits tumor growth and metastasis through inhibition of PI3K/AKT pathways [85,140]. Down regulation of miR-7 in cancer cells including glioblastoma and its inhibitory effects on EMT and metastasis is well documented [141]. Activation of this pathway could be further aided by over expression of MiR-22 in all treated cells, which exerts a proto oncogenic effect through down regulation of PTEN in AI prostate cancer cells [142]. Additionally, activation of VEGF and DOK4 could be predicted, as these proteins are over expressed in treated -104S cells possibly as a result of down regulation of their regulatory miRNA, miR-205. Earlier studies showed an association between poor prognosis of localized prostate cancer and epigenetic repression of miR-205 [77], and thus confirms the relevance of the loss of miR-205 in development of CDX resistance. DOK4 is a newly identified substrate

of ligand-bound insulin receptor (IRS-5), which upon phosphorylation translocates to mitochondria and recruits c-Src kinase to the mitochondria. Up regulation of DOK4 is also noted in renal cell carcinoma [128]. VEGFA is a target of miR-15b-5p also, which showed 2-10-fold reduction in expression in treated -104S cells and in chemotherapy-resistant squamous cell carcinoma [67].

Other than modulation of specific signaling axis, altered expression of miRNA clusters is also evident in our study. Members of the miR-17-92 and its paralogous miR-106a-363 clusters, miR-17, miR-18a, miR-18b, miR-20a and miR-106a showed ~9-10-fold down regulation upon CDX treatment and androgen blockade. In support of our observation, loss of expression of miR-17 [71], miR-18a [73], miR-20a [78] and miR-106a [65] are reported in breast and other cancers. Loss of expression of miR-106a, miR-17 and miR-20a are further supported by an increased expression of their target protein FGD4 in these cells. Contrary to the published study [143], over expression of miR-34b was noted in AI and CDXR cells, however, an inverse relationship between miR-34b expression and disease free survival of triple negative breast cancer has been reported [61], which suggests that miR-34b expression may be dependent on the status of hormone responsiveness. Among the other up regulated miRNAs, over expression of let-7f1 [42], miR-143 [46], miR-218 [50], miR-29a [55], miR-302a [57], miR-3144 [59], miR-493 [62] and miR-664 [63] in cancer cells has been reported earlier. Our study also identified a number of miRNAs with > 2-fold difference in expression such as miR-3138, miR-3192, miR-3199, and a subset of miR-548 series, which are not yet known to be involved in development of CRPC.

Additional miRNAs such as miR-518b, miR-205 and miR-596 showed > 10-fold loss of expression upon CDX treatment or androgen withdrawal. In support of our observation, loss of expression and tumor suppressor functions of miR-518b and miR-596 has been documented in other cancers [83]. MiR-1244 and miR-759 are two other miRNAs that are significantly down regulated in all treated cells. This is substantiated by an increased expression of their common target ABHD3 in these cells [118]. Among the other down regulated miRNAs, miR-9 and miR-422a are known to have tumor suppressor roles in various cancer cells [81,86], whereas miRNAs -454, -3131 and -3185 are noted for the first time to be deregulated during progression of CRPC.

Functional contribution of some of the identified microRNAs in development of CRPC has been previously reported. Over expression of miR-222/221 in AI LAPC4 cells was shown to promote androgen independent cell growth, which was abrogated upon expression of anti-miR-222/221 inhibitors [21]. Down regulation of miR-205 has been correlated with advanced prostate cancer and ectopic expression of miR-205 suppressed AR and

MAPK signaling and inhibited cell growth [144]. Down regulation of miR-17 in AI prostate cancer cells also has been demonstrated. Expression of pre-miR-17 in these cells prevented AR induced gene transcription and inhibited cell proliferation [145].

Analysis of the altered cellular processes during progression towards CDX resistance and androgen independence showed a decreased percentage of miRNAs involved in cancer but an increased percentage in reproductive system, endocrine system, hepatic system and metabolic diseases. It can be speculated that up regulated oncomirs at earlier stages aid in transformation of cells through suppression of tumor suppressors. Whereas, at later stages accumulation of abnormal cellular events triggers expression of additional sets of miRNA, which inhibit key regulatory proteins involved in metabolic process, hormone response and other cellular events. Our qRT-PCR FC data indicate differential expression of miRNAs between 1wk and 3wks treatment, which would have been undetected had the profiling been done only in CDX sensitive/AD and -resistant/AI cells. In silico analysis identified a number of targets that are potentially regulated by one or more altered miRNAs. This includes, two mitosis regulatory proteins CCNJ and CHAMP1 (ZNF828) [120,121], two oncogenic proteins PIK3CD [85] and MYB that are over expressed in CRPC [122], a protein trafficking regulatory protein RAB9B, a ubiquitination promoting protein SPOPL involved in the Hedgehog/Gli signaling pathway [124] and E2F1 transcription factor [127].

In summary, our results and in silico network analysis suggest that inhibition of expression of TP53, BRCA, Toll like receptors, IRAK1, STAT1, CHUK and FADD by the up regulated miRNAs, and increased synthesis of EGFR, NFkB1/RelA, E2F family members, BCL2L2, ZBTB7A, EGO2 (EIF2C2) and ZEB2 as the targets of down regulated miRNAs are part of the events that support growth and survival of AI LNCaP cells. During treatment with AR antagonist in androgen-deprived condition, additional inhibition of expression of DDX20, URF1, IRF5 and CDKN3 as the targets of the up regulated miRNAs and increased expression of PRDM1, DOK4, TNFSF9 and NOTCH2 as a result of down regulated miRNAs may provide additional protection against CDX induced cell death. In-depth studies are needed to accurately determine the activation and inactivation of specific signaling pathways during development of insensitivities of prostate cancer cells to AR antagonistic drugs.

Conclusion

The overall evaluation of the changes in expression profiles of miRNAs during transition of CDX sensitive and AD cells to the CDX resistant and AI ones demonstrates that not any one or two miRNAs are responsible for development of drug-resistant prostate cancer. Instead, a complex

network of activation and inactivation of specific signaling pathways aided by degradation or accumulation of the target mRNAs as a result of differential expression of a significant number of miRNAs plays the pivotal role. Also, there are transient changes in the expression of miRNAs as well as their target proteins during transition of cells towards ADT resistance, which may provide growth and survival advantage to a subset of cancer cells; and these changes in miRNA/mRNA signature may be missed in already developed castration resistant prostate cancer. This study provides a predictive tool for monitoring the susceptibility of development of anti-androgen therapy resistant prostate cancer.

Materials and methods

Cell culture, and treatments

The androgen responsive LNCaP-104S and androgen independent LNCaP-104R1 cells were generous gifts from Dr. Shutsung Liao, University of Chicago. These LNCaP sublines were isolated and characterized as previously described [146]. LNCaP-104S cells were maintained in DMEM (Gibco) containing 10% FBS (Atlanta Biologicals), 1nM DHT (Sigma-Aldrich), 1% Antibiotic/Antimycotic (Invitrogen). LNCaP-104R1 cells were maintained in DMEM containing 10% charcoal-stripped FBS (CSFBS), 1% Antibiotic/Antimycotic. Isolation of androgen independent cells was conducted by passaging LNCaP-104S cells in DMEM/10% CSFBS, supplemented with or without 5 μ M Bicalutamide (Fluka) (Casodex, CDX) for 3 weeks (detail treatment method is in the Additional file 12 supplemental method section). Cells were harvested for protein or RNA extraction at 7 and 21 days post treatment.

RNA extraction and cDNA synthesis

Total RNA was extracted from untreated and treated cells using the Cell-to-Cts kit (System Biosciences) according to the manufacturer's instructions. Total RNA was converted to cDNA utilizing the QuantiMir RT Kit (System Biosciences) according to the manufacturer's instructions. Briefly, small RNAs were first tagged with polyA-tails; oligo-dT primers were annealed next, and converted to cDNA by reverse transcription.

Quantitative real-time PCR

Expression of mature miRNAs in untreated and treated LNCaP cells was determined by quantitative real-time PCR (qRT-PCR) using the miRNome microRNA Profiling Kit (System Biosciences) and cDNAs according to the manufacturer's instruction. The kit provides specific primers for 1,113 mature miRNAs and 3 internal control snRNAs. MiRNA IDs listed in the text are based on Sanger miRBase identifiers. Primers were designed to maintain uniform amplification efficiencies. qRT-PCR was conducted using the Applied Biosystems 7900HT thermal cycler and data

analyzed using and SDS2.3 software. DNA concentrations were reported through SYBR Green fluorescence and normalized to that of the passive reference dye, ROX.

Statistical analysis of miRNA expression

Ct values generated by the SDS2.3 software were normalized according to the average Ct values of the three internal controls provided with the miRNome Profiler kit using qbasePLUS software (Biogazelle). In order to ensure the integrity of the Δ Ct values, we utilized the Genorm software (Biogazelle) to identify 7 additional miRNAs displaying stable expressions between samples. The Ct values of all miRNAs were then normalized to these 10 controls. The relative expression values were then generated using qbasePLUS software and used in additional analysis (detail explanation is in the Additional file 12 supplemental method section). The Ct values were used to derive $\Delta\Delta$ Ct values using the miRNome analysis software (SBI). Candidate miRNAs with higher fold change values in each treatment conditions were determined by z score calculation as described in the Additional file 12 supplemental method section.

Clustering was performed using \log_2 transformed average $\Delta\Delta$ Ct values and the Cluster 3.0 software (Michiel de Hoon, Univ of Tokyo, based on Eisen Lab Cluster software). Hierarchical clustering of the miRNAs was based on the average linkage of the Pearson's correlation values. The relative expression values were \log_2 transformed and used for evaluation of differences among groups of treated and untreated cells by performing a Welch t-test using MultExperiment Viewer (MEV) software. Results of the t-test were displayed in Volcano plots using $-\log_{10}$ P values of the \log_2 transformed values. The K-median clustering of the normalized values were performed using MEV software.

Next, identification of target mRNAs, creation of miRNA-protein networks, and identification of altered cellular processes were conducted using the IPA software (Ingenuity Systems). The $\Delta\Delta$ Ct values were used for a core analysis by selecting all tissue and cell types and using a stringent filter and generating direct relationships only. From the core analysis, mRNA targets were identified for each sample and these mRNAs were used to generate the Venn diagrams using online tool (Venny: <http://bioinfogp.cnb.csic.es/tools/venny/index.html>). The miRNA-protein network generated by the core analysis was overlaid with different cancer types and members of the network that displayed alterations in those cancers were identified using the Interactive Pathway Analysis (IPA) software (Ingenuity Systems). All connections made in the networks are based on previously published results.

Western blotting

Cells harvested at different time points were lysed and total protein extracts were used for western blotting

using antibodies specific for AR (US Biological, Salem, MA), PSA (Santa Cruz Biotechnology, Dallas, TX), Cbl (C-15) (Santa Cruz Biotechnology, Dallas, TX), TRAF6 (Millipore, Temecula, CA), p27Kip1 (C-19) (Santa Cruz, Biotechnology), IRAK1 (F-4) (Santa Cruz, Biotechnology), ZFAND1 (A-14) (Santa Cruz Biotechnology), FGD4 (Epitomics, Burlingame, CA), ABHD3 (Biorbyt, Cambridge, UK), DOK4 (C-16) (Santa Cruz Biotechnology), EGFR (1005) (Santa Cruz Biotechnology), VEGFA (A-20) (Santa Cruz Biotechnology), α -tubulin (Cell Signaling, Danvers, MA), GAPDH (Sigma-Aldrich, St. Louis, MO). Total extracts (30–50 μ g) were directly mixed with Lammeli sample buffer and separated on SDS-PAGE. Immunoblotting was performed using appropriate primary and horseradish peroxidase conjugated respective secondary antibodies. Positive signals were detected using a chemiluminescence ECL kit (Pierce, Rockford, IL).

Additional files

Additional file 1: Figure S1. Light micrograph images of LNCaP cells before and during treatment with CS-FBS and CDX.

Additional file 2: Table S1. Listing normalized and log-transformed values of the miRNA expression.

Additional file 3: Figure S2. Hierarchical clustering of the data from genome wide miRNA profiling.

Additional file 4: Table S2. Listing the p-values, expression patterns and IDs of significant miRNAs identified in t-tests (supple Figure 2).

Additional file 5: Figure S3. Volcano plots of the two samples t-tests of the normalized values of untreated and treated LNCaP cells.

Additional file 6: Table S3. Listing the log-transformed fold change values of the up regulated and down regulated miRNAs identified in miRNA profiling.

Additional file 7: Table S4. Listing miRNAs in specific clusters identified in K-median cluster analysis.

Additional file 8: Table S5. Listing the p-values, expression profile and IDs of significant miRNAs from the list of validated miRNAs identified in two samples t-tests.

Additional file 9: Table S6. Listing the log-transformed values of the fold change in expression of the validated miRNAs.

Additional file 10: Table S7. Listing the up and down regulated subset of the validated miRNAs in specific clusters identified in K-median cluster analysis.

Additional file 11: Figure S4. Analysis of association of deregulated miRNAs with canonical pathways and cellular processes.

Additional file 12: Supplemental methods and figure legends.

Abbreviations

ADT: Androgen deprivation therapy; AR: Androgen receptor; MiRNA: MicroRNA; AD: androgen dependent; AI: Androgen independent; CRPC: castration resistant prostate cancer; AS: Androgen sensitive; -104S: LNCaP-104S cells; -104R1: LNCaP-104R1 cells; CDX: Casodex; CDXR: Casodex resistant; V plot: Volcano plot; CSFBS: Charcoal stripped FBS; FC: Fold change; PSA: Prostate specific antigen.

Competing interests

The author(s) declare that they have no competing interests.

Authors' contributions

RO performed cell treatments, profiling and all validation experiments including western blots. He also did the hierarchical clustering and in silico analysis of the targets. He participated in manuscript writing. CN analyzed data using IPA software and performed cell treatment related experiments. RL analyzed data including data normalization, z score calculation and Venn diagrams. RC conceived the idea, wrote the manuscript and performed data analysis. All authors read and approved the final manuscript.

Acknowledgements

This study is supported by a grant from the Department of Defense PCRIP (W81XWH-11-1-0563) (RC). We are thankful to Dr. Liao, University of Chicago for the generous gift of the LNCaP cell lines.

Received: 6 August 2013 Accepted: 11 November 2013

Published: 3 January 2014

References

1. Javidan J, Deitch AD, Shi XB, de Vere White RW: **The androgen receptor and mechanisms for androgen independence in prostate cancer.** *Cancer Invest* 2005, **23**:520–528.
2. Culig Z, Hoffmann J, Erdel M, Eder IE, Hobisch A, Hittmair A, Bartsch G, Utermann G, Schneider MR, Parczyk K, Klocker H: **Switch from antagonist to agonist of the androgen receptor bicalutamide is associated with prostate tumour progression in a new model system.** *Br J Cancer* 1999, **81**:242–251.
3. Seruga B, Ocana A, Tannock IF: **Drug resistance in metastatic castration-resistant prostate cancer.** *Nat Rev Clin Oncol* 2011, **8**:12–23.
4. Scher HI, Buchanan G, Gerald W, Butler LM, Tilley WD: **Targeting the androgen receptor: improving outcomes for castration-resistant prostate cancer.** *Endocr Relat Cancer* 2004, **11**:459–476.
5. Feldman BJ, Feldman D: **The development of androgen-independent prostate cancer.** *Nat Rev Cancer* 2001, **1**:34–45.
6. Gao M, Ossowski L, Ferrari AC: **Activation of Rb and decline in androgen receptor protein precede retinoic acid-induced apoptosis in androgen-dependent LNCaP cells and their androgen-independent derivative.** *J Cell Physiol* 1999, **179**:336–346.
7. Powell SM, Christiaens V, Voulgaraki D, Waxman J, Claessens F, Bevan CL: **Mechanisms of androgen receptor signalling via steroid receptor coactivator-1 in prostate.** *Endocr Relat Cancer* 2004, **11**:117–130.
8. Jenster G: **Ligand-independent activation of the androgen receptor in prostate cancer by growth factors and cytokines.** *J Pathol* 2000, **191**:227–228.
9. Berrevoets CA, Umar A, Brinkmann AO: **Antiandrogens: selective androgen receptor modulators.** *Mol Cell Endocrinol* 2002, **198**:97–103.
10. Chen CD, Welsbie DS, Tran C, Baek SH, Chen R, Vessella R, Rosenfeld MG, Sawyers CL: **Molecular determinants of resistance to antiandrogen therapy.** *Nat Med* 2004, **10**:33–39.
11. Reid AH, Attard G, Danila DC, Oommen NB, Olmos D, Fong PC, Molife LR, Hunt J, Messiou C, Parker C, et al: **Significant and sustained antitumor activity in post-docetaxel, castration-resistant prostate cancer with the CYP17 inhibitor abiraterone acetate.** *J Clin Oncol* 2010, **28**:1489–1495.
12. Lee YF, Lin WJ, Huang J, Messing EM, Chan FL, Wilding G, Chang C: **Activation of mitogen-activated protein kinase pathway by the antiandrogen hydroxyflutamide in androgen receptor-negative prostate cancer cells.** *Cancer Res* 2002, **62**:6039–6044.
13. Everley PA, Krijgsveld J, Zetter BR, Gygi SP: **Quantitative cancer proteomics: stable isotope labeling with amino acids in cell culture (SILAC) as a tool for prostate cancer research.** *Mol Cell Proteomics* 2004, **3**:729–735.
14. Bartel DP: **MicroRNAs: target recognition and regulatory functions.** *Cell* 2009, **136**:215–233.
15. Rodriguez A, Griffiths-Jones S, Ashurst JL, Bradley A: **Identification of mammalian microRNA host genes and transcription units.** *Genome Res* 2004, **14**:1902–1910.
16. Calin GA, Croce CM: **MicroRNA signatures in human cancers.** *Nat Rev Cancer* 2006, **6**:857–866.
17. Stefani G: **Roles of microRNAs and their targets in cancer.** *Expert Opin Biol Ther* 2007, **7**:1833–1840.
18. Filipowicz W, Bhattacharyya SN, Sonenberg N: **Mechanisms of post-transcriptional regulation by microRNAs: are the answers in sight?** *Nat Rev Genet* 2008, **9**:102–114.

19. Lim LP, Lau NC, Garrett-Engle P, Grimson A, Schelter JM, Castle J, Bartel DP, Linsley PS, Johnson JM: **Microarray analysis shows that some microRNAs downregulate large numbers of target mRNAs.** *Nature* 2005, **433**:769–773.
20. Ribas J, Ni X, Haffner M, Wentzel EA, Salmasi AH, Chowdhury WH, Kudrolli TA, Yegnasubramanian S, Luo J, Rodriguez R, et al: **miR-21: an androgen receptor-regulated microRNA that promotes hormone-dependent and hormone-independent prostate cancer growth.** *Cancer Res* 2009, **69**:7165–7169.
21. Sun T, Wang Q, Balk S, Brown M, Lee GS, Kantoff P: **The role of microRNA-221 and microRNA-222 in androgen-independent prostate cancer cell lines.** *Cancer Res* 2009, **69**:3356–3363.
22. Shi XB, Xue L, Yang J, Ma AH, Zhao J, Xu M, Tepper CG, Evans CP, Kung HJ, deVere White RW: **An androgen-regulated miRNA suppresses Bak1 expression and induces androgen-independent growth of prostate cancer cells.** *Proc Natl Acad Sci U S A* 2007, **104**:19983–19988.
23. Jalava SE, Urbanucci A, Latonen L, Walthering KK, Sahu B, Janne OA, Seppala J, Lahdesmaki H, Tammela TL, Visakorpi T: **Androgen-regulated miR-32 targets BTG2 and is overexpressed in castration-resistant prostate cancer.** *Oncogene* 2012, **31**:4460–4471.
24. Kokontis JM, Hsu S, Chuu CP, Dang M, Fukuchi J, Hiipakka RA, Liao S: **Role of androgen receptor in the progression of human prostate tumor cells to androgen independence and insensitivity.** *Prostate* 2005, **65**:287–298.
25. Wang Z, Liu M, Zhu H, Zhang W, He S, Hu C, Quan L, Bai J, Xu N: **miR-106a is frequently upregulated in gastric cancer and inhibits the extrinsic apoptotic pathway by targeting FAS.** *Molecular carcinogenesis* 2012, **52**:634–646.
26. Satzger I, Mattern A, Kuettler U, Weinspach D, Voelker B, Kapp A, Gutzmer R: **MicroRNA-15b represents an independent prognostic parameter and is correlated with tumor cell proliferation and apoptosis in malignant melanoma.** *Int J Cancer* 2010, **126**:2553–2562.
27. Chen L, Jiang M, Yuan W, Tang H: **miR-17-5p as a novel prognostic marker for hepatocellular carcinoma.** *J Invest Surg: Off J Acad Surg Res* 2012, **25**:156–161.
28. Wu CW, Dong YJ, Liang QY, He XQ, Ng SS, Chan FK, Sung JJ, Yu J: **MicroRNA-18a attenuates DNA damage repair through suppressing the expression of ataxia telangiectasia mutated in colorectal cancer.** *PLoS ONE* 2013, **8**:e57036.
29. Xie H, Zhao Y, Caramuta S, Larsson C, Lui WO: **miR-205 expression promotes cell proliferation and migration of human cervical cancer cells.** *PLoS ONE* 2012, **7**:e46990.
30. Kang HW, Wang F, Wei Q, Zhao YF, Liu M, Li X, Tang H: **miR-20a promotes migration and invasion by regulating TNKS2 in human cervical cancer cells.** *FEBS Lett* 2012, **586**:897–904.
31. Chou YT, Lin HH, Lien YC, Wang YH, Hong CF, Kao YR, Lin SC, Chang YC, Lin SY, Chen SJ, et al: **EGFR promotes lung tumorigenesis by activating miR-7 through a Ras/ERK/Myc pathway that targets the Ets2 transcriptional repressor ERF.** *Cancer Res* 2010, **70**:8822–8831.
32. Shibahara Y, Miki Y, Onodera Y, Hata S, Chan MS, Yiu CC, Loo TY, Nakamura Y, Akahira J, Ishida T, et al: **Aromatase inhibitor treatment of breast cancer cells increases the expression of let-7f, a microRNA targeting CYP19A1.** *J Pathol* 2012, **227**:357–366.
33. Yang Y, Wu J, Guan H, Cai J, Fang L, Li J, Li M: **MiR-136 promotes apoptosis of glioma cells by targeting AEG-1 and Bcl-2.** *FEBS Lett* 2012, **586**:3608–3612.
34. Lin T, Dong W, Huang J, Pan Q, Fan X, Zhang C, Huang L: **MicroRNA-143 as a tumor suppressor for bladder cancer.** *J Urol* 2009, **181**:1372–1380.
35. Li Y, Vandenboom TG 2nd, Wang Z, Kong D, Ali S, Philip PA, Sarkar FH: **miR-146a suppresses invasion of pancreatic cancer cells.** *Cancer Res* 2010, **70**:1486–1495.
36. Venkataraman S, Birks DK, Balakrishnan I, Alimova I, Harris PS, Patel PR, Handler MH, Dubuc A, Taylor MD, Foreman NK, Vibhakar R: **MicroRNA 218 acts as a tumor suppressor by targeting multiple cancer phenotype-associated genes in medulloblastoma.** *J Biol Chem* 2013, **288**:1918–1928.
37. Li B, Song Y, Liu TJ, Cui YB, Jiang Y, Xie ZS, Xie SL: **miRNA-22 suppresses colon cancer cell migration and invasion by inhibiting the expression of T-cell lymphoma invasion and metastasis 1 and matrix metalloproteinases 2 and 9.** *Oncol Rep* 2013, **29**:1932–1938.
38. Fuse M, Kojima S, Enokida H, Chiyomaru T, Yoshino H, Nohata N, Kinoshita T, Sakamoto S, Naya Y, Nakagawa M, et al: **Tumor suppressive microRNAs (miR-222 and miR-31) regulate molecular pathways based on microRNA expression signature in prostate cancer.** *J Human Genet* 2012, **57**:691–699.
39. Wang F, Wang XS, Yang GH, Zhai PF, Xiao Z, Xia LY, Chen LR, Wang Y, Wang XZ, Bi LX, et al: **miR-29a and miR-142-3p downregulation and diagnostic implication in human acute myeloid leukemia.** *Mol Biol Rep* 2012, **39**:2713–2722.
40. Majid S, Dar AA, Saini S, Shahryari V, Arora S, Zaman MS, Chang I, Yamamura S, Tanaka Y, Chiyomaru T, et al: **miRNA-34b inhibits prostate cancer through demethylation, active chromatin modifications, and AKT pathways.** *Clin Cancer Res: Off J Am Assoc Cancer Res* 2013, **19**:73–84.
41. Ueno K, Hirata H, Majid S, Yamamura S, Shahryari V, Tabatabai ZL, Hinoda Y, Dahiya R: **Tumor suppressor microRNA-493 decreases cell motility and migration ability in human bladder cancer cells by downregulating RhoC and FZD4.** *Mol Cancer Ther* 2012, **11**:244–253.
42. Gottardo F, Liu CG, Ferracin M, Calin GA, Fassan M, Bassi P, Sevignani C, Byrne D, Negrini M, Pagano F, et al: **Micro-RNA profiling in kidney and bladder cancers.** *Urol Oncol* 2007, **25**:387–392.
43. Vaksman O, Stavnes HT, Kaern J, Trope CG, Davidson B, Reich R: **miRNA profiling along tumour progression in ovarian carcinoma.** *J Cell Mol Med* 2011, **15**:1593–1602.
44. Liu X, Sempere LF, Ouyang H, Memoli VA, Andrew AS, Luo Y, Demidenko E, Korc M, Shi W, Preis M, et al: **MicroRNA-31 functions as an oncogenic microRNA in mouse and human lung cancer cells by repressing specific tumor suppressors.** *J Clin Invest* 2010, **120**:1298–1309.
45. McIver SC, Stanger SJ, Santarelli DM, Roman SD, Nixon B, McLaughlin EA: **A unique combination of male germ cell miRNAs coordinates gonocyte differentiation.** *PLoS ONE* 2012, **7**:e35553.
46. Fan X, Chen X, Deng W, Zhong G, Cai Q, Lin T: **Up-regulated microRNA-143 in cancer stem cells differentiation promotes prostate cancer cells metastasis by modulating FNDC3B expression.** *BMC Cancer* 2013, **13**:61.
47. Peng X, Guo W, Liu T, Wang X, Tu X, Xiong D, Chen S, Lai Y, Du H, Chen G, et al: **Identification of miR-143 and -145 that is associated with bone metastasis of prostate cancer and involved in the regulation of EMT.** *PLoS ONE* 2011, **6**:e20341.
48. Xiao B, Zhu ED, Li N, Lu DS, Li W, Li BS, Zhao YL, Mao XH, Guo G, Yu PW, Zou QM: **Increased miR-146a in gastric cancer directly targets SMAD4 and is involved in modulating cell proliferation and apoptosis.** *Oncol Rep* 2012, **27**:559–566.
49. Wang S, Wu HT, Fischer A, Miller DH, McLaughlin S, Douglass E, Steinhoff M, Collins C, Smith PJ, Brard L, Brodsky AS: **Identification of Ovarian Cancer Metastatic miRNAs.** *PLoS ONE* 2013, **8**:e58226.
50. Leite KR, Sousa-Canavez JM, Reis ST, Tomiyama AH, Camara-Lopes LH, Sanudo A, Antunes AA, Srougi M: **Change in expression of miR-let7c, miR-100, and miR-218 from high grade localized prostate cancer to metastasis.** *Urol Oncol* 2011, **29**:265–269.
51. Hassan MQ, Maeda Y, Taipaleenmaki H, Zhang W, Jafferji M, Gordon JA, Li Z, Croce CM, van Wijnen AJ, Stein JL, et al: **miR-218 directs a Wnt signaling circuit to promote differentiation of osteoblasts and osteomimicry of metastatic cancer cells.** *J Biol Chem* 2012, **287**:42084–42092.
52. Jiang R, Deng L, Zhao L, Li X, Zhang F, Xia Y, Gao Y, Wang X, Sun B: **miR-22 promotes HBV-related hepatocellular carcinoma development in males.** *Clin Cancer Res: Off J Am Assoc Cancer Res* 2011, **17**:5593–5603.
53. Wang Z, Zhang H, He L, Dong W, Li J, Shan Z, Teng W: **Association between the expression of four upregulated miRNAs and extrathyroidal invasion in papillary thyroid carcinoma.** *Oncotargets Ther* 2013, **6**:281–287.
54. Visone R, Russo L, Pallante P, De Martino I, Ferraro A, Leone V, Borbone E, Petrocca F, Alder H, Croce CM, Fusco A: **MicroRNAs (miR)-221 and miR-222, both overexpressed in human thyroid papillary carcinomas, regulate p27Kip1 protein levels and cell cycle.** *Endocr Relat Cancer* 2007, **14**:791–798.
55. Han YC, Park CY, Bhagat G, Zhang J, Wang Y, Fan JB, Liu M, Zou Y, Weissman IL, Gu H: **microRNA-29a induces aberrant self-renewal capacity in hematopoietic progenitors, biased myeloid development, and acute myeloid leukemia.** *J Exp Med* 2010, **207**:475–489.
56. Wu Q, Lu Z, Li H, Lu J, Guo L, Ge Q: **Next-generation sequencing of microRNAs for breast cancer detection.** *J Biomed Biotech* 2011, **2011**:597145.
57. Watson JA, Bryan K, Williams R, Popov S, Vujanic G, Coulomb A, Boccon-Gibod L, Graf N, Pritchard-Jones K, O'Sullivan M: **miRNA profiles as a predictor of chemoresponsiveness in Wilms' tumor blastema.** *PLoS ONE* 2013, **8**:e53417.
58. Yao Y, Suo AL, Li ZF, Liu LY, Tian T, Ni L, Zhang WG, Nan KJ, Song TS, Huang C: **MicroRNA profiling of human gastric cancer.** *Mol Med Rep* 2009, **2**:963–970.

59. Hamfjord J, Stangeland AM, Hughes T, Skrede ML, Tveit KM, Ikdahl T, Kure EH: **Differential expression of miRNAs in colorectal cancer: comparison of paired tumor tissue and adjacent normal mucosa using high-throughput sequencing.** *PLoS ONE* 2012, **7**:e34150.
60. Dutta KK, Zhong Y, Liu YT, Yamada T, Akatsuka S, Hu Q, Yoshihara M, Ohara H, Takehashi M, Shinohara T, et al: **Association of microRNA-34a overexpression with proliferation is cell type-dependent.** *Cancer Sci* 2007, **98**:1845–1852.
61. Svoboda M, Sana J, Redova M, Navratil J, Palacova M, Fabian P, Slaby O, Vyzula R: **Mir-34b is associated with clinical outcome in triple-negative breast cancer patients.** *Diagn Pathol* 2012, **7**:31.62.
62. Lehmann U, Streichert T, Otto B, Albat C, Hasemeier B, Christgen H, Schipper E, Hille U, Kreipe HH, Langer F: **Identification of differentially expressed microRNAs in human male breast cancer.** *BMC Cancer* 2010, **10**:109.
63. Yang H, Cho ME, Li TW, Peng H, Ko KS, Mato JM, Lu SC: **MicroRNAs regulate methionine adenosyltransferase 1A expression in hepatocellular carcinoma.** *J Clin Invest* 2013, **123**:285–298.
64. Yang G, Zhang R, Chen X, Mu Y, Ai J, Shi C, Liu Y, Sun L, Rainov NG, Li H, et al: **Mir-106a inhibits glioma cell growth by targeting E2F1 independent of p53 status.** *J Mol Med* 2011, **89**:1037–1050.
65. Hummel R, Hussey DJ, Michael MZ, Haier J, Bruewer M, Senninger N, Watson DI: **MiRNAs and their association with locoregional staging and survival following surgery for esophageal carcinoma.** *Ann Surg Oncol* 2011, **18**:253–260.
66. Zheng X, Chopp M, Lu Y, Buller B, Jiang F: **Mir-15b and miR-152 reduce glioma cell invasion and angiogenesis via NRP-2 and MMP-3.** *Cancer Lett* 2013, **329**:146–154.
67. Sun L, Yao Y, Liu B, Lin Z, Lin L, Yang M, Zhang W, Chen W, Pan C, Liu Q, et al: **Mir-200b and miR-15b regulate chemotherapy-induced epithelial-mesenchymal transition in human tongue cancer cells by targeting BMI1.** *Oncogene* 2012, **31**:432–445.
68. Lui WO, Pourmand N, Patterson BK, Fire A: **Patterns of known and novel small RNAs in human cervical cancer.** *Cancer Res* 2007, **67**:6031–6043.
69. Michael MZ, SMOG, van Holst Pellekaan NG, Young GP, James RJ: **Reduced accumulation of specific microRNAs in colorectal neoplasia.** *Mol Cancer Res* 2003, **1**:882–891.
70. Wei Q, Li YX, Liu M, Li X, Tang H: **Mir-17-5p targets TP53INP1 and regulates cell proliferation and apoptosis of cervical cancer cells.** *IUBMB Life* 2012, **64**:697–704.
71. Hossain A, Kuo MT, Saunders GF: **Mir-17-5p regulates breast cancer cell proliferation by inhibiting translation of AIB1 mRNA.** *Mol Cell Biol* 2006, **26**:8191–8201.
72. Suh MR, Lee Y, Kim JY, Kim SK, Moon SH, Lee JY, Cha KY, Chung HM, Yoon HS, Moon SY, et al: **Human embryonic stem cells express a unique set of microRNAs.** *Dev Biol* 2004, **270**:488–498.
73. Tao J, Wu D, Li P, Xu B, Lu Q, Zhang W: **microRNA-18a, a member of the oncogenic miR-17-92 cluster, targets Dicer and suppresses cell proliferation in bladder cancer T24 cells.** *Mol Med Rep* 2012, **5**:167–172.
74. O'Donnell KA, Wentzel EA, Zeller KJ, Dang CV, Mendell JT: **c-Myc-regulated microRNAs modulate E2F1 expression.** *Nature* 2005, **435**:839–843.
75. Landgraf P, Rusu M, Sheridan R, Sewer A, Iovino N, Aravin A, Pfeffer S, Rice A, Kamphorst AO, Landthaler M, et al: **A mammalian microRNA expression atlas based on small RNA library sequencing.** *Cell* 2007, **129**:1401–1414.
76. Wu H, Zhu S, Mo YY: **Suppression of cell growth and invasion by miR-205 in breast cancer.** *Cell Res* 2009, **19**:439–448.
77. Hulf T, Sibbritt T, Wiklund ED, Patterson K, Song JZ, Stirzaker C, Qu W, Nair S, Horvath LG, Armstrong NJ, et al: **Epigenetic-induced repression of microRNA-205 is associated with MED1 activation and a poorer prognosis in localized prostate cancer.** *Oncogene* 2012, **32**:2892–2899.
78. Chang CC, Yang YJ, Li YJ, Chen ST, Lin BR, Wu TS, Lin SK, Kuo MY, Tan CT: **MicroRNA-17/20a functions to inhibit cell migration and can be used a prognostic marker in oral squamous cell carcinoma.** *Oral oncology* 2013, **49**:923–931.
79. Sewer A, Paul N, Landgraf P, Aravin A, Pfeffer S, Brownstein MJ, Tuschl T, van Nimwegen E, Zavolan M: **Identification of clustered microRNAs using an ab initio prediction method.** *BMC Bioinformatics* 2005, **6**:267.
80. Stark MS, Tyagi S, Nancarrow DJ, Boyle GM, Cook AL, Whiteman DC, Parsons PG, Schmidt C, Sturm RA, Hayward NK: **Characterization of the Melanoma miRNAome by Deep Sequencing.** *PLoS ONE* 2010, **5**:e9685.
81. Faltejskova P, Svoboda M, Srutova K, Milcochova J, Besse A, Nekvindova J, Radova L, Fabian P, Slaba K, Kiss I, et al: **Identification and functional screening of microRNAs highly deregulated in colorectal cancer.** *J Cell Mol Med* 2012, **16**:2655–2666.
82. Berezikov E, van Tetering G, Verheul M, van de Belt J, van Laake L, Vos J, Verloop R, van de Wetering M, Guryev V, Takada S, et al: **Many novel mammalian microRNA candidates identified by extensive cloning and RAKE analysis.** *Genome Res* 2006, **16**:1289–1298.
83. Zhang M, Zhou S, Zhang L, Zhang J, Cai H, Zhu J, Huang C, Wang J: **miR-518b is down-regulated, and involved in cell proliferation and invasion by targeting Rap1b in esophageal squamous cell carcinoma.** *FEBS Lett* 2012, **586**:3508–3521.
84. Cummins JM, He Y, Leary RJ, Pagliarini R, Diaz LA Jr, Sjoblom T, Barad O, Bentwich Z, Szafrańska AE, Labourier E, et al: **The colorectal microRNAome.** *Proc Natl Acad Sci U S A* 2006, **103**:3687–3692.
85. Fang Y, Xue JL, Shen Q, Chen J, Tian L: **MicroRNA-7 inhibits tumor growth and metastasis by targeting the phosphoinositide 3-kinase/Akt pathway in hepatocellular carcinoma.** *Hepatology* 2012, **55**:1852–1862.
86. Guo LM, Pu Y, Han Z, Liu T, Li YX, Liu M, Li X, Tang H: **MicroRNA-9 inhibits ovarian cancer cell growth through regulation of NF-kappaB1.** *FEBS J* 2009, **276**:5537–5546.
87. Gallagher DJ, Gaudet MM, Pal P, Kirchoff T, Balistreri L, Vora K, Bhatia J, Stadler Z, Fine SW, Reuter V, et al: **Germline BRCA mutations denote a clinicopathologic subset of prostate cancer.** *Clin Cancer Res* 2010, **16**:2115–2121.
88. Martin P, Liu YN, Pierce R, Abou-Kheir W, Casey O, Seng V, Camacho D, Simpson RM, Kelly K: **Prostate epithelial Pten/TP53 loss leads to transformation of multipotential progenitors and epithelial to mesenchymal transition.** *Am J Pathol* 2011, **179**:422–435.
89. Matsuda M, Miyagawa K, Takahashi M, Fukuda T, Kataoka T, Asahara T, Inui H, Watatani M, Yasutomi M, Kamada N, et al: **Mutations in the RAD54 recombination gene in primary cancers.** *Oncogene* 1999, **18**:3427–3430.
90. Bi X, Hameed M, Mirani N, Pimenta EM, Anari J, Barnes BJ: **Loss of interferon regulatory factor 5 (IRF5) expression in human ductal carcinoma correlates with disease stage and contributes to metastasis.** *Breast Cancer Res* 2011, **13**:R111.
91. Lin SC, Chien CW, Lee JC, Yeh YC, Hsu KF, Lai YY, Lin SC, Tsai SJ: **Suppression of dual-specificity phosphatase-2 by hypoxia increases chemoresistance and malignancy in human cancer cells.** *J Clin Invest* 2011, **121**:1905–1916.
92. Liu B, Xia X, Zhu F, Park E, Carbajal S, Kiguchi K, DiGiovanni J, Fischer SM, Hu Y: **IKKalpha is required to maintain skin homeostasis and prevent skin cancer.** *Cancer Cell* 2008, **14**:212–225.
93. Gao Q, Steine EJ, Barrasa MI, Hockemeyer D, Pawlak M, Fu D, Reddy S, Bell GW, Jaenisch R: **Deletion of the de novo DNA methyltransferase Dnmt3a promotes lung tumor progression.** *Proc Natl Acad Sci U S A* 2011, **108**:18061–18066.
94. Jang MS, Lee SJ, Kim CJ, Lee CW, Kim E: **Phosphorylation by polo-like kinase 1 induces the tumor-suppressing activity of FADD.** *Oncogene* 2011, **30**:471–481.
95. Choucair K, Ejdelman J, Brimo F, Aprikian A, Chevalier S, Lapointe J: **PTEN genomic deletion predicts prostate cancer recurrence and is associated with low AR expression and transcriptional activity.** *BMC Cancer* 2012, **12**:543.
96. Qi W, Weber CR, Wasland K, Roy H, Wali R, Joshi S, Savkovic SD: **Tumor suppressor FOXO3 mediates signals from the EGF receptor to regulate proliferation of colonic cells.** *Am J Physiol Gastrointest Liver Physiol* 2011, **300**:G264–G272.
97. Takata A, Otsuka M, Yoshikawa T, Kishikawa T, Kudo Y, Goto T, Yoshida H, Koike K: **A miRNA machinery component DDX20 controls NF-kappaB via microRNA-140 function.** *Biochem Biophys Res Commun* 2012, **420**:564–569.
98. Zhang Y, Linn D, Liu Z, Melamed J, Tavora F, Young CY, Burger AM, Hamburger AW: **EBP1, an ErbB3-binding protein, is decreased in prostate cancer and implicated in hormone resistance.** *Mol Cancer Ther* 2008, **7**:3176–3186.
99. Hakariya T, Shida Y, Sakai H, Kanetake H, Igawa T: **EGFR signaling pathway negatively regulates PSA expression and secretion via the PI3K-Akt pathway in LNCaP prostate cancer cells.** *Biochem Biophys Res Commun* 2006, **342**:92–100.
100. Cao YEG, Wang E, Pal K, Dutta SK, Bar-Sagi D, Mukhopadhyay D: **VEGF exerts an angiogenesis-independent function in cancer cells to promote their malignant progression.** *Cancer Res* 2012, **72**:3912–3918.
101. Aggarwal H, Aggarwal A, Agrawal DK: **Epidermal growth factor increases LRF/Pokemon expression in human prostate cancer cells.** *Exp Mol Pathol* 2011, **91**:496–501.

102. Bhatt DK, Nagda G: **Modulation of acid phosphatase and lactic dehydrogenase in hexachlorocyclohexane-induced hepatocarcinogenesis in mice.** *J Biochem Mol Toxicol* 2012, **26**:439–444.
103. Domingo-Domenech J, Mellado B, Ferrer B, Truan D, Codony-Servat J, Sauleda S, Alcover J, Campo E, Gascon P, Rovira A, et al: **Activation of nuclear factor-kappaB in human prostate carcinogenesis and association to biochemical relapse.** *Br J Cancer* 2005, **93**:1285–1294.
104. Bonci D, Coppola V, Musumeci M, Addario A, Giuffrida R, Memeo L, D'Urso L, Pagliuca A, Biffoni M, Labbaye C, et al: **The miR-15a-miR-16-1 cluster controls prostate cancer by targeting multiple oncogenic activities.** *Nat Med* 2008, **14**:1271–1277.
105. Berger E, Vega N, Vidal H, Geloan A: **Gene network analysis leads to functional validation of pathways linked to cancer cell growth and survival.** *Biotechnol J* 2012, **7**:1395–1404.
106. Derdak Z, Mark NM, Beldi G, Robson SC, Wands JR, Baffy G: **The mitochondrial uncoupling protein-2 promotes chemoresistance in cancer cells.** *Cancer Res* 2008, **68**:2813–2819.
107. Liu P, Ramachandran S, Ali Seyed M, Scharer CD, Laycock N, Dalton WB, Williams H, Karanam S, Datta MW, Jaye DL, Moreno CS: **Sex-determining region Y box 4 is a transforming oncogene in human prostate cancer cells.** *Cancer Res* 2006, **66**:4011–4019.
108. Jin F, Irshad S, Yu W, Belakavadi M, Chekmareva M, Ittmann MM, Abate-Shen C, Fondell JD: **ERK and AKT signaling drive MED1 overexpression in prostate cancer in association with elevated proliferation and tumorigenicity.** *Mol Cancer Res* 2013, **11**:736–747.
109. Wei J, Yuan Y, Jin C, Chen H, Leng L, He F, Wang J: **The ubiquitin ligase TRAF6 negatively regulates the JAK-STAT signaling pathway by binding to STAT3 and mediating its ubiquitination.** *PLoS One* 2012, **7**:e49567.
110. Tiacci E, Grossmann V, Martelli MP, Kohlmann A, Haferlach T, Falini B: **The corepressors BCOR and BCORL1: two novel players in acute myeloid leukemia.** *Haematologica* 2012, **97**:3–5.
111. Iourov IY, Vorsanova SG, Liehr T, Kolotii AD, Yurov YB: **Increased chromosome instability dramatically disrupts neural genome integrity and mediates cerebellar degeneration in the ataxia-telangiectasia brain.** *Hum Mol Genet* 2009, **18**:2656–2669.
112. Park SJ, Jang HR, Kim M, Kim JH, Kwon OH, Park JL, Noh SM, Song KS, Kim SY, Kim YH, Kim YS: **Epigenetic alteration of CCDC67 and its tumor suppressor function in gastric cancer.** *Carcinogenesis* 2012, **33**:1494–1501.
113. Srougi MC, Burrige K: **The nuclear guanine nucleotide exchange factors Ect2 and Net1 regulate RhoB-mediated cell death after DNA damage.** *PLoS One* 2011, **6**:e17108.
114. Maity B, Stewart A, O'Malley Y, Askeland RW, Sugg SL, Fisher RA: **Regulator of G Protein Signaling 6 (RGS6) is a novel suppressor of breast tumor initiation and progression.** *Carcinogenesis* 2013, **34**:1747–1755.
115. Sharma P, Patel D, Chaudhary J: **Id1 and Id3 expression is associated with increasing grade of prostate cancer: Id3 preferentially regulates CDKN1B.** *Cancer Med* 2012, **1**:187–197.
116. Ha S, Iqbal NJ, Mita P, Ruoff R, Gerald WL, Lepor H, Taneja SS, Lee P, Melamed J, Garabedian MJ, Logan SK: **Phosphorylation of the androgen receptor by PIM1 in hormone refractory prostate cancer.** *Oncogene* 2013, **32**:3992–4000.
117. Bisbal C, Silverman RH: **Diverse functions of RNase L and implications in pathology.** *Biochimie* 2007, **89**:789–798.
118. Long JZ, Cisar JS, Milliken D, Niessen S, Wang C, Trauger SA, Siuzdak G, Cravatt BF: **Metabolomics annotates ABHD3 as a physiologic regulator of medium-chain phospholipids.** *Nat Chem Biol* 2011, **7**:763–765.
119. Hayashi A, Hiattari R, Tsuji T, Ohashi K, Mizuno K: **p63RhoGEF-mediated formation of a single polarized lamellipodium is required for chemotactic migration in breast carcinoma cells.** *FEBS Lett* 2013, **587**:698–705.
120. Ting HJ, Messing J, Yasmin-Karim S, Lee YF: **Identification of microRNA-98 as a therapeutic target inhibiting prostate cancer growth and a biomarker induced by vitamin D.** *J Biol Chem* 2013, **288**:1–9.
121. Itoh G, Kanno S, Uchida KS, Chiba S, Sugino S, Watanabe K, Mizuno K, Yasui A, Hirota T, Tanaka K: **CAMP (C13orf8, ZNF828) is a novel regulator of kinetochore-microtubule attachment.** *EMBO J* 2011, **30**:130–144.
122. Srivastava SK, Bhardwaj A, Singh S, Arora S, McClellan S, Grizzle WE, Reed E, Singh AP: **Myb overexpression overrides androgen depletion-induced cell cycle arrest and apoptosis in prostate cancer cells, and confers aggressive malignant traits: potential role in castration resistance.** *Carcinogenesis* 2012, **33**:1149–1157.
123. Yang P, Guo L, Duan ZJ, Tepper CG, Xue L, Chen X, Kung HJ, Gao AC, Zou JX, Chen HW: **Histone methyltransferase NSD2/MMSET mediates constitutive NF-kappaB signaling for cancer cell proliferation, survival, and tumor growth via a feed-forward loop.** *Mol Cell Biol* 2012, **32**:3121–3131.
124. Errington WJ, Khan MQ, Bueler SA, Rubinstein JL, Chakrabartty A, Prive GG: **Adaptor protein self-assembly drives the control of a cullin-RING ubiquitin ligase.** *Structure* 2012, **20**:1141–1153.
125. Yoon S, De Micheli G: **Prediction and Analysis of Human microRNA Regulatory Modules.** *Conf Proc IEEE Eng Med Biol Soc* 2005, **5**:4799–4802.
126. Cai C, Portnoy DC, Wang H, Jiang X, Chen S, Balk SP: **Androgen receptor expression in prostate cancer cells is suppressed by activation of epidermal growth factor receptor and ErbB2.** *Cancer Res* 2009, **69**:5202–5209.
127. Gandellini P, Folini M, Longoni N, Pennati M, Binda M, Colecchia M, Salvioni R, Supino R, Moretti R, Limonta P, et al: **miR-205 Exerts tumor-suppressive functions in human prostate through down-regulation of protein kinase Cepsilon.** *Cancer Res* 2009, **69**:2287–2295.
128. Al-Sarraf J, Reiff JN, Hinrichsen J, Mahmood S, Teh BT, McGovern E, De Meyts P, O'Byrne KJ, Gray SG: **DOK4/IRS-5 expression is altered in clear cell renal cell carcinoma.** *Int J Cancer* 2007, **121**:992–998.
129. Lin SL, Chiang A, Chang D, Ying SY: **Loss of mir-146a function in hormone-refractory prostate cancer.** *RNA* 2008, **14**:417–424.
130. Taganov KD, Boldin MP, Chang KJ, Baltimore D: **NF-kappaB-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses.** *Proc Natl Acad Sci U S A* 2006, **103**:12481–12486.
131. Pacifico F, Crescenzi E, Mellone S, Iannetti A, Porrino N, Liguoro D, Moscato F, Grieco M, Formisano S, Leonardi A: **Nuclear factor-(kappa)B contributes to anaplastic thyroid carcinomas through up-regulation of miR-146a.** *J Clin Endocrinol Metab* 2010, **95**:1421–1430.
132. Boldin MP, Taganov KD, Rao DS, Yang L, Zhao JL, Kalwani M, Garcia-Flores Y, Luong M, Devrekanli A, Xu J, et al: **miR-146a is a significant brake on autoimmunity, myeloproliferation, and cancer in mice.** *J Exp Med* 2011, **208**:1189–1201.
133. Bazzoni F, Rossato M, Fabbri M, Gaudiosi D, Mirolo M, Mori L, Tamassia N, Mantovani A, Cassatella MA, Locati M: **Induction and regulatory function of miR-9 in human monocytes and neutrophils exposed to proinflammatory signals.** *Proc Natl Acad Sci U S A* 2009, **106**:5282–5287.
134. Shahab SW, Matyunina LV, Hill CG, Wang L, Mezenцев R, Walker LD, McDonald JF: **The effects of MicroRNA transfections on global patterns of gene expression in ovarian cancer cells are functionally coordinated.** *BMC Med Genomics* 2012, **5**:33.
135. Li M, Collins R, Jiao Y, Ouillet P, Bixby D, Erba H, Vogelstein B, Kinzler KW, Papadopoulos N, Malek SN: **Somatic mutations in the transcriptional corepressor gene BCORL1 in adult acute myelogenous leukemia.** *Blood* 2011, **118**:5914–5917.
136. Al-Ahmadi W, Al-Haj L, Al-Mohanna FA, Silverman RH, Khabar KS: **RNase L downmodulation of the RNA-binding protein, HuR, and cellular growth.** *Oncogene* 2009, **28**:1782–1791.
137. Ravid T, Heidinger JM, Gee P, Khan EM, Goldkorn T: **c-Cbl-mediated ubiquitylation is required for epidermal growth factor receptor exit from the early endosomes.** *J Biol Chem* 2004, **279**:37153–37162.
138. Truitt L, Freywald T, DeCoteau J, Sharfe N, Freywald A: **The EphB6 receptor cooperates with c-Cbl to regulate the behavior of breast cancer cells.** *Cancer Res* 2010, **70**:1141–1153.
139. Baek KH, Hong ME, Jung YY, Lee CH, Lee TJ, Park ES, Kim MK, Yoo JH, Lee SW: **Correlation of AR, EGFR, and HER2 Expression Levels in Prostate Cancer: Immunohistochemical Analysis and Chromogenic In Situ Hybridization.** *Cancer Res Treat* 2012, **44**:50–56.
140. Zhao X, Dou W, He L, Liang S, Tie J, Liu C, Li T, Lu Y, Mo P, Shi Y, et al: **MicroRNA-7 functions as an anti-metastatic microRNA in gastric cancer by targeting insulin-like growth factor-1 receptor.** *Oncogene* 2013, **32**:1363–1372.
141. Kong X, Li G, Yuan Y, He Y, Wu X, Zhang W, Wu Z, Chen T, Wu W, Lobie PE, Zhu T: **MicroRNA-7 inhibits epithelial-to-mesenchymal transition and metastasis of breast cancer cells via targeting FAK expression.** *PLoS ONE* 2012, **7**:e41523.
142. Poliseno L, Salmena L, Riccardi L, Fornari A, Song MS, Hobbs RM, Sportoletti P, Varmeh S, Egia A, Fedele G, et al: **Identification of the miR-106b 25 microRNA cluster as a proto-oncogenic PTEN-targeting intron that cooperates with its host gene MCM7 in transformation.** *Sci Signal* 2010, **3**:ra29.

143. Majid S, Dar AA, Saini S, Shahryari V, Arora S, Zaman MS, Chang I, Yamamura S, Tanaka Y, Chiyomaru T, *et al*: **miRNA-34b inhibits prostate cancer through demethylation, active chromatin modifications, and AKT pathways.** *Clin Cancer Res* 2013, **19**:73–84.
144. Boll K, Reiche K, Kasack K, Morbt N, Kretzschmar AK, Tomm JM, Verhaegh G, Schalken J, von Bergen M, Horn F, Hackermuller J: **MiR-130a, miR-203 and miR-205 jointly repress key oncogenic pathways and are downregulated in prostate carcinoma.** *Oncogene* 2013, **32**:277–285.
145. Gong AY, Eischeid AN, Xiao J, Zhao J, Chen D, Wang ZY, Young CY, Chen XM: **miR-17-5p targets the p300/CBP-associated factor and modulates androgen receptor transcriptional activity in cultured prostate cancer cells.** *BMC Cancer* 2012, **12**:492.
146. Kokontis J, Takakura K, Hay N, Liao S: **Increased androgen receptor activity and altered c-myc expression in prostate cancer cells after long-term androgen deprivation.** *Cancer Res* 1994, **54**:1566–1573.

doi:10.1186/1476-4598-13-1

Cite this article as: Ottman *et al*: MicroRNA expressions associated with progression of prostate cancer cells to antiandrogen therapy resistance. *Molecular Cancer* 2014 **13**:1.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

