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miR deregulation in CLL

Veronica Balatti, **Yuri Pekarky**, **Lara Rizzotto**, and **Carlo M. Croce**
Department of Molecular Virology, Immunology and Medical Genetics, Comprehensive Cancer Center and the Wexner Medical Center, The Ohio State University, Columbus, OH, USA

Abstract

B-cell chronic lymphocytic leukemia (CLL) is the most frequent human leukemia and it occurs in two forms, indolent and aggressive. Although clinical features and genetic abnormalities in CLL are well documented, molecular details underlying the disease are still under investigation.

MicroRNAs are small non-coding RNAs involved in a variety of cellular processes and expressed in a tissue specific manner. MicroRNAs have the ability to regulate gene expression. In physiological conditions, microRNAs act as gene expression controllers by targeting the mRNA or inhibiting its translation. Their deregulation can lead to an alteration of the expression level of many genes which can induce the development or promote the progression of tumors.

In CLL microRNAs can function as oncogenes, tumor suppressor genes or and can be used as markers for disease onset/progression. For example, in indolent CLL, 13q14 deletions targeting *miR-15/16* initiate the disease, while in aggressive CLL *miR-181* targets the critical *TCL1* oncogene and can also be used as a progression marker. Here we discuss the foremost findings about the role of microRNAs in CLL pathogenesis, and how this knowledge can be used to identify new approaches to treat CLL.

Keywords

CLL; microRNA; miR-15/16; Tcl1

CLL: characteristics and outcomes

Chronic lymphocytic leukemia (CLL) is the most common human leukemia, accounting for ~30% of all cases of adult leukemia. In the United States, almost 15,000 new cases are diagnosed each year (Jemal A et al 2010). CLL is mostly a disease of elderly people, and the incidence increases linearly with each decade (Bullrich F 2001). This disease occurs in two forms, aggressive and indolent, both characterized by the progressive accumulation of functionally incompetent B lymphocytes expressing CD5 antigen on their surface (Bullrich F 2001). More than 90% of the leukemic cells are non dividing and are at the G_0/G_1 phase of the cell cycle (Bullrich F 2001). However, several reports showed that high lymphocyte count in CLL patients is also triggered by the presence of proliferating cells from the bone morrow, spleen or lymph nodes (Messmer BT et al 2005; Chiorazzi N 2007; Sieklucka M et al 2008). CLL cells are also quite resistant to apoptosis (Bullrich F 2001).

The clinical course of CLL is highly variable but several prognostic markers have been identified so far to facilitate the clinical management of CLL patients, such as the mutational

status of the immunoglobulin heavy-chain variable-region gene (IgH V_H), the expression levels of the 70 kD zeta-associated protein (ZAP-70), and the presence of different chromosomal alterations (Orchard JA et al 2004; Rassenti LZ et al 2004). CLLs with unmutated IgH V_H gene and high expression of the ZAP-70 usually have an aggressive course, whereas patients with mutated V_H clones and low ZAP-70 expression have an indolent course (Chiorazzi N et al 2005). Genomic alterations in CLL are also important independent predictors of disease progression and survival (Dohner H et al 2000), however, the molecular basis of these associations was largely unknown until recently. Genomic aberrations are detected by fluorescence *in situ* hybridization (FISH) in over 80% of CLL cases and include 13q, 11q, 17p and 6q deletions, and trisomy 12 (Dohner H et al 2000). The incidence of these genetic abnormalities are ~50% for deletion of 13q14, ~10% for deletion of 11q23, ~15% for trisomy 12, 7–10% for deletion of 17p, and 2–5% for deletion of 6q (Neilson JR et al 1997; Stilgenbauer S et al 1999). Prognosis is worst in patients with 17p deletion, followed by 11q deletion, trisomy 12 and normal karyotype (negative FISH panel), while patients with deletion of 13q as the only abnormality have the best prognosis (Neilson JR et al 1997; Zenz T et al 2008). Cytogenetic abnormalities can be used to identify subsets of patients with different clinical course, time to progression, and survival rates. According to recent studies, three risk groups can be differentiated: (i) low-risk: patients with a normal karyotype or isolated 13q deletion; (ii) intermediate-risk: subjects with del11q deletion, trisomy 12 or 6q deletion; and (iii) high-risk: patients with 17p deletion or a complex karyotype (Moreno C, Montserrat E 2010). Approximately one third of patients never require treatment; in another third the initial indolent phase is followed by progression of the disease, and the remaining third has aggressive disease at the onset and needs immediate treatment (Dighiero G, Binet JL 2000). Because several CLL cases show discordant prognostic factors, the identification of new parameters able to relate disease activity and clinical outcome is essential for patient management.

Signatures of microRNAs in CLLs

The miRNAs are a large family of highly conserved non-coding genes thought to be involved in temporal and tissue specific gene regulation (Ambros V 2004). miRNAs represent an evolving class of gene products with generally unknown function, and are usually excised from 70- to 80-nt stem-loop RNA precursor structures. Derived from transcripts transcribed by RNA polymerase II (Cai et al. 2004), microRNAs are made via a two step processing mechanism from a primary transcript (pri-miRNA) through an intermediate 60–90 nucleotide stem-loop structure (pre-mRNA) to the final mature microRNA. Dicer and Argonaute family members are required for the miRNA precursor processing reaction (Ambros V 2003). In mammals, single-stranded microRNA binds specific messenger RNA (mRNA) through sequences that are significantly, though not completely, complementary to the target mRNA, mainly to the 3’ untranslated region (3’ UTR) (Ambros V 2003). By a mechanism that is not fully characterized, the bound mRNA remains untranslated, resulting in reduced levels of the corresponding protein; alternatively, the bound mRNA can be degraded, resulting in reduced levels of both the corresponding transcript and consequently the protein. It was estimated that there could be from 300 to 1,000 microRNA genes in the mammalian genome (~1–3% of known genes are represented

by microRNAs). The function of most microRNAs is not known. However, recent reports revealed functions of several microRNAs: hematopoietic B-cell lineage fate (*miR-181*), B-cell survival (*miR-15a* and *miR-16-*), cell proliferation control (*miR-125b* and *let-7*), brain patterning (*miR-430*), pancreatic cell insulin secretion (miR-375) and adipocyte development (*miR-375*), reviewed in (Harfe BD 2005). Recently several the microRNAs were also linked to several types of cancer (Bartel DP 2004) and DNA methylation (Fabbri M et al 2007). Moreover, MicroRNAs can modulate gene expression in a tissue specific manner and are able to bind target messenger RNAs (mRNAs) either inhibiting their translation or promoting their degradation (Iorio MV, Croce CM 2012).

Since the first association between microRNA and cancer has been demonstrated by Calin and collegues (Calin GA et al 2002), it was clear that these genes could play a role in the clinical management of cancer patients. Numerous reports further confirmed that microRNAs are differentially expressed in cancers, thus suggesting that their deregulation could play tumor suppressor or oncogenenic roles in cancer pathogenesis (Volinia S et al 2006).

MicroRNA expression profiles revealed several remarkable outcomes that could be applied to clinic. MicroRNAs profiles can be used to distinguish normal B-cells from malignant CLL cells and, more importantly, they are associated with prognosis, progression and drug resistance in CLL (Ferracin M et al 2010). In particular, a signature profile was reported, describing 13 microRNAs that differentiate aggressive and indolent CLL (Calin GA et al 2004). Another report showed that the expression profile of 32 microRNAs is able to discriminate between cytogenetic subgroups (Visone R et al 2009). For instance, patients with high levels of *miR-21* had a higher risk of death compared to patients with low expression levels (Rossi S et al 2010). Likewise, high expression of *miR-155* was reported in the aggressive form of CLL (Calin GA et al 2007). Intriguingly, we recently found that expression levels of *miR-181b* can not only distinguish between indolent and aggressive cohorts of patients but can also predict time to treatment, acting as a biomarker of the disease progression. We studied serial time points derived from the same patients and found that expression of *miR-181b* decreases along with the severity of the disease. These new findings highlight the importance of *miR-181b* in clinics, suggesting that expression levels of microRNAs can be used not only to classify patients according to the gravity of the pathology, but also tracking the disease course (Visone R et al 2011). Moreover, microRNA signature can be also used to predict refractoriness to Fludarabine treatment in CLL (Ferracin M et al 2010). To clarify if microRNAs are directly involved in the development of fludarabine resistance, Ferracin et al. analyzed the expression of microRNAs before and after Fludarabine therapy in patients classified as responder or refractory and identified a microRNA signature able to distinguish between these two classes. Expression levels of several microRNAs were also able to predict fludarabine resistance in an independent test cohort. Among these microRNAs, *miR-148a*, *miR-222* and *miR-21* exhibited a significantly higher expression in non-responders either before or after treatment. Recently, Zenz and colleagues found that fludarabine refractory CLLs are frequently characterized by lower levels of *miR-34a* (Zenz T et al 2009), and low expression of miR-34a was associated with fludarabine resistance even in the absence of p53 aberrations (Zenz T et al 2009).

To conclude, microRNA expression levels can distinguish normal B-cells from CLL, discriminate between indolent and aggressive CLL forms, indicate the progression of the disease, and separate responder and refractory cohorts of patients. These findings provide new roles for microRNAs as markers for CLL development/sensitivity to treatment (Ferracin M et al 2010) and potential predictors of time to treatment (Visone R et al 2011).

Role of microRNAs in CLL

Besides using microRNAs expression levels as tools to discriminate different CLL forms or to keep track of disease progression, researchers have recently focused on the molecular impact of microRNA deregulation in CLL. Interestingly, the *miR-15/16* cluster, *miR-29*, *miR-181* family members, and *miRs-34b/c*, were found as the most deregulated microRNAs in CLL. The same microRNAs were found to regulate gene expression patterns, helping to clarify molecular steps that lead to the onset of the disease or drive its progression.

MicroRNA 15a/16-1

In CLL, deletion at chromosome 13q14.3 is the most frequent genomic aberration (about 50% of cases) and is associated with the longest treatment-free interval (Dohner H et al 2000). The first attempts to identify tumor suppressor genes at 13q14 locus by using positional cloning and sequencing of a region of more than 1 Mb failed (Bullrich F et al 2001; Migliazza A et al 2001); moreover none of the known genes in this region were found to be down-regulated in CLL by deletions or mutations (Bullrich F et al 2001; Migliazza A et al 2001; Rondeau G et al 2001; Mertens D et al 2002). In 2001 we generated somatic cell hybrids using mouse and CLL cells carrying 13q14 deletion and translocations and we identified a 30-kb region of deletion between exon 2 and exon 5 of *LEU2* gene (Calin GA et al 2002; Pekarsky Y et al 2005). Interestingly, the translocation breakpoint was mapped to the same region (Calin GA et al 2002; Pekarsky Y et al 2005). Since *LEU2* had previously been sequenced and excluded as a candidate tumor suppressor gene in 13q14 (Bullrich F et al 2001; Migliazza A et al 2001; Wolf S et al 2001; Mertens D et al 2002), we continued to investigate that region and finally discovered a cluster of two noncoding microRNA genes, *miR-15a* and *miR-16-1*, located exactly within the deleted region and near the translocation breakpoint (Calin GA et al 2002). Accordingly, *miR-15a/16-1* cluster was found deleted or its expression down-regulated in ~66% of CLL cases (Calin GA et al 2002, Smonskey MT et al 2012). In contrast, expression levels of the other genes in the region (*DLEU1*, *DLEU2* and *RFP5*) were not affected by the 13q14 deletions (Bullrich F et al 2001; Migliazza A et al 2001; Pekarsky Y et al 2005).

The first genetic manipulation in mice that confirmed the importance of *miR-15a/16-1* deletion in CLL was carried out by Dr. Dalla-Favera and colleagues (Klein U et al 2010). These authors designed a model with conditional alleles that either resembled the loss of the minimal deleted region (*Mdr*), already characterized in human CLL and that spans entirely the *DLEU2* gene (Migliazza A et al 2001), or the specific *miR-15a/16-1* cluster deletion, without altering Dleu2 expression (Klein U et al 2010). Both *Mdr* and *miR-15a/16-1* knockout strains at one year of age presented approximately 50% of $CD5^+$ $B220^+$ B-cells among mononuclear cells in the peritoneum versus 15% in control animals. In total, mice with CLL were 27% of *Mdr* KO and 21% of *miR-15a/16-1* KO, while some type of clonal

B-cell proliferation affected 42% of *Mdr* KO and 26% of *miR-15a/16-1* KO mice between 15 and 18 months of age. *Mdr* KO animals lived less than WT siblings and eventually succumbed to leukemias, while the differential survival between *miR-15a/16-1* and their WT littermates was not statistically significant, providing evidence that the latter were affected by a phenotype milder than the former. Because of the more aggressive disease shown by *Mdr* KO mice, it is likely that other elements included in the *Mdr* locus, like the *DLEU2* gene itself, may participate to CLL tumor suppression (Klein U et al 2010). Mechanisms leading to B-cell proliferations were investigated with different approaches. *MiR-15a/16-1* KO B-cells were shown to begin DNA synthesis earlier than WT B-cells (Klein U et al 2010). The authors also analyzed levels of phosphorylated retinoblastoma (pRb) protein, an indicator of entry into the cell cycle, in mitogen-stimulated B-cells isolated from *miR-15a/16-1* KO or *Mdr* KO and WT animals. PRb was produced in both KO B-cells at earlier time points than in WT B-cells. Individual contributions of *miR-15a/16-1* cluster versus *DLEU2* gene to the lympho-proliferation were dissected generating an inducible system where these two genetic elements underwent separate *in vitro* re-expression in a human cell line derived from a 13q14 KO CLL. These findings demonstrated that impaired proliferation occurred in *miR-15a/16-1* expressing cells, with higher fraction of cells in G0/G1 phase, but not in those expressing Dleu2, thus suggesting a possible control of the inhibition of G0/G1 phase transition by *miR-15a/16-1* (Klein U et al 2010).

The importance of *miR-15a/16-1* cluster in CLL was confirmed in a study of CLL development in New Zealand black (NZB) mice, the only mouse strain that naturally develops CLL (Raveche ES et al 2007). In NZB mice, CLL arises late in life, with an autoimmune phenotype and B-cell hyper-proliferation followed by slow progression to late-onset CLL (Raveche ES 1990; Zanesi N et al 2010). Older NZB animals show a clonal expansion of the sub-population of B-1 B-cells similar to that found in human CLL (Raveche ES 1990; Zanesi N et al 2010). Linkage analysis has found that the mouse genomic region homologous to 13q14 is one of the *loci* associated with CLL development. Subsequent DNA sequencing resulted in the identification of a point mutation in *miR-15a/16-1* precursor causing a decrease of *miR-16-1* expression in NZB lymphoid tissues, accompanied by elevated levels of Bcl-2 (Raveche ES et al 2007). Accordingly, lymphoid tissues from NZB mice were analyzed for the levels of mature *miR-16-1* and showed reduced expression of this microRNA. Finally, delivery of exogenous *miR-16-1* to a NZB malignant cell line led to cell cycle alterations such as decrease in S phase cells and G1 arrest (Raveche ES et al 2007). Other strains of mice including the NZW strain, the closest relative of NZB, did not show the mutation in *miR-15a/16-1* precursor.

B cell lymphoma 2 (*BCL2*) is a central player in the genetic program of eukaryotic cells, promoting survival by inhibiting cell death (Cory S Adams JM 2002). Over-expression of Bcl2 protein has been reported in many types of human cancers, including leukemias, lymphomas, and carcinomas (Sanchez-Beato M et al 2003). In follicular lymphomas and in a fraction of diffuse large B cell lymphomas, *BCL2* is activated due to the translocation t(14,18)(q32;q21), which places the *BCL2* gene under the control of Ig heavy chain enhancers, resulting in the over-expression of the gene (Tsujimoto Y et al 1984; Tsujimoto Y et al 1985). In CLL, malignant B cells over-express Bcl2 (Kitada S et al 1998); however,

with the exception of less than 5% of cases, in which the *BCL2* gene is juxtaposed to Ig loci (Adachi M et al 1990), no mechanism has been discovered to explain *BCL2* up-regulation in CLL. M*iR-15a* and *miR-16-1* expression is inversely correlated to Bcl2 expression in CLL and these microRNAs negatively regulate *BCL2* at post-transcriptional level (Cimmino A et al 2005). Since *BCL2* is a predicted target of both *miR-15a* and *miR-16-1*, the down-regulation of these microRNAs in a leukemic cell line resulted in an increase of Bcl2 expression with consequent inhibition of apoptosis (Cimmino A et al 2005). Interestingly, *miR-15a/16-1* expression also resulted in growth inhibition of tumor engraftment of leukemic cells in nude mice, confirming the tumor suppression properties of these microRNAs (Calin GA et al 2008). In summary, Bcl2 over-expression driven by down-regulation of *miR-15a* and *miR-16-1* seems to be a regulatory mechanism involved in the pathogenesis of a large part of human CLL. These studies determined that *miR-15a/16-1* cluster functions as a tumor suppressor in CLL by inhibiting Bcl2, and deletions at 13q14 represent an initializing step in CLL development (Cimmino A et al 2005). In this respect *miR-15a/16-1* have promise to be used as a drugs for CLL.

Since the indolent form of the disease is often characterized by 13q14 deletion, it is likely that up-regulation of *BCL2* plays a major role in this subset of CLLs. Evidence for this hypothesis came from Dr. Reed and colleagues, who used two previously described mouse models, one with Bcl2 over-expression in the lymphoid system (Katsumata M et al 1992), and the second with up-regulation of a specific isoform of *TRAF2* (TNF receptor associated factor 2) in B and T-cells (Lee SY et al 1997). *TRAF2* can bind to TNF receptor family and mediate the activation of NF-kB by TNF proteins (Chung JY et al 2002); TNF-mediated signaling increased lymphocyte proliferation and survival (Haiat S et al 2006).

TRAF2 transgenic mice failed to develop a frank leukemia, but showed an increased number of B-cells accompanied by lympho-adenopathy and splenomegaly (Lee SY et al 1997). *BCL2* transgenic animals, which were designed with a construct mimicking t(14;18) translocation juxtaposing *BCL2* gene with the immunoglobulin heavy-chain locus at 14q32 as reported in human follicular lymphomas, did not develop malignancies either, presenting only prolonged *in vitro* B-cell survival and *in vivo* polyclonal B-cell expansions (Katsumata M et al 1992).

TRAF2DN-BCL2 double transgenic mice, on the other hand, displayed severe splenomegaly and most animals were affected by a CLL-like disease with high B-cell blood count (Zapata JM et al 2004). While single transgenics showed a normal lifespan, the double ones survived only between 6 and 14 months. Because of their complex features, it was not clear whether *TRAF2DN-BCL2* transgenics were a model of indolent or aggressive CLL (Pekarsky Y et al 2007).

Based on these findings, 13q14 deletions could induce CLL development by a molecular mechanism resembling the oncogenic events in *TRAF2DN/BCL2* transgenics(Palamarchuk A et al 2010). In fact, in addition to *miR-15a/16-1*, the 13q14 region deleted in indolent CLL contains *DLEU7* gene, located telomeric to *miR-15a/16-1* (Ouillette P et al 2008). Our report showed that *DLEU7* is a cooperating tumor suppressor along with miR-15a/16- and we

recently confirmed that *DLEU7* deletions result in the induction of TNF signaling through TRAFs, while *miR-15a/16-1* deletions cause a constitutive increase of Bcl2 expression.

DLEU7 was previously identified as a candidate tumor suppressor gene at 13q14 (Hammarsund M et al 2004). Recently, Ouillette *et al*., by using microarray technology, have displayed that the minimal deleted region at 13q14 in CLL contains *DLEU7* gene (Ouillette P et al 2008). Since *DLEU7* is the only protein coding gene located within reported minimal deleted region at 13q14, we investigated whether *DLEU7* can cooperate with *miR-15a/16-1* (Palamarchuk A et al 2010). Sequencing of *DLEU7* coding exons failed to find mutations in CLL samples, although a previous study reported hyper-methylation of *DLEU7* promoter, with consequent silencing of this gene in 61% of CLL cases (Hammarsund M et al 2004). Real time RT-PCR experiments confirmed that expression of *DLEU7* in CLL samples is decreased when compared to normal CD19+ B-cells. *MiR-15a/16-1* was also found down regulated in the same CLL samples (Palamarchuk A et al 2010).

Since recent studies confirmed a significant role for the NF-kB pathway in the pathogenesis of CLL (Pekarsky Y et al 2007), we examined whether Dleu7 might function as an inhibitor of NF-kB. In the inactive state, NF-κB proteins are bound to IκB proteins in the cytoplasm; after stimulation, IκB is degraded and NF-κB translocate the nucleus (Brockman JA et al 1995; Chen Z et al 1995; Ghosh S et al 1998). Induction of NF-κB can be driven by a variety of stimuli, including exposure to members of the tumor necrosis factors superfamily (TNF), chemotherapy, and ionizing radiation (Beg AA Baltimore D 1996; Van Antwerp DJ et al 1996; Wang CY et al 1996). Activation of NF-kB prevents B-cells from undergoing apoptosis and regulates growth and differentiation (Beg AA Baltimore D 1996; Van Antwerp DJ et al 1996; Wang CY et al 1996). In B-cells, it has been shown that transgenic expression of the TNF ligand APRIL resulted in an expansion of $B220^{+}CD5^{+}$ cells (Planelles L et al 2004). APRIL binds BCMA (B-cell maturation antigen) and TACI (Haiat S et al 2006), which stimulate the NF-kB pathway thus suggesting that NF-kB activation through TACI and BCMA is important in the pathogenesis of CLL (Palamarchuk A et al 2010). Moreover, nuclear factor of activated T-cells (NFAT) can also be activated by TACI and BCMA (Mackay F et al 2003); NFAT was previously reported as a hallmark of unstimulated CLL cells (Schuh K et al 1996; Berland R Wortis HH 1998).

Since *DLEU7* is located within the 13q14 deleted region and NF-kB/NFAT activation can be critical in CLL pathogenesis, we studied whether Dleu7 expression has an effect on NF-kB and NFAT activation by TACI and BCMA. Our experiments showed that Dleu7 expression inhibits NF-kB activation by BCMA over 5 fold, while activation by TACI was inhibited over 4 fold (Palamarchuk A et al 2010). Also, Dleu7 expression can inhibit NFAT activation by TACI and BCMA ~8 fold. Thus, we concluded that Dleu7 functions as NFAT and NF-kB inhibitor (Palamarchuk A et al 2010).

In conclusion, *miR-15a/16-1*, deletion is an initializing step in CLL development, eliciting the control on Bcl2 expression level and cooperating with DLEU7 in promoting the activation of NF-kB and NFAT via TACI and BCMA. Moreover, we also recently discovered a *miR-15a/16-1-TP53* feedback circuitry, in which p53 directly transactivates

miR-15a/16-1 promoter, while *miR-15a/16-1* cluster targets *TP53* expression (Fabbri M et al 2011).

MicroRNA 34b/c

It is currently unknown how the 11q, 17p and 13q deletions contribute to CLL pathogenesis and progression (Dohner H et al 2000). However, it has been proved that the loss of the long arm of chromosome 11 includes the region where the *miR-34b/c* cluster is located (Auer RL et al 2007), while deletion of 17p leads to abrogation of the p53 tumor suppressor (Merkel O et al 2010) and 13q deletion involves *miR15a/16*- down-regulation. To establish the possible existence of molecular interactions between these chromosomal alterations, we investigated if the *miR-15a/16-1* cluster, tumor protein p53, and *miR-34b/c* cluster are connected in a molecular pathway that could explain the prognostic implications (aggressive vs indolent form) of 11q, 17p, and 13q deletions in CLL (Fabbri M et al 2011).

Several *TP53* binding sites were found upstream of the *miR-15a/16-1* on chromosome 13 and of the *miR-34b/c* on chromosome 11. Chromatin immunoprecipitation analysis revealed that *TP53* directly binds to its predicted binding sites on both chromosomes 13 and 11. Thus, *TP53* can induce the expression of both these microRNAs (Fabbri M et al 2011). On the other hand, *miR-15a/16-1* target *TP53* while a binding site for the *miR-34* family was predicted in ZAP-70 mRNA (Fabbri M et al 2011). These interactions could lead to different outcomes via feedback circuits involving protein coding genes and microRNAs (Fabbri M et al 2011). In this model, *TP53* (on chromosome 17p) represents the molecular connection between *miR-15a/16-1* (on chromosome 13q) and *miR-34b/c* (on chromosome 11q) (Fabbri M et al 2011).

In 13q deleted patients the loss of *miR-15a/16-1* expression shifts the balance not only toward higher levels of anti-apoptotic proteins Bcl2 (Cimmino A et al 2005; Calin GA et al 2008), but also toward higher levels of the tumor suppressor protein p53. Consequently, in 13q patients, while the number of apoptotic cells may decrease because of the increased levels of Bcl2, the p53 tumor suppressor pathway remains intact, thus keeping the increase in tumor burden relatively low. This finding could explain how 13q deletions are associated with the indolent form of CLL. Moreover, increased p53 levels in patients with 13q deletions are associated with transactivation of *miR-34b/c* and with reduced levels of ZAP-70 (Rassenti LZ et al 2004), and further supporting the indolent course of CLLs carrying 13q deletions.

CLL patients with 11q deletion, instead, express significantly lower levels of *miR-34b/c* and significantly higher levels of ZAP-70, both at mRNA and protein level. These patients show poorer overall survival than patients with normal cytogenetic profiles and lower levels of ZAP-70. In these patients TP53 is not upregulated because *miR-15a/16-1* are not deleted and this condition is associated with lower control on apoptosis (Fabbri M et al 2011).

In conclusion, we demonstrated that a microRNA/TP53 feedback circuitry is associated with pathogenesis of CLL. These results also showed that restoring expression of *miR-15a/16-1* indirectly affects expression of *miR-34* family by modulating levels of TP53

expression. Moreover, *miR-34* family is a downstream target of p53 and its overexpression can cause p53-like effects on apoptosis or cell cycle arrest (Fabbri M et al 2011).

MicroRNA 29

In both indolent and aggressive CLLs, *miR-29* is over expressed compared to normal B-cells, but its role in development/progression of CLLs is still unclear. In addition, expression levels of *miR-29* is higher in indolent than in aggressive CLLs (Calin GA et al 2005; Pekarsky Y et al 2006; Santanam U et al 2010). These results prompted us to evaluate the role of this microRNA in CLL. The up-regulation of *miR-29* in indolent CLL compared to normal B-cells implies an oncogenic function for this microRNA, initiating or at least significantly contributing to the pathogenesis of CLL (Calin GA et al 2005; Pekarsky Y et al 2006; Santanam U et al 2010). On the other hand, we showed that expression levels of *TCL1* and *miR-29* are inversely correlated, and that *miR-29* targets *TCL1* expression (Pekarsky Y et al 2006), thus suggesting a possible tumor suppressor function for *miR-29* in aggressive CLL. Furthermore, a microRNA signature was published with 13 microRNAs that differentiate aggressive and indolent CLL (Calin GA et al 2004). Intriguingly, of the four down-regulated microRNAs in aggressive CLL, three are different isoforms of *miR-29* (*miR-29a-2, miR-29b-2* and *miR-29c*) (Calin GA et al 2004), strongly suggesting that deregulation of *miR-29* can play a role in the pathogenesis of aggressive CLLs. In addition, expression of members of *miR-29* family could discriminate between CLL samples with good and bad prognosis (Calin GA et al 2005).

In order to study the role of *miR-29* in B-cell leukemias, we designed a transgenic mouse characterized by over-expression of *miR-29* in B cells. In splenocytes from these transgenics we reported an increase in $CD5^+$ $CD19^+$ IgM^+ B-cell populations, an hallmark of CLL (Santanam U et al 2010). Eighty-five percent of *miR-29* animals showed a marked growth of $CD5^+$ B-cells that, between 12 and 14 months of age, represented up to 50% of total B-cells. Only 20% of the transgenics died because of leukemia between 24 and 26 months of age. These data led us to conclude that *miR-29* mice mimicked the indolent form of CLL. In fact, the percentage of leukemic cells increased with age, from 20% of all B-cells in mice below 15 months of age to more than 65% in mice above 20 months of age, indicating a gradual progression of indolent CLL (Santanam U et al 2010). Using BrdU incorporation experiments to measure the proliferative capacity of leukemic cells, we confirmed a significantly increased proliferation in *miR-29* transgenic B-cells compared to wild type $CD19^+$ cells, where no proliferation was found. Thus, *miR-29* over-expression seems to play a role in promoting B-cell proliferation. Furthermore, since immune incompetence and progressive hypogammaglobulinemia are typical features of human CLL, immune response to SRBC antigen and serum levels of immunoglobulins were analyzed in *miR-29* mice and their wild type littermates. Both parameters were drastically decreased in transgenic animals, confirming that *miR-29* transgenics mimicks the indolent course of human CLL (Santanam U et al 2010).

In aggressive CLLs, the down-regulation of *miR-29* appears to be involved in Tcl1 over-expression, along with *miR-181* (Pekarsky Y et al 2006). Activation of the *TCL1* oncogene is a central initiating event in the pathogenesis of aggressive CLL. *TCL1* (T cell leukemia/

lymphoma 1) was originally identified as a target of translocations and inversions at 14q32.1 in T-cell prolymphocytic leukemias (T-PLL) (Virgilio L et al 1994). High Tcl1 expression in human CLL correlates with aggressive phenotype (Herling M et al 2006). Tcl1 functions as a promoter of PI3K - Akt(PKB) oncogenic pathway (Laine J et al 2000; Pekarsky Y et al 2000) activating Akt, driving its nuclear translocation and leading to an increased proliferation, inhibition of apoptosis and transformation (Pekarsky Y et al 2000). At the same time Tcl1 activates NF-kB, inhibits AP-1 (Pekarsky Y et al 2008) and restrains *DNMT3a* (Palamarchuk A et al 2012), which is involved in epigenetic deregulation of gene expression. This leads to defects in cell death, increased survival, and CLL pathogenesis.

Recently we investigated whether *TCL1* expression in CLL is regulated by microRNAs (Pekarsky Y et al 2006). *MiR-29b* and *miR-181b* are down-regulated in aggressive CLLs with 11q deletions and are predicted to target Tcl1 (Pekarsky Y et al 2006). Interestingly, *miR-181* is differentially expressed in B-cells and *TCL1* is mostly a B-cell specific gene (Ramkissoon SH et al 2006), thus suggesting that Tcl1 might be a target of *miR-181* not only in CLL cells but also in normal B-lymphocytes. We therefore proceeded to verify if these microRNAs really target Tcl1 expression. Our experiments revealed that co-expression of Tcl1 with *miR-29* and *miR-181* significantly decreased Tcl1 expression (Pekarsky Y et al 2006) and we consequently concluded that *miR-29b* and *miR-181b* target *TCL1* expression on mRNA and protein levels (Pekarsky Y et al 2006). Concordantly, we found inverse correlation between *miR-29b* and *miR-181b* expression and Tcl1 protein expression in CLL samples, which further support the idea that Tcl1 expression in CLL is, at least in part, regulated by *miR-29* and *miR-181* (Pekarsky Y et al 2006).

Since *TCL1* expression is regulated by microRNAs, like *miR-29* and *miR-181*, that target the 3'UTR region of the gene, we generated transgenic mice of Eμ-*TCL1* Full Length (Eμ-*TCL1* FL) including both the 3' and 5' UTRs of *TCL1* under a B-cell specific promoter (Efanov A et al 2010). These animals showed the development of a CLL-like leukemia between sixteen and twenty months of age and a population of $CD5^+$ $CD23^+$ B-cells accumulated in spleens and lymph nodes of these mice. Immunological abnormalities like hypoimmunoglobulinemia, impaired immune response, and abnormal levels of cytokines were also found in Eμ-*TCL1* FL animals and were similar to those observed in human CLL (Efanov A et al 2010). In conclusion, both classical Eμ-*TCL1* and Eμ-*TCL1* FL transgenic mouse model of CLL displayed important biological similarities with their human counterpart that went beyond the simple resemblance between the two leukemias. Our study demonstrated that *TCL1* upregulation in mouse B-cells results in aggressive CLL (Bichi R et al 2002).

In conclusion, the current idea of the role of *miR-29* in CLL is associated with its effect on Tcl1 expression levels in both indolent and aggressive forms. Since *TCL1* is generally not expressed in indolent CLL (Pekarsky Y et al 2006), it likely does not play an important function in indolent CLL and its down-regulation due to *miR-29* over-expression does not slow indolent CLL development. Up-regulation of *miR-29* expression is not sufficient to cause aggressive CLL; on the other hand, up-regulation of Tcl1 is absolutely required for the initiation of the aggressive form of CLL. Down-regulation of *miR-29* expression in

aggressive CLL (compared to the indolent form) contributes to up-regulation of Tcl1 and development of aggressive CLL (Pekarsky Y Croce CM 2010).

Effects of polymorphisms and epigenetic regulation on microRNAs expression

The complexity of the pathways involving microRNAs in CLL development/progression was found to extend beyond their ability to directly regulate gene expression. MicroRNA expression can respond to the presence of single nucleotide polymorphisms (SNPs) and can also be altered by trasactivator factors (Asslaber D et al 2010). Moreover, deregulation of epigenetic processes can modify microRNA expression, leading to a diverse progression of the disease and a different prognosis (Sampath D et al 2012).

A good example of SNPs being involved in altered microRNA expression is offered by *miR-34a* (Asslaber D et al 2010). *MiR-34a* has been implicated in the CLL response to DNA damage through a p53-mediated induction (Dijkstra MK et al 2009; Mraz M et al 2009; Zenz T et al 2009). TP53 protein transactivates *miR-34a* on chromosome 1p36 inducing tumor suppressor effects, enhancing apoptosis and cycle arrest (Bommer GT et al 2007; Chang TC et al 2007; He L et al 2007; Tarasov V et al 2007). The presence of a single nucleotide polymorphism (SNP 309) in the intronic region of the promoter of ubiquitine ligase *MDM2* leads to increased expression of *MDM2*, which binds p53 (Asslaber D et al 2010). In patients with intact p53, it has been reported that the presence of this SNP inhibits p53 transactivation effects on *miR-34a* and can induce down-regulation of *miR34a* (Asslaber D et al 2010). In many types of cancer this SNP has been associated with accelerated tumor formation and poor prognosis (Menin C et al 2006; Ohmiya N et al 2006; Gryshchenko I et al 2008). Asslaber et al. has shown that the GG genotype of *MDM2* SNP 309 is associated with reduced overall survival and treatment-free survival in CLL. CLL cells of patients with the GG-genotype had a significantly lower mean expression of *miR-34a* as compared with the TT-genotype, suggesting attenuation of the p53 pathway by the SNP 309. *MiR-34a* levels in cells with the heterozygous GT-genotype were found between those with the GG and the TT genotype. Thus, the presence of this SNP restrains p53 activity on the *miR-34a* expression in CLL patients without p53 deletion/mutation (Asslaber D et al 2010).

MicroRNAs can be also involved in epigenetic gene regulation with positive and negative feedbeck circuits (Sampath D et al 2012). The histone deacetylases (HDACs) are chromatin-modulating enzymes that catalyze the removal of acetyl groups on specific lysines around gene promoters (van der Vlag J Otte AP 1999). Moreover, they can trigger the demethylation of lysine 4 on histones (H3K4me2/3), thus promoting chromatin compaction and leading to epigenetic gene silencing (van der Vlag J Otte AP 1999). Recent data established that HDACs can also silence microRNAs. In particular, it has been observed that *miR-15a/16-1* are silenced by epigenetic mechanisms in 30%-35% of CLL samples, therefore cooperating with 13q14 deletion to account for the low expression levels of these microRNAs in CLL (Sampath D et al 2012). Indeed, it has been found that HDAC1–3 is over-expressed in CLL but not in normal lymphocytes, hence identifying an independent mechanism for the silencing of *miR-15a/16-1* (Sampath D et al 2012).

In samples with monoallelic 13q14 deletion it has been observed that the HDACs repressed *miR-15a/16-1* expression on the residual allele, providing an example of functional cooperation between a genetic and epigenetic mechanism to achieve gene repression. Induction of *miR-15a/16-1* in response to HDAC inhibition is associated with activation of cell death. Future prospective trials should evaluate the specific impact of epigenetic silencing of *miR-15a/16-1* on disease behavior and progression, that could represent a new therapeutic strategy to antagonize an important survival mechanism in cells. CLL patients who exhibit such epigenetic silencing may represent a group that will possibly benefit from HDAC inhibitor–based therapy (Sampath D et al 2012).

CONCLUSIONS

Chronic lymphocytic leukemia (CLL) is a heterogeneous disease. Karyotypic aberrations are strongly prognostic of survival as well as IgH V_H mutational status and ZAP70 expression. Lately, microRNAs expression has been considered as a new important tool in the management of the desease. In order of highest to lowest risk, the genomic categories so far identified are: 17p deletion, 11q deletion, trisomy 12, normal FISH and 13q deletion. Patients with 17p deletion respond poorly to treatment while patients with 11q deletion CLL show a better response to treatment, even if they progress early. Moreover, unmutated IgH V_H/ZAP70 positives patients have increased rates of progression and reduced remission durations.

MicroRNAs differentially expressed in cancers and their deregulation could play tumor suppressor or oncogenenic roles in cancer pathogenesis. MicroRNA expression profiles have been found to be useful tools to distinguish normal B-cells from malignant CLL cells and can be correlated with prognosis, progression and drug resistance of CLL. MicroRNAs modify gene expression and their deregulation involves downstream effects on cell cycle and proliferation. Deletion of *miR-15a/16-1* has been correlated to Bcl2 up-regulation in indolent CLL, while down-regulation of *miR-29* and *miR-181* has been correlated to Tcl1 upregulation in aggressive CLL. On the other hand, over-expression of *miR-29* in B-cells results in development of indolent CLL. *MiR-34* family members are involved in a fine regulated feedback circuitry with p53 and *miR-15a/16-1* in 13q deleted CLL, thus suggesting that the interplay between microRNAs and genes is bidirectional.

Deregulation of microRNAs can be a consequence of chromosomal alteration, epigenetic modulation, or interaction with other genes. In fact, microRNAs can be epigenetically silenced, suggesting a new cooperating system of abnormal regulation of these molecules. The study of these mechanisms can clarify the role of microRNAs in development and progression of CLL and allow the identification of new targets for therapy.

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