

Open access • Journal Article • DOI:10.2174/1381612823666170926123550

# Modelling Bronchopulmonary Dysplasia in Animals: Arguments for the Preterm Rabbit Model — Source link

<u>Thomas Salaets, Andre Gie, Bieke Tack, Jan Deprest</u>...+2 more authors **Institutions:** <u>Katholieke Universiteit Leuven, University College London</u> **Published on:** 01 Jan 2017 - <u>Current Pharmaceutical Design</u> (Curr Pharm Des) **Topics:** Bronchopulmonary dysplasia, Lung injury and Intensive care

#### Related papers:

- · Functional assessment of hyperoxia-induced lung injury after preterm birth in the rabbit
- · Looking ahead: where to next for animal models of bronchopulmonary dysplasia?
- Animal models of bronchopulmonary dysplasia. The preterm baboon models.
- Recent Advances in Bronchopulmonary Dysplasia
- Bronchopulmonary dysplasia: clinical aspects and preventive and therapeutic strategies.

### TITLEPAGE

Modelling bronchopulmonary dysplasia in animals: arguments for the preterm rabbit model.

Thomas Salaets<sup>1,2</sup>, Andre Gie<sup>1</sup>, Bieke Tack<sup>2</sup>, Jan Deprest<sup>1,3</sup>, Jaan Toelen<sup>1,2</sup>

<sup>1</sup> Department of Development and Regeneration, Cluster Organ Systems, Faculty of Medicine, University of Leuven, Herestraat 49, Leuven 3000, Belgium.

<sup>2</sup> Department of Pediatrics, Division Woman and Child, University Hospitals Leuven, Herestraat 49, Leuven 3000, Belgium.

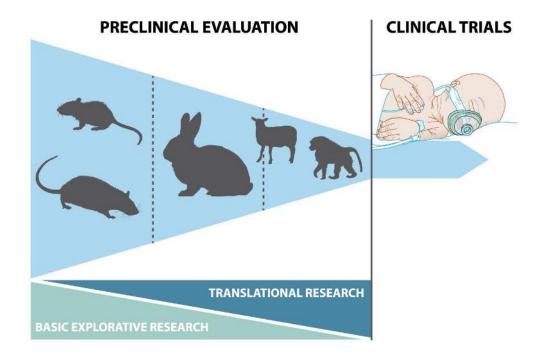
<sup>3</sup> Department of Obstetrics and Gynaecology, Division Woman and Child, University Hospitals Leuven, Herestraat 49, Leuven 3000, Belgium.

CORRESPONDING AUTHOR Jaan Toelen, MD, PhD Department of Pediatrics, Division Woman and Child University Hospitals Leuven Herestraat 49, 3000 Leuven, Belgium Email: jaan.toelen@uzleuven.be Phone: +32 16 34 20 97, Fax: +32 16 34 42 05

### ABSTRACT

Bronchopulmonary dysplasia (BPD) remains a frequent and disabling consequence of preterm birth, despite the recent advances in neonatal intensive care. There is a need to further improve outcomes and many novel therapeutic or preventive strategies are therefore investigated in animal models. We discuss in this review the aspects of human BPD pathophysiology and phenotype, which ideally should be mimicked by an animal model for this disease. Prematurity remains the common denominator in the heterogeneous spectrum of human BPD, and preterm animal models thus have a clear translational advantage. Additional factors, like excessive oxygen, mechanical ventilation and infection, which frequently have been studied in animal models, can contribute to preterm lung injury however are not indispensable to develop BPD. The phenotype of human BPD is characterized by alveolar developmental arrest with extracellular matrix remodeling, signs of obstructive airway disease and pulmonary vascular disease. Many animal models mimic this phenotype and have their place in BPD research, but results should be interpreted bearing in mind the specific advantages and disadvantages of the model. Term mice and rats are well suited for basic explorative research on specific disease mechanisms, essential for the generation of new hypotheses, while the larger ventilated preterm baboons and lambs provide a good platform for the ultimate translation of these strategies towards clinical application. The preterm rabbit model seems a promising model as it the smallest model that includes a factor of prematurity and has a unique position between the small and large animal models.

### **GRAPHICAL ABSTRACT**



### **KEYWORDS**

bronchopulmonary dysplasia; animal models; rabbit; preclinical research; translational research; prematurity

#### 1 MAIN TEXT

#### 2 Introduction

3

4 Premature birth affects 11% of all pregnancies and is the leading cause of death in neonates worldwide [1]. Fifty 5 years ago Northway was the first to describe the fibrotic changes in the lungs of preterm infants who could only 6 be kept alive with aggressive mechanical ventilation and supraphysiological oxygen administration [2]. Even 7 though advances in perinatal medicine have increased the survival of ever more premature infants, these were 8 unable to reduce long-term respiratory morbidity. At present 45% of survivors of extremely preterm birth (<28 9 weeks of gestation) still develop bronchopulmonary dysplasia (BPD) [3]. This lung disease has disabling 10 consequences throughout life as it is associated with respiratory morbidity (lung function abnormalities, episodes 11 of wheezing and frequent hospital admissions), pulmonary hypertension (which is associated with a mortality rate 12 up to 38% [4, 5]) and even abnormal neurological development [6, 7]. Also economically BPD remains an 13 important problem [8]. As the current clinical care fails to prevent BPD, the development of effective preventive 14 and therapeutic strategies remains imperative.

15 Jensen et al. have reviewed the evidence behind the current pharmacological strategies for prevention of BPD in 16 2015 [9]. There is high-quality evidence that vitamin A [10] and caffeine [11] reduce BPD, but their effect size is 17 limited (number needed to treat of 11,5 and 9,5 respectively). Systemic corticosteroids also lower the incidence of 18 BPD, but serious concerns exist on their safety profile. Recently however, a number of clinical trials have shown 19 success with local delivery [12, 13] and low dose regimens [14, 15] of steroids. However, the uncertainty about 20 their safety remains as for instance inhaled budesonide resulted in a trend towards increased mortality [12]. Jensen 21 et al. also reported on the currently ongoing clinical trials. Besides intratracheal stem cells, only few novel 22 medicinal products are being investigated. Yet a recent trial, where recombinant insulin-like growth factor 1 (IGF1) 23 was given to preterm neonates with the primary aim to reduce the incidence of retinopathy of prematurity, 24 illustrates the potential benefits of exploring novel therapeutic strategies, as the researchers noticed a significant 25 reduction in BPD (Ley et al., abstract presented at the EAPS 2016 congress).

In order to develop innovative therapeutic strategies to improve the chronic respiratory outcome of survivors of preterm birth, the knowledge on BPD pathophysiology needs to increase. On this point the limitations of exploratory research on humans become clear. Research on biomaterial of human cases is limited to what can be easily obtained: mostly serum [18, 19] or tracheal aspirates, bronchoalveolar lavage fluid or exhaled air [20-22]. While studies on lung tissue of patients with BPD would be able to provide important information, they are 31 understandably rare (especially in the post surfactant era) and mostly represent the lethal end of the disease 32 spectrum [23-26]. Even though they are interesting for biomarker identification [27] and the generation of new 33 hypotheses, studies on human material often do not prove causality. Therefore, researchers employ animal models 34 as they provide the ability to use a plethora of experimental techniques to measure, control and influence relevant 35 factors. A good animal model can increase the understanding of pathophysiology of a disease and can enable the 36 development of novel therapeutic strategies.

37 However the translation of therapeutic findings in animal models have been proven difficult in the past [28] 38 (recently illustrated by a clinical trial where the use of docosahexaenoic acid (DHA) supplements failed to reduce 39 BPD [29, 30]). Therefore there is still a need to improve our models for this disease. In this review we will discuss 40 the characteristics of an 'ideal BPD model'. We will first describe the factors contributing to the development of 41 human BPD which can be used as insults in animal models, and we will then discuss the phenotype of the disease 42 which should be mimicked. We will subsequently outline the characteristics and practicalities of existing animal 43 models. Finally we will make the case for the preterm rabbit model, as this model provides a good balance between 44 translational and practical considerations. In the last few years, many excellent reviews on animal models for BPD 45 have been published [28, 31-38]. In this review we look at this subject from a different perspective: we focus on 46 the utility of the different models (and more specifically the rabbit model) for preclinical drug evaluation.

#### Translating the pathophysiology of BPD into animal models 47

48

49 Bronchopulmonary dysplasia is a unique disease as it is not defined by its pathophysiology but by its treatment. This disorder is currently defined as 'the requirement for oxygen therapy on the 28<sup>th</sup> postnatal day or 36<sup>th</sup> week 50 51 post conception'. As this is a very 'clinical definition', it is no wonder that it invokes increasing critique in this 52 age of 'omics' and systems biology [39]. The phenotype of BPD is very heterogeneous and reflects the complex 53 interplay of individually varying insults on the developing lung, which goes through a critical adaptation process 54 [40]. This makes BPD, in contrast to for instance a genetic surfactant deficiency, particularly hard to mimic in an 55 animal model. Additionally the phenotype and pathophysiology of BPD has changed significantly over time. 56 Where the so-called 'old BPD' was a fibrotic disease, occurring in moderately preterm infants exposed to 57 aggressive respiratory support, 'new BPD' is described as a developmental arrest, that occurs in extremely preterm 58 infants benefiting from antenatal steroids, surfactant and minimal invasive supportive measures [41]. This 59 spectrum continues to evolve, in parallel with recent advances in perinatal care. In the following paragraphs we 60 will, based on recent data, review the factors that are involved in the etiology of BPD, as encountered by

61 neonatologists today. These factors could be used as insults to create experimental BPD in animal models.

#### 62 <u>Prematurity</u>

The primary and essential factor in the development of human BPD is premature birth. The incidence of BPD depends strongly on gestational age and birth weight. A recent follow-up study revealed an incidence of about 75% in a population of periviable infants (survivors of birth at 22-24 weeks of gestation) [42]. Another dataset revealed an incidence of moderate to severe BPD in 69% of infants born in the 24<sup>th</sup> week of gestation, while it only occurred in 24% of the infants born in the 28<sup>th</sup> week [3]. In general, incidences of BPD tend to vary between centers (depending on the definitions being used and the age limits of the study population) with the common relationship of increasing incidence of BPD with decreasing gestational age and birth weight [41, 43].

70 Babies at the highest risk for BPD (22-28 weeks of gestation) are born in the late canalicular to early saccular stage 71 of lung development (figure 1) [44]. While in healthy pregnancies the saccular stage proceeds into alveolarization 72 in utero, both pre- and postnatal factors disturb this developmental program in preterm birth (figure 2). It thus 73 seems sensible to favor an animal model in which birth occurs in the same late canalicular to early saccular stage 74 of lung development (figure 1).

75 Besides structural prematurity of the lung, it is logical to also take functional prematurity into account. For 76 example, the surfactant system develops during the saccular stage of lung development, lowering alveolar surface 77 tension and thus allowing expansion of the lung at birth. Preterm neonates are thus at birth at least partially deficient 78 for surfactant, and therefore often exhibit signs of respiratory distress or failure (so called respiratory distress 79 syndrome (RDS)) [45]. Below we will also discuss the antioxidant defense enzymes, which mature in parallel with 80 the surfactant system, as do probably many other pathways, to prepare a fetus for its pulmonary adaptation at term 81 birth. The lack of this preparation in the preterm neonate, requires endogenous adaptive strategies and medical 82 interventions (antenatal steroids, surfactant administration and respiratory support), which allow for survival by 83 facilitating the transition to postnatal life. However this process alters the normal development of the lung at longer 84 term. An interesting concept is to look at BPD as the adaptation of a functionally preterm lung to the new reality 85 of postnatal life [40]. This adaptation seems especially crucial in new BPD, where additional iatrogenic insults are 86 often non-aggressive and tapered to individual needs, but many preterm birth survivors still develop BPD. We 87 therefore hypothesize that the preterm animal models, as they allow to study the long term respiratory 88 consequences of preterm birth, have an important translational advantage.

#### 89 Insults to the developing lung

90 Superimposed on this adaptation of the preterm lung to postnatal life, a variety of endogenous and exogenous 91 factors interferes with lung development through injury and consecutive repair (summarized in figure 2) [40]. A 92 complete review of BPD related noxiae is beyond the scope of this text and we will limit ourselves to the major 93 contributors.

94 Oxygen

95 Signs of oxidative injury are a frequently reported finding in samples of BPD neonates: e.g. protein and lipid 96 peroxidation in exhaled gases and bronchoalveolar lavage fluid [51-53]. This form of injury can in the preterm 97 neonate be seen as the result of an imbalance between antioxidant defense mechanisms on the one hand and on the 98 other hand reactive oxygen species (ROS) overproduction due to hyperoxia, mechanical and infection.

99 During prenatal life the oxygen exposure in the lungs is low. Oxygenated blood enters the fetal circulation through 100 the umbilical vein delivering a partial oxygen pressure of about 20mmHg to the fetal lung through the vasculature. 101 After birth, the lungs are filled with air, resulting in alveolar partial oxygen pressures of 100mmHg even in normal 102 room air (a factor 5 increase in oxygen exposure). Term neonates seem to respond to this marked increase with an 103 upregulation of antioxidants and oxygen scavenging enzymes, especially in conditions with high oxidative stress 104 [46, 47]. Animal data suggest that these high enzyme levels are the result of a preparatory process during the final 105 part of gestation [48]. Preterm birth will thus untimely expose unprepared lungs to room air, which already contains 106 a supraphysiological concentration of oxygen. As preterm neonates appear to have weaker antioxidant defense 107 mechanisms than term babies [49-51], this might lead to oxidative damage even in the absence of inspired oxygen 108 fractions (FiO<sub>2</sub>) above 21%.

109 Raising the fraction of inspired oxygen (FiO<sub>2</sub>) above 21% room air - often crucial for maintaining viable arterial 110 oxygen pressures –might further increase the amount of ROS and subsequent oxidative damage in the developing 111 lung. ).Studies published in the early 2000's, have revealed an excess of BPD in patients where supplementary 112 oxygen was administered to target oxygen saturations above 95% [52-54]. This has led to the current practice to 113 target lower oxygen saturation levels (90-94%). More recent trials have gone even further in lowering saturation 114 targets to 85-89%, thereby significantly decreasing the FiO<sub>2</sub> to which babies were exposed. This resulted in 115 increased mortality, and a decreased ROP rate, but interestingly no further decrease in BPD prevalence was noted 116 in comparison to the higher target group (90-94%) [55, 56]. These findings pare down the role of oxygen supplementation as the most important contributor to the pathophysiology of today's BPD, where extremesaturation targets of >95% are no longer standard of care.

119 This is in contrast to many neonatal animal models, in which solely the exposure to excessive  $FiO_2$  has been used 120 to create a damage reminiscent of human BPD, making use of different timings and intensities [31-33, 36]. 121 Recently a robust study compared different oxygen exposure protocols in neonatal mice, highlighting advantages 122 and disadvantages of each protocol [57]. Generally, in these models the exposure to very high oxygen 123 concentrations results in comparable oxidative damage and subsequent interference with lung maturation, as seen 124 in infants developing BPD [58-61]. However, due to the lack of prematurity and other factors contributing in 125 oxidative damage (for instance ventilation or infection), the mechanisms leading to this phenotype in term 126 hyperoxic animal models might be different.

127 Ventilation

Another intervention, which might add damage to the already abnormal adaptation of the preterm lung to postnatal life, is mechanical ventilation. In a large preterm birth cohort of infants born between 25 and 28 weeks of gestation, 83% required some form of mechanical ventilation. Even when CPAP was the initial treatment modality, 43% of the patients required switch-over to mechanical ventilation at some point [62]. Mechanical ventilation thus remains a very frequent intervention, and ventilator induced lung injury is therefore thought to be an important contributor to BPD [63].

The injury induced by mechanical ventilation is probably caused by strain and distention, more than by pressure itself. Volutrauma thus seems a more correct denominator than barotrauma [64, 65]. In preterm infants the presence of marked differences in ventilation within the lung, adds on even more strain through atelectrauma and unequal distribution of distention [63, 64]. Overall, excessive distention of the parenchymal tissue leads to disruption of the epithelium and increased permeability of the alveolo-capillary membrane, exudation of proteins, release of cytokines and enzymes, and subsequently an inflammatory reaction [66, 67]. Even in the absence of increased FiO<sub>2</sub> the stretch of lung tissue might also lead to the formation of ROS [68, 69].

141 It is also established that the avoidance of mechanical ventilation in preterm infants reduces BPD prevalence, albeit 142 only modestly. A meta-analysis of 4 clinical trials comparing the use of nasal continuous positive airway pressure 143 (nCPAP) and intubation as initial respiratory strategy revealed that one additional infant could survive to 36 weeks 144 without BPD for every 25 neonates intended to be treated with nCPAP instead of intubation and mechanical 145 ventilation [70]. An explanation for the low effect size, might be the high rate of nCPAP-failures in the very immature lung and the need for cross-over to conventional ventilation. The modest effect of nCPAP is in contrast
to earlier observational data on the role of mechanical ventilation, as lung immaturity is an important confounding
factor here [71]. One might thus hypothesize that being born with very immature lungs, necessitating mechanical
ventilation, is more essential in the pathophysiology of BPD than mechanical ventilation itself. This point is also
supported by data which show that also preterm neonates who never required mechanical ventilation, develop BPD
(18,5% of neonates born between 25 and 28 weeks of gestation in group of nCPAP successes) [62].

In baboons and lambs researchers have been able to combine extreme lung immaturity with mechanical ventilation to create experimental BPD [34, 38]. Both invasive and non-invasive strategies of respiratory management have been used successfully in these larger animal models [72, 73], accurately mimicking current practice in neonatology. Term small animals have also been ventilated conventionally for shorter periods of time to study the effects of volutrauma on the developing lung, however translation to the modern human disease - in an era of increasingly immature babies and less invasive supportive measures - is obviously more difficult [74, 75].

#### 158 Other postnatal factors

Besides the exposure to supplemental oxygen and mechanical ventilation, many other postnatal factors add to the developmental arrest occurring when preterm lungs are exposed to a postnatal environment. Postnatal infections for instance (pneumonia, sepsis) will increase inflammatory damage in the lung, and determine the long term respiratory outcome [76, 77]. There also seems to be a relationship between malnutrition and BPD [78] or fluid overload and BPD [79]. Although controversial some authors also suggest a link between patent ductus arteriosus and BPD [80]. These clinically relevant risk factors have occasionally been studied in animal models as well [81-84].

ELBW infants (at risk of BPD) are currently also exposed to surfactant and caffeine treatment, and most of them have had a course of antenatal steroids as well, since these interventions are all part of the current standard of care. It is likely that the widespread availability of these interventions has changed the phenotype of BPD, in terms of survival of more preterm children and less-invasive supportive requirements ("new BPD" [39, 41]), but maybe also in terms of altered physiology [85-87]. In the context of drug development, these current pharmacological therapies, might also affect the pharmacokinetics and –dynamics of an investigated drug. The addition of these standard of care interventions could thus be advantageous for a preclinical BPD model.

173 Prenatal factors

There is increasing interest in prenatal biomarkers for the early prediction of BPD [27], as prenatal factors determine the lung (im)maturity at birth, and subsequently the long term respiratory outcome. Previously, it has been suggested that the absence of pregnancy related complications in many animal models, explains the difficult translation of findings to human BPD [28].

*Prenatal infection or chorioamnionitis* is a frequent cause of preterm birth and seems to be associated to BPD [88].
In utero inflammation might interfere with normal lung development, however its direct effect, independent of
gestational age remains unclear [77]. In animal models the effect of prenatal inflammation on lung development
has been studied by injecting E. Coli lipopolysaccharides or Ureaplasma organisms in the amniotic sac [89, 90].

When normal *functioning of the placenta* is disturbed, growth and development of the fetal lung will be affected.
Associations between pre-eclampsia, intrauterine growth retardation (IUGR) and BPD have been described [88,
91]. An interesting finding is also the association between placental vascularity and pulmonary vascular disease in
BPD [92]. In animal models, this state of decreased angiogenesis has been mimicked by intra-amniotic injections
with an antagonist of vascular endothelial growth factor (VEGF) [93].

187 There has been increasing interest in *maternal smoking*, as a risk factor for the development of BPD. Smoking 188 during pregnancy interacts with BPD in a complex manner, as it is both associated to preterm delivery and impaired 189 lung development [94]. In a Canadian cohort a significant increase in BPD was noted in preterm babies of which 190 mothers smoked during pregnancy [95]. Maternal smoking has also been used as an insult in animal models, to 191 create a BPD-like phenotype [96, 97].

192 The genetic constitution might also predispose to more immature and underdeveloped lungs at the start of life (e.g. 193 polymorphisms in surfactant proteins or VEGF), but might also increase the susceptibility to postnatal damage 194 (e.g. polymorphisms in scavengers of ROS) [98]. On the other hand the genetic constitution might also determine 195 the response to an (experimental) treatment. It seems therefore important that a translational drug evaluation occurs 196 in an animal model reflecting the genetic heterogeneity of human newborns, which is not the case in inbred strains. 197 On the other hand, for basic explorative research, the genetic homogeneity can be an advantage as it increases 198 reproducibility. The importance of the genetic background is also nicely illustrated by the variation in responses 199 of adult mice from different strains to hyperoxia [99].

#### 200 Mimicking the BPD phenotype in animal models

201

By using comparable insults, animal models should be able to mimic the phenotype of BPD. In this paragraph we
will describe the histological, functional and vascular manifestations of the human disease, and the options to study
this in animal models.

#### 205 <u>Histology</u>

206 The histological hallmark of BPD is impaired alveolar and vascular development, with a variable degree of 207 extracellular matrix remodeling. Being the only extensive histological evaluation in the post surfactant era (yet 208 before antenatal steroids were introduced), Husain et al. found inhibition of acinar development (lower radial 209 alveolar count (RAC) and increased mean linear intercept (Lm)) in 22 BPD autopsy specimens, compared to 15 210 lungs of age-matched controls [100]. The question remains whether findings in autopsies can represent the 211 structural changes in BPD survivors. Nevertheless, a study of lung biopsy specimens from surviving BPD infants 212 (without surfactant or antenatal steroids) showed comparable enlarged air spaces with simplified alveoli and 213 variable alveolar wall cellularity or fibrosis [26]. Since those studies have been published, care in neonatology has 214 evolved significantly, changing the phenotype of BPD. For understandable reasons there are no recent reports on 215 biopsy specimens of BPD infants which benefited from antenatal steroids, non-invasive ventilation strategies and 216 other current practices. We could only identify a few more recent autopsy series reports, all of them with a very 217 specific scope (mainly vascular manifestations of BPD) [24, 25, 101, 102]. Despite the gaps in knowledge on 218 current human BPD histology, animal models are still generally accepted if they display an alveolar developmental 219 arrest, with increased septal thickness, as described in the aforementioned historical case series.

220 Impaired alveolarization can be quantified in experimental animals by measuring alveolar size, surface area and 221 number, with appropriate unbiased stereological techniques after correct processing of the tissue [103]. The most 222 reported measure is probably the mean linear intercept (Lm), reflecting the mean length of a random line between 223 to gas exchange surfaces. Increased Lm as shown by Husain et al. in BPD autopsy specimens has been reproduced 224 in many animal models [57, 104-107]. Other none stereological measurements can complete the picture: radial 225 alveolar count (RAC), septal crest counting and acute lung injury score [108-110]. Manual counting of 226 morphometric parameters is a time consuming effort, and is prone to bias, certainly if not properly blinded. 227 Computer-aided morphometry can make analysis less labor intensive and biased, however some evaluations might 228 still require a trained eye [111, 112]. Alternatively, direct 3D assessment of alveolar architecture by means of 229 micro CT and subsequent automated analysis will gain importance in the future [113, 114].

Another interesting finding in the limited histological data, is the disorganization of the elastin fiber network [115].

231 Quantitatively more elastin is present, but the fibers occur more tortuous and less organized [116]. This is

particularly interesting because of elastin's key role in the process of secondary septation [117, 118], as a
dysfunctional elastin matrix might explain the alveolar developmental arrest described above. Furthermore,
because of elastin's function in tethering of the airways [119], this finding could also clarify the lung function
abnormalities mentioned below.

#### 236 *Lung function*

237 Pulmonary function testing has been performed in short- and long-term studies of BPD, applying a variety of 238 techniques [120]. The major finding in these studies is that BPD patients depict signs of obstructive airway disease. 239 Classical spirometry showed decreased forced expiratory volumes (FEV) and increased responsiveness to 240 bronchodilation in older children surviving BPD [121]. In neonates developing BPD, more complex techniques 241 have to be used, however the same conclusions are reached: at both single breath occlusion and forced oscillation 242 increased airway resistance is noted [122, 123]. Additionally higher functional residual capacities and residual 243 volumes are measured with body plethysmography and helium dilution, suggesting hyperinflation and air trapping 244 [120, 124, 125]. These findings suggest an important factor of small airway pathology in BPD, which is often 245 neglected in research.

Besides airway disease, preterm (ventilated) infants developing BPD also have lower lung compliance [122, 124], indicating increased stiffness of the lung in the acute phase of their disease. A final functional characteristic of the BPD lung is a decreased gas exchange efficiency. Diffusion capacity measurements reveal a reduction of both pulmonary membrane diffusing capacity and pulmonary capillary blood volume [126, 127], which supports the paradigms of impaired alveolarization (see above) and vascular maldevelopment (see below).

251 Evaluation of lung function in experimental animal BPD, revealing changes in line with observations in humans, 252 can thus improve the strength of the model. Measuring lung functions in animals can be challenging, and there are 253 often parallels between the techniques used for non-cooperative neonates and laboratory animals [128]. Invasive 254 forced ventilatory maneuvers (pressure-volume loops and recruitment maneuvers) are commonly used to look for 255 decreased lung capacities and compliance in BPD models [129, 130], however do not provide information on 256 airway obstruction. Forced oscillation is therefore often added, to estimate airway (and tissue) resistance [129, 257 130]. An alternative is to add adapted harnesses or negative pressure chambers, to measure forced expiratory 258 volume and flow [131], however this technique has, by our knowledge not been applied in neonatal animals until 259 now. Both techniques can also be performed after methacholine instillation, to assess the hyperreactivity of the 260 airways [130]. A final addition can be gas dilution and diffusion techniques: by using those researchers have

revealed changes in diffusion or residual capacities in line with human observations [107, 132]. On the other hand,
there are non-invasive measurements of lung function of which plethysmography has been used in many animal
models of BPD [133-135].

#### 264 Pulmonary vascular disease

265 About 14% of BPD-infants has pulmonary hypertension, as diagnosed on ultrasound at 36 weeks post conception 266 [4], however they are thought to be only the worst end of a spectrum. Probably all BPD infants have pulmonary 267 vascular disease in more subtle forms [4, 136], reflecting the close interaction between acinar and vascular 268 development [104, 137, 138]. Clinical echocardiographic data demonstrate a high incidence of both early and late 269 pulmonary hypertension in BPD and an association with BPD severity [4, 139]. In addition, Thibeault et al. studied 270 arterial histopathological changes in autopsy specimens from infants with severe BPD and found at the level of 271 terminal bronchioles an increased intra-acinar arterial wall thickness due to muscularization [102]. Autopsy and 272 biopsy specimens from BPD infants also demonstrated a dysmorphic microvascular pattern with variable 273 abnormalities in the pulmonary capillaries [101, 140].

274 It thus seems essential that an animal model for bronchopulmonary dysplasia also exhibits pulmonary vascular 275 disease (functional or structural signs of pulmonary hypertension and/or a dysmorphic capillary bed). A higher 276 medial thickness index and a decreased peripheral vascular density have been described consistently in BPD-277 phenotypes in mice [141-143], rats [104, 144], rabbits [145], lambs [146] and baboons [106]. Also in animal 278 models pulmonary hypertension can be evaluated by echocardiography. Right ventricular free wall thickness, 279 pulmonary artery acceleration time (PAAT) and pulmonary artery acceleration time/pulmonary artery ejection 280 time ratio (PAAT/PAET) have been used as surrogates for pulmonary artery pressure in small BPD animal models 281 [141, 142, 147, 148]. Good correlation between these surrogates and invasive measures by catheterization have 282 been reported [145, 149]. Finally, also micro CT angiograms after the (lethal) injection of barium in the pulmonary 283 artery have provided interesting data on the 3D structure of the pulmonary arterial tree in experimental BPD [104, 284 141, 145].

286

#### 285 Overview of the available animal models

287 Multiple animal models have been developed to investigate the pathophysiology of BPD. Each animal model has
288 a unique set of translational and practical advantages and limitations which should be considered carefully in the
289 context of preclinical research on BPD.

#### 290 <u>Small animal models: mice and rats</u>

291 Mice and rats have often been used for BPD research and have well established models that use term neonates, in 292 whom damage is induced by hyperoxia, hypoxia, prenatal inflammation, transgenesis and even short term 293 ventilation [31, 33] (table 1). The animals are small (birth weight mice: 0.5–1.5 g and rats: 5-6g), cheap to house 294 and have a short gestation (21 days for mice, 22 days for rats) with a large litter size making experiments relatively 295 easy to perform and plan. The major advantage however of the mouse models (and to a lesser extent rats) is the 296 availability of reagents and technology for extensive pathway analysis: e.g. (conditional) gene knock-out strains; 297 validated antibodies for flow cytometry, western blot or immunohistochemistry; transcriptomics, proteomics and 298 advanced microscopic techniques [150, 151]. At term both mice and rats are in the saccular phase of lung 299 development, which is morphologically similar to human premature infants at 28-30 weeks gestation (figure 1). 300 This means that they are good models to study alveolarization, but not necessarily the transition of the canalicular 301 to saccular stage as occurring in human preterm babies developing BPD. Additionally these pups do not encounter 302 the gas exchange limitations seen in premature infants. Rats and mice seem to have developed adaptive 303 mechanisms to function normally, with structurally preterm lungs (e.g. they express sufficient levels of surfactant 304 and antioxidant defense enzymes [48, 152]). Truly preterm mice and rat models have not been described so far. 305 Moreover the small size of the pups limits the possibility of performing long term monitored mechanical 306 ventilation experiments. Overall the term mice and rats provide good models for basic explorative research and 307 pathway analysis of specific mechanisms involved in the pathogenesis of human BPD. Resulting insights are 308 essential in the conception of innovative treatment ideas (a good example is the research on the relationship 309 between angiogenesis and alveolarization in hyperoxia [104]). However, the lack of prematurity, and the need for 310 insults exceeding what is common practice in neonatal medicine, questions the translatability of findings in mice 311 and rats to current human BPD.

#### 312 Large animal models: baboons and lambs

Large animal models in the field of BPD research are the premature lamb and the premature baboon model. The lung development of lambs and especially baboons reflects human lung development closely, as they start alveolarization before term birth, and the length of the developmental stages is proportionate to the length in humans (figure 1). The animals can be delivered prematurely and have respiratory failure as seen in human preterms, necessitating mechanical ventilation or non-invasive support measures [72, 73]. Standard of care interventions (exogenous surfactant, antenatal steroids, titrated oxygen supplementation and caffeine) have been 319 added to the models. Their size (mean 382g for preterm baboons at day 125, mean 3kg for preterm lambs at day 320 131), also allows for intensive monitoring along with repeated blood sampling and catheterization, simulating 321 nicely the clinical neonatal intensive care setting [34, 38]. These models thus reproduce the human disease and 322 context very closely. An additional advantage is that, similar to human BPD, signs of obstructive airway disease 323 have been well described. The ability to closely mimic the airway pathology in BPD, can be explained by the 324 similarities between human and large animal airway anatomy (the presence of respiratory bronchioles, the presence 325 of cartilage in the intrapulmonary airways and symmetrical splitting of airways) [34, 38, 157]. On the other hand 326 the intensity of care and the high cost involved are major disadvantages. The scientific community and the public 327 might also have ethical constraints with large animal experiments, especially with non-human primates, however 328 these considerations should also be taken in to account when using smaller animals. Furthermore the availability 329 of reagents is rather limited, when compared to research in the murine models. A final disadvantage is translational, 330 as most models are created by cesarean section in a formerly healthy pregnancy. This does not parallel most human 331 preterm births, where prenatal pathology leads to preterm birth, and predisposes the lung towards BPD. In both 332 baboons and sheep the influence of perinatal infection or IUGR in the etiology of BPD has also been studied [90, 333 156, 158] (table 1), and one could argument to include these prenatal factors in preclinical studies evaluating 334 therapeutic options for BPD.

335 Many slightly different variations on these models have been published. The preterm baboon model has closely 336 followed the evolving clinical practice in neonatology, with both a model for old BPD (moderately preterm animals 337 delivered at day 140, supported with aggressive mechanical ventilation) and new BPD (severely preterm animals 338 delivered at day 125, supported with surfactant administration and non-invasive ventilation). Preterm (ventilated) 339 lambs between 125 and 132 days of gestation, have been used by many groups for a wide range purposes: to study 340 the effects of short term in utero ventilation [153], to study respiratory stability during different ventilation 341 modalities [154] and recently also for trials with an ex vivo uterine environment [155]. Fewer groups however 342 have in sheep focused on the effect of interventions on longer term BPD related outcomes, mainly in models born 343 around day 125 [38]. It is important to note that there is only a narrow time window to mimic the human setting 344 of birth in the early saccular stage of lung development, as sheep have a relatively short saccular stage, and around 345 day 130 alveolarization has already started in many of them (figure 1).

346 <u>Rabbits</u>

347 Rabbits are medium sized animals that have the main advantage that their lung development closely mirrors that 348 of humans. In contrast to smaller rodents, alveolarization starts in-utero in rabbits (figure 1). They also have large 349 litter sizes, relatively short gestations (31 days), are easy to house and have a size that allows for certain technical 350 manipulations (30-60g) [32]. Many different insults have been used to mimic BPD in term rabbit models, including 351 different hyperoxia protocols [159-161], short term ventilation [162] and chorioamnionitis [163]. However, term 352 rabbit pups are in the alveolar phase of lung development, and thus lack both structural and functional prematurity 353 (table 1). The major asset of the rabbit over the smaller rodent models, is the possibility to induce prematurity 354 [105, 129]. Below we will discuss the preterm rabbit model in more detail.

355 356

### The case for a preterm rabbit model

357 The stage for the development of a preterm rabbit model for BPD has been set in the 1980s. At that time researchers 358 could show a doubling in fetal lung antioxidant enzyme levels during the last 3-5 days of normal term gestation in 359 rabbits [164], as well as in other laboratory animal species [48]. This physiological increase was hypothesized to 360 prepare the fetuses for the upsurge in oxygen pressure related with a normal fetal to postnatal life transition at term 361 birth. Consequently, they could demonstrate that preterm rabbit pups (day 29) had more severe lung injury in 362 response to 48-72h of hyperoxia (FiO<sub>2</sub>>90%) than term pups. Preterm pups were, besides their uncompleted fetal 363 maturation, also unable to upregulate their antioxidant enzymes in defense to a hyperoxia insult [165]. These 364 findings illustrate that prematurity is a major contributor to neonatal lung disease, and emphasize the functional 365 prematurity of the preterm rabbit models. Additionally, the fetal increase in antioxidant levels in rabbits coincides 366 with a steep maturation of the surfactant system and a swift development of the fetal lung from late canalicular, 367 over saccular, to the alveolar stage (figure 1) [166, 167]. Preterm rabbits are thus born in a comparable stage of 368 lung development as human preterms developing BPD, and have a comparable immaturity of the antioxidant and 369 surfactant systems.

Additional injury to the developing lung of the preterm rabbit, has mainly been administered under the form of hyperoxia. Initial studies with the preterm rabbit model, were limited to short term exposures (FiO<sub>2</sub> 90-100%, during 1-4 days), mainly using pups after 29 days of gestation [165, 168, 169]. These short-term experiments, focused on acute consequences of hyperoxia (lipid peroxidation, pulmonary edema and inflammation), and did not evaluate the effects on architectural changes in the lung related to lung development. The first longer term experiment was published by Mascaretti *et al.* in 2009. In pups delivered at 28 days of gestation and exposed to FiO<sub>2</sub>>95% for 11 days, they could demonstrate an arrested lung development, however survival was extremely low (11% in hyperoxia, 31% in normoxia) [170]. In later work they showed that the developmental arrest is
preserved in less preterm pups (29 days of gestation), exposed to lower oxygen concentrations (FiO<sub>2</sub> 80%),
however mortality remained high [105]. Furthermore the additional effect of postnatal malnutrition on the
pulmonary phenotype of the model was explored [83].

Our group developed a comparable preterm rabbit model for BPD (figure 3) [129]. Preterm rabbit pups (New Zealand White and Dendermonde hybrids) are delivered by cesarean section after 28 days of gestation. Pups are hand raised in either hyperoxia (FiO<sub>2</sub>>95%) or normoxia (FiO<sub>2</sub> 21%) in an incubator at a constant temperature (32°C) and humidity (75%). Standardized gavage-feeding is administered to all pups, eliminating the variability associated with maternal feeding: a milk formula adapted for neonatal rabbits (30% protein, 50% fat), supplemented with colostrum and probiotics is administered twice daily. Pups are also injected prophylactically with vitamin K and antibiotics. After 7 days all outcome measures are obtained and lungs are harvested.

388 Hyperoxia exposed preterm rabbit pups exhibit morphological, functional and vascular manifestations comparable 389 to human BPD. Morphologically signs of alveolar simplification and developmental arrest are present: increased 390 alveolar size (Lm) and increased thickness of the alveolar septa. Furthermore increased amounts of collagen, but 391 not elastin, and increased numbers of proliferating cells were noted when compared to normoxia preterm controls 392 [129, 170]. Signs of increased inflammation were also present [171]. Functionally the lungs of the hyperoxia 393 exposed pups had a lower total lung capacity, decreased compliance and increased tissue resistance (tissue 394 damping). Signs of obstructive airway disease, as seen in humans, were not observed [129], however no 395 provocation studies have been undertaken to study bronchial hyperreactivity. At the vascular level increased 396 arterial wall thickness has been observed, together with signs of pulmonary hypertension (decreased PAAT/PAET 397 ratio) at cardiac echo Doppler [145].

398 The preterm rabbit model combines some crucial advantages of both the large and small animal models. First, its 399 major asset is without doubt that it consists of premature animals (both structurally and functionally), that need to 400 go through a precocious adaptation process to postnatal life, even without the addition of extra insults. The preterm 401 pups at day 28 depict signs of respiratory distress at birth, however can survive without surfactant administration 402 or mechanical ventilation. Second, unlike other preterm animal models they are easy to house and handle, have 403 large litter sizes, short gestations and relatively low cost, facilitating many practical aspects of research. Third, the 404 size of the pups allows for certain technical manipulations (tracheostomy for ventilation and surfactant 405 administration, cardiac ultrasound, gavage feeding). Long term ventilation has not been described, yet many

406 investigators have evaluated the effect of exogenous surfactant administration on ventilation of preterm pups407 delivered after 27 days of gestation [172, 173].

408 Obviously working with preterm rabbit also has disadvantages. The paucity of antibodies and absence of genetic 409 knock-outs limits in depth pathway analysis in rabbits, though measuring messenger RNA expression (qPCR, 410 RNA sequencing) can compensate partially [129]. Furthermore, the high mortality is often considered as an issue, 411 but workable survival rates of 56% in hyperoxia and 83% in normoxia at day 7 have been reported [129]. As often 412 only surviving pups can be included for outcome measures, it is important to note that this high mortality might 413 bias study results. Another drawback of the preterm rabbit model however is that mortality might fluctuate because 414 of maternal and seasonal influences that are often difficult to control for [174, 175]. On the other hand, the mortality 415 can also be considered as a translational advantage, as it illustrates the functional prematurity of the rabbit pups. 416 Finally, the lack of prenatal pathology leading to prematurity in this cesarean section based model, can also be 417 considered as a translational deficit.

418 Based on the arguments mentioned above, we consider the preterm rabbit model a valuable model for preclinical 419 drug research in BPD. Until now, the effect of caffeine and omeprazole on the phenotype have been assessed in 420 our model [174, 176]. An important consideration for drug research are the pharmacokinetics and 421 pharmacodynamics of the tested pharmaceuticals. When considering a preclinical study, it should be evaluated if 422 the translation of the findings is not hindered by important differences in absorption, distribution, metabolism or 423 excretion between the animal model and the clinical target population. Of course this depends on the specific 424 characteristics of every individual drug, but in general the rabbit bears some interesting similarities to the human. 425 In terms of absorption, it is mainly the variety in administration methods that mimics clinical practice. In the 426 smaller neonatal rodents the intraperitoneal route is often used as the only option for systemic drug delivery, 427 however many factors may determine (and limit) subsequent absorption [177]. The size of the rabbit pups allows, 428 besides intraperitoneal administration, for more translatable methods: either subcutaneous, intramuscular, 429 intravenous or enteral administration by gavage. Recently we also optimized a method for localized intratracheal 430 pulmonary delivery, which will importantly expand the applications of the model (unpublished work). In terms of 431 metabolism and excretion, it is important to notice that there are significant developmental differences between 432 the adult and (preterm) neonate [178]. As in humans, both phase I (e.g. cytochrome P450 monooxygenases) and 433 II (e.g. glucuronidation enzymes) hepatic drug metabolism mature over the lifespan of a rabbit [179-181]. Equally, 434 renal clearance of neonatal rabbits is very low, and increases with aging [182]. For instance, for caffeine, this 435 results in an accumulation of the drug and its metabolites in neonatal rabbits, as it is also the case in preterm infants [183]. In studies testing prenatal drug therapies, a final aspect of pharmacokinetics that should be considered is
placental transfer of the drug. The structure of the placental barrier of the rabbit (hemodichorial) is closer to the
human placenta (hemomonochorial in the third trimester), than the placenta of sheep (epitheliochorial) and rats or
mice (hemotrichorial) [184], making the rabbit a popular model for fetal toxicity studies [185]. Recent data also
suggest functional similarities in placental drug transporter spectra between humans and rabbits [186].
Transplacental treatment strategies in the preterm rabbit model have been carried out with rosiglitazone (PPARγagonist) and omeprazole (inducer of CYP1A1) [174, 175].

#### 443 Discussion

444

445 The purpose of this review is to discuss the boundaries and characteristics of an ideal animal model that both 446 mimics the morphological, functional and vascular manifestations of BPD and provides a good representation of 447 the human pathophysiology. Neonatology is a constantly changing field and this affects the utility of the different 448 animal models. Where aggressive iatrogenic insults (high FiO<sub>2</sub> and mechanical ventilation) were the cornerstones 449 of the so called "old BPD", this may no longer hold true for the BPD that neonatologists currently encounter. 450 Modern BPD remains a multifactorial disease, but the consistent common denominator in the whole disease 451 spectrum is extreme prematurity. We therefore advocate the use of preterm animals because of their translational 452 value.

453 Yet biomedical research always has to allow a certain degree of limitations in the models that it uses and as such 454 all sub-entities of the disease will probably never be covered by one single animal model. Research models in 455 general only incorporate stable and reproducible (mostly single and exaggerated) insults, while a variable plethora 456 of insults contribute to human preterm lung injury, making every case different. Novel pathophysiological insights 457 and derived therapeutic strategies from an animal model with a specific insult, might therefore only be applicable 458 to the subset of BPD cases resulting from a comparable etiology. We hypothesize that this explains why very few 459 therapeutic findings in animal models could be reproduced in humans. Precision medicine and the identification 460 of different BPD subtypes might improve translation of findings in experimental animals to clinical care in the 461 future [39].

We will thus conclude that a perfect animal model for bronchopulmonary dysplasia does not exist. However all models have their place in BPD research, and for every research question there is one that is ideal (figure 4). The small animal models of mice and rats are particularly suited for basic explorative research on pathways involved 465 in specific aspects of the disease, because of the availability of state of the art molecular techniques. On the other 466 hand larger animal models such are preterm primates and lambs provide a good platform for the translational study 467 of innovative therapeutic strategies in a setting similar to the neonatal intensive care unit. Practical and ethical 468 constraints however limit an extensive use, so only the most promising therapies can be tested out in these models. 469 In this review we focused on the preterm rabbit model. This is an elegant model that sits in between the small and 470 large animals and can be used for both explorative and translational research, as it is the smallest model that 471 combines structural and functional prematurity. The use of the appropriate animal model for each research 472 question, and the evaluation of preclinical study results in the context of the limitations of its model, will benefit 473 the development of novel therapeutic strategies for BPD.

474

475

## LIST OF ABBREVIATIONS

BPD	bronchopulmonary dysplasia
CYP1A1	cytochrome P450 monooxygenase 1A1
DHA	docosahexaenoic acid
ELBW	extremely low birth weight
FEV	forced expiratory volume
FiO <sub>2</sub>	fraction in inspired oxygen
HFV	high frequency ventilation
HFNV	high frequency nasal ventilation
IGF1	insulin-like growth factor 1
IUGR	intra-uterine growth retardation
Lm	mean linear intercept
(n)CPAP	(nasal) continuous positive airway pressure
(n)IPPV	(nasal) intermittent positive pressure ventilation
PAAT	pulmonary artery acceleration time
PAAT/PAET	pulmonary artery acceleration over ejection time ratio
PPARγ	peroxisome proliferator-activated receptor gamma
RAC	radial alveolar count
RDS	respiratory distress syndrome
ROS	reactive oxygen species

### **CONFLICT OF INTEREST**

The authors report no conflicts of interest.

J. D. is beneficiary of a fundamental clinical research grant of the Fonds Wetenschappelijk Onderzoek Vlaanderen (1801207). J. T. is supported by the Klinische Opleidings- en Onderzoeks-Raad of the University Hospitals Leuven, F. L. by Internal Funding of KU Leuven "Onderzoekstoelage" (OT/13/115) and A. G. by the European Commission via its Erasmus Joint Doctoral program (2013-0040).

### ACKNOWLEDGEMENTS

All individuals listed as authors have contributed substantially to the writing of this paper. T. S., A. G. and B. T. performed a literature study and wrote different parts of the text. T. S. merged the manuscript. J. T. and J. D. guided and corrected the work.

### REFERENCES

- 1. Blencowe, H., et al., *Born too soon: the global epidemiology of 15 million preterm births.* Reprod Health, 2013. **10 Suppl 1**: p. S2.
- Northway, W.H., Jr., R.C. Rosan, and D.Y. Porter, *Pulmonary disease following respirator* therapy of hyaline-membrane disease. Bronchopulmonary dysplasia. N Engl J Med, 1967.
   276(7): p. 357-68.
- 3. Stoll, B.J., et al., *Trends in Care Practices, Morbidity, and Mortality of Extremely Preterm Neonates, 1993-2012.* Jama, 2015. **314**(10): p. 1039-51.
- 4. Mourani, P.M., et al., *Early Pulmonary Vascular Disease in Preterm Infants at Risk for Bronchopulmonary Dysplasia.* Am J Respir Crit Care Med, 2014.
- 5. Khemani, E., et al., *Pulmonary artery hypertension in formerly premature infants with bronchopulmonary dysplasia: clinical features and outcomes in the surfactant era*. Pediatrics, 2007. **120**(6): p. 1260-9.
- 6. Hilgendorff, A. and M.A. O'Reilly, *Bronchopulmonary dysplasia early changes leading to longterm consequences.* Front Med (Lausanne), 2015. **2**: p. 2.
- 7. Synnes, A., et al., *Determinants of developmental outcomes in a very preterm Canadian cohort.* Arch Dis Child Fetal Neonatal Ed, 2016.
- 8. Johnson, T.J., et al., *Cost of morbidities in very low birth weight infants.* J Pediatr, 2013. **162**(2): p. 243-49.e1.
- 9. Jensen, E.A., E.E. Foglia, and B. Schmidt, *Evidence-Based Pharmacologic Therapies for Prevention of Bronchopulmonary Dysplasia: Application of the Grading of Recommendations Assessment, Development, and Evaluation Methodology.* Clin Perinatol, 2015. **42**(4): p. 755-79.
- 10. Darlow, B.A. and P.J. Graham, *Vitamin A supplementation to prevent mortality and short- and long-term morbidity in very low birthweight infants.* Cochrane Database Syst Rev, 2011(10): p. Cd000501.
- 11. Schmidt, B., et al., *Caffeine therapy for apnea of prematurity*. N Engl J Med, 2006. **354**(20): p. 2112-21.
- 12. Bassler, D., et al., *Early Inhaled Budesonide for the Prevention of Bronchopulmonary Dysplasia*. N Engl J Med, 2015. **373**(16): p. 1497-506.
- 13. Yeh, T.F., et al., *Intratracheal Administration of Budesonide/Surfactant to Prevent Bronchopulmonary Dysplasia*. Am J Respir Crit Care Med, 2016. **193**(1): p. 86-95.
- 14. Baud, O., et al., *Effect of early low-dose hydrocortisone on survival without* bronchopulmonary dysplasia in extremely preterm infants (PREMILOC): a double-blind, placebo-controlled, multicentre, randomised trial. Lancet, 2016. **387**(10030): p. 1827-36.
- Baud, O., et al., Association Between Early Low-Dose Hydrocortisone Therapy in Extremely Preterm Neonates and Neurodevelopmental Outcomes at 2 Years of Age. Jama, 2017.
   317(13): p. 1329-1337.
- 16. Lal, C.V. and N. Ambalavanan, *Genetic predisposition to bronchopulmonary dysplasia*. Semin Perinatol, 2015. **39**(8): p. 584-91.
- 17. Wang, H., et al., *A genome-wide association study (GWAS) for bronchopulmonary dysplasia.* Pediatrics, 2013. **132**(2): p. 290-7.
- 18. Baker, C.D., et al., *Cord blood angiogenic progenitor cells are decreased in bronchopulmonary dysplasia.* Eur Respir J, 2012. **40**(6): p. 1516-22.
- 19. Kim, D.H. and H.S. Kim, Serial changes of serum endostatin and angiopoietin-1 levels in preterm infants with severe bronchopulmonary dysplasia and subsequent pulmonary artery hypertension. Neonatology, 2014. **106**(1): p. 55-61.
- 20. Tramper, J., et al., *The Association between Positive Tracheal Aspirate Cultures and Adverse Pulmonary Outcomes in Preterm Infants with Severe Bronchopulmonary Dysplasia*. Am J Perinatol, 2017. **34**(1): p. 96-104.

- Vento, G., et al., Bronchoalveolar lavage fluid peptidomics suggests a possible matrix metalloproteinase-3 role in bronchopulmonary dysplasia. Intensive Care Med, 2009. 35(12): p. 2115-24.
- 22. May, C., et al., *Relation of exhaled nitric oxide levels to development of bronchopulmonary dysplasia.* Arch Dis Child Fetal Neonatal Ed, 2009. **94**(3): p. F205-9.
- 23. De Paepe, M.E., D. Greco, and Q. Mao, *Angiogenesis-related gene expression profiling in ventilated preterm human lungs.* Exp Lung Res, 2010. **36**(7): p. 399-410.
- 24. Galambos, C., S. Sims-Lucas, and S.H. Abman, *Histologic evidence of intrapulmonary anastomoses by three-dimensional reconstruction in severe bronchopulmonary dysplasia*. Ann Am Thorac Soc, 2013. **10**(5): p. 474-81.
- 25. De Paepe, M.E., et al., *Pulmonary dendritic cells in lungs of preterm infants: neglected participants in bronchopulmonary dysplasia*? Pediatr Dev Pathol, 2011. **14**(1): p. 20-7.
- 26. Coalson, J.J., *Pathology of new bronchopulmonary dysplasia*. Semin Neonatol, 2003. **8**(1): p. 73-81.
- 27. Rivera, L., et al., *Biomarkers for Bronchopulmonary Dysplasia in the Preterm Infant.* Front Pediatr, 2016. **4**: p. 33.
- 28. Jobe, A.H., *Animal Models, Learning Lessons to Prevent and Treat Neonatal Chronic Lung Disease*. Front Med (Lausanne), 2015. **2**: p. 49.
- 29. Collins, C.T., et al., *Docosahexaenoic Acid and Bronchopulmonary Dysplasia in Preterm Infants.* N Engl J Med, 2017. **376**(13): p. 1245-1255.
- 30. Ma, L., et al., *Arginyl-glutamine dipeptide or docosahexaenoic acid attenuate hyperoxiainduced lung injury in neonatal mice.* Nutrition, 2012. **28**(11-12): p. 1186-91.
- 31. O'Reilly, M. and B. Thebaud, *Animal models of bronchopulmonary dysplasia. The term rat models*, in *Am J Physiol Lung Cell Mol Physiol*. 2014, Copyright (c) 2014 the American Physiological Society. p. L948-I958.
- 32. D'Angio, C.T. and R.M. Ryan, *Animal models of bronchopulmonary dysplasia. The preterm and term rabbit models*, in *Am J Physiol Lung Cell Mol Physiol*. 2014, Copyright (c) 2014 the American Physiological Society. p. L959-I969.
- 33. Berger, J. and V. Bhandari, *Animal models of bronchopulmonary dysplasia. The term mouse models.* Am J Physiol Lung Cell Mol Physiol, 2014. **307**(12): p. L936-47.
- 34. Yoder, B.A. and J.J. Coalson, *Animal models of bronchopulmonary dysplasia. The preterm baboon models.* Am J Physiol Lung Cell Mol Physiol, 2014. **307**(12): p. L970-7.
- 35. Ambalavanan, N. and R.E. Morty, *Searching for better animal models of BPD: a perspective.* Am J Physiol Lung Cell Mol Physiol, 2016: p. ajplung.00355.2016.
- 36. Nardiello, C., I. Mizikova, and R.E. Morty, *Looking ahead: where to next for animal models of bronchopulmonary dysplasia?* Cell Tissue Res, 2016.
- 37. Silva, D.M., et al., *Recent advances in the mechanisms of lung alveolarization and the pathogenesis of bronchopulmonary dysplasia*. Am J Physiol Lung Cell Mol Physiol, 2015. **309**(11): p. L1239-72.
- 38. Albertine, K.H., *Utility of large-animal models of BPD: chronically ventilated preterm lambs.* Am J Physiol Lung Cell Mol Physiol, 2015. **308**(10): p. L983-I1001.
- 39. Day, C.L. and R.M. Ryan, *Bronchopulmonary dysplasia: new becomes old again!* Pediatr Res, 2017. **81**(1-2): p. 210-213.
- 40. Jobe, A.H., *Mechanisms of Lung Injury and Bronchopulmonary Dysplasia*. Am J Perinatol, 2016. **33**(11): p. 1076-8.
- 41. Jobe, A.H., *The new bronchopulmonary dysplasia*. Curr Opin Pediatr, 2011. **23**(2): p. 167-72.
- 42. Younge, N., et al., *Survival and Neurodevelopmental Outcomes among Periviable Infants.* N Engl J Med, 2017. **376**(7): p. 617-628.
- 43. Jensen, E.A. and B. Schmidt, *Epidemiology of bronchopulmonary dysplasia*. Birth Defects Res A Clin Mol Teratol, 2014. **100**(3): p. 145-57.
- 44. Langston, C., et al., *Human lung growth in late gestation and in the neonate*. Am Rev Respir Dis, 1984. **129**(4): p. 607-13.

- 45. Carnielli, V.P., et al., *Pulmonary surfactant kinetics of the newborn infant: novel insights from studies with stable isotopes.* J Perinatol, 2009. **29 Suppl 2**: p. S29-37.
- 46. Lurie, S., et al., *Different degrees of fetal oxidative stress in elective and emergent cesarean section.* Neonatology, 2007. **92**(2): p. 111-5.
- 47. Vento, M., et al., *Oxidative stress in asphyxiated term infants resuscitated with 100% oxygen.* J Pediatr, 2003. **142**(3): p. 240-6.
- 48. Frank, L. and I.R. Sosenko, *Prenatal development of lung antioxidant enzymes in four species.* J Pediatr, 1987. **110**(1): p. 106-10.
- 49. Baydas, G., et al., Antioxidant vitamin levels in term and preterm infants and their relation to maternal vitamin status. Arch Med Res, 2002. **33**(3): p. 276-80.
- 50. Abdel Ghany, E.A., et al., *Anti-oxidant profiles and markers of oxidative stress in preterm neonates.* Paediatr Int Child Health, 2016.
- 51. Negi, R., et al., Evaluation of biomarkers of oxidative stress and antioxidant capacity in the cord blood of preterm low birth weight neonates. J Matern Fetal Neonatal Med, 2012. 25(8): p. 1338-41.
- 52. Tin, W., et al., *Pulse oximetry, severe retinopathy, and outcome at one year in babies of less than 28 weeks gestation*. Arch Dis Child Fetal Neonatal Ed, 2001. **84**(2): p. F106-10.
- 53. Askie, L.M., et al., *Oxygen-saturation targets and outcomes in extremely preterm infants.* N Engl J Med, 2003. **349**(10): p. 959-67.
- 54. group, S.-R.s., Supplemental Therapeutic Oxygen for Prethreshold Retinopathy Of Prematurity (STOP-ROP), a randomized, controlled trial. I: primary outcomes. Pediatrics, 2000. **105**(2): p. 295-310.
- 55. Saugstad, O.D. and D. Aune, *Optimal oxygenation of extremely low birth weight infants: a meta-analysis and systematic review of the oxygen saturation target studies.* Neonatology, 2014. **105**(1): p. 55-63.
- 56. Darlow, B.A. and C.J. Morley, *Oxygen Saturation Targeting and Bronchopulmonary Dysplasia*. Clin Perinatol, 2015. **42**(4): p. 807-23.
- 57. Nardiello, C., et al., *Standardisation of oxygen exposure in the development of mouse models for bronchopulmonary dysplasia.* Dis Model Mech, 2017. **10**(2): p. 185-196.
- 58. Wang, L., et al., *Disruption of cytochrome P4501A2 in mice leads to increased susceptibility to hyperoxic lung injury.* Free Radic Biol Med, 2015. **82**: p. 147-59.
- 59. Berkelhamer, S.K., et al., *Developmental differences in hyperoxia-induced oxidative stress and cellular responses in the murine lung*. Free Radic Biol Med, 2013. **61**: p. 51-60.
- 60. Schock, B.C., et al., *Oxidative stress in lavage fluid of preterm infants at risk of chronic lung disease.* Am J Physiol Lung Cell Mol Physiol, 2001. **281**(6): p. L1386-91.
- 61. Collard, K.J., et al., *Pulmonary antioxidant concentrations and oxidative damage in ventilated premature babies.* Arch Dis Child Fetal Neonatal Ed, 2004. **89**(5): p. F412-6.
- 62. Dargaville, P.A., et al., *Incidence and Outcome of CPAP Failure in Preterm Infants*. Pediatrics, 2016. **138**(1).
- 63. Keszler, M. and G. Sant'Anna, *Mechanical Ventilation and Bronchopulmonary Dysplasia*. Clin Perinatol, 2015. **42**(4): p. 781-96.
- 64. Slutsky, A.S. and V.M. Ranieri, *Ventilator-induced lung injury.* N Engl J Med, 2013. **369**(22): p. 2126-36.
- 65. Dreyfuss, D., et al., *High inflation pressure pulmonary edema. Respective effects of high airway pressure, high tidal volume, and positive end-expiratory pressure.* Am Rev Respir Dis, 1988. **137**(5): p. 1159-64.
- 66. Ricard, J.D., D. Dreyfuss, and G. Saumon, *Ventilator-induced lung injury*. Eur Respir J Suppl, 2003. **42**: p. 2s-9s.
- 67. Copland, I.B., et al., *High tidal volume ventilation causes different inflammatory responses in newborn versus adult lung.* Am J Respir Crit Care Med, 2004. **169**(6): p. 739-48.
- 68. Chapman, K.E., et al., *Cyclic mechanical strain increases reactive oxygen species production in pulmonary epithelial cells.* Am J Physiol Lung Cell Mol Physiol, 2005. **289**(5): p. L834-41.

- 69. Spassov, S.G., et al., Hydrogen Sulfide Prevents Formation of Reactive Oxygen Species through PI3K/Akt Signaling and Limits Ventilator-Induced Lung Injury. Oxid Med Cell Longev, 2017.
   2017: p. 3715037.
- 70. Schmolzer, G.M., et al., *Non-invasive versus invasive respiratory support in preterm infants at birth: systematic review and meta-analysis.* Bmj, 2013. **347**: p. f5980.
- 71. Van Marter, L.J., et al., *Do clinical markers of barotrauma and oxygen toxicity explain interhospital variation in rates of chronic lung disease? The Neonatology Committee for the Developmental Network.* Pediatrics, 2000. **105**(6): p. 1194-201.
- 72. Thomson, M.A., et al., *Delayed extubation to nasal continuous positive airway pressure in the immature baboon model of bronchopulmonary dysplasia: lung clinical and pathological findings.* Pediatrics, 2006. **118**(5): p. 2038-50.
- 73. Reyburn, B., et al., *Nasal ventilation alters mesenchymal cell turnover and improves alveolarization in preterm lambs.* Am J Respir Crit Care Med, 2008. **178**(4): p. 407-18.
- 74. Kroon, A.A., J. Wang, and M. Post, *Alterations in expression of elastogenic and angiogenic genes by different conditions of mechanical ventilation in newborn rat lung.* Am J Physiol Lung Cell Mol Physiol, 2015. **308**(7): p. L639-49.
- 75. Mokres, L.M., et al., *Prolonged mechanical ventilation with air induces apoptosis and causes failure of alveolar septation and angiogenesis in lungs of newborn mice.* Am J Physiol Lung Cell Mol Physiol, 2010. **298**(1): p. L23-35.
- 76. Shah, J., et al., *Risk Factors and Outcomes of Late-Onset Bacterial Sepsis in Preterm Neonates Born at < 32 Weeks' Gestation.* Am J Perinatol, 2015. **32**(7): p. 675-82.
- Balany, J. and V. Bhandari, Understanding the Impact of Infection, Inflammation, and Their Persistence in the Pathogenesis of Bronchopulmonary Dysplasia. Front Med (Lausanne), 2015. 2: p. 90.
- 78. Uberos, J., et al., *Nutrition in extremely low birth weight infants: impact on bronchopulmonary dysplasia.* Minerva Pediatr, 2016. **68**(6): p. 419-426.
- 79. Van Marter, L.J., et al., *Hydration during the first days of life and the risk of bronchopulmonary dysplasia in low birth weight infants.* J Pediatr, 1990. **116**(6): p. 942-9.
- 80. Harkin, P., et al., *Morbidities associated with patent ductus arteriosus in preterm infants. Nationwide cohort study.* J Matern Fetal Neonatal Med, 2017: p. 1.
- 81. Polglase, G.R., et al., *Lung and systemic inflammation in preterm lambs on continuous positive airway pressure or conventional ventilation.* Pediatr Res, 2009. **65**(1): p. 67-71.
- 82. Tillema, M.S., et al., *Sublethal endotoxemia promotes pulmonary cytokine-induced neutrophil chemoattractant expression and neutrophil recruitment but not overt lung injury in neonatal rats.* Biol Neonate, 2000. **78**(4): p. 308-14.
- 83. Mataloun, M.M., et al., *Effect of postnatal malnutrition on hyperoxia-induced newborn lung development.* Braz J Med Biol Res, 2009. **42**(7): p. 606-13.
- 84. Chang, L.Y., et al., *Ductus arteriosus ligation and alveolar growth in preterm baboons with a patent ductus arteriosus.* Pediatr Res, 2008. **63**(3): p. 299-302.
- 85. Fehrholz, M., et al., *Caffeine modulates glucocorticoid-induced expression of CTGF in lung epithelial cells and fibroblasts.* Respir Res, 2017. **18**(1): p. 51.
- 86. Nakamura, T., et al., *Mechanical strain and dexamethasone selectively increase surfactant protein C and tropoelastin gene expression.* Am J Physiol Lung Cell Mol Physiol, 2000. **278**(5): p. L974-80.
- 87. Ervin, M.G., et al., *Direct fetal glucocorticoid treatment alters postnatal adaptation in premature newborn baboons.* Am J Physiol, 1998. **274**(4 Pt 2): p. R1169-76.
- 88. Eriksson, L., et al., *Prenatal inflammatory risk factors for development of bronchopulmonary dysplasia.* Pediatr Pulmonol, 2014. **49**(7): p. 665-72.
- 89. Koroglu, O.A., et al., *Anti-inflammatory effect of caffeine is associated with improved lung function after lipopolysaccharide-induced amnionitis.* Neonatology, 2014. **106**(3): p. 235-40.
- 90. Yoder, B.A., et al., *Effects of antenatal colonization with ureaplasma urealyticum on pulmonary disease in the immature baboon.* Pediatr Res, 2003. **54**(6): p. 797-807.

- 91. Soliman, N., et al., *Preeclampsia and the Risk of Bronchopulmonary Dysplasia in Preterm Infants Less Than 32 Weeks' Gestation.* Am J Perinatol, 2016.
- 92. Yallapragada, S.G., et al., *Placental Villous Vascularity Is Decreased in Premature Infants with Bronchopulmonary Dysplasia-Associated Pulmonary Hypertension.* Pediatr Dev Pathol, 2016. **19**(2): p. 101-7.
- 93. Tang, J.R., et al., *Excess soluble vascular endothelial growth factor receptor-1 in amniotic fluid impairs lung growth in rats: linking preeclampsia with bronchopulmonary dysplasia*. Am J Physiol Lung Cell Mol Physiol, 2012. **302**(1): p. L36-46.
- 94. Wagijo, M.A., et al., *Reducing tobacco smoking and smoke exposure to prevent preterm birth and its complications.* Paediatr Respir Rev, 2017. **22**: p. 3-10.
- 95. Isayama, T., et al., *Adverse Impact of Maternal Cigarette Smoking on Preterm Infants: A Population-Based Cohort Study.* Am J Perinatol, 2015. **32**(12): p. 1105-11.
- 96. Campos, K.K., et al., *Exposure to cigarette smoke during pregnancy causes redox imbalance and histological damage in lung tissue of neonatal mice.* Exp Lung Res, 2014. **40**(4): p. 164-71.
- 97. Singh, S.P., et al., *HIF-1alpha Plays a Critical Role in the Gestational Sidestream Smoke-Induced Bronchopulmonary Dysplasia in Mice.* PLoS One, 2015. **10**(9): p. e0137757.
- 98. Shaw, G.M. and H.M. O'Brodovich, *Progress in understanding the genetics of bronchopulmonary dysplasia.* Semin Perinatol, 2013. **37**(2): p. 85-93.
- 99. Whitehead, G.S., et al., *Genetic basis of murine responses to hyperoxia-induced lung injury.* Immunogenetics, 2006. **58**(10): p. 793-804.
- 100. Husain, A.N., N.H. Siddiqui, and J.T. Stocker, *Pathology of arrested acinar development in postsurfactant bronchopulmonary dysplasia.* Hum Pathol, 1998. **29**(7): p. 710-7.
- 101. De Paepe, M.E., et al., *Growth of pulmonary microvasculature in ventilated preterm infants.* Am J Respir Crit Care Med, 2006. **173**(2): p. 204-11.
- 102. Thibeault, D.W., W.E. Truog, and Ekekezie, II, *Acinar arterial changes with chronic lung disease of prematurity in the surfactant era*. Pediatr Pulmonol, 2003. **36**(6): p. 482-9.
- 103. Hsia, C.C., et al., An official research policy statement of the American Thoracic Society/European Respiratory Society: standards for quantitative assessment of lung structure. Am J Respir Crit Care Med, 2010. **181**(4): p. 394-418.
- 104. Thebaud, B., et al., Vascular endothelial growth factor gene therapy increases survival, promotes lung angiogenesis, and prevents alveolar damage in hyperoxia-induced lung injury: evidence that angiogenesis participates in alveolarization. Circulation, 2005. **112**(16): p. 2477-86.
- 105. Manzano, R.M., et al., *A hyperoxic lung injury model in premature rabbits: the influence of different gestational ages and oxygen concentrations.* PLoS One, 2014. **9**(4): p. e95844.
- 106. Coalson, J.J., et al., *Neonatal chronic lung disease in extremely immature baboons.* Am J Respir Crit Care Med, 1999. **160**(4): p. 1333-46.
- 107. Albertine, K.H., et al., *Chronic lung injury in preterm lambs. Disordered respiratory tract development.* Am J Respir Crit Care Med, 1999. **159**(3): p. 945-58.
- 108. Cooney, T.P. and W.M. Thurlbeck, *The radial alveolar count method of Emery and Mithal: a reappraisal 2--intrauterine and early postnatal lung growth.* Thorax, 1982. **37**(8): p. 580-3.
- 109. Matute-Bello, G., et al., *An official American Thoracic Society workshop report: features and measurements of experimental acute lung injury in animals*, in *Am J Respir Cell Mol Biol*. 2011: United States. p. 725-38.
- 110. Brew, N., et al., *Mechanical ventilation injury and repair in extremely and very preterm lungs.* PLoS One, 2013. **8**(5): p. e63905.
- 111. Davey, M.G., et al., *Computer-assisted stereology: point fraction of lung parenchyma and alveolar surface density in fetal and newborn sheep.* Scanning, 2003. **25**(1): p. 37-44.
- 112. Sallon, C., et al., *Automated High-Performance Analysis of Lung Morphometry*. Am J Respir Cell Mol Biol, 2015. **53**(2): p. 149-58.

- 113. Vasilescu, D.M., et al., *Stereological assessment of mouse lung parenchyma via nondestructive, multiscale micro-CT imaging validated by light microscopic histology.* J Appl Physiol (1985), 2013. **114**(6): p. 716-24.
- 114. Vasilescu, D.M., et al., Assessment of morphometry of pulmonary acini in mouse lungs by nondestructive imaging using multiscale microcomputed tomography. Proc Natl Acad Sci U S A, 2012. **109**(42): p. 17105-10.
- 115. Bruce, M.C., et al., *Risk factors for the degradation of lung elastic fibers in the ventilated neonate. Implications for impaired lung development in bronchopulmonary dysplasia.* Am Rev Respir Dis, 1992. **146**(1): p. 204-12.
- 116. Thibeault, D.W., et al., *Lung elastic tissue maturation and perturbations during the evolution of chronic lung disease.* Pediatrics, 2000. **106**(6): p. 1452-9.
- 117. Wendel, D.P., et al., *Impaired distal airway development in mice lacking elastin*. Am J Respir Cell Mol Biol, 2000. **23**(3): p. 320-6.
- 118. Bostrom, H., et al., *PDGF-A signaling is a critical event in lung alveolar myofibroblast development and alveogenesis.* Cell, 1996. **85**(6): p. 863-73.
- 119. Khan, M.A., et al., *Influence of airway wall stiffness and parenchymal tethering on the dynamics of bronchoconstriction*. Am J Physiol Lung Cell Mol Physiol, 2010. **299**(1): p. L98-1108.
- 120. Greenough, A. and A. Pahuja, *Updates on Functional Characterization of Bronchopulmonary Dysplasia The Contribution of Lung Function Testing*. Front Med (Lausanne), 2015. **2**: p. 35.
- 121. Fawke, J., et al., *Lung function and respiratory symptoms at 11 years in children born extremely preterm: the EPICure study.* Am J Respir Crit Care Med, 2010. **182**(2): p. 237-45.
- 122. Baraldi, E., et al., *Pulmonary function until two years of life in infants with bronchopulmonary dysplasia*. Am J Respir Crit Care Med, 1997. **155**(1): p. 149-55.
- 123. Vendettuoli, V., et al., *Positional effects on lung mechanics of ventilated preterm infants with acute and chronic lung disease.* Pediatr Pulmonol, 2015. **50**(8): p. 798-804.
- 124. May, C., et al., *Lung function abnormalities in infants developing bronchopulmonary dysplasia*. Arch Dis Child, 2011. **96**(11): p. 1014-9.
- 125. Wauer, R.R., et al., *Assessment of functional residual capacity using nitrogen washout and plethysmographic techniques in infants with and without bronchopulmonary dysplasia.* Intensive Care Med, 1998. **24**(5): p. 469-75.
- 126. Balinotti, J.E., et al., *Growth of lung parenchyma in infants and toddlers with chronic lung disease of infancy.* Am J Respir Crit Care Med, 2010. **181**(10): p. 1093-7.
- 127. Chang, D.V., et al., *Membrane and Capillary Components of Lung Diffusion in Infants with Bronchopulmonary Dysplasia*. Am J Respir Crit Care Med, 2015.
- 128. Bates, J.H. and C.G. Irvin, *Measuring lung function in mice: the phenotyping uncertainty principle.* J Appl Physiol (1985), 2003. **94**(4): p. 1297-306.
- 129. Richter, J., et al., *Functional assessment of hyperoxia-induced lung injury after preterm birth in the rabbit.* Am J Physiol Lung Cell Mol Physiol, 2014. **306**(3): p. L277-83.
- 130. Faksh, A., et al., *Effects of antenatal lipopolysaccharide and postnatal hyperoxia on airway reactivity and remodeling in a neonatal mouse model.* Pediatr Res, 2016. **79**(3): p. 391-400.
- 131. Shalaby, K.H., et al., *Combined forced oscillation and forced expiration measurements in mice for the assessment of airway hyperresponsiveness.* Respir Res, 2010. **11**: p. 82.
- 132. Cox, A.M., et al., *Cumulative effects of neonatal hyperoxia on murine alveolar structure and function*. Pediatr Pulmonol, 2017.
- 133. Warner, B.B., et al., *Functional and pathological effects of prolonged hyperoxia in neonatal mice.* Am J Physiol, 1998. **275**(1 Pt 1): p. L110-7.
- 134. Jones, C.V., et al., *The effect of CSF-1 administration on lung maturation in a mouse model of neonatal hyperoxia exposure*. Respir Res, 2014. **15**: p. 110.
- McCurnin, D.C., et al., *Postnatal estradiol up-regulates lung nitric oxide synthases and improves lung function in bronchopulmonary dysplasia*. Am J Respir Crit Care Med, 2009. **179**(6): p. 492-500.

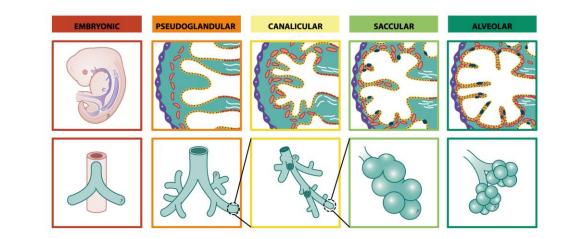
- 136. Mourani, P.M. and S.H. Abman, *Pulmonary vascular disease in bronchopulmonary dysplasia: pulmonary hypertension and beyond.* Curr Opin Pediatr, 2013. **25**(3): p. 329-37.
- 137. Thebaud, B., Angiogenesis in lung development, injury and repair: implications for chronic lung disease of prematurity. Neonatology, 2007. **91**(4): p. 291-7.
- Thebaud, B. and S.H. Abman, Bronchopulmonary dysplasia: where have all the vessels gone? Roles of angiogenic growth factors in chronic lung disease. Am J Respir Crit Care Med, 2007. 175(10): p. 978-85.
- 139. Bhat, R., et al., *Prospective analysis of pulmonary hypertension in extremely low birth weight infants.* Pediatrics, 2012. **129**(3): p. e682-9.
- 140. Bhatt, A.J., et al., *Disrupted pulmonary vasculature and decreased vascular endothelial growth factor, Flt-1, and TIE-2 in human infants dying with bronchopulmonary dysplasia.* Am J Respir Crit Care Med, 2001. **164**(10 Pt 1): p. 1971-80.
- 141. Hansmann, G., et al., *Mesenchymal stem cell-mediated reversal of bronchopulmonary dysplasia and associated pulmonary hypertension*. Pulm Circ, 2012. **2**(2): p. 170-81.
- 142. Reynolds, C.L., et al., *Phenotypic assessment of pulmonary hypertension using high-resolution echocardiography is feasible in neonatal mice with experimental bronchopulmonary dysplasia and pulmonary hypertension: a step toward preventing chronic obstructive pulmonary disease.* Int J Chron Obstruct Pulmon Dis, 2016. **11**: p. 1597-605.
- 143. Chen, S., et al., *CTGF disrupts alveolarization and induces pulmonary hypertension in neonatal mice: implication in the pathogenesis of severe bronchopulmonary dysplasia.* Am J Physiol Lung Cell Mol Physiol, 2011. **300**(3): p. L330-40.
- 144. Wedgwood, S., et al., Postnatal growth restriction augments oxygen-induced pulmonary hypertension in a neonatal rat model of bronchopulmonary dysplasia. Pediatr Res, 2016.
   80(6): p. 894-902.
- 145. Jimenez, J., et al., *Progressive Vascular Functional and Structural Damage in a Bronchopulmonary Dysplasia Model in Preterm Rabbits Exposed to Hyperoxia.* Int J Mol Sci, 2016. **17**(10).
- 146. Bland, R.D., et al., *Chronic lung injury in preterm lambs: abnormalities of the pulmonary circulation and lung fluid balance.* Pediatr Res, 2000. **48**(1): p. 64-74.
- 147. Jimenez, J., et al., *Prenatal interventions to prevent bronchopulmonary dysplasia in animal models: a systematic review.* J Matern Fetal Neonatal Med, 2016. **29**(16): p. 2555-62.
- 148. Ladha, F., et al., *Sildenafil improves alveolar growth and pulmonary hypertension in hyperoxia-induced lung injury.* Am J Respir Crit Care Med, 2005. **172**(6): p. 750-6.
- 149. Thibault, H.B., et al., *Noninvasive assessment of murine pulmonary arterial pressure: validation and application to models of pulmonary hypertension.* Circ Cardiovasc Imaging, 2010. **3**(2): p. 157-63.
- 150. Xia, H., et al., *Foxm1 regulates resolution of hyperoxic lung injury in newborns*. Am J Respir Cell Mol Biol, 2015. **52**(5): p. 611-21.
- 151. Sureshbabu, A., et al., *Conditional overexpression of TGFbeta1 promotes pulmonary inflammation, apoptosis and mortality via TGFbetaR2 in the developing mouse lung.* Respir Res, 2015. **16**: p. 4.
- 152. Schmiedl, A., et al., *Distribution of surfactant proteins in type II pneumocytes of newborn, 14day old, and adult rats: an immunoelectron microscopic and stereological study.* Histochem Cell Biol, 2005. **124**(6): p. 465-76.
- 153. Melville, J.M., et al., *Human amnion epithelial cells modulate the inflammatory response to ventilation in preterm lambs.* PLoS One, 2017. **12**(3): p. e0173572.
- 154. Dargaville, P.A., et al., *An authentic animal model of the very preterm infant on nasal continuous positive airway pressure.* Intensive Care Med Exp, 2015. **3**(1): p. 51.
- 155. Partridge, E.A., et al., *An extra-uterine system to physiologically support the extreme premature lamb.* Nat Commun, 2017. **8**: p. 15112.

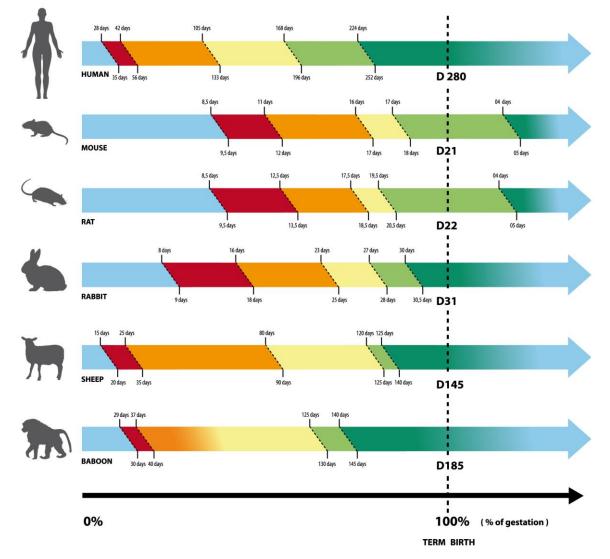
- Hanita, T., et al., Potential Role of Prenatal Inflammation in the Impairment of Lung Development Following Mechanical Ventilation of Preterm Lambs. Reprod Sci, 2017. 24(3): p. 478-487.
- 157. Tyler, W.S., *Comparative subgross anatomy of lungs. Pleuras, interlobular septa, and distal airways.* Am Rev Respir Dis, 1983. **128**(2 Pt 2): p. S32-6.
- 158. Allison, B.J., et al., *Ventilation-induced lung injury is not exacerbated by growth restriction in preterm lambs.* Am J Physiol Lung Cell Mol Physiol, 2016. **310**(3): p. L213-23.
- 159. Frank, L., J.R. Bucher, and R.J. Roberts, *Oxygen toxicity in neonatal and adult animals of various species*. J Appl Physiol Respir Environ Exerc Physiol, 1978. **45**(5): p. 699-704.
- 160. Ward, J.A. and R.J. Roberts, *Vitamin E inhibition of the effects of hyperoxia on the pulmonary surfactant system of the newborn rabbit.* Pediatr Res, 1984. **18**(4): p. 329-34.
- 161. D'Angio, C.T., et al., *Changes in surfactant protein gene expression in a neonatal rabbit model of hyperoxia-induced fibrosis.* Am J Physiol, 1997. **272**(4 Pt 1): p. L720-30.
- 162. Hua, S., et al., *Effects of different ventilation strategies on lung injury in newborn rabbits*. Pediatr Pulmonol, 2012. **47**(11): p. 1103-12.
- 163. Gras-Le Guen, C., et al., *Antenatal infection in the rabbit impairs post-natal growth and lung alveolarisation*. Eur Respir J, 2008. **32**(6): p. 1520-8.
- 164. Frank, L. and E.E. Groseclose, *Preparation for birth into an O2-rich environment: the antioxidant enzymes in the developing rabbit lung.* Pediatr Res, 1984. **18**(3): p. 240-4.
- 165. Frank, L. and I.R. Sosenko, *Failure of premature rabbits to increase antioxidant enzymes during hyperoxic exposure: increased susceptibility to pulmonary oxygen toxicity compared with term rabbits.* Pediatr Res, 1991. **29**(3): p. 292-6.
- 166. Ross, G.F., et al., *Surfactant protein C in fetal and ventilated preterm rabbit lungs.* Am J Physiol, 1999. **277**(6 Pt 1): p. L1104-8.
- 167. Pringle, K.C., *Human fetal lung development and related animal models.* Clin Obstet Gynecol, 1986. **29**(3): p. 502-13.
- 168. Walther, F.J., R. David-Cu, and S.L. Lopez, *Antioxidant-surfactant liposomes mitigate hyperoxic lung injury in premature rabbits.* Am J Physiol, 1995. **269**(5 Pt 1): p. L613-7.
- 169. Bany-Mohammed, F.M., S. Slivka, and M. Hallman, *Recombinant human erythropoietin: possible role as an antioxidant in premature rabbits.* Pediatr Res, 1996. **40**(3): p. 381-7.
- 170. Mascaretti, R.S., et al., *Lung morphometry, collagen and elastin content: changes after hyperoxic exposure in preterm rabbits.* Clinics (Sao Paulo), 2009. **64**(11): p. 1099-104.
- 171. Salaets, T., et al., *Transcriptome Analysis of the Preterm Rabbit Lung after Seven Days of Hyperoxic Exposure*. PLoS One, 2015. **10**(8): p. e0136569.
- 172. Ricci, F., et al., *In vitro and in vivo comparison between poractant alfa and the new generation synthetic surfactant CHF5633.* Pediatr Res, 2017. **81**(2): p. 369-375.
- 173. Calkovska, A., et al., *Phospholipid Composition in Synthetic Surfactants Is Important for Tidal Volumes and Alveolar Stability in Surfactant-Treated Preterm Newborn Rabbits*. Neonatology, 2016. **109**(3): p. 177-85.
- 174. Richter, J., et al., *Proton-pump inhibitor omeprazole attenuates hyperoxia induced lung injury.* J Transl Med, 2016. **14**(1): p. 247.
- 175. Richter, J., et al., *Transplacental Administration of Rosiglitazone Attenuates Hyperoxic Lung Injury in a Preterm Rabbit Model.* Fetal Diagn Ther, 2016. **39**(4): p. 297-305.
- 176. Nagatomo, T., et al., *Caffeine Prevents Hyperoxia-Induced Functional and Structural Lung Damage in Preterm Rabbits*. Neonatology, 2016. **109**(4): p. 274-81.
- 177. Chaudhary, K., et al., *Intraperitoneal drug therapy: an advantage*. Curr Clin Pharmacol, 2010. **5**(2): p. 82-8.
- 178. Smits, A., et al., *Pharmacokinetics of drugs in neonates: pattern recognition beyond compound specific observations.* Curr Pharm Des, 2012. **18**(21): p. 3119-46.
- 179. Fouts, J.R. and T.R. Devereux, *Developmental aspects of hepatic and extrahepatic drugmetabolizing enzyme systems: microsomal enzymes and components in rabbit liver and lung during the first month of life.* J Pharmacol Exp Ther, 1972. **183**(2): p. 458-68.

- 180. Lucier, G.W., B.R. Sonawane, and O.S. McDaniel, *Glucuronidation and deglucuronidation reactions in hepatic and extrahepatic tissues during perinatal development*. Drug Metab Dispos, 1977. **5**(3): p. 279-87.
- 181. Peng, H.M., et al., *Differences in the developmental expression of rabbit cytochromes P-450* 2E1 and 2E2. Mol Pharmacol, 1991. **40**(1): p. 58-62.
- 182. Matos, P., et al., *Creatinine reabsorption by the newborn rabbit kidney*. Pediatr Res, 1998.
  44(5): p. 639-41.
- 183. McNamara, P.J., D. Burgio, and S.D. Yoo, *Pharmacokinetics of caffeine and its demethylated metabolites in lactating adult rabbits and neonatal offspring. Predictions of breast milk to serum concentration ratios.* Drug Metab Dispos, 1992. **20**(2): p. 302-8.
- 184. Enders, A., et al., *Morphological variation in the interhemal areas of chorioallantoic placentae: A review.* Placenta, 1998. **Vol. 19**(Supplement 2): p. 1-19.
- 185. Fischer, B., et al., *Rabbit as a reproductive model for human health*. Reproduction, 2012. **144**(1): p. 1-10.
- 186. Halwachs, S., et al., *The ABCG2 efflux transporter from rabbit placenta: Cloning and functional characterization.* Placenta, 2016. **38**: p. 8-15.
- 187. Ornitz, D.M. and Y. Yin, *Signaling networks regulating development of the lower respiratory tract.* Cold Spring Harb Perspect Biol, 2012. **4**(5).
- 188. Hendrickx, A., *Embryology of the baboon*. 1971, Chicago (III.): University of Chicago press.
- 189. Beaudoin, S., P. Barbet, and F. Bargy, *Developmental stages in the rabbit embryo: guidelines to choose an appropriate experimental model.* Fetal Diagn Ther, 2003. **18**(6): p. 422-7.
- 190. Choi, C.W., et al., *Protective effect of chorioamnionitis on the development of bronchopulmonary dysplasia triggered by postnatal systemic inflammation in neonatal rats.* Pediatr Res, 2016. **79**(2): p. 287-94.
- 191. Joss-Moore, L.A., et al., Alveolar formation is dysregulated by restricted nutrition but not excess sedation in preterm lambs managed by noninvasive support. Pediatr Res, 2016. 80(5): p. 719-728.

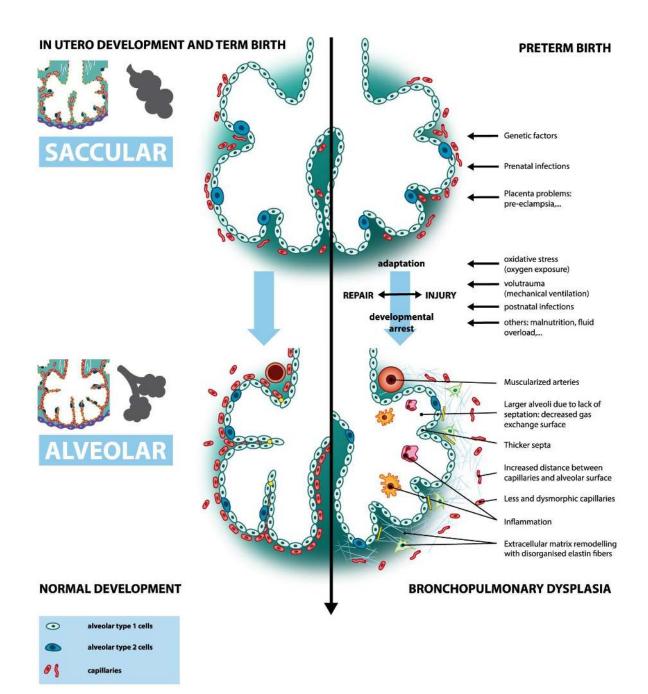
### **FIGURES**

**Figure 1: Mammalian lung development.** The upper pane shows the 5 stages of mammalian lung development. In the embryonic phase budding of the primitive lung occurs from the foregut. This buds expands to a bronchial tree in the pseudoglandular phase. In the canalicular phase the primitive lung parenchyma branches out of the bronchioles, evolving to saccular structures in the saccular phase, divided by primary septa. In the alveolar phase secondary septation occurs increasing the surface available for gas exchange. The lower pane shows the timelines of the 5 phases in humans and experimental animals frequently used to mimick BPD. Adapted from [31, 33, 34, 167, 187-189]





*Figure 2: Pathophysiology of human bronchopulmonary dysplasia.* Schematic overview of the factors involved in chronic lung disease after preterm birth and schematic representation of BPD histology.



*Figure 3: Methodology of an example of the preterm rabbit model. Schematic representation of the preterm rabbit model as described in [129]* 

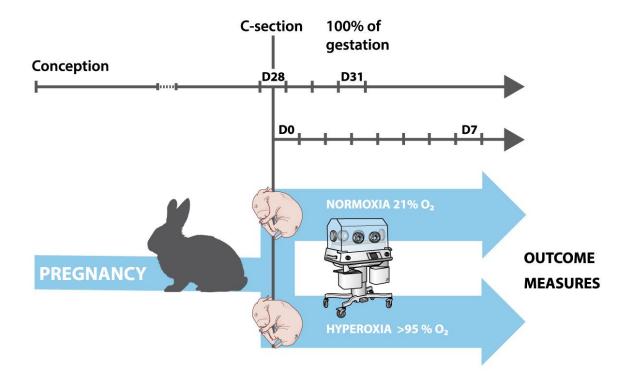
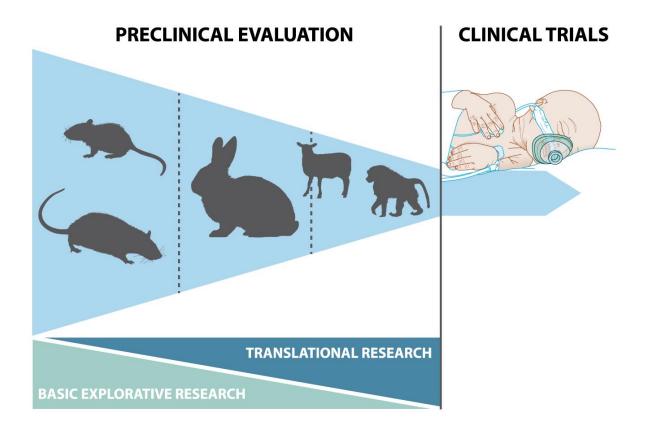


Figure 4: Place of the different animal models for BPD research. Each animal for BPD has a unique set of practical and translational advantages and disadvantages, determining their place in BPD research, schematically represented in this figure.



### TABLES

 Table 1: Animal models for BPD. Non-exhaustive summary of the factors used in the different animal models to

 mimic BPD pathophysiology. References: [31-34, 38, 74, 81, 93, 156, 158, 190, 191]

	Factors used to mimic human BPD
Mice	- Hyperoxia
	- Short term and intermittent mechanical ventilation
	- Pre- and postnatal hypoxia
	- Prenatal inflammation: LPS
	- Transgenic mice
	- Pre-eclampsia: sFlt1
Rats	- Hyperoxia
	- Short term and intermittent mechanical ventilation
	- Pre- and postnatal inflammation: LPS
Rabbits	- Prematurity
	- Hyperoxia
	- Malnutrition
	- Prenatal inflammation: E. Coli
	- Short term mechanical ventilation
Sheep	Combination of:
	- Prematurity
	- Long term mechanical ventilation or non-invasive ventilatory support: HFNV
	- Oxygen (titrated)
	- Prenatal inflammation: LPS
	- Postnatal inflammation: intratracheal LPS
	- Malnutrition
	- IUGR
D 1	- Antenatal steroids, postnatal surfactant and caffeine
Baboons	Combination of:
	- Prematurity
	<ul> <li>Long term mechanical ventilation or non-invasive ventilatory support: CPAP, HFV</li> </ul>
	- Oxygen (titrated)
	- Prenatal inflammation: Ureaplasma species
	- Antenatal steroids, postnatal surfactant