Frontiers in Nephrology

Molecular Control of Iron Transport

Tomas Ganz

Departments of Medicine and Pathology, David Geffen School of Medicine, University of California, Los Angeles, California

The iron-regulatory hormone hepcidin is a 25-amino acid peptide that is synthesized in hepatocytes. Hepcidin binds to the cellular iron export channel ferroportin and causes its internalization and degradation and thereby decreases iron efflux from iron exporting tissues into plasma. By this mechanism, hepcidin inhibits dietary iron absorption, the efflux of recycled iron from splenic and hepatic macrophages, and the release of iron from storage in hepatocytes. Hepcidin synthesis is stimulated by plasma iron and iron stores and is inhibited by erythropoietic activity, ensuring that extracellular plasma iron concentrations and iron stores remain stable and the erythropoietic demand for iron is met. During inflammation, increased hepcidin concentrations cause iron sequestration in macrophages, resulting in hypoferremia and eventually anemia of inflammation. Hepcidin deficiency plays a central role in most iron overload disorders. The role of hepcidin abnormalities in anemias that are associated with renal disease and in resistance to erythropoietic therapies remains to be elucidated.

J Am Soc Nephrol 18: 394-400, 2007. doi: 10.1681/ASN.2006070802

Iron Economy

Iron is an essential element that is required for oxidative energy metabolism in nearly all species. In humans, iron is an essential component of the oxygen carriers hemoglobin and myoglobin and of cytochromes and other enzymes that are involved in oxidation or reduction of biologic substrates. The average male adult contains approximately 4 g of iron, a little more than 2 g of which is in hemoglobin (each 1 ml of packed erythrocytes contains approximately 1 mg of iron), 1 g in body stores predominantly in the liver, and the rest in myoglobin and other iron-containing proteins. Approximately 1 to 2 mg of iron is lost each day by epithelial shedding in the gastrointestinal tract and the skin and through blood loss in menstruating women. There is no physiologic mechanism for excreting larger amounts of iron, even in severely iron-overloaded individuals. The normal losses are balanced by absorption of iron from the diet. Western diets contain a much greater amount of iron (10 to 20 mg) than what is absorbed daily under normal circumstances (1 to 2 mg). Iron absorption increases several-fold in iron deficiency and is suppressed partly when iron stores are excessive. Approximately 20 mg/d iron is recycled from senescent erythrocytes by macrophages. Erythrocytes have a life span of 120 d, so each day, approximately 1% of erythrocytes is removed by macrophages from the circulation and iron is extracted from hemoglobin. Recycled and absorbed iron is delivered to transferrin in blood, and most of it is destined for the nascent erythrocytes in the bone marrow, whereas a much smaller portion is distributed to other tissues. The plasma transferrin compartment is relatively small, containing only approximately 3 mg of iron, which therefore must turn over every few hours.

Iron plays an important role in host defense responses to infectious agents. Several host defense proteins, including lactoferrin, siderocalin (also called neutrophil gelatinase–associated lipocalin, lipocalin 2, or NGAL), and the macrophage divalent metal transporter natural resistance associated macrophage protein 2, seem to function in infected tissues to sequester iron from invading microorganisms. On a systemic level, hypoferremia develops early during infection and, if prolonged, eventually leads to the characteristic anemia of inflammation (anemia of chronic disease).

Iron absorption, recycling, and movement in and out of stores is subject to regulation by three major influences: Systemic iron status (referred to as the "stores" regulator), the iron requirements of erythropoiesis ("erythropoietic" regulator), and the pathologic effects of inflammation ("inflammatory" regulator). As is discussed in detail, it has become clear that all three of these influences converge on a single iron-regulatory hormone, hepcidin, whose biology is central to systemic iron homeostasis.

Iron Absorption

Dietary iron is presented to the duodenum either as ferric iron complexed with macromolecules such as ferritin or phytates or in the form of heme or heme-containing proteins. At the epithelial surface, ferric iron is reduced to ferrous iron with the assistance of one or more apical ferric reductases, possibly including the duodenal cytochrome dCytB, and crosses into the cytoplasm *via* the apical enterocyte transporter

Published online ahead of print. Publication date available at www.jasn.org.

Address correspondence to: Dr. Tomas Ganz, Departments of Medicine and Pathology, David Geffen School of Medicine, 10833 Le Conte Avenue, CHS 37-055, University of California, Los Angeles, CA 90095-1690. Phone: 310-825-6112; Fax: 310-206-8766; E-mail: tganz@mednet.ucla.edu

divalent metal transporter 1 (DMT-1), as reviewed recently (1). DMT-1-mediated iron transport requires an inward proton gradient. Dietary heme is taken up by apical heme transporters, one of which was characterized recently (2). In the cytoplasm, iron is stored in ferritin. On the basolateral surface of duodenal enterocytes, cellular iron is exported to plasma by ferroportin (also called iron regulated transporter 1, mental transport protein 1, or SLC40A1) (3-5). Ferroportin cooperates with the multicopper ferroxidase hephaestin (6), which converts ferrous to ferric iron for uptake by plasma transferrin. As shown in mice with homozygous disruption of ferroportin genes, little or no iron transfer from enterocytes to plasma takes place without ferroportin, despite massive iron accumulation in enterocytes (7). Ferroportin therefore is a natural control point for intestinal iron absorption. The short lifespan of intestinal enterocytes (approximately 2 d) ensures that iron that is not transferred to plasma is shed with the enterocytes into the fecal stream.

Iron Recycling and Storage

As indicated in a recent review, despite its fundamental importance, the biology of iron recycling by macrophages is "one of the least well understood areas of iron metabolism" (8). At the end of their 4-mo lifespan, human erythrocytes undergo surface alterations that mark them to be phagocytosed and digested by macrophages in the spleen and the liver. In macrophages, iron is recovered from heme by the action of heme oxygenase (predominantly the inducible heme oxygenase HO-1). It is not certain at what subcellular location HO-1 performs its activity. Both DMT-1 and the closely related divalent metal transporter natural resistance associated macrophage protein 2 are expressed in macrophages and could transport iron across the phagosomal membrane to the cytoplasm, but it is not clear that either is required for this function. Macrophages, like other cells, store iron in ferritin. Iron eventually is exported to plasma transferrin by ferroportin with the aid of the multicopper ferroxidase ceruloplasmin (9), but it remains to be established whether ceruloplasmin interacts directly with ferroportin. Ceruloplasmin-deficient patients have a mild impairment of iron mobilization from macrophages as indicated by a mild anemia and gradual iron retention in macrophages, suggesting that the ferroxidase function of ceruloplasmin is partially redundant. Conversely, ferroportin is essential for iron recycling; ferroportin-deficient mice become severely anemic and rapidly accumulate iron in macrophages (7).

Hepatocytes are a major site of iron storage and express relatively low amounts of the iron exporter ferroportin on surfaces that face the sinusoids (5). In states of genetic or acquired iron overload, hepatocytes become a major site of iron deposition, presumably because their iron uptake exceeds the capacity for export. Hepatocytes display particularly high uptake rates for non-transferrin-bound iron, a form of iron that is present when iron load exceeds the iron-binding capacity of transferrin (10).

Hepcidin

The bioactive form of hepcidin is a 25–amino acid cationic peptide that contains four disulfide bonds (11–15). It is encoded

as an 84–amino acid prepropeptide and is synthesized, processed, and secreted predominantly by hepatocytes. Injection of synthetic hepcidin into mice induces profound hypoferremia within 1 h (14). The essential role of hepcidin in iron homeostasis was established in transgenic mouse models and human diseases that result in hepcidin deficiency (16–18) or excessive production of the peptide (19,20). Complete deficiency of hepcidin causes juvenile hemochromatosis, a severe form of genetic iron overload in which dietary iron absorption is dysregulated so that iron is taken up at a high rate despite excessive iron stores. Excessive production of hepcidin causes iron deficiency anemia as a result of the individual's inability to absorb iron, despite normal or even iron-enriched diet. The severity of both conditions indicates that any mechanisms that override the effects of hepcidin are relatively ineffective.

Hepcidin Assays

The small size and evolutionary conservation of hepcidin has hampered the development of sensitive immunoassays. In humans, urinary hepcidin measurements detect the bioactive form of hepcidin, have provided physiologically meaningful information (21,22), and have correlated well with hepcidin mRNA concentrations in liver biopsies from the same patients (23). A human serum prohepcidin assay (24) has been less reflective of iron or inflammatory physiology probably because prohepcidin is a processing intermediate without a direct biologic role in iron metabolism. In mice, hepcidin-1 mRNA is used as a proxy for the measurement of the bioactive hepcidin form. Although further studies are necessary to clarify this issue, it seems that hepcidin is regulated predominantly by the concentrations of its mRNA so that the conclusions from mRNA-based mouse studies and peptide-based human studies are very similar. More recently, semiquantitative assays that were based on mass spectrometry were developed for human urinary (25) and serum (26) hepcidin, and these hold promise especially for measurements of elevated hepcidin concentrations.

Regulation of Hepcidin Synthesis

Hepcidin is induced by iron loading ("stores" signal in the older literature) and by inflammation and is suppressed by erythropoietic activity (12,21,22,27,28). The effects of acute inflammation are best understood and are mediated at least in part by IL-6 (21,29) through the induction and binding of STAT-3 to the hepcidin promoter (30). Although the acute hypoferremic response and hepcidin induction both are impaired in IL-6-deficient mice (21), chronic inflammatory stimulation elicits hepcidin even in IL-6-deficient mice (31). Potential candidates for chronic stimulatory factors include bone morphogenic proteins and TGF- β and other ligands of the receptors of the TGF- β family (32–34). It is not yet known how iron and erythropoiesis regulate hepcidin. Iron ingestion or parenteral iron administration results in the induction of hepcidin (12,21), which in turn blocks iron absorption until the normal plasma iron levels are restored. Hepcidin deficiency as a result of dysregulation of its synthesis would be expected to lead to iron accumulation and hemochromatosis. Indeed, recent studies indicate that hepcidin is deficient in hereditary hemochromatosis as a result of mutations in transferrin receptor 2, HFE, or hemojuvelin (35–38). These genes therefore must encode regulators of hepcidin synthesis. Recent studies showed that these molecules form a hepcidin-regulating complex, possibly also involving bone morphogenic proteins and their receptor (32). However, how iron is sensed by these molecules and which signaling pathways they activate to regulate hepcidin in response to iron remain to be elucidated.

Hepcidin also is regulated by anemia (27), but it seems that the effects of anemia largely are mediated by an unknown factor that is produced during erythropoietic activity (28,39). Hepcidin is suppressed appropriately in mice that are made anemic by phlebotomy or hemolysis, but this suppression is reversed when reactive erythropoiesis is inhibited by cytotoxic agents, radiation, or erythropoietin-neutralizing antibodies (28,39). The hepcidin-suppressive effect of erythropoietin also is inhibited by cytotoxic agents and therefore is substantially dependent on erythropoietic activity (39). Although hepcidin suppression during anemia and reactive erythropoiesis could be due in part to increased iron use and systemic iron depletion, hepcidin suppression also is seen in mouse models and human diseases in which severe systemic iron overload coexists with anemia and reactive erythropoiesis. Urinary hepcidin concentrations are very low in untransfused anemic patients with thalassemia intermedia and systemic iron overload (22), and hepcidin mRNA concentrations are low in anemic mice with hypotransferrinemia and severe iron overload (20). The hepcidin-regulatory factor that is generated during erythropoiesis and acts on hepatocytes remains to be identified.

Ferroportin

All cells require iron for energy metabolism and other metabolic processes. Depending on the cell type, multiple mechanisms exist for iron uptake, whether in the form of transferrinbound iron, nontransferrin iron, heme, hemoglobin, or entire red blood cells. In contrast, the ability to export iron is limited to tissues that are engaged in iron transport, including small intestinal (mainly duodenal) epithelium, macrophages, hepatocytes, and embryonic or placental cells that interface to the maternal circulation. As shown in ferroportin knockout mice, ferroportin is the sole or predominant efflux channel for iron in all of these tissues (7). Autosomal dominant ferroportin mutations that lead to mislocalization and degradation of ferroportin cause accumulation of iron in macrophages (ferroportin disease), confirming the critical role of ferroportin in iron recycling by macrophages (40–42). Ferroportin is a multipass membrane protein whose topology has not yet been established with certainty. Moreover, little is known about how it transports iron.

Hepcidin Regulates Ferroportin

Hepcidin regulates iron efflux by binding to ferroportin and inducing its internalization and lysosomal degradation (43,44) (Figure 1). In cells that express ferroportin, hepcidin has a concentration-dependent inhibitory effect on iron export that parallels the hepcidin-induced loss of ferroportin from the plasma membrane (15,43). Therefore, ferroportin is both a hep-



Figure 1. Hepcidin regulates cellular iron export into plasma. When hepcidin concentrations are low, ferroportin (Fpn) molecules are displayed on the plasma membrane and export iron. When hepcidin concentrations increase, hepcidin binds to ferroportin molecules and induces their internalization and degradation, and iron release is decreased progressively. Illustration by Josh Gramling—Gramling Medical Illustration.

cidin-regulated iron efflux channel and the hepcidin receptor. Several autosomal dominant mutations of ferroportin interfere with its ability to bind and internalize hepcidin (40-42,45,46) and cause systemic iron overload similar to that from hepcidin deficiency (45,47). The molecular details of hepcidin binding by ferroportin and of the ferroportin internalization pathway remain to be elucidated. On the hepcidin molecule, the five N-terminal amino acids seem to be required for the interaction with ferroportin (15), and hepcidin-20, a naturally occurring form that lacks these amino acids, essentially is inactive.

Hepcidin Metabolism

Hepcidin is detectable in urine predominantly as the bioactive 25–amino acid form and, to a lesser extent, as N-terminally truncated 20– and 22–amino acid forms (11) that biologically are much less active (15) and probably represent degradation products. Renal clearance of hepcidin may be metabolically important, as suggested by its accumulation in patients with renal failure (26). Presumably because of its very small size, hepcidin is cleared efficiently by hemodialysis (26,48). When injected into mice, radiolabeled hepcidin is excreted rapidly in urine but also accumulates in ferroportin-rich tissues (14), including the proximal duodenum, the spleen, and the liver, where its uptake and degradation also could contribute to its removal from circulation.

Systemic Iron Homeostasis

Despite large fluxes of iron through the plasma compartment, human plasma iron concentrations normally are maintained in a relatively narrow range of 10 to 30 μ M. This suggests that a homeostatic mechanism must exist to regulate extracellular iron concentrations. Such a mechanism would be expected to contain an iron sensor, transduction machinery, and messenger molecules that regulate iron efflux from various cellular reservoirs into plasma. Recent studies suggest that the hepcidin-ferroportin interaction is at the core of this regulatory system (Figure 2). Intraperitoneal injection of 50 μ g of hepcidin into mice causes profound hypoferremia as soon as 1 h after the injection (14). Serum iron does not return to normal until more than 48 h later, consistent with resynthesis of ferroportin that has been degraded in lysosomes under the influence of hepcidin. The key role of the hepcidin–ferroportin interaction in systemic iron homeostasis is supported by the phenotypic similarity of hemochromatosis as a result of hepcidin deficiency or mutations in ferroportin that interfere with hepcidin binding or ferroportin internalization (45,47).

Anemia of Inflammation

Anemia of inflammation (also called anemia of chronic disease) is defined by hypoferremia despite normal iron stores. Operationally, normal iron stores used to be established by the presence of stainable iron in bone marrow macrophages, but, more recently, adequate iron stores are assessed by normal or elevated serum ferritin. The anemia usually is mild to moderate and normocytic/normochromic. The pathophysiology of ane-



Figure 2. Systemic iron homeostasis. By regulating ferroportin, hepcidin controls the entry of iron into plasma. The major iron flows that are regulated by hepcidin–ferroportin interactions include the release of iron from macrophages that recycle iron in the spleen and other organs, dietary iron absorption in the duodenum, and the release of iron from storage in hepatocytes. The feedback stimulation of hepcidin by plasma iron saturation and iron stores ensures that extracellular iron concentration and iron stores stay within normal limits. Hepcidin synthesis is suppressed by erythropoietic activity, ensuring a sufficient supply of iron to the bone marrow when demand for erythrocytes is high. During inflammation, hepcidin production is stimulated and iron entry into plasma is inhibited, causing the hypoferremia and anemia of inflammation. Illustration by Josh Gramling–Gramling Medical Illustration.

mia of inflammation centers on iron-limited erythropoiesis that is exacerbated to a varying extent by shortened erythrocyte lifespan. The iron limitation is evidenced by hypoferremia, which in turn reflects the sequestration of iron in macrophages, intestinal enterocytes, and possibly hepatocytes. Other correlates of iron-limited erythropoiesis include elevation of zinc protoporphyrin as a result of substitution of zinc for iron during the formation of heme. It has been assumed that the hypoferremia of inflammation has a role in host defense against microbial infection, together with a number of other mechanisms that limit iron availability to invading microorganisms (49).

Role of Hepcidin in Anemia of Inflammation

The inducibility of hepcidin by inflammatory stimuli (12,27,29,50) suggested that hepcidin, by limiting iron export from macrophages, could have a key role in anemia of inflammation. Excess hepcidin production in transgenic mice causes iron-restricted erythropoiesis (19), and anemia with hypoferremia also is seen with tumors that overproduce hepcidin in humans or mice (20,51). Patients with anemia of inflammation had elevated urinary hepcidin concentrations (50) that correlated with serum ferritin, a marker of inflammation. These studies strongly suggest that the final common pathway for anemia of inflammation involves cytokine stimulation of hepcidin synthesis, iron sequestration as a result of hepcidin-induced loss of ferroportin from macrophages, and iron-limited erythropoiesis. Inflammatory cytokines (including IL-1, TNF- α , IL-6, and IFN- γ) also may affect erythropoiesis through hepcidin-independent effects on erythroid development in the bone marrow (52) and by suppressing the production of erythropoietin (53). Moreover, ferroportin mRNA levels seem to be regulated by cytokines and toll-like receptor-dependent pathways by hepcidin-independent mechanisms (54). The relative impact of these mechanisms on erythropoiesis and iron metabolism in vivo remains to be determined.

Role of Hepcidin in Iron Overload Disorders

Acute or chronic dietary supplementation or iron injections increased hepcidin mRNA in normal mice (12,21), and the administration of 65 mg of oral ferrous iron to a group of human volunteers increased urinary hepcidin excretion more than five-fold within 24 h. Therefore, increased production of hepcidin represents the normal response to iron loading. The rapid response of urinary hepcidin to iron load in human volunteers (as little as 6 h; T. Ganz and E. Nemeth, unpublished observations) suggests that the system senses the iron saturation of transferrin, but both the form of iron and the specific molecular sensors that detect it remain to be identified. The expected effect of increased hepcidin after iron challenge would be homeostatic: To decrease further absorption of iron from the diet and inhibit its release from macrophages. Pathologic iron overload occurs in hereditary hemochromatoses, in anemias with massively increased erythropoiesis, or after repeated erythrocyte transfusions. In these settings, the normal response to iron loading is modified by genetic lesions that decrease or ablate hepcidin synthesis (hereditary hemochromatoses) or by the suppressive effects of anemia and erythropoietic activity on hepcidin production. Hereditary hemochromatoses (55) are a group of diseases that are caused by autosomal recessive mutations in genes that encode the molecules HFE, human transferrin receptor 2, hemojuvelin, and hepcidin itself and certain autosomal dominant mutations in the hepcidin target ferroportin. Decreased production or absence of hepcidin or, in the case of ferroportin mutations, insensitivity to hepcidin seems to be the common feature of the disease, regardless of the specific genetic lesion involved. Hepcidin deficiency or resistance to hepcidin therefore is the fundamental cause of hemochromatosis.

In further support of the central involvement of hepcidin in the pathogenesis of hemochromatosis, transgenic correction of hepcidin deficiency in a mouse model of HFE hemochromatosis prevents the development of iron overload pathology (56). If hepcidin synthesis is switched on after hemochromatosis already developed, then increased hepcidin causes redistribution of iron from parenchymal cells to macrophages, where iron is relatively nontoxic (57). In iron-loading anemias (β -thalassemia and congenital dyserythropoietic anemia), the suppressive effect of erythropoiesis on hepcidin production (22,58) and the resulting increase in dietary iron absorption is sufficient to cause systemic iron overload and iron-mediated damage to the liver and myocardium even without blood transfusions. It remains to be seen whether exogenous hepcidin can correct the iron pathology in iron-loading anemias as well.

Role of Hepcidin in Anemia of Renal Disease

Anemia of renal disease has a multifactorial pathogenesis, including erythropoietin deficiency, inflammatory effects of the primary disease, inflammatory effects of its complications and of its treatments, and the potential consequences of decreased renal clearance of hepcidin. Both inflammation and the decreased clearance of hepcidin could raise blood hepcidin concentrations and lead to iron-restricted erythropoiesis. In milder cases, iron restriction could become manifest only when erythropoietic activity and iron demand increase as a result of treatment with recombinant erythropoietin. In these settings, iron restriction may lead to erythropoietin resistance with partial reversibility by parenteral iron therapy (59). Fundamental understanding of these processes is desirable but awaits further studies with improved serum hepcidin assays.

Conclusion

Hepcidin is the iron-regulatory hormone responsible for systemic iron homeostasis. It regulates intestinal iron absorption and the release of iron from macrophages and hepatic stores. Hepcidin acts by binding to the sole cellular iron exporter ferroportin and causing its internalization and degradation. Pathologic alterations of hepcidin regulation are central to disorders of iron metabolism, including hereditary hemochromatosis, iron-loading anemias, and anemia of inflammation. The role of hepcidin in anemias that are associated with renal diseases remains to be explored.

Acknowledgment

This work was supported by National Institutes of Health grant HL 46809 and the Will Rogers Fund.

Disclosures

None.

References

- Mackenzie B, Garrick MD: Iron imports. II. Iron uptake at the apical membrane in the intestine. *Am J Physiol Gastrointest Liver Physiol* 289: G981–G986, 2005
- Shayeghi M, Latunde-Dada GO, Oakhill JS, Laftah AH, Takeuchi K, Halliday N, Khan Y, Warley A, McCann FE, Hider RC, Frazer DM, Anderson GJ, Vulpe CD, Simpson RJ, McKie AT: Identification of an intestinal heme transporter. *Cell* 122: 789–801, 2005
- Donovan A, Brownlie A, Zhou Y, Shepard J, Pratt SJ, Moynihan J, Paw BH, Drejer A, Barut B, Zapata A, Law TC, Brugnara C, Lux SE, Pinkus GS, Pinkus JL, Kingsley PD, Palis J, Fleming MD, Andrews NC, Zon LI: Positional cloning of zebrafish ferroportin1 identifies a conserved vertebrate iron exporter. *Nature* 403: 776–781, 2000
- McKie AT, Marciani P, Rolfs A, Brennan K, Wehr K, Barrow D, Miret S, Bomford A, Peters TJ, Farzaneh F, Hediger MA, Hentze MW, Simpson RJ: A novel duodenal iron-regulated transporter, IREG1, implicated in the basolateral transfer of iron to the circulation. *Mol Cell* 5: 299–309, 2000
- 5. Abboud S, Haile DJ: A novel mammalian iron-regulated protein involved in intracellular iron metabolism. *J Biol Chem* 275: 19906–19912, 2000
- Vulpe CD, Kuo YM, Murphy TL, Cowley L, Askwith C, Libina N, Gitschier J, Anderson GJ: Hephaestin, a ceruloplasmin homologue implicated in intestinal iron transport, is defective in the sla mouse. *Nat Genet* 21: 195–199, 1999
- Donovan A, Lima CA, Pinkus JL, Pinkus GS, Zon LI, Robine S, Andrews NC: The iron exporter ferroportin/ Slc40a1 is essential for iron homeostasis. *Cell Metabolism* 1: 191–200, 2005
- Knutson M, Wessling-Resnick M: Iron metabolism in the reticuloendothelial system. Crit Rev Biochem Mol Biol 38: 61–88, 2003
- Harris ZL, Durley AP, Man TK, Gitlin JD: Targeted gene disruption reveals an essential role for ceruloplasmin in cellular iron efflux. *Proc Natl Acad Sci U S A* 96: 10812– 10817, 1999
- Craven CM, Alexander J, Eldridge M, Kushner JP, Bernstein S, Kaplan J: Tissue distribution and clearance kinetics of non-transferrin-bound iron in the hypotransferrinemic mouse: A rodent model for hemochromatosis. *Proc Natl Acad Sci U S A* 84: 3457–3461, 1987
- Park CH, Valore EV, Waring AJ, Ganz T: Hepcidin, a urinary antimicrobial peptide synthesized in the liver. *J Biol Chem* 276: 7806–7810, 2001
- Pigeon C, Ilyin G, Courselaud B, Leroyer P, Turlin B, Brissot P, Loreal O: A new mouse liver-specific gene, encoding a protein homologous to human antimicrobial peptide hepcidin, is overexpressed during iron overload. *J Biol Chem* 276: 7811–7819, 2001
- 13. Hunter HN, Fulton DB, Ganz T, Vogel HJ: The solution structure of human hepcidin, a peptide hormone with

antimicrobial activity that is involved in iron uptake and hereditary hemochromatosis. *J Biol Chem* 277: 37597–37603, 2002

- 14. Rivera S, Nemeth E, Gabayan V, Lopez MA, Farshidi D, Ganz T: Synthetic hepcidin causes rapid dose-dependent hypoferremia and is concentrated in ferroportin-containing organs. *Blood* 106: 2196–2199, 2005
- Nemeth E, Preza GC, Jung CL, Kaplan J, Waring AJ, Ganz T: The N-terminus of hepcidin is essential for its interaction with ferroportin: Structure-function study. *Blood* 107: 328– 333, 2006
- Nicolas G, Bennoun M, Devaux I, Beaumont C, Grandchamp B, Kahn A, Vaulont S: Lack of hepcidin gene expression and severe tissue iron overload in upstream stimulatory factor 2 (USF2) knockout mice. *Proc Natl Acad Sci U S A* 98: 8780–8785, 2001
- 17. Viatte L, Lesbordes-Brion JC, Lou DQ, Bennoun M, Nicolas G, Kahn A, Canonne-Hergaux F, Vaulont S: Deregulation of proteins involved in iron metabolism in hepcidin-deficient mice. *Blood* 105: 4861–4864, 2005
- Roetto A, Papanikolaou G, Politou M, Alberti F, Girelli D, Christakis J, Loukopoulos D, Camaschella C: Mutant antimicrobial peptide hepcidin is associated with severe juvenile hemochromatosis. *Nat Genet* 33: 21–22, 2003
- Nicolas G, Bennoun M, Porteu A, Mativet S, Beaumont C, Grandchamp B, Sirito M, Sawadogo M, Kahn A, Vaulont S: Severe iron deficiency anemia in transgenic mice expressing liver hepcidin. *Proc Natl Acad Sci U S A* 99: 4596–4601, 2002
- 20. Weinstein DA, Roy CN, Fleming MD, Loda MF, Wolfsdorf JI, Andrews NC: Inappropriate expression of hepcidin is associated with iron refractory anemia: Implications for the anemia of chronic disease. *Blood* 100: 3776–3781, 2002
- Nemeth E, Rivera S, Gabayan V, Keller C, Taudorf S, Pedersen BK, Ganz T: IL-6 mediates hypoferremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. J Clin Invest 113: 1271–1276, 2004
- 22. Papanikolaou G, Tzilianos M, Christakis JI, Bogdanos D, Tsimirika K, MacFarlane J, Goldberg YP, Sakellaropoulos N, Ganz T, Nemeth E: Hepcidin in iron overload disorders. *Blood* 105: 4103–4105, 2005
- Detivaud L, Nemeth E, Boudjema K, Turlin B, Troadec MB, Leroyer P, Ropert M, Jacquelinet S, Courselaud B, Ganz T, Brissot P, Loreal O: Hepcidin levels in humans are correlated with hepatic iron stores, hemoglobin levels, and hepatic function. *Blood* 106: 746–748, 2005
- 24. Kulaksiz H, Gehrke SG, Janetzko A, Rost D, Bruckner T, Kallinowski B, Stremmel W: Pro-hepcidin: Expression and cell specific localisation in the liver and its regulation in hereditary haemochromatosis, chronic renal insufficiency, and renal anaemia. *Gut* 53: 735–743, 2004
- 25. Kemna E, Tjalsma H, Laarakkers C, Nemeth E, Willems H, Swinkels D: Novel urine hepcidin assay by mass spectrometry. *Blood* 106: 3268–3270, 2005
- 26. Tomosugi N, Kawabata H, Wakatabe R, Higuchi M, Yamaya H, Umehara H, Ishikawa I: Detection of serum hepcidin in renal failure and inflammation by using ProteinChip System. *Blood* 108: 1381–1387, 2006
- 27. Nicolas G, Chauvet C, Viatte L, Danan JL, Bigard X, Devaux I, Beaumont C, Kahn A, Vaulont S: The gene encoding the iron regulatory peptide hepcidin is regulated by

anemia, hypoxia, and inflammation. *J Clin Invest* 110: 1037–1044, 2002

- 28. Vokurka M, Krijt J, Sulc K, Necas E: Hepcidin mRNA levels in mouse liver respond to inhibition of erythropoiesis. *Physiol Res* February 23, 2006 [epub ahead of print]
- 29. Kemna E, Pickkers P, Nemeth E, van der HH, Swinkels D: Time-course analysis of hepcidin, serum iron, and plasma cytokine levels in humans injected with LPS. *Blood* 106: 1864–1866, 2005
- Wrighting DM, Andrews NC: Interleukin-6 induces hepcidin expression through STAT3. *Blood* 108: 3204–3209, 2006
- 31. Rivera S, Gabayan V, Ganz T: In chronic inflammation, there exists an IL-6 independent pathway for the induction of hepcidin. *ASH Annual Meeting Abstracts* 104: 3205, 2004
- 32. Babitt JL, Huang FW, Wrighting DM, Xia Y, Sidis Y, Samad TA, Campagna JA, Chung RT, Schneyer AL, Woolf CJ, Andrews NC, Lin HY: Bone morphogenetic protein signaling by hemojuvelin regulates hepcidin expression. *Nat Genet* 38: 531–539, 2006
- 33. Truksa J, Peng H, Lee P, Beutler E: Bone morphogenetic proteins 2, 4, and 9 stimulate murine hepcidin 1 expression independently of Hfe, transferrin receptor 2 (Tfr2), and IL-6. *Proc Natl Acad Sci U S A* 103: 10289–10293, 2006
- Wang RH, Li C, Xu X, Zheng Y, Xiao C, Zerfas P, Cooperman S, Eckhaus M, Rouault T, Mishra L, Deng CX: A role of SMAD4 in iron metabolism through the positive regulation of hepcidin expression. *Cell Metab* 2: 399–409, 2005
- 35. Gehrke SG, Kulaksiz H, Herrmann T, Riedel HD, Bents K, Veltkamp C, Stremmel W: Expression of hepcidin in hereditary hemochromatosis: Evidence for a regulation in response to the serum transferrin saturation and non-transferrin-bound iron. *Blood* 102: 371–376, 2003
- 36. Muckenthaler M, Roy CN, Custodio AO, Minana B, de-Graaf J, Montross LK, Andrews NC, Hentze MW: Regulatory defects in liver and intestine implicate abnormal hepcidin and Cybrd1 expression in mouse hemochromatosis. *Nat Genet* 34: 102–107, 2003
- 37. Papanikolaou G, Samuels ME, Ludwig EH, MacDonald ML, Franchini PL, Dube MP, Andres L, MacFarlane J, Sakellaropoulos N, Politou M, Nemeth E, Thompson J, Risler JK, Zaborowska C, Babakaiff R, Radomski CC, Pape TD, Davidas O, Christakis J, Brissot P, Lockitch G, Ganz T, Hayden MR, Goldberg YP: Mutations in HFE2 cause iron overload in chromosome 1q-linked juvenile hemochromatosis. *Nat Genet* 36: 77–82, 2004
- Nemeth E, Roetto A, Garozzo G, Ganz T, Camaschella C: Hepcidin is decreased in TFR2 hemochromatosis. *Blood* 105: 1803–1806, 2005
- Pak M, Lopez MA, Gabayan V, Ganz T, Rivera S: Suppression of hepcidin during anemia requires erythropoietic activity. *Blood* 108: 3730–3735, 2006
- De Domenico I, Ward DM, Nemeth E, Vaughn MB, Musci G, Ganz T, Kaplan J: The molecular basis of ferroportinlinked hemochromatosis. *Proc Natl Acad Sci U S A* 102: 8955–8960, 2005
- De Domenico I, Ward DM, Musci G, Kaplan J: Iron overload due to mutations in ferroportin. *Haematologica* 91: 92–95, 2006
- 42. Schimanski LM, Drakesmith H, Merryweather-Clarke AT, Viprakasit V, Edwards JP, Sweetland E, Bastin JM, Cowley D, Chinthammitr Y, Robson KJ, Townsend AR: In vitro functional analysis of human ferroportin (FPN) and hemo-

chromatosis-associated FPN mutations. Blood 105: 4096-4102, 2005

- Nemeth E, Tuttle MS, Powelson J, Vaughn MB, Donovan A, Ward DM, Ganz T, Kaplan J: Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* 306: 2090–2093, 2004
- Knutson MD, Oukka M, Koss LM, Aydemir F, Wessling-Resnick M: Iron release from macrophages after erythrophagocytosis is up-regulated by ferroportin 1 overexpression and down-regulated by hepcidin. *Proc Natl Acad Sci U S A* 102: 1324–1328, 2005
- 45. Drakesmith H, Schimanski LM, Ormerod E, Merryweather-Clarke AT, Viprakasit V, Edwards JP, Sweetland E, Bastin JM, Cowley D, Chinthammitr Y, Robson KJ, Townsend AR: Resistance to hepcidin is conferred by hemochromatosis-associated mutations of ferroportin. *Blood* 106: 1092–1097, 2005
- Liu XB, Yang F, Haile DJ: Functional consequences of ferroportin 1 mutations. *Blood Cells Mol Dis* 35: 33–46, 2005
- 47. Sham RL, Phatak PD, West C, Lee P, Andrews C, Beutler E: Autosomal dominant hereditary hemochromatosis associated with a novel ferroportin mutation and unique clinical features. *Blood Cells Mol Dis* 34: 157–161, 2005
- Krause A, Neitz S, Magert HJ, Schulz A, Forssmann WG, Schulz-Knappe P, Adermann K: LEAP-1, a novel highly disulfide-bonded human peptide, exhibits antimicrobial activity. *FEBS Lett* 480: 147–150, 2000
- 49. Jurado RL: Iron, infections, and anemia of inflammation. *Clin Infect Dis* 25: 888–895, 1997
- Nemeth E, Valore EV, Territo M, Schiller G, Lichtenstein A, Ganz T: Hepcidin, a putative mediator of anemia of inflammation, is a type II acute-phase protein. *Blood* 101: 2461–2463, 2003
- 51. Rivera S, Liu L, Nemeth E, Gabayan V, Sorensen OE, Ganz T:

Hepcidin excess induces the sequestration of iron and exacerbates tumor-associated anemia. *Blood* 105: 1797–1802, 2005

- Means RT Jr, Krantz SB: Inhibition of human erythroid colony-forming units by interferons alpha and beta: Differing mechanisms despite shared receptor. *Exp Hematol* 24: 204–208, 1996
- 53. Theurl I, Mattle V, Seifert M, Mariani M, Marth C, Weiss G: Dysregulated monocyte iron homeostasis and erythropoietin formation in patients with anemia of chronic disease. *Blood* 107: 4142–4148, 2006
- Liu XB, Nguyen NB, Marquess KD, Yang F, Haile DJ: Regulation of hepcidin and ferroportin expression by lipopolysaccharide in splenic macrophages. *Blood Cells Mol Dis* 35: 47–56, 2005
- 55. Pietrangelo A: Hereditary hemochromatosis. Annu Rev Nutr 26: 251–270, 2006
- Nicolas G, Viatte L, Lou DQ, Bennoun M, Beaumont C, Kahn A, Andrews NC, Vaulont S: Constitutive hepcidin expression prevents iron overload in a mouse model of hemochromatosis. *Nat Genet* 34: 97–101, 2003
- Viatte L, Nicolas G, Lou DQ, Bennoun M, Lesbordes-Brion JC, Canonne-Hergaux F, Schonig K, Bujard H, Kahn A, Andrews NC, Vaulont S: Chronic hepcidin induction causes hyposideremia and alters the pattern of cellular iron accumulation in hemochromatotic mice. *Blood* 107: 2952– 2958, 2006
- Kearney SL, Nemeth E, Neufeld EJ, Thapa D, Ganz T, Weinstein DA, Cunningham MJ: Urinary hepcidin in congenital chronic anemias. *Pediatr Blood Cancer* 48: 57–63, 2007
- 59. Slotki I: Intravenous iron supplementation in the anaemia of renal and cardiac failure: A double-edged sword? *Nephrol Dial Transplant* 20[Suppl 7]: vii16–vii23, 2005