Clinical Cancer Research

Wade T. lams¹ and Christine M. Lovly^{1,2,3}

Abstract

The IGF1R signaling pathway is a complex and tightly regulated network that is critical for cell proliferation, growth, and survival. IGF1R is a potential therapeutic target for patients with many different malignancies. This brief review summarizes the results of clinical trials targeting the IGF1R pathway in patients with breast cancer, sarcoma, and non-small cell lung cancer (NSCLC). Therapeutic agents discussed include both monoclonal antibodies to IGF1R (dalotuzumab, figitumumab, cixutumumab, ganitumab, R1507, AVE1642) and newer IGF1R pathway targeting strategies, including monoclonal antibodies to IGF1 and IGF2 (MEDI-573 and BI 836845) and a small-molecule tyrosine kinase inhibitor of IGF1R (linsitinib). The pullback of trials in patients with breast cancer and NSCLC based on several large negative

trials is noted and contrasted with the sustained success of IGF1R inhibitor monotherapy in a subset of patients with sarcoma. Several different biomarkers have been examined in these trials with varying levels of success, including tumor expression of IGF1R and its pathway components, serum IGF ligand levels, alternate pathway activation, and specific molecular signatures of IGF1R pathway dependence. However, there remains a critical need to define predictive biomarkers in order to identify patients who may benefit from IGF1R-directed therapies. Ongoing research focuses on uncovering such biomarkers and elucidating mechanisms of resistance, as this therapeutic target is currently being analyzed from the bedside to bench. *Clin Cancer Res; 21(19); 4270-7.* ©2015 AACR.

Disclosure of Potential Conflicts of Interest

C.M. Lovly reports receiving commercial research grants from AstraZeneca and Novartis, and is a consultant/advisory board member for ARIAD Pharmaceuticals, Genoptix, Harrison and Star, Novartis, and Sequenom. No potential conflicts of interest were disclosed by the other author.

Editor's Disclosures

The following editor(s) reported relevant financial relationships: P.S. Steeg reports receiving a commercial research grant from Sanofi.

CME Staff Planners' Disclosures

The members of the planning committee have no real or apparent conflicts of interest to disclose.

Learning Objectives

Upon completion of this activity, the participant should be able to explain the unique outcomes of the clinical application of IGF1R inhibition in patients with sarcoma, breast cancer, and non-small cell lung cancer, and the participant should also understand the pathophysiologic basis of current strategies that can identify patients who are more likely to benefit from IGF1R inhibition.

Acknowledgment of Financial or Other Support

This activity does not receive commercial support.

¹Department of Medicine, Vanderbilt University Medical Center, Nashville, Tennessee. ²Department of Cancer Biology, Vanderbilt University Medical Center, Nashville, Tennessee. ³Vanderbilt-Ingram Cancer Center, Nashville, Tennessee.

Corresponding Author: Christine M. Lovly, Vanderbilt-Ingram Cancer Center, 2220 Pierce Avenue, 777 Preston Research Building, Nashville, TN 37232-6307. Phone: 615-936-3457; Fax: 615-343-7602; E-mail: christine.lovly@vanderbilt.edu

doi: 10.1158/1078-0432.CCR-14-2518

©2015 American Association for Cancer Research.

Background

The insulin-like growth factor (IGF) signaling pathway is a complex and tightly regulated network that is critical for cell proliferation and survival (1). This pathway (Fig. 1) is composed of three receptor tyrosine kinases—insulin-like growth factor-1 receptor (IGF1R), insulin-like growth factor-2 receptor (IGF2R), and insulin receptor (INSR)—three ligands (insulin, IGF1, and IGF2; refs. 2, 3), and six serum insulin-like growth factor binding proteins (IGFBP), which serve as regulators of the pathway by



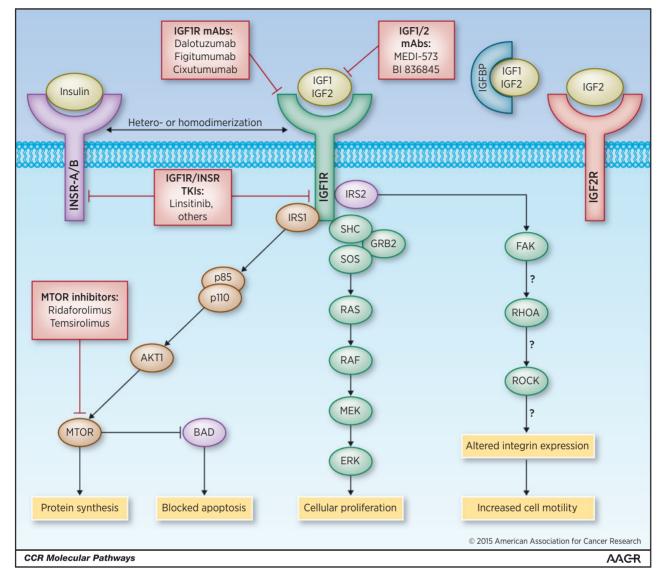


Figure 1.

Schematic representation of the IGFIR signaling network and nodes of therapeutic blockade. The IGFIR signaling pathway is composed of three receptor tyrosine kinases (IGFIR, IGF2R, and INSR), three ligands (insulin, IGF1, and IGF2; formerly known as somatomedins; refs. 1, 2), and six serum insulin-like growth factor binding proteins (IGFBP). The IGFBPs, of which IGFBP3 is the most common, serve as regulators of the pathway by determining the bioavailability of IGF1 and IGF2 ligands (4). Both IGF1 and IGF2 exert their effects through autocrine, paracrine, and endocrine mechanisms, and both can activate the IGFIR pathway. For simplification, IGF1 ligand only is shown binding to IGF1R. IGF1 binding to IGF1R promotes receptor homodimerization or heterodimerization with INSR. Ligand-activated IGF1R first binds to intracellular adaptor proteins, such as IRS1 and SHC. These adaptor proteins transmit signals through the PI3K-AKT1-mTOR pathway and through the MAPK pathway. Activated IGF1R promotes cellular motility through activation of IRS2, which alters integrin expression through poorly understood mechanisms involving the small G protein RHOA, FAK, ROCK, PI3K, and other signaling molecules. Of note, IGF2R is a repository for IGF2, and it has no intracellular signaling activity. IGF2R acts as a tumor suppressor gene, as when IGF2R function is lost, IGF2 is able to bind IGF1R and promote tumorigenesis (17). Targets for potential monotherapy and combinatorial therapeutic strategies are noted in the figure. TKI, tyrosine kinase inhibitor.

determining ligand bioavailability (4). The most prevalent of the IGFBPs is IGFBP3 (5). Both IGF1 and IGF2 exert their effects through autocrine, paracrine, and endocrine mechanisms, and both can activate IGF1R signaling.

IGF1R is a type 2 tyrosine kinase transmembrane receptor that is normally found as a heterotetramer with two alpha and two beta subunits (6, 7). IGF1R binding to IGF1 or IGF2 can occur with IGF1R as a homodimer or as a heterodimer with insulin receptor isoforms A or B (INSR-A, INSR-B; refs. 2, 8). While the heterodimer IGF1R/INSR can bind insulin, it has been shown to preferentially favor IGF1-mediated signaling (9, 10).

Once activated, IGF1R activates numerous downstream pathways within the cell. In order to propagate these signals, ligandactivated IGF1R first binds to intracellular adaptor proteins, predominantly insulin receptor substrate1 (IRS1; ref. 11), although other intracellular proteins, such as SHC1 (12), GAB (13), and CRK (14), can interact with activated IGF1R. These adaptor proteins are necessary for IGF1R to transmit signals downstream in the cell through the PI3K-AKT1-mTOR pathway and through the MAPK pathway. Ligand-activated IGF1R binds to IRS1, which then binds to the p85 regulatory subunit of PI3K, which then transmits signals to AKT1 and MTOR. Activation of the PI3K-AKT1-MTOR pathway results in pleiotropic effects, including inactivation of the proapoptotic protein BAD (15-19). Concurrently, IGF1R binds to SHC, which interacts with growth factor receptor-bound-2 (GRB2)-son-of-sevenless (SOS) to activate the MAPK pathway (14). Finally, activated IGF1R is thought to promote cellular motility through activation of IRS2, which acts to alter integrin expression through poorly understood mechanisms involving the small G protein RHOA, focal adhesion kinase (FAK), and Rho-kinase (ROCK; refs. 15, 16). Of note, IGF2R is a repository for IGF2, and it has no intracellular signaling activity. In this capacity, IGF2R acts as a tumor suppressor gene, as when IGF2R function is lost, IGF2 is able to bind IGF1R and promote tumorigenesis (17).

Serum IGF1 and IGFBP3 levels are normally regulated by the pituitary gland (18, 19). Elevated serum levels of IGF1 and IGF2 and overactivation of the mitogenic, antiapoptotic, and promotility signaling cascades induced by IGF1R have been implicated in many tumor types, including epithelial malignancies (breast, lung, colorectal, prostate, ovarian), mesenchymal tumors (osteosarcoma, rhabdomyosarcoma), and hematologic malignancies (1, 2, 17, 20, 21). Furthermore, IGF1R pathway dysregulation acts as an oncogenic signal in the context of both initial tumorigenesis and resistance to cytotoxic and targeted anticancer therapies (2, 3, 22, 23).

Herein, we focus on the role of the IGF1R pathway in breast cancer, sarcoma, and non-small cell lung cancer (NSCLC), as it is in these three malignancies that IGF1R pathway blockade has been most extensively studied. In patients with breast cancer, it has been noted that the IGF1R pathway has extensive cross-talk with the estrogen receptor (ER) and epidermal growth factor receptor 2 (ERBB2) signaling pathways, and IGF1R has been implicated in resistance to hormonal therapy (24, 25). Furthermore, IGF1R is directly upstream of the PI3K-AKT1-mTOR pathway, which is aberrantly activated in more than half of human breast cancers (26). Preclinical data in sarcoma tumor models have shown that the IGF1R pathway is particularly important in tumor growth, metastasis, and angiogenesis in patients with Ewing sarcoma and rhabdomyosarcoma, leading to the initial application of IGF1R inhibitors in patients with these tumor types (27). Finally, IGF1R protein levels have been shown to be high in NSCLC cell lines and patient samples, both in adenocarcinoma and squamous histologies (28, 29). Also, IGF1R expression is associated with poor prognosis in patients with NSCLC (28). It is worth mentioning that IGF1R expression levels have been evaluated in small cell lung cancer (SCLC); however, we only discuss NSCLC

Numerous therapeutic agents targeting the IGF1R pathway have been developed. These agents include IGF1R monoclonal antibodies (mAb), IGF1R/INSR tyrosine kinase inhibitors (TKI), and, more recently, IGF1- and IGF2-specific mAbs (Fig. 1). Furthermore, several rational combination therapeutic strategies have been used to attempt to more potently inhibit IGF1R signaling. To date, the most widely tested combination strategy involves the use of IGF1R antibodies with mTOR allosteric inhibitors, such as temsirolimus (30) or ridaforolimus (3). There is an established preclinical rationale for this approach, as numer-

ous studies have now shown that mTOR inhibition paradoxically results in activation of the IGF1R pathway (31).

In the following sections, we describe the current state and future directions of the application of IGF1R targeting agents in patients with breast cancer, sarcoma, and NSCLC, with a summary of the high-impact trials provided in Table 1.

Clinical-Translational Advances

IGF1R pathway inhibition in patients with breast cancer

Four different anti-IGF1R mAbs have been tested in early clinical trials involving small numbers of patients with advanced, treatment-refractory breast cancer with largely unimpressive results (5, 32-35). Consequently, three phase I clinical trials assessing the combination of IGF1R mAbs with mTOR inhibitors in patients with advanced, treatment-refractory breast cancer have been completed (19, 36, 37). In a phase I clinical trial with dalotuzumab and the mTOR inhibitor ridaforolimus, a subset of patients with ER-positive (ER⁺), highly proliferative disease was shown to have exceptional responses, experiencing a disease control rate [stable disease (SD) plus partial response (PR)] of 55% (6/11 patients). These promising results created momentum for a recently completed phase II clinical trial involving patients with advanced luminal B breast cancer treated with dalotuzumab, ridaforolimus, and hormonal therapy (NCT01234857; ref. 36). In a phase I clinical trial with cixutumumab and the mTOR inhibitor temsirolimus, among 26 patients with breast cancer (86% with ER⁺ disease), 4 patients (15%) had SD, and no PRs or complete responses (CR) were observed. The results of this trial, in which the median number of prior chemotherapeutic regimens was three, stimulated interest in testing the combination of cixutumumab and temsirolimus in patients with metastatic breast cancer and no more than two prior lines of chemotherapy, but initial trial results have shown no tumor responses (NCT00699491; ref. 37). Finally, unlike combination with mTOR inhibition, the combination of IGF1R inhibition with exemestane or fulvestrant in patients with advanced breast cancer was unsuccessful in a phase II trial (38), halting the application of combination hormonal therapy and IGF1R inhibition in patients with breast cancer.

IGF1R pathway inhibition in patients with sarcoma

Because of successful initial clinical trials in patients with advanced, treatment-refractory sarcoma treated with IGF1R mAbs (5, 32, 34, 35, 39), larger trials with a combined total of 362 patients have been completed (27, 40–42). In summation of these clinical trials, disease stabilization rates have been 16% to 40%, PRs have ranged from 2% to 12% of patients, and 2 of 362 patients have achieved a CR. Overall, the exceptional response of some patients to IGF1R inhibitor monotherapy has led to speculation that a subset of patients with sarcoma, especially Ewing sarcoma, are uniquely dependent on IGF1R signaling (19).

The combination of mTOR inhibition with IGF1R inhibition in patients with advanced sarcoma has yielded results similar to those from IGF1R monotherapy (19, 30, 36, 43, 44), and the combination of IGF1R inhibition with cytotoxic chemotherapy has yielded provocative results in patients with leiomyosarcoma (18, 45). Overall, the clinical trials of anti-IGF1R mAbs in patients with sarcoma have shown occasionally profound responses and disease stabilization rates ranging from 16% to up to 70% when IGF1R mAbs have been combined with mTOR inhibitors (43). Table 1. Published clinical trials involving IGFIR pathway inhibition in patients with breast cancer, sarcoma, or lung cancer

References	Phase	n	Tumor types	Therapy	Disease control rates
Atzori et al., 2011 (32)	I	80	Colorectal (24%), breast (21%), sarcoma (11%), other (43%)	Dalotuzumab (MK-0646)	SD 8%, PR 4%, CR 0%
Di Cosimo et al., 2015 (36)	Ι	87	Breast (26%), colorectal (22%), NSCLC (18%), sarcoma (16%), other (18%)	Dalotuzumab (MK-0646) + ridaforolimus	SD 46%, PR 7%, CR 0%
Higano et al., 2015 (33)	Ι	40	Lung (20%), colon (15%), breast (7.5%), other (57.5%)	Cixutumumab (IMC-A12)	SD 25%, PR 0%, CR 0%
Ma et al., 2013 (37)	Ι	26	Breast (100%); ER positive (86%)	Cixutumumab (IMC-A12) $+$ temsirolimus	SD 15%, PR 0%, CR 0%
Naing et al., 2011 (19)	I	42	Adrenocortical (24%), breast (21%), sarcoma (21%), other (41%)	Cixutumumab (IMC-A12) + temsirolimus	SD 43%, PR 0%, CR 0%
Tolcher et al., 2009 (34)	Ι	53	Sarcoma (42%), other (58%)	Ganitumab (AMG-479)	SD NA, PR 4%, CR 2%
Goto et al., 2012 (47)	Ι	19	NSCLC (100%)	Figitumumab (CP-751,871) + carboplatin and paclitaxel	SD 42%, PR 37%, CR 0%
Molife et al., 2010 (45)	Ι	46	Prostate (48%), esophageal (20%), sarcoma (6.5%), NSCLC (4.3%), other (21.2%)	Figitumumab (CP-751,871) + docetaxel	SD 26%, PR 9%, CR 0%
Murakami et al., 2012 (5)	Ι	19	Breast (21%), gastric (16%), NSCLC (10%), sarcoma (10%), other (43%)	Ganitumab (AMG-479)	SD 37%, PR 0%, CR 0%
Kurzrock et al., 2010 (35)	Ι	35	Sarcoma (51%), lung (5.5%), breast (5.5%), other (38%)	R1507	SD 35%, PR 5%, CR 0%
Macaulay et al., 2013 (18)	I	58	Ovarian (21%), sarcoma (9%), breast (7%), NSCLC (5%), other (58%)	AVE1642 + docetaxel OR gemcitabine/ erlotinib OR doxorubicin	SD 40%-70%, PR 2.5%-20%, CR 0%
Puzanov et al., 2014 (8)	Ι	86	Colorectal (49%), NSCLC (4%), sarcoma (4%), other (43%)	Linsitinib (OSI-906)	SD 36%, PR 1%, CR 0%
Haluska et al., 2014 (50)	Ι	43	Urothelial (46.5%), sarcoma (9%), colorectal (5%), breast (2.5%), NSCLC (2.5%), other (34.5%)	MEDI-573	SD 30%, PR 0%, CR 0%
Haluska et al., 2007 (39)	Ι	24	Colorectal (25%), lung (17%), sarcoma (17%), other (41%)	Figitumumab (CP-751,871)	SD 41%, PR 0%, CR 0%
Olmos et al., 2010 (27)	Ι	29	Sarcoma (100%); Ewing sarcoma (55%)	Figitumumab (CP-751,871)	SD 28.5%, PR 3.5% CR 3.5%
Naing et al., 2012 (30)	Ι	20	Ewing sarcoma (85%), desmoplastic small round cell tumor (15%)	Cixutumumab (IMC-A12) + temsirolimus	SD 25%, PR 0%, CR 10%
Quek et al., 2011 (43)	Ι	21	Sarcoma (90%), adrenal cortical (5%), colorectal (5%)	Figitumumab (CP-751,871) + everolimus	SD 71%, PR 5%, CR 0%
Juergens et al., 2011 (40)	1/11	31 (I) 107 (II)	Sarcoma (100%); Ewing sarcoma (89%)	Figitumumab (CP-751,871)	SD 24%, PR 14%, CR 0%
Schoffski et al., 2013 (42)	II	111	Sarcoma (100%); Ewing sarcoma (18%)	Cixutumumab (IMC-A12)	SD 40%, PR 2%, CR 0%
Schwartz et al., 2013 (44)	II	174	Sarcoma (100%); Ewing sarcoma (15.5%)	Cixutumumab (IMC-A12) + temsirolimus	SD 38%, PR 5%, CR 0%
^D appo et al., 2011 (41)	II	115	Ewing sarcoma (100%)	R1507	SD 16%, PR 9%, CR 1%
(arp et al., 2009 (48)	II	98	NSCLC (100%)	Figitumumab (CP-751,871) + carboplatin and paclitaxel	SD 10%-20%, PR + CR 54% -: 37% corrected
Robertson et al., 2013 (38)	II	63	Breast (100%); ER positive (94%)	Ganitumab (AMG-479) + fulvestrant OR exemestane	SD 27%, PR 8%, CR 0%
2010 (00) Ramalingam et al., 2011 (49)	II	172	NSCLC (100%)	Erlotinib \pm R1507	12-week PFS: 41%/43.5% OS: 8.1 mo/10 mo
Langer et al., 2014 (46)		338	NSCLC (nonadenocarcinoma 100%)	Figitumumab (CP-751,871) + carboplatin and paclitaxel	SD not noted PR + CR 33%

Abbreviations: mo, months; NA, not available; OS, overall survival; PFS, progression-free survival; PR, partial response; SD, stable disease.

However, larger trials are needed to determine the optimal therapeutic strategy (monotherapy vs. combination therapy with mTOR inhibitors) and also to parse out which subsets of patients are most likely to benefit.

IGF1R pathway inhibition in patients with NSCLC

The combination of IGF1R inhibition with cytotoxic chemotherapy has been tested in several large clinical trials in patients with NSCLC (46–48). The most well-studied IGF1R mAb in lung cancer is figitumumab. When the combination of figitumumab, carboplatin, and paclitaxel was used as first-line therapy in 98 patients with advanced NSCLC, the objective response rate (ORR) was initially reported to be 57%, with an additional 10% to 20% of patients experiencing SD (48). These encouraging results prompted the completion of a phase III trial comparing figitumumab plus carboplatin/paclitaxel to carboplatin/paclitaxel alone in patients with treatment-naïve advanced NSCLC. This clinical trial was closed early due to increased rates of serious adverse events and treatment-related deaths in patients treated with figitumumab (46). The phase III trial showed an ORR of 33% for the figitumumab-plus-carboplatin/paclitaxel arm, and rather than the initially reported 54% ORR in the phase II trial, the actual observed rate was 37% (46). The serious adverse events that were observed more commonly in patients receiving figitumumab compared with chemotherapy alone included pneumonia (6% vs. 4%), hyperglycemia (3% vs. <1%), asthenia (3% vs. 1%), and dehydration (4% vs. 1%). The etiologies of the 17 treatment-related deaths in patients treated with figitumumab included pulmonary hemorrhage, pneumonia, septic shock, hypovolemic shock, sepsis in a neutropenic patient, renal failure, hemorrhage, and etiologies listed as cardiorespiratory arrest, toxicity to various agents, and decrease of performance status (46).

The combination of EGFR plus IGF1R inhibition has been tested in a cohort of unselected patients with lung adenocarcinoma or squamous cell carcinoma, but there was no improvement in PFS or OS compared to treatment with EGFR inhibition alone. Importantly, in this study, less than 5% of patients had an *EGFR* mutation, as it was proposed that based on preclinical models IGF1R and EGFR cross-talk was a key mechanism of tumorigenesis and resistance to isolated EGFR inhibition in patients with NSCLC, independent of *EGFR* mutation status (49).

Other therapeutic agents that target the IGF1R pathway

The growing appreciation of INSR-mediated signaling in the IGF pathway has led to two novel strategies to target the IGF1R pathway in patients with advanced breast cancer, sarcoma, and NSCLC: combined IGF1R and insulin receptor inhibition (8) and therapeutic antibodies directed against the IGF1 and IGF2 ligands (50, 51).

On the basis of antitumor activity demonstrated in preclinical models in several tumor types, linsitinib, an oral smallmolecule TKI of IGF1R and INSR, has been evaluated in 86 patients with advanced, treatment-refractory solid tumors. When patients were treated with linsitinib monotherapy, the overall disease stabilization rate was 36%, and 1 patient with melanoma achieved a PR (8). Recently completed phase II trials have evaluated linsitinib combination therapies with paclitaxel in patients with recurrent ovarian cancer (NCT00889382) and with erlotinib in patients with metastatic *EGFR*-mutant NSCLC (NCT01221077). Results from these trials are pending.

MEDI-573, a mAb to both IGF1 and IGF2, has demonstrated the ability to suppress IGF signaling through both IGF1R and INSR-A without affecting normal INSR-B-mediated signaling in cancer cell lines, leading to its use in an early clinical trial in patients with advanced, heavily pretreated solid tumors (50). In this trial, the disease stabilization rate was 30% with no PRs or CRs observed. On the basis of preclinical studies showing increased INSR-A:INSR-B mRNA ratios in tumor tissue from patients with hormone receptor-positive, ERBB2-negative tumors, a phase I/II clinical trial is now under way assessing the impact of MEDI-573 combined with hormonal therapy in this subset of breast cancer patients (NCT01446159; ref. 50).

A second mAb to both IGF1 and IGF2, BI 836845, has been tested in phase I clinical trials involving 81 patients with advanced solid tumors (52, 53). The results have demonstrated tolerability, and 2 patients have experienced a PR, resulting in additional ongoing clinical trials involving the combination of BI 836845 with afatinib in patients with *EGFR*-mutant NSCLC in East Asia (NCT02191891) and in combination with ever-

olimus and exemestane in patients with ER⁺ breast cancer (NCT02123823).

Challenges to clinical applications

The most pressing and as yet undefined challenge to the appropriate clinical application of IGF1R pathway blockade is the identification of predictive markers that are able to identify patients likely to respond to this therapeutic strategy. As the clinical trial data show, some treatment combinations have shown disease stabilization rates of one-quarter to one-half of patients, and there has been some intriguing antitumor activity, especially in patients with sarcoma. However, what is now critically needed is development of predictive biomarkers that can guide future clinical trials in applying this therapeutic strategy to the patient populations most likely to benefit.

The identification of predictive biomarkers can be divided into four main categories that have seen varying levels of success: tumor expression of IGF1R and its pathway components, serum IGF ligand levels, assessment of alternate pathway activation, and attempts at identifying specific molecular signatures of IGF1R pathway dependence.

Pretreatment IGF1R expression as assessed by immunohistochemistry has not consistently been correlated with disease control in heterogeneous groups of patients treated with anti-IGF1R mAbs (32, 35, 44). It is important to note that when tumor expression of a target of a therapeutic agent does not correlate with response, there are many possible etiologies of false-negative signals, including sampling bias, variability in sample handling, limited assay sensitivity and specificity, and tumor mutations between the time a sample is obtained and the time when treatment is administered (32).

In the case of IGF ligand assessments, serum ligand levels have consistently demonstrated predictive value in patients with sarcoma and NSCLC, although it must be noted that the degree of correlation between IGF ligand levels in the serum versus in the tumor microenvironment is unknown. In two clinical trials involving patients with sarcoma treated with IGF1R inhibitor monotherapy, elevated pretreatment and ontreatment serum IGF1 levels were associated with improved OS (40, 41). In patients with NSCLC, both a phase I (47) and a phase III clinical trial (46) have demonstrated improved disease control and overall survival in patients with elevated pretreatment serum total IGF1 (46) and greater elevations in serum IGF1 when treated with figitumumab plus carboplatin/ paclitaxel (46, 47). In contrast to serum IGF1 levels, pretreatment levels and on-treatment changes in serum IGFBP3 have not been associated with disease control in isolated IGF1R inhibition (5), combination IGF1R and mTOR inhibition (19, 37), or IGF1R inhibition in combination with cytotoxic chemotherapy (18, 47).

The assessment of alternative pathway activation mediating resistance to IGF1R-targeted therapies was the impetus for the combination trials described above. However, the interpretation of the heterogeneous responses to combination therapy necessitates a better understanding of the cross-talk between the IGF1R pathway and other important signaling molecules such as EGFR, SRC, and ER (54–56) and downstream molecules such as mTOR, PI3K, and AKT1, which have been shown to mediate IGF1R resistance in preclinical models (57).

Finally, specific gene expression profiles associated with IGF1R sensitivity or resistance have been identified in models of

breast cancer and Ewing sarcoma (58–60). This characteristic IGF1-dependent gene expression profile includes upregulation of transcriptional targets of ER, MAPK3, MAPK1, and components of the PI3K–AKT1–mTOR pathway (58). The assessment of these molecular signatures and alternative pathways mediating IGF1R resistance within the context of the significant clinical trial data described above is an important next step in improving the patient-specific application of IGF1R-targeting therapies (61).

Conclusions

In conclusion, the IGF1R pathway is important in the development and maintenance of many different types of malignancies. Drug development targeting this pathway has taken unique routes by different tumor types, from preferential combination therapy in the case of patients with breast cancer to impressive antitumor efficacy in patients with sarcoma treated with anti-IGF1R monotherapy to a negative phase III trial in combination with cytotoxic chemotherapy in patients with NSCLC. The most pressing needs for the future development of this therapeutic strategy are identifying biomarkers of response by applying a bedside-to-bench approach with the existing clinical trials data, including an in-depth analysis of tumor samples from patients who have responded to IGF1R-directed therapies. These critical

References

- Samani AA, Yakar S, LeRoith D, Brodt P. The role of the IGF system in cancer growth and metastasis: overview and recent insights. Endocr Rev 2007; 28:20–47.
- Casa AJ, Dearth RK, Litzenburger BC, Lee AV, Cui X. The type I insulin-like growth factor receptor pathway: a key player in cancer therapeutic resistance. Front Biosci 2008;13:3273–87.
- Brana I, Berger R, Golan T, Haluska P, Edenfield J, Fiorica J, et al. A parallelarm phase I trial of the humanised anti-IGF-1R antibody dalotuzumab in combination with the AKT inhibitor MK-2206, the mTOR inhibitor ridaforolimus, or the NOTCH inhibitor MK-0752, in patients with advanced solid tumours. Br J Cancer 2014;111:1932–44.
- Massague J, Czech MP. The subunit structures of two distinct receptors for insulin-like growth factors I and II and their relationship to the insulin receptor. J Biol Chem 1982;257:5038–45.
- Murakami H, Doi T, Yamamoto N, Watanabe J, Boku N, Fuse N, et al. Phase 1 study of ganitumab (AMG 479), a fully human monoclonal antibody against the insulin-like growth factor receptor type I (IGF1R), in Japanese patients with advanced solid tumors. Cancer Chemother Pharmacol 2012;70:407–14.
- Steele-Perkins G, Turner J, Edman JC, Hari J, Pierce SB, Stover C, et al. Expression and characterization of a functional human insulin-like growth factor I receptor. J Biol Chem 1988;263:11486–92.
- Tollefsen SE, Stoszek RM, Thompson K. Interaction of the alpha beta dimers of the insulin-like growth factor I receptor is required for receptor autophosphorylation. Biochemistry 1991;30:48–54.
- Puzanov I, Lindsay CR, Goff LW, Sosman JA, Gilbert J, Berlin J, et al. A phase I study of continuous oral dosing of OSI-906, a dual inhibitor of insulinlike growth factor-1 and insulin receptors in patients with advanced solid tumors. Clin Cancer Res 2014;21:701–11.
- 9. Morgan DO, Edman JC, Standring DN, Fried VA, Smith MC, Roth RA, et al. Insulin-like growth factor II receptor as a multifunctional binding protein. Nature 1987;329:301–7.
- MacDonald RG, Pfeffer SR, Coussens L, Tepper MA, Brocklebank CM, Mole JE, et al. A single receptor binds both insulin-like growth factor II and mannose-6-phosphate. Science 1988;239:1134–7.
- 11. Peruzzi F, Prisco M, Dews M, Salomoni P, Grassilli E, Romano G, et al. Multiple signaling pathways of the insulin-like growth factor 1 receptor in protection from apoptosis. Mol Cell Biol 1999;19: 7203–15.

analyses will serve as the foundation to guide the most appropriate application of IGF1R blockade in the clinic.

Disclaimer

The content is solely the responsibility of the authors and does not necessarily represent the official views of the NIH.

Authors' Contributions

Conception and design: W.T. Iams, C.M. Lovly

Development of methodology: C.M. Lovly Acquisition of data (provided animals, acquired and managed patients,

provided facilities, etc.): C.M. Lovly

Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): C.M. Lovly

Writing, review, and/or revision of the manuscript: W.T. lams, C.M. Lovly Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): W.T. lams, C.M. Lovly Study supervision: C.M. Lovly

Grant Support

C.M. Lovly is supported in part by the NIH and the NCI under award numbers R01CA121210 and P01CA129243, a Damon Runyon Clinical Investigator Award, and a LUNGevity Career Development Award.

Received May 21, 2015; revised July 4, 2015; accepted July 21, 2015; published online October 1, 2015.

- Koval AP, Blakesley VA, Roberts CT Jr, Zick Y, Leroith D. Interaction in vitro of the product of the c-Crk-II proto-oncogene with the insulin-like growth factor I receptor. Biochem J 1998;330:923–32.
- 13. Dearth RK, Cui X, Kim HJ, Hadsell DL, Lee AV. Oncogenic transformation by the signaling adaptor proteins insulin receptor substrate (IRS)-1 and IRS-2. Cell Cycle 2007;6:705–13.
- Yamauchi K, Pessin JE. Insulin receptor substrate-1 (IRS1) and Shc compete for a limited pool of Grb2 in mediating insulin downstream signaling. J Biol Chem 1994;269:31107–14.
- Stewart AJ, Johnson MD, May FE, Westley BR. Role of insulin-like growth factors and the type I insulin-like growth factor receptor in the estrogenstimulated proliferation of human breast cancer cells. J Biol Chem 1990;265:21172–8.
- Lee AV, Jackson JG, Gooch JL, Hilsenbeck SG, Coronado-Heinsohn E, Osborne CK, et al. Enhancement of insulin-like growth factor signaling in human breast cancer: estrogen regulation of insulin receptor substrate-1 expression *in vitro* and *in vivo*. Mol Endocrinol 1999;13:787–96.
- Rowinsky EK, Youssoufian H, Tonra JR, Solomon P, Burtrum D, Ludwig DL. IMC-A12, a human IgG1 monoclonal antibody to the insulin-like growth factor I receptor. Clin Cancer Res 2007;13:5549s–55s.
- Macaulay VM, Middleton MR, Protheroe AS, Tolcher A, Dieras V, Sessa C, et al. Phase I study of humanized monoclonal antibody AVE1642 directed against the type 1 insulin-like growth factor receptor (IGF-1R), administered in combination with anticancer therapies to patients with advanced solid tumors. Ann Oncol 2013;24:784–91.
- 19. Naing A, Kurzrock R, Burger A, Gupta S, Lei X, Busaidy N, et al. Phase I trial of cixutumumab combined with temsirolimus in patients with advanced cancer. Clin Cancer Res 2011;17:6052–60.
- Singh P, Alex JM, Bast F. Insulin receptor (IR) and insulin-like growth factor receptor 1 (IGF-1R) signaling systems: novel treatment strategies for cancer. Med Oncol 2014;31:805.
- Avnet S, Sciacca L, Salerno M, Gancitano G, Cassarino MF, Longhi A, et al. Insulin receptor isoform A and insulin-like growth factor II as additional treatment targets in human osteosarcoma. Cancer Res 2009;69:2443–52.
- 22. Pollak M. Insulin and insulin-like growth factor signalling in neoplasia. Nat Rev Cancer 2008;8:915–28.
- Gao J, Chang YS, Jallal B, Viner J. Targeting the insulin-like growth factor axis for the development of novel therapeutics in oncology. Cancer Res 2012;72:3–12.

- Gee JM, Robertson JF, Gutteridge E, Ellis IO, Pinder SE, Rubini M, et al. Epidermal growth factor receptor/HER2/insulin-like growth factor receptor signalling and oestrogen receptor activity in clinical breast cancer. Endocr Relat Cancer 2005;12:S99–S111.
- 25. Fox EM, Miller TW, Balko JM, Kuba MG, Sanchez V, Smith RA, et al. A kinome-wide screen identifies the insulin/IGF-I receptor pathway as a mechanism of escape from hormone dependence in breast cancer. Cancer Res 2011;71:6773–84.
- 26. Miller TW, Balko JM, Arteaga CL. Phosphatidylinositol 3-kinase and antiestrogen resistance in breast cancer. J Clin Oncol 2011;29:4452-61.
- 27. Olmos D, Postel-Vinay S, Molife LR, Okuno SH, Schuetze SM, Paccagnella ML, et al. Safety, pharmacokinetics, and preliminary activity of the anti-IGF-1R antibody figitumumab (CP-751,871) in patients with sarcoma and Ewing's sarcoma: a phase 1 expansion cohort study. Lancet Oncol 2010; 11:129–35.
- Gong Y, Yao E, Shen R, Goel A, Arcila M, Teruya-Feldstein J, et al. High expression levels of total IGF-1R and sensitivity of NSCLC cells in vitro to an anti-IGF-1R antibody (R1507). PLoS ONE 2009;4:e7273.
- 29. Kim YH, Sumiyoshi S, Hashimoto S, Masago K, Togashi Y, Sakamori Y, et al. Expressions of insulin-like growth factor receptor-1 and insulin-like growth factor binding protein 3 in advanced non-small-cell lung cancer. Clin Lung Cancer 2012;13:385–90.
- 30. Naing A, LoRusso P, Fu S, Hong DS, Anderson P, Benjamin RS, et al. Insulin growth factor-receptor (IGF-1R) antibody cixutumumab combined with the mTOR inhibitor temsirolimus in patients with refractory Ewing's sarcoma family tumors. Clin Cancer Res 2012;18:2625–31.
- O'Reilly KE, Rojo F, She QB, Solit D, Mills GB, Smith D, et al. mTOR inhibition induces upstream receptor tyrosine kinase signaling and activates Akt. Cancer Res 2006;66:1500–8.
- 32. Atzori F, Tabernero J, Cervantes A, Prudkin L, Andreu J, Rodriguez-Braun E, et al. A phase I pharmacokinetic and pharmacodynamic study of dalotuzumab (MK-0646), an anti-insulin-like growth factor-1 receptor monoclonal antibody, in patients with advanced solid tumors. Clin Cancer Res 2011;17:6304–12.
- 33. Higano CS, Berlin J, Gordon M, LoRusso P, Tang S, Dontabhaktuni A, et al. Safety, tolerability, and pharmacokinetics of single and multiple doses of intravenous cixutumumab (IMC-A12), an inhibitor of the insulin-like growth factor-I receptor, administered weekly or every 2 weeks in patients with advanced solid tumors. Invest New Drugs 2015;33:450–62.
- 34. Tolcher AW, Sarantopoulos J, Patnaik A, Papadopoulos K, Lin CC, Rodon J, et al. Phase I, pharmacokinetic, and pharmacodynamic study of AMG 479, a fully human monoclonal antibody to insulin-like growth factor receptor 1. J Clin Oncol 2009;27:5800–7.
- 35. Kurzrock R, Patnaik A, Aisner J, Warren T, Leong S, Benjamin R, et al. A phase I study of weekly R1507, a human monoclonal antibody insulin-like growth factor-I receptor antagonist, in patients with advanced solid tumors. Clin Cancer Res 2010;16:2458–65.
- 36. Di Cosimo S, Sathyanarayanan S, Bendell JC, Cervantes A, Stein MN, Brana I, et al. Combination of the mTOR inhibitor ridaforolimus and the anti-IGF1R monoclonal antibody dalotuzumab: preclinical characterization and phase I clinical trial. Clin Cancer Res 2015;21:49–59.
- Ma CX, Suman VJ, Goetz M, Haluska P, Moynihan T, Nanda R, et al. A phase I trial of the IGF-1R antibody cixutumumab in combination with temsirolimus in patients with metastatic breast cancer. Breast Cancer Res Treat 2013;139:145–53.
- Robertson JF, Ferrero JM, Bourgeois H, Kennecke H, de Boer RH, Jacot W, et al. Ganitumab with either exemestane or fulvestrant for postmenopausal women with advanced, hormone-receptor-positive breast cancer: a randomised, controlled, double-blind, phase 2 trial. Lancet Oncol 2013;14: 228–35.
- Haluska P, Shaw HM, Batzel GN, Yin D, Molina JR, Molife LR, et al. Phase I dose escalation study of the anti insulin-like growth factor-I receptor monoclonal antibody CP-751,871 in patients with refractory solid tumors. Clin Cancer Res 2007;13:5834–40.
- Juergens H, Daw NC, Geoerger B, Ferrari S, Villarroel M, Aerts I, et al. Preliminary efficacy of the anti-insulin-like growth factor type 1 receptor antibody figitumumab in patients with refractory Ewing sarcoma. J Clin Oncol 2011;29:4534–40.
- 41. Pappo AS, Patel SR, Crowley J, Reinke DK, Kuenkele KP, Chawla SP, et al. R1507, a monoclonal antibody to the insulin-like growth factor 1 receptor, in patients with recurrent or refractory Ewing sarcoma family of tumors:

results of a phase II Sarcoma Alliance for Research through Collaboration study. J Clin Oncol 2011;29:4541-7.

- 42. Schoffski P, Adkins D, Blay JY, Gil T, Elias AD, Rutkowski P, et al. An openlabel, phase 2 study evaluating the efficacy and safety of the anti-IGF-1R antibody cixutumumab in patients with previously treated advanced or metastatic soft-tissue sarcoma or Ewing family of tumours. Eur J Cancer 2013;49:3219–28.
- 43. Quek R, Wang Q, Morgan JA, Shapiro GI, Butrynski JE, Ramaiya N, et al. Combination mTOR and IGF-1R inhibition: phase I trial of everolimus and figitumumab in patients with advanced sarcomas and other solid tumors. Clin Cancer Res 2011;17:871–9.
- Schwartz GK, Tap WD, Qin LX, Livingston MB, Undevia SD, Chmielowski B, et al. Cixutumumab and temsirolimus for patients with bone and softtissue sarcoma: a multicentre, open-label, phase 2 trial. Lancet Oncol 2013;14:371–82.
- 45. Molife LR, Fong PC, Paccagnella L, Reid AH, Shaw HM, Vidal L, et al. The insulin-like growth factor-I receptor inhibitor figitumumab (CP-751,871) in combination with docetaxel in patients with advanced solid tumours: results of a phase lb dose-escalation, open-label study. Br J Cancer 2010; 103:332–9.
- 46. Langer CJ, Novello S, Park K, Krzakowski M, Karp DD, Mok T, et al. Randomized, phase III trial of first-line figitumumab in combination with paclitaxel and carboplatin versus paclitaxel and carboplatin alone in patients with advanced non-small-cell lung cancer. J Clin Oncol 2014; 32:2059–66.
- 47. Goto Y, Sekine I, Tanioka M, Shibata T, Tanai C, Asahina H, et al. Figitumumab combined with carboplatin and paclitaxel in treatmentnaive Japanese patients with advanced non-small cell lung cancer. Invest New Drugs 2012;30:1548–56.
- 48. Karp DD, Paz-Ares LG, Novello S, Haluska P, Garland L, Cardenal F, et al. Phase II study of the anti-insulin-like growth factor type 1 receptor antibody CP-751,871 in combination with paclitaxel and carboplatin in previously untreated, locally advanced, or metastatic non-small-cell lung cancer. J Clin Oncol 2009;27:2516–22.
- Ramalingam SS, Spigel DR, Chen D, Steins MB, Engelman JA, Schneider CP, et al. Randomized phase II study of erlotinib in combination with placebo or R1507, a monoclonal antibody to insulin-like growth factor-1 receptor, for advanced-stage non-small-cell lung cancer. J Clin Oncol 2011;29:4574–80.
- Haluska P, Menefee M, Plimack ER, Rosenberg J, Northfelt D, LaVallee T, et al. Phase I dose-escalation study of MEDI-573, a bispecific, antiligand monoclonal antibody against IGFI and IGFII, in patients with advanced solid tumors. Clin Cancer Res 2014;20:4747–57.
- Friedbichler K, Hofmann MH, Kroez M, Ostermann E, Lamche HR, Koessl C, et al. Pharmacodynamic and antineoplastic activity of BI 836845, a fully human IGF ligand-neutralizing antibody, and mechanistic rationale for combination with rapamycin. Mol Cancer Ther 2014;13:399–409.
- 52. Lin CC, Chang K-Y, Huang DC, Marriott V, Van Beijsterveldt L, Chen L-T, et al. A phase I dose escalation study of weekly BI 836845, a fully human, affinity-optimized, insulin-like growth factor (IGF) ligand neutralizing antibody, in patients with advanced solid cancers. J Clin Oncol 32:5s, 2014 (suppl; abstr 2617).
- 53. Rihawi K, Ong M, Michalarea V, Bent L, Buschke S, Bogenrieder T, et al. Phase I dose escalation study of 3-weekly BI 836845, a fully human, affinity optimized, insulin-like growth factor (IGF) ligand neutralizing antibody, in patients with advanced solid tumors. J Clin Oncol 32:5s, 2014 (suppl; abstr 2622).
- Huang F, Hurlburt W, Greer A, Reeves KA, Hillerman S, Chang H, et al. Differential mechanisms of acquired resistance to insulin-like growth factor-i receptor antibody therapy or to a small-molecule inhibitor, BMS-754807, in a human rhabdomyosarcoma model. Cancer Res 2010; 70:7221–31.
- Hou X, Huang F, Macedo LF, Harrington SC, Reeves KA, Greer A, et al. Dual IGF-1R/InsR inhibitor BMS-754807 synergizes with hormonal agents in treatment of estrogen-dependent breast cancer. Cancer Res 2011;71: 7597–607.
- Haluska P, Carboni JM, TenEyck C, Attar RM, Hou X, Yu C, et al. HER receptor signaling confers resistance to the insulin-like growth factor-I receptor inhibitor, BMS-536924. Mol Cancer Ther 2008;7: 2589–98.

- Cao L, Yu Y, Darko I, Currier D, Mayeenuddin LH, Wan X, et al. Addiction to elevated insulin-like growth factor I receptor and initial modulation of the AKT pathway define the responsiveness of rhabdomyosarcoma to the targeting antibody. Cancer Res 2008;68:8039–48.
 Creighton CJ, Casa A, Lazard Z, Huang S, Tsimelzon A, Hilsenbeck SG, et al.
- Creighton CJ, Casa A, Lazard Z, Huang S, Tsimelzon A, Hilsenbeck SG, et al. Insulin-like growth factor-I activates gene transcription programs strongly associated with poor breast cancer prognosis. J Clin Oncol 2008;26:4078–85.
- Litzenburger BC, Creighton CJ, Tsimelzon A, Chan BT, Hilsenbeck SG, Wang T, et al. High IGF-IR activity in triple-negative breast cancer cell lines

and tumorgrafts correlates with sensitivity to anti-IGF-IR therapy. Clin Cancer Res 2011;17:2314-27.

- Garofalo C, Mancarella C, Grilli A, Manara MC, Astolfi A, Marino MT, et al. Identification of common and distinctive mechanisms of resistance to different anti-IGF-IR agents in Ewing's sarcoma. Mol Endocrinol 2012; 26:1603–16.
- King H, Aleksic T, Haluska P, Macaulay VM. Can we unlock the potential of IGF-1R inhibition in cancer therapy? Cancer Treat Rev 2014;40: 1096–105.