

Molecular phylogeny of the stingless bees (Apidae, Apinae, Meliponini) inferred from mitochondrial 16S rDNA sequences

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Abstract – Sequence data from the mitochondrial 16S rDNA of 34 species from 22 genera of stingless bees plus outgroup sequences from 11 species of other corbiculate bees were used to investigate the phylogenetic relationships among the Meliponini. Equally weighted parsimony and maximum likelihood analyses were performed. Four main clades were recognized in the parsimony consensus tree: (A) *Hypotrigona*, (B) *Austroplebeia*, (C) remaining African genera (*Plebeina*, *Meliplebeia*, and *Axestotrigona*) plus the two Oriental genera (*Lepidotrigona* and *Heterotrigona*), and (D) Neotropical genera. The African genus *Hypotrigona* was placed as the most basal branch in the tribe, followed by *Austroplebeia* as the sister group of other two major clades (C and D). Our results did not support traditional groups with intercontinental composition, e.g. *Trigona* sensu lato or *Plebeia* sensu lato.

stingless bees / Meliponini / 16S rDNA / phylogeny

1. INTRODUCTION

Meliponini is a tribe of highly social stingless honey bees. Although the tribe has a pantropical distribution, most species are restricted to the Neotropical region (ca. 75% of the approximately 500 known species). Meliponini are easily distinguished from other tribes of the subfamily Apinae by the reduced wing venation, presence of a penicillum (a brush of long setae on the outer, apical surface of the hind tibia), and a vestigial sting (Wille, 1979, 1983; Michener, 1990, 2000), among

many other features. A major character that separates Meliponini from other corbiculate tribes is the absence of an auricle on the hind basitarsus. The species included in the tribe show considerable variety in size, nesting sites, and nest architecture (Michener, 1974; Sakagami, 1982). The female castes, queen and worker, differ strikingly in morphology (Michener, 1974; Wille, 1979).

Different generic classifications have been proposed for this tribe over the past 50 years (e.g. Michener, 1944; Schwarz, 1948; Moure, 1951, 1961). The first cladistic hypothesis

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based on an analysis of 17 morphological characters was proposed by Michener (1990). He recognized a total of 35 taxa at the genus and subgenus level. Camargo and Pedro (1992a) reanalyzed the same characters and proposed an alternative hypothesis in which 55 genera were recognized. More recently a new genus, *Meliwillea*, endemic to Central American cloud forests was described by Roubik et al. (1997). Both phylogenetic hypotheses recognize groups with intercontinental composition. *Trigona* sensu Michener (1990, 2000), for example, includes Neotropical, as well as Indomalayan taxa.

Michener's and Camargo and Pedro's hypotheses have several major disagreements. In Michener's (1990) analysis, the Neotropical genus *Melipona* comes out as the sister group of the remaining Meliponini, and the Australian genus *Austroplebeia* appears related to the African forms. In Camargo and Pedro (1992a), these two genera are placed among several Neotropical genera related to *Plebeia*. Moreover, doubtful relationships appear among the minute *Hypotrigona*-like bees that include Neotropical, African and Indomalayan forms and which are placed as closely related taxa in Michener's cladograms. According to Moure (1961), character convergence associated with reduction of size could have occurred within this group.

Despite these recent studies, the relationships among the meliponine genera remain poorly understood. The utility of DNA sequence data to resolve Meliponini phylogenetic relationships is still to be evaluated. Sequence data from the mitochondrial 16S rRNA gene have been used previously at higher-level phylogenetic studies within the order Hymenoptera (Cameron, 1991, 1993; Derr et al. 1992a, b; Downton and Austin, 1994, 2001; Downton et al., 1997; Koulianos et al., 1999; Cameron and Mardulyn, 2001). In the present study, we used 16S rDNA sequence data to investigate the phylogenetic relationships among 22 of the 56 genera of Meliponini recognized by Camargo and Pedro (1992a) and Roubik et al. (1997).

2. MATERIALS AND METHODS

We examined sequence data from the mitochondrial 16S rDNA of 34 species of Meliponini, repre-

senting 22 genera. Available GenBank sequences from four species of *Apis*, four of *Bombus* and three of euglossine bees (*Euglossa*, *Eufriesea* and *Eulaema*) were included as outgroups in the analysis. A list of the species analyzed is shown in Table 1.

Total nucleic acid extraction followed the protocol of Sheppard and McPherson (1991), with a few modifications. For each extraction, a single bee thorax was homogenized in 100 μ L of buffer in a 1.5 mL microcentrifuge tube. The final pellet was resuspended in 50 μ L of TE buffer (10 mM Tris, 0.1 mM EDTA, pH 8.0). DNA templates were amplified by polymerase chain reaction on an ERI-COMP thermocycler performing 40 amplification cycles (94 $^{\circ}$ C, 1 min; 42 $^{\circ}$ C, 1.5 min; and 64 $^{\circ}$ C, 1.5 min), followed by a final extension step at 72 $^{\circ}$ C for 5 min. PCR amplifications were carried out in 25 μ L total reaction volumes using 2.5 μ L of reaction buffer, 4 μ L of dNTP mixture (final concentration of 200 μ M each), 1 μ L of each primer, 1 unit of Taq polymerase (Promega), 1 or 2 μ L of DNA template, and sterile water.

For DNA amplification, we designed the following two primers based on available sequences from the honey bee 16S rDNA: LR13943F 5'-CACCTGTTTATCAAAAACAT-3' and LR13392R 5'-CGTCGATTTGAACTCAAATC-3'. Sequences corresponding to nucleotides 13392 through 13943 of the honey bee mitochondrial genome (Crozier and Crozier, 1993) were obtained by direct sequencing of double-stranded amplified DNA fragments of about 550 bp. Sequencing followed the Sanger method (Sanger et al., 1977) using T7 Sequenase version 2.0 (Amersham Co.). Electrophoresis was carried out on 6% polyacrylamide sequencing gels (Long Ranger Gel - AT Biochem, Malvern, PA), 0.5X and 1X TBE upper and lower tray respectively, 2400 V at 50 $^{\circ}$ C for 2 hours. Afterwards, gels were rinsed in deionized water, dried and exposed to X-ray film at room temperature for 24 hours. The new sequences obtained were deposited in GenBank (<http://www.ncbi.nlm.nih.gov>) under accession numbers AF343091-AF343118 (see Tab. 1).

Alignment of the sequences was performed using the program CLUSTAL W (Thompson et al., 1994) and adjusted by hand to optimize the alignments of gaps. Gaps were treated as missing data in the parsimony analyses. Maximum parsimony analysis was performed using the program PAUP* 4 b8a (Swofford, 2001) for Windows. Branch support was evaluated by jackknife [37% of characters removed per search, as suggested by Farris et al. (1996)] and bootstrap proportions, calculated on a data matrix with invariant and autapomorphic positions removed. The following command strings were used in PAUP

Table I. List of the species, origin and type of sample used for DNA extraction or source of sequence data used in the analysis. E = ethanol-preserved sample; D = dry sample.

Taxon	Abbreviated name	Accession Number	Locality data or Original reference
<i>Austroplebeia australis</i>	<i>A. australis</i>	AF343112	Duaringa, Australia [E]
<i>Austroplebeia symei</i>	<i>A. symei</i>	AF343113	GinGin, Australia [E]
<i>Axestotrigona togoensis</i>	<i>Axestotrigona</i>	AF343117	Harare, Zimbabwe [D]
<i>Friesella schrotkyi</i>	<i>Friesella</i>	AF343099	Viçosa, MG, Brazil [E]
<i>Frieseomelitta doederleini</i>	<i>Frieseomelitta</i>	AF343101	Bom Jesus, PI, Brazil [E]
<i>Heterotrigona itama</i>	<i>Heterotrigona</i>	AF343115	Chartaburi, Thailand [D]
<i>Hypotrigona gribodoi</i>	<i>Hypotrigona</i>	AF343114	Northern Prov., South Africa [D]
<i>Lepidotrigona ventralis</i>	<i>Lepidotrigona</i>	AF343118	Chiang Mai, Thailand [D]
<i>Lestrimelitta sp.*</i>	<i>Lestrimelitta</i>	AF181586	Cameron and Mardulyn (2001)
<i>Lestrimelitta limao</i>	<i>L. limao</i>	AF343106	Ribeirão Preto, SP, Brazil [E]
<i>Meliplebeia becarii</i>	<i>Meliplebeia</i>	AF343109	Mgahinga, Uganda [D]
<i>Melipona compressipes</i>	<i>M. compressipes</i>	AF181589	Cameron and Mardulyn (2001)
<i>M. quadrifasciata</i>	<i>M. quadrifasciata</i>	AF343100	Viçosa, MG, Brazil [E]
<i>Meliwillea bivea</i>	<i>Meliwillea</i>	AF343108	Zurqui, Costa Rica [D]
<i>Nannotrigona testaceicornis</i>	<i>Nannotrigona</i>	AF343102	Viçosa, MG, Brazil [E]
<i>Oxytrigona tataira</i>	<i>Oxytrigona</i>	AF343104	Cajuru, SP, Brazil [E]
<i>Paratrigona subnuda</i>	<i>Paratrigona</i>	AF343105	C. do Castelo, ES, Brazil [D]
<i>Partamona helleri</i>	<i>Partamona</i>	AF343098	Viçosa, MG, Brazil [E]
<i>Plebeia droryana</i>	<i>P. droryana</i>	AF343092	Viçosa, MG, Brazil [E]
<i>P. julianii</i>	<i>P. julianii</i>	AF343096	Londrina, PR, Brazil [E]
<i>P. nigriceps</i>	<i>P. nigriceps</i>	AF343094	Pres. Prudente, SP, Brazil [E]
<i>P. remota</i>	<i>P. remota</i>	AF343097	Cunha, SP, Brazil [E]
<i>P. saiqui</i>	<i>P. saiqui</i>	AF343093	Cunha, SP, Brazil [E]
<i>P. wittmanni</i>	<i>P. wittmanni</i>	AF343095	Canguçu, RS, Brazil [E]
<i>Plebeia denoiti</i>	<i>Plebeina</i>	AF343116	Harare, Zimbabwe [D]
<i>Scaptotrigona depilis</i>	<i>S. depilis</i>	AF181588	Cameron and Mardulyn (2001)
<i>S. luteipennis</i>	<i>S. luteipennis</i>	L22900	Cameron (1993)
<i>S. subobscuripennis</i>	<i>S. subobscuripennis</i>	AF343103	Zurqui, Costa Rica [D]
<i>Scaura latitarsis</i>	<i>Scaura</i>	AF343111	Ribeirão Preto, SP, Brazil [E]
<i>Schwarziana quadripunctata</i>	<i>Schwarziana</i>	AF343110	Viçosa, MG, Brazil [E]
<i>Tetragonisca angustula</i>	<i>Tetragonisca</i>	AF343107	Viçosa, MG, Brazil [E]
<i>Trigona amalthea</i>	<i>T. amalthea</i>	AF214667	Tanaka et al. (2001)
<i>T. fuscipennis</i>	<i>T. fuscipennis</i>	AF343091	Ribeirão Preto, SP, Brazil [E]
<i>T. hypogea</i>	<i>T. hypogea</i>	L22901	Cameron (1993)
<i>Apis cerana</i>	<i>A. cerana</i>	AF153089	Tanaka et al. (2001)
<i>A. dorsata</i>	<i>A. dorsata</i>	AF153098	Tanaka et al. (2001)
<i>A. florea</i>	<i>A. florea</i>	L22894	Cameron (1993)
<i>A. mellifera</i>	<i>A. mellifera</i>	AF214666	Tanaka et al. (2001)
<i>Bombus avinoviellus</i>	<i>B. avinoviellus</i>	L22897	Cameron (1993)
<i>B. pennsylvanicus</i>	<i>B. pennsylvanicus</i>	L22896	Cameron (1993)
<i>B. terrestris</i>	<i>B. terrestris</i>	AF181582	Cameron and Mardulyn (2001)
<i>B. variabilis</i>	<i>B. variabilis</i>	L22898	Cameron (1993)
<i>Eufriesea caerulescens</i>	<i>E. caerulescens</i>	L22904	Cameron (1993)
<i>Euglossa imperialis</i>	<i>E. imperialis</i>	AF181584	Cameron and Mardulyn (2001)
<i>Eulaema polychroma</i>	<i>E. polychroma</i>	L22903	Cameron (1993)

* Cited as *Lestrimelitta limao* by Cameron and Mardulyn (2001), but this species does not occur in Central America, being restricted to southeastern Brazil.

to obtain, respectively, the jackknife and bootstrap proportions: “*jackknife pctdelete = 37 nreps = 100 search = heuristic/ nreps = 2 addseq = random*” and “*bootstrap nreps = 100 search = heuristic/ nreps = 2 addseq = random*”.

Model selection for the maximum likelihood (ML) analyses was carried out using the program Modeltest Version 3.06 (Posada and Crandall, 1998, 2001). The likelihood settings in PAUP were adjusted to implement a transversional model with gamma distributed rates (TVM+G), the best-fit model selected by hierarchical likelihood ratio tests in Modeltest. The following PAUP block was added at the end of the input file: *lset base = (0.4306 0.0499 0.1209) nst = 6 rmat = (1.8757 7.0389 5.0851 3.8966 7.0389) rates = gamma shape = 0.3421 pinvar = 0.*

3. RESULTS

The length of the 16S rDNA fragments sequenced in the present study and used in the phylogenetic analyses varied from 320 (for *Hypotrigona*) to 421 bp (for *Frieseomelitta*). Most species had between 410 and 420 bp sequenced and only three species had less than 390 bp. The length of the sequences taken from GenBank varied from 458 (*T. amalthea*) to 479 bp (*Euglossa imperialis*). The partial sequences of the 45 taxa resulted in a data matrix of 462 aligned bases. The aligned sequences can be obtained from the first author upon request. Variation among the taxa occurred in 259 base positions, with 72 characters being autapomorphic. A high A+T content was observed (81.4%). The uncorrected sequence divergence among Meliponini ranged from 1 to 19%.

Exclusion of autapomorphies and invariant characters from the data set left 187 informative sites for the parsimony analysis. Equally weighted parsimony analysis resulted in 6 most parsimonious trees of 901 steps with a consistency index of 0.33 and a retention index of 0.55. The strict consensus tree is shown in Figure 1. Previous hypotheses of relationships among the corbiculate Apinae were not taken into consideration for tree rooting: the tree root was simply placed between the ingroup and outgroup. Previous molecular and morphological studies on the phylogeny of the corbiculate Apinae (Apini, Euglossini, Bombini and Meliponini) have produced con-

flicting results and readers interested in these relationships should consult recent studies on this subject, e.g. those by Ascher et al. (2001), Cameron and Mardulyn (2001), Engel (2001a, b), Schultz et al. (2001) and Noll (2002). Therefore, the arrangements shown here for the outgroup taxa should not be taken as evidence for or against any of these hypotheses, especially because our taxa sampling is rather inadequate for this purpose.

Meliponini came out as a monophyletic group, with a high jackknife and bootstrap support (100%). Four main clades within Meliponini can be recognized in the consensus tree: (A) *Hypotrigona*, (B) *Austroplebeia*, (C) the remaining African genera (*Plebeina*, *Meliplebeia* and *Axestotrigona*) plus the two Oriental genera (*Lepidotrigona* and *Heterotrigona*) and (D) the Neotropical genera. However, none of the basal arrangements are supported by jackknife and bootstrap values higher than 50%.

The maximum likelihood analysis resulted in one tree (Fig. 2) with a $-\ln$ score of 4817.06398. The relationships within Meliponini differed in several aspects from those reconstructed under parsimony, especially for the arrangements at the base of the tree. Despite these incongruencies, the large Neotropical clade and several other smaller groups were recovered in both analyses (compare Figs. 1 and 2).

The main difference between the results from the parsimony and ML analyses was the basal group formed by *Hypotrigona*, *Lepidotrigona* and *Austroplebeia* in the ML tree. However, *Hypotrigona*, *Lepidotrigona* and *Austroplebeia symei* contributed the shortest sequences in the dataset, (320, 340 and 338 bp respectively). Additional sequence data from multiple genes will be needed to clearly resolve these basal relationships. Considering that morphological and biogeographical evidence also supports a closer relationship between *Lepidotrigona* and *Heterotrigona* than for *Lepidotrigona* and *Hypotrigona*, our discussion focuses on the parsimony results.

4. DISCUSSION

The analyses of the 16S rDNA partial sequences conducted in the present study

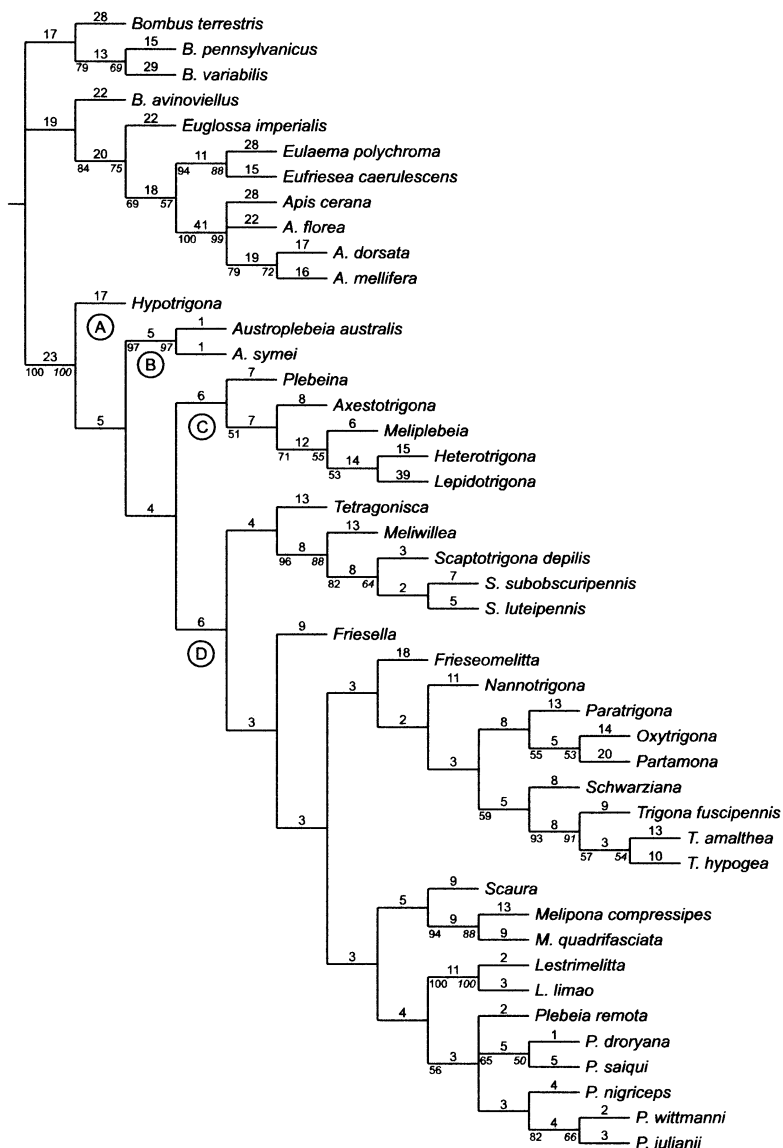


Figure 1. Strict consensus of the 6 most parsimonious trees. The four main clades of the ingroup are indicated within the circles [A. *Hypotrigona* (Africa), B. *Austroplebeia* (Australia), C. remaining African and Oriental genera, D. Neotropical genera]. The numbers above lines are branch lengths and those below are jackknife (left) and bootstrap values above 50% (right). Tree length = 907, CI = 0.33, RI = 0.55.

indicated phylogenetic relationships for the major groups of Meliponini that differ significantly from previous hypotheses based on adult worker morphology. Traditional groups with intercontinental composition, e.g. *Plebeia* sensu lato (Moore, 1951, 1961 –

including Neotropical and African genera) or *Trigona* sensu lato (Michener, 1990, 2000 – including Neotropical and Oriental genera) were not recovered in our analyses, nor were sister group relationships between Neotropical and African or Oriental faunas.

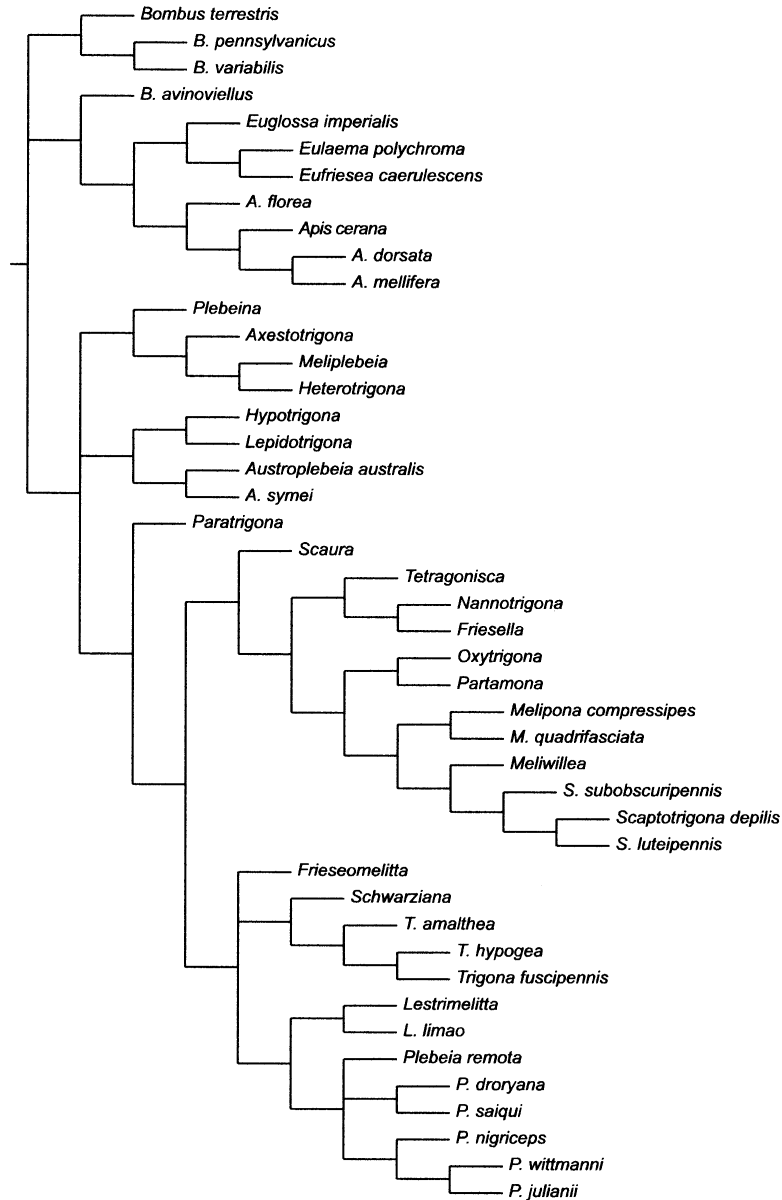


Figure 2. Maximum-likelihood tree obtained under a transversional model with gamma distributed rates (TVM+G). $-\ln$ score = 4817.06398.

For comparison, the stingless bee taxa used in the present study were arranged accordingly to the hypotheses presented by Michener (1990) (Fig. 3) and Camargo and Pedro (1992a) (Fig. 4). Optimizing the 16S data set on these trees revealed that their arrangements

are considerably less parsimonious (613 and 596 steps in length, respectively) when compared to the hypothesis obtained with the 16S sequences (515 steps).

The basalmost position for the African genus *Hypotrigona* (branch A, Fig. 1) supports

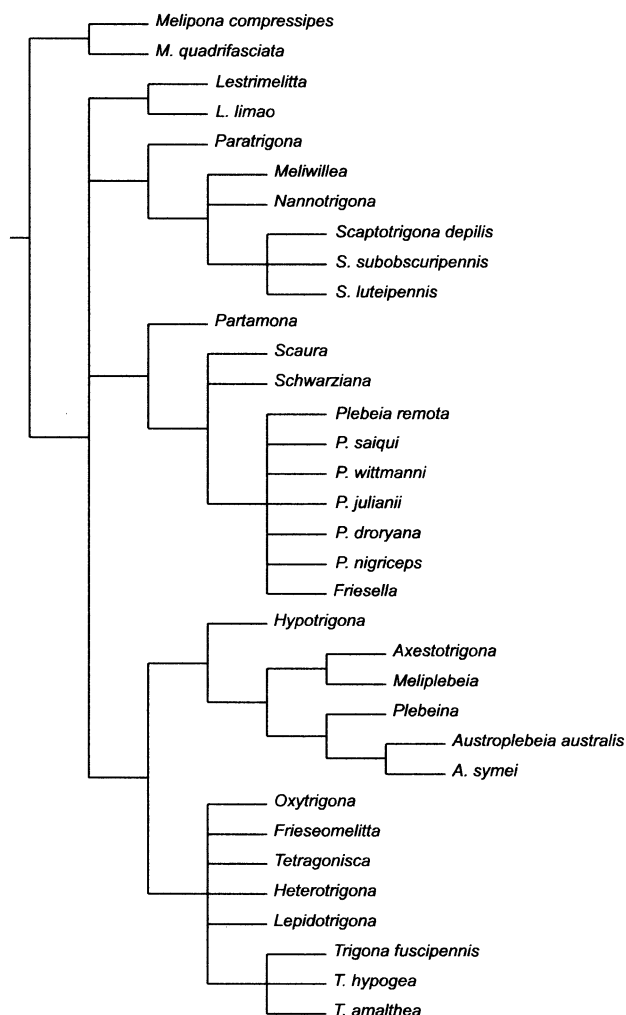


Figure 3. Tree showing relationships among stingless bee taxa used in the present study arranged according to the hypothesis presented by Michener (1990).

the previous suggestion by Michener (1990) that this genus could be the sister group to all the other genera of Meliponini, a hypothesis based on the putatively plesiomorphic mandibular and sternal features of the male. Unfortunately, other genera likely to come out as basal groups within Meliponini, as e.g. *Liotrigona* (African), *Pariotrigona* (Oriental) and *Trigonisca* (Neotropical), could not be included in our analysis due to lack of adequate material for DNA extraction. Although all Meliponini lacks the mesoscutal supra-alar carina (Michener, 1990), these

genera contain the only stingless bee species in which the groove associated with the carina is still present (Melo, unpublished data).

Austroplebeia, a genus proposed by Moure (1961) for a group of species restricted to Australia and New Guinea, came out in the parsimony analysis as the second most basal branch within the Meliponini (branch B, Fig. 1). Its isolated geographic distribution lends support to this placement. This result disagrees with Moure's original suggestion of a close relationship between *Austroplebeia* and the Neotropical *Plebeia*. His view,

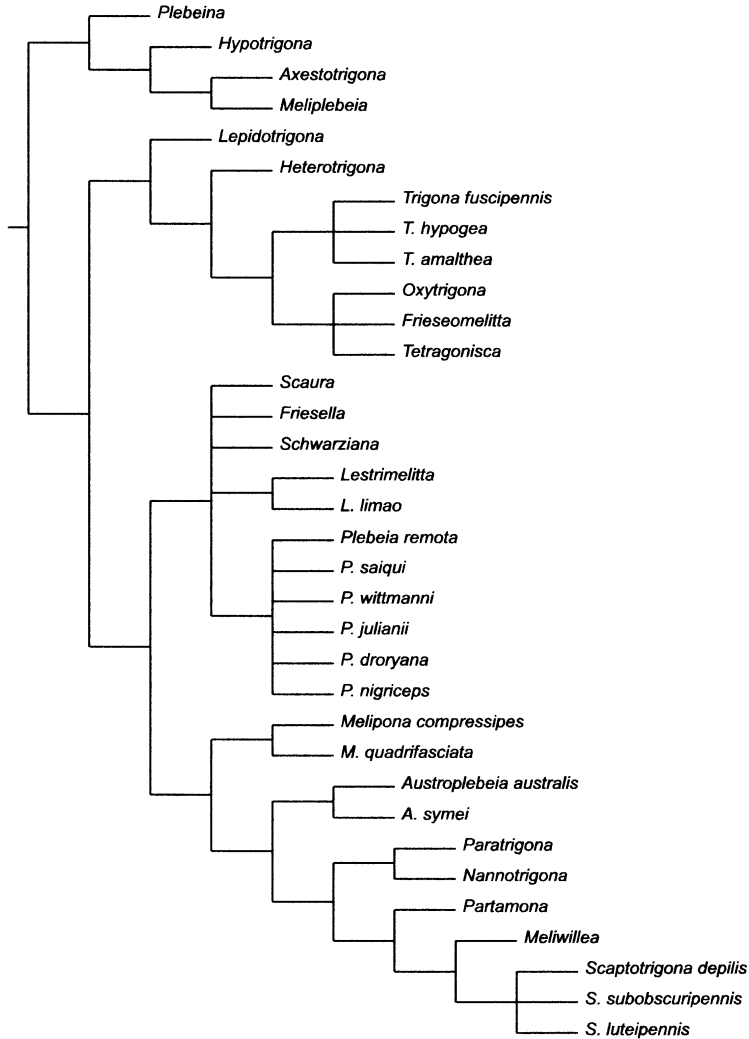


Figure 4. Tree showing relationships among stingless bee taxa used in the present study arranged accordingly to the hypothesis presented by Camargo and Pedro (1992a).

somewhat supported by Camargo and Pedro (1992a, b), was mainly based on the shape of the keirotrichiate area on the inner surface of the worker hind tibia. Michener (1990), however, suggested a different hypothesis placing *Austroplebeia* as closely related to the African genera based on morphology of the worker gonostyli. Our result, on the other hand, does not support either of the previous hypotheses and suggests that the similarities in the morphology of the keirotrichiate area and gonostylus might represent plesiomorphies.

Heterotrigona and *Lepidotrigona*, the two Oriental representatives of the *Trigona* sensu Michener (1990) in our data set, came out as sister groups in the parsimony analysis. These two taxa were nested within a larger clade containing the African genera *Plebeina*, *Axestotrigona* and *Meliplebeia* (branch C in Fig. 1). Although the whole clade had neither jackknife nor bootstrap support higher than 50%, two of the internal branches did, in particular the branch connecting *Meliplebeia* with *Heterotrigona* + *Lepidotrigona*.

This close association between the Oriental and African taxa might seem unlikely considering the morphological features used by Michener (1990) to support the monophyly of both an African clade and *Trigona sensu lato*. Nevertheless, the morphological evidence should be interpreted with caution. *Dactylurina*, for example, was for a long time considered the only African representative of *Trigona* s.l. because of its elongated body shape, shiny integument, narrow keitrichiate ridge and plumose setae along posterior margin of hind tibia (e.g. Moure, 1961). The morphology of the worker sting rudiments and male genitalia, however, led Michener (1990) to include *Dactylurina* among the other African taxa as the sister group of *Plebeina*.

Additional support for a closer relationship between African and Oriental taxa also comes from the relatively high number of paleotropical groups of bees with no representatives in the Neotropical region, e.g. the tribes Allopini and Ctenoplectrini and the genus *Apis*. There is no known case of a bee group occurring only in the Oriental and Neotropical regions and lacking from Africa. In this regard, *Trigona sensu Michener* would be an exception. Therefore, recovering a monophyletic Neotropical clade (branch D, Fig. 1) was not surprising. This does not mean, however, that the whole Neotropical meliponine fauna would constitute a monophyletic group, since *Trigonisca* and related forms were not included in the analysis. There is morphological evidence that this Neotropical group might represent one of the basal branches in Meliponini (see above).

Regarding the arrangements among the 15 Neotropical genera, our results do not support the main groups traditionally recognized within the Neotropical fauna (see Figs. 3 and 4), although some smaller groups with strong morphological support were recovered in the present analysis, e.g. the clade formed by the genera *Meliwillea* and *Scaptotrigona*. This relationship had a high jackknife and bootstrap support corroborating the hypothesis proposed by Roubik et al. (1997). In addition, alternative phylogenetic relationships were suggested for some Neotropical taxa as shown in Figure 1: (1) *Lestrimelitta*, a genus of obligatory robber bees, was placed as closely related to the genus

Plebeia s. str.; (2) the *Plebeia*-like genus *Schwarziana* was placed close to the genus *Trigona* s. str.; (3) *Friesella*, another *Plebeia*-like genus, was placed distantly from *Plebeia*; (4) *Oxytrigona*, a group traditionally included among the *Trigona* s.l. (e.g. Moure, 1951; Michener, 1990, 2000; Camargo, 1996) came out together with *Partamona*; and (5) *Melipona*, a morphologically and behaviorally isolated group within Meliponini, was deeply nested within the Neotropical clade.

The placement of *Lestrimelitta* as sister group of *Plebeia* s. str. in our cladograms also has some support from morphological data. The inner surface of the worker hind tibia of *Lestrimelitta* possesses a narrow, slightly depressed posterior rim, resembling that of *Plebeia*, a condition that, according to Michener (1990), may be indicative of derivation from a common ancestor.

Plebeia sensu Michener includes several taxa recognized as distinct genera by other authors (e.g., Moure, 1951; Camargo and Pedro, 1992a, b). Michener (1990, 2000) emphasized the morphological similarities of these taxa and downweighted the small number of characters used by Moure (1951) to separate the groups. Four of these genera were included in our analysis, *Plebeia*, *Schwarziana*, *Friesella*, and *Scaura*. The results obtained here favor recognition of generic status for these groups since they did not form a monophyletic clade as one would expect if Michener's character weighting was correct.

The position of *Melipona* in our phylogenetic analysis, as one of the most apical branches, is congruent with Camargo and Pedro's (1992a, b) hypothesis (see Fig. 4). This is one of the most specialized groups within Meliponini, exhibiting several autapomorphic morphological and behavioral features. *Melipona* has been traditionally placed as an isolated group within the tribe because of its unique features (e.g. Moure, 1951; Michener, 1990, 2000). Our results, therefore, do not support the somewhat aberrant position for *Melipona* as the most basal branch of the Meliponini in Michener's phylogeny.

This first investigation into the molecular phylogeny of the stingless bees provided only limited support to the previous morphology-based hypotheses. More importantly, it

suggested new alternative relationships among the included taxa. We are aware that some of these relationships are poorly supported and should be carefully reevaluated in future studies. An improved resolution of the present phylogeny is likely to be obtained by analyzing a larger number of meliponine taxa, larger fragments, and by obtaining sequence information from other regions of the genome, in particular nuclear genes. In addition, molecular and morphological data should be jointly analyzed in future investigations. Unfortunately, the currently available information on morphological characters for Meliponini are not detailed enough to allow a robust combined analysis and gathering the necessary morphological data was beyond the scope of the present study.

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Résumé – Phylogénèse moléculaire des abeilles sans aiguillon (Apidae, Apinae, Meliponini) déduite des séquences d'ADN mitochondrial codant pour l'ARNr 16S. Les données des séquences de l'ADN mitochondrial codant pour l'ARN ribosomal 16S de 34 espèces recouvrant 22 genres ont été utilisées pour étudier les relations phylogénétiques parmi les abeilles sans aiguillon (tribu des Meliponini). On a inclus comme groupes extérieurs les séquences disponibles de quatre espèces d'*Apis*, quatre espèces de *Bombus* et trois espèces d'Euglossini (Tab. I). Des analyses de parcimonie sans pondération et du maximum de vraisemblance ont été faites sur une série de données constituées de 462 positions de bases alignées. L'analyse de parcimonie a fourni six arbres également parcimonieux (901 pas, CI = 0,33 et RI = 0,55). L'analyse du maximum de vraisemblance a fourni un arbre dans lequel les relations entre Meliponini différaient sous plusieurs aspects de celles reconstruites par la parcimonie, principalement dans l'organisation à la base de l'arbre. En dépit de ces incongruences, le grand rameau néotropical et plusieurs autres groupes plus petits se ret-

rouvaient dans les deux analyses (Figs. 1 et 2). Dans l'arbre consensus on peut reconnaître quatre rameaux principaux (Fig. 1) : (A) *Hypotrigona*, (B) *Austroplebeia*, (C) les autres genres africains (*Plebeina*, *Meliplebeia* et *Axestotrigona*) plus les deux genres orientaux (*Lepidotrigona* et *Heterotrigona*) et (D) les genres néotropicaux. Le genre africain *Hypotrigona* a été placée sur la branche la plus basale dans la tribu, suivie d'*Austroplebeia* en tant que groupe sœur des deux principaux rameaux (C et D). Ces résultats ne concordent pas avec les groupes traditionnels à la composition intercontinentale, par exemple *Trigona* sensu lato ou *Plebeia* sensu lato. Si l'on considère les groupements parmi les 15 genres néotropicaux, nos résultats ne concordent pas avec les principaux groupes traditionnellement reconnus au sein de cette faune, bien qu'on ait retrouvé certains petits groupes aux caractéristiques morphologiques fortes, par exemple le rameau formé par le genre *Meliwillea* et *Scaptotrigona*. Les relations phylogénétiques alternatives suivantes ont été par ailleurs suggérées : (1) *Lestrimelitta*, genre d'abeilles voleuses obligatoires, a été placée comme étroitement apparentée à *Plebeia* s. str., (2) le genre semblable à *Plebeia*, *Schwarziana*, a été placé près du genre *Trigona* s. str., (3) *Friesella*, autre genre semblable à *Plebeia*, a été placé loin de *Plebeia*, (4) *Oxytrigona*, groupe traditionnellement inclus dans *Trigona* s.l. (Moure, 1951 ; Michener, 1990, 2000 ; Camargo, 1996) formait un groupe avec *Partamona* et (5) *Melipona*, groupe isolé du point de vue morphologique et comportemental au sein des Meliponini, était situé au milieu du rameau néotropical.

abeilles sans aiguillon / Meliponini / phylogénie / ADNr 16S

Zusammenfassung – Ableitung der molekularen Phylogenie der Stachellosen Bienen (Apidae, Apinae, Meliponini) auf Grund von mitochondrialen 16S rDNA Sequenzen. Mit Sequenzdaten mitochondrialer 16S rDNA von 34 Arten (22 Gattungen) wurde die phylogenetische Verwandtschaft bei Stachellosen Bienen (Stamm Tribus Meliponini) untersucht. Die Sequenzen von 4 Arten von *Apis*, 4 Arten von *Bombus* und 3 Arten der Euglossini wurden als "outgroups" zur Untersuchung herangezogen (Tab. I). Gleich gewichtete (equally weighted) Parsimony und Maximum-Likelihood-Analysen wurde bei einem Datensatz von 462 aligned Basenpositionen angewendet. Die Parsimony Analyse ergab 6 Bäume höchster Parsimony (901 Schritte; CI = 0.33 und RI = 0.55). Die Maximum-Likelihood-Analyse ergab einen Baum, in dem sich die Verwandtschaft innerhalb der Meliponini in mehreren Aspekten von der durch Parsimony erhaltenen Verwandtschaft unterschied, vor allem

in der Anordnung in den Basisgruppen. Trotz dieser Inkongruenz wurde der große neotropische Zweig und verschiedene andere kleinere Gruppen in beiden Analysen aufgetrennt (Abb. 1 und 2). Im Parsimony Consensus Baum können vier Hauptzweige erkannt werden (Abb. 1): (A) *Hypotrigona*, (B) *Austroplebeia*, (C) noch vorhandene afrikanische Gattungen (*Plebeina*, *Meliplebeia*, und *Axestotrigona*) zusammen mit den beiden orientalischen Gattungen (*Lepidotrigona* and *Heterotrigona*), und (D) neotropischen Gattungen. Die afrikanische Gattung *Hypotrigona* befand sich am untersten Ast innerhalb des Tribus, gefolgt von *Austroplebeia* als Schwestergruppe der beiden Hauptäste (C und D). Diese Ergebnisse unterstützen die traditionelle Gruppierung nach Kontinenten nicht, e.g. *Trigona sensu lato* oder *Plebeia sensu lato*. Wurde die Gruppierung bei den 15 neotropischen Gattungen betrachtet, unterstützten unsere Ergebnisse auch nicht die Hauptgruppen, die allgemein innerhalb dieser Fauna anerkannt sind, auch wenn einige kleinere Gruppen mit starken morphologisch bedeutenden Daten erkannt wurden, e.g. der Ast, der aus den Gattungen *Meliwillea* und *Scaptotrigona* besteht. Zusätzlich wurden folgende alternative phylogenetische Verwandtschaften vorgeschlagen: (1) *Lestrimelitta*, eine Gattung von obligatorischen Räuberbienen, lag als nahe verwandt bei der Gattung *Plebeia s. str.*; (2) die *Plebeia*-ähnliche Gattung *Schwarziana* wurde nahe an der Gattung *Trigona s. str.* plaziert; (3) *Friesella*, eine andere *Plebeia*-ähnliche Gattung lag entfernt von *Plebeia*; (4) *Oxytrigona*, eine traditionell innerhalb der *Trigona s.l.* liegende Gruppe (e.g. Moure, 1951; Michener, 1990, 2000; Camargo, 1996) bildete einen Cluster mit *Partamona*; und (5) *Melipona*, eine innerhalb der Meliponini morphologisch und vom Verhalten isolierte Gruppe lag mitten in dem neotropischen Ast.

Stachellose Bienen / 16S rDNA / Phylogenie / Meliponini

REFERENCES

- Ascher J.S., Danforth B.N., Ji S. (2001) Phylogenetic utility of the major opsin in bees (Hymenoptera: Apoidea): a reassessment, *Mol. Phylogenet. Evol.* 19, 76–93.
- Camargo J.M.F. (1996) Meliponini Neotropicais: o gênero *Camargoia* Moure, 1989 (Apinae, Apidae, Hymenoptera), *Arq. Zool.* 33, 71–92.
- Camargo J.M.F., Pedro S.R.M. (1992a) Sistemática de Meliponinae (Hymenoptera, Apidae): sobre a polaridade e significado de alguns caracteres morfológicos, in: Cruz-Landim C., Chaud-Netto J. (Eds.), *Anais do Encontro Brasileiro de Biologia de Abelhas e outros Insetos Sociais*, Ed. UNESP, São Paulo, pp. 45–49.
- Camargo J.M.F., Pedro S.R.M. (1992b) Systematics, phylogeny and biogeography of the Meliponinae (Hymenoptera, Apidae): a mini review, *Apidologie* 23, 293–314.
- Cameron S.A. (1991) A new tribal phylogeny of the Apidae inferred from mitochondrial sequences, in: Smith D.R. (Ed.), *Diversity in the Genus Apis*, Westview Press, Boulder, pp. 71–78.
- Cameron S.A. (1993) Multiple origins of advanced eusociality in bees inferred from mitochondrial DNA sequences, *Proc. Natl. Acad. Sci. USA* 90, 8687–8691.
- Cameron S.A., Mardulyn P. (2001) Multiple molecular data sets suggest independent origins of highly eusocial behavior in bees (Hymenoptera: Apinae), *Syst. Biol.* 50, 194–214.
- Crozier R.H., Crozier Y.C. (1993) The mitochondrial genome of the honeybee *Apis mellifera*: complete sequence and genome organization, *Genetics* 133, 97–117.
- Derr J.N., Davis S.K., Wooley J.B., Wharton R.A. (1992a) Variation and the phylogenetic utility of the large ribosomal subunit of mitochondrial DNA from the insect order Hymenoptera, *Mol. Phylogenet. Evol.* 1, 136–147.
- Derr J.N., Davis S.K., Wooley J.B., Wharton R.A. (1992b) Reassessment of the 16S rRNA nucleotide sequence from members of the parasitic Hymenoptera, *Mol. Phylogenet. Evol.* 1, 338–341.
- Dowton M., Austin A.D. (1994) Molecular phylogeny of the insect order Hymenoptera: Apocritan relationships, *Proc. Natl. Acad. Sci. USA* 91, 9911–9915.
- Dowton M., Austin A.D. (2001) Simultaneous analysis of 16S, 28S, COI and morphology in the Hymenoptera: Apocrita: Evolutionary transitions among parasitic wasps, *Biol. J. Linn. Soc.* 74, 87–111.
- Dowton M., Austin A.D., Dillon N., Bartowsky E. (1997) Molecular phylogeny of the apocritan wasps: the Proctotrupomorpha and Evaniomorpha, *Syst. Entomol.* 22, 245–255.
- Engel M.S. (2001a) A monograph of the Baltic amber bees and evolution of the Apoidea (Hymenoptera), *Bull. Am. Mus. Nat. Hist.* 259, 1–192.
- Engel M.S. (2001b) Monophyly and extensive extinction of advanced eusocial bees: Insights from an unexpected Eocene diversity, *Proc. Natl. Acad. Sci. USA* 98, 1661–1664.
- Farris J.S., Albert V.A., Källersjö M., Lipscomb D., Kluge A.G. (1996) Parsimony jackknifing outperforms neighbor-joining, *Cladistics* 12, 99–124.
- Koulianos S., Schmid-Hempel R., Roubik D.W., Schmid-Hempel P. (1999) Phylogenetic relationships within the corbiculate Apinae (Hymenoptera) and the evolution of eusociality, *J. Evol. Biol.* 2, 380–384.

- Michener C.D. (1944) Comparative external morphology, phylogeny and classification of the bees, *Bull Am. Mus. Nat. Hist.* 82, 151–326.
- Michener C.D. (1974) *The Social Behavior of the Bees*, Harvard Univ. Press, Cambridge.
- Michener C.D. (1990) Classification of the Apidae (Hymenoptera), *Univ. Kansas Sci. Bull.* 54, 75–164.
- Michener C.D. (2000) *The Bees of the World*, Johns Hopkins Univ. Press, Baltimore.
- Moure J.S. (1951) Notas sobre Meliponinae (Hymenoptera, Apoidea), *Dusenía* 2, 25–50.
- Moure J.S. (1961) A preliminary supra-specific classification of the old world Meliponinae bees (Hymenoptera, Apidae), *Studia Entomol.* 4, 181–242.
- Noll F.B. (2002) Behavioral phylogeny of corbiculate Apidae (Hymenoptera; Apinae), with special reference to social behavior, *Cladistics* 18, 137–153.
- Posada D., Crandall K.A. (1998) MODELTEST: testing the model of DNA substitution, *Bioinformatics* 14, 817–818.
- Posada D., Crandall K.A. (2001) Selecting the best-fit model of nucleotide substitution, *Syst. Biol.* 50, 580–601.
- Roubik D.W., Segura J.A.L., Camargo J.M.F. (1997) New stingless genus endemic to Central American cloudforests: phylogenetic and biogeographic implications (Hymenoptera: Apidae: Meliponini), *Syst. Entomol.* 22, 67–80.
- Sakagami S.F. (1982) Stingless bees, in: Hermann H.R. (Ed.), *Social Insects*, Vol. III, Academic Press, New York, pp. 361–423.
- Sanger F.S., Nicklen S., Coulson A.R. (1977) DNA sequencing with chain terminator inhibitors, *Proc. Natl. Acad. Sci. USA* 74, 5463–5467.
- Schultz T.R., Engel M.S., Ascher J.S. (2001) Evidence for the origin of eusociality in the corbiculate bees (Hymenoptera: Apidae), *J. Kans. Entomol. Soc.* 74, 10–16.
- Schwarz H.F. (1948) Stingless bees (Meliponidae) of the Western Hemisphere, *Bull. Am. Mus. Nat. Hist.* 90, 1–546.
- Sheppard W.S., McPherson B.A. (1991) Ribosomal DNA diversity in *Apis*, in: Smith D.R. (Ed.), *Diversity in the Genus Apis*, Oxford, Westview Press, Boulder, pp. 89–107.
- Swofford D.L. (2001) PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods), Version 4, Sinauer Associates, Sunderland, Massachusetts.
- Tanaka H., Roubik D.W., Kato M., Liew F., Gunsalam G. (2001) Phylogenetic position of *Apis nuluensis* of northern Borneo and phylogeography of *A. cerana* as inferred from mitochondrial DNA sequences, *Insectes Soc.* 48, 44–51.
- Thompson J.D., Higgins D.G., Gibson T.J. (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, positions-specific gap penalties and weight matrix choice, *Nucl. Acids Res.* 22, 4673–4680.
- Wille A. (1979) Phylogeny and relationships among the genera and subgenera of stingless bees (Meliponinae) of the world, *Rev. Biol. Trop.* 27, 241–277.
- Wille A. (1983) Biology of the stingless bees, *Annu. Rev. Entomol.* 28, 41–64.