



## Molecular Similarity of MDR Inhibitors

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**Abstract:** The molecular similarity of multidrug resistance (MDR) inhibitors was evaluated using the point centred atom charge approach in an attempt to find some common features of structurally unrelated inhibitors. A series of inhibitors of bacterial MDR were studied and there is a high similarity between these in terms of their shape, presence and orientation of aromatic ring moieties. A comparison of the lipophilic properties of these molecules has also been conducted suggesting that this factor is important in MDR inhibition.

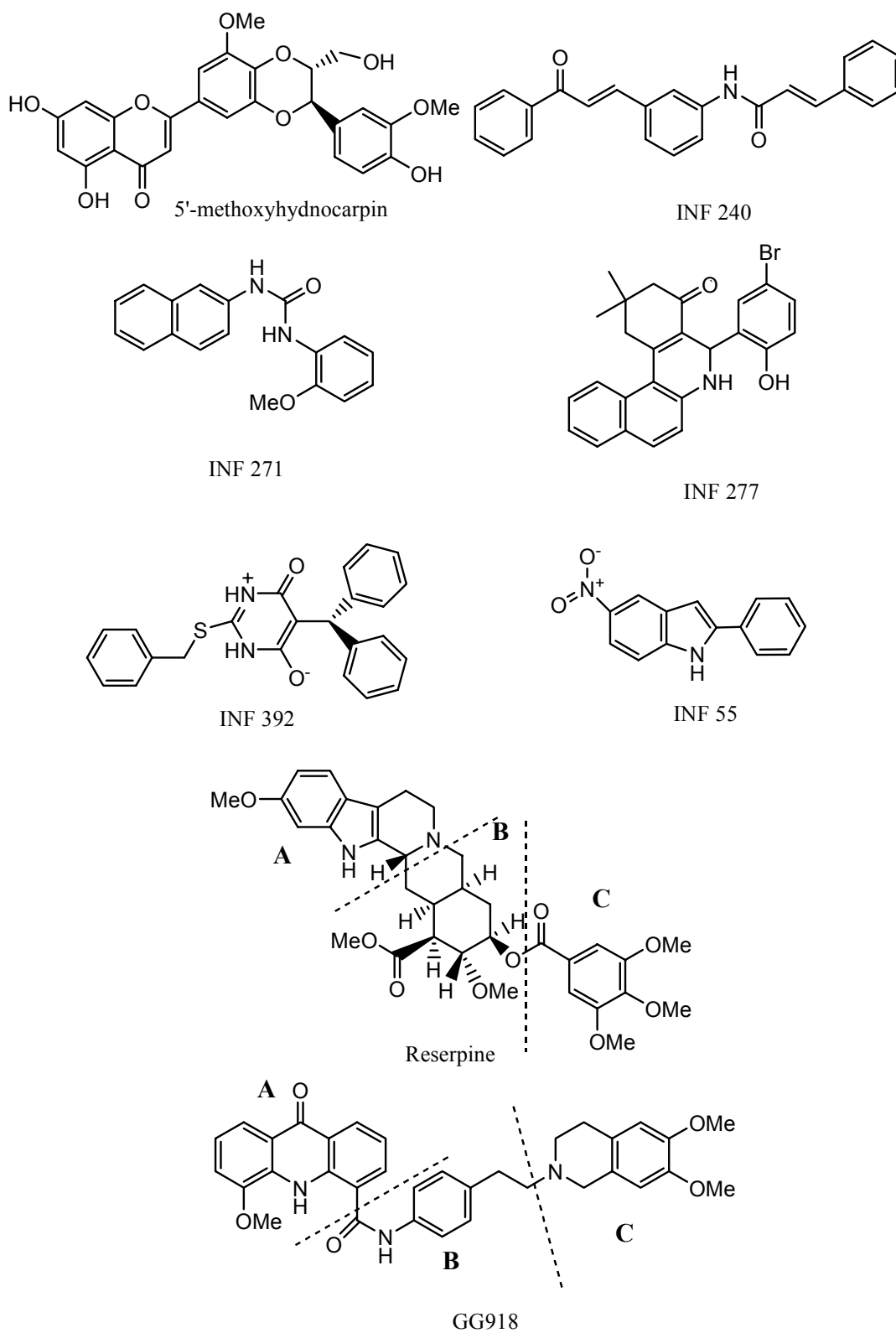
**Keywords:** MDR, multidrug resistance, inhibitor, molecular similarity, SAR, *ab initio*.

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### Introduction

MDR or multidrug resistance is responsible for many forms of resistance in bacteria, fungi and human tumours [1]. This resistance functions by the presence of membrane bound efflux pumps, which actively export therapeutics from the cell resulting in a low intracellular ineffective concentration of the drug [2]. These pumps recognize a wide variety of structurally unrelated compounds [3] and it is believed that MDR inhibitors bind directly to the hydrophobic region of the efflux pump thus preventing the drug transport [4].

There has been much research conducted to find inhibitors of these proteins, particular in human tumour resistance as reviewed by Stouch and Gudmundsson [5]. Some progress has been reported on



the structure-activity relationships (SARs) for inhibitors of bacterial efflux pumps [6], but further work is necessary to fully explain the mechanism of MDR efflux inhibition, since most potent inhibitors of the NorA MDR pump of *Staphylococcus aureus* come from totally different chemical classes. This is unusual since it seems that there is no common pharmacophore that causes inhibition of MDR.

We have modelled and explored biomolecular similarities of a series of representative MDR inhibitors of the NorA pump from different classes (Scheme 1) on the basis of molecular interaction potentials and report here some structural features required for MDR inhibition.

## Computational Methods

The inhibitors studied were optimized by Gamess-US *ab initio* software package [7] and HF/6-311G(\*) basis set (except for the INF 277 where we have used HF/6-31G(\*) basis set). The molecular similarity was evaluated by MIPSIM software [8] using COMP module and a classical atom-centred point-charge distribution (PTC\_MEP) approach. The reserpine and GG918 molecules were too big for the MIPSIM calculations, and were split into 3 units for comparison with other inhibitors (denoted as A, B and C in Scheme 1). The theoretical values of logP, surface area and volume were calculated by SciLogP 3.0 [9], Vega [10] and Chem3D [11] software packages. Visualization of the results was achieved by ViewerLite [12] and ICM Lite [13] software packages.

## Results and Discussion

The efflux pump NorA plays an important role in resistance to fluoroquinolone antibiotics of the major human pathogen *Staphylococcus aureus*, which is highly problematic in the clinical environment [14]. The restoration of antibiotic efficacy could be achieved by using inhibitors, molecules that potentiate the activity of standard antibiotics against MDR cells. Efflux inhibitors are from a wide range of structural classes and representative molecules for different classes were studied here. The experimental results of MDR modulation for chosen inhibitors are shown in Table 1. These results are taken from the literature [15] or from one of the authors [16, 17]. Compounds with no potential for MDR inhibition are shown in Scheme 2. It is believed that inhibitors of these transport processes act by directly binding to hydrophobic regions of MDR proteins causing inhibition of antibiotic removal [4]. Since a wide range of MDR inhibitors have been discovered and with no apparent pharmacophore detected, we have assumed that interactions important for MDR inhibition must be non-specific. The molecular electrostatic potential could be very important for the formation of a potential hydrogen bond network and other interactions between MDR inhibitor and efflux pump, and it could also play a significant role in molecular recognition.

In the absence of an explanation for the MDR inhibition mechanism, we have decided to evaluate the importance of molecular electrostatic potential in these processes. The *ab initio* optimised geometries of selected inhibitors of the NorA efflux pump were compared using atom-centred point-

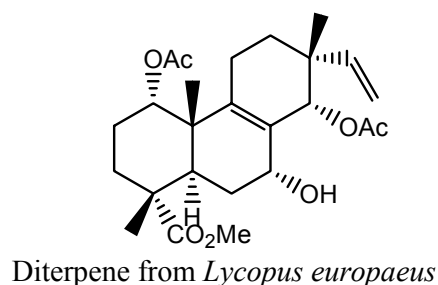
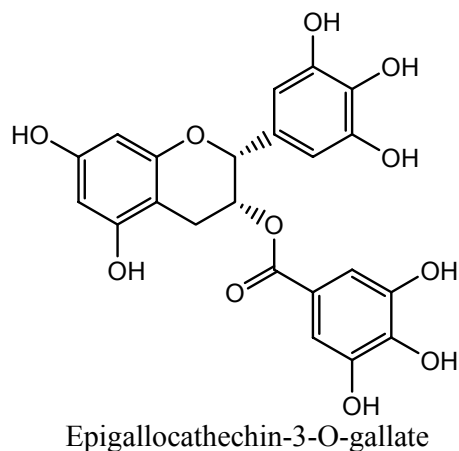
**Table 1.** MDR modulation results for NorA inhibitors (5'-MHC depicted 5'-methoxyhydnocarpin).

Inhibitor	Drug	Inhibition	Reference
INF 240	Ciprofloxacin	0.12*	Markham et al., 1999. [15]
INF 271	Ciprofloxacin	0.12*	Markham et al., 1999. [15]
INF 277	Ciprofloxacin	0.15*	Markham et al., 1999. [15]
INF 392	Ciprofloxacin	0.28*	Markham et al., 1999. [15]
INF 55	Ciprofloxacin	0.25*	Markham et al., 1999. [15]
5'-MHC	Norfloxacin	4**	Stermitz et al., 2000. [18]
Reserpine	Norfloxacin	4**	Gibbons et al., 2003. [16]
GG918	Norfloxacin	8**	Gibbons et al., 2003a. [16]
Diterpene from <i>Lycopus europaeus</i>	Norfloxacin	ND	Gibbons et al., 2003b. [16]
Epigallocatechin- 3-O-gallate	Norfloxacin	ND	Gibbons, unpublished data [2003]

\*FIC index - <0.5 is considered to be indicator of synergistic activity.

\*\*Fold reduction in minimum inhibitory concentration (MIC) of antibiotic in the presence of inhibitor.

ND - no drug potentiation.

**Scheme 2**

charge distribution and results are presented in Table 2 in the form of a similarity matrix. Reserpine and GG918 had to be split into three portions, and those are defined in the Scheme 1.

The similarity index for all pairs of inhibitors was between 0.644 and 0.932, depending on the size of the compared molecules. The results for the parts of reserpine and GG918 are to be taken with caution due to the relatively small sizes of examined moieties. Direct correlation between similarity and fold of modulation cannot be fully examined due to differences in the representation of experimental data, however, the following could be emphasised:

- a) INF 271 and INF 277 have a similar potency of MDR inhibition and similarity index is also high – 0.817;
- b) 5'-methoxyhydnocarpin (5'-MHC) and INF 240 are potent MDR inhibitors and have a high similarity index.

Due to the difference in the size of molecules, a more detailed analysis was carried out by visually comparing inhibitors.

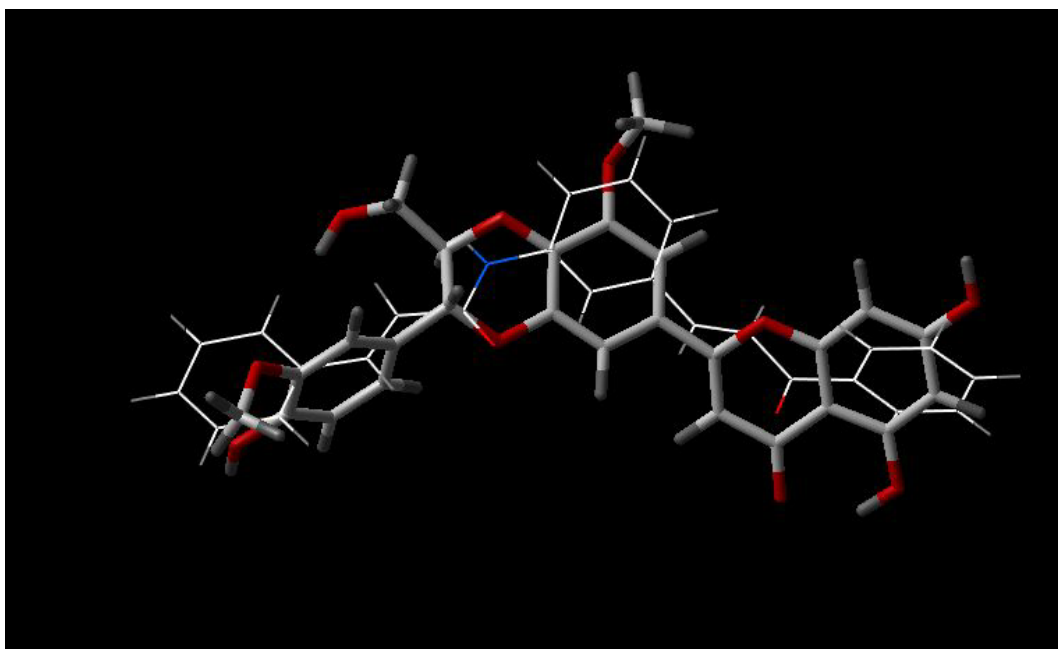
**Table 2.** Final similarity matrix for MDR inhibitors of the NorA pump calculated by MIPSIM using point charges. Three parts of reserpine were denoted as Reserpine A, Reserpine B and Reserpine C, and three parts of GG918 were denoted as GG918 A, GG918 B, and GG918 C. Similarity between parts that belong to the same molecule were not considered.

	5'-MHC	INF 240	INF 271	INF 277	INF 392	INF 55
5'-MHC	1.000					
INF 240	0.840	1.000				
INF 271	0.735	0.733	1.000			
INF 277	0.704	0.673	0.817	1.000		
INF 392	0.740	0.751	0.781	0.722	1.000	
INF 55	0.729	0.707	0.866	0.803	0.725	1.000
Reserpine A	0.690	0.735	0.870	0.754	0.719	0.932
Reserpine B	0.691	0.616	0.817	0.813	0.739	0.808
Reserpine C	0.699	0.649	0.717	0.695	0.677	0.737
GG918 A	0.669	0.713	0.818	0.755	0.707	0.915
GG918 B	0.644	0.703	0.759	0.699	0.670	0.839
GG918 C	0.716	0.692	0.795	0.722	0.646	0.824
Diterpene from <i>Lycopus europaeus</i>	0.681	0.566	0.763	0.807	0.722	0.720
Epigallocatechin-3-O-gallate	0.682	0.670	0.825	0.868	0.733	0.750

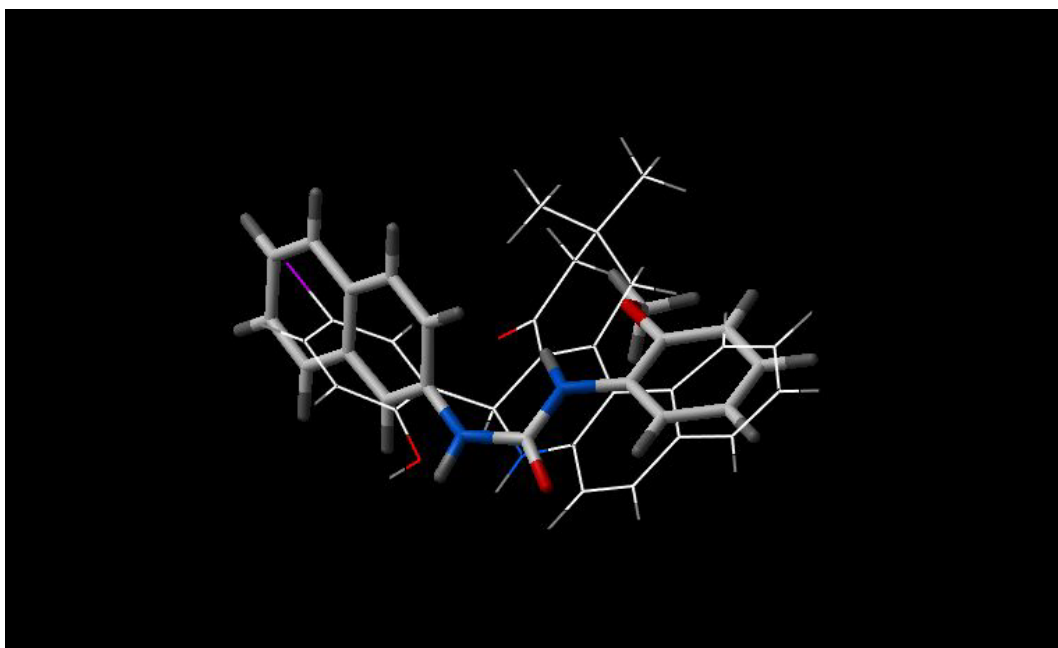
From the figures of the best fit between pairs of inhibitors, some observations are apparent:

- 5'-methoxyhydnocarpin and INF 240 have the same shape and some polar groups in a similar position (Figure 1). Note the absence of nitrogen atom in 5'-MHC;
- INF 271 and INF 277 have a similar shape and both have a nitrogen atom in the middle of the molecule (Figure 2);
- INF 55 is planar and different in shape compared to INF 271, however both molecules have a nitrogen atom in the middle of molecule and aromatic rings from both molecules are almost parallel to each other (Figure 3);
- there is high similarity between INF 271 and parts of Reserpine and GG918 (Figure 4 and Figure 5, respectively), again with a nitrogen atom in the middle of the molecule;
- all potent MDR inhibitors have aromatic rings in the areas that contain high similarity.

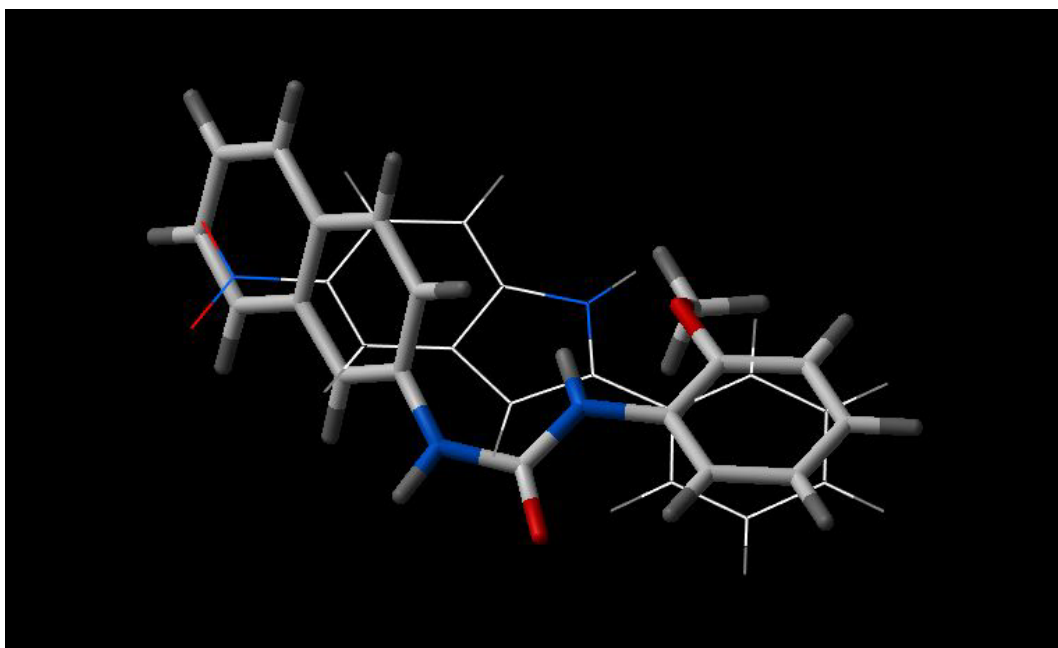
In Figures 6 and 7, similarities between epigallocatechin-3-O-gallate (EGG) and two Influx compounds are depicted. However, EGG is not a potentiator of drugs in bacterial MDR processes, and possibly is therefore not an inhibitor of bacterial MDR efflux. This can possibly be explained by visually examining the best fit between EGG and the two Influx compounds (INF271 and INF277). The similarity index is very high for both of these combinations but it can be observed that there is a poor fit between EGG and the aromatic moieties of both of these compounds. Looking at the fit between INF277 and the diterpene from *Lycopus europaeus*, there is a high calculated similarity but again this diterpene is not a potentiator of MDR drugs and this is probably due to the poor fit that this has with aromatic moieties of other MDR inhibitors e.g. INF277 (Figure 8).



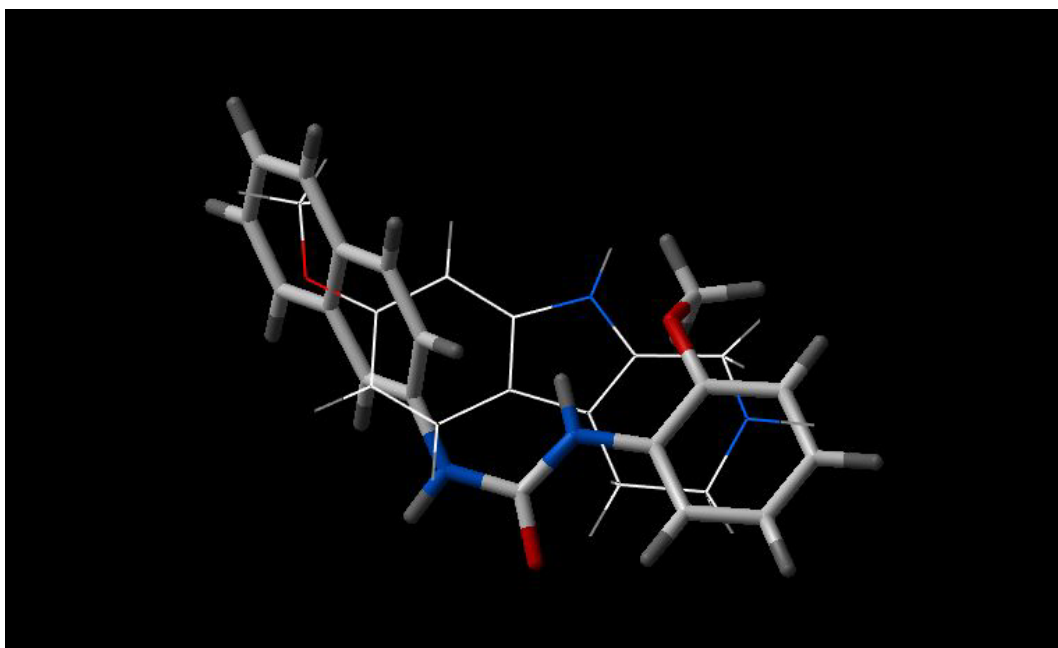
**Figure 1.** Best fit of optimized 5'-methoxyhydnocarpin (sticks) and INF 240 (wireframe) structures.



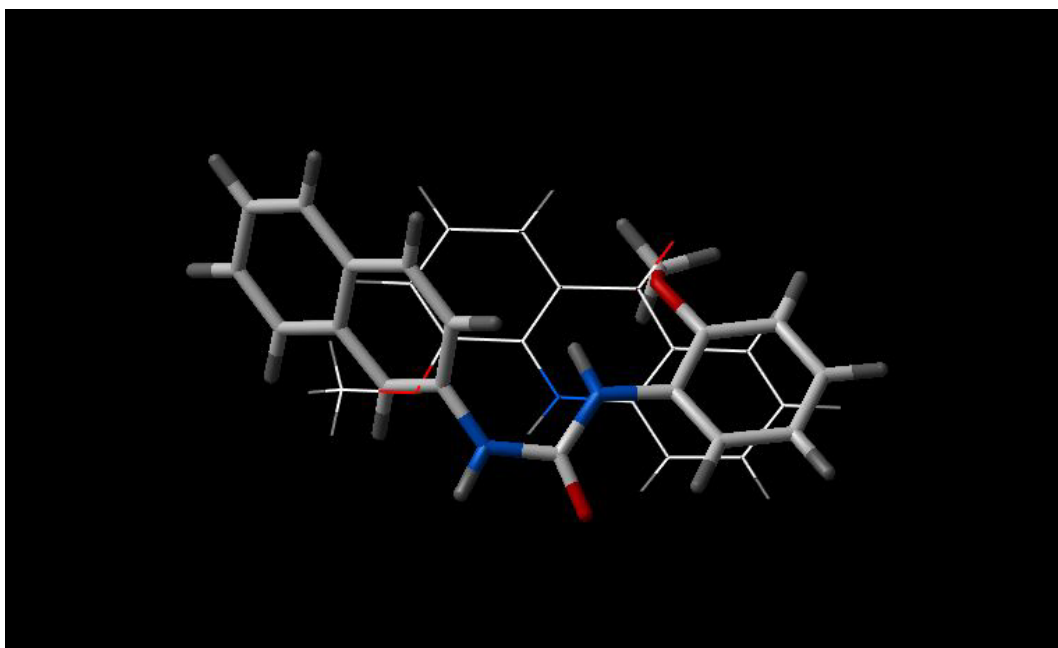
**Figure 2.** Best fit of optimised INF 271 (sticks) and INF 277 (wireframe) structures.



**Figure 3.** Best fit of optimised INF 271 (sticks) and INF 55 (wireframe) structures.

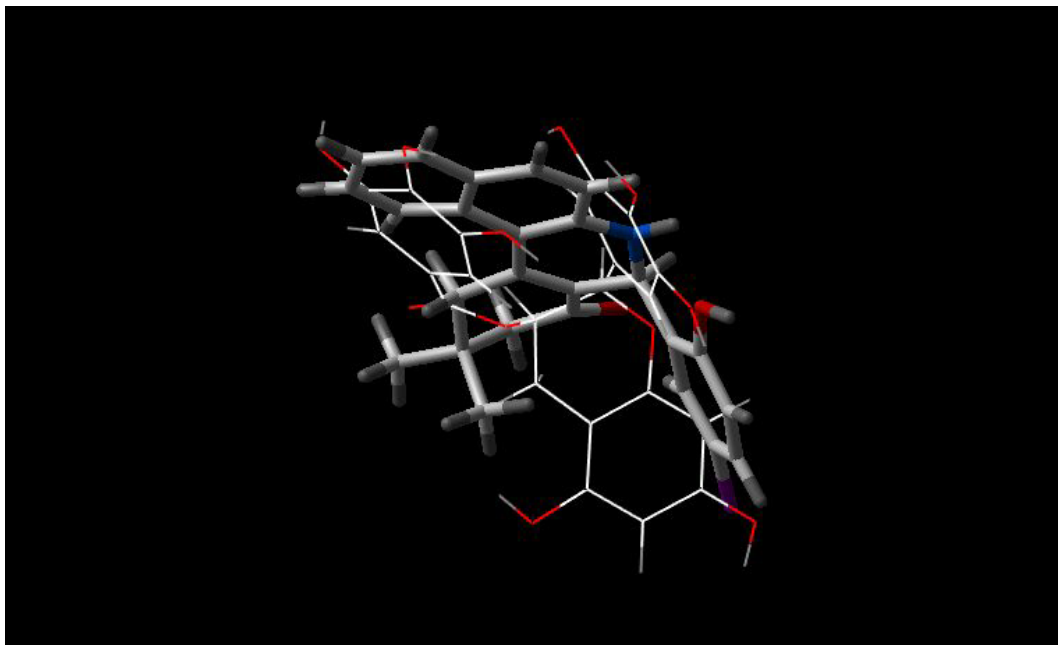


**Figure 4.** Best fit of optimised INF 271 (sticks) and Reserpine A (wireframe) structures.

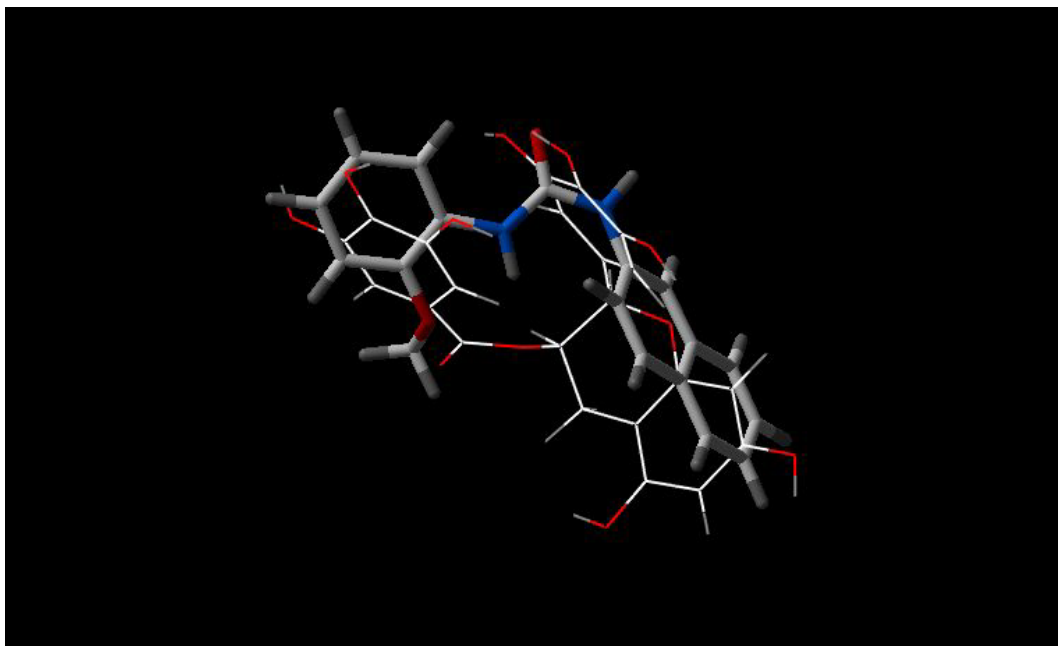


**Figure 5.** Best fit of optimised INF 271 (sticks) and GG918 A (wireframe) structures.

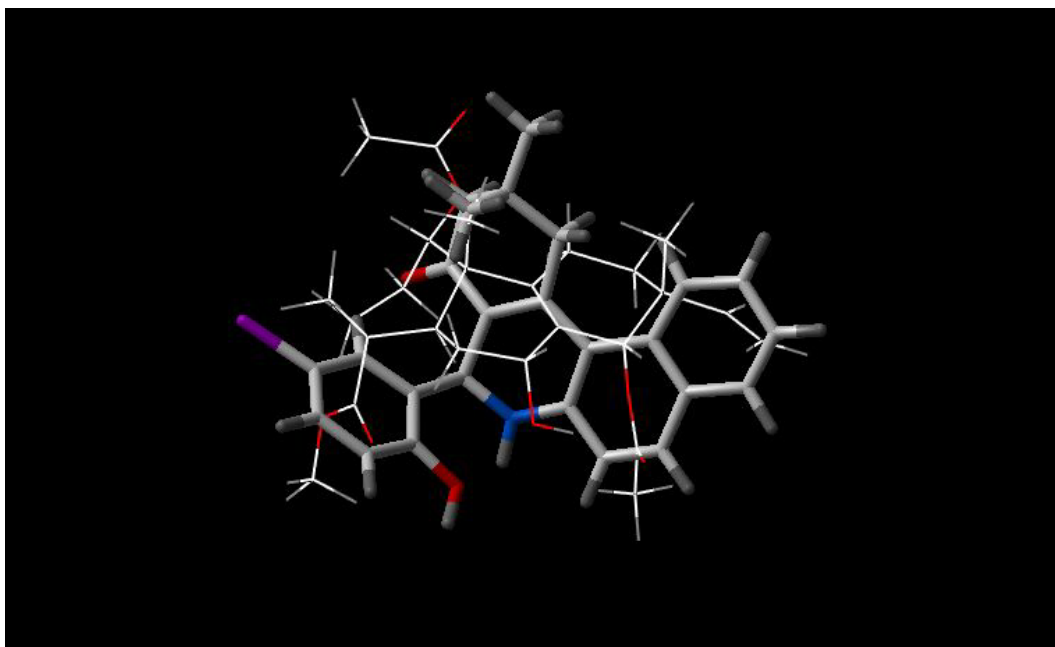




**Figure 6.** Best fit of optimised INF277 (sticks) and Epigallocatechin-3-O-gallate (wireframe) structures.



**Figure 7.** Best fit of optimised INF271 (sticks) and Epigallocatechin-3-O-gallate (wireframe) structures.



**Figure 8.** Best fit of optimised INF277 (sticks) and diterpene from *Lycopodium europaeus* (wireframe) structures.

We have also studied some theoretical parameters i.e. clogP and logP, (Table 3), however there is no obvious correlation with MDR modulation results. However clogP is generally lower for non-MDR inhibitors, the exception to this is INF55, which is a small molecule and one of the poorest MDR inhibitor of the INF series.

**Table 3.** Theoretically calculated values of clogP, logP, surface area and volume of all studied MDR inhibitors and non potentiators by different methods.

Inhibitor	clogP (Chem3D)	logP (Chem3D)	logP (SciLogP)	logP (Vega)	lipole (Vega)	Surface area (Å <sup>2</sup> )	Volume (Å <sup>3</sup> )
5'-MHC	3.47	2.33	6.23	5.43	2.92	482.9	410.5
INF 240	5.14	4.73	6.04	3.03	1.43	396.5	328.5
INF 271	4.28	3.38	5.67	3.76	2.77	316.9	263.9
INF 277	6.02	5.12	6.02	6.73	2.29	408.0	361.2
INF 392	4.25	5.60	6.15	4.48	1.29	419.8	354.0
INF 55	0.93	3.68	5.09	4.92	5.12	248.7	207.2
Reserpine	3.85	2.69	6.08	4.38	1.10	656.0	546.6
GG918	4.21	5.03	6.12	5.60	0.89	629.9	536.0
Diterpene from <i>Lycopodium europaeus</i>	2.45	3.65	6.23	3.34	0.73	497.84	425.59
Epigallocatechin- 3-O-gallate	1.49	2.07	6.21	6.62	2.61	429.3	363.6

## Conclusions

This study has shown that there is a high similarity between inhibitors of the NorA MDR transporter with similarity index higher than 0.6. However, there is no obvious correlation between similarity index and potential as MDR inhibitor, since some non-potentiators have high similarity index with MDR inhibitors. The important feature that differentiates inhibitors and non-inhibitors is the shape of the molecule and relative position of the aromatic moieties present in the molecule.

Although in most inhibitors there is a nitrogen atom in the middle of the molecule, it is not essential, for example 5'-MHC has no such feature. This confirms the assumption that the interactions occurring during MDR inhibition must be non-specific. The shape of the molecule, aromatic rings and presence of some polar atoms will determine the potency of MDR inhibition. This study should be expanded to encompass a further series of inhibitor and non-inhibitor molecules of MDR processes of NorA in order to derive rules for the *in silico* screening for MDR inhibitors.

## References

1. Ling, V. *Cancer Chemotherapy and Pharmacology* 1997, 40, Suppl:S3-8.
2. Bradley, G.; Ling, V. *Cancer Metastasis Rev.* 1994, 13, 223-233.
3. Neyfakh, A.A. *J. Mol. Microbiol. Biotechnol.* 2001, 3, 151-154.
4. Neyfakh, A.A. *Molecular Microbiology* 2002, 44, 1123-1130.
5. Stouch, T.R.; Gudmundsson, O. *Advanced Drug Delivery Reviews* 2002, 54, 315-328.
6. Guz, N. R.; Stermitz, F.R.; Johnson, J.B.; Beeson, T.D.; Willen, S.; Hsiang, J.F.; Lewis, K. *J. Med. Chem.* 2001, 44, 261-268.
7. Schmidh, M.W.; Baldridge, K.K.; Boatz, J.A.; Elbert, S.T.; Gordon, M.S.; Jensen, J.H.; Koseki, S.; Matsunaga, N.; Nguyen, K.A.; Su, S.J.; Windus, T.L.; Dupuis, M.; Montgomery, J.A. *Journal of Computational Chemistry* 1993, 14, 1347-1363.
8. Sanz, F.; Manaut, F.; Rodriguez, J.; Lozoya E.; Lopez-de-Briñas, E. *Journal of Computer-Aided Molecular Design* 1993, 7, 337-347.
9. SciLogP 3.0, SciVision, Inc. 1999.
10. Pedretti, A.; Villa, L.; Vistoli, G. *J. Mol. Graph.*, 2002, 21, 47-49.
11. Chem3D Ultra, CambridgeSoft 2001.
12. ViewerLite 5.0, Accelerlys Inc 2002.
13. ICM Lite, MolSoft, 2002.
14. Kaatz, G.W.; Seo, S.M.; Ruble, C.A. *Antimicrob. Agents. Chemother.* 1993, 37, 1086-1094.
15. Markham, P.N.; Westhaus, E.; Klyachko, K.; Johnson, M.E.; Neyfakh, A.A. *Antimicrobial Agents and Chemotherapy* 1999, 43, 2404-2408.
16. Gibbons, S.; Oluwatuyi, M. Kaatz, G.W. *Journal of Antimicrobial Chemotherapy* 2003, 51, 13-17.
17. Gibbons, S.; Oluwatuyi, M.; Veitch, N.C. Gray, A.I. *Phytochemistry* 2003, 62, 83-87.
18. Stermitz, F.R.; Lorenz, P.; Tawara, J.N.; Zenewicz, L.A. Lewis, K. *PNAS* 2000, 97, 1433-1437.