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Monoamine oxidase B inhibitor, selegiline, reduces ¹⁸F-THK5351 uptake in the human brain

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Abstract

Background: ¹⁸F-THK5351 is a quinoline-derived tau imaging agent with high affinity to paired helical filaments (PHF). However, high levels of ¹⁸F-THK5351 retention in brain regions thought to contain negligible concentrations of PHF raise questions about the interpretation of the positron emission tomography (PET) signals, particularly given previously described interactions between quinolone derivatives and monoamine oxidase B (MAO-B). Here, we tested the effects of MAO-B inhibition on ¹⁸F-THK5351 brain uptake using PET and autoradiography.

Methods: Eight participants (five mild cognitive impairment, two Alzheimer's disease, and one progressive supranuclear palsy) had baseline ¹⁸F-AZD4694 and ¹⁸F-THK5351 scans in order to quantify brain amyloid and PHF load, respectively. A second ¹⁸F-THK5351 scan was conducted 1 week later, 1 h after a 10-mg oral dose of selegiline. Three out of eight patients also had a third ¹⁸F-THK5351 scan 9–28 days after the selegiline administration. The primary outcome measure was standardized uptake value (SUV), calculated using tissue radioactivity concentration from 50 to 70 min after ¹⁸F-THK5351 injection, normalizing for body weight and injected radioactivity. The SUV ratio (SUVR) was determined using the cerebellar cortex as the reference region. ¹⁸F-THK5351 competition autoradiography studies in postmortem tissue were conducted using 150 and 500 nM selegiline.

Results: At baseline, ¹⁸F-THK5351 SUVs were highest in the basal ganglia (0.64 ± 0.11) and thalamus (0.62 ± 0.14). In the post-selegiline scans, the regional SUVs were reduced on average by 36.7% to 51.8%, with the greatest reduction noted in the thalamus (51.8%) and basal ganglia (51.4%). MAO-B inhibition also reduced ¹⁸F-THK5351 SUVs in the cerebellar cortex (41.6%). The SUVs remained reduced in the three patients imaged at 9–28 days. Tissue autoradiography confirmed the effects of MAO-B inhibition on ¹⁸F-THK5351 uptake.

Conclusions: These results indicate that the interpretation of ¹⁸F-THK5351 PET images, with respect to tau, is confounded by the high MAO-B availability across the entire brain. In addition, the heterogeneous MAO-B availability across the cortex may limit the interpretation of ¹⁸F-THK5351 scans using reference region methods.

Keywords: ¹⁸F-THK5351 tau tracer, Monoamine oxidase-B, Selegiline, Alzheimer's disease, Positron emission tomography

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Background

In vivo characterization of tau pathology using positron emission tomography (PET) constitutes a new frontier in Alzheimer's disease (AD) research [1]. It is expected that tau imaging agents will provide the means for staging disease progression as well as selecting patients appropriate for a given therapy, confirming the mechanism of action of pharmacological interventions affecting tau aggregates and monitoring treatment efficacy for AD [2].

¹⁸F-THK5351 is a guinoline-derivative tau imaging tracer with affinity to paired helical filaments (PHF), a typical tau aggregate present in neurofibrillary tangles (NFTs). ¹⁸F-THK5351 retention in the temporal lobes in vivo is able to distinguish AD patients from healthy individuals [3]. However, ¹⁸F-THK5351 is also retained in the basal ganglia and other brain regions known to express negligible amounts of PHF, a phenomenon attributable to off-target binding, which raises concerns about the specificity of ¹⁸F-THK5351 for PHF. Rather than PHF, the basal ganglia typically express high concentrations of aminergic projections, neuroreceptors, and their degrading enzymes such as monoamine oxidase (MAO) [4, 5]. As such, it is plausible to hypothesize that MAO-B (EC 1.4.3.4) constitutes a ¹⁸F-THK5351 off-target binding site, particularly given its previous reported affinity to quinolone derivatives [6]. Furthermore, the presence of high concentrations of MAO-B in the human cortex demands a careful examination of the impact of MAO-B binding on ¹⁸F-THK5351 images.

Selegiline is an irreversible MAO-B inhibitor utilized to enhance dopaminergic neurotransmission in Parkinson's disease patients [7]. Previous PET studies revealed a persistent MAO-B inhibition following a single administration of MAO-B inhibitors [4]. As such, an acute selegiline challenge constitutes an interesting strategy to probe the contribution of brain MAO-B availability to the ¹⁸F-THK5351 uptake observed in patient populations.

Here, in a longitudinal observation of patients with baseline and post-selegiline ¹⁸F-THK5351 PET scans, we tested the hypothesis that a reduction of MAO-B availability also reduces ¹⁸F-THK5351 uptake. We further tested this hypothesis in postmortem human tissue using autoradiography.

Methods

Subjects

Eight individuals (five mild cognitive impairment (MCI), two AD, and one progressive supranuclear palsy (PSP)) were recruited. All study participants were screened to exclude any underlying neuropsychiatric diseases such as depression and concomitant intake of MAO inhibitors. Study participants then underwent a Mini-Mental State Examination (MMSE) [8] and Montreal Cognitive Assessment (MoCA) [9]. The MCI, AD, and PSP diagnoses were made using the Petersen's [10], National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's disease and Related Disorders Association (NINCDS-ADRDA) [11], and the National Institute of Neurological Disorders and Stroke and the Society for PSP (NINDS-SPSP) [12] criteria, respectively.

Scanning protocol

All eight participants had baseline ¹⁸F-AZD4694 and ¹⁸F-THK5351 PET scans to quantify brain amyloid and PHF load, respectively. A second ¹⁸F-THK5351 scan was conducted 1 week later, 1 h after an oral dose of 10 mg selegiline. Participants were also invited to undergo an optional third ¹⁸F-THK5351 PET scan 10 days after the selegiline administration. Each ¹⁸F-THK5351 acquisition consisted of dynamic images $(4 \times 5 \text{ min})$ acquired at 50-70 min after intravenous bolus injection. The mean ± standard deviation injected radioactivity of ¹⁸F-THK5351 was 6.6 ± 0.3 mCi for the baseline scan, 6.7 ± 0.4 mCi for the post-selegiline scan, and 6.9 ± 0.4 mCi for the third scan. ¹⁸F-AZD4694 acquisition consisted of dynamic images $(6 \times 5 \text{ min})$ acquired at 40–70 min after intravenous bolus injection of ¹⁸F-AZD4694. The injected radioactivity of ¹⁸F-AZD4694 was 6.3 ± 0.3 mCi. A 6-min transmission scan was acquired at the end of each PET scan. All PET scans were performed using the Siemens High Resolution Research Tomograph (HRRT). A magnetic resonance imaging (MRI) anatomical scan was also performed for all patients using the 1.5 T Siemens Sonata scanner for co-registration purposes.

¹⁸F-THK5351 and ¹⁸F-AZD4694 synthesis

¹⁸F-THK5351 was synthesized according to a previously published paper [3], with an average specific activity of 24.3 Ci/μmol for the baseline scan, 19.2 Ci/μmol for the post-selegiline scan, and 13.5 Ci/μmol for the third scan. The average injected mass dose was 5.58 ng/kg for the baseline scan, compared with 6.68 ng/kg for the postselegiline scan (P = 0.61, not significant), and 4.21 ng/kg for the third scan (P = 0.39, not significant). ¹⁸F-AZD4694 was synthesized using a modified synthesis procedure previously described [13], with an average specific activity of 6.1 Ci/μmol.

PET and MRI image processing

T1-weighted MRI images were processed using the CIVET image processing pipeline [14, 15] for field distortions correction, and the PET images were processed using an established image processing pipeline [16]. Briefly, using transformations obtained from PET native to MRI native space and the MRI native to the Montreal Neurological Institute (MNI) 152 space, the PET images were linearly registered and subsequently

spatially normalized to the MNI 152 standardized space. The primary outcome measure was the standardized uptake value (SUV). SUVs were calculated using tissue radioactivity concentration data from 50 to 70 min following ¹⁸F-THK5351 injection, and 40 to 70 min following ¹⁸F-AZD4694 injection, normalized by body weight and injected radioactivity. The baseline and postselegiline ¹⁸F-THK5351 SUVs in the cortical gray matter of the frontal, parietal, lateral temporal, and occipital lobes, hippocampus, cingulate gyri, basal ganglia, thalamus, and cerebellar cortex of each individual were measured. We then calculated the average percentage reduction of mean regional SUV in the post-selegiline scans. The ¹⁸F-THK5351 SUV ratio (SUVR) and ¹⁸F-AZD4694 SUVR were obtained using the cerebellar cortex and cerebellum as reference regions, respectively. The ¹⁸F-AZD4694 SUVR images were assessed by two independent experts (JPS and PRN) to visually classify the participants as amyloid-positive or amyloid-negative after full agreement was reached. In a clinical validation of ¹⁸F-AZD4694 using the cerebellum as the reference region, the SUVRs in AD patients varied between 2.10 and 2.88 across regions [13].

Autoradiography

In vitro autoradiography was performed on 14 postmortem brain sections (seven AD patients with a diagnosis confirmed by pathological assessment, and seven agematched healthy controls) as previously described [17]. In brief, baseline and R-(–)-deprenyl hydrochloride competition experiments were performed on adjacent brain sections (20 µm thickness). Prepared frozen brain tissues were warmed to room temperature, air-dried, and preincubated in a buffer saline solution (138 mM NaCl and 27 mM KCl, pH 7.4 adjusted by NaOH) and 1% bovine serum albumin for 20 min. Brain tissues were then airdried and incubated with ¹⁸F-THK5351 (1.85 µCi) and the R-(-)-deprenyl hydrochloride challenge was performed at concentrations of 150 nM and 500 nM for 150 min. After the incubation, brain tissues were dipped three times in the buffer solution for 3 min each time, once in 4 °C water for 30 s, and dried under a stream of cool air. The brain sections were subsequently exposed to a radioluminographic imaging plate (Fujifilm BAS-MS2025) for 20 min and obtained using FUJIFILM BAS-5000. Activity in photostimulated luminescence units per mm² was measured using Image Gauge 4.0 (Fujifilm) and the percentage of ¹⁸F-THK5351 total binding after R-(–)-deprenyl hydrochloride challenge was calculated.

Statistical analysis

The statistical analysis was performed using GraphPad Prism Version 7.0b (GraphPad Software, La Jolla, California, USA; www.graphpad.com). The baseline and the post-selegiline regional ¹⁸F-THK5351 SUVs were compared using paired *t* test analyses. The Bonferroni correction was used to correct the aforementioned analyses for multiple comparisons.

Results

Baseline demographics and global ¹⁸F-AZD4694 SUVR of the study participants are summarized in Table 1. Patients 4, 5 and 8 are classified as amyloid-negative and patients 1, 2, 3, 6, and 7 are classified as amyloid-positive (Fig. 1).

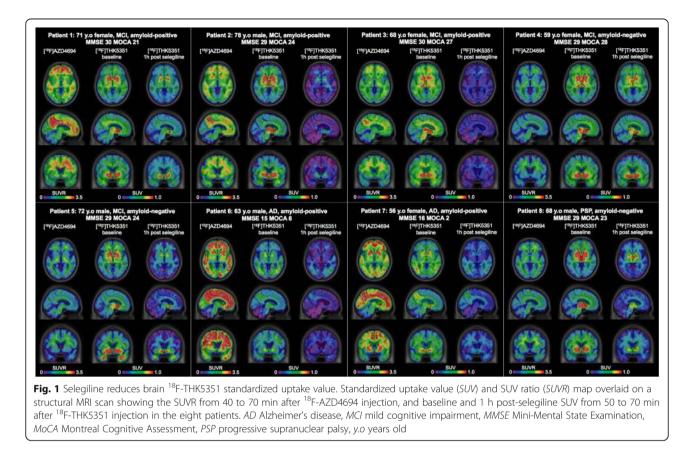
The ¹⁸F-THK5351 SUV map demonstrated an average of 36.7 to 51.8% regional uptake reduction in the post-selegiline scans from the baseline scans (Fig. 1). At baseline, the mean SUVs were highest in the basal ganglia (0.64 ± 0.11) followed by the thalamus (0.62 ± 0.14) (Fig. 2). In the post-selegiline scans, there were statistically significant regional SUV declines compared to the baseline scans. The SUV reduction was greatest in the thalamus (51.8%), followed by the basal ganglia (51.4%). MAO-B inhibition also reduced ¹⁸F-THK5351 SUVs in the cerebellar cortex (41.6%).

In the three MCI individuals who underwent a third ¹⁸F-THK5351 PET scan 9–28 days after the selegiline administration, the SUV remained reduced (Fig. 3). There was no consistent regional ¹⁸F-THK5351 SUVR reduction in the post-selegiline scans from baseline (Figs. 3 and 4), which is in line with the anticipated differences in the PHF to MAO-B ratio in different brain regions of

Table 1 Baseline demographics

	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient 6	Patient 7	Patient 8
Diagnosis	MCI	MCI	MCI	MCI	MCI	AD	AD	PSP
Age (years)	71	78	68	59	72	63	55	68
Gender	Female	Male	Female	Female	Male	Male	Female	Male
MMSE	30	29	30	29	29	15	16	29
MoCA	21	24	27	28	24	6	2	23
Global ¹⁸ F-AZD4694 SUVR	2.19	1.89	1.73	0.96	0.94	2.64	2.24	1.04

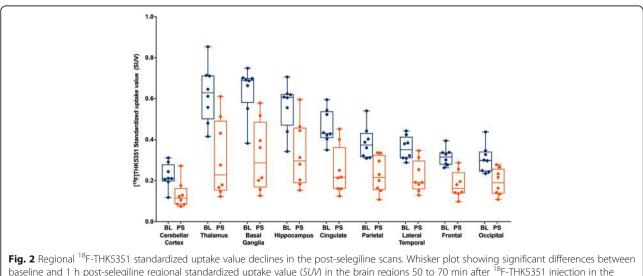
AD Alzheimer's disease, MCI mild cognitive impairment, MMSE Mini-Mental State Examination, MoCA Montreal Cognitive Assessment, PSP progressive supranuclear palsy, SUVR standardized uptake value ratio

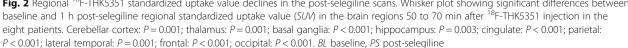


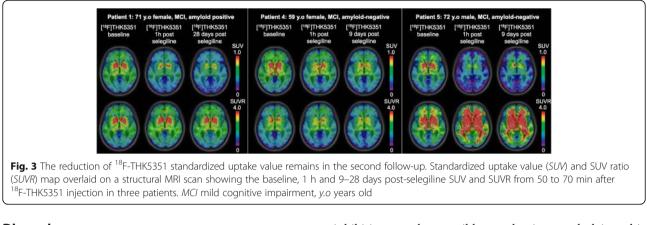
different participants, including the cerebellar cortex reference region.

Autoradiography in postmortem brain sections of AD patients and healthy controls further demonstrated a reduction of total ¹⁸F-THK5351 uptake following 150

nM and 500 nM R-(–)-deprenyl hydrochloride challenge (Fig. 5a). This reduction was greater in the striatum, a region rich in MAO-B but with negligible PHF, than in the prefrontal cortex and hippocampus where both PHF and MAO-B are present (Fig. 5b).



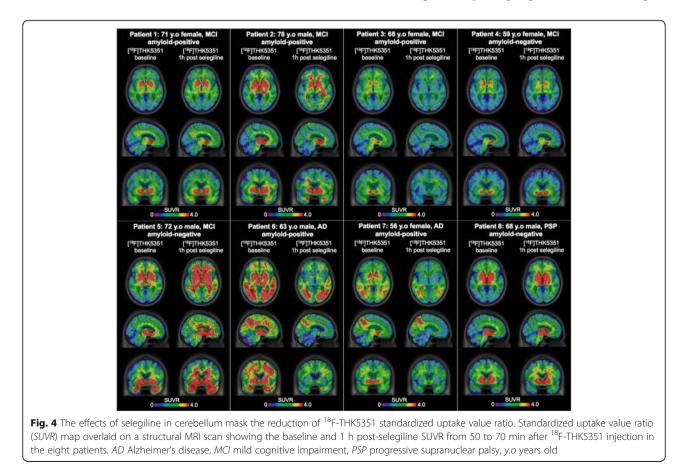


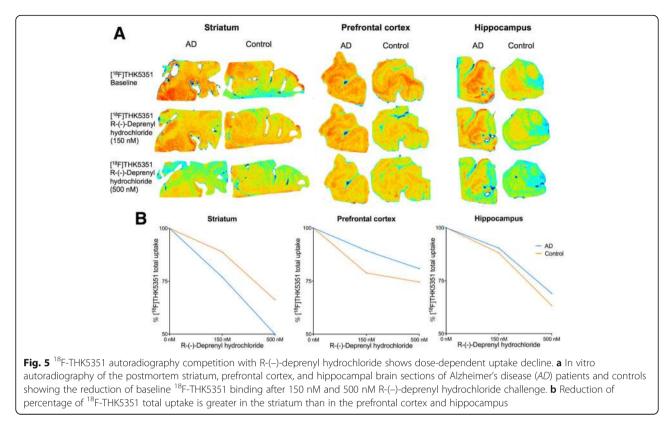


Discussion

In summary, we showed that brain and cerebellar cortex MAO-B availability affects ¹⁸F-THK5351 SUV. Following a single 10 mg oral dose of selegiline, ¹⁸F-THK5351 SUVs decreased by approximately 30 to 50% depending on the brain region, with the highest decline noted in the basal ganglia and thalamus which are known to express the highest concentrations of MAO-B in the brain [4]. The in vitro autoradiography blocking experiments showed similar effects. The long-lasting selegilineevoked ¹⁸F-THK5351 SUV decline indicates MAO-B inhibition as the possible mechanism underlying this effect.

MAO-B is a protein highly expressed in all brain regions. It is compartmentalized in the outer membrane of the astrocytes mitochondria [18]. During the normal aging process, global MAO-B availability increases at the rate of nearly 9% per decade [19, 20]. In fact, MAO-B imaging has been proposed as a biomarker for astrocytosis in various neurodegenerative conditions associated with cell death or activation of immune responses, with some investigators reporting higher MAO-B binding in





AD patients versus aged-matched controls [21–23]. Moreover, MAO-B inhibition has significant therapeutic effects for mood disorders and Parkinson's disease, and it is considered a therapeutic target for various neurodegenerative conditions [24].

The high retention of tau imaging agents such as ¹⁸F-THK5351 and ¹⁸F-flortaucipir to brain structures known to be devoid of PHF has been attributed to off-target binding sites. It is important to emphasize that off-target binding implies specific (saturable) binding of the imaging agent to proteins other than the primary target for which it was intended. Generally, it is desirable that the affinity of the target binding sites be higher by at least one order of magnitude as compared to that of the offtarget binding sites, although the lack of such a specificity might be acceptable if the target and off-target binding sites do not coexist in the same brain region [25]. Our results suggest that ¹⁸F-THK5351 binds to MAO-B and PHF with similar affinity, which might be detrimental to PHF quantification in the brain. However, given that ¹⁸F-THK5351 is biochemically different from ¹⁸F-flortaucipir, our findings cannot be generalized.

Interestingly, the acute selegiline challenge conducted in this study evoked the highest ¹⁸F-THK5351 SUV decline in the thalamus and striatum, which express high levels of MAO-B [4], and are also known to be devoid of PHF. However, it is worth mentioning that the magnitude of MAO-B inhibition in the aforementioned regions was variable, as described by previous studies [26, 27]. This variability can be attributed to individual differences in selegiline absorption, biodistribution, or pharmacodynamics. Furthermore, the exposure to MAO-B inhibitors present in tobacco, certain antidepressants, or herbal therapies may also influence such variability [28, 29].

The long-lasting effects of selegiline support MAO-B inhibition as the main mechanism involved in the effects observed by this study. Indeed, previous studies conducted in vivo estimate that the de novo synthesis half-life of MAO-B is 40 days [30]. Our autoradiography results conducted in postmortem brain of AD patients (Fig. 5) also support ¹⁸F-THK5351 binding to both MAO-B and PHF. Importantly, the enrolment of a clinically heterogeneous population in this study was intended to test whether MAO-B availability affects ¹⁸F-THK5351 SUV independently of other specific pathophysiological processes.

The selegiline challenge in the present study affected cortical ¹⁸F-THK5351 SUVs (Fig. 1) in regions known to accumulate PHF, such as the mesial basal temporal, temporal, and parietal cortices [31]. These regions also express high concentrations of MAO-B [5]. The visual assessments of ¹⁸F-THK5351 SUV images obtained in amyloid-positive individuals revealed the signature previously described in AD patients [2], which remained after the challenge with selegiline. Similarly, the high uptake

in the midbrain and basal ganglia of our patient with PSP remained after the post-selegiline scan. This may support the concept that ¹⁸F-THK5351 binds to 4-R tau pathology in the midbrain and basal ganglia, which is consistent with the imaging signature recently described [32]. The AD and PSP signatures observed after the selegiline challenge represent images with decreased effects of MAO-B availability.

We found that cerebellar cortex ¹⁸F-THK5351 SUVs declined to a lesser extent as compared to cerebral cortical regions following the selegiline challenge. This observation is consistent with previous studies showing declines in cerebellar MAO-B availability following pharmacological inhibition [5, 33]. In our study, the quantification of ¹⁸F-THK5351 uptake using SUVR with the cerebellar cortex as a reference region was also sensitive to the selegiline challenge. Although cerebellar MAO-B availability invalidates the use of this region as a reference to derive ¹⁸F-THK5351 SUVR in the context of this study, future research should focus on methodology to correct MAO-B bias in ¹⁸F-THK5351 SUVR measurements [34].

The use of imaging agents with low specificity is acceptable to analyze PET images from brain regions without off-target binding sites [35, 36]. However, the interpretation of PET data obtained using tracers with low specificity becomes challenging when target and offtarget binding sites co-localize within specific anatomical regions. In this situation, the major issue is obtaining PET signals of interest without the confounding effects from off-target sites. Tracer specificity becomes even more important in the case of imaging agents targeting abnormal protein aggregates. At early stages of the disease, when the load of abnormal protein aggregates is negligible, as compared to the availability of off-target binding sites, one could expect a higher confounding signal from the off-target binding [5, 37]. By contrast, in later stages of the disease, the relative contribution of off-target binding is reduced given the higher concentrations of protein aggregates in the tissue, despite high concentrations of MAO-B in the hippocampus, temporal neocortex, and cingulate gyrus [5]. It is also worth noting that the visual appearance of ¹⁸F-THK5351 PET images is not inconsistent with the pathological distribution of NFTs reported in Braak staging [38]. The potential reason for this is that, as neurons are compromised by NFTs, astrocytosis ensues in these regions [31]. The increase of MAO-B is likely due to astrocytosis in response to NFTs, and the PET signals here could be considered as a combination of both PHF and astrocytosis. This would need to be verified by future studies.

The interpretation of these results should take into consideration the following methodological limitations. Firstly, the present study was not conducted using full dynamic scans with an arterial input function, but relied on the SUV measured on ¹⁸F-THK5351 images obtained from 50 to 70 min after tracer injection [3] which is susceptible to physiological sources of variability. Secondly, although previous studies show that chronic administration of selegiline does not significantly affect cerebral blood flow, it has been reported that acute selegiline administration may induce rapid brain vasodilation via nitric oxide release [39]. Hence, it is possible that the selegiline dose in the present study may induce cerebral blood flow changes and affect accurate quantification of ¹⁸F-THK5351 uptake.

Conclusions

The present study supports the concept that MAO-B is an ¹⁸F-THK5351 off-target binding site. As such, the current methods for analyzing ¹⁸F-THK5351 inevitably incorporate undesired confounding signal from MAO-B availability, which probably dominates PET signals in early stages of AD. Therefore, it would be appropriate for the methods used to quantify ¹⁸F-THK5351 to take into account the cortical MAO-B signal. Importantly, heterogeneous MAO-B availability across the cortex may also limit the quantification of ¹⁸F-THK5351 uptake using reference brain regions. In addition, with regards to the use of ¹⁸F-THK5351 in longitudinal studies and clinical trials in AD, one should be aware that both MAO-B and tau binding may increase with age at different rates in different brain regions of different individuals, whilst tau binding will increase in the same fashion with disease progression, or potentially decrease in response to an efficacious treatment. As such, experiments should be designed and interpreted with care to account for these processes. Finally, it is important to emphasize that the confounding effect of MAO-B availability in patients with high loads of PHF should be further evaluated by subsequent studies.

Abbreviations

AD: Alzheimer's disease; HRRT: High-Resolution Research Tomograph; MAO: Monoamine oxidase; MAO-B: Monoamine oxidase B; MCI: Mild cognitive impairment; MMSE: Mini-Mental State Examination; MNI: Montreal Neurological Institute; MOCA: Montreal Cognitive Assessment; MRI: Magnetic resonance imaging; NFT: Neurofibrillary tangle; NINCDS-ADRDA: National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's disease and Related Disorders Association; NINDS-SPSP: National Institute of Neurological Disorders and Stroke and the Society for PSP; PET: Positron emission tomography; PHF: Paired helical filaments; PSP: Progressive supranuclear palsy; SUV: Standardized uptake value; SUVR: Standardized uptake value ratio

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Availability of data and materials

The data supporting the conclusions of this article are available from the corresponding author upon request.

Authors' contributions

KPN performed the design and execution of the study, processing and analysis of the image data, composed the figures, and drafted the manuscript. TAP, SM, and MS performed processing and analysis of the image data, and revised the manuscript. JT performed the execution of the study, and revised the manuscript. JT performed the design of the study, processing and analysis of the image data, autoradiography, and revised the manuscript. MCG performed postmortem brain section diagnoses, and revised the manuscript. JPS assisted with the interpretation and analysis of the image data, and revised the manuscript. QG, RH, RAC, and NO revised the manuscript. SG assisted with the study design, execution of the study, and revised the manuscript. PRN performed the conception and design of the study, analysis of the image data, and analysis of and approved the final version of the study. All authors read and approved the final version of the manuscript.

Authors' information

Not applicable.

Competing interests

QG and RAC are employees of AbbVie Inc. SG received honoraria for serving on the scientific advisory boards of Alzheon, Axovant, Lilly, Lundbeck, Novartis, Schwabe, and TauRx, on the DSMB of a study sponsored by Eisai, and studies run by ADCS and by ATRI. All other authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

The study has been approved by the McGill institutional review board, study number A10-M38-16A, and all participants signed an informed consent form. The use of postmortem brain sections in this study has been approved by the Douglas Institute's Research Ethics Board and the Douglas-Bell Canada Brain Bank's Scientific Review Committee.

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References

- Villemagne VL, Fodero-Tavoletti MT, Masters CL, Rowe CC. Tau imaging: early progress and future directions. Lancet Neurol. 2015;14(1):114–24.
- Johnson KA, Schultz A, Betensky RA, Becker JA, Sepulcre J, Rentz D, et al. Tau positron emission tomographic imaging in aging and early Alzheimer disease. Ann Neurol. 2016;79:110–9.
- Harada R, Okamura N, Furumoto S, Furukawa K, Ishiki A, Tomita N, et al. 18F-THK5351: a novel PET radiotracer for imaging neurofibrillary pathology in Alzheimer's disease. J Nucl Med. 2015;57:1–43.
- Fowler JS, MacGregor RR, Wolf AP, Arnett CD, Dewey SL, Schlyer D, et al. Mapping human brain monoamine oxidase A and B with 11C-labeled suicide inactivators and PET. Science. 1987;235:481–5.
- Tong J, Meyer JH, Furukawa Y, Boileau I, Chang L-J, Wilson AA, et al. Distribution of monoamine oxidase proteins in human brain: implications for brain imaging studies. J Cereb Blood Flow Metab. 2013;33:863–71.
- Lee MK, Hwang BY, Lee SA, Oh GJ, Choi WH, Hong SS, et al. 1-methyl-2undecyl-4(1H)-quinolone as an irreversible and selective inhibitor of type B monoamine oxidase. Chem Pharm Bull (Tokyo). 2003;51:409–11.
- Mahmood I. Clinical pharmacokinetics and pharmacodynamics of selegiline. An update. Clin Pharmacokinet. 1997;33:91–102.
- Folstein MF, Folstein SE, McHugh PR. "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. J Psychiatr Res. 1975;12:189–98.
- Nasreddine ZS, Phillips NA, Bédirian V, Charbonneau S, Whitehead V, Collin I, et al. The Montreal Cognitive Assessment, MoCA: A Brief Screening Tool For Mild Cognitive Impairment. J. Am. Geriatr. Soc. Blackwell Science Inc; 2005; 53:695–9.
- Petersen RC. Mild cognitive impairment as a diagnostic entity. J Intern Med. 2004;256(3):183–94.
- McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM. Clinical diagnosis of Alzheimer's disease: Report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. Neurology. 2011;77:333.
- Litvan I, Agid Y, Calne D, Campbell G, Dubois B, Duvoisin RC, et al. Clinical research criteria for the diagnosis of progressive supranuclear palsy (Steele-Richardson-Olszewski syndrome): report of the NINDS-SPSP international workshop. Neurology. 1996;47:1–9.
- Cselényi Z, Jönhagen ME, Forsberg A, Halldin C, Julin P, Schou M, et al. Clinical validation of 18F-AZD4694, an amyloid-β-specific PET radioligand. J Nucl Med. 2012;53:415–24.
- Ad-Dab'bagh Y, Lyttelton O, Muehlboeck JS, Lepage C, Einarson D, Mok K, et al. The CIVET image-processing environment: a fully automated comprehensive pipeline for anatomical neuroimaging research. Proc. 12th Annu. Meet. Organ. Hum. Brain Mapp. 2006. p. 2266.
- Zijdenbos AP, Forghani R, Evans AC. Automatic "pipeline" analysis of 3-D MRI data for clinical trials: application to multiple sclerosis. IEEE Trans Med Imaging. 2002;21:1280–91.
- Pascoal TA, Mathotaarachchi S, Mohades S, Benedet AL, Chung C-O, Shin M, et al. Amyloid-β and hyperphosphorylated tau synergy drives metabolic decline in preclinical Alzheimer's disease. Mol. Psychiatry. Nature Publishing Group; 2017;22:306–11.
- Parent MJ, Bedard M-A, Aliaga A, Minuzzi L, Mechawar N, Soucy J-P, et al. Cholinergic depletion in Alzheimer's disease shown by [¹⁸F]FEOBV autoradiography. Int J Mol Imaging. 2013;2013:1–6.
- Levitt P, Pintar JE, Breakefield XO. Immunocytochemical demonstration of monoamine oxidase B in brain astrocytes and serotonergic neurons. Proc Natl Acad Sci U S A. 1982;79:6385–9.
- Saura J, Andrés N, Andrade C, Ojuel J, Eriksson K, Mahy N, et al. Biphasic and region-specific MAO-B response to aging in normal human brain. Neurobiol Aging. 1980;18:497–507.
- Fowler JS, Volkow ND, Wang GJ, Logan J, Pappas N, Shea C, et al. Agerelated increases in brain monoamine oxidase B in living healthy human subjects. Neurobiol Aging. 1997;18:431–5.
- Gulyás B, Pavlova E, Kása P, Gulya K, Bakota L, Várszegi S, et al. Activated MAO-B in the brain of Alzheimer patients, demonstrated by [11C]-I-deprenyl using whole hemisphere autoradiography. Neurochem Int. 2011;58:60–8.

- Schöll M, Carter SF, Westman E, Rodriguez-Vieitez E, Almkvist O, Thordardottir S, et al. Early astrocytosis in autosomal dominant Alzheimer's disease measured in vivo by multi-tracer positron emission tomography. Sci Rep. 2015;5:16404.
- Carter SF, Scholl M, Almkvist O, Wall A, Engler H, Langstrom B, et al. Evidence for astrocytosis in prodromal Alzheimer disease provided by 11Cdeuterium-L-deprenyl: a multitracer PET paradigm combining 11C-Pittsburgh Compound B and 18F-FDG. J Nucl Med. 2012;53:37–46.
- Foley P, Gerlach M, Youdim MB, Riederer P, Johnston JP, Kochersperger LM, et al. MAO-B inhibitors: multiple roles in the therapy of neurodegenerative disorders? Parkinsonism Relat Disord. 2000;6:25–47.
- Takano A, Halldin C, Farde L. Neuroimaging in psychiatric drug development and radioligand development for new targets. PET SPECT Psychiatry. Berlin, Heidelberg: Springer Berlin Heidelberg; 2014. p. 3–14.
- Logan J, Fowler JS, Volkow ND, Wang GJ, MacGregor RR, Shea C, et al. Reproducibility of repeated measures of deuterium substituted [11C]Ldeprenyl ([11C]L-deprenyl-D2) binding in the human brain. Nucl Med Biol. 2000;27:43–9.
- Freedman NMT, Mishani E, Krausz Y, Weininger J, Lester H, Blaugrund E, et al. In vivo measurement of brain monoamine oxidase B occupancy by rasagiline, using (11)C-I-deprenyl and PET. J Nucl Med. 2005;46:1618–24.
- Fowler JS, Volkow ND, Wang G-J, Pappas N, Logan J, MacGregor R, et al. Inhibition of monoamine oxidase B in the brains of smokers. Nature. 1996;379:733–6.
- Vina D, Serra S, Lamela M, Delogu G. Herbal natural products as a source of monoamine oxidase inhibitors: a review. Curr Top Med Chem. 2012;12:2131–44.
- Fowler JS, Volkow ND, Logan J, Wang G-J, Mac Gregor RR, Schlyer D, et al. Slow recovery of human brain MAO B after L-deprenyl (Selegeline) withdrawal. Synapse. 1994;18:86–93.
- Serrano-Pozo A, Frosch MP, Masliah E, Hyman BT. Neuropathological alterations in Alzheimer disease. Cold Spring Harb Perspect Med. 2011;1:a006189.
- Ishiki A, Harada R, Okamura N, Tomita N, Rowe CC, Villemagne VL, et al. Tau imaging with [18 F]THK-5351 in progressive supranuclear palsy. Eur. J. Neurol. 2017;24:130–6.
- Bench CJ, Price GW, Lammertsma AA, Cremer JC, Luthra SK, Turton D, et al. Measurement of human cerebral monoamine oxidase type B (MAO-B) activity with positron emission tomography (PET): a dose ranging study with the reversible inhibitor Ro 19-6327. Eur J Clin Pharmacol. 1991;40:169–73.
- Lammertsma AA, Bench CJ, Hume SP, Osman S, Gunn K, Brooks DJ, et al. Comparison of methods for analysis of clinical [11C]raclopride studies. J Cereb Blood Flow Metab. 1996;16:42–52.
- Ekelund J, Slifstein M, Narendran R, Guillin O, Belani H, Guo N-N, et al. In vivo DA D1 receptor selectivity of NNC 112 and SCH 23390. Mol Imaging Biol. 2007;9:117–25.
- Kiss B, Horti F, Bobok A. In vitro and in vivo comparison of [3H](+)-PHNO and [3H]raclopride binding to rat striatum and lobes 9 and 10 of the cerebellum: a method to distinguish dopamine D3 from D2 receptor sites. Synapse. 2011;65:467–78.
- Marquié M, Normandin MD, Vanderburg CR, Costantino IM, Bien EA, Rycyna LG, et al. Validating novel tau positron emission tomography tracer [F-18]-AV-1451 (T807) on postmortem brain tissue. Ann Neurol. 2015;78:787–800.
- Braak H, Braak E. Staging of Alzheimer's disease-related neurofibrillary changes. Neurobiol Aging. 1995;16:271–8.
- Thomas T, McLendon C, Thomas G. L-deprenyl: nitric oxide production and dilation of cerebral blood vessels. Neuroreport. 1998;9:2595–600.

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