

Most Highly Cytokinergic IgEs Have Polyreactivity to Autoantigens

Jun-ichi Kashiwakura,^{1,2} Yoshimichi Okayama,² Masutaka Furue,³ Kenji Kabashima,^{4†} Shinji Shimada,⁵ Chisei Ra,² Reuben P. Siraganian,⁶ Yuko Kawakami,¹ Toshiaki Kawakami^{1*}

¹Division of Cell Biology, La Jolla Institute for Allergy and Immunology, La Jolla, CA, USA

²Division of Molecular Cell Immunology and Allergology, Advanced Medical Research Center, Nihon University Graduate School of Medical Science, Itabashi-ku, Tokyo, Japan

³Department of Dermatology, Kyushu University, Fukuoka, Japan

⁴Department of Dermatology, University of Occupational and Environmental Health, Kitakyushu, Japan

⁵Department of Dermatology, University of Yamanashi, Yamanashi, Japan

⁶Receptors and Signal Transduction Section, OIIB, National Institute for Dental and Craniofacial Research, National Institutes of Health, Bethesda, MD, USA

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Purpose: Monomeric IgE molecules, when bound to the high-affinity receptor, exhibit a vast heterogeneity in their ability to induce survival promotion and cytokine production in mast cells. At one end of this spectrum, highly cytokinergic (HC) IgEs can induce potent survival promotion, degranulation, cytokine production, migration, etc., whereas at the other end, poorly cytokinergic (PC) IgEs can do so inefficiently. In this study, we investigated whether IgEs recognize autoantigens and whether IgEs' binding of autoantigens correlates with difference in HC versus PC properties. **Methods:** Enzyme-linked immunosorbent assays were performed to test whether IgEs bind antigens. Histamine-releasing factor in human sera was quantified by western blotting. Cultured mast cells derived from human cord blood were used to test the effects of human sera on cytokine production. **Results:** Most (7/8) of mouse monoclonal HC IgEs exhibited polyreactivity to double-stranded DNA (dsDNA), single-stranded DNA (ssDNA), β -galactosidase, thyroglobulin and/or histamine-releasing factor. By contrast, mouse PC IgEs failed to react with these antigens. A human monoclonal HC IgE also showed polyreactivity to histamine-releasing factor, dsDNA and ssDNA. Interestingly, sera from atopic dermatitis patients showed increased reactivity to ssDNA and β -galactosidase and increased levels of histamine-releasing factor. Some atopic dermatitis patients, but not healthy individuals, had substantial serum levels of HRF-reactive IgE. Sera from atopic dermatitis patients with high titers of DNA-reactive IgE could induce several fold more IL-8 secretion in human mast cells than sera from healthy individuals. **Conclusions:** The results show that most HC, but not PC, IgEs exhibit polyreactivity to autoantigens, supporting the autoimmune mechanism in the pathogenesis of atopic dermatitis.

Key Words: Atopic dermatitis; mast cell; cytokine; IgE; autoantigen; histamine-releasing factor

INTRODUCTION

Mast cells are the key effector cell type in IgE-mediated immediate hypersensitivity and allergic disorders.^{1,2} Mast cells bound by antigen-specific IgE via the high-affinity receptor for IgE (Fc ϵ RI) must encounter multivalent antigen for their activation.³ Activated mast cells secrete preformed proinflammatory mediators, such as histamine, serotonin, nucleotides, proteases, and TNF- α , and synthesize and secrete lipid mediators (such as leukotrienes and prostaglandins), various cytokines and chemokines. In contrast to this traditional view, we and others showed that IgE binding to Fc ϵ RI in the absence of specific antigen engenders several biological outcomes in mast cells: upregulation of cell surface expression of Fc ϵ RI,⁴⁻⁶ survival,^{7,8} increase in his-

tamine content,⁹ histamine release, leukotriene release, receptor internalization, DNA synthesis,¹⁰ increased responses to compound 48/80 and substance P,¹¹ increase in filamentous actin content,¹² membrane ruffling,¹³ adhesion to fibronectin,¹⁴ and migration.¹⁵ These effects are due to binding to Fc ϵ RI by monomeric, but not aggregate, IgE molecules.^{7,8,12} Fc ϵ RI cross-

Correspondence to: Toshiaki Kawakami, MD, PhD, Division of Cell Biology, La Jolla Institute for Allergy and Immunology, 9420 Athena Circle, La Jolla, California 92037, USA.

Tel: +1-858-752-6814; Fax: +1-858-752-6986; E-mail: toshi@liai.org

Received: February 6, 2012; Revised: April 13, 2012; Accepted: May 2, 2012

• There are no financial or other issues that might lead to conflict of interest.

• [†]Present address: Department of Dermatology, Kyoto University, Kyoto, Japan.

linking appears essential for mast cell activation induced by monomeric IgE,¹⁶ similar to that by IgE+antigen or IgE+anti-IgE.^{17,18}

Not all IgE molecules can induce all the activation events in the absence of antigen: IgEs display a wide spectrum of heterogeneity in their ability to induce the production and secretion of IL-6 and TNF- α , two tested cytokines produced by mouse mast cells, with HC IgEs at one extreme end and PC IgEs at the other.¹⁰ More extensive receptor aggregation can be induced by HC IgEs than by PC IgEs. Consistent with such a difference, strong HC IgEs can induce each of the tested activation events, whereas PC IgEs induce only a limited set of activation events, and to a lesser extent.¹⁹ Polyclonal IgE from mice and humans suffering from atopic dermatitis (AD) can enhance survival and cytokine production in mast cell cultures, indicating that monomeric IgE effects are operative in polyclonal situations as well.²⁰

Compared to numerous reports on effects of monomeric IgE on mouse mast cells, effects of human IgE on human mast cells have been less studied. However, using human cord blood derived mast cells (CBMCs), Gilcrest et al.²¹ reported that the chemokine I-309 RNA and protein levels were upregulated not only in response to IgE+anti-IgE stimulation but also by IgE alone and these responses were further augmented in the presence of IL-4. Cruse et al.²² also reported that human monomeric myeloma IgE stimulated cultured human lung mast cells to release histamine, leukotriene C4, and IL-8 in the absence of a specific antigen. Matsuda et al.¹⁶ showed that human monomeric IgE in the absence of specific antigen enhanced IL-8 and monocyte chemoattractant protein 1 (MCP-1) production in cultured human mast cells, and this response was augmented by preincubation of the cells in IL-4. However, it is still unclear whether the effects of monomeric HC versus PC IgEs exist in the human system. Since heterogeneity of human IgEs in the ability to prime basophils to stimulation with histamine-releasing factor (HRF) has been reported,²³ it is possible that some highly allergic patients might produce HC IgEs, which may activate mast cells or basophils in the absence of antigen.

Our recent study demonstrated that ~30% of the tested mouse IgE mAbs bind to HRF and that HRF+HRF-reactive IgE can activate mast cells.²⁴ Since HRF has two IgE-binding sites and can be present as a dimer, it is assumed that IgE-bound receptors can be cross-linked by HRF. Similar situations could occur if receptor-bound IgE react with multivalent (auto)antigen. Therefore, we investigated whether HC IgEs react with various antigens.

MATERIALS AND METHODS

IgE preparations

Mouse monoclonal IgEs were purified by ammonium sulfate precipitation from serum-free culture supernatants of hybridomas or peritoneal exudates of hybridoma-bearing mice, followed by DEAE column chromatography.²⁵ Some ELISA exper-

iments were performed with hybridoma culture supernatants. Sources of IgEs: H1 DNP- ϵ -26 (26) and H1 DNP- ϵ -206 (206) from Fu-Tong Liu, University of California, Davis; AR40EA, BE1BD (BE1BD2D5), BE2, BE4 (BE4C2C3), DNP48BC, DNP48BD, HB30, and R25 from R.P.S.; C38-2, C48-2, and 27-74 from BD Biosciences; SPE-7 from Sigma-Aldrich; IGELa2 from American Type Culture Collection (ATCC) (TIB142).

Human IgEs: SKO-007 monoclonal IgE was purified by ammonium sulfate precipitation from serum-free culture supernatants of hybridomas (ATCC: CRL-8033-1). HE1 monoclonal IgE from a hybridoma was purchased from Diatec Monoclonals AS (Oslo, Norway). IgEs derived from multiple myeloma patients (BS- κ and ART) were purchased from The Binding Site Inc. (San Diego, CA, USA) or Athens Research & Technology (Athens, GA, USA). IgE from the multiple myeloma patient PS was a gift from Dr. Kimishige Ishizaka, LIAI. All IgE samples were dialyzed extensively against PBS, and ultracentrifuged at 100,000 \times g for 30 minutes before use to remove possible aggregates.

Sera from AD patients and normal subjects

After the informed consent was obtained, the sera of AD patients diagnosed according to the criteria of Hanifin and Rajka²⁶ were collected. Total numbers of AD patients and normal controls are 41 and 25, respectively. All studies involving human subjects were conducted in accordance with the guidelines of the World Medical Association's Declaration of Helsinki and approved by the Institutional Review Boards of La Jolla Institute for Allergy and Immunology, Kyushu University, University of Occupational and Environmental Health and University of Yamanashi.

Enzyme-linked immunosorbent assay (ELISA)

Calf thymus double-stranded DNA (dsDNA) and single-stranded DNA (ssDNA), β -galactosidase from bovine liver, thyroglobulin from bovine thyroid, insulin from bovine pancreas, LPS (all from Sigma-Aldrich) and recombinant HRF tagged with 6x histidine (all 10 μ g/mL in PBS) were coated in 96-well plates for overnight at 4°C. After the wells were blocked with 10% FCS-PBS, mouse IgE molecules (10 μ g/mL) were added. Bound mouse IgEs were detected by biotinylated anti-mouse IgE mAb (BD Bioscience PharMingen, San Diego, CA, USA), followed by streptavidin-HRP conjugates. Color was developed and absorbance at 450 nm was measured. In some experiments, human IgEs (10 μ g/mL) or sera from healthy subjects and AD patients (1/10 dilution) were incubated in the well. Bound human IgE molecules were detected by biotinylated anti-human IgE mAb (BD Bioscience PharMingen), followed by streptavidin-HRP.

Human mast cell culture

Human CBMCs were generated as described.²⁷ Briefly, CD34⁺ cells were purified from umbilical cord blood mononuclear cells

and cultured in serum-free Iscove's methylcellulose medium (IMDM; Stem Cell Technologies Inc., Vancouver, BC, Canada) containing 200 ng/mL SCF (PeproTech Inc., Rocky Hill, NJ, USA), 50 ng/mL IL-6 (PeproTech Inc.), 1 ng/mL IL-3 (PeproTech Inc.), 1% insulin-transferrin-selenium (Invitrogen, Carlsbad, CA, USA), 50 μ M 2-mercaptoethanol, 1% penicillin-streptomycin (Invitrogen), and 0.1% bovine serum albumin (Sigma-Aldrich, St Louis, MO, USA). On day 42 of culture, methylcellulose was dissolved in PBS and the cells were then suspended and cultured in IMDM (Invitrogen) supplemented with 100 ng/mL SCF, 50 ng/mL IL-6, and 5% fetal calf serum (Cansera, Rexdale, ON, Canada). After 15 weeks, the purity of the resulting CBMCs was ~98% (c-Kit⁺Fc ϵ RI⁺ by flow cytometry). The use of human cord blood was approved by the Institutional Review Board of Nihon University Graduate School of Medical Science.

Cytokine measurement

CBMCs were incubated with 10 ng/mL of IL-4 in the presence or absence of 1 μ g/mL of IgE for sensitization. After 24 hours, the cells were washed and resuspended in culture medium. The cells (1×10^5 /100 μ L) were then stimulated with IgE molecules for the indicated times. After stimulation, supernatants and cell pellets were collected for analysis of IL-8 and IL-6 expression and production. IL-8 production was measured using a human IL-8 ELISA kit (BD PharMingen, San Diego, CA, USA).

Cell survival

CBMCs were incubated with IL-4 in the presence or absence of IgE for sensitization. After 24 hours, the cells were washed and resuspended in SCF-free culture medium. The cells (1×10^5 /100 μ L) were then stimulated with anti-IgE, SCF, or IgE molecules for 72 hours. After 72 hours, cell survival was analyzed after staining with Trypan blue.

Real-time RT-PCR

Total cellular RNA was isolated from CBMCs with an RNeasy mini kit (Qiagen, Valencia, CA, USA) according to the manufacturer's instructions. An equal amount of total RNA (100 ng) was used for reverse transcription. Real-time RT-PCR was performed as follows: cDNA (10 ng) was amplified in 20 μ L in the presence of 1 μ L of "Assay-on-Demand" primer sets for IL-8, IL-6, and GAPDH purchased from Applied Biosystems (Foster City, CA, USA) in 7700 ABI thermal cyclers (Applied Biosystems). Relative expression levels were determined using Ct method.

Measurement of serum HRF

Human sera (10 μ L) were precipitated first with acetone. Precipitated materials were dissolved in SDS sample buffer and, together with dilution series of recombinant HRF-His6, analyzed by SDS-PAGE and blotted onto PVDF membranes. The blots were probed with anti-HRF antibody followed by HRP-conjugated secondary antibody. HRF bands were detected by

ECL reagent (Perkin-Elmer). The concentrations of HRF in the samples were determined by densitometry.

Statistical analysis

Data are expressed as mean \pm SD. Paired student *t* test was used for mean comparisons. Differences were considered statistically significant at *P* values < 0.05.

RESULTS

Mouse HC, but not PC, IgE molecules react with various autoantigens

HC IgEs are defined operationally by the ability to induce secretion of IL-6 and/or TNF- α from BMMCs, whereas PC IgEs lack this ability.¹⁰ We have recently shown that some of the tested IgEs interact with HRF.²⁴ We extended this line of study by testing whether mouse IgE mAbs interact with other antigens, such as dsDNA, ssDNA, β -galactosidase, thyroglobulin, insulin and LPS. ELISA data (Fig. 1) showed that most HC IgEs react with two or more of these antigens. For example, the most potent HC IgE SPE-7 reacted with dsDNA, ssDNA and thyroglobulin, in addition to DNP (hapten used to generate this mAb) and thioredoxin, which was previously shown to bind to SPE-7 IgE.²⁸ H1 DNP- ϵ -26 IgE reacted with dsDNA and ssDNA; C38-2 IgE reacted with HRF, dsDNA, ssDNA, β -galactosidase and thyroglobulin; IGELa2 IgE reacted with HRF, β -galactosidase and thyroglobulin. The possibility of non-specific binding was ruled out. For example, interactions of TNP-specific C38-2 and IGE-La2 IgEs with HRF were inhibited by TNP-glycine,²⁴ suggesting that the HRF-binding site in IgE overlaps at least in part with the antigen-binding site. By contrast, none of the 13 tested PC IgE mAbs reacted with the antigens under the same conditions. Neither HC nor PC IgEs reacted with insulin or LPS. These experiments clearly demonstrate that most of mouse HC, but not PC, IgEs react with multiple autoantigens.

Human IgE molecules also show the HC vs. PC heterogeneity

Human CBMCs express low levels of Fc ϵ RI on their cell surface.²⁹ Previous studies showed that incubation of human CBMCs with IL-4 enhances Fc ϵ RI expression²⁹ and renders the cells sensitive to stimulation with human IgE, which induces the production of chemokines, IL-8 and monocyte chemoattractant protein 1.¹⁶ Therefore, we used IL-4-preincubated human CBMCs in this study. To determine whether human IgE molecules show heterogeneity in inducing cytokine production in human mast cells, human CBMCs were incubated with 5 μ g/mL of human monoclonal and myeloma-derived IgE molecules for 2 hours, to measure cytokine mRNAs and proteins. As shown previously,¹⁶ quantitative RT-PCR showed that IL-8 mRNA expression is induced strongly by a monoclonal (HE1) IgE and weakly by another myeloma IgE, BS- κ (Fig. 2A). We could also show secretion of IL-8 protein induced by the HE1 IgE (Fig. 2B).

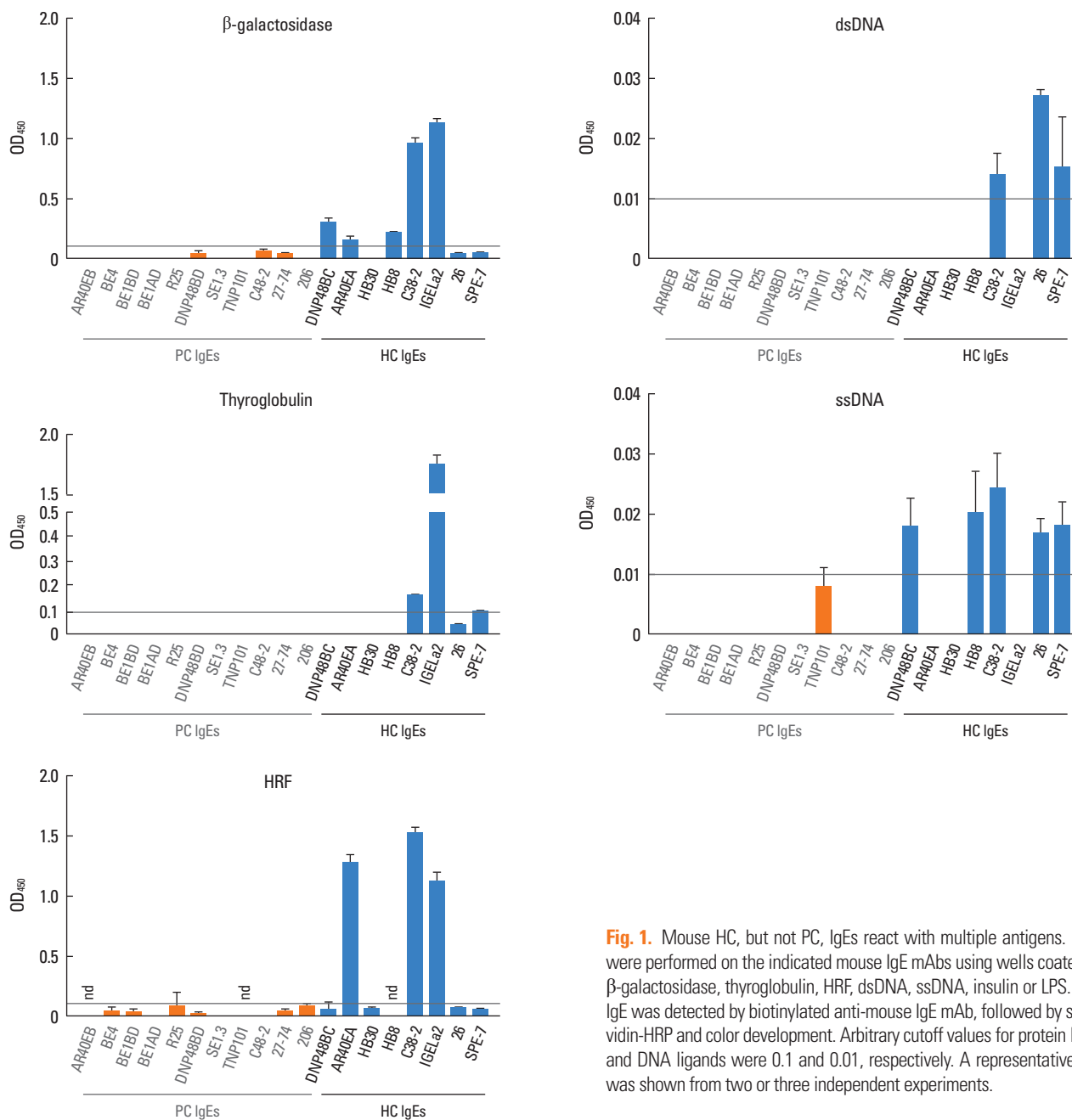


Fig. 1. Mouse HC, but not PC, IgEs react with multiple antigens. ELISAs were performed on the indicated mouse IgE mAbs using wells coated with β -galactosidase, thyroglobulin, HRF, dsDNA, ssDNA, insulin or LPS. Bound IgE was detected by biotinylated anti-mouse IgE mAb, followed by streptavidin-HRP and color development. Arbitrary cutoff values for protein ligands and DNA ligands were 0.1 and 0.01, respectively. A representative result was shown from two or three independent experiments.

However, the other tested monoclonal IgE preparations failed to induce IL-8 mRNA. Similarly, only HE1 IgE induced IL-6 mRNA (Fig. 2C). Secreted IL-6 levels could not be measured because CBMCs were cultured in IL-6-containing medium. HE1 IgE, but not SKO IgE, enhanced survival of CBMCs (Fig. 2D). Thus, these results indicate that HE1 is an HC IgE and others are PC IgEs.

A human HC IgE, but not PC IgEs, react with various autoantigens

We next performed ELISA to test whether human HC and PC

IgEs react with dsDNA and ssDNAs. As shown in Fig. 3, HE1 IgE, which was previously shown to bind to HRF²⁴ reacted with both dsDNA and ssDNA. By contrast, none of the human PC IgEs reacted with either DNA.

Sera from AD patients contain higher levels of HRF and ssDNA- or β -galactosidase-binding IgEs

The ability of some IgEs to bind to multivalent autoantigens such as HRF or DNA raised the possibility that mast cells and basophils can be activated by interacting with such an autoantigen via Fc ϵ RI-bound IgE. Given the importance of IgE in the

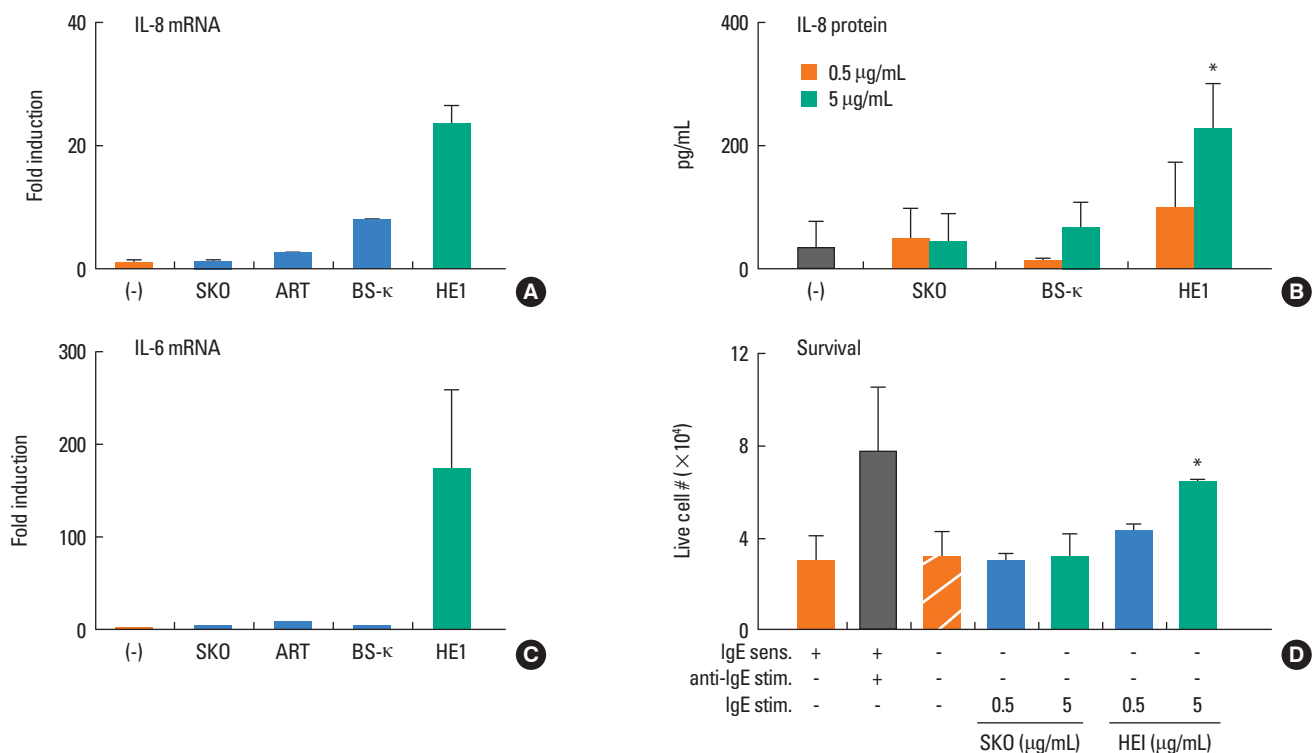


Fig. 2. A human IgE preparation can induce IL-8 and IL-6 production and promote survival in human mast cells. (A-C) Human CBMCs were incubated with 0 (PBS, [-]), 0.5 (white bar), or 5 (gray or black bar) μg/mL of the indicated human IgEs for 2 hr. (A,C) IL-8, IL-6, and GAPDH mRNA levels were measured by quantitative RT-PCR. The amount of cytokine mRNA was normalized against GAPDH mRNA and the relative value in PBS-treated cells was set at 1. (B) IL-8 protein was measured by ELISA. **P*<0.05 vs PBS control by Student's *t*-test. (D) Human CBMCs (1×10^5 cells) were incubated with 0 (PBS), 0.5, or 5 μg/mL of the indicated human IgEs in the absence of SCF and IL-6 for 72 hr. As controls, CBMCs were pre-incubated with 1 μg/mL of PS IgE in the presence of IL-4 for 24 hr, and washed cells were incubated with 0 or 1 μg/mL anti-IgE for 72 hr. Trypan blue excluding live cell numbers were counted. **P*<0.05 vs control (no IgE sensitization/no IgE stimulation) by Student's *t*-test.

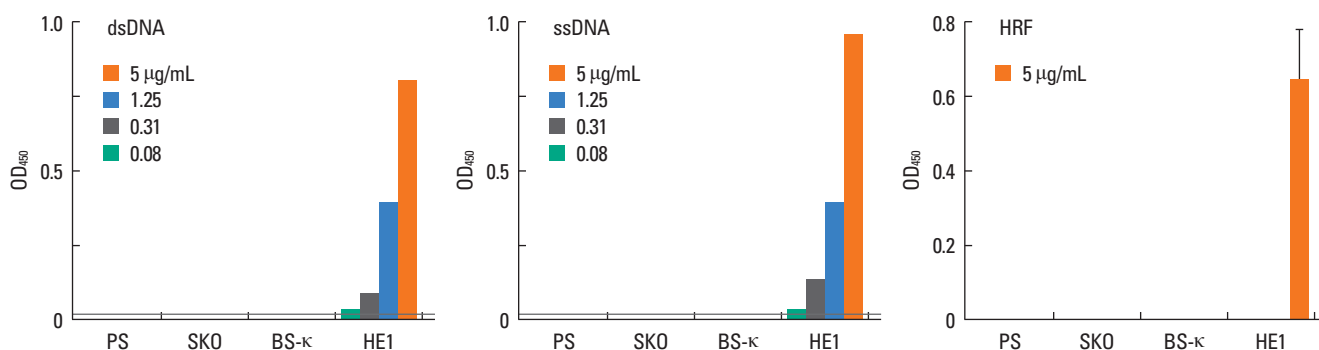


Fig. 3. A human HC IgE HE1 reacts with dsDNA, ssDNA and HRF. ELISAs were performed on the indicated human IgE samples, as shown in Fig. 1 except that anti-human IgE was used in place of anti-mouse IgE.

pathogenesis of AD, asthma and other allergic diseases³⁰⁻³³ and preferential IgE reactivity with autoantigens in AD,³⁴ we measured levels of IgE that reacted with dsDNA, ssDNA, β-galactosidase, and LPS in sera of AD patients. AD patients had significantly higher serum levels of ssDNA-reactive and β-galactosidase-reactive IgE than healthy controls (Fig. 4A and 4C). Levels of dsDNA-reactive IgE also tended to be higher than healthy con-

trols, although the difference did not reach statistical significance (Fig. 4B). AD patients also had higher serum levels of HRF than healthy controls (Fig. 4D). Four of 34 AD patients had measurable levels of IgE that reacted with HRF (Fig. 4E). Two of these patients had extremely high IgE levels of 48.7 and 384 μg/mL, whereas another patient had 0.17 μg/mL and IgE levels were not known for the other patient. By contrast, none of 25

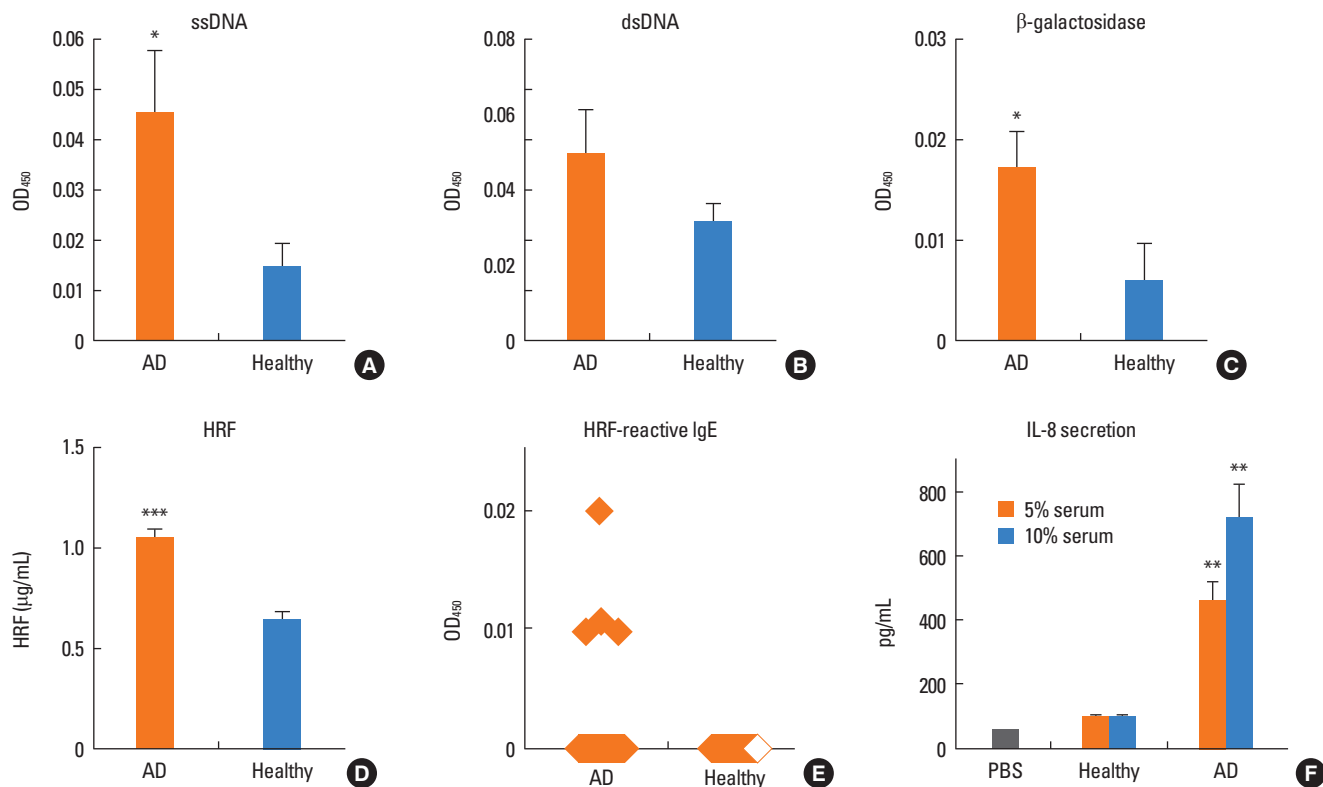


Fig. 4. AD patients have higher serum levels of ssDNA- or β -galactosidase-reactive IgE and HRF. ELISAs were performed on human sera using wells coated with ssDNA (A), dsDNA (B), β -galactosidase (C) and HRF (E). Sera were taken from cohorts of AD patients and healthy individuals. Note that low OD_{450} values, compared to those in Fig. 3, were obtained with sera diluted tenfold. (D) Serum HRF levels were quantified by immunoblotting. (A-D) $n=27$, AD patients; $n=23$, healthy subjects; (E) $n=34$, AD patients; $n=25$, healthy subjects. (F) Human CBMCs were pretreated with 10 ng/mL IL-4 for 24 hr and stimulated with sera (5% or 10%) from normal subjects ($n=2$) or AD patients ($n=8$) for 24 hr. Supernatants were collected and IL-8 was quantified by ELISA.

healthy individuals had detectable levels of anti-HRF IgE. However, we found no increase in anti-LPS IgE titers ($OD_{450} < 0.01$) in AD or healthy individuals, ruling out the possibility that the measured high O.D. values reflected antigen-non-specific bindings due to high concentrations of total IgE.

We previously showed that polyclonal IgE in AD sera can induce IL-8 production, a signature of HC IgEs, in human CBMCs.²⁰ Therefore, we tested this ability of AD sera ($n=8$) with high titers (≥ 0.01 OD_{450}) of dsDNA- and ssDNA-reactive IgEs, compared to sera from healthy subjects. As shown in Fig. 4F, the AD sera exhibited 4.7- and 7.2-fold higher IL-8-producing activity at 5% and 10% serum concentrations. Therefore, these results strongly suggest that AD patients have autoantigen-reactive IgEs and these IgEs may be HC IgEs.

DISCUSSION

This study demonstrates that most of mouse HC IgEs react with multiple autoantigens. Since some of the autoantigens, e.g., HRF and dsDNA and ssDNA, have multivalency in terms of the IgE-binding ability, it seems reasonable to assume that such an autoantigen can trigger cross-linking of IgE-bound receptors,

thus activation of mast cells and basophils, eventually leading to allergic reactions. Indeed, HRF was shown to amplify allergic inflammation by this mechanism in passive cutaneous anaphylaxis and airway inflammation in mice.²⁴ Similar to HRF, dsDNA and ssDNA released from damaged tissues during allergic reactions may contribute to exacerbate allergic inflammation by interacting with HC IgE bound to Fc ϵ RI on mast cells and basophils. One of the eight mouse IgE mAbs tested failed to react with autoantigens. This negative result might be due to the fact that the number of autoantigens tested was not exhaustive enough or appropriate autoantigens were not tested.

As encountered by western blotting or other immunoassays on a daily basis, immunologists have long known that an antibody recognizes more than one antigen.³⁵ Particularly, monoclonal antibodies were shown to have multispecificity.³⁶ Conformational diversity of antigen recognition, whereby one antibody (e.g., IgE) sequence adopts multiple structures, can increase the effective size of the antibody repertoire and may lead to autoimmunity.²⁸ Autoimmunity underlies allergic diseases.³⁴ Epidemiological data indicate that the prevalence of allergies and autoimmune diseases have increased in parallel.^{37,38} Recent studies support the concept that IgE autoreactivity may

play a pathogenic role in severe and chronic forms of atopy.^{34,39} While acute allergic reactions following exposure to exogenous allergens are understood as immediate type inflammation triggered by degranulation of mast cells via allergen-IgE-FcεRI interactions, chronic allergic inflammation with Th1 characteristics can occur and persist in the absence of exogenous allergens. IgE-mediated presentation of autoallergens may activate auto-reactive Th1 cells to release proinflammatory cytokines.^{40,41} Several environmental allergens have striking structural and immunologic similarities with human proteins.⁴²⁻⁴⁶ A broad spectrum (≥ 140 proteins) of IgE-reactive autoantigens were found in AD patients and some of them were shown to have the ability to activate basophils.⁴⁷ As shown in this study, some AD patients who had high levels of ssDNA- (or dsDNA-) and/or β -galactosidase-reactive IgEs also had high levels of HRF-reactive IgEs, the autoimmune mechanism could be an important part of the disease pathogenesis in this subset of patients. This observation also suggests the heterogeneity of AD pathogenesis, as not all AD patients have increased IgE reactivity to autoantigens. Interestingly, NC/Nga mice spontaneously develop AD-like skin lesions⁴⁸ and IgE autoantibodies including anti-histone H3 IgE;⁴⁹ this age-dependent increase in the serum anti-histone H3 IgE was in parallel with the onset of dermatitis, and correlated well with dermatitis severity at 12-16 weeks of age. As global gene expression patterns are similar in lesional skin between NC/Nga mice and human AD patients (Ando et al., in preparation), these data and our present study support the notion that IgE autoreactivity may play a pathogenic role in AD. Thus, further characterization of the role of IgE autoantibodies in the NC/Nga model will be interesting.

This study also demonstrates that human IgE molecules exhibit heterogeneity in the ability to induce cytokine production and survival promotion in human mast cells in a similar way that mouse HC versus PC IgE molecules exhibit differences in inducing various activation events in mouse mast cells.^{19,50} Thus, a human HC IgE induces cytokine/chemokine production more strongly than do human PC IgEs. However, there are some differences between human and mouse systems. For example, significant survival promotion was seen only with a human HC IgE, in contrast with mouse IgE effects showing that even PC IgEs can promote mast cell survival albeit with weak potency.^{7,10} Degranulation could not be induced by either type of human IgE. These results could be due to the type of human mast cells used in this study. Indeed, even IgE+anti-IgE-stimulated human CBMCs degranulate only 10%-20% of the granule content (data not shown). This low responsiveness may be related to low expression levels of FcεRI in CBMCs, which can be enhanced by IL-4 stimulation.²⁹ Unlike CBMCs pretreated or not with IL-4, human monomeric myeloma IgE induced dose-dependent histamine release, leukotriene C4 production, and IL-8 synthesis in cultured human lung mast cells.²² That study suggests that cultured human lung mast cells are more responsive to IgE than

human CBMCs, although this and our studies were performed using different IgE molecules. Despite the above-mentioned minor differences in monomeric IgE effects between mouse and human, a human HC IgE bound autoantigens such as dsDNA, ssDNA and HRF, like mouse HC IgEs.

In conclusion, HC, but not PC, IgEs exhibit reactivity to various autoantigens. Some of the autoantigens are multivalent, thus potentially capable of activating HC IgE-bound mast cells and basophils. Therefore, these results indicate an autoimmune component in the pathogenic mechanisms of allergic diseases.

ACKNOWLEDGMENTS

The authors thank Drs. Shane Crotty and Rachel Soloff for providing normal sera. This study was funded in part by an NIH grant R01 AI050209 (T.K.). RPS was supported by the Intramural Research Program of the National Institutes of Health, NIDCR. This is Publication No. 1465 from the La Jolla Institute for Allergy and Immunology.

REFERENCES

- Galli SJ. Mast cells and basophils. *Curr Opin Hematol* 2000;7:32-9.
- Galli SJ, Maurer M, Lantz CS. Mast cells as sentinels of innate immunity. *Curr Opin Immunol* 1999;11:53-9.
- Metzger H. The receptor with high affinity for IgE. *Immunol Rev* 1992;125:37-48.
- Furuichi K, Rivera J, Isersky C. The receptor for immunoglobulin E on rat basophilic leukemia cells: effect of ligand binding on receptor expression. *Proc Natl Acad Sci U S A* 1985;82:1522-5.
- Hsu C, MacGlashan D Jr. IgE antibody up-regulates high affinity IgE binding on murine bone marrow-derived mast cells. *Immunol Lett* 1996;52:129-34.
- Yamaguchi M, Lantz CS, Oettgen HC, Katona IM, Fleming T, Miyajima I, Kinet JP, Galli SJ. IgE enhances mouse mast cell FcεRI expression in vitro and in vivo: evidence for a novel amplification mechanism in IgE-dependent reactions. *J Exp Med* 1997;185:663-72.
- Asai K, Kitaura J, Kawakami Y, Yamagata N, Tsai M, Carbone DP, Liu FT, Galli SJ, Kawakami T. Regulation of mast cell survival by IgE. *Immunity* 2001;14:791-800.
- Kalesnikoff J, Huber M, Lam V, Damen JE, Zhang J, Siraganian RP, Krystal G. Monomeric IgE stimulates signaling pathways in mast cells that lead to cytokine production and cell survival. *Immunity* 2001;14:801-11.
- Tanaka S, Takasu Y, Mikura S, Satoh N, Ichikawa A. Antigen-independent induction of histamine synthesis by immunoglobulin E in mouse bone marrow-derived mast cells. *J Exp Med* 2002;196:229-35.
- Kitaura J, Song J, Tsai M, Asai K, Maeda-Yamamoto M, Mocsai A, Kawakami Y, Liu FT, Lowell CA, Barisac BG, Galli SJ, Kawakami T. Evidence that IgE molecules mediate a spectrum of effects on mast cell survival and activation via aggregation of the FcεRI. *Proc Natl Acad Sci U S A* 2003;100:12911-6.
- Yamada N, Matsushima H, Tagaya Y, Shimada S, Katz SI. Generation of a large number of connective tissue type mast cells by culture of murine fetal skin cells. *J Invest Dermatol* 2003;121:1425-32.

12. Oka T, Hori M, Tanaka A, Matsuda H, Karaki H, Ozaki H. IgE alone-induced actin assembly modifies calcium signaling and degranulation in RBL-2H3 mast cells. *Am J Physiol Cell Physiol* 2004;286:C256-63.
13. Pandey V, Mihara S, Fensome-Green A, Bolsover S, Cockcroft S. Monomeric IgE stimulates NFAT translocation into the nucleus, a rise in cytosol Ca²⁺, degranulation, and membrane ruffling in the cultured rat basophilic leukemia-2H3 mast cell line. *J Immunol* 2004;172:4048-58.
14. Lam V, Kalesnikoff J, Lee CW, Hernandez-Hansen V, Wilson BS, Oliver JM, Krystal G. IgE alone stimulates mast cell adhesion to fibronectin via pathways similar to those used by IgE + antigen but distinct from those used by Steel factor. *Blood* 2003;102:1405-13.
15. Kitaura J, Eto K, Kinoshita T, Kawakami Y, Leitges M, Lowell CA, Kawakami T. Regulation of highly cytokinergic IgE-induced mast cell adhesion by Src, Syk, Tec, and protein kinase C family kinases. *J Immunol* 2005;174:4495-504.
16. Matsuda K, Piliponsky AM, Iikura M, Nakae S, Wang EW, Dutta SM, Kawakami T, Tsai M, Galli SJ. Monomeric IgE enhances human mast cell chemokine production: IL-4 augments and dexamethasone suppresses the response. *J Allergy Clin Immunol* 2005;116:1357-63.
17. Beaven MA, Metzger H. Signal transduction by Fc receptors: the FcεRI case. *Immunol Today* 1993;14:222-6.
18. Macglashan D Jr. IgE and Fc{epsilon}RI regulation. *Ann N Y Acad Sci* 2005;1050:73-88.
19. Kawakami T, Kitaura J. Mast cell survival and activation by IgE in the absence of antigen: a consideration of the biologic mechanisms and relevance. *J Immunol* 2005;175:4167-73.
20. Kashiwakura J, Kawakami Y, Yuki K, Zajonc DM, Hasegawa S, Tomimori Y, Caplan B, Saito H, Furue M, Oettgen HC, Okayama Y, Kawakami T. Polyclonal IgE induces mast cell survival and cytokine production. *Allergol Int* 2009;58:411-9.
21. Gilchrist H, Cheewatrakoolpong B, Billah M, Egan RW, Anthes JC, Greenfeder S. Human cord blood-derived mast cells synthesize and release I-309 in response to IgE. *Life Sci* 2003;73:2571-81.
22. Cruse G, Kaur D, Yang W, Duffy SM, Brightling CE, Bradding P. Activation of human lung mast cells by monomeric immunoglobulin E. *Eur Respir J* 2005;25:858-63.
23. MacDonald SM, Lichtenstein LM, Proud D, Plaut M, Naclerio RM, MacGlashan DW, Kagey-Sobotka A. Studies of IgE-dependent histamine releasing factors: heterogeneity of IgE. *J Immunol* 1987;139:506-12.
24. Kashiwakura JC, Ando T, Matsumoto K, Kimura M, Kitaura J, Matho MH, Zajonc DM, Ozeki T, Ra C, MacDonald SM, Siraganian RP, Broide DH, Kawakami Y, Kawakami T. Histamine-releasing factor has a proinflammatory role in mouse models of asthma and allergy. *J Clin Invest* 2012;122:218-28.
25. Ishizaka K. Immunoglobulin E (IgE). *Methods Enzymol* 1985;116:76-94.
26. Hanifin JM, Rajka G. Diagnostic features of atopic dermatitis. *Acta Derm Venereol Suppl (Stockh)* 1980;59:44-7.
27. Iida M, Matsumoto K, Tomita H, Nakajima T, Akasawa A, Ohtani NY, Yoshida NL, Matsui K, Nakada A, Sugita Y, Shimizu Y, Wakahara S, Nakao T, Fujii Y, Ra C, Saito H. Selective down-regulation of high-affinity IgE receptor (FcεRI) alpha-chain messenger RNA among transcriptome in cord blood-derived versus adult peripheral blood-derived cultured human mast cells. *Blood* 2001;97:1016-22.
28. James LC, Roversi P, Tawfik DS. Antibody multispecificity mediated by conformational diversity. *Science* 2003;299:1362-7.
29. Toru H, Ra C, Nonoyama S, Suzuki K, Yata J, Nakahata T. Induction of the high-affinity IgE receptor (FcεRI) on human mast cells by IL-4. *Int Immunol* 1996;8:1367-73.
30. Lane JE, Cheyney JM, Lane TN, Kent DE, Cohen DJ. Treatment of recalcitrant atopic dermatitis with omalizumab. *J Am Acad Dermatol* 2006;54:68-72.
31. Park SY, Choi MR, Na JI, Youn SW, Park KC, Huh CH. Recalcitrant atopic dermatitis treated with omalizumab. *Ann Dermatol* 2010;22:349-52.
32. Sheinkopf LE, Rafi AW, Do LT, Katz RM, Klaustermeyer WB. Efficacy of omalizumab in the treatment of atopic dermatitis: a pilot study. *Allergy Asthma Proc* 2008;29:530-7.
33. Strunk RC, Bloomberg GR. Omalizumab for asthma. *N Engl J Med* 2006;354:2689-95.
34. Valenta R, Mittermann I, Werfel T, Garn H, Renz H. Linking allergy to autoimmune disease. *Trends Immunol* 2009;30:109-16.
35. Talmage DW. Immunological specificity, unique combinations of selected natural globulins provide an alternative to the classical concept. *Science* 1959;129:1643-8.
36. Varga JM, Konigsberg WH, Richards FF. Antibodies with multiple binding functions. Induction of single immunoglobulin species by structurally dissimilar haptens. *Proc Natl Acad Sci U S A* 1973;70:3269-74.
37. Black P. Why is the prevalence of allergy and autoimmunity increasing? *Trends Immunol* 2001;22:354-5.
38. Rabin RL, Levinson AI. The nexus between atopic disease and autoimmunity: a review of the epidemiological and mechanistic literature. *Clin Exp Immunol* 2008;153:19-30.
39. Valenta R, Seiberler S, Natter S, Mahler V, Mossabeh R, Ring J, Stingl G. Autoallergy: a pathogenetic factor in atopic dermatitis? *J Allergy Clin Immunol* 2000;105:432-7.
40. Aichberger KJ, Mittermann I, Reininger R, Seiberler S, Swoboda I, Spitzauer S, Kopp T, Stingl G, Sperr WR, Valent P, Repa A, Bohle B, Kraft D, Valenta R. Hom s 4, an IgE-reactive autoantigen belonging to a new subfamily of calcium-binding proteins, can induce Th cell type 1-mediated autoreactivity. *J Immunol* 2005;175:1286-94.
41. Mittermann I, Reininger R, Zimmermann M, Gangl K, Reisinger J, Aichberger KJ, Greisenegger EK, Niederberger V, Seipelt J, Bohle B, Kopp T, Akdis CA, Spitzauer S, Valent P, Valenta R. The IgE-reactive autoantigen Hom s 2 induces damage of respiratory epithelial cells and keratinocytes via induction of IFN-γ. *J Invest Dermatol* 2008;128:1451-9.
42. Cramer R, Faith A, Hemmann S, Jaussi R, Ismail C, Menz G, Blaser K. Humoral and cell-mediated autoimmunity in allergy to *Aspergillus fumigatus*. *J Exp Med* 1996;184:265-70.
43. Mayer C, Appenzeller U, Seelbach H, Achatz G, Oberkofler H, Breitenbach M, Blaser K, Cramer R. Humoral and cell-mediated autoimmune reactions to human acidic ribosomal P2 protein in individuals sensitized to *Aspergillus fumigatus* P2 protein. *J Exp Med* 1999;189:1507-12.
44. Schmid-Grendelmeier P, Flückiger S, Disch R, Trautmann A, Wüthrich B, Blaser K, Scheynius A, Cramer R. IgE-mediated and T cell-mediated autoimmunity against manganese superoxide dismutase in atopic dermatitis. *J Allergy Clin Immunol* 2005;115:1068-75.
45. Spitzauer S, Schweiger C, Sperr WR, Pandjaitan B, Valent P, Mühl S, Ebner C, Scheiner O, Kraft D, Rumpold H, Valenta R. Molecular characterization of dog albumin as a cross-reactive allergen. *J Allergy Clin Immunol* 1994;93:614-27.

46. Valenta R, Duchêne M, Pettenburger K, Sillaber C, Valent P, Bettelheim P, Breitenbach M, Rumpold H, Kraft D, Scheiner O. Identification of profilin as a novel pollen allergen; IgE autoreactivity in sensitized individuals. *Science* 1991;253:557-60.
47. Zeller S, Rhyner C, Meyer N, Schmid-Grendelmeier P, Akdis CA, Cramer R. Exploring the repertoire of IgE-binding self-antigens associated with atopic eczema. *J Allergy Clin Immunol* 2009;124:278-85, 285.e1-7.
48. Matsuda H, Watanabe N, Geba GP, Sperl J, Tsudzuki M, Hiroi J, Matsumoto M, Ushio H, Saito S, Askenase PW, Ra C. Development of atopic dermatitis-like skin lesion with IgE hyperproduction in NC/Nga mice. *Int Immunol* 1997;9:461-6.
49. Matsubara T, Aoki N, Hino S, Okajima T, Nadano D, Matsuda T. Serum and monoclonal immunoglobulin E antibodies from NC/Nga mice with severe atopic-like dermatitis recognize an auto-antigen, histone H3. *Clin Exp Allergy* 2009;39:579-90.
50. Kashiwakura J, Otani IM, Kawakami T. Monomeric IgE and mast cell development, survival and function. *Adv Exp Med Biol* 2011;716:29-46.