## SCISPACE <br> formerly Typeset

〇 Open access • Posted Content • DOI:10.1101/682633

## Multi-tissue probabilistic fine-mapping of transcriptome-wide association study identifies cis-regulated genes for miserableness - Source link $\square$

Calwing Liao, Calwing Liao, Vuokila, Alexandre D. Laporte ...+5 more authors
Institutions: Montreal Neurological Institute and Hospital, McGill University
Published on: 26 Jun 2019 - bioRxiv (Cold Spring Harbor Laboratory)
Topics: Genome-wide association study

Related papers:

- A powerful fine-mapping method for transcriptome-wide association studies
- Transcriptome-wide association studies: opportunities and challenges
- A tissue-specific collaborative mixed model for jointly analyzing multiple tissues in transcriptome-wide association studies.
- Inferring Relevant Cell Types for Complex Traits by Using Single-Cell Gene Expression.
- The anatomical distribution of genetic associations

Share this paper: 9 in $\square$
View more about this paper here: https://typeset.io/papers/multi-tissue-probabilistic-fine-mapping-of-transcriptome14dtz9ucln

# Multi-tissue probabilistic fine-mapping of transcriptome-wide association study identifies cis-regulated genes for miserableness 

Calwing Liao ${ }^{1,2}$ BSc, Veikko Vuokila ${ }^{2}$, Alexandre D Laporte ${ }^{2}$ BSc, Dan Spiegelman ${ }^{2}$ MSc, Patrick A. Dion ${ }^{2,3}$ PhD, Guy A. Rouleau ${ }^{1,2,3^{*}}$ MD, PhD<br>${ }^{1}$ Department of Human Genetics, McGill University, Montréal, Quebec, Canada<br>${ }^{2}$ Montreal Neurological Institute, McGill University, Montréal, Quebec, Canada<br>${ }^{3}$ Department of Neurology and Neurosurgery, McGill University, Montréal, Quebec, Canada

Short summary: The first transcriptome-wide association study of miserableness identifies many genes including c7orf50 implicated in the trait.

Word count: 1,522 excluding abstract and references
Tables: 3

Keywords: Miserableness, transcriptome-wide association study, TWAS

*Correspondence: | Dr. Guy A. Rouleau |  |
| :--- | :--- |
|  | Montreal Neurological Institute and Hospital |
|  | Department of Neurology and Neurosurgery |
|  | 3801 University Street, Montreal, QC |
|  | Canada H3A 2B4. |
|  | Tel: +1 514 3982690 |
|  | Fax: +1 514 3988248 |
|  | E-mail: guy.rouleau@mcgill.ca |

## Abstract (141 words)

Miserableness is a behavioural trait that is characterized by strong negative feelings in an individual. Although environmental factors tend to invoke miserableness, it is common to feel miserable 'for no reason', suggesting an innate, potential genetic component. Currently, little is known about the functional relevance of common variants associated with miserableness. To further characterize the trait, we conducted a transcriptome-wide association study (TWAS) on 373,733 individuals and identified 104 signals across brain tissue panels with 37 unique genes. Subsequent probabilistic fine-mapping prioritized 95 genes into 90\%-credible sets. Amongst these prioritized hits, C7orf50 had the highest posterior inclusion probability of 0.869 in the brain cortex. Furthermore, we demonstrate that many GWAS hits for miserableness are driven by expression. To conclude, we successfully identified several genes implicated in miserableness and highlighted the power of TWAS to prioritize genes associated with a trait.

## Introduction

Miserableness is characterized by emotional distress, typically caused by an event invoking negative feelings in an individual ${ }^{1}$. Although environmental factors may cause feelings of miserableness, it is common for certain individuals to feel miserable independent of environmental influences, suggesting that underlying biologic and genetic factors might play a role in miserableness. Several large-scale genome-wide association studies (GWAS) have focused on characterizing the genetics of neuroticism items such as miserableness ${ }^{2,3}$. No genetic factors fully overlap both miserableness and other neuroticism traits, which suggests that there are variants and/or genes that are specific to miserableness ${ }^{4}$. For instance, previous interrogation of neuroticism items found that miserableness had the highest genetic correlation with feeling "fedup" $(0.88)$ and "experiencing mood swings" $(0.86)$, suggesting that there are genetic differences ${ }^{4}$. However, identifying the biological functionality of these variants remains difficult.

Transcriptomic imputation has recently been used to integrate genotype and expression data to identify predictive cis-eQTLs, which in turn can be applied to independent datasets in a tissuespecific manner ${ }^{5}$. Transcriptome-wide association studies (TWAS) can identify cis-eQTL regulated
genes by integrating expression panels from large consortia such as GTEx and the CommonMind Consortium (CMC) with GWAS data ${ }^{6}$. Furthermore, probabilistic fine-mapping of these TWAS hits can be done by modelling correlation among TWAS hits and assigning a probability for each gene within the risk region to prioritize genes ${ }^{7}$.

To understand cis-eQTL regulated candidate genes in miserableness, we conducted a TWAS consisting of 373,733 individuals using the summary statistics from a recent study dissecting the genetic heterogeneity of neuroticism ${ }^{4}$. The participants were asked whether they tend to 'feel miserable' for no particular reason. A total of 104 TWAS signals with 37 unique genes were transcriptome-wide significant across brain tissue panels. Probabilistic fine-mapping identified a number of putatively causal genes, including C7orf50 in GPX1 in the frontal cortex with a posterior inclusion probability (PIP) of 0.869 and MTCH2 in the nucleus accumbens with a PIP of 0.573. Conditioning on the transcriptome-wide significant hits showed that the hits accounted for $>75 \%$ of the GWAS signal present. To conclude, miserableness genetics is explained partially by cis-eQTLs driving the expression of certain genes. Additional studies are needed to characterize other sources of non-coding variation such as $3^{\prime}$-aQTLs.

## Results

## Transcriptome-wide association study identifies 104 signals associated with miserableness

A multi-tissue TWAS using FUSION identified 104 TWAS signals associated with miserableness after Bonferroni-correction (Table 1, Figure 1). Across all signals, there were 37 genes identified in at least one imputation panel (Supplementary table 1). Amongst all signals, the top five hits in chronological order consisted of GPX1 ( $\mathrm{P}_{\text {Bonferroni }}=5.18 \times 10^{-10}$ ) in the frontal cortex, RNF123 ( Panferroni $=3.67 \times 10^{-8}$ ) in the cerebellum, GPX1 ( Pbonferroni $=3.09 \times 10^{-7}$ ) in the cerebellum, RBM6 ( $\mathrm{P}=5.12 \times 10^{-7}$ ) in the caudate basal ganglia, and MST1R ( $\mathrm{P}_{\text {Bonferroni }}=1.29 \times 10^{-6}$ ) in the prefrontal cortex. Amongst the signals, several implicated RNA-coding genes, reinforcing the notion that non-coding genes are relevant to complex traits (Table 1).

## Top GWAS signals are largely explained by expression

To determine how much GWAS signal remains after the expression association from TWAS is removed, conditional testing was done for transcriptome-wide significant TWAS signals. For most of the GWAS signals, the expression accounted for $>75 \%$ of the variance (Table 2, Supplementary Figures 1-14). For RP11-74E22.6, RP11-798G7.5, and DNF-AS, conditioning on the expression accounted for $100 \%$ of the signal. There were less TWAS signals compared to the GWAS significant signals for miserableness, suggesting that other genetic mechanisms are driving those signals.

## Fine-mapping of TWAS association causally implicates several genes with miserableness

To identify causal genes, FOCUS was used to assign a posterior inclusion probability for genes at each TWAS region and for relevant tissue types. Across all panels, there were 95 hits included in the $90 \%$ credible gene set (Table 3). The gene, C7orf50, had the highest posterior inclusion probability (PIP) of 0.869 in the brain cortex. Additionally, the top multi-tissue TWAS hit from the FUSION results, GPX1, was included in the credible sets with a PIP of 0.192 in the frontal cortex. Several other genes also had high PIPs, such as RP11-127L20.3 in the hippocampus with a PIP of 0.304, MTCH2 in the nucleus accumbens with a PIP of 0.573 , ORC4 in the hypothalamus with a PIP of 0.524, ATAD2B in the cerebellar hemisphere with a PIP of 0.264, FANCL in the dorsolateral prefrontal cortex with a PIP of 0.446, and CTC-467M3.3 in the frontal cortex with a PIP of 0.567.

## Discussion

With the influx of several large-scale biobanks and GWAS studies, many loci are being identified. The next step is to identify biologically- and phenotypically- relevant genes found by GWAS. To date, few studies have attempted to understand the genetics of miserableness, despite the high prevalence of the trait. Here, we conducted the largest TWAS to date using the summary statistics of 373,733 individuals to further understand this neuroticism item. A total of 104 TWAS signals were transcriptome-wide significant across brain tissues with 37 unique genes.

We identified a total of 104 transcriptome-wide signals across brain tissues with 37 unique genes. The top two signals included GPX1 frontal cortex and RNF123 in the cerebellum. The gene, GPX1, encodes for a cytosolic enzyme, glutathione peroxidase-1, expressed in many different tissues ${ }^{8}$.

This gene has previously been implicated in Alzheimer's disease affecting cortical neurons ${ }^{9}$. Furthermore, neurons lacking GPX1 leads have a greater susceptibility to oxidative-driven cell death ${ }^{8}$. The Z-scores for all GPX1 hits were positive, suggesting that increased expression leads to susceptibility to miserableness. Currently, little is known about the effects of increased GPX1 in the nervous system. The RNF123 gene, encodes for a ubiquitin-protein ligase, and expression has been correlated with depressive disorder, which likely has genetic overlap with miserableness ${ }^{10,11}$.

Often, an implicated GWAS locus contains many genes. Common GWAS mapping techniques would assign the SNP association to the closest gene, however, this has been shown to be suboptimal ${ }^{12-14}$. After conditioning on the TWAS signal for each transcriptome-wide significant hit, most of the signal was explained by the expression of the conditioned gene.

Fine-mapping of the TWAS signals identified many genes included in the credible set, where C7orf50 had the highest PIP of 0.869 in the brain cortex. Currently, the literature is sparse on C7orf50, however, querying the gene using the tissue-specific gene network (GIANT), showed potential implications in autism spectrum disorder and epilepsy ${ }^{15}$. For the brain cortex, GIANT implicated CCDC85B as the top functionally related gene to C7orf50. Previous studies have shown that CCDC85B is implicated in neural tube development ${ }^{16}$.

We conclude this study with some caveats and potential follow-up ideas. First, TWAS only measures the effects of cis-eQTLs and may not capture other genetic regulatory effects that contribute towards miserableness. Second, future large studies with wide ranges of phenotypes such as the All of Us initiative will allow to successfully measure the genetic susceptibility to miserableness in other ethnic populations. Here, we successfully demonstrated that behavioural traits such as miserableness have a strong genetic basis and many signals are driven by cis-eQTL. Genes such as GPX1, RNF123, C7orf50 and MTCH2 should be further investigated to understand the molecular consequences of dysregulated expression.

## Methods

Genotype data and patient information

Summary statistics were obtained from Nagel et al. (2018) ${ }^{4}$. Details pertaining to participant ascertainment and quality control were previously reported by Nagel et al. (2018) ${ }^{4}$. Succinctly, the data was derived from the UK BioBank and miserableness was determined by asking "Do you ever feel 'just miserable' for no reason?". A total of 373,733 individuals were included, with $45 \%$ of them saying "yes". There were $47 \%$ of females who responded "yes", and $43 \%$ of males who responded "yes".

## Transcriptomic imputation

Transcriptomic imputation (TI) was done using eQTL panels created from tissue-specific gene expression coupled with genotypic data ${ }^{17}$. Here, we used all the brain tissue types from GTEx 53 v7 and the CommonMind Consortium (CMC) ${ }^{6}$. A strict Bonferroni-corrected study-wise threshold was used: $\mathrm{P}=4.97 \mathrm{E}-07(0.05 / 100,572)$ (total number of genes across panels). FUSION was used to conduct the transcriptome-wide association testing ${ }^{17}$. The 1000 Genomes v3 LD panel was used for the TWAS. FUSION utilizes several penalized linear models such as GBLUP, LASSO, Elastic Net ${ }^{17}$. Furthermore, a Bayesian sparse linear mixed model (BSLMM) is used. FUSION works by computing an out-sample $R^{2}$ to determine the best model by performing a fivefold crossvalidating of every model. Further details can be found in the original manuscript.

## Conditionally testing GWAS signals

To determine how much GWAS signal remains after the expression association from TWAS is removed, joint and conditional testing was done for genome-wide Bonferroni-corrected TWAS signals. The joint and conditional analyses help to determine genes with independent genetic predictors associated with miserableness from genes that are simply co-expressed with a genetic predictor. Each miserableness GWAS SNP association was conditioned on the joint gene model one SNP at a time.

## Fine-mapping of TWAS associations

To address the issue of co-regulation in TWAS, we used the program FOCUS (Fine-mapping of causal gene sets) to directly model predicted expression correlations and to provide a posterior
causal probability for genes in relevant tissue types ${ }^{7}$. FOCUS identifies genes for each TWAS signal to be part of a $90 \%$-credible set while simultaneously controlling for pleiotropic effects of SNPs. Furthermore, the same TWAS reference panels for FUSION were used.

## References

1. Vandercammen, L., Hofmans, J. \& Theuns, P. Relating Specific Emotions to Intrinsic Motivation: On the Moderating Role of Positive and Negative Emotion Differentiation. PLoS One 9, e115396 (2014).
2. Luciano, M. et al. Association analysis in over 329,000 individuals identifies 116 independent variants influencing neuroticism. Nat. Genet. 50, 6-11 (2018).
3. Nagel, M. et al. Meta-analysis of genome-wide association studies for neuroticism in 449,484 individuals identifies novel genetic loci and pathways. Nat. Genet. 50, 920-927 (2018).
4. Nagel, M., Watanabe, K., Stringer, S., Posthuma, D. \& van der Sluis, S. Item-level analyses reveal genetic heterogeneity in neuroticism. Nat. Commun. 9, 905 (2018).
5. Gusev, A. et al. Integrative approaches for large-scale transcriptome-wide association studies. Nat. Genet. 48, 245-252 (2016).
6. Consortium, Gte. Genetic effects on gene expression across human tissues. Nature 550, 204-213 (2017).
7. Mancuso, N. et al. Probabilistic fine-mapping of transcriptome-wide association studies. Nat. Genet. 51, 675-682 (2019).
8. de Haan, J. B. et al. Mice with a homozygous null mutation for the most abundant glutathione peroxidase, Gpx1, show increased susceptibility to the oxidative stressinducing agents paraquat and hydrogen peroxide. J. Biol. Chem. 273, 22528-36 (1998).
9. Crack, P. J., Cimdins, K., Ali, U., Hertzog, P. J. \& lannello, R. C. Lack of glutathione peroxidase-1 exacerbates $A \beta$-mediated neurotoxicity in cortical neurons. J. Neural Transm. 113, 645-657 (2006).
10. Teyssier, J.-R., Rey, R., Ragot, S., Chauvet-Gelinier, J.-C. \& Bonin, B. Correlative gene expression pattern linking RNF123 to cellular stress-senescence genes in patients with depressive disorder: Implication of DRD1 in the cerebral cortex. J. Affect. Disord. 151, 432-438 (2013).
11. Chaturvedi, P., Khanna, R. \& Parnaik, V. K. Ubiquitin ligase RNF123 Mediates Degradation of Heterochromatin Protein $1 \alpha$ and $\beta$ in Lamin A/C Knock-Down Cells. PLoS One 7, e47558 (2012).
12. Liao, C. et al. Transcriptome-wide association study of attention deficit hyperactivity disorder identifies associated genes and phenotypes. bioRxiv 642231 (2019). doi:10.1101/642231
13. Mancuso, N. et al. Large-scale transcriptome-wide association study identifies new prostate cancer risk regions. Nat. Commun. 9, 4079 (2018).
14. Gusev, A. et al. Transcriptome-wide association study of schizophrenia and chromatin activity yields mechanistic disease insights. Nat. Genet. 50, 538-548 (2018).
15. Greene, C. S. et al. Understanding multicellular function and disease with human tissue-
specific networks. Nat. Genet. 47, 569-576 (2015).
16. Markham, N. O. et al. DIPA-family coiled-coils bind conserved isoform-specific head domain of p120-catenin family: potential roles in hydrocephalus and heterotopia. Mol. Biol. Cell 25, 2592-603 (2014).
17. Gusev, A. et al. Integrative approaches for large-scale transcriptome-wide association studies. Nat. Genet. 48, 245-252 (2016).

## Acknowledgements

This work was supported by a Canadian Institutes of Health Research Foundation Scheme grant (\#332971). G.A.R. holds a Canada Research Chair in Genetics of the Nervous System and the Wilder Penfield Chair in Neurosciences. C.L. is a recipient of the Frederick Banting and Charles Best Canada Graduate Scholarship from the Canadian Institutes of Health Research (CIHR). C.L. conducted the experiments, analyses and drafted the manuscript. V.V. helped with analyses. A.D.L, D.S. helped with bioinformatics. P.A.D. and G.A.R. oversaw the analyses and helped draft the manuscript.

## Conflicts of Interest

We report no conflicts of interest.

Table 1. Multi-tissue significant TWAS hits for miserableness.

| Gene | Tissue | eQTL | Z-score | Uncorrected <br> P-value | Bonferroni P- <br> value |
| :--- | :--- | :--- | :--- | :--- | :--- |
| AF131215.2 | ROSMAP <br> Brain | rs17724226 | -5.57 | $2.51 \mathrm{E}-08$ | $3.92 \mathrm{E}-03$ |
|  | Cerebellar <br> Hemisphere | rs17724226 | -5.93 | $3.11 \mathrm{E}-09$ | $4.86 \mathrm{E}-04$ |
| AF131215.3 | ROSMAP <br> Brain | rs2409745 | -6.13 | $8.79 \mathrm{E}-10$ | $1.37 \mathrm{E}-04$ |
|  | Brain Cortex | $\mathrm{rs3448}$ | -3.49 | $2.05 \mathrm{E}-07$ | $3.20 \mathrm{E}-02$ |
|  | Caudate Basal <br> Ganglia | rs9834003 | -4.89 | $7.18 \mathrm{E}-08$ | $1.12 \mathrm{E}-02$ |
|  | ROSMAP <br> Brain | rs7619016 | -11.57 | $6.45 \mathrm{E}-08$ | $1.01 \mathrm{E}-02$ |
|  | Hippocampus | rs 17689471 | -4.07 | $1.89 \mathrm{E}-08$ | $2.95 \mathrm{E}-03$ |
| ARHGAP27 | Nucleus <br> Accumbens <br> Basal Ganglia | rs 17689471 | 5.94 | $2.91 \mathrm{E}-09$ | $4.54 \mathrm{E}-04$ |
|  | Brain Cortex | rs12286721 | 6.41 | $1.41 \mathrm{E}-10$ | $2.20 \mathrm{E}-05$ |
| CRHR1-IT1 | Brain Cortex | $\mathrm{rs17689471}$ | 5.17 | $2.37 \mathrm{E}-07$ | $3.70 \mathrm{E}-02$ |


|  | Nucleus Accumbens Basal Ganglia | rs8072451 | 5.74 | $9.67 \mathrm{E}-09$ | $1.51 \mathrm{E}-03$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  | Caudata Basal Ganglia | rs17763086 | 5.88 | 4.10E-09 | 6.40E-04 |
|  | Hypothalamus | rs17763086 | 5.99 | $2.06 \mathrm{E}-09$ | $3.22 \mathrm{E}-04$ |
|  | Frontal Cortex BA9 | rs17763086 | 6.02 | $1.71 \mathrm{E}-09$ | $2.67 \mathrm{E}-04$ |
|  | Putamen basal ganglia | rs17689471 | 6.05 | 1.43E-09 | 2.23E-04 |
|  | Hippocampus | rs17689471 | 6.43 | $1.31 \mathrm{E}-10$ | $2.05 \mathrm{E}-05$ |
| ERI1 | Dorsolateral prefrontal cortex | rs7826478 | 5.95 | $2.74 \mathrm{E}-09$ | $4.28 \mathrm{E}-04$ |
| FAM167A | Dorsolateral prefrontal cortex | rs7844834 | 5.44 | 5.47E-08 | 8.54E-03 |
|  | ROSMAP <br> Brain | rs10102600 | 5.93 | 3.08E-09 | $4.81 \mathrm{E}-04$ |
| FAM212A | Cerebellum | rs7372966 | 5.860692 | 4.61E-09 | $7.20 \mathrm{E}-04$ |
| FAM66A | ROSMAP <br> Brain | rs12676145 | -5.39 | 7.16E-08 | $1.12 \mathrm{E}-02$ |
| FAM66D | ROSMAP <br> Brain | rs12550733 | 5.21 | 1.90E-07 | $2.97 \mathrm{E}-02$ |
| FAM85B | ROSMAP <br> Brain | rs2945251 | 5.64 | $1.72 \mathrm{E}-08$ | $2.69 \mathrm{E}-03$ |
| FAM86B1 | Dorsolateral prefrontal cortex | rs11998678 | -5.46 | 4.90E-08 | 7.65E-03 |
| FAM86B3P | Frontal Cortex BA9 | rs2948286 | 5.5 | 3.69E-08 | 5.76E-03 |
|  | ROSMAP <br> brain | rs2945230 | 5.65 | $1.57 \mathrm{E}-08$ | $2.45 \mathrm{E}-03$ |
|  | Cerebellar hemisphere | rs876954 | 6.25 | 4.14E-10 | 6.47E-05 |
|  | Cerebellum | rs2980436 | 6.5 | 7.99E-11 | $1.25 \mathrm{E}-05$ |
|  | Brain Cortex | rs2945230 | 6.5 | $7.91 \mathrm{E}-11$ | $1.24 \mathrm{E}-05$ |
| FNBP4 | ROSMAP <br> Brain | rs7929014 | -6.23 | 4.77E-10 | 7.45E-05 |
| FTSJ2 | Brain Cortex | rs3757440 | 5.805551 | $6.42 \mathrm{E}-09$ | $1.00 \mathrm{E}-03$ |
| GPX1 | Caudate Basal Ganglia | rs11130186 | 5.680812 | $1.34 \mathrm{E}-08$ | $2.09 \mathrm{E}-03$ |



|  | Hypothalamus | rs199439 | 5.81 | 6.14E-09 | 9.59E-04 |
| :---: | :---: | :---: | :---: | :---: | :---: |
| LRRC37A4P | Brain cortex | rs17689918 | -5.38 | 7.49E-08 | $1.17 \mathrm{E}-02$ |
|  | Caudate basal ganglia | rs8072451 | -5.86 | $4.68 \mathrm{E}-09$ | 7.31E-04 |
|  | Putamen basal ganglia | rs8072451 | -5.86 | 4.68E-09 | 7.31E-04 |
|  | Hippocampus | rs8072451 | -5.86 | 4.56E-09 | $7.12 \mathrm{E}-04$ |
|  | Nucleus accumbens basal ganglia | rs8072451 | -5.86 | 4.51E-09 | 7.04E-04 |
|  | Cerebellar hemisphere | rs8072451 | -5.87 | 4.45E-09 | 6.95E-04 |
|  | Cerebellum | rs17689471 | -5.87 | 4.44E-09 | 6.93E-04 |
|  | Frontal cortex BA9 | rs17763086 | -5.91 | 3.45E-09 | 5.39E-04 |
|  | Hypothalamus | rs8072451 | -6.01 | $1.83 \mathrm{E}-09$ | $2.86 \mathrm{E}-04$ |
| MAPT | Cerebellar hemisphere | rs17763086 | -5.88 | 4.10E-09 | 6.40E-04 |
|  | Frontal cortex BA9 | rs17689882 | -5.9 | 3.55E-09 | 5.54E-04 |
|  | Dorsolateral prefrontal cortex | rs17689471 | -6.24 | 4.40E-10 | 6.87E-05 |
|  | Brain cortex | rs17689882 | -6.36 | $2.04 \mathrm{E}-10$ | $3.19 \mathrm{E}-05$ |
| MST1R | Dorsolateral prefrontal cortex | rs2526388 | 5.226389 | $1.73 \mathrm{E}-07$ | $2.70 \mathrm{E}-02$ |
|  | Cerebellum | rs2856236 | 6.563222 | 5.27E-11 | 8.23E-06 |
|  | ROSMAP brain | rs11713193 | 6.833558 | 8.28E-12 | 1.29E-06 |
| MTCH2 | Brain cortex | rs7947730 | -5.15 | $2.55 \mathrm{E}-07$ | 3.98E-02 |
|  | ROSMAP brain | rs4752856 | -5.55 | $2.85 \mathrm{E}-08$ | $4.45 \mathrm{E}-03$ |
| MTMR9 | ROSMAP brain | rs2246606 | 5.44 | 5.39E-08 | 8.42E-03 |
| NICN1 | Nucleus accumbens basal ganglia | rs11130186 | 5.260183 | $1.44 \mathrm{E}-07$ | $2.25 \mathrm{E}-02$ |
| PLEKHM1 | Brain Cortex | rs11012 | 5.21 | $1.94 \mathrm{E}-07$ | $3.03 \mathrm{E}-02$ |
|  | Cerebellar hemisphere | rs8072451 | -5.9 | 3.63E-09 | 5.67E-04 |


|  | Cerebellum | rs17689918 | -6.39 | $1.65 \mathrm{E}-10$ | $2.58 \mathrm{E}-05$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
| RBM6 | ROSMAP <br> brain | rs6765484 | -5.375061 | 7.66E-08 | 1.20E-02 |
|  | Dorsolateral prefrontal cortex | rs9311446 | -5.785099 | 7.25E-09 | $1.13 \mathrm{E}-03$ |
|  | Putamen basal ganglia | rs2240329 | -5.843777 | 5.10E-09 | 7.96E-04 |
|  | Frontal cortex BA9 | rs2681780 | 5.852691 | 4.84E-09 | 7.56E-04 |
|  | Cerebellar hemisphere | rs11713193 | -5.870895 | 4.33E-09 | 6.76E-04 |
|  | Nucleus accumbens basal ganglia | rs4688755 | -5.912526 | 3.37E-09 | 5.26E-04 |
|  | Cerebellum | rs6446193 | -6.056184 | 1.39E-09 | 2.17E-04 |
|  | Brain cortex | rs6765484 | -6.185421 | 6.19E-10 | 9.67E-05 |
|  | Caudate basal ganglia | rs2014830 | -6.965004 | $3.28 \mathrm{E}-12$ | 5.12E-07 |
| RNF123 | Dorsolateral prefrontal cortex | rs2352974 | 5.465525 | 4.62E-08 | 7.21E-03 |
|  | Cerebellar hemisphere | rs7634902 | 5.474334 | 4.39E-08 | 6.86E-03 |
|  | Brain cortex | rs7634902 | 5.559422 | 2.71E-08 | $4.23 \mathrm{E}-03$ |
|  | Frontal cortex BA9 | rs7634902 | 5.924265 | 3.14E-09 | 4.90E-04 |
|  | ROSMAP <br> brain | rs2352974 | 6.269043 | $3.63 \mathrm{E}-10$ | 5.67E-05 |
|  | Caudate basal ganglia | rs11716575 | 6.449052 | 1.13E-10 | 1.76E-05 |
|  | Cerebellum | rs11716575 | 7.327323 | $2.35 \mathrm{E}-13$ | 3.67E-08 |
| RP11-I65J3.6 | Cerebellum | rs7025805 | 5.16009 | 2.47E-07 | 3.86E-02 |
| $\begin{aligned} & \text { RP11- } \\ & \text { 481A20.11 } \\ & \hline \end{aligned}$ | ROSMAP <br> brain | rs4841662 | -5.51 | 3.63E-08 | 5.67E-03 |
| $\begin{array}{\|l\|} \hline \text { RP11- } \\ 750 \mathrm{H} 9.5 \\ \hline \end{array}$ | Cerebellar hemisphere | rs896817 | -6.05 | $1.46 \mathrm{E}-09$ | $2.28 \mathrm{E}-04$ |
| SEMA3F | ROSMAP brain | rs2247510 | -5.798856 | 6.68E-09 | $1.04 \mathrm{E}-03$ |
| SLC38A3 | ROSMAP brain | rs1061474 | -5.503226 | 3.73E-08 | 5.82E-03 |


| UBXN2A | Dorsolateral <br> prefrontal <br> cortex | rs12616678 | -5.354554 | $8.58 \mathrm{E}-08$ | $1.34 \mathrm{E}-02$ |
| :--- | :--- | :--- | :--- | :--- | :--- |
|  | ROSMAP <br> brain | rs7588286 | -5.926861 | $3.09 \mathrm{E}-09$ | $4.83 \mathrm{E}-04$ |
|  | ROSMAP <br> brain | rs7821914 | -5.52 | $3.41 \mathrm{E}-08$ | $5.33 \mathrm{E}-03$ |

Table 2. Variance explained of GWAS signals by conditioning on expression for miserableness.

| $\begin{aligned} & \text { Lead GWAS } \\ & \text { SNP } \end{aligned}$ | Conditioned on | Tissue | Before conditioning $P$-value | After conditioning P-value | Variance explained |
| :---: | :---: | :---: | :---: | :---: | :---: |
| rs12612492 | AC008073.7 | Hippocampus | $4.68 \mathrm{E}-08$ | 0.0717 | 0.893 |
| Rs9586 | AMT | Hypothalamus | $1.57 \mathrm{E}-08$ | 0.173 | 0.942 |
| rs1850344 | $\begin{aligned} & \hline \text { RP11- } \\ & \text { 115H18.1 } \end{aligned}$ | Putamen Basal Ganglia | 1.84E-08 | 0.0344 | 0.859 |
| rs4840157 | RP3- <br> 467N11.1 | Cerebellum | $1.02 \mathrm{E}-07$ | 0.0278 | 0.829 |
| rs11764590 | FTSJ2 | Caudate basal ganglia | $9.48 \mathrm{E}-11$ | 1.3E-06 | 0.441 |
| rs6463710 | RPA3 | Caudate basal ganglia | $1.56 \mathrm{E}-06$ | 0.148 | 0.909 |
| rs2715147 | PCLO | Nucleus accumbens basal banglia | 5.5E-07 | 0.141 | 0.914 |
| rs7853605 | RP11-165J3.5 | Cerebellum | 1.9E-07 | 0.499 | 0.983 |
| rs11030009 | DNF-AS | Frontal Cortex | 1.9E-06 | 1.000 | 1.000 |
| rs11039149 | LRP4 | Nucleus accumbens basal ganglia | $1.88 \mathrm{E}-13$ | 0.00204 | 0.824 |
| rs174549 | FADS3 | Cerebellar Hemisphere | 0.00035 | 0.129 | 0.82 |
| rs2013515 | RP11-74E22.6 | Caudate basal ganglia | $1.27 \mathrm{E}-10$ | 1.000 | 1.000 |
| rs12945855 | TTC19 | Nucleus accumbens basal ganglia | 6.75E-09 | 0.00767 | 0.788 |
| rs8072451 | $\begin{aligned} & \text { RP11- } \\ & 798 G 7.5 \end{aligned}$ | Cerebellar Hemisphere | $2.04 \mathrm{E}-12$ | 1.000 | 1.000 |
| Rs1292060 | VMP1 | Cerebellar Hemisphere | 1.2E-07 | 0.0889 | 0.897 |

Table 3. Fine-mapped genes for miserableness.

| Region | Gene | Tissue | TWAS Z-score | Posterior <br> probabilit <br> y for <br> causality |
| :---: | :---: | :---: | :---: | :---: |
| $\begin{aligned} & 1: 173097989- \\ & 1: 175089768 \end{aligned}$ | $\begin{aligned} & \text { RP11- } \\ & \text { 160H22.5 } \\ & \hline \end{aligned}$ | brain cortex | -4.76 | 0.0074 |
| $\begin{aligned} & \hline \text { 1:7247335- } \\ & \text { 1:9365093 } \end{aligned}$ | H6PD | brain hippocampus | 1.86 | 0.0189 |
|  | RERE | brain cortex | -4.39 | 0.0176 |
|  | UTS2 | brain cerebellar hemisphere | -2.94 | 0.0106 |
|  | H6PD | brain cortex | 2.62 | 0.00839 |
|  | PARK7 | brain dorsolateral prefrontal cortex | 2.79 | 0.00442 |
| $\begin{aligned} & 10: 106695048 \\ & - \\ & 10: 108726491 \end{aligned}$ | $\begin{aligned} & \hline \text { RP11- } \\ & \text { 127L20.3 } \end{aligned}$ | brain hippocampus | -5.26 | 0.304 |
| $\begin{aligned} & \hline 11: 44694135- \\ & 11: 47003304 \end{aligned}$ | C1QTNF4 | brain cerebellar hemisphere | 5.26 | 0.0275 |
|  | LRP4 | brain dorsolateral prefrontal cortex | -5.28 | 0.0269 |
|  | HSD17B12 | brain amygdala | -2.55 | 0.0153 |
|  | F2 | brain cerebellum | 5.14 | 0.0104 |
|  | PRDM11 | brain dorsolateral prefrontal cortex | -2.91 | 0.01 |
| $\begin{aligned} & \hline \text { 11:47007278- } \\ & \text { 11:49865926 } \end{aligned}$ | MTCH2 | brain caudate basal ganglia | -5.68 | 0.127 |
|  | FAM180B | brain cerebellar hemisphere | -6.23 | 0.0153 |
|  | MTCH2 | brain nucleus accumbens basal ganglia | -6.18 | 0.573 |
| $\begin{array}{\|l\|} \hline 12: 109025901 \\ - \\ 12: 110336719 \\ \hline \end{array}$ | KCTD10 | brain cerebellum | 5.12 | 0.177 |
|  | MMAB | brain dorsolateral prefrontal cortex | -4.69 | 0.0216 |
|  | VPS29 | brain cortex | 2.32 | 0.00618 |


|  | $\begin{aligned} & \hline \text { RP11- } \\ & \text { 423G4.7 } \\ & \hline \end{aligned}$ | brain caudate basal ganglia | -2.66 | 0.00387 |
| :---: | :---: | :---: | :---: | :---: |
|  | KCTD10 | brain cerebellar hemisphere | 4.12 | 0.00361 |
| $\begin{aligned} & \hline 15: 76398987- \\ & 15: 78516053 \end{aligned}$ | ADAMTS7 | brain caudate basal ganglia | -3.74 | 0.0717 |
|  | LINGO1 | brain cerebellar hemisphere | -2.48 | 0.0618 |
|  | $\begin{aligned} & \hline \text { RP11- } \\ & 762 \mathrm{H} 8.2 \end{aligned}$ | brain cerebellar hemisphere | -2.18 | 0.0148 |
|  | LINGO1 | brain cerebellum | -1.29 | 0.00406 |
|  | CHRNA5 | brain hypothalamus | 2.9 | 0.00391 |
| $\begin{aligned} & \hline \text { 2:147277162- } \\ & \text { 2:150210292 } \end{aligned}$ | ORC4 | brain cerebellar hemisphere | 3.78 | 0.0822 |
|  | LYPD6 | brain cortex | -3.21 | 0.0187 |
|  | ORC4 | brain hypothalamus | 4.33 | 0.524 |
| $\begin{aligned} & \hline \text { 2:171226245- } \\ & \text { 2:173138562 } \end{aligned}$ | DYNC112 | brain cerebellar hemisphere | -3.21 | 0.01 |
|  | SLC25A12 | brain cerebellum | 3.13 | 0.0184 |
|  | DYNC112 | brain cerebellum | -2.99 | 0.00374 |
|  | SLC25A12 | brain cortex | 3.39 | 0.0402 |
|  | SLC25A12 | brain nucleus accumbens basal ganglia | 3.21 | 0.0147 |
| $\begin{aligned} & \text { 2:23342019- } \\ & 2: 24686630 \end{aligned}$ | ATAD2B | brain cerebellar hemisphere | 5.08 | 0.264 |
|  | ATAD2B | brain cerebellum | 5 | 0.175 |
|  | SUCLA2P3 | brain cerebellum | -2.67 | 0.00204 |
|  | UBXN2A | brain dorsolateral prefrontal cortex | -4.74 | 0.0559 |
|  | KLHL29 | brain substantia nigra | 3.23 | 0.00312 |
| $\begin{aligned} & \hline 2: 57429100- \\ & 2: 58296890 \end{aligned}$ | FANCL | brain dorsolateral prefrontal cortex | -4.28 | 0.446 |


|  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
| $\begin{aligned} & \hline 3: 33255592- \\ & 3: 35282963 \end{aligned}$ | RP11- <br> 10C24.1 | brain cortex | -3.51 | 0.0575 |
| $\begin{aligned} & 3: 47727379- \\ & 3: 49316164 \end{aligned}$ | GPX1 | brain frontal cortex ba9 | 5.7 | 0.192 |
|  | AMT | brain hypothalamus | -5.4 | 0.0558 |
|  | AMT | brain hippocampus | -5.31 | 0.0289 |
|  | PRSS45 | brain cerebellar hemisphere | -3.51 | 0.0123 |
|  | AMT | brain frontal cortex ba9 | -5.03 | 0.00828 |
| $\begin{aligned} & \hline 3: 49317338- \\ & 3: 51830565 \end{aligned}$ | AMT | brain hypothalamus | -6.18 | 0.108 |
|  | NCKIPSD | brain cerebellar hemisphere | -5.55 | 0.047 |
|  | GPX1 | brain dorsolateral prefrontal cortex | 5.2 | 0.0389 |
|  | GPX1 | brain frontal cortex ba9 | 5.3 | 0.0353 |
|  | AMT | brain frontal cortex ba9 | -5.35 | 0.033 |
| $\begin{array}{\|l\|} \hline 5: 87390784- \\ 5: 88891530 \\ \hline \end{array}$ | CTC467M3.3 | brain frontal cortex ba9 | -4.37 | 0.567 |
| $\begin{array}{\|l\|} \hline 7: 1353968- \\ 7: 2061783 \\ \hline \end{array}$ | FTSJ2 | brain amygdala | 5.71 | 0.134 |
|  | AC110781.3 | brain nucleus accumbens basal ganglia | 4.19 | 0.00391 |
|  | TMEM184A | brain cerebellar hemisphere | 1.1 | 0.00317 |
|  | GPR146 | brain cerebellar hemisphere | -2.83 | 0.00247 |
|  | PSMG3-AS1 | brain cerebellum | -2.64 | 0.00199 |
| $\begin{aligned} & \hline 7: 2062621- \\ & 7: 2772047 \end{aligned}$ | C7orf50 | brain cortex | 6 | 0.869 |
|  | FTSJ2 | brain caudate basal ganglia | 4.92 | 0.0634 |
|  | FTSJ2 | brain cortex | 5.17 | 0.0358 |
|  | FTSJ2 | brain cerebellar hemisphere | 4.64 | 0.0122 |


|  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: |
|  | INTS1 | brain hypothalamus | -3.59 | 0.0119 |
| $\begin{aligned} & \hline 8: 1163443- \\ & 8: 2042942 \end{aligned}$ | $\begin{aligned} & \text { RP11- } \\ & 439 \mathrm{C} 15.5 \end{aligned}$ | brain cerebellar hemisphere | -3.15 | 0.012 |
| $\begin{aligned} & \hline \text { 17:15020965- } \\ & \text { 17:16411522 } \end{aligned}$ | CENPV | brain spinal cord cervical c-1 | -5.31 | 0.027 |
|  | CENPV | brain anterior cingulate cortex ba24 | -5.17 | 0.0161 |
|  | ADORA2B | brain cerebellum | -4.92 | 0.00452 |
|  | ADORA2B | brain cortex | -4.8 | 0.00289 |
|  | ADORA2B | brain dorsolateral prefrontal cortex | -4.78 | 0.00281 |
| $\begin{aligned} & \hline 17: 1930037- \\ & 17: 3701935 \end{aligned}$ | $\begin{aligned} & \text { RP11- } \\ & \text { 74E22.6 } \end{aligned}$ | brain caudate basal ganglia | 6.13 | 0.0504 |
|  | SERPINF2 | brain nucleus accumbens basal ganglia | 0.0731 | 0.0384 |
|  | RPA1 | brain spinal cord cervical c-1 | -3.45 | 0.00857 |
| $\begin{aligned} & \hline \text { 17:36809344- } \\ & \text { 17:38877404 } \end{aligned}$ | KRT23 | brain caudate basal ganglia | -5.09 | 0.0457 |
|  | RAPGEFL1 | brain cortex | -4.8 | 0.0257 |
|  | MSL1 | brain hypothalamus | -4.66 | 0.0163 |
|  | KRT40 | brain cerebellum | -4.44 | 0.00738 |
|  | MSL1 | brain cerebellum | -4.4 | 0.0065 |
| $\begin{aligned} & 17: 43056905- \\ & 17: 45876022 \end{aligned}$ | FMNL1 | brain cerebellar hemisphere | -7.07 | 0.0255 |
|  | PLEKHM1 | brain cerebellar hemisphere | -7.03 | 0.0253 |
|  | LRRC37A4P | brain nucleus accumbens basal ganglia | -7.05 | 0.0221 |
|  | MAPT | brain dorsolateral prefrontal cortex | -7.04 | 0.0212 |
|  | $\begin{aligned} & \hline \text { RP11- } \\ & \text { 259G18.3 } \end{aligned}$ | brain caudate basal ganglia | 7.01 | 0.0175 |
|  | MICE | brain putamen basal ganglia | -5.5 | 0.167 |


| $\begin{aligned} & \hline 6: 28018353- \\ & 6: 28917091 \end{aligned}$ | IFITM4P | brain cerebellum | -5.35 | 0.104 |
| :---: | :---: | :---: | :---: | :---: |
|  | ZSCAN9 | brain cerebellum | -4.57 | 0.1 |
|  | ZSCAN23 | brain hippocampus | -1.49 | 0.0201 |
|  | PRSS16 | brain cerebellum | -3.97 | 0.0102 |
| $\begin{aligned} & \hline 6: 28917832- \\ & 6: 29737971 \end{aligned}$ | TRIM26 | brain cerebellum | -5.56 | 0.145 |
|  | HLA-H | brain nucleus accumbens basal ganglia | 5.45 | 0.0872 |
|  | HLA-H | brain putamen basal ganglia | 5.31 | 0.0444 |
|  | ZSCAN31 | brain putamen basal ganglia | -2.64 | 0.0438 |
|  | HLA-H | brain caudate basal ganglia | 5.08 | 0.0342 |
| $\begin{aligned} & \hline \text { 6:30798168- } \\ & \text { 6:31570931 } \end{aligned}$ | DXO | brain nucleus accumbens basal ganglia | 5.18 | 0.175 |
|  | STK19P | brain hypothalamus | 5.01 | 0.0529 |
|  | HLA-L | brain nucleus accumbens basal ganglia | 4.88 | 0.0151 |
|  | MSH5 | brain putamen basal ganglia | 3.3 | 0.0136 |
|  | DDAH2 | brain dorsolateral prefrontal cortex | 3.32 | 0.00593 |

# Multi-tissue probabilistic fine-mapping of transcriptome-wide association study identifies cis-regulated genes for miserableness 

Calwing Liao*, Veikko Vuokila*, Alexandre D Laporte, Dan Spiegelman, Patrick A. Dion, Guy A. Rouleau
${ }^{1}$ Department of Human Genetics, McGill University, Montréal, Quebec, Canada
${ }^{2}$ Montreal Neurological Institute, McGill University, Montréal, Quebec, Canada
${ }^{3}$ Department of Neurology and Neurosurgery, McGill University, Montréal, Quebec, Canada

## Supplementary



Supplementary Figure 1. Regional association plot of chromosome 2 conditioned on ACOO8073.7 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.



Supplementary Figure 2. Regional association plot of chromosome 3 conditioned on AMT expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.



Supplementary Figure 3. Regional association plot of chromosome 3 conditioned on RP11115H18.1 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.


Supplementary Figure 4. Regional association plot of chromosome 6 conditioned on RP3467N11.1 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.



## Supplementary Figure 5. Regional association plot of chromosome 7 conditioned on FTSJ2

 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.


## Supplementary Figure 6. Regional association plot of chromosome 7 conditioned on RPA3

 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.

Supplementary Figure 7. Regional association plot of chromosome 7 conditioned on PCLO expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.


Supplementary Figure 8. Regional association plot of chromosome 9 conditioned on RP11165J3.5 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.


Supplementary Figure 9. Regional association plot of chromosome 11 conditioned on BDNF-AS expression. The top panel highlights all genes in the region. The marginally associated TWAS
genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.


Supplementary Figure 10. Regional association plot of chromosome 11 conditioned on LRP4 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.


Supplementary Figure 11. Regional association plot of chromosome 11 conditioned on FADS3 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.



Supplementary Figure 12. Regional association plot of chromosome 17 conditioned on RP11-
74E22.6 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.



Supplementary Figure 13. Regional association plot of chromosome 17 conditioned on TTC19 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.


Supplementary Figure 14. Regional association plot of chromosome 17 conditioned on VMP1 expression. The top panel highlights all genes in the region. The marginally associated TWAS genes are shown in blue and the jointly significant genes are shown in green. The bottom panel shows a regional Manhattan plot of the GWAS data before (grey) and after (blue) conditioning on the predicted expression of the green genes.

Supplementary Table 1. Within-tissue panel significance thresholds

| TI Panel | Number of genes | Within-tissue significance <br> threshold |
| :--- | :--- | :--- |
| GTEx Brain Anterior Cingulate <br> Cortex ba24 | 8731 | $5.73 \mathrm{E}-06$ |
| GTEx Caudate Basal Ganglia | 9145 | $5.47 \mathrm{E}-06$ |


| GTEx Cerebellar Hemisphere | 9451 | $5.29 \mathrm{E}-06$ |
| :--- | :--- | :--- |
| GTEx Cerebellum | 10002 | $5.00 \mathrm{E}-06$ |
| GTEx Cortex | 9162 | $5.46 \mathrm{E}-06$ |
| GTEx Frontal Cortex BA9 | 9031 | $5.54 \mathrm{E}-06$ |
| GTEx Hippocampus | 8535 | $5.86 \mathrm{E}-06$ |
| GTEx Hypothalamus | 8551 | $5.85 \mathrm{E}-06$ |
| GTEx Nucleus Accumbens <br> Basal Ganglia | 8913 | $5.61 \mathrm{E}-06$ |
| GTEx Brain Putamen Basal <br> Ganglia | 8759 | $5.71 \mathrm{E}-06$ |
| CMC DLPFC | 10292 | $4.86 \mathrm{E}-06$ |
| Total | 100572 | $4.9 \mathrm{E}-07$ |

