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Myasthenia and related disorders of the neuromuscular junction

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Abtract

Our understanding of transmission at the neuromuscular junction has increased greatly in recent years. We now recognise a wide variety of autoimmune and genetic diseases that affect this specialised synapse, causing muscle weakness and fatigue. These disorders greatly affect quality of life and rarely can be fatal. Myasthenia Gravis is the most common disorder and is most commonly caused by auto-antibodies targeting postsynaptic acetylcholine receptors (AChRs). Antibodies to muscle-specific kinase (MuSK) are detected in a variable proportion of the remainder. Treatment is symptomatic and immunomodulatory. Lambert-Eaton Myasthenic Syndrome is caused by antibodies to presynaptic calcium channels, and approximately 50% of cases are paraneoplastic, most often related to small cell carcinoma of the lung. Botulism is an acquired disorder caused by neurotoxins produced by clostridium botulinum, impairing acetylcholine release into the synaptic cleft. In addition several rare congenital myasthenic syndromes have been identified, caused by inherited defects in presynaptic, synaptic basal lamina, and postsynaptic proteins necessary for neuromuscular transmission. This review focuses on recent advances in the diagnosis and treatment of these disorders.

Introduction

Abnormalities of synaptic function contribute to many neurological and psychiatric diseases. The archetypical "synaptopathies" are a group of relatively rare disorders affecting the neuromuscular junction (NMJ). Both inherited and acquired disorders of the NMJ cause weakness and fatigability. Much progress has been made in understanding the pathogenesis and diagnosis of these disorders in recent years. This review highlights some recent advances in the understanding of this group of diseases and also considers the emerging evidence regarding their optimal management.

Neuromuscular transmission

The fundamental principles of synaptic transmission were first identified at the NMJ (Fig. 1). Many of the individual steps are directly implicated in neuromuscular disease. Voltage-gated calcium channels open in the presynaptic end-plate in response to the nerve action potential. The subsequent influx of calcium ions triggers exocytosis of vesicles containing acetylcholine (ACh) into the synaptic cleft, in a mechanism that depends on co-ordinated assembly of proteins known as the SNAREs (Soluble NSF Attachment protein Receptors). The SNARE proteins include syntaxin 1, SNAP 25 and synaptobrevin, and are capable of forming a tight complex that brings the vesicle and the plasma membranes into close apposition, in preparation for exoctytosis. Binding of ACh to the postsynaptic receptors opens a cation-permeable pore, which depolarises the muscle fibre. This depolarization can be measured experimentally as an end-plate potential (EPP). Voltage-gated sodium channels open in response to the EPP, triggering a muscle action potential, which leads to release of calcium ions from the sarcoplasmic reticulum. Calcium ions bind to troponin C, which leads to muscle contraction mediated by actin and myosin filaments.

Under normal conditions, neuromuscular transmission has a high safety factor; that is, the EPP is more than sufficient to trigger the action potential (1,2). However, under conditions of impaired transmission, due to insufficient presynaptic ACh release or alternatively because of a defect of postsynaptic receptors, the EPP can become sub-threshold, leading to muscle weakness. The NMJ is a target of several autoimmune diseases, caused by antibodies to pre- or postsynaptic proteins. It is also affected in botulism, where SNARE proteins are cleaved. And several congenital myasthenic syndromes (CMS) result from inherited defects of proteins at all stages of neuromuscular transmission.

Myasthenia gravis (MG)

Epidemiology

MG is the commonest disorder of the NMJ, with a prevalence of around 15 per 100,000, although it is probably under-diagnosed in the elderly population. The incidence is bimodal, with a female to male ratio of approximately 2:1 in young adults, and a reversed sex ratio in the older group that accounts for 60% of cases (3-6).

Immunopathogenesis

MG is firmly established as an autoimmune disease (1,2). In the commonest form of MG, antibodies that bind to ACh receptors are detected in the synaptic cleft (7-9). These antibodies are of the IgG1 or IgG3 subtypes and activate complement, resulting in focal destruction of the postsynaptic membrane by the membrane attack complex (10). In addition, they cross-link AChRs, leading to an increase in the rate of destruction, and may also inhibit receptor function directly (7).

A distinct form of MG is caused by antibodies targeting muscle-specific kinase (MuSK) and will be discussed separately.

The thymus has an important role in the pathogenesis of early onset MG associated with anti-AChR antibodies, and is frequently enlarged with lymphocytic infiltrates, germinal centres and myoid cells, which express AChRs. (7) The thymus gland in patients developing MG later in life is however typically atrophic, and its immunogenic role is less clear.

A thymoma is present in 10% of patients with MG, with a peak incidence in the 4th to 6th decades and an equal frequency in males and females. Patients with thymoma often have antibodies to various components of striated muscle: titin, myosin, actin and ryanodine receptors which are associated with a more severe clinical MG phenotype (11,12).

Clinical Manifestations

In common with other acquired and inherited disorders of the NMJ, MG manifests as painless fatigable muscle weakness. The pattern of muscle involvement varies, but extraocular muscles are most commonly affected, causing diplopia and ptosis. Approximately 80% of patients presenting with ocular symptoms will eventually develop generalised MG(13). Up to 20% of patients with MG have prominent bulbar symptoms early in their disease (14). Limb weakness can affect any muscle group, although proximal muscles are most often affected. Symptoms typically worsen towards the end of the day and with exercise. Extreme heat, emotional stress, infection, pregnancy and menstruation can all exacerbate myasthenia, as can many drugs, including aminoglycoside antibiotics and penicillamine.

Prognosis/ Natural History

MG has a highly variable course. If symptoms remain purely ocular for more than 2 years, the risk of progression to generalised MG is less than 10% (13). However, approximately 39% of patients have severe MG and half of these will require invasive ventilation during the course of their disease (13). The forced vital capacity should be measured in all patients to look out for

this complication. The mortality of MG has nevertheless dropped significantly from over 30% in the 1950s to less than 5%. This is most likely due to improved ventilatory care and increased availability of immunomodulatory treatments(15,3). Assessment of outcome has been helped by the establishment of a task force by the Myasthenia Gravis Foundation of America in 1997, which led to recommendations for clinical research standards, including uniformity in the reporting of clinical trials (16,17).

Diagnosis

Relevant questions to the patient include pattern of muscle weakness, fatigability and exacerbating factors. Sensory deficits and absent/increased reflexes are not features of MG. Fatigability can be elicited by observing for the development of ptosis and diplopia during prolonged upgaze or by testing neck flexion/extension, and shoulder abduction before and after unilateral repetitive movement. A relatively sensitive and specific bedside test that distinguishes myasthenia from other causes of ptosis involves application of crushed ice in a latex glove to the eye. This leads to improvement of ptosis in MG and has been reported to have a sensitivity of 89% (18).

Edrophonium test

Although this test is uncommonly performed, it can be useful if there are delays in obtaining other investigations, and has a high sensitivity for generalised MG. The acetylcholinesterase inhibitor is administered intravenously, observing for transient improvement in muscle strength (such as resolution of ptosis) (19). Oral acetylcholinesterase inhibitors should be withheld for at least 24 hours before testing, and atropine and resuscitation facilities should be available.

Neurophysiological tests

Routine nerve conduction tests and EMG are not usually informative, although the compound muscle action potential (CMAP) can be reduced in severe cases. However, repetitive nerve

stimulation (RNS) typically elicits a decremental response at 3-10 Hz, with a decrement >10% considered abnormal (20). The sensitivity of RNS for diagnosing MG ranges from 53% to 100%. It is however frequently negative in ocular MG (21).

Single fibre EMG (SFEMG) is more sensitive, and especially useful in ocular MG, although it is less specific Jitter, the trial to trial variation in the latency from stimulus to response, is increased, and there may also be intermittent failure of excitation of muscle fibres ('block').

Immunology

Not all patients with MG will have demonstrable antibodies but in those that do, these tests are highly specific. Approximately 85% of patients with generalised MG, and almost all patients with an associated thymoma, have detectable circulating autoantibodies to AChRs (22) A variable proportion of patients who do not have anti-AChR antibodies instead have antibodies to MuSK, and the remaining patients are described as 'seronegative', although a seroconversion rate of 15% has been reported. Immunosuppression itself can occasionally lead to disappearance of antibodies (23).

MuSK

MuSk (Muscle-Specific tyrosine Kinase) antibodies occur in up to 70% of patients who test negative for AChR antibodies, and are more prevalent in countries close to the equator (24). MuSK is essential for AChR clustering at the NMJ (25). MuSK positive patients are more likely to have early bulbar and respiratory symptoms with less severe limb involvement and women are more commonly affected (26). The clinical neurophysiology in MuSK usually shows abnormal SFEMG of facial muscles but RNS of limb muscles can be normal. Thymic pathology in MuSK patients does not show lymphocytic infiltrates or complement deposition In addition muscle biopsy does not show reduced AChR density as it does in AChR antibody positive patients (27). The pathogenicity of anti-MuSK antibodies, although confirmed by passive transfer experiments is incompletely understood. However, it has been shown that interfering with MuSk synthesis can cause declustering of AChRs (28).

Seronegative MG

A small proportion of patients remain consistently negative for both AchR and MuSK antibodies. Clinical features and thymic pathology imply that seronegative MG (SNMG) is similar to MG caused by AChR antibodies. This has led to the hypothesis that the patients have antibodies to AChR which are undetectable with conventional assays. Indeed, many such patients can be shown to have antibodies that bind AChRs expressed in a cell line together with rapsyn, which contributes to clustering AChRs (29). It remains to be determined whether other antigens are recognized by circulating antibodies in the remaining 5% of so of patients with MG in whom this improved assay is negative.

Treatment of MG

Treatment of MG is symptomatic (AChE inhibition) or immunomodulatory, although there is a paucity of controlled randomised controlled trials to provide definitive guidance.

Acetylcholinesterase (AChE) inhibitors are usually first line therapy, providing symptomatic relief in ocular and generalised myasthenia. There are no placebo-controlled randomized studies but clinical experience and case reports support their effectiveness (30). Patients with anti MuSK antibodies may not respond as well as those with anti-AChR MG. Side-effects include hyperhidrosis, salivation and lacrimation. Cholinergic crisis can occur with high doses.

AChE inhibitors have a short half life and thus require regular dosing. A controlled-release form of pyridostigmine is available in some countries, but not in the UK. Most patients will require immunomodulatory treatment but given the adverse effects associated with these drugs, it may be reasonable to treat with an AChE alone initially.

Immunosuppression

Corticosteroids are generally the first line immunosuppressive treatment. A Cochrane review found only limited evidence from randomised controlled trials; nonetheless several observational trials support their efficacy (31). Early use of steroids in ocular MG may prevent generalisation (32,33). High initial doses of prednisolone (prednisone) may transiently exacerbate weakness and even trigger a crisis, and it may be prudent to admit patients to hospital for escalation of treatment. Bone protection strategies and alternate day dosing are often used to counteract steroid-related side-effects.

Azathioprine is the most widely used second-line immunosuppressant. Although a Cochrane review did not demonstrate any significant improvement in quantitative MG (QMG) score at six month, its use does allow a lower steroid dose (34-37). Simultaneously commencing azathioprine and prednisolone may allow more rapid steroid tapering. Adverse effects include hepatotoxicity, cytopenia, and lymhoproliferative disease. One case of progressive multifocal leukoencephalopathy (PML) has been reported (38). Thiopurine methyltransferase (TPMT) activity should be measured before commencement as patients with reduced activity of this enzyme will become toxic at normal doses. Regular monitoring of full blood count (FBC) and liver and renal function is warranted.

Cyclosporine has been shown to be effective in improving muscle strength and reducing steroid dose and indeed has the strongest evidence base of the immunosupressants used in MG (39,40). Despite this, concerns about side-effects, mainly nephrotoxicity, have limited its widespread use. Tacrolimus has also been shown to be efficacious in initial treatment of MG (41). Use of cyclophosphamide improves muscle weakness but because of severe side-effects it is generally reserved for patients who are intolerant of other immunosuppressants (42). Despite a lack of randomised controlled trials, methotrexate is increasingly used in the management of MG. Its use has been recommended as a second line agent by the European Federation of Neurological Societies (43), although it is relatively contra-indicated in women of child-bearing age.

Mycophenolate mofetil has been used in transplant medicine and in other autoimmune diseases without producing major organ toxicity (44). It has been adopted widely in MG (45-47). However, two recent randomized studies have failed to demonstrate additional benefit of mycophenolate with prednisolone compared to prednisolone alone (48,49). The negative outcome of these studies has not deterred many clinicians from using it in MG, mainly because of its favourable side-effect profile, and further evidence is needed to determine its role.

Intravenous Immunoglobulin (IVIG) and plasma exchange are both effective in acute exacerbations of MG, although their long term effectiveness has not been demonstrated in clinical trials (50-52). Their efficacy is similar although IVIG is better tolerated. Their main role is in bringing about a rapid improvement in patients with worsening myasthenic symptoms (53). A small minority of patients remain refractory to oral medication require frequent courses of IVIG or plasma exchange.

Rituximab is a monoclonal antibody to the B cell marker CD20. Its use in MG has been described in case reports (54-57) It has been used both in AChR- and MuSK-antibody positive patients, and in patients with thymoma, although there is no randomised evidence. Side-effects include neutropenia and increased susceptibility to infections, Other B cell directed approaches include a humanized anti CD20 antibody (ofatumumab) and atacicept which sequesters B cell survival factors (58). Use of etanercept, soluble recombinant TNF receptor FC protein, in MG has been disappointing (59). A novel complement inhibitor REV576 was recently used in an experimental rodent model of MG rats with promising results, and a trial of eculuzimab, which has a similar mechanism of action, is under way (60).

Surgery

Surgery with or without *radiotherapy* is indicated in the treatment of thymoma. It is sometimes necessary to stabilise myasthenic symptoms preoperatively, and IVIG and plasma exchange have been used in this situation.

The value of *thymectomy* in patients with non-thymomatous MG is less clear. Retrospective follow up studies suggest benefit but clinical effects may not be seen for 6-12 months (61-63).

Despite its use since the 1930s no randomized trials have been published; one is currently underway (64).

The outcomes of different surgical approaches appear equivalent but the routine use of minimally invasive procedures is controversial since thymoma or thymic carcinoma may only be identified intraoperatively (65). Consensus is lacking regarding the age limits for thymectomy, its use in ocular myasthenia or the stage of disease that thymectomy should be performed (58). In general thymectomy is offered to young AChR antibody-positive patients with generalised MG. There is no evidence that patients with MuSK antibodies benefit from thymectomy

Ocular MG

Extraocular muscles are affected in at least 85% of patients with MG but it is unclear why they are so vulnerable. They have a high blood flow, mitochondrial content and metabolic rate. They also have small motor units that fire at high firing frequencies, and possibly a smaller safety factor, making them prone to fatigue. There may also be differences in intrinsic regulators of the complement cascade, making them more susceptible to complement-mediated injury(66)

AChE inhibitors are first line therapy in ocular myasthenia and in very mild disease may be all that is required. However, ocular symptoms can be disabling and 60-80% of patients will require additional therapy. Prednisolone therapy has been shown to reduce ocular symptoms of ptosis and diplopia and observational studies suggest benefit (67-70). Steroids have been reported in retrospective studies to delay the onset of generalised MG but there is no definitive evidence regarding this.

Myasthenic Crisis

Myasthenic crisis is a neurological emergency requiring intensive care unit admission for ventilatory support, and removal of circulating autoantibodies with IVIG or plasma exchange

can be highly effective (71). Cholinesterase inhibitors have a limited role in the treatment of myasthenic crisis, and indeed, a cholinergic crisis an occasionally mimic it. Case-control studies have suggested the long term beneficial effect of immunosuppressive therapy in preventing myasthenic crisis (72).

Lambert-Eaton Myasthenic Syndrome

Lambert-Eaton Myasthenic Syndrome (LEMS) is much less common than MG (73). It usually begins in mid to late life and affects males and females equally ((74). It is associated with a malignancy, most often small cell carcinoma of the lung (SCLC), in approximately 50% of cases. Conversely, 1-3% of SCLC patients have LEMS. In the vast majority of cases, the tumour is detected within a year of diagnosis of LEMS (75). In patients who do not have an associated tumour, there a female preponderance and a strong association with HLA-B8. Around 25% of non tumour LEMS patients have a past history of organ specific autoimmune disease (ref*).

Most patients develop proximal limb weakness, especially of the legs. Areflexia, dry mouth and signs of autonomic failure also occur (74). Some patients, most often those with small cell carcinoma of the lung, may have associated paraneoplastic cerebellar degeneration.

Antibodies in LEMS are directed against presynaptic calcium channels, most likely of the P/Q-type (CaV2.1), and interfere with the release of ACh. They are detectable in 85% of patients and are very specific (2). In a recent study antibodies to SOX1, a DNA binding transcription factor were found in paraneoplastic LEMS patients but not in those who did not have a tumour (76).

In contrast to MG, the CMAP amplitude increases in LEMS in response to high frequency stimulation (77). This is thought to be due to presynaptic accumulation of calcium, which slowly recruits additional vesicles for exocytosis. The same phenomenon can be demonstrated clinically by the return of previously absent deep tendon reflexes after muscle contraction.

The diagnosis of LEMS should spark an intensive search for malignancy, including CT thorax, bronchoscopy and PET. Patients with a history of smoking, cerebellar dysfunction and older patients with rapid progression of symptoms are more likely to have an underlying malignancy(78). The prognosis in patients with both LEMS and SCLC is poor, with a 2 year survival of less than 10%, albeit better than in patients without LEMS, suggesting a possible anti-tumour effect of the antibodies.

Treatment of muscle weakness in LEMS is symptomatic or immune-based. Aminopyridines such as *3,4-diaminopyridine* block potassium channels, and prolong the duration of the nerve action potential increasing neurotransmitter release at the synapse. Their efficacy has been summarized in a Cochrane review of two randomized trial with up to 79% of patients demonstrating significant symptomatic improvement (79-81). AChE inhibitors provide a moderate response at best and are generally not used as monotherapy. Guanidine is an alternative treatment that increases free intracellular calcium and ACh release. It has not been subjected to randomized controlled trials. Side-effects include gastrointestinal disturbance, bone marrow suppression and renal failure. It is generally less well tolerated than the aminopyridines.

IVIG has been shown to be useful in LEMS, leading to clinical improvement and a decrease in antibody titre. It is generally used as an adjuvant therapy in treatment-resistant patients (82) Oral prednisolone has been used alone and in combination with azathioprine, and case reports have suggested benefit(79). More recently a single case report has highlighted a beneficial response to rituximab in a non-paraneoplastic LEMS patients who had failed conservative therapy (83)

Botulism

Botulism is a rare disease caused by exposure to the neurotoxins produced by the anaerobe clostridium botulinum that is found in soil and aquatic sediment. The three toxin subtypes A, B

and E all act by cleaving SNARE proteins, although they recognize different sequences. Their action prevents the close apposition of vesicles to the presynaptic membrane, leading to a failure of ACh release (84,85) (Fig. 1). There are four types of botulism, classified according to the mode of exposure.

Food-borne botulism is caused by consumption of toxin formed under anaerobic conditions, such as in inadequately sterilized home-made preserves (86). Wound botulism is particularly seen in drug users who use 'skin popping' to self-administer heroin. The organism does not normally survive ingestion but can do so in neonates, where it causes infant botulism (84). Rarely intestinal colonization occurs in adults with functional bowel abnormality. Finally, iatrogenic botulism has been reported in patients treated with toxin for movement disorders or cosmetic purposes (87,88)

Botulism typically presents with a descending paralysis, causing symmetrical ptosis, opthalmoplegia, dysarthia and dysphagia. This progresses to proximal limb weakness and respiratory compromise. Autonomic involvement, including anhidrosis and postural hypotension also occur, and gastrointestinal symptoms may be the first sign of food-borne botulism. Reflexes are typically lost but sensation is normal.

Neurophysiological tests reveal changes similar to those in LEMS. An enzyme-linked immunosorbent assay, detecting toxin in serum, stool or in a food sample, has now displaced a mouse bioassay.

Supportive intensive care management has decreased mortality from 79% to 3-5%. It is essential to administer antitoxin early because the toxin is internalized and continues to cleave SNARE proteins after it has been cleared from the circulation. Public health authorities must be informed if a botulism outbreak is suspected.

Congenital Myasthenic Syndromes

A heterogenous group of genetic disorders can affect the NMJ. These are typically inherited in an autosomal recessive fashion. Early studies on patients with inherited myasthenic disorders used electrophysiology, electron microscopy, histochemistry and biochemical techniques to show that variable synaptic proteins were likely to be affected in different families. The congenital myasthenic syndromes (CMSs) were first classified on the basis of the site of the defective neuromuscular transmission – presynaptic, synaptic and postsynaptic. Within these headings different genes may be defective and the clinical picture varies. Currently, definitive diagnosis depends upon electrophysiological tests, morphological studies of the end-plate region in muscle biopsy specimens, and increasingly on identification of the specific genetic defect. To date 13 genes have been identified (Table 1). However, mutations have yet to be identified in approximately 30% of patients referred to the Oxford CMS diagnostic service whose clinical features strongly suggest a genetic myasthenic syndrome. As new CMS-associated genes are identified a revised classification is evolving, based on the gene defects and the underlying molecular mechanisms of the disorder.

AChR subunits

Initial studies identified mutations in the genes encoding the AChR subunits that impair ion channel gating, reduce the number of endplate receptors, or a combination of the two, leading to "slow channel", "fast channel" or AChR deficiency syndromes (89). Slow channel syndrome is the only dominantly inherited CMS.

Presynaptic and synaptic CMS

Mutations were subsequently identified in the acetylcholinesterase collagen-like tail subunit gene (COLQ) and also in the enzyme choline acetyltransferase (ChAT) (90-92).

AChR clustering disorders

More recently, mutations have been found in the genes encoding key molecules involved in clustering of the AChR at the endplate and in maintaining synaptic structure (Fig 1). Agrin released from the nerve terminal activates MuSK on the postsynaptic membrane through low density lipoprotein receptor-related protein 4 (LRP4). This, in turn, leads to aggregation of AChRs through an interaction with rapsyn (93,94). The activation of MuSK is enhanced by the cytoplasmic protein Dok-7 (95). With the exception of LRP4, mutations in each of these components have been reported in CMS, emphasising their critical roles in this pathway (Table 1). To date there have been only isolated case reports of AGRIN or MUSK mutations (96). However, RAPSN and DOK 7 mutations are well recognized as major causes of CMS (97-103).

Dok-7 (gene DOK7) binds to MuSK and plays a role in amplifying the MuSK signaling to downstream pathways that control clustering, partially through phosphorylation of the AChR β subunit, and maintenance of the NMJ. The syndrome associated with DOK7 mutations is characterized by a limb girdle pattern of weakness. Children typically have normal initial motor milestones followed by an abnormal waddling or lordotic gait and frequent falls soon after learning to walk. Ptosis, stridor and respiratory problems may occur but eye movements are usually spared. Bulbar problems often develop later. Fluctuation in symptoms is common and patients may have been previously diagnosed with an unspecified congenital myopathy (102). There is a remarkable variation in disease severity with symptom onset ranging from birth to adolescence or even in adulthood.

Rapsyn is an AChR-clustering protein and mutations in the RAPSN gene cause a deficiency of AChR at the motor endplate. Both early and late onset cases have been reported. (98) Early onset cases are associated with hypotonia and bulbar dysfunction, which may require assisted feeding and ventilation. Mild facial malformations and contractures of hands and ankles are common. Patients may suffer severe exacerbations with life threatening respiratory failure; however, the condition does tend to improve over time with minimal

disability in adulthood. Interestingly, weakness of ankle dorsiflexion tends to remain into adult life and may provide a diagnostic clue. The overwhelming majority of patients harbour the missense mutation N88K, suggesting an original founder mutation (104). Other mutations have been identified in the promoter and coding regions of the RAPSN gene. In addition, a missense mutation in the AChR δ subunit gene δ E381K has been identified that does not affect the function of the receptor but rather impairs rapsyn-induced AChR clustering. These patients bear, phenotypically, the hallmarks of rapsyn deficiency rather than AChR subunit deficiency and this mutation may prove useful for studying the AChR-rapysn interaction.

The facial malformations and joint contractures seen in rapsyn mutations are thought to result from lack of movement in utero. Neuromuscular transmission is generally mediated by the adult AChR, which has the stoichiometry $(\alpha_1)2\beta_1\delta\epsilon$. There is however a fetal form of the AChR, $(\alpha_1)2\beta_1\gamma\delta$, which is crucial in certain periods of fetal development. Mutations in the fetal γ subunit can lead to severe developmental abnormalities or death. Surprisingly, some patients with γ subunit null alleles survive, suggesting that early expression of the adult ϵ subunit may compensate. It is likely that loss of function mutations in other subunits of the fetal AChR or other components of the developing NMJ are likely to be fatal. Indeed null mutations have now been found in *CHRNA1*, *CHRND*, *RAPSN* and *DOK7* (105,106).

Mutations are not identified in approximately 30% of patients with a clinical diagnosis of CMS. A form of limb girdle CMS that does not have a demonstrable DOK7 mutation and differs from this condition in its response to anticholinesterase and in the presence of tubular aggregates on muscle biopsy exists. Studies are currently underway to define the affected gene for this characteristic phenotype.

A mutation was recently described in the LAMB2 gene that codes the laminin $\beta 2$ subunit. Mutations in Lamb2 have previously been shown to be involved in Pierson syndrome which causes ocular abnormalities and nephrosis. Mice lacking laminin $\beta 2$ do not develop normal neuromuscular synapses (107).

Treatment

With increased understanding of the molecular mechanisms that underlie individual CMSs, therapy can be tailored to the individual. Most CMSs such as AChR deficiency, fast channel syndrome and rapsyn deficiency result from "loss of function" mutations that reduce signal transmission. These patients tend to respond to therapies that enhance neurotransmitter release such as AChE inhibitors and 3,4 diaminopyridine (3,4-DAP), although theoretically ChAT CMS could worsen in response to 3,4 DAP due to depletion of the presynaptic ACh.

By contrast in patients with slow channel syndrome or COLQ mutations, there is overstimulation of endplate receptors, calling for quite different therapy. The prolonged opening time of the AChR seen in slow channel syndrome can be curtailed by drugs that block the channel in its open state, so called "open channel blockers". Fluoxetine and quinidine sulphate have both been used for this purpse (108,109). In patients with COLQ mutations, prolonged exposure to ACh casues receptor desensitization and an endplate myopathy. This has been found to respond to ephedrine(110,111).

Dok-7 mutations may respond well to appropriate treatment. Cholinesterase inhibitors often elicit a brief initial response but subsequently cause worsening of symptoms.

Response to 3,4-diaminopyridine is mixed, with some patients reporting benefits and others worsening.(80) However, patients usually respond well to ephedrine (101), although the basis for this is unknown. Patients may also respond to salbutamol (112) suggesting that

stimulation of $\beta 2$ -adrenergic receptors may be able to partially stabilize the end plate region.

Summary

Increased understanding of the fundamental mechanisms of synaptic transmission and of the development and maintenance of the end-plate has greatly enhanced the ability to diagnose patients with acquired or inherited diseases of the neuromuscular junction. Nevertheless, there remains a lack of robust evidence to guide the treatment of even the most common of the neuromuscular junction diseases, myasthenia gravis. Further research in the form of randomized controlled trials of currently used treatments is indicated. As we understand more about the pathogenic mechanisms that interrupt individual proteins at the neuromuscular junction, we will be able to develop more specific treatments for both the acquired and congenital disorders of this highly specialized synapse.

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Competing Interest: None declared.

Presynaptic CMS	Gene
CMS with episodic apnea	CHAT
Synaptic CMS	
Congenital endplate acetylcholinesterase deficiency	COLQ
CMS due to mutations in Agrin	AGRN
CMS due to mutations in the laminin β2 chain	LAMB2
Postsynaptic CMS	
AChR deficiency syndromes	CHRNA, CHRNB,
	CHRND, CHRNE
AChR deficiency syndromes due to mutations in rapsyn	RAPSN
Slow-channel CMS	CHRNA, CHRNB,
	CHRND, CHRNE
Fast-channel CMS	CHRNA, CHRND,
	CHRNE
CMS due to voltage-gated sodium channel mutations	SCN4A
CMS due to mutations in MuSK	MUSK
CMS due to mutations in Dok-7	DOK7
Multiple pterygium/Escobar syndromes due to AChR γ-subunit	CHRNG
mutations	
Fatal fetal akinesia deformation sequence due to NMJ protein	CHRNA, CHRND,
mutations	RAPSN, DOK7

Table 1. Classification of congenital myasthenic syndromes (CMS). The genetic causes of some cases of presynaptic and postsynaptic CMS remain to be defined.

References

- 1. Conti-Fine BM, Milani M, Kaminski HJ. Myasthenia gravis: past, present, and future. J. Clin. Invest. 2006 Nov;116(11):2843-2854.
- 2. Vincent. Autoimmune disorders of the neuromuscular junction [Internet]. 2008 Jul 1 [cited 2009 Jul 8]; Available from: http://www.neurologyindia.com/article.asp?issn=0028-3886; year=2008; volume=56; issue=3; spage=305; epage=313; aulast=Vincent
- 3. Alshekhlee A, Miles JD, Katirji B, Preston DC, Kaminski HJ. Incidence and mortality rates of myasthenia gravis and myasthenic crisis in US hospitals. Neurology. 2009 May 5;72(18):1548-1554.
- 4. Robertson NP, Deans J, Compston DA. Myasthenia gravis: a population based epidemiological study in Cambridgeshire, England. J. Neurol. Neurosurg. Psychiatr. 1998 Oct;65(4):492-496.
- 5. Phillips LH. The epidemiology of myasthenia gravis. Ann. N. Y. Acad. Sci. 2003 Sep;998:407-412.
- 6. Vincent A, Clover L, Buckley C, Grimley Evans J, Rothwell PM. Evidence of underdiagnosis of myasthenia gravis in older people. J. Neurol. Neurosurg. Psychiatr. 2003 Aug;74(8):1105-1108.
- 7. Vincent A. Unravelling the pathogenesis of myasthenia gravis. Nat Rev Immunol. 2002 Oct;2(10):797-804.
- 8. Fambrough DM, Drachman DB, Satyamurti S. Neuromuscular junction in myasthenia gravis: decreased acetylcholine receptors. Science. 1973 Oct 19;182(109):293-295.
- 9. Toyka KV, Drachman DB, Griffin DE, Pestronk A, Winkelstein JA, Fishbeck KH, et al. Myasthenia gravis. Study of humoral immune mechanisms by passive transfer to mice. N. Engl. J. Med. 1977 Jan 20;296(3):125-131.
- 10. Meriggioli MN, Sanders DB. Autoimmune myasthenia gravis: emerging clinical and biological heterogeneity. Lancet Neurol. 2009 May;8(5):475-490.
- 11. Mygland A, Aarli JA, Matre R, Gilhus NE. Ryanodine receptor antibodies related to severity of thymoma associated myasthenia gravis. J. Neurol. Neurosurg. Psychiatr. 1994 Jul;57(7):843-846.
- 12. Skeie GO, Romi F. Paraneoplastic myasthenia gravis: immunological and clinical aspects. Eur. J. Neurol. 2008 Oct;15(10):1029-1033.

- 13. Grob D, Brunner N, Namba T, Pagala M. Lifetime course of myasthenia gravis. Muscle Nerve. 2008 Feb;37(2):141-149.
- 14. Grob D, Arsura EL, Brunner NG, Namba T. The course of myasthenia gravis and therapies affecting outcome. Ann. N. Y. Acad. Sci. 1987;505:472-499.
- 15. Barohn RJ. Treatment and clinical research in myasthenia gravis: how far have we come? Ann. N. Y. Acad. Sci. 2008;1132:225-232.
- 16. Barohn RJ. Standards of Measurements in Myasthenia Gravis. Annals of the New York Academy of Sciences. 2003 9;998(1):432-439.
- 17. Jaretzki A, Barohn RJ, Ernstoff RM, Kaminski HJ, Keesey JC, Penn AS, et al. Myasthenia gravis: recommendations for clinical research standards. Task Force of the Medical Scientific Advisory Board of the Myasthenia Gravis Foundation of America. Neurology. 2000 Jul 12;55(1):16-23.
- 18. Sethi KD, Rivner MH, Swift TR. Ice pack test for myasthenia gravis. Neurology. 1987 Aug;37(8):1383-1385.
- 19. Meriggioli MN, Sanders DB. Advances in the diagnosis of neuromuscular junction disorders. Am J Phys Med Rehabil. 2005 Aug;84(8):627-638.
- 20. Literature review of the usefulness of repetitive nerve stimulation and single fiber EMG in the electrodiagnostic evaluation of patients with suspected myasthenia gravis or Lambert-Eaton myasthenic syndrome. Muscle Nerve. 2001 Sep;24(9):1239-1247.
- 21. Padua L, Stalberg E, LoMonaco M, Evoli A, Batocchi A, Tonali P. SFEMG in ocular myasthenia gravis diagnosis. Clin Neurophysiol. 2000 Jul;111(7):1203-1207.
- 22. Meriggioli MN. Myasthenia gravis with anti-acetylcholine receptor antibodies. Front Neurol Neurosci. 2009;26:94-108.
- 23. Chan KH, Lachance DH, Harper CM, Lennon VA. Frequency of seronegativity in adultacquired generalized myasthenia gravis. Muscle Nerve. 2007 Nov;36(5):651-658.
- 24. Vincent A, Leite MI, Farrugia ME, Jacob S, Viegas S, Shiraishi H, et al. Myasthenia gravis seronegative for acetylcholine receptor antibodies. Ann. N. Y. Acad. Sci. 2008;1132:84-92.
- 25. Vincent A, Leite MI. Neuromuscular junction autoimmune disease: muscle specific kinase antibodies and treatments for myasthenia gravis. Curr. Opin. Neurol. 2005 Oct;18(5):519-525.
- 26. Lavrnic D, Losen M, Vujic A, De Baets M, Hajdukovic LJ, Stojanovic V, et al. The

- features of myasthenia gravis with autoantibodies to MuSK. J. Neurol. Neurosurg. Psychiatr. 2005 Aug;76(8):1099-1102.
- 27. Shiraishi H, Motomura M, Yoshimura T, Fukudome T, Fukuda T, Nakao Y, et al. Acetylcholine receptors loss and postsynaptic damage in MuSK antibody-positive myasthenia gravis. Ann. Neurol. 2005 Feb;57(2):289-293.
- 28. Kong XC, Barzaghi P, Ruegg MA. Inhibition of synapse assembly in mammalian muscle in vivo by RNA interference. EMBO Rep. 2004 Feb;5(2):183-188.
- 29. Leite MI, Jacob S, Viegas S, Cossins J, Clover L, Morgan BP, et al. IgG1 antibodies to acetylcholine receptors in 'seronegative' myasthenia gravis. Brain. 2008 Jul;131(Pt 7):1940-1952.
- 30. Skeie GO, Apostolski S, Evoli A, Gilhus NE, Hart IK, Harms L, et al. Guidelines for the treatment of autoimmune neuromuscular transmission disorders. Eur. J. Neurol. 2006 Jul;13(7):691-699.
- 31. Schneider-Gold C, Gajdos P, Toyka KV, Hohlfeld RR. Corticosteroids for myasthenia gravis. Cochrane Database Syst Rev. 2005;(2):CD002828.
- 32. Sathasivam S. Steroids and immunosuppressant drugs in myasthenia gravis. Nat Clin Pract Neurol. 2008 Jun;4(6):317-327.
- 33. Sommer N, Sigg B, Melms A, Weller M, Schepelmann K, Herzau V, et al. Ocular myasthenia gravis: response to long-term immunosuppressive treatment. J. Neurol. Neurosurg. Psychiatr. 1997 Feb;62(2):156-162.
- 34. Hart IK, Sharshar T, Sathasivam S. Immunosuppressant drugs for myasthenia gravis. J. Neurol. Neurosurg. Psychiatr. 2009 Jan;80(1):5-6; discussion 6.
- 35. A randomised clinical trial comparing prednisone and azathioprine in myasthenia gravis. Results of the second interim analysis. Myasthenia Gravis Clinical Study Group. J. Neurol. Neurosurg. Psychiatr. 1993 Nov;56(11):1157-1163.
- 36. Palace J, Newsom-Davis J, Lecky B. A randomized double-blind trial of prednisolone alone or with azathioprine in myasthenia gravis. Myasthenia Gravis Study Group. Neurology. 1998 Jun;50(6):1778-1783.
- 37. Sathasivam S. Steroids and immunosuppressant drugs in myasthenia gravis. Nat Clin Pract Neurol. 2008 Jun;4(6):317-327.
- 38. Gedizlioglu M, Coban P, Ce P, Sivasli IE. An unusual complication of immunosuppression in myasthenia gravis: progressive multifocal leukoencephalopathy. Neuromuscul. Disord. 2009 Feb;19(2):155-157.

- 39. Tindall RS, Phillips JT, Rollins JA, Wells L, Hall K. A clinical therapeutic trial of cyclosporine in myasthenia gravis. Ann. N. Y. Acad. Sci. 1993 Jun 21;681:539-551.
- 40. Hart IK, Sathasivam S, Sharshar T. Immunosuppressive agents for myasthenia gravis. Cochrane Database Syst Rev. 2007;(4):CD005224.
- 41. Nagane Y, Utsugisawa K, Obara D, Kondoh R, Terayama Y. Efficacy of low-dose FK506 in the treatment of Myasthenia gravis--a randomized pilot study. Eur. Neurol. 2005;53(3):146-150.
- 42. Drachman DB, Jones RJ, Brodsky RA. Treatment of refractory myasthenia: "rebooting" with high-dose cyclophosphamide. Ann. Neurol. 2003 Jan;53(1):29-34.
- 43. Skeie GO, Apostolski S, Evoli A, Gilhus NE, Hart IK, Harms L, et al. Guidelines for the treatment of autoimmune neuromuscular transmission disorders. Eur. J. Neurol. 2006 Jul;13(7):691-699.
- 44. Heatwole C, Ciafaloni E. Mycophenolate mofetil for myasthenia gravis: a clear and present controversy. Neuropsychiatr Dis Treat. 2008 Dec;4(6):1203-1209.
- 45. Hauser RA, Malek AR, Rosen R. Successful treatment of a patient with severe refractory myasthenia gravis using mycophenolate mofetil. Neurology. 1998 Sep;51(3):912-913.
- 46. Chaudhry V, Cornblath DR, Griffin JW, O'Brien R, Drachman DB. Mycophenolate mofetil: a safe and promising immunosuppressant in neuromuscular diseases. Neurology. 2001 Jan 9;56(1):94-96.
- 47. Meriggioli MN, Ciafaloni E, Al-Hayk KA, Rowin J, Tucker-Lipscomb B, Massey JM, et al. Mycophenolate mofetil for myasthenia gravis: an analysis of efficacy, safety, and tolerability. Neurology. 2003 Nov 25;61(10):1438-1440.
- 48. A trial of mycophenolate mofetil with prednisone as initial immunotherapy in myasthenia gravis. Neurology. 2008 Aug 5;71(6):394-399.
- 49. Sanders DB, Hart IK, Mantegazza R, Shukla SS, Siddiqi ZA, De Baets MHV, et al. An international, phase III, randomized trial of mycophenolate mofetil in myasthenia gravis. Neurology. 2008 Aug 5;71(6):400-406.
- 50. Gajdos P, Chevret S, Clair B, Tranchant C, Chastang C. Clinical trial of plasma exchange and high-dose intravenous immunoglobulin in myasthenia gravis. Myasthenia Gravis Clinical Study Group. Ann. Neurol. 1997 Jun;41(6):789-796.
- 51. Gajdos P, Chevret S, Toyka K. Intravenous immunoglobulin for myasthenia gravis. Cochrane Database Syst Rev. 2003;(2):CD002277.

- 52. Gajdos P, Chevret S, Toyka K. Plasma exchange for myasthenia gravis. Cochrane Database Syst Rev. 2002;(4):CD002275.
- 53. Zinman L, Ng E, Bril V. IV immunoglobulin in patients with myasthenia gravis: a randomized controlled trial. Neurology. 2007 Mar 13;68(11):837-841.
- 54. Illa I, Diaz-Manera J, Rojas-Garcia R, Pradas J, Rey A, Blesa R, et al. Sustained response to Rituximab in anti-AChR and anti-MuSK positive Myasthenia Gravis patients. J. Neuroimmunol. 2008 Sep 15;201-202:90-94.
- 55. Stübgen J. B cell-targeted therapy with rituximab and autoimmune neuromuscular disorders. J. Neuroimmunol. 2008 Nov 15;204(1-2):1-12.
- 56. Lebrun C, Bourg V, Tieulie N, Thomas P. Successful treatment of refractory generalized myasthenia gravis with rituximab. Eur. J. Neurol. 2009 Feb;16(2):246-250.
- 57. Nelson RP, Pascuzzi RM, Kessler K, Walsh LE, Faught PP, Ramanuja S, et al. Rituximab for the treatment of thymoma-associated and de novo myasthenia gravis: 3 cases and review. J Clin Neuromuscul Dis. 2009 Jun;10(4):170-177.
- 58. Gold R, Schneider-Gold C. Current and future standards in treatment of myasthenia gravis. Neurotherapeutics. 2008 Oct;5(4):535-541.
- 59. Sieb JP. Myasthenia gravis: emerging new therapy options. Curr Opin Pharmacol. 2005 Jun;5(3):303-307.
- 60. Soltys J, Kusner LL, Young A, Richmonds C, Hatala D, Gong B, et al. Novel complement inhibitor limits severity of experimentally myasthenia gravis. Ann. Neurol. 2009 Jan;65(1):67-75.
- 61. Bachmann K, Burkhardt D, Schreiter I, Kaifi J, Schurr P, Busch C, et al. Thymectomy is more effective than conservative treatment for myasthenia gravis regarding outcome and clinical improvement. Surgery. 2009 Apr;145(4):392-398.
- 62. Gronseth GS, Barohn RJ. Practice parameter: thymectomy for autoimmune myasthenia gravis (an evidence-based review): report of the Quality Standards Subcommittee of the American Academy of Neurology. Neurology. 2000 Jul 12;55(1):7-15.
- 63. Gronseth G, Barohn R. Thymectomy for Myasthenia Gravis. Curr Treat Options Neurol. 2002 May;4(3):203-209.
- 64. Díaz-Manera J, Rojas-García R, Illa I. Treatment strategies for myasthenia gravis. Expert Opin Pharmacother. 2009 Jun;10(8):1329-1342.

- 65. Meyer DM, Herbert MA, Sobhani NC, Tavakolian P, Duncan A, Bruns M, et al. Comparative clinical outcomes of thymectomy for myasthenia gravis performed by extended transsternal and minimally invasive approaches. Ann. Thorac. Surg. 2009 Feb;87(2):385-390; discussion 390-391.
- 66. Soltys J, Gong B, Kaminski HJ, Zhou Y, Kusner LL. Extraocular muscle susceptibility to myasthenia gravis: unique immunological environment? Ann. N. Y. Acad. Sci. 2008;1132:220-224.
- 67. Kupersmith MJ, Ying G. Ocular motor dysfunction and ptosis in ocular myasthenia gravis: effects of treatment. Br J Ophthalmol. 2005 Oct;89(10):1330-1334.
- 68. Benatar M, Kaminski HJ. Evidence report: the medical treatment of ocular myasthenia (an evidence-based review): report of the Quality Standards Subcommittee of the American Academy of Neurology. Neurology. 2007 Jun 12;68(24):2144-2149.
- 69. Kupersmith M. Ocular myasthenia gravis: treatment successes and failures in patients with long-term follow-up. J. Neurol [Internet]. 2009 Apr 18 [cited 2009 Jul 17]; Available from: http://www.ncbi.nlm.nih.gov/pubmed/19377863
- 70. Benatar M, Kaminski H. Medical and surgical treatment for ocular myasthenia. Cochrane Database Syst Rev. 2006;(2):CD005081.
- 71. Jani-Acsadi A, Lisak RP. Myasthenic crisis: guidelines for prevention and treatment. J. Neurol. Sci. 2007 Oct 15;261(1-2):127-133.
- 72. Rózsa C, Mikor A, Kasa K, Illes Z, Komoly S. Long-term effects of combined immunosuppressive treatment on myasthenic crisis. Eur. J. Neurol [Internet]. 2009 Apr 21 [cited 2009 Jul 17]; Available from: http://www.ncbi.nlm.nih.gov/pubmed/19453406
- 73. Petty R. Lambert Eaton myasthenic syndrome. PRACTICAL NEUROLOGY. 2007 Aug 1;7(4):265-267.
- 74. Sanders DB. Lambert-eaton myasthenic syndrome: diagnosis and treatment. Ann. N. Y. Acad. Sci. 2003 Sep;998:500-508.
- 75. Titulaer MJ, Wirtz PW, Willems LNA, van Kralingen KW, Smitt PAES, Verschuuren JJGM. Screening for small-cell lung cancer: a follow-up study of patients with Lambert-Eaton myasthenic syndrome. J. Clin. Oncol. 2008 Sep 10;26(26):4276-4281.
- 76. Tschernatsch M, Gross O, Kneifel N, Kaps M, Blaes F. SOX-1 autoantibodies in patients with paraneoplastic neurological syndromes. Autoimmun Rev. 2009 Jun;8(7):549-551.
- 77. Oh SJ, Kurokawa K, Claussen GC, Ryan HF. Electrophysiological diagnostic criteria of Lambert-Eaton myasthenic syndrome. Muscle Nerve. 2005 Oct;32(4):515-520.

- 78. Lang B, Vincent A. Autoimmune disorders of the neuromuscular junction. Curr Opin Pharmacol. 2009 Jun;9(3):336-340.
- 79. Maddison P, Newsom-Davis J. Treatment for Lambert-Eaton myasthenic syndrome. Cochrane Database Syst Rev. 2003;(2):CD003279.
- 80. Verschuuren JJGM, Wirtz PW, Titulaer MJ, Willems LNA, van Gerven J. Available treatment options for the management of Lambert-Eaton myasthenic syndrome. Expert Opin Pharmacother. 2006 Jul;7(10):1323-1336.
- 81. Lang B, Pinto A, Giovannini F, Newsom-Davis J, Vincent A. Pathogenic autoantibodies in the lambert-eaton myasthenic syndrome. Ann. N. Y. Acad. Sci. 2003 Sep;998:187-195.
- 82. Illa I. IVIg in myasthenia gravis, Lambert Eaton myasthenic syndrome and inflammatory myopathies: current status. J. Neurol. 2005 May;252 Suppl 1:I14-18.
- 83. Pellkofer HL, Voltz R, Kuempfel T. Favorable response to rituximab in a patient with anti-VGCC-positive Lambert-Eaton myasthenic syndrome and cerebellar dysfunction. Muscle Nerve. 2009 Jul 16;40(2):305-308.
- 84. Sobel J. Botulism. Clin. Infect. Dis. 2005 Oct 15;41(8):1167-1173.
- 85. Shapiro RL, Hatheway C, Swerdlow DL. Botulism in the United States: a clinical and epidemiologic review. Ann. Intern. Med. 1998 Aug 1;129(3):221-228.
- 86. McLauchlin J, Grant KA, Little CL. Food-borne botulism in the United Kingdom. J Public Health (Oxf). 2006 Dec;28(4):337-342.
- 87. Crowner BE, Brunstrom JE, Racette BA. Iatrogenic botulism due to therapeutic botulinum toxin a injection in a pediatric patient. Clin Neuropharmacol. 2007 Oct;30(5):310-313.
- 88. Chertow DS, Tan ET, Maslanka SE, Schulte J, Bresnitz EA, Weisman RS, et al. Botulism in 4 adults following cosmetic injections with an unlicensed, highly concentrated botulinum preparation. JAMA. 2006 Nov 22;296(20):2476-2479.
- 89. Engel A, Shen X, Selcen D, Sine S. What Have We Learned from the Congenital Myasthenic Syndromes. J. Mol. Neurosci [Internet]. 2009 Aug 18 [cited 2009 Oct 11]; Available from: http://www.ncbi.nlm.nih.gov/pubmed/19688192
- 90. Ohno K, Brengman J, Tsujino A, Engel AG. Human endplate acetylcholinesterase deficiency caused by mutations in the collagen-like tail subunit (ColQ) of the asymmetric enzyme. Proc. Natl. Acad. Sci. U.S.A. 1998 Aug 4;95(16):9654-9659.
- 91. Donger C, Krejci E, Serradell AP, Eymard B, Bon S, Nicole S, et al. Mutation in the human

- acetylcholinesterase-associated collagen gene, COLQ, is responsible for congenital myasthenic syndrome with end-plate acetylcholinesterase deficiency (Type Ic). Am. J. Hum. Genet. 1998 Oct;63(4):967-975.
- 92. Ohno K, Tsujino A, Brengman JM, Harper CM, Bajzer Z, Udd B, et al. Choline acetyltransferase mutations cause myasthenic syndrome associated with episodic apnea in humans. Proc. Natl. Acad. Sci. U.S.A. 2001 Feb 13;98(4):2017-2022.
- 93. Sanes JR, Lichtman JW. Induction, assembly, maturation and maintenance of a postsynaptic apparatus. Nat. Rev. Neurosci. 2001 Nov;2(11):791-805.
- 94. Kummer TT, Misgeld T, Sanes JR. Assembly of the postsynaptic membrane at the neuromuscular junction: paradigm lost. Curr. Opin. Neurobiol. 2006 Feb;16(1):74-82.
- 95. Okada K, Inoue A, Okada M, Murata Y, Kakuta S, Jigami T, et al. The muscle protein Dok-7 is essential for neuromuscular synaptogenesis. Science. 2006 Jun 23;312(5781):1802-1805.
- 96. Chevessier F, Faraut B, Ravel-Chapuis A, Richard P, Gaudon K, Bauché S, et al. MUSK, a new target for mutations causing congenital myasthenic syndrome. Hum. Mol. Genet. 2004 Dec 15;13(24):3229-3240.
- 97. Ohno K, Sadeh M, Blatt I, Brengman JM, Engel AG. E-box mutations in the RAPSN promoter region in eight cases with congenital myasthenic syndrome. Hum. Mol. Genet. 2003 Apr 1;12(7):739-748.
- 98. Burke G, Cossins J, Maxwell S, Owens G, Vincent A, Robb S, et al. Rapsyn mutations in hereditary myasthenia: distinct early- and late-onset phenotypes. Neurology. 2003 Sep 23;61(6):826-828.
- 99. Müller JS, Mildner G, Müller-Felber W, Schara U, Krampfl K, Petersen B, et al. Rapsyn N88K is a frequent cause of congenital myasthenic syndromes in European patients. Neurology. 2003 Jun 10;60(11):1805-1810.
- 100. Maselli RA, Dunne V, Pascual-Pascual SI, Bowe C, Agius M, Frank R, et al. Rapsyn mutations in myasthenic syndrome due to impaired receptor clustering. Muscle Nerve. 2003 Sep;28(3):293-301.
- 101. Beeson D, Higuchi O, Palace J, Cossins J, Spearman H, Maxwell S, et al. Dok-7 mutations underlie a neuromuscular junction synaptopathy. Science. 2006 Sep 29;313(5795):1975-1978.
- 102. Palace J, Lashley D, Newsom-Davis J, Cossins J, Maxwell S, Kennett R, et al. Clinical features of the DOK7 neuromuscular junction synaptopathy. Brain. 2007 Jun;130(Pt 6):1507-1515.

- 103. Müller JS, Herczegfalvi A, Vilchez JJ, Colomer J, Bachinski LL, Mihaylova V, et al. Phenotypical spectrum of DOK7 mutations in congenital myasthenic syndromes. Brain. 2007 Jun;130(Pt 6):1497-1506.
- 104. Müller JS, Abicht A, Burke G, Cossins J, Richard P, Baumeister SK, et al. The congenital myasthenic syndrome mutation RAPSN N88K derives from an ancient Indo-European founder. J. Med. Genet. 2004 Aug;41(8):e104.
- 105. Michalk A, Stricker S, Becker J, Rupps R, Pantzar T, Miertus J, et al. Acetylcholine receptor pathway mutations explain various fetal akinesia deformation sequence disorders. Am. J. Hum. Genet. 2008 Feb;82(2):464-476.
- 106. Vogt J, Harrison BJ, Spearman H, Cossins J, Vermeer S, ten Cate LN, et al. Mutation analysis of CHRNA1, CHRNB1, CHRND, and RAPSN genes in multiple pterygium syndrome/fetal akinesia patients. Am. J. Hum. Genet. 2008 Jan;82(1):222-227.
- 107. Maselli RA, Ng JJ, Anderson JA, Cagney O, Arredondo J, Williams C, et al. Mutations in LAMB2 causing a severe form of synaptic congenital myasthenic syndrome. J. Med. Genet. 2009 Mar;46(3):203-208.
- 108. Harper CM, Engel AG. Quinidine sulfate therapy for the slow-channel congenital myasthenic syndrome. Ann. Neurol. 1998 Apr;43(4):480-484.
- 109. Harper CM, Fukodome T, Engel AG. Treatment of slow-channel congenital myasthenic syndrome with fluoxetine. Neurology. 2003 May 27;60(10):1710-1713.
- 110. Bestue-Cardiel M, Sáenz de Cabezón-Alvarez A, Capablo-Liesa JL, López-Pisón J, Peña-Segura JL, Martin-Martinez J, et al. Congenital endplate acetylcholinesterase deficiency responsive to ephedrine. Neurology. 2005 Jul 12;65(1):144-146.
- 111. Mihaylova V, Müller JS, Vilchez JJ, Salih MA, Kabiraj MM, D'Amico A, et al. Clinical and molecular genetic findings in COLQ-mutant congenital myasthenic syndromes. Brain. 2008 Mar;131(Pt 3):747-759.
- 112. Selcen D, Milone M, Shen X, Harper CM, Stans AA, Wieben ED, et al. Dok-7 myasthenia: phenotypic and molecular genetic studies in 16 patients. Ann. Neurol. 2008 Jul;64(1):71-87.

Figure legend

Figure 1.

Key steps in ACh synthesis and release at the neuromuscular junction and the core pathway responsible for ACh receptor clustering. AcCoA, Acetyl Coenzyme A; AChE, Acetylcholinesterase; ChAT, Choline Acetyltransferase; Dok-7, Downstream of tyrosine kinase 7; ColQ, AChE collagen-like tail subunit; LRP4, low density lipoprotein receptor-related protein 4; MuSK, Muscle-Specific Kinase; SNARE, Soluble NSF Attachment protein Receptor.



