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Myeloidderived suppressor cells: Escorts at the maternal– fetal interface

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Myeloid-derived suppressor cells (MDSCs) are a novel heterogenous group of immunosuppressive cells derived from myeloid progenitors. Their role is well known in tumors and autoimmune diseases. In recent years, the role and function of MDSCs during reproduction have attracted increasing attention. Improving the understanding of their strong association with recurrent implantation failure, pathological pregnancy, and neonatal health has become a focus area in research. In this review, we focus on the interaction between MDSCs and other cell types (immune and non-immune cells) from embryo implantation to postpartum. Furthermore, we discuss the molecular mechanisms that could facilitate the therapeutic targeting of MDSCs. Therefore, this review intends to encourage further research in the field of maternal–fetal interface immunity in order to identify probable pathways driving the accumulation of MDSCs and to effectively target their ability to promote embryo implantation, reduce pathological pregnancy, and increase neonatal health.

KEYWORDS

myeloid-derived suppressor cells (MDSCs), maternal-fetal immune tolerance, embryo implantation, pregnancy complications, newborn

1 Introduction

Recently, a population of immature cells derived from myeloid progenitors, i.e., myeloidderived suppressor cells (MDSCs), has entered the research spotlight (1). The molecular phenotypes of MDSCs are CD11b and Gr1 in mice, whereas it is HLA-DR^{low/-}CD33⁺ in humans. These can be subdivided into the monocytic and granulocytic subtypes (M-MDSCs and G-MDSCs, respectively). Traditionally, they are defined as CD11b⁺Ly6C^{high}Ly6G⁻ and CD11b⁺Ly6G⁺Ly6C^{low} in mice and CD11b⁺CD33⁺CD14⁺HLA-DR^{-/low} and CD11b⁺CD33⁺CD15⁺/CD66⁺HLA-DR⁻ in humans, respectively (2). Recently, a number of novel markers and subsites of MDSCs have emerged, as shown in Table 1. MDSCs exhibit significant heterogeneity because M-MDSCs and G-MDSCs can further differentiate into myeloid cells. Furthermore, it is difficult to distinguish them from mature monocytes and granulocytes based on immune molecular markers (30). In addition, a great deal of attention has been focused on the immunosuppressive function of MDSCs in tumors, autoimmune diseases, organ transplantation, and infectious diseases (31–34). In tumors, MDSCs inhibit the antitumor functions of T cells and natural killer (NK) cells and promote tumor progression by accelerating angiogenesis and cell invasion (2, 35, 36). Moreover, the proportion of MDSCs is closely associated with the clinical outcomes and therapeutic effects in patients with cancer (37). Although MDSCs are short-lived cells, they play crucial roles in many diseases.

The function of MDSCs during reproduction remains controversial. Embryo implantation, including the balance of immune cells at the maternal-fetal interface, reconstruction of the endometrial spiral arteries, and moderate invasion of trophoblasts, is a prerequisite for a successful pregnancy. Several studies have described how maternal-fetal tolerance is enhanced by accumulating MDSCs to suppress T-cell responses (38-40). Furthermore, the proportion of MDSCs was decreased in spontaneously aborting mice compared with controls, whereas their depletion resulted in the increased cytotoxicity of decidual NK cells (41). Even in infertile couples, the ratio of MDSCs in the peripheral blood of women who undergo in vitro fertilization (IVF) is considered an essential factor in predicting the outcomes of pregnancy (42, 43). These findings might provide new insights into immune-related miscarriage and IVF failure. In addition, several studies have further demonstrated the accumulation and augmentation of MDSCs in newborn humans and mice (44-46). There is growing acceptance that MDSCs manipulate the maternalfetal immune microenvironment to promote and sustain embryo implantation, protect the fetus during gestation, and keep the newborn healthy.

In this review, we discuss the evidence concerning the association of MDSCs with embryo implantation, pregnancy complications, and newborn health and explore the potential relationship between MDSCs and the maternal-fetal immune microenvironment.

2 MDSCs in embryo implantation

The first steps to a successful pregnancy are embryo localization, adhesion, and invasion of the endometrium. As a type of semiallogeneic transplantation, the fetal trophoblasts in the maternal–fetal interface interact with the maternal decidual stromal cells, decidual glandular epithelial cells, and numerous immune cells (47, 48). The maternal–fetal immune microenvironment plays a decisive role during these events. A balance between the pro- and antiinflammatory cells (T cells, NK cells, and MDSCs) is crucial for successful implantation and placentation (40, 49–53). In addition to protecting the fetus from maternal immune attacks, immune cells play critical roles in the migration and invasion of trophoblasts and in forming decidual blood vessels (54). The maternal corpus luteum and fetal placenta can signal the production of high levels of estrogen and progesterone (55), and interactions involving hormones and immune cells cannot be ignored.

2.1 Immunosuppressive function of MDSCs

Immunotolerance is vital in the maternal immune system in order to accept the embryo. As a population of novel immunoregulatory cells, MDSCs have received increasing attention during early pregnancy rather than in cancer and autoimmune diseases. Besides investigations into the frequency and induction of MDSCs in pregnant human and mouse models, studies have also focused on understanding their immunosuppressive effects during pregnancy. G-MDSCs and M-MDSCs exert immunosuppressive functions via different molecular mechanisms (56). M-MDSCs function primarily through the secretion of nitric oxide and arginase-1 (Arg-1) (57), whereas G-MDSCs produce reactive oxygen species (ROS) and hydrogen peroxide (58). Cytokines, growth factors, and microorganisms enhance the immunosuppressive functions of MDSCs in tumors or the inflammatory microenvironment through mediators and subsequently activate nuclear factor kappa B, signal transducer and activator of transcriptions (STATs), and other signaling pathways (59–61).

An earlier study reported that G-MDSCs comprise the dominant MDSC subset in the peripheral blood of pregnant women, producing immunosuppressive enzymes such as Arg-1 and inducible nitric oxide synthase (iNOS) (62). In an MDSC depletion pregnancy mouse model, T cells showed higher proliferation capacity without Arg-1 and ROS (63). In our preliminary research, the T cells in the peripheral blood of infertile patients exhibited higher immunosuppressive function than those in the MDSC-depleted group (43). Furthermore, it has been suggested that MDSCs reduce the T-cell responses in a cell-cell contact manner (40). In humans, a subpopulation of decidual MDSCs was first recognized by Bartmann et al. in 2015. These cells secrete high levels of immunosuppressive products, including Arg-1, iNOS, and indoleamine-2,3-dioxygenase (IDO), and produce anti-inflammatory cytokines, such as interleukin-10 (IL-10) and transforming growth factor beta (TGF- β), with the ability to inhibit T-cell proliferation (64). Although the relationship between endometriosis and embryo implantation has received increasing attention, the underlying molecular mechanisms remain unclear. Elevated G-MDSC counts positively regulate the immunosuppressive activity (increased Arg-1 and ROS levels and suppressed T-cell proliferation) and the increased number of endometrial lesions. However, MDSCs depleted with the anti-Gr-1 antibody in mice exhibited dramatically fewer lesions (65). This indicates that MDSCs could possibly reduce endometrial receptivity so as to affect embryo implantation through immune attack.

As a form of semi-allogeneic transplantation, the embryo is considered as a foreign antigen by the maternal immune system; therefore, appropriate immunosuppressive products are essential in inducing immune tolerance.

2.2 MDSCs shift T-cell differentiation

The maternal immune system is a complex immune paradox. The mother is sensitized by foreign embryos in early pregnancy under

TABLE 1 Phenotypes of myeloid-derived suppressor cells (MDSCs) in mice and humans.

		G-MDSCs		M-MDSCs		Other types of MDSCs	Reference
	Mice	Human	Mice	Human	Mice	Human	
Traditional	CD11b ⁺ Ly6G ⁺ Ly6C ^{low}		CD11b ⁺ Ly6G ⁻ Ly6C ^{high}				(2-4)
markers		CD11b ⁺ CD33 ⁺ CD14 ⁻ CD15 ⁺ CD66 ⁺ HLA- DR ^{low/-}		CD11b ⁺ CD33 ⁺ CD14 ⁺ CD15 ⁻ CD66 ⁻ HLA- DR ^{low/-}			(2, 4–6)
Novel	Side scatter (SSC)						(7, 8)
markers	CD244						(9)
		CD84					(10)
		CD36					(11)
		FATP2					(12)
		CD13					(13, 14)
		CD16					(15)
		LOX1					(16)
			CD49d (VLA4)				(17)
			CD115				(18)
				CCR2			(19)
				CD11c			(20, 21)
				IL-4Rα (CD124)			(22)
				CD68 ⁺ CD163 ⁺			(23)
					Early-stage MDSCs (e- MDSCs) CD11b ⁺ Gr- 1 ⁻ F4/ 80 ⁻ MHCII ⁻		(24)
						e-MDSCs Lin ⁻ CD3 ⁻ CD14 ⁻ CD15 ⁻ CD19 ⁻ CD56 ⁻ CD33 ⁺ HLA- DR ⁻	(25)
						MDSC-like fibrocytes Collagen type-1 ⁺ CD15 ⁺ CD45 ^{dim} CD34 ⁻ CD14 ⁻ / CD11c ^{hi} CD123 ⁻ CD14 ⁻	(26, 27)
					Fibrocytic- MDSCs (f- MDSCs)		(28)

estrogen and progesterone stimulation. Subsequently, immune
tolerance is induced to protect the fetus from maternal immune
attack. In recent years, the research on maternal-fetal immunity has
focused on the balance between T helper (Th) 1/Th2 cells and Th17/
regulatory T cells (Tregs) (66-68). The hyperfunction of pro-
inflammatory T cells, such as Th1 and Th17, has been confirmed to
be associated with recurrent implantation failure (69, 70).

It is well documented that MDSCs in the decidua isolated from pregnant women can induce the expression of forkhead box P3 (Foxp3) in CD4⁺ T cells (Tregs) through the TGF- β/β -catenin signaling pathway. Furthermore, CD4⁺Foxp3⁺ T cells are associated with increased MDSC number and recruitment, resulting in a positive feedback loop (71). In addition, MDSCs have been reported to induce a shift toward the anti-inflammatory subtype of Th2 cells via cell-cell interactions (72). Exosomes released by G-MDSCs in pregnant women have the ability to suppress T-cell proliferation, polarize Th cells to Th2 and Tregs, and inhibit lymphocyte cytotoxicity (73). Although the exact reason remains unclear, the reduction of L-selectin expression in naive T cells has been considered to inhibit their trafficking toward lymph nodes (38). Reducing the frequency of MDSCs leads to Tcell responses that enhance and elevate the ratio of Th1 to total T cells in pregnant women (74). In addition to T cells, multiple types of effector immune cells, including macrophages, uterine natural killer (uNK) cells, and immature dendritic cells (iDCs), play substantial roles in the induction of maternal-fetal immunotolerance (39, 75, 76).

2.3 MDSCs affect NK cell cytotoxicity

NK cells represent one of the dominant innate lymphoid cell types that exert cytotoxic effects and primarily contribute to the killing of pathogen-infected cells and tumor cells (77). These cells can primarily be categorized into two subtypes: peripheral blood NK (pbNK) cells and tissue-resident NK (trNK) cells. pbNK cells are identified as CD3⁻CD56⁺ in humans, whereas trNK cells exhibit various phenotypes and signatures (78, 79). Therefore, the CD45⁺CD56⁺Lin⁻ cells in humans or the CD45⁺Lin⁻NK1.1⁺NKp46⁺ cells in mice are defined as uNK cells. Owing to the unique structure of the uterus, uNK cells are mostly distributed in the endometrium. uNK cells are also known as decidual NK cells (80). In contrast to the cytotoxicity in pbNK cells, uNK cells facilitate decidualization and placenta formation by remodeling the extracellular matrix and endometrial stroma vessels (81).

NK cells can trigger targeted cell death by releasing cytotoxic granules, such as granzymes and perforin. Furthermore, NK cells are also employed to recognize and eliminate target cells through the NK group protein 2D-activating NK receptor (NKG2C). A study on mouse models identified that MDSC depletion results in the upregulation of the embryo resorption rates and a decrease in the uNK cell counts. Furthermore, MDSC depletion increased the cytotoxicity of uNK cells by upregulating perforin and granzyme B in the cytoplasm and NKG2C on the cell surface (38, 41, 63). It cannot be ignored that MDSCs also represent something of a double-edged sword during pregnancy because of the immunosuppressive function of pbNK cells. In a pregnant mouse

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		G-MDSCs		M-MDSCs		Other types of MDSCs	Reference
	Mice	Human	Mice	Human	Mice	Human	
					CD11b ⁺ Gr- 1 ⁺ FAP ⁺		
						$f-MDSCs$ CD 33 ⁺ IL -4R α^+	(28, 29)
G-MDSCs, granul	ocytic myeloid-derived suppre	essor cells; M-MDSCs, monocytic myeloid-deri	ved suppressor cells.				

model with breast cancer, MDSC accumulation led to the inhibition of NK cell activity and the subsequent promotion of tumor metastasis and disease progression (82).

2.4 MDSCs promote angiogenesis

Epidemiological and experimental research confirmed that an imbalance in circulating angiogenic factors directly leads to embryo implantation failure and maternal pregnancy complications, such as preeclampsia (83, 84). Although several potential cellular immunities associated with MDSCs at the maternal-fetal interface have been identified, MDSCs are related to vascular remodeling during embryo implantation and placentation. For instance, it is well documented that MDSCs support angiogenesis in tumor environments (36, 85, 86). Adequate invasion of extravillous trophoblasts into the endometrium with dilated and reconstructed spiral arteries in the myometrium so as to maintain an adequate blood supply is a prerequisite for a successful pregnancy. It has been shown that MDSCs play crucial roles during placental formation, similar to that during tumorigenesis (87). However, there is no direct evidence suggesting that MDSCs promote angiogenesis during human placentation. In rats, the MDSC-derived vascular endothelial growth factor (VEGF) has been confirmed to promote the development of maternal uterine spiral arteries and placenta to elevate the reception of implantation (88). In patients undergoing IVF, elevated serum VEGF levels were positively related to the MDSC ratio. Furthermore, the VEGF level and MDSC ratio positively correlated with the pregnancy rates (43). In addition to VEGF, MDSCs have also been discovered to secrete various bioactive factors, including CXC chemokine receptor 2 (CXCR2), CXCR4, IL-6, TGF- β , and metalloproteinases, that could facilitate tumor migration and metastasis and promote angiogenesis (36). G-MDSCs in the decidua with a high expression of CXCR2 have also been found at the maternal-fetal interface and supposedly induced angiogenesis (89, 90). In patients with spontaneous miscarriage, VEGF activated the VEGF or MEK/ERK signaling pathways in the villi to govern uterine angiogenesis and vascular remodeling during pregnancy (91, 92). In general, promoting angiogenesis may be another role of MDSCs in promoting embryo implantation.

2.5 Mutual promotion of MDSCs and trophoblasts

The embryo must experience the cleavage stage and the blastocyst stage and finally implant into the endometrium. MDSCs are more abundant in the peripheral blood and decidua of pregnant women and mice than in those of non-pregnant individuals (74, 81). In one of our preliminary experiments to investigate the relationship between MDSCs and trophoblasts, we showed that the proportion of MDSCs in the peripheral blood of IVF patients was positively correlated with pregnancy outcomes (43, 62). These findings suggest that MDSCs enhance trophoblast cell activity. Trophoblasts are also effective players in the expansion of

MDSCs. An *in vitro* study using a trophoblast cell line (HTR8/ SVneo) reported the successful triggering of peripheral CD14⁺ myelomonocytic cell differentiation into MDSCs with high expression of IDO1 and Arg-1. Notably, these cells also expressed higher levels of STAT3, indicating the potential mechanisms underlying trophoblast-induced MDSC differentiation (93). Together, these findings reveal the positive feedback loop of trophoblast implantation and MDSC recruitment at the maternalfetal interface.

2.6 Expansion and differentiation of MDSCs under conditions of hormone stimulation

During gestation, the maternal corpus luteum and fetal placenta can relay to produce high levels of estrogen and progesterone (55), and the interactions between hormones and MDSCs cannot be ignored. The MDSC count and frequency steadily increase during pregnancy and peak in the second trimester, showing parallel changes with the circulating estradiol and progesterone levels. Exosomes isolated from the peripheral blood MDSCs of pregnant women exhibited various abilities involving differentiation and suppressive functions according to the stage of gestation (73). Growing evidence has shown that the recruitment and the accumulation of MDSCs depend on estradiol signaling in cancer patients (94). During human gestation, high estradiol and progesterone levels considerably facilitate the ratio and suppressive functions in both G-MDSCs and M-MDSCs via the STAT3 signaling pathway (40, 95). Estradiol alone may induce an increased VEGF expression in MDSCs to promote maternal uterine spiral arteries and placental development in rats (88, 96). Similar to the above-mentioned hormones, human leukocyte antigen G (HLA-G), a molecule secreted and expressed by several cells in the maternal-fetal interface, is also a crucial player in MDSC accumulation in the decidua (97). It usually binds to immunoglobulin-like transcript 4 and induces the expansion and differentiation of MDSCs in peripheral blood mononuclear cells through STAT3 signaling (98, 99). A recent study has shown that the estradiol receptor negatively modulated the expression of hypoxia-inducible factor-1 alpha (HIF-1 α) (100). HIF-1 α deficiency at the maternal-fetal interface leads to a decrease in the MDSC counts. Furthermore, silencing the expression of HIF-1 α in MDSCs results in an impaired immunosuppressive function and renders them susceptible to apoptosis (101). Therefore, hormones protect and ensure embryo implantation and regulate the function of MDSCs through multiple pathways.

2.7 Lipid metabolism of MDSCs

Cellular energy metabolism is currently a critical area of focus in scientific research. Understanding the role of energy metabolism could help in better comprehending the fate of MDSCs during gestation or in pathological processes. An association between MDSCs and lipid metabolism was first recognized in 2011 by Xia et al. (102). The authors identified that MDSCs accumulated in the fat tissue, the liver, and peripheral blood, with pro-inflammation and reestablishment of metabolic homeostatic functions, even increasing insulin resistance. In contrast, the adoptive transfer of MDSCs reduced obesity-associated inflammation and improved insulin sensitivity (103). In humans, the M-MDSC counts were higher in the peripheral blood of obese men compared to those in the control group (104). Mechanistically, excessive adipocytes secreted pro-inflammatory mediators, including prostaglandin E2, tumor necrosis factor alpha (TNF- α), interleukin 6 (IL-6), and IL- 1β (105), and MDSCs were recruited to the adipose tissue, subsequently resulting in a substantially worse immune response cascade (106-108). Dyslipidemia may also lead to the accumulation and immunosuppressive capacity enhancement of MDSCs, indicating that exogenous lipids may be taken up by MDSCs (11). In patients experiencing unexplained recurrent pregnancy loss, G-MDSC differentiation was associated with the pSTAT3/FABP5/ PGE2 pathway (109). These findings are instrumental in understanding the physiological mechanisms of immune tolerance in pregnancy.

3 MDSCs during pregnancy

Recently, the numbers of studies investigating the role of MDSCs during pregnancy have increased. MDSCs are accumulated in a time-dependent manner during gestation (62, 74). Abnormal pregnancies and pregnancy complications, including spontaneous miscarriage, intrauterine growth restriction (IUGR), and preeclampsia (PE), are also closely related to an imbalance in the maternal-fetal immunological tolerance (51, 110, 111). As reviewed and discussed previously, MDSC deficiency may participate in such adverse events.

3.1 MDSC deficiency leads to spontaneous miscarriage

Spontaneous miscarriage is the loss of a pregnancy before viability with no manual intervention. This condition occurs in up to 20% of recognized pregnancies (109). As listed above, there is a reciprocal causation between MDSC deficiency and spontaneous miscarriage in the first or the second trimester. A reduction in the MDSC ratio, accompanied by a decrease in the levels of Arg-1, iNOS, IL-10, and TGF-B, has been observed in a spontaneous abortion mouse model compared to controls (41). The proportion of MDSCs was significantly increased in the decidua of early pregnant women compared to non-pregnant women (71, 112). Decidual G-MDSC apoptosis is mediated by TNF-related apoptosis-induced ligand (TRAIL) in a caspase 3-dependent manner. Notably, downregulated expression of the TRAIL receptor DcR2 leads to increased G-MDSC apoptosis, and this is responsible for spontaneous miscarriage (113). Generally, high levels of estradiol and progesterone lead to the recruitment of MDSCs, and an increase in their counts, during gestation. MDSCs

manipulate the outcomes of pregnancy by producing immunosuppressive molecules (62, 74), regulating immune tolerance (96, 114), promoting angiogenesis (95), and inducing trophoblast implantation (40, 63).

3.2 MDSC deficiency results in pregnancy complications

IUGR represents an abnormal pregnancy and is the second leading cause of perinatal morbidity and mortality, affecting 5% of pregnancies (114). IUGR is one of the major causes of spontaneous miscarriage, prematurity, stillbirth, and even infant mortality (115). On the other hand, PE is characterized by new-onset hypertension, usually occurring after 20 weeks of gestation and affecting 3%-8% of pregnancies (116). The causes of IUGR and PE are multifactorial, including maternal age and nutrition, thrombophilia, placental dysfunction, and fetal factors (117). Placental dysfunction includes uteroplacental insufficiency, vessel thrombosis, placenta or spiral artery arteritis, chronic villitis, and umbilical cord abnormalities (118, 119). There is still a lack of IUGR and PE animal models; therefore, investigations have been primarily conducted in humans. As mentioned previously, G-MDSCs are observed in all stages of pregnancy and are accompanied by high cellular levels of Arg-1 and ROS activity; however, patients with IUGR exhibit a significantly lower suppressive activity. Moreover, the frequency of G-MDSCs is negatively correlated with adverse outcomes in newborns from pregnancies with IUGR (120). The MDSC ratio and serum Arg-1 levels in patients with PE are much lower than those in healthy pregnant women (111). Furthermore, the frequency of T-cell immunoglobulin and mucin domain-containing protein 3 (TIM3)⁺ MDSCs is also more elevated in patients with PE than in healthy controls, and the Tim-3/galectin-9 pathway is considered to modulate the function of MDSCs so as to affect the pathogenesis of PE (121). We speculate that adverse pregnancy outcomes associated with IUGR and PE may be related to immune imbalance, angiogenesis disorders, and shallow placental implantation caused by MDSC deficiency during early pregnancy.

4 MDSCs in postpartum and newborns

MDSCs play an immunosuppressive role in tumors (122, 123) and a pro-inflammatory role in autoimmune diseases (124, 125). Similarly, there is a shift from the early anti-inflammatory profiles that contribute to embryo implantation to a late pro-inflammatory cytokine profile that promotes fetal delivery (126). Notably, the MDSC counts decline rapidly in the peripheral blood of women after parturition (62). A transcriptome study that compared pregnancy-induced MDSCs to tumor-induced MDSCs revealed similar transcriptomes in both conditions. However, the antimicrobial-associated protein levels were higher in pregnancy-induced MDSCs (127). Furthermore, the MDSCs in neonates exhibited antimicrobial activity by secreting higher levels of cathepsin G, neutrophil cytosolic factor 1, S100 calcium-binding protein A8/A9, myeloperoxidase, lysozyme, neutrophil elastase, and lipocalin 2 (127). Similarly, the MDSCs in breast milk have also been found to protect infants from necrotizing enterocolitis and nosocomial bacterial infections (128). It appears that the MDSCs transferred from the mother to the fetus during delivery *via* the umbilical cord and breast milk harbor antimicrobial effects in infants.

In addition to this protective function, MDSCs may also play a negative role in postnatal immune development, increasing susceptibility to infections in neonates. Estradiol levels are high in newborns, and there is estradiol-dominant MDSC recruitment and augmentation. Among infants, preterm babies show elevated estradiol levels in serum compared to full-term babies, while the MDSCs in umbilical cord blood also exhibit an abnormal increase (129). Premature infants are more susceptible to necrotizing enterocolitis, bronchopulmonary dysplasia, and sepsis (44, 127, 128). The underlying mechanisms may be associated with immune regulation activity and the immunosuppressive characteristics of MDSCs.

During the neonatal period, a balance between immune attack and tolerance is extremely important (130). Generally, the immunosuppressive functions of umbilical cord blood MDSCs are to inhibit the differentiation and proliferation of Th1 and Th17 cells by producing Arg-1 and ROS and inducing cell-cell contact and to enhance the accumulation of immune regulatory Th2 cells and Tregs (131–134). Furthermore, MDSCs also suppress the cytotoxic NK cells through cell-cell interactions (131). When an anti-inflammatory immune reaction is activated (represented by Th2 and Treg hyperfunction), the newborn will not exhibit excessive immune attack when exposed to various antigens, bacteria, and viruses. However, in preterm infants, the high frequency of MDSCs enhances immune tolerance, leading to insufficient immune responses and neonatal diseases (46, 134–136).

5 Conclusion

The immune environment at the maternal-fetal interface is complex and mysterious. MDSCs, as immunoregulatory cells with both pro-inflammatory and anti-inflammatory functions, are well documented to act as an escort throughout pregnancy (schematic diagram shown in Figure 1). Identifying the molecular mechanisms underlying the angiogenesis promotion and trophoblast implantation-inducing functions of these cells will enable precision therapy. During the window of implantation, intra-uterus transfusion of induced MDSCs or augment MDSCs in the decidua using multiple cytokines may provide tangible clinical benefits, such as elevate the live birth rate in recurrent embryo implantation failure, decrease pregnancy complications, and



FIGURE 1

Schematic diagram. Myeloid-derived suppressor cells (MDSCs) secrete high levels of immunosuppressive products, including arginase-1 (Arg-1), reactive oxygen species (ROS), inducible nitric oxide synthase (iNOS), and indoleamine-2,3-dioxygenase (IDO), and produce anti-inflammatory cytokines, such as interleukin 10 (IL-10) and transforming growth factor beta (TGF- β), with the ability to inhibit T-cell proliferation. In addition, MDSCs shift the T helper (Th) 1/Th2 cell and Th17/regulatory T cell (Treg) subsets to exert immunoregulation function. They also enhance uterine natural killer (uNK) cells to facilitate decidualization and placenta formation by downregulating perforin and granzyme B in the cytoplasm and NK group protein 2 D-activating NK receptor (NKG2C) on the cell surface and promote angiogenesis and placental implantation through the VEGF and TIM3 pathway. During pregnancy, high levels of estrogen and trophoblasts may recruit, accumulate, or even induce the differentiation of MDSCs. MDSCs deficiency is considered to be associated with embry implantation failure, spontaneous miscarriage, intrauterine growth restriction, and preeclampsia. Furthermore, MDSCs even play crucial roles to enhance immune tolerance in neonates.

promote neonatal health. However, whether immunosuppression during pregnancy promotes the occurrence and progression of tumors remains uncertain. Further studies focusing on this aspect still need to be conducted.

Author contributions

BP, CH, HL, XN, and KW wrote the manuscript. CZ and HY revised the manuscript. All authors contributed to the article and approved the submitted version.

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References

1. Parker KH, Beury DW, Ostrand-Rosenberg S. Myeloid-derived suppressor cells: Critical cells driving immune suppression in the tumor microenvironment. *Adv Cancer Res* (2015) 128:95-139. doi: 10.1016/bs.acr.2015.04.002

2. Hu C, Pang B, Lin G, Zhen Y, Yi H. Energy metabolism manipulates the fate and function of tumour myeloid-derived suppressor cells. *Br J Cancer* (2020) 122(1):23–9. doi: 10.1038/s41416-019-0644-x

3. Li Q, Pan PY, Gu P, Xu D, Chen SH. Role of immature myeloid gr-1+ cells in the development of antitumor immunity. *Cancer Res* (2004) 64(3):1130–9. doi: 10.1158/0008-5472.can-03-1715

4. Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. Nat Rev Immunol (2009) 9(3):162-74. doi: 10.1038/nri2506

5. Almand B, Clark JI, Nikitina E, van Beynen J, English NR, Knight SC, et al. Increased production of immature myeloid cells in cancer patients: A mechanism of immunosuppression in cancer. *J Immunol* (2001) 166(1):678–89. doi: 10.4049/ jimmunol.166.1.678

6. Schmielau J, Finn OJ. Activated granulocytes and granulocyte-derived hydrogen peroxide are the underlying mechanism of suppression of t-cell function in advanced cancer patients. *Cancer Res* (2001) 61(12):4756–60.

7. Damuzzo V, Pinton L, Desantis G, Solito S, Marigo I, Bronte V, et al. Complexity and challenges in defining myeloid-derived suppressor cells. *Cytometry B Clin Cytom* (2015) 88(2):77–91. doi: 10.1002/cyto.b.21206

8. Gabrilovich DI, Ostrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. *Nat Rev Immunol* (2012) 12(4):253-68. doi: 10.1038/nri3175

9. Youn JI, Collazo M, Shalova IN, Biswas SK, Gabrilovich DI. Characterization of the nature of granulocytic myeloid-derived suppressor cells in tumor-bearing mice. *J Leukoc Biol* (2012) 91(1):167–81. doi: 10.1189/jlb.0311177

10. Alshetaiwi H, Pervolarakis N, McIntyre LL, Ma D, Nguyen Q, Rath JA, et al. Defining the emergence of myeloid-derived suppressor cells in breast cancer using single-cell transcriptomics. *Sci Immunol* (2020) 5(44):eaay6017. doi: 10.1126/sciimmunol.aay6017

 Al-Khami AA, Zheng L, Del Valle L, Hossain F, Wyczechowska D, Zabaleta J, et al. Exogenous lipid uptake induces metabolic and functional reprogramming of tumorassociated myeloid-derived suppressor cells. *Oncoimmunology* (2017) 6(10):e1344804. doi: 10.1080/2162402X.2017.1344804

12. Veglia F, Tyurin VA, Blasi M, De Leo A, Kossenkov AV, Donthireddy L, et al. Fatty acid transport protein 2 reprograms neutrophils in cancer. *Nature* (2019) 569(7754):73–8. doi: 10.1038/s41586-019-1118-2

13. Zhang J, Xu X, Shi M, Chen Y, Yu D, Zhao C, et al. CD13(hi) neutrophil-like myeloid-derived suppressor cells exert immune suppression through arginase 1 expression in pancreatic ductal adenocarcinoma. *Oncoimmunology* (2017) 6(2): e1258504. doi: 10.1080/2162402x.2016.1258504

14. Resheq YJ, Menzner AK, Bosch J, Tickle J, Li KK, Wilhelm A, et al. Impaired transmigration of myeloid-derived suppressor cells across human sinusoidal endothelium is associated with decreased expression of CD13. *J Immunol* (2017) 199(5):1672–81. doi: 10.4049/jimmunol.1600466

15. Lang S, Bruderek K, Kaspar C, Höing B, Kanaan O, Dominas N, et al. Clinical relevance and suppressive capacity of human myeloid-derived suppressor cell subsets. *Clin Cancer Res* (2018) 24(19):4834–44. doi: 10.1158/1078-0432.Ccr-17-3726

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16. Condamine T, Dominguez GA, Youn JI, Kossenkov AV, Mony S, Alicea-Torres K, et al. Lectin-type oxidized LDL receptor-1 distinguishes population of human polymorphonuclear myeloid-derived suppressor cells in cancer patients. *Sci Immunol* (2016) 1(2):aaf8943. doi: 10.1126/sciimmunol.aaf8943

17. Haile LA, Gamrekelashvili J, Manns MP, Korangy F, Greten TF. CD49d is a new marker for distinct myeloid-derived suppressor cell subpopulations in mice. *J Immunol* (2010) 185(1):203–10. doi: 10.4049/jimmunol.0903573

18. Geissmann F, Auffray C, Palframan R, Wirrig C, Ciocca A, Campisi L, et al. Blood monocytes: distinct subsets, how they relate to dendritic cells, and their possible roles in the regulation of T-cell responses. *Immunol Cell Biol* (2008) 86(5):398–408. doi: 10.1038/ icb.2008.19

19. Lesokhin AM, Hohl TM, Kitano S, Cortez C, Hirschhorn-Cymerman D, Avogadri F, et al. Monocytic CCR2(+) myeloid-derived suppressor cells promote immune escape by limiting activated CD8 T-cell infiltration into the tumor microenvironment. *Cancer Res* (2012) 72(4):876–86. doi: 10.1158/0008-5472.Can-11-1792

20. Roussel M, Ferrell PBJr., Greenplate AR, Lhomme F, Le Gallou S, Diggins KE, et al. Mass cytometry deep phenotyping of human mononuclear phagocytes and myeloidderived suppressor cells from human blood and bone marrow. *J Leukoc Biol* (2017) 102 (2):437–47. doi: 10.1189/jlb.5MA1116-457R

21. Zhao T, Liu S, Ding X, Johnson EM, Hanna NH, Singh K, et al. Lysosomal acid lipase, CSF1R, and PD-L1 determine functions of CD11c+ myeloid-derived suppressor cells. JCI Insight (2022) 7(17):e156623. doi: 10.1172/jci.insight.156623

22. Mandruzzato S, Solito S, Falisi E, Francescato S, Chiarion-Sileni V, Mocellin S, et al. IL4Ralpha+ myeloid-derived suppressor cell expansion in cancer patients. *J Immunol* (2009) 182(10):6562–8. doi: 10.4049/jimmunol.0803831

23. Shao X, Wu B, Cheng L, Li F, Zhan Y, Liu C, et al. Distinct alterations of CD68(+) CD163(+) M2-like macrophages and myeloid-derived suppressor cells in newly diagnosed primary immune thrombocytopenia with or without CR after high-dose dexamethasone treatment. *J Transl Med* (2018) 16(1):48. doi: 10.1186/s12967-018-1424-8

24. Zhang W, Jiang M, Chen J, Zhang R, Ye Y, Liu P, et al. SOCS3 suppression promoted the recruitment of CD11b(+)Gr-1(-)F4/80(-)MHCII(-) early-stage myeloid-derived suppressor cells and accelerated interleukin-6-Related tumor invasion *via* affecting myeloid differentiation in breast cancer. *Front Immunol* (2018) 9:1699. doi: 10.3389/fimmu.2018.01699

25. Khan ANH, Emmons TR, Wong JT, Alqassim E, Singel KL, Mark J, et al. Quantification of early-stage myeloid-derived suppressor cells in cancer requires excluding basophils. *Cancer Immunol Res* (2020) 8(6):819–28. doi: 10.1158/2326-6066.Cir-19-0556

26. Wright AK, Newby C, Hartley RA, Mistry V, Gupta S, Berair R, et al. Myeloidderived suppressor cell-like fibrocytes are increased and associated with preserved lung function in chronic obstructive pulmonary disease. *Allergy* (2017) 72(4):645–55. doi: 10.1111/all.13061

27. Zhang H, Maric I, DiPrima MJ, Khan J, Orentas RJ, Kaplan RN, et al. Fibrocytes represent a novel MDSC subset circulating in patients with metastatic cancer. *Blood* (2013) 122(7):1105–13. doi: 10.1182/blood-2012-08-449413

28. Mu X, Wu K, Zhu Y, Zhu Y, Wang Y, Xiao L, et al. Intra-arterial infusion chemotherapy utilizing cisplatin inhibits bladder cancer by decreasing the fibrocytic

myeloid-derived suppressor cells in an m6A-dependent manner. *Mol Immunol* (2021) 137:28–40. doi: 10.1016/j.molimm.2021.06.012

29. Zoso A, Mazza EM, Bicciato S, Mandruzzato S, Bronte V, Serafini P, et al. Human fibrocytic myeloid-derived suppressor cells express IDO and promote tolerance *via* tregcell expansion. *Eur J Immunol* (2014) 44(11):3307–19. doi: 10.1002/eji.201444522

30. Hegde S, Leader AM, Merad M. MDSC: Markers, development, states, and unaddressed complexity. *Immunity* (2021) 54(5):875-84. doi: 10.1016/j.immuni.2021.04.004

31. Gabrilovich DI. Myeloid-derived suppressor cells. Cancer Immunol Res (2017) 5 (1):3–8. doi: 10.1158/2326-6066.Cir-16-0297

32. Pang B, Zhen Y, Hu C, Ma Z, Lin S, Yi H. Myeloid-derived suppressor cells shift Th17/Treg ratio and promote systemic lupus erythematosus progression through arginase-1/miR-322-5p/TGF- β pathway. *Clin Sci (London Engl 1979)* (2020) 134 (16):2209–22. doi: 10.1042/cs20200799

33. Liu H, Ling CC, Yeung WHO, Pang L, Liu J, Zhou J, et al. Monocytic MDSC mobilization promotes tumor recurrence after liver transplantation *via* CXCL10/ TLR4/MMP14 signaling. *Cell Death Dis* (2021) 12(5):489. doi: 10.1038/s41419-021-03788-4

34. Schneider KM, Mohs A, Gui W, Galvez EJC, Candels LS, Hoenicke L, et al. Imbalanced gut microbiota fuels hepatocellular carcinoma development by shaping the hepatic inflammatory microenvironment. *Nat Commun* (2022) 13(1):3964. doi: 10.1038/ s41467-022-31312-5

35. Zhang S, Ma X, Zhu C, Liu L, Wang G, Yuan X. The role of myeloid-derived suppressor cells in patients with solid tumors: A meta-analysis. *PloS One* (2016) 11(10): e0164514. doi: 10.1371/journal.pone.0164514

36. Rahma OE, Hodi FS. The intersection between tumor angiogenesis and immune suppression. *Clin Cancer Res* (2019) 25(18):5449–57. doi: 10.1158/1078-0432.Ccr-18-1543

37. Hinshaw DC, Shevde LA. The tumor microenvironment innately modulates cancer progression. *Cancer Res* (2019) 79(18):4557–66. doi: 10.1158/0008-5472.Can-18-3962

38. Ostrand-Rosenberg S, Sinha P, Figley C, Long R, Park D, Carter D, et al. Frontline science: Myeloid-derived suppressor cells (MDSCs) facilitate maternal-fetal tolerance in mice. *J Leukoc Biol* (2017) 101(5):1091–101. doi: 10.1189/jlb.1HI1016-306RR

39. Ghaebi M, Nouri M, Ghasemzadeh A, Farzadi L, Jadidi-Niaragh F, Ahmadi M, et al. Immune regulatory network in successful pregnancy and reproductive failures. *BioMed Pharmacother* (2017) 88:61–73. doi: 10.1016/j.biopha.2017.01.016

40. Pan T, Liu Y, Zhong LM, Shi MH, Duan XB, Wu K, et al. Myeloid-derived suppressor cells are essential for maintaining feto-maternal immunotolerance *via* STAT3 signaling in mice. *J Leukocyte Biol* (2016) 100(3):499–511. doi: 10.1189/ jlb.1A1015-481RR

41. Ren J, Zeng W, Tian F, Zhang S, Wu F, Qin X, et al. Myeloid-derived suppressor cells depletion may cause pregnancy loss *via* upregulating the cytotoxicity of decidual natural killer cells. *Am J Reprod Immunol* (2019) 81(4):e13099. doi: 10.1111/aji.13099

42. Zhu M, Huang X, Yi S, Sun H, Zhou J. High granulocytic myeloid-derived suppressor cell levels in the peripheral blood predict a better IVF treatment outcome. J Matern Fetal Neona (2019) 32(7):1092–7. doi: 10.1080/14767058.2017.1400002

43. Hu C, Zhen Y, Pang B, Lin X, Yi H. Myeloid-derived suppressor cells are regulated by estradiol and are a predictive marker for IVF outcome. *Front Endocrinol* (2019) 10:521. doi: 10.3389/fendo.2019.00521

44. Liu Y, Perego M, Xiao Q, He Y, Fu S, He J, et al. Lactoferrin-induced myeloidderived suppressor cell therapy attenuates pathologic inflammatory conditions in newborn mice. *J Clin Invest* (2019) 129(10):4261–75. doi: 10.1172/JCI128164

45. Leiber A, Schwarz J, Köstlin N, Spring B, Fehrenbach B, Katava N, et al. Neonatal myeloid derived suppressor cells show reduced apoptosis and immunosuppressive activity upon infection with escherichia coli. *Eur J Immunol* (2017) 47(6):1009–21. doi: 10.1002/eji.201646621

46. Schwarz J, Scheckenbach V, Kugel H, Spring B, Pagel J, Härtel C, et al. Granulocytic myeloid-derived suppressor cells (GR-MDSC) accumulate in cord blood of preterm infants and remain elevated during the neonatal period. *Clin Exp Immunol* (2018) 191(3):328–37. doi: 10.1111/cei.13059

47. Yang F, Zheng Q, Jin L. Dynamic function and composition changes of immune cells during normal and pathological pregnancy at the maternal-fetal interface. *Front Immunol* (2019) 10:2317. doi: 10.3389/fimmu.2019.02317

48. Vento-Tormo R, Efremova M, Botting RA, Turco MY, Vento-Tormo M, Meyer KB, et al. Single-cell reconstruction of the early maternal-fetal interface in humans. *Nature* (2018) 563(7731):347–53. doi: 10.1038/s41586-018-0698-6

49. Cai D, Tang Y, Yao X. Changes of $\gamma\delta T$ cell subtypes during pregnancy and their influences in spontaneous abortion. J Reprod Immunol (2019) 131:57–62. doi: 10.1016/ j.jri.2019.01.003

50. Miller D, Gershater M, Slutsky R, Romero R, Gomez-Lopez N. Maternal and fetal T cells in term pregnancy and preterm labor. *Cell Mol Immunol* (2020) 17(7):693–704. doi: 10.1038/s41423-020-0471-2

51. Lu H, Yang H-L, Zhou W-J, Lai Z-Z, Qiu X-M, Fu Q, et al. Rapamycin prevents spontaneous abortion by triggering decidual stromal cell autophagy-mediated NK cell residence. *Autophagy* (2021) 17(9):2511–27. doi: 10.1080/15548627.2020.1833515

52. Fu B, Tian Z, Wei H. TH17 cells in human recurrent pregnancy loss and preeclampsia. *Cell Mol Immunol* (2014) 11(6):564–70. doi: 10.1038/cmi.2014.54 53. Zhang Y, Wang X, Zhang R, Wang X, Fu H, Yang W. MDSCs interactions with other immune cells and their role in maternal-fetal tolerance. *Int Rev Immunol* (2022) 41 (5):534–51. doi: 10.1080/08830185.2021.1938566

54. Aplin JD, Myers JE, Timms K, Westwood M. Tracking placental development in health and disease. *Nat Rev Endocrinol* (2020) 16(9):479–94. doi: 10.1038/s41574-020-0372-6

55. Yu B, Guo F, Yang Y, Long W, Zhou J. Steroidomics of pregnant women at advanced age. *Front Endocrinol (Lausanne)* (2022) 13:796909. doi: 10.3389/fendo.2022.796909

56. Veglia F, Perego M, Gabrilovich D. Myeloid-derived suppressor cells coming of age. Nat Immunol (2018) 19(2):108–19. doi: 10.1038/s41590-017-0022-x

57. Cassetta L, Bruderek K, Skrzeczynska-Moncznik J, Osiecka O, Hu X, Rundgren IM, et al. Differential expansion of circulating human MDSC subsets in patients with cancer, infection and inflammation. *J Immunother Cancer* (2020) 8(2):e001223. doi: 10.1136/jitc-2020-001223

58. Perez C, Botta C, Zabaleta A, Puig N, Cedena MT, Goicoechea I, et al. Immunogenomic identification and characterization of granulocytic myeloid-derived suppressor cells in multiple myeloma. *Blood* (2020) 136(2):199–209. doi: 10.1182/ blood.2019004537

59. Deng LJ, Qi M, Li N, Lei YH, Zhang DM, Chen JX. Natural products and their derivatives: Promising modulators of tumor immunotherapy. *J Leukoc Biol* (2020) 108 (2):493–508. doi: 10.1002/jlb.3mr0320-444r

60. Salminen A, Kauppinen A, Kaarniranta K. AMPK activation inhibits the functions of myeloid-derived suppressor cells (MDSC): impact on cancer and aging. *J Mol Med* (*Berl*) (2019) 97(8):1049–64. doi: 10.1007/s00109-019-01795-9

61. Taki M, Abiko K, Baba T, Hamanishi J, Yamaguchi K, Murakami R, et al. Snail promotes ovarian cancer progression by recruiting myeloid-derived suppressor cells *via* CXCR2 ligand upregulation. *Nat Commun* (2018) 9(1):1685. doi: 10.1038/s41467-018-03966-7

62. Köstlin N, Kugel H, Spring B, Leiber A, Marmé A, Henes M, et al. Granulocytic myeloid derived suppressor cells expand in human pregnancy and modulate T-cell responses. *Eur J Immunol* (2014) 44(9):2582–91. doi: 10.1002/eji.201344200

63. Zhao H, Kalish F, Schulz S, Yang Y, Wong RJ, Stevenson DK. Unique roles of infiltrating myeloid cells in the murine uterus during early to midpregnancy. J Immunol (Baltimore Md 1950) (2015) 194(8):3713–22. doi: 10.4049/jimmunol.1401930

64. Bartmann C, Junker M, Segerer SE, Häusler SF, Krockenberger M, Kämmerer U. CD33(+) /HLA-DR(neg) and CD33(+) /HLA-DR(+/-) cells: Rare populations in the human decidua with characteristics of MDSC. *Am J Reprod Immunol* (2016) 75(5):539–56. doi: 10.1111/aji.12492

65. Zhang T, Zhou J, Man GCW, Leung KT, Liang B, Xiao B, et al. MDSCs drive the process of endometriosis by enhancing angiogenesis and are a new potential therapeutic target. *Eur J Immunol* (2018) 48(6):1059–73. doi: 10.1002/eji.201747417

66. Suah AN, Tran D-KV, Khiew SH, Andrade MS, Pollard JM, Jain D, et al. Pregnancy-induced humoral sensitization overrides T cell tolerance to fetus-matched allografts in mice. J Clin Invest (2021) 131(1):e140715. doi: 10.1172/JCI140715

67. Kinder JM, Stelzer IA, Arck PC, Way SS. Immunological implications of pregnancy-induced microchimerism. *Nat Rev Immunol* (2017) 17(8):483-94. doi: 10.1038/nri.2017.38

68. Pollard JM, Chong AS. Semiallogeneic pregnancy: A paradigm change for T-cell transplantation tolerance. *Transplantation* (2022) 106(6):1098–100. doi: 10.1097/TP.000000000004139

69. Piccinni MP, Raghupathy R, Saito S, Szekeres-Bartho J. Cytokines, hormones and cellular regulatory mechanisms favoring successful reproduction. *Front Immunol* (2021) 12:717808. doi: 10.3389/fimmu.2021.717808

70. AbdulHussain G, Azizieh F, Makhseed M, Raghupathy R. Effects of progesterone, dydrogesterone and estrogen on the production of Th1/Th2/Th17 cytokines by lymphocytes from women with recurrent spontaneous miscarriage. *J Reprod Immunol* (2020) 140:103132. doi: 10.1016/j.jri.2020.103132

71. Kang X, Zhang X, Liu Z, Xu H, Wang T, He L, et al. Granulocytic myeloid-derived suppressor cells maintain feto-maternal tolerance by inducing Foxp3 expression in CD4 +CD25-T cells by activation of the TGF- β/β -catenin pathway. *Mol Hum Reprod* (2016) 22 (7):499–511. doi: 10.1093/molehr/gaw026

72. Köstlin N, Hofstädter K, Ostermeir A-L, Spring B, Leiber A, Haen S, et al. Granulocytic myeloid-derived suppressor cells accumulate in human placenta and polarize toward a Th2 phenotype. J Immunol (Baltimore Md 1950) (2016) 196(3):1132–45. doi: 10.4049/jimmunol.1500340

73. Dietz S, Schwarz J, Ruhle J, Schaller M, Fehrenbacher B, Marme A, et al. Extracellular vesicles released by myeloid-derived suppressor cells from pregnant women modulate adaptive immune responses. *Cell Immunol* (2021) 361:104276. doi: 10.1016/j.cellimm.2020.104276

74. Nair RR, Sinha P, Khanna A, Singh K. Reduced myeloid-derived suppressor cells in the blood and endometrium is associated with early miscarriage. *Am J Reprod Immunol* (2015) 73(6):479–86. doi: 10.1111/aji.12351

75. PrabhuDas M, Bonney E, Caron K, Dey S, Erlebacher A, Fazleabas A, et al. Immune mechanisms at the maternal-fetal interface: perspectives and challenges. *Nat Immunol* (2015) 16(4):328–34. doi: 10.1038/ni.3131

 Nasri F, Doroudchi M, Namavar Jahromi B, Gharesi-Fard B. T Helper cells profile and CD4+CD25+Foxp3+Regulatory T cells in polycystic ovary syndrome. *Iran J Immunol* (2018) 15(3):175–85. doi: 10.22034/IJI.2018.39387 77. Jabrane-Ferrat N. Features of human decidual NK cells in healthy pregnancy and during viral infection. Front In Immunol (2019) 10:1397. doi: 10.3389/fimmu.2019.01397

78. Huhn O, Zhao X, Esposito L, Moffett A, Colucci F, Sharkey AM. How do uterine natural killer and innate lymphoid cells contribute to successful pregnancy? *Front Immunol* (2021) 12:607669. doi: 10.3389/fimmu.2021.607669

79. Björkström NK, Ljunggren HG, Michaëlsson J. Emerging insights into natural killer cells in human peripheral tissues. *Nat Rev Immunol* (2016) 16(5):310-20. doi: 10.1038/nri.2016.34

80. Zhang X, Wei H. Role of decidual natural killer cells in human pregnancy and related pregnancy complications. *Front In Immunol* (2021) 12:728291. doi: 10.3389/fimmu.2021.728291

81. Wang F, Qualls AE, Marques-Fernandez L, Colucci F. Biology and pathology of the uterine microenvironment and its natural killer cells. *Cell Mol Immunol* (2021) 18 (9):2101–13. doi: 10.1038/s41423-021-00739-z

82. Mauti LA, Le Bitoux M-A, Baumer K, Stehle J-C, Golshayan D, Provero P, et al. Myeloid-derived suppressor cells are implicated in regulating permissiveness for tumor metastasis during mouse gestation. *J Clin Invest* (2011) 121(7):2794–807. doi: 10.1172/JCI41936

83. Phipps EA, Thadhani R, Benzing T, Karumanchi SA. Pre-eclampsia: pathogenesis, novel diagnostics and therapies. *Nat Rev Nephrol* (2019) 15(5):275–89. doi: 10.1038/ s41581-019-0119-6

84. Huang Z, Huang S, Song T, Yin Y, Tan C. Placental angiogenesis in mammals: A review of the regulatory effects of signaling pathways and functional nutrients. *Adv Nutr* (2021) 12(6):2415–34. doi: 10.1093/advances/nmab070

85. Weber R, Fleming V, Hu X, Nagibin V, Groth C, Altevogt P, et al. Myeloid-derived suppressor cells hinder the anti-cancer activity of immune checkpoint inhibitors. *Front In Immunol* (2018) 9:1310. doi: 10.3389/fimmu.2018.01310

86. Wang SC, Li YH, Piao HL, Hong XW, Zhang D, Xu YY, et al. PD-1 and Tim-3 pathways are associated with regulatory CD8+ T-cell function in decidua and maintenance of normal pregnancy. *Cell Death Dis* (2015) 6:e1738. doi: 10.1038/ cddis.2015.112

87. Costanzo V, Bardelli A, Siena S, Abrignani S. Exploring the links between cancer and placenta development. *Open Biol* (2018) 8(6):180081. doi: 10.1098/rsob.180081

88. Rabbani ML, Rogers PA. Role of vascular endothelial growth factor in endometrial vascular events before implantation in rats. *Reproduction* (2001) 122(1):85–90. doi: 10.1530/rep.0.1220085

89. Kang X, Zhang X, Liu Z, Xu H, Wang T, He L, et al. CXCR2-mediated granulocytic myeloid-derived suppressor cells' functional characterization and their role in maternal fetal interface. *DNA Cell Biol* (2016) 35(7):358–65. doi: 10.1089/dna.2015.2962

90. Ahmadi M, Mohammadi M, Ali-Hassanzadeh M, Zare M, Gharesi-Fard B. MDSCs in pregnancy: Critical players for a balanced immune system at the feto-maternal interface. *Cell Immunol* (2019) 346:103990. doi: 10.1016/j.cellimm.2019.103990

91. Jing G, Yao J, Dang Y, Liang W, Xie L, Chen J, et al. The role of β -HCG and VEGF-MEK/ERK signaling pathway in villi angiogenesis in patients with missed abortion. *Placenta* (2021) 103:16–23. doi: 10.1016/j.placenta.2020.10.005

92. Lai ZZ, Yang HL, Shi JW, Shen HH, Wang Y, Chang KK, et al. Protopanaxadiol improves endometriosis associated infertility and miscarriage in sex hormones receptorsdependent and independent manners. *Int J Biol Sci* (2021) 17(8):1878–94. doi: 10.7150/ ijbs.58657

93. Zhang Y, Qu D, Sun J, Zhao L, Wang Q, Shao Q, et al. Human trophoblast cells induced MDSCs from peripheral blood CD14(+) myelomonocytic cells via elevated levels of CCL2. Cell Mol Immunol (2016) 13(5):615–27. doi: 10.1038/cmi.2015.41

94. Svoronos N, Perales-Puchalt A, Allegrezza MJ, Rutkowski MR, Payne KK, Tesone AJ, et al. Tumor cell-independent estrogen signaling drives disease progression through mobilization of myeloid-derived suppressor cells. *Cancer Discov* (2017) 7(1):72–85. doi: 10.1158/2159-8290.CD-16-0502

95. Pan T, Zhong L, Wu S, Cao Y, Yang Q, Cai Z, et al. 17β-oestradiol enhances the expansion and activation of myeloid-derived suppressor cells *via* signal transducer and activator of transcription (STAT)-3 signalling in human pregnancy. *Clin Exp Immunol* (2016) 185(1):86–97. doi: 10.1111/cei.12790

96. Verma P, Verma R, Nair RR, Budhwar S, Khanna A, Agrawal NR, et al. Altered crosstalk of estradiol and progesterone with myeloid-derived suppressor cells and Th1/Th2 cytokines in early miscarriage is associated with early breakdown of maternal-fetal tolerance. *Am J Reprod Immunol* (2019) 81(2):e13081. doi: 10.1111/aji.13081

97. Dietz S, Schwarz J, Velic A, Gonzalez-Menendez I, Quintanilla-Martinez L, Casadei N, et al. Human leucocyte antigen G and murine qa-2 are critical for myeloid derived suppressor cell expansion and activation and for successful pregnancy outcome. *Front Immunol* (2021) 12:787468. doi: 10.3389/fimmu.2021.787468

98. Köstlin N, Ostermeir A-L, Spring B, Schwarz J, Marmé A, Walter CB, et al. HLA-G promotes myeloid-derived suppressor cell accumulation and suppressive activity during human pregnancy through engagement of the receptor ILT4. *Eur J Immunol* (2017) 47 (2):374–84. doi: 10.1002/eji.201646564

99. Zare M, Namavar Jahromi B, Gharesi-Fard B. Analysis of the frequencies and functions of CD4CD25CD127, CD4HLA-G, and CD8HLA-G regulatory T cells in preeclampsia. *J Reprod Immunol* (2019) 133:43–51. doi: 10.1016/j.jri.2019.06.002

100. Morita M, Sato Y, Iwasaki R, Kobayashi T, Watanabe R, Oike T, et al. Selective estrogen receptor modulators suppress Hif1 α protein accumulation in mouse osteoclasts. *PloS One* (2016) 11(11):e0165922. doi: 10.1371/journal.pone.0165922

101. Köstlin-Gille N, Dietz S, Schwarz J, Spring B, Pauluschke-Fröhlich J, Poets CF, et al. HIF-1 α -Deficiency in myeloid cells leads to a disturbed accumulation of myeloid derived suppressor cells (MDSC) during pregnancy and to an increased abortion rate in mice. *Front In Immunol* (2019) 10:161. doi: 10.3389/fimmu.2019.00161

102. Xia S, Sha H, Yang L, Ji Y, Ostrand-Rosenberg S, Qi L. Gr-1+ CD11b+ myeloidderived suppressor cells suppress inflammation and promote insulin sensitivity in obesity. *J Biol Chem* (2011) 286(26):23591–9. doi: 10.1074/jbc.M111.237123

103. Clements VK, Long T, Long R, Figley C, Smith DMC, Ostrand-Rosenberg S. Frontline science: High fat diet and leptin promote tumor progression by inducing myeloid-derived suppressor cells. *J Leukocyte Biol* (2018) 103(3):395–407. doi: 10.1002/JLB.4HI0517-210R

104. Bao Y, Mo J, Ruan L, Li G. Increased monocytic CD14⁺HLADRlow/- myeloidderived suppressor cells in obesity. *Mol Med Rep* (2015) 11(3):2322–8. doi: 10.3892/ mmr.2014.2927

105. Chan P-C, Liao M-T, Hsieh P-S. The dualistic effect of COX-2-Mediated signaling in obesity and insulin resistance. Int J Mol Sci (2019) 20(13):3115. doi: 10.3390/ijms20133115

106. Bunt SK, Yang L, Sinha P, Clements VK, Leips J, Ostrand-Rosenberg S. Reduced inflammation in the tumor microenvironment delays the accumulation of myeloid-derived suppressor cells and limits tumor progression. *Cancer Res* (2007) 67(20):10019–26. doi: 10.1158/0008-5472.CAN-07-2354

107. Sade-Feldman M, Kanterman J, Ish-Shalom E, Elnekave M, Horwitz E, Baniyash M. Tumor necrosis factor- α blocks differentiation and enhances suppressive activity of immature myeloid cells during chronic inflammation. *Immunity* (2013) 38(3):541–54. doi: 10.1016/j.immuni.2013.02.007

108. Tomić S, Joksimović B, Bekić M, Vasiljević M, Milanović M, Čolić M, et al. Prostaglanin-E2 potentiates the suppressive functions of human mononuclear myeloidderived suppressor cells and increases their capacity to expand IL-10-Producing regulatory T cell subsets. *Front In Immunol* (2019) 10:475. doi: 10.3389/ fmmu.2019.00475

109. Wang Q, Zhang X, Li C, Xiong M, Bai W, Sun S, et al. Intracellular lipid accumulation drives the differentiation of decidual polymorphonuclear myeloid-derived suppressor cells *via* arachidonic acid metabolism. *Front In Immunol* (2022) 13:868669. doi: 10.3389/fimmu.2022.868669

110. Li YH, Zhou WH, Tao Y, Wang SC, Jiang YL, Zhang D, et al. The galectin-9/Tim-3 pathway is involved in the regulation of NK cell function at the maternal-fetal interface in early pregnancy. *Cell Mol Immunol* (2016) 13(1):73–81. doi: 10.1038/cmi.2014.126

111. Wang Y, Liu Y, Shu C, Wan J, Shan Y, Zhi X, et al. Inhibition of pregnancyassociated granulocytic myeloid-derived suppressor cell expansion and arginase-1 production in preeclampsia. *J Reprod Immunol* (2018) 127:48–54. doi: 10.1016/ j.jri.2018.05.002

112. Li C, Chen C, Kang X, Zhang X, Sun S, Guo F, et al. Decidua-derived granulocyte macrophage colony-stimulating factor induces polymorphonuclear myeloid-derived suppressor cells from circulating CD15+ neutrophils. *Hum Reprod* (2020) 35(12):2677–91. doi: 10.1093/humrep/deaa217

113. Li C, Zhang X, Kang X, Chen C, Guo F, Wang Q, et al. Upregulated TRAIL and reduced DcR2 mediate apoptosis of decidual PMN-MDSC in unexplained recurrent pregnancy loss. *Front In Immunol* (2020) 11:1345. doi: 10.3389/fimmu.2020.01345

114. Mandruzzato G, Antsaklis A, Botet F, Chervenak FA, Figueras F, Grunebaum A, et al. Intrauterine restriction (IUGR). *J Perinat Med* (2008) 36(4):277–81. doi: 10.1515/JPM.2008.050

115. Pilliod RA, Cheng YW, Snowden JM, Doss AE, Caughey AB. The risk of intrauterine fetal death in the small-for-gestational-age fetus. *Am J Obstet Gynecol* (2012) 207(4):318.e1-6. doi: 10.1016/j.ajog.2012.06.039

116. Ma'ayeh M, Costantine MM. Prevention of preeclampsia. Semin Fetal Neonatal Med (2020) 25(5):101123. doi: 10.1016/j.siny.2020.101123

117. Marsál K. Intrauterine growth restriction. Curr Opin Obstet Gynecol (2002) 14 (2):127–35. doi: 10.1097/00001703-200204000-00005

118. Veerbeek JHW, Nikkels PGJ, Torrance HL, Gravesteijn J, Post Uiterweer ED, Derks JB, et al. Placental pathology in early intrauterine growth restriction associated with maternal hypertension. *Placenta* (2014) 35(9):696–701. doi: 10.1016/j.placenta.2014.06.375

119. John RM. Imprinted genes and the regulation of placental endocrine function: Pregnancy and beyond. *Placenta* (2017) 56:86–90. doi: 10.1016/j.placenta.2017.01.099

120. Shi M, Chen Z, Chen M, Liu J, Li J, Xing Z, et al. Continuous activation of polymorphonuclear myeloid-derived suppressor cells during pregnancy is critical for fetal development. *Cell Mol Immunol* (2021) 18(7):1692–707. doi: 10.1038/s41423-021-00704-w

121. Dong S, Shah NK, He J, Han S, Xie M, Wang Y, et al. The abnormal expression of Tim-3 is involved in the regulation of myeloid-derived suppressor cells and its correlation with preeclampsia. *Placenta* (2021) 114:108–14. doi: 10.1016/j.placenta.2021.08.060

122. Guha P, Heatherton KR, O'Connell KP, Alexander IS, Katz SC. Assessing the future of solid tumor immunotherapy. *Biomedicines* (2022) 10(3):655. doi: 10.3390/biomedicines10030655

123. Dong S, Guo X, Han F, He Z, Wang Y. Emerging role of natural products in cancer immunotherapy. *Acta Pharm Sin B* (2022) 12(3):1163-85. doi: 10.1016/j.apsb.2021.08.020

124. Schroeter A, Roesel MJ, Matsunaga T, Xiao Y, Zhou H, Tullius SG. Aging affects the role of myeloid-derived suppressor cells in alloimmunity. *Front Immunol* (2022) 13:917972. doi: 10.3389/fimmu.2022.917972

125. Park Y, Kwok SK. Recent advances in cell therapeutics for systemic autoimmune diseases. *Immune Netw* (2022) 22(1):e10. doi: 10.4110/in.2022.22.e10

126. Zhao A-M, Xu H-J, Kang X-M, Zhao A-M, Lu L-M. New insights into myeloidderived suppressor cells and their roles in feto-maternal immune cross-talk. *J Reprod Immunol* (2016) 113:35-41. doi: 10.1016/j.jri.2015.11.001

127. He Y-M, Li X, Perego M, Nefedova Y, Kossenkov AV, Jensen EA, et al. Transitory presence of myeloid-derived suppressor cells in neonates is critical for control of inflammation. *Nat Med* (2018) 24(2):224–31. doi: 10.1038/nm.4467

128. Köstlin N, Schoetensack C, Schwarz J, Spring B, Marmé A, Goelz R, et al. Granulocytic myeloid-derived suppressor cells (GR-MDSC) in breast milk (BM); GR-MDSC accumulate in human BM and modulate T-cell and monocyte function. *Front In Immunol* (2018) 9:1098. doi: 10.3389/fimmu.2018.01098

129. Budhwar S, Verma R, Verma P, Bala R, Rai S, Singh K. Estradiol correlates with the accumulation of monocytic myeloid-derived suppressor cells in pre-term birth: A possible explanation of immune suppression in pre-term babies. *J Reprod Immunol* (2021) 147:103350. doi: 10.1016/j.jri.2021.103350

130. Levy O. Innate immunity of the newborn: basic mechanisms and clinical correlates. *Nat Rev Immunol* (2007) 7(5):379-90. doi: 10.1038/nri2075

131. Rieber N, Gille C, Köstlin N, Schäfer I, Spring B, Ost M, et al. Neutrophilic myeloid-derived suppressor cells in cord blood modulate innate and adaptive immune responses. *Clin Exp Immunol* (2013) 174(1):45–52. doi: 10.1111/cei.12143

132. Dabagh-Gorjani F, Anvari F, Zolghadri J, Kamali-Sarvestani E, Gharesi-Fard B. Differences in the expression of TLRs and inflammatory cytokines in pre-eclamptic compared with healthy pregnant women. *Iran J Immunol* (2014) 11(4):233–45

133. Gervassi A, Lejarcegui N, Dross S, Jacobson A, Itaya G, Kidzeru E, et al. Myeloid derived suppressor cells are present at high frequency in neonates and suppress *in vitro* T cell responses. *PloS One* (2014) 9(9):e107816. doi: 10.1371/journal.pone.0107816

134. Köstlin N, Vogelmann M, Spring B, Schwarz J, Feucht J, Härtel C, et al. Granulocytic myeloid-derived suppressor cells from human cord blood modulate Thelper cell response towards an anti-inflammatory phenotype. *Immunology* (2017) 152 (1):89–101. doi: 10.1111/imm.12751

135. Budhwar S, Verma P, Verma R, Rai S, Singh K. The yin and yang of myeloid derived suppressor cells. *Front Immunol* (2018) 9:2776. doi: 10.3389/fimmu.2018.02776

136. Leaphart CL, Cavallo J, Gribar SC, Cetin S, Li J, Branca MF, et al. A critical role for TLR4 in the pathogenesis of necrotizing enterocolitis by modulating intestinal injury and repair. *J Immunol* (2007) 179(7):4808–20. doi: 10.4049/jimmunol.179.7.4808