Narcolepsy is strongly associated with the T-cell receptor alpha locus

Joachim Hallmayer^{1,2,32}, Juliette Faraco^{1,2,32}, Ling Lin^{1,2}, Stephanie Hesselson³, Juliane Winkelmann^{4–6}, Minae Kawashima^{1,2,7}, Geert Mayer^{8,9}, Giuseppe Plazzi¹⁰, Sona Nevsimalova¹¹, Patrice Bourgin¹², Seung-Chul Hong¹³, Yutaka Honda¹⁴, Makoto Honda¹⁴, Birgit Högl¹⁵, William T Longstreth Jr^{16,17}, Jacques Montplaisir¹⁸, David Kemlink¹¹, Mali Einen^{1,2}, Justin Chen³, Stacy L Musone³, Matthew Akana³, Taku Miyagawa⁷, Jubao Duan¹⁹, Alex Desautels¹⁸, Christine Erhardt¹², Per Egil Hesla²⁰, Francesca Poli¹⁰, Birgit Frauscher¹⁵, Jong-Hyun Jeong¹³, Sung-Pil Lee¹³, Thanh G N Ton^{16,17}, Mark Kvale³, Libor Kolesar²¹, Marie Dobrovolná²², Gerald T Nepom²³, Dan Salomon²⁴, H-Erich Wichmann^{25,26}, Guy A Rouleau²⁷, Christian Gieger²⁵, Douglas F Levinson², Pablo V Gejman^{19,28}, Thomas Meitinger^{4,6}, Terry Young²⁹, Paul Peppard²⁹, Katsushi Tokunaga⁷, Pui-Yan Kwok³, Neil Risch^{3,30} & Emmanuel Mignot^{1,2,31}

rapid onset into REM sleep, affects 1 in 2,000 individuals^{1,2}. Narcolepsy was first shown to be tightly associated with HLA-DR2 (ref. 3) and later sublocalized to DOB1*0602 (ref. 4). Following studies in dogs⁵ and mice⁶, a 95% loss of hypocretin-producing cells in postmortem hypothalami from narcoleptic individuals was reported^{7,8}. Using genome-wide association (GWA) in Caucasians with replication in three ethnic groups, we found association between narcolepsy and polymorphisms in the TRA@ (T-cell receptor alpha) locus, with highest significance at rs1154155 (average allelic odds ratio 1.69, genotypic odds ratios 1.94 and 2.55, *P* < 10⁻²¹, 1,830 cases, 2,164 controls). This is the first documented genetic involvement of the TRA@ locus, encoding the major receptor for HLA-peptide presentation, in any disease. It is still unclear how specific HLA alleles confer susceptibility to over 100 HLA-associated disorders⁹; thus, narcolepsy will provide new insights on how HLA-TCR interactions contribute to

Narcolepsy with cataplexy, characterized by sleepiness and

organ-specific autoimmune targeting and may serve as a model for over 100 other HLA-associated disorders9.

An autoimmune etiology has been suggested for narcolepsy but never proven despite decades of intensive research^{10,11}. Narcolepsy is recognized to be familial, and despite the strong association with HLA-DQB1*0602, is not fully explained by the HLA locus¹. To identify additional susceptibility loci for narcolepsy, we undertook a genome-wide association study. We selected Caucasian cases from North America and Europe, together with geographically and ethnically matched controls. All cases were HLA-DQB1*0602 positive and all had clear-cut cataplexy. Among the 23% for whom we had measurements of hypocretin-1, all were found to be hypocretin deficient. Potential controls were typed using sequence-specific PCR, and only those who were also HLA-DQB1*0602 positive were included. The sample was comprised of 807 cases and 1,074 controls of mixed European ancestry: 415 cases and 753 controls were recruited from the United States and Canada; 392 cases and 321 controls were

Received 24 December 2008; accepted 5 March 2009; published online 3 May 2009; corrected after print 26 June 2009; doi:10.1038/ng.372

¹Center for Sleep Sciences and ²Department of Psychiatry, Stanford University School of Medicine, Palo Alto, California, USA. ³Institute for Human Genetics, University of California San Francisco School of Medicine, San Francisco, California, USA. ⁴Institute for Human Genetics, Technische Universität München, Munich, Germany. ⁵Department of Neurology, Technische Universität München, Munich, Germany. ⁶Institute of Human Genetics Helmholtz Zentrum München, Munich Germany. ⁷Department of Human Genetics, University of Tokyo, Tokyo, Japan. ⁸Hephata-Klinik, Schwalmstadt-Treysa, Germany. ⁹Department of Neurology, Philipps University of Marburg, Marburg, Germany. ¹⁰University of Bologna, Bologna, Italy. ¹¹Department of Neurology, Charles University in Prague, 1st Faculty of Medicine and General Teaching Hospital, Pague, Czech Republic, ¹²Sleep Clinic, Hôpital Civil, Louis Pasteur University, Strasbourg, France. ¹³Department of Neuropsychiatry, St. Vincent's Hospital, The Catholic University of Korea, Suwon, Korea. ¹⁴Tokyo Institute of Psychiatry, Setagaya, Japan. ¹⁵Department of Neurology, Innsbruck Medical University, Innsbruck, Austria. ¹⁶Departments of Neurology and ¹⁷Epidemiology, University of Washington, Seattle, Washington, USA. ¹⁸Sleep Disorders Center, Université de Montréal, Montréal, Québec, Canada. ¹⁹Department of Psychiatry, Feinberg School of Medicine, Northwestern University, Evanston, Illinois, USA. ²⁰Coliseum on Majorstua Clinic, Oslo, Norway. ²¹Department of Immunogenetics, Institute for Clinical and Experimental Medicine, Videnska, Prague, Czech Republic. ²²HLA typing lab, National Reference Laboratory for DNA Diagnostics, Institute of Hematology and Blood Transfusion, Prague, Czech Republic. ²³The Benaroya Research Institute at Virginia Mason, Seattle, Washington, USA. ²⁴Department of Molecular and Experimental Medicine, The Scripps Research Institute, La Jolla, California, USA. ²⁵Institute of Epidemiology Helmholtz Zentrum München, Munich, Germany. ²⁶Institute of Medical Informatics, Biometry and Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany.²⁷Center of Excellence in Neuromics, Université de Montréal, Montréal, Canada.²⁸NorthShore University HealthSystem, Evanston, Illinois, USA. ²⁹Department of Population Health Sciences, University of Wisconsin Medical School, Madison, Wisconsin, USA. ³⁰Kaiser Permanente Northern California Division of Research, Oakland, California, USA. ³¹Howard Hughes Medical Institute, Stanford University, California, USA. ³²These authors contributed equally to this work. Correspondence should be addressed to E.M. (mignot@stanford.edu).

Table 1 SNP markers of interest from the genome-wide association study

SNP	Chr	Position (bp)	Minor allele	Freq controls (<i>n</i>)	Freq cases (n)	χ ² (MH)	<i>P</i> (MH)	OR (95% CI)	χ^2 (BD)	<i>P</i> (BD)
rs1154155	14	22072524	С	0.14 (1,067)	0.24 (796)	54.11	1.90×10^{-13}	1.87 (1.58–2.21)	2.49	0.29
rs12587781	14	22069457	С	0.15 (917)	0.25 (622)	53.19	3.03×10^{-13}	1.96 (1.63–2.35)	1.66	0.20
				0.14 (1,066) ^a	0.24 (794) ^a	60.42 ^a	7.65×10^{-15a}	1.93 (1.63–2.28) ^a	1.61ª	0.45ª
rs1263646	14	22087370	G	0.16 (1,069)	0.25 (797)	47.74	4.86×10^{-12}	1.77 (1.50–2.09)	0.40	0.82
rs5770917 ^b	22	49364219	G	0.05 (1,063)	0.04 (796)	1.07	0.30	0.84 (0.61–1.16)	1.90	0.39

The top three genome-wide significant markers are listed, together with data obtained for rs5770917, previously found to be associated with narcolepsy in a Japanese population¹⁷. We genotyped 1,074 controls and 807 cases using SNP Affymetrix Array platforms (500K and 6.0). MH, Mantel-Haenszel; BD, Breslow Day heterogeneity test; OR, odds ratio. ^aAffymetrix 6.0K marker after genotypes were completed using TaqMan (see text). ^bNote that 388 of the 796 narcolepsy genotypes were previously reported for this marker by Miyagawa *et al.*¹⁷.

recruited from European centers. For the GWA study, subjects were genotyped using the Affymetrix Mapping 500K array set or Genome-Wide SNP Array 6.0. Homogeneity in ancestry and case-control matching was verified by cluster and principal component analysis¹². In addition, we compared the allele frequency of 107 of 400 SNPs known to predict European substructure and found no significant differences after Bonferroni correction¹³.

We conducted allele-based association tests in SNPs with allele frequency above 5% in controls using the Mantel-Haenszel test¹⁴ in three groups of subjects defined by platform and location of typing (Affymetrix 500K typed at UCSF; Affymetrix 6.0 typed at UCSF; Affymetrix 6.0 typed at Institut fur Humangenetik, Munich, Germany). The χ^2 quantile-quantile plot showed a slight deviation from the expected χ^2 distribution, and an inflation factor λ of 1.11 was estimated (**Supplementary Fig. 1** online). However, the plot also showed the presence of three extreme outlier χ^2 values of 47.7, 54.1 and 60.4 (**Table 1** and **Supplementary Fig. 1**). These three SNPs, all on chromosome 14, clearly exceeded the genome-wide significance level of 9.1×10^{-8} . Other nominally significant associations ($P < 10^{-6}$) are reported in **Supplementary Table 1** online.

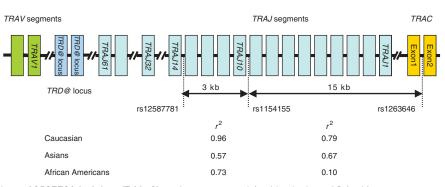
The three top markers were in high linkage disequilibrium (LD) and are located within an 18-kb segment of the *TRA@* locus containing the TRA joining (J) segment subregion (14q11.2; **Fig. 1**). Of interest, one of the markers of nominal significance ($P = 5.2 \times 10^{-7}$), rs17231, is located within the V segment region of the *TRB@* (T-cell receptor beta) locus (7q34). Genome-wide significant SNPs were genotyped using TaqMan assays (Applied Biosystems) in an independent sample of 1,057 cases (using the same diagnostic criteria) and 1,104 controls (matched by ancestry) as a replication study. The Caucasian replication sample contained 718 individuals, of whom 542 were recruited from the United States and Canada (259 cases, 283 controls), and 176 from Europe (104 cases, 72 controls). The Asian sample included 866 Japanese (433 cases, 433 controls) and 300 Koreans (128 cases, 172 controls). Finally, we studied 277 African Americans (133 cases, 144 controls).

As shown in **Table 2**, the three SNPs located within the *TRA@* locus replicated with high significance across the three major ethnic groups combined and showed significant effects individually in the Caucasian and Asian subsamples. In the African American sample, although the odds ratios (ORs) trended in the same direction, formal significance was not reached owing to small sample size and low allele frequencies (**Table 2**).

According to HapMap data¹⁵, the three SNPs are located within a 37-kb region of increased LD across ancestry groups (CEU, YRI, CHB-JPT). The localized haplotype block structure among these populations differs, with highest LD between rs12587781 and rs1154155 extending in opposite directions in Europeans versus Asians. In all ethnic groups, rs1263646, a SNP located closer to the *TRAC* gene, showed a smaller OR, suggesting that the association peaks in the *TRAJ* segment region (**Fig. 1**). Further, ORs differed significantly for rs12587781 but not rs1154155 between Caucasians and Asians (**Table 2**). This was likely explained by the difference in LD patterns across the two ethnicities. Whereas rs1154155 and rs12587781 are in almost complete LD in Caucasians ($r^2 = 0.96$), LD is substantially weaker in Asians ($r^2 = 0.57$; **Fig. 1**). In Asians, rs1154155 had a stronger impact on risk (OR = 1.54) than did rs12587781

To further evaluate this pattern, we estimated the frequency of haplotypes AA, AC, CA, CC for rs12587781 and rs1154155 in Asian cases and controls. For cases, the frequencies were 0.318, 0.003, 0.109 and 0.571, respectively. For controls, the frequencies were 0.381, 0.005, 0.154 and 0.460, respectively. We note that the OR is increased for

Figure 1 Schematic representation of the *TRA@* locus and of SNPs associated with narcolepsy. The *TRA@* locus consists of clusters of V and J segments and exons of the C region. The T-cell receptor delta locus (*TRD@*) resides within the *TRA@* locus. A 40-kb region of LD encompasses half of the *TRAJ* segments and is flanked by *TRAJ32* and the second exon of the *TRAC* gene. Within this region, three SNPs are highly associated with narcolepsy, separated by 3 and 15 kb, successively. In Caucasians, the association is equivalent with rs12587781 and rs1154155 (**Tables 1** and **2**), and LD is extremely high ($r^2 = 0.97$ and 0.94; 1,154 cases and 1,425 controls, respectively). In



contrast, the association is stronger with rs1154155 than rs12587781 in Asians (**Table 2**), a phenomenon explained by the lower LD in this ancestry group ($r^2 = 0.62$ and 0.52; 553 cases and 603 controls, respectively). Intermediate LD was seen in African-American individuals ($r^2 = 0.74$ and 0.71; 124 cases and 142 controls, respectively). The association with rs1263646 is weaker across all ancestry groups, most notably Asians and African Americans (**Table 2**). These results, depicted as values for cases and controls combined in this figure, illustrate the value of trans-ethnic mapping.

Region of LD -//

Table 2 Replication	of SNP markers	discovered in	the GWA study
---------------------	----------------	---------------	---------------

-			
Ancestry	rs12587781	rs1154155	rs1263646
Caucasian	С	С	G
Freq controls (n)	0.14 (352)	0.14 (348)	0.16 (351)
Freq cases (n)	0.22 (353)	0.22 (343)	0.24 (353)
χ^2	17.08	17.04	13.66
Ρ	3.58×10^{-5}	3.67×10^{-5}	2.19×10^{-4}
OR (95% CI)	1.79 (1.36–2.37)	1.80 (1.36–2.39)	1.65 (1.26–2.15)
Asians	С	С	G
Freq controls (n)	0.61 (601)	0.47 (599)	0.45 (600)
Freq cases (n)	0.68 (552)	0.57 (549)	0.51 (553)
χ^2	11.09	26.76	9.81
Ρ	8.70×10^{-4}	2.30×10^{-7}	1.73×10^{-3}
OR (95% CI)	1.34 (1.13–1.59)	1.54 (1.31–1.82)	1.30 (1.10–1.53)
African Americans	С	С	G
Freq controls (n)	0.11 (142)	0.08 (138)	0.133 (139)
Freq cases (n)	0.13 (124)	0.10 (113)	0.165 (124)
χ^2	0.70	0.74	1.08
Ρ	0.40	0.39	0.30
OR (95% CI)	1.25 (0.74–2.13)	1.31 (0.71–2.42)	1.29 (0.80–2.09)

Within the subset of Caucasian controls with HLA information, allele frequencies at the three SNPs did not differ between DQB1*0602 positive (n = 469) and negative (n = 1,352) individuals.

haplotype CC versus AA (1.49, 95% CI = 1.24-1.79) but not for haplotype CA versus AA (0.85, 95% CI = 0.64-1.12). Thus, rs12587781 seems to have no effect after controlling for rs1154155, suggesting that rs1154155 may have functional significance or is in high LD with another causative SNP nearby. SNPs with $r^2 > 0.8$ with rs1154155 are known to exist from HapMap data. This SNP is located 176 bp 3' to TRAJ10, a J segment without known coding polymorphisms. Genotype analysis suggested a dosage effect (CC versus AA Mantel-Haenszel OR = 2.55, 95% CI = 1.92-3.38; AC versus AA Mantel-Haenszel OR = 1.94, 95% CI = 1.68-2.25) (Table 3).

Population attributable risks¹⁶ for TRA@ rs1154155C in Caucasians and in Asians were 20% and 42%, respectively. The increased frequency of rs1154155[C] in Asians likely contributes to the reported increased prevalence in Japan¹ despite lower HLA-DQB1*0602 frequency⁴. Our identified TRA@ rs1154155[C] polymorphism showed no interaction with the nominally significant TRB@ rs17231[T] polymorphism in the GWA data in our preliminary analyses (OR interaction = 1.2, P = 0.24). In our much larger sample, we also did not replicate a previously published rs5770917 association in Japanese narcolepsy (Table 1), suggesting an ancestryspecific effect177. Further, interactions between rs5770917 and rs1154155 were nonsignificant in Caucasians, Asians and African Americans (OR interaction = 0.88, P = 0.66 in all samples).

The TRA@ locus encodes the α -chain of the TCR $\alpha\beta$ -heterodimer, a protein expressed by T lymphocytes¹⁸. The T-cell receptor is a unique protein that interacts with both HLA class I (CD8 in cytotoxic T cells) and HLA class II (CD4 in helper T cells), including the DQaß heterodimer denoted DQ0602, encoded by HLA-DQB1*0602 and the closely linked HLA-DQA1*0102 allele. The TRA@ locus, like the TRB@ and the immunoglobulin variable heavy and light chain loci, is unusual in undergoing somatic cell recombination. TRA@ and TRB@ recombination occurs in the thymus, resulting, after deletion of autoreactive clones and positive selection, in the generation of T-cell clones with unique TRA@ and TRB@ recombined loci. In the TRA@ locus, recombination occurs between the 5' area of one of the 46 functional variable (V) segments¹⁹ and the 3' area of one of the 49 functional J segments^{20–22}, with additional amino-acid junctional diversity generated by N and P additions in the V-J border region. In the TRB@ locus, diversity is even more complex and generated by recombination of 48V, 2D and 13J segments²². This mechanism produces a diverse repertoire of distinct TCRaß idiotype-bearing T cells²¹, which can be called upon to recognize antigens presented by HLA class I or class II molecules²³.

Unlike most other autoimmune diseases9, narcolepsy is almost completely associated with a single HLA allele, DQB1*0602, across Caucasians, Asians and African Americans⁴. Considering the tight DQB1*0602 association in narcolepsy, it is logical to hypothesize that the DQB0602 heterodimer should interact with a specific TCR $\alpha\beta$ receptor subtype whose occurrence is marked by rs1154155[C], and less strongly by rs17231[T] at both TCR loci. This TCR idiotype would bear specific VJ α and VDJ β recombinants, with recognition of a peptide that also binds DQ0602, mediating further immune reaction leading to the destruction of hypocretin-producing cells. Precisely how a J segment region polymorphism such as rs1154155[C] could increase the risk of occurrence of this narcolepsy associated T-cell clone is unknown, but could involve nonrandom VJ α choices in recombination²¹, as previously reported. Similarly, a polymorphism in the TRB@ V region could influence VDJ recombination for the complementary TCRB chain. Less probably, the TCR-DQ association could also occur without the need for peptide binding, through superantigenlike bridging of TCR and DQ, although most known superantigens interact with TCRB rather than TCRa chains²⁴. Further, superantigen bridging typically results in stimulation of large systemic lymphocyte populations carrying specific TRB@ segments such as that seen in toxic shock syndrome.

Notably, of over ten HLA associated autoimmune diseases that have been subjected to genome-wide analyses and candidate gene

Table 3	Analysis	of rs1154155	genotypes	in	three	replication	cohorts a	and combined

Ancestry	AA Case/Ctrl	AC Case/Ctrl	CC Case/Ctrl	OR _{AC}	OR _{CC}	OR _C
African American	90/117	23/20	0/1	1.50 (0.74,3.04)	0.00 (0.00,22.90)	1.31 (0.68,2.52)
Asian	86/161	296/318	167/120	1.74 (1.27,2.39)	2.61 (1.81,3.76)	1.54 (1.30,1.83)
Caucasian	201/259	132/83	10/6	2.05 (1.45,2.89)	2.15 (0.70,6.77)	1.80 (1.35,2.41)
Three replication samples (MH)			1.83 (1.48,2.27)	2.50 (1.80,3.48)	1.59 (1.38,1.83) ^a	
All samples (MH)				1.94 (1.68,2.25)	2.55 (1.92,3.38)	1.69 (1.52,1.88) ^b

 OR_{AC} is the odds ratio for genotype AC versus AA; OR_{CC} is the odds ratio for genotype CC versus AA; OR_{C} is the odds ratio for allele C versus A. $a\chi^2 = 42.9$, $P = 5.9 \times 10^{-11}$. $b\chi^2 = 94.2$, $P = 2.8 \times 10^{-22}$.

studies, none has shown consistent association with either TCR locus²⁵. Further studies of the TCR loci in narcolepsy may for the first time reveal a role for a specific TCR receptor idiotype in the pathophysiology of an autoimmune disorder.

METHODS

Subjects. Narcolepsy cases were selected as described, 98% of whom are predicted to be hypocretin deficient. The initial sample was comprised of 807 cases and 1,074 Caucasian controls: 415 cases and 753 controls were recruited from the United States and Canada; 392 cases and 321 controls were recruited from European centers.

The Caucasian replication sample contained 718 individuals, of whom 542 were recruited from the United States and Canada (259 cases, 283 controls) and 176 from Europe (104 cases 72 controls). The Asian sample included 866 Japanese (433 cases, 433 controls) and 300 Koreans (128 cases, 172 controls). Finally, we studied 277 African Americans (133 cases, 144 controls). All subjects had given written informed consent approval.

HLA-DQB1*0602 typing. The presence or absence of DQB1*0602 was determined using DQB1 exon 2 sequence-specific primers (Supplementary Table 2 online). These primers amplify DQB1*0602 and a few exceptionally rare DQB1*06 alleles (allele frequency <0.5%) as a 218-bp PCR product. The assay includes a DRB1 internal positive control.

Analysis of Affymetrix data. We obtained Cel file data for all samples and carried out genotyping using the Birdseed-dev algorithm for Affy 6.0 (Affymetrix Power Tools \apt-1.8.5) (n = 1544), and BRLMM for Affy 500K array set chips (n = 337). In each genotype-calling group, individual chips with poor call rates (typically <97%) or high heterozygosity were excluded from further analysis. For each Birdseed calling run, SNPs with call rates <0.9, or Hardy-Weinberg P < 0.01 in controls were excluded. A total of 549,596 SNPs passed all quality control filters and were included in the final analysis. Genotype data was maintained in our database (Progeny Lab 7), and analyses were done using the PLINK software package (v1.04 26/Aug/2008)¹⁴. Interaction studies were conducted in the initial set and in replication sets (cases and controls) using PLINK epistasis, which performs a logistic regression including main genotype effects plus an interaction term.

URLs. Birdseed-dev algorithm, http://www.affymetrix.com/products/software/ specific/birdseed_algorithm.affx; BRLMM, http://www.affymetrix.com/support/ technical/whitepapers/brlmm_whitepaper.pdf; Progeny Lab 7, http://www. progenygenetics.com; PLINK, http://pngu.mgh.harvard.edu/~purcell/plink/.

Note: Supplementary information is available on the Nature Genetics website.

ACKNOWLEDGMENTS

We are most indebted to all the participants of the study, most notably the subjects with narcolepsy. This study was supported primarily by the US National Institutes of Neurological Disease and Stroke grant P50 NS2372. Additional funding included National Institutes of Mental Health R01 MH080957 to E.M., 5U01 MH079470 to D.F.L., 5U01 MH079469-02 to P.V.G., R01 HL62252 to T.Y. Czech Ministry of Education MSM0021620849 and MZO 0002373601 to S.N., National institutes of Allergic and Infectious diseases 5U19 AI063603 to D.S., National Institute of Neurological Disorders and Stroke R01 NS38523 to W.T.L., MIUR PRIN Grant 2005065029 to G.P. and a grant from Grants-in-Aid for Scientific Research on Comprehensive Genomics from the Ministry of Education, Culture, Sports, Science and Technology of Japan to K.T. G.A.R. is supported by the Canadian Institutes of Health Research. Grant MGC 77493 to J. Montplaisir. E.M. is an HHMI supported investigator. We are also grateful to GAIN (the Genetic Association Information Network, NIH) and KORA (Kooperative Gesundheitsforschung in der Region Augsburg, Germany). The KORA research platform was initiated and financed by the German Federal Ministry of Education and Research and by the State of Bavaria. The authors extend their thanks to E. Wan, C. Chu, C. Ha, J. Zhang and A. Voros for technical assistance, and C. Grumet for brainstorming and constant support.

AUTHOR CONTRIBUTIONS

J.H. and J.F. contributed equally to this work. E.M., J.H. and N.R. designed the study. P.-Y.K., L.L., S.H., T.M., J.C., M.A., J.D. and M.K. generated molecular data. J.H., J.F., E.M., N.R., L.L., J.W. and M.K. performed the data analysis. E.M., J.H., J.F. and N.R. wrote the manuscript. E.M., G.M., G.P., S.N., S.H., P.B., Y.H., M.H., B.H., J.M., W.T.L., D.K., M.E., A.D., G.A.R., P.E.H., F.P., B.F., J.-H.J. and S.-P.L. contributed narcolepsy samples. T.G.N.T., L.K., G.T.N., D.S., H.-E.W., G.A.R., C.G., D.E.L., P.V.G., P.P., T.Y., and T.M. provided samples and/or genotypes. E.M. provided financial support.

Published online at http://www.nature.com/naturegenetics/

Reprints and permissions information is available online at http://npg.nature.com/ reprintsandpermissions/

- Mignot, E. Genetic and familial aspects of narcolepsy. *Neurology* 50, S16–S22 (1998).
- Longstreth, W.T. Jr., Koepsell, T.D., Ton, T.G., Hendrickson, A.F. & van Belle, G. The epidemiology of narcolepsy. Sleep 30, 13–26 (2007).
- Juji, T., Satake, M., Honda, Y. & Doi, Y. HLA antigens in Japanese patients with narcolepsy. All the patients were DR2 positive. *Tissue Antigens* 24, 316–319 (1984).
- Mignot, E. *et al.* Complex HLA-DR and -DQ interactions confer risk of narcolepsycataplexy in three ethnic groups. *Am. J. Hum. Genet.* 68, 686–699 (2001).
- Lin, L. *et al.* The sleep disorder canine narcolepsy is caused by a mutation in the hypocretin (orexin) receptor 2 gene. *Cell* 98, 365–376 (1999).
- Chemelli, R.M. *et al.* Narcolepsy in orexin knockout mice: molecular genetics of sleep regulation. *Cell* 98, 437–451 (1999).
- Peyron, C. *et al.* A mutation in a case of early onset narcolepsy and a generalized absence of hypocretin peptides in human narcoleptic brains. *Nat. Med.* 6, 991–997 (2000).
- Thannickal, T.C. *et al.* Reduced number of hypocretin neurons in human narcolepsy. *Neuron* 27, 469–474 (2000).
- Shiina, T., Inoko, H. & Kulski, J.K. An update of the HLA genomic region, locus information and disease associations: 2004. *Tissue Antigens* 64, 631–649 (2004).
- Scammell, T.E. The frustrating and mostly fruitless search for an autoimmune cause of narcolepsy. Sleep 29, 601–602 (2006).
- Overeem, S., Black, J.L. III & Lammers, G.J. Narcolepsy: immunological aspects. Sleep Med. Rev. 12, 95–107 (2008).
- Price, A.L. *et al.* Principal components analysis corrects for stratification in genomewide association studies. *Nat. Genet.* 38, 904–909 (2006).
- Seldin, M.F. et al. European population substructure: clustering of northern and southern populations. PLoS Genet. 2, e143 (2006).
- Purcell, S. et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. Am. J. Hum. Genet. 81, 559–575 (2007).
- Frazer, K.A. *et al.* A second generation human haplotype map of over 3.1 million SNPs. *Nature* 449, 851–861 (2007).
- Ng, M.C. et al. Replication and identification of novel variants at TCF7L2 associated with type 2 diabetes in Hong Kong Chinese. J. Clin. Endocrinol. Metab. 92, 3733–3737 (2007).
- Miyagawa, T. et al. Variant between CPT1B and CHKB associated with susceptibility to narcolepsy. Nat. Genet. 40, 1324–1328 (2008).
- Garcia, K.C. & Teyton, L. T-cell receptor peptide-MHC interactions: biological lessons from structural studies. *Curr. Opin. Biotechnol.* 9, 338–343 (1998).
- Haynes, M.R. & Wu, G.E. Evolution of the variable gene segments and recombination signal sequences of the human T-cell receptor alpha/delta locus. *Immunogenetics* 56, 470–479 (2004).
- Koop, B.F. *et al.* The human T-cell receptor *TCRAC/TCRDC* (C alpha/C delta) region: organization, sequence, and evolution of 97.6 kb of DNA. *Genomics* 19, 478–493 (1994).
- 21. Fuschiotti, P. et al. Analysis of the TCR alpha-chain rearrangement profile in human T lymphocytes. Mol. Immunol. 44, 3380–3388 (2007).
- Lefranc, M.P. et al. IMGT, the international ImMunoGeneTics information system. Nucleic Acids Res. 37, D1006–D1012 (2009).
- Nikolich-Zugich, J., Slifka, M.K. & Messaoudi, I. The many important facets of T-cell repertoire diversity. *Nat. Rev. Immunol.* 4, 123–132 (2004).
- 24. Sundberg, E.J., Deng, L. & Mariuzza, R.A. TCR recognition of peptide/MHC class II complexes and superantigens. *Semin. Immunol.* **19**, 262–271 (2007).
- Lettre, G. & Rioux, J.D. Autoimmune diseases: insights from genome-wide association studies. *Hum. Mol. Genet.* 17, R116–R121 (2008).

Corrigendum: Tiny RNAs associated with transcription start sites in animals

Ryan J Taft, Evgeny A Glazov, Nicole Cloonan, Cas Simons, Stuart Stephen, Geoffrey J Faulkner, Timo Lassmann, Alistair R R Forrest, Sean M Grimmond, Kate Schroder, Katharine Irvine, Takahiro Arakawa, Mari Nakamura, Atsutaka Kubosaki, Kengo Hayashida, Chika Kawazu, Mitsuyoshi Murata, Hiromi Nishiyori, Shiro Fukuda, Jun Kawai, Carsten O Daub, David A Hume, Harukazu Suzuki, Valerio Orlando, Piero Carninci, Yoshihide Hayashizaki & John S Mattick *Nat. Genet.* 41, 572–578 (2009); published online 19 April 2009; corrected after print 26 June 2009

In the version of this article initially published, some author affiliations were incorrectly stated. The error has been corrected in the HTML and PDF versions of the article.

Erratum: Narcolepsy is strongly associated with the T-cell receptor alpha locus

Joachim Hallmayer, Juliette Faraco, Ling Lin, Stephanie Hesselson, Juliane Winkelmann, Minae Kawashima, Geert Mayer, Giuseppe Plazzi, Sona Nevsimalova, Patrice Bourgin, Sheng Seung-Chul Hong, Yutaka Honda, Makoto Honda, Birgit Högl, William T Longstreth Jr, Jacques Montplaisir, David Kemlink, Mali Einen, Justin Chen, Stacy L Musone, Matthew Akana, Taku Miyagawa, Jubao Duan, Alex Desautels, Christine Erhardt, Per Egil Hesla, Francesca Poli, Birgit Frauscher, Jong-Hyun Jeong, Sung-Pil Lee, Thanh G N Ton, Mark Kvale, Libor Kolesar, Marie Dobrovolná, Gerald T Nepom, Dan Salomon, H-Erich Wichmann, Guy A Rouleau, Christian Gieger, Douglas F Levinson, Pablo V Gejman, Thomas Meitinger, Terry Young, Paul Peppard, Katsushi Tokunaga, Pui-Yan Kwok, Neil Risch & Emmanuel Mignot

Nat. Genet. 41, 708–711 (2009); published online 3 May 2009; corrected after print 26 June 2009

In the version of this article initially published, Seung-Chul Hong was incorrectly listed as Sheng Seung-Chul Hong. The error has been corrected in the HTML and PDF versions of the article.