

Nerves in gastrointestinal cancer

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REVIEWS



Nerves in gastrointestinal cancer: from mechanism to modulations

Nathalie Vaes \mathbb{D}^1 , Musa Idris $\mathbb{D}^{1,2}$, Werend Boesmans $\mathbb{D}^{1,3}$, Maria M. Alves \mathbb{D}^2 and Veerle Melotte $\mathbb{D}^{1,2} \boxtimes$

Abstract | Maintenance of gastrointestinal health is challenging as it requires balancing multifaceted processes within the highly complex and dynamic ecosystem of the gastrointestinal tract. Disturbances within this vibrant environment can have detrimental consequences, including the onset of gastrointestinal cancers. Globally, gastrointestinal cancers account for ~19% of all cancer cases and ~22.5% of all cancer-related deaths. Developing new ways to more readily detect and more efficiently target these malignancies are urgently needed. Whereas members of the tumour microenvironment, such as immune cells and fibroblasts, have already been in the spotlight as key players of cancer initiation and progression, the importance of the nervous system in gastrointestinal cancers has only been highlighted in the past few years. Although extrinsic innervations modulate gastrointestinal cancers, cells and signals from the gut's intrinsic innervation also have the ability to do so. Here, we shed light on this thriving field and discuss neural influences during gastrointestinal carcinogenesis. We focus on the interactions between neurons and components of the gastrointestinal tract and tumour microenvironment, on the neural signalling pathways involved, and how these factors affect the cancer hallmarks, and discuss the neural signatures in gastrointestinal cancers. Finally, we highlight neural-related therapies that have potential for the management of gastrointestinal cancers.

With more than 3.6 million diagnoses and 2.2 million related deaths in 2020 worldwide, gastrointestinal cancers (oesophagus, stomach and intestine) represent a major health burden, accounting for 22.5% of all cancer-related deaths worldwide1. Proper management of gastrointestinal cancers is challenging because the gastrointestinal tract is a vibrant and dynamic ecosystem wherein each component — epithelial and various other cell types, luminal contents, the microbiota and extracellular matrix (ECM) — can be an accomplice to the pathogenesis of these cancers, as they serve various biological roles. Collectively, these components constitute the so-called tumour microenvironment (TME)^{2,3}. In the early days of cancer research, scientists believed that a tumour was a homogeneous mass of cancer cells. However, a plethora of long-term and in-depth investigations uncovered that tumour development closely resembles normal organogenesis and should be regarded as a heterogeneous entity resulting from the dynamic, reciprocal interactions between cancer cells and their surrounding microenvironment^{4,5}. Moreover, the coordinated cellular and molecular processes that enable the body to manage its homeostatic balance will be governed by the tumour itself^{4,6}. A well-understood phenomenon driving this feature is the ability of tumours to foster the

formation of blood and lymph vessels by which necessary nutrients are received and by which spreading to distant sites is conferred^{4,6}. To this end, the 'established' TME strongly influences the behaviour of a primary tumour, determines whether it will disseminate to other organs and affects responses to therapy^{6,7}. Whereas immune, endothelial and cancer (epithelial) cells have been in the spotlight for many years, justifying the development of targeted therapies, the role of the nervous system only entered the limelight in the past couple of years, which is remarkable given the extensive innervation of the gastrointestinal tract.

The gastrointestinal tract is innervated by the three main divisions of the autonomic nervous system, which provide both extrinsic and intrinsic neural control of gut function^{8,9}. The parasympathetic and sympathetic nervous systems supply the gastrointestinal tract with extrinsic innervation, that is, parasympathetic and sympathetic neurons have their cell bodies outside the gut wall, whereas cells of the enteric nervous system (ENS) are located within the bowel itself, providing intrinsic control of gut function (FIG. 1 and BOX 1). In mammals, the ENS forms an intricate neural network embedded along the gastrointestinal wall that consists of different types of enteric neurons and glia, which are

Department of Pathology, GROW-School for Oncology and Reproduction, Maastricht University Medical Centre, Maastricht, Netherlands.

²Department of Clinical Genetics, Erasmus University Medical Centre, Rotterdam, Netherlands.

³Biomedical Research Institute (BIOMED), Hasselt University, Hasselt, Belgium.

™e-mail: veerle.melotte@ maastrichtuniversity.nl https://doi.org/10.1038/s41575-022-00669-9

Key points

- The gut is a highly innervated, dynamic ecosystem wherein nerves are key for intestinal functioning and homeostasis by communicating with a variety of cell types and the gut microbiome.
- Neural contributions to gastrointestinal cancers represent a flourishing area of investigation as both intrinsic and extrinsic nerves influence gastrointestinal tumorigenesis via their interplay with cancer cells.
- Neural-related signals and pathways can influence the cancer hallmarks, interfering
 with several cancer cell characteristics (metabolism and (epi)genomic stability)
 and/or supporting a cancer-promoting microenvironment (immune infiltration,
 extracellular matrix).
- While neoneurogenesis and axonogenesis are emerging within the gastrointestinal cancer field, both topics require in-depth investigation to identify their exact origin and driving mechanisms.
- Cancer cells are able to hijack (embryonic) neural pathways to promote their own fitness.
- Targeting neural cell-derived messengers and their respective receptors holds great promise in the treatment of gastrointestinal cancers.

predominantly clustered within interconnected ganglia of the submucosal and myenteric plexus¹⁰. Although the inner workings of the gut are modulated by the ENS, extensive extrinsic inputs fine-tune gut function¹¹. By collaborating with a variety of cell types, such as smooth muscle cells and interstitial cells of Cajal as well as epithelial, endothelial and immune cells, the enteric neuron-glial network controls motility patterns, supports mucosal barrier function, aids in immunological defence and controls mucosal secretions12. The ENS has been implicated in the pathogenesis of various gastrointestinal disorders, with an obvious role in the aetiology of enteric neuropathies¹³. Moreover, increased understanding of its contribution to several other diseases, including those associated with intestinal inflammation, is currently arising14. Despite the recognized importance of neurons in cancer4, the role of the nervous system in gastrointestinal carcinogenesis is only now emerging^{15,16}.

In this Review, we focus on the blossoming area of neural contributions to the pathogenesis of gastrointestinal cancers. We address how neurons interact with the gut microbiota and various TME components and how these interactions affect the hallmarks of cancer. Neural and neural-related intracellular signalling pathways and the neural signature in gastrointestinal cancers are also highlighted. Supported by the resulting insights, we discuss current and potential future therapeutic strategies.

Crosstalk between cancer and neural cells

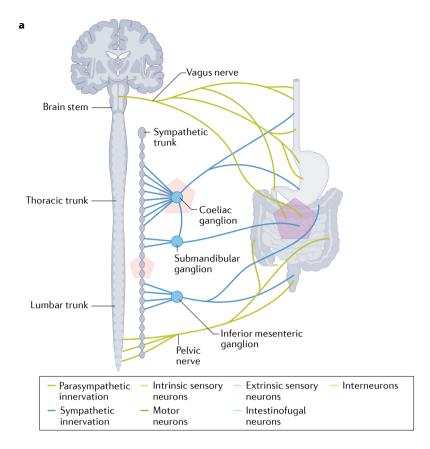
In the past decade, several landmark papers have shown that tumour innervation promotes the malignant phenotype of cancer cells. For oesophageal, stomach and colorectal cancers, it has been established that perineural invasion (BOX 2) is an independent prognostic factor associated with worse prognosis and poor clinical outcome^{17–22}. Similarly, axonogenesis (that is, the formation of new axons or increased nerve density^{23,24}) and (neo)neurogenesis (that is, de novo formation of nerve cells^{25,26}) confer tumours a growth advantage. From the early 2000s onwards, pioneering in vivo studies started to use a variety of tools to interfere with tumour innervation

and explore their mechanistic and functional importance for the aetiology of gastrointestinal cancers.

Denervation

Both surgical cutting of nerves and injection of neurotoxic drugs have been used to study the effect of denervation in cancer. In humans, extrinsic vagal denervation, termed vagotomy, either unilateral or bilateral, was first used in the management of gastric and duodenal ulcers^{27,28}. This treatment seemed to increase the risk of developing gastric cancer due to delayed gastric emptying and concomitant hypochlorhydria, which could increase the levels of N-nitroso compounds²⁹⁻³¹. Similarly, unilateral vagotomy increased the numbers of gastric tumours that were formed upon treatment of Wistar rats with the nitrosoguanidine derivative N-methyl-N-nitro-N-nitrosoguanidine (MNNG)32. However, later papers have shown that, for gastric and small intestinal cancers, vagal denervation has antitumorigenic effects. Using a spontaneous (Ins-Gas), carcinogen-induced (MNU) and bacteria-infected (Helicobacter pylori–H⁺/K⁺–ATPase-Il1β) mouse model of gastric cancer, Zhao et al. demonstrated that surgical denervation (bilateral or unilateral truncal vagotomy) or a botulin toxin injection (which blocks acetylcholine release from axon endings) suppressed tumorigenesis²³. In agreement, gastric cancer is more prevalent in the lesser as opposed to the greater curvature in mice and humans^{33,34}, which parallels the higher nerve density in the lesser curvature²³. The discrepant outcomes between earlier and later studies are most likely explained by the fact that later studies applied vagotomy with an accompanying drainage procedure (pyloroplasty) to improve gastric emptying³¹. In a genetic intestinal cancer mouse model (APC^{Min/+}), tumour development in the small intestine was suppressed by subdiaphragmatic vagotomy³⁵. Interestingly, for colorectal cancer (CRC), truncal vagotomy with accompanying drainage procedure had no clear effect on carcinogenesis in rats36,37 whereas, in humans, this treatment for gastric or duodenal ulcers increased the risk of developing CRC across a range of studies of varying cohort sizes^{38–40}. Further research is needed to specify if this increased risk results from delayed emptying and increased levels of gastrin, because these findings were observed in patient cohorts treated with different gastric surgeries with or without accompanying drainage.

Conflicting data have also been reported following surgical and pharmacological sympathetic denervation. The most widely used compound to achieve chemical sympathectomy is 6-hydroxydopamine, a neurotoxin that destroys catecholaminergic neurons at the injection site. Using Wistar rats, Tatsuta et al. found that prolonged administration of 6-hydroxydopamine markedly reduced azoxymethane-induced colonic tumour incidence as well as gastric tumorigenesis (MNNG-induced)41,42. Even though Sadighparvar et al. observed substantially fewer aberrant crypt foci following sympathetic denervation (coeliac-mesenteric ganglionectomy and guanethidine sulfate administration) in Wistar rats during 1,2-dimethylhydrazine (DMH)induced carcinogenesis, no effect on colon tumour incidence or size was observed⁴³. Surgical sympathectomy



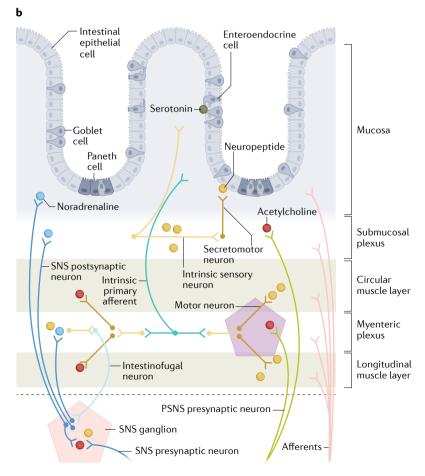


Fig. 1 | Nervous signalling paths and modulators of the gastrointestinal tract in mammals. Each part of the gastrointestinal tract (that is, oesophagus, stomach, small and large intestine) is differentially innervated, both extrinsically and intrinsically, a | Extrinsic innervation of the gastrointestinal tract. Parasympathetic (cholinergic) innervation (green trajectories) via the vagus nerve is very dense at the beginning of the gastrointestinal tract, yet becomes sparse whilst moving distally, with most parasympathetic innervation of the distal colon originating from preganglionic neurons within S1-S4 lumbosacral spinal cord regions. Parasympathetic preganglionic neurons synapse with myenteric postganglionic neurons in the gastrointestinal tract (purple pentagon). Sympathetic (adrenergic) innervation (blue trajectories) varies per organ of the gastrointestinal tract, with the cervical and thoracic trunk from spinal segments T1-T10 innervating the oesophagus, T6-T9 and T9-10 thoracic neurons supplying the stomach and small intestine, respectively, and L2-L5 lumbar regions innervating the colon. Sympathetic preganglionic neurons synapse with postganglionic neurons in the prevertebral ganglia or, yet less likely, in the paravertebral ganglia (rose pentagons). Visceral afferent fibres travel along the vagus and spinal nerves as well as the sympathetic nerves and transduce sensory signals from the enteric nervous system (ENS) (not shown). **b** | Intrinsic innervation and major synapses in the gastrointestinal tract. Schematic overview of a variety of neurotransmitters and synapses in the ENS with a specific focus on those that have been shown to be involved in gastrointestinal carcinogenesis. The ENS is embedded along the entire gastrointestinal wall with a variable design depending on the intestinal segment. The myenteric plexus continues from the upper oesophagus to the internal anal sphincter, whereas the submucosal plexuses are absent in the oesophagus, contain few ganglia in the stomach, and are only fully established in the small and large intestine. ENS neurons are depicted in various colours to distinguish subpopulations (see key). Enteric glial cells are not shown in this figure. PSNS, parasympathetic nervous system; SNS, sympathetic nervous system.

of the small intestine did not affect *Apc*-driven intestinal carcinogenesis in mice³⁵. Compared with normal tissue, human gastric^{44–46} and colon cancer⁴⁷ tissues were characterized by markedly reduced sympathetic nerve fibres or noradrenaline levels, which were gradually restored with increasing distance from the tumour site. However, these conclusions were drawn from studies with small patient cohorts (82 and 5, respectively) and do not provide conclusive data regarding potential positive effects of sympathetic innervation on gastrointestinal carcinogenesis.

For intrinsic denervation in animal models, benzalkonium chloride is the chemical that has mostly been used to locally damage the myenteric plexus⁴⁸. Whereas application of benzalkonium chloride to the serosal surface of the gastrointestinal tract initially induces hyperplasia of gastric G cells, oesophageal cells and descending colonic epithelial cells^{49–51}, it hampers gastric and colon tumour development and growth in an MNNG and DMH rat model of gastric⁵² and colon cancer⁵³, respectively. This finding could be mediated by an imbalance in the levels of several neural factors as well as by the interplay between benzalkonium chloride, the acidic environment and various neuropeptides. Upon

Box 1 | Innervation of the gastrointestinal tract

The nervous system in vertebrates is built from two main parts: the central nervous system and the peripheral nervous system, with the first comprising the brain and spinal cord, and the latter consisting of neural cells located outside of the brain and spinal cord. The peripheral nervous system can be separated into the somatic and autonomic nervous systems, with the autonomic nervous system further subdivided into the parasympathetic (FIG. 1a, green trajectories), sympathetic (FIG. 1a, blue trajectories) and enteric nervous systems ¹².

Acetylcholine is the primary neurotransmitter to relay signals in parasympathetic preganglionic and postganglionic synapses as well as from preganglionic sympathetic fibres. Noradrenaline, on the other hand, is the main neurotransmitter released from postganglionic sympathetics (FIG. 1b). The native receptors via which acetylcholine and noradrenaline exert their actions are muscarinic or nicotinic cholinergic receptors, and the α -adrenoceptor and β -adrenoceptor, respectively. Acetylcholine released within ganglionic synapses primarily activates nicotinic receptors on postganglionic neurons to transmit autonomic signals from preganglionic to postganglionic neurons. Postganglionic neurons extend further to visceral organs, including the gastrointestinal tract, where parasympathetic acetylcholine binds muscarinic receptors (M_1-M_5) and sympathetic noradrenaline binds α -adrenergic receptors $(\alpha_1$ and $\alpha_2)$ and β -adrenergic receptors (especially β_1 and β_3) on target gastrointestinal cells, including epithelial, immune and other stromal cells. Next to neuronal sources of these neurotransmitters, systemic adrenaline produced by adrenal glands or members of the tumour microenvironment can reach target cells in the gastrointestinal tract 286 .

Sensory information is conveyed via visceral afferents that make up most of the extrinsic gastrointestinal innervation and largely follow autonomic fibres to reach the central nervous system following two distinct routes. Splanchnic and pelvic afferents, which have their cell bodies in thoracolumbar and lumbosacral dorsal root ganglia, mainly follow the sympathetic and parasympathetic chain, respectively, to enter the brain via the spinal cord. The nodose or jugular ganglia harbour cell bodies of vagal afferents and project to the nucleus tractus solitarius within the brainstem^{287–290} (FIG. 1a). The enteric nervous system harbours region-specific circuits that integrate information from the gut lumen and gut wall via intrinsic primary afferent neurons, interneurons and motor neurons to regulate gut motility and secretomotor and vasomotor responses 287,291,292 (FIG. 1b). In addition, intestinofugal neurons project outside the gut wall and convey afferent information to sympathetic prevertebral ganglia²⁹². Enteric neuron subtypes communicate with one another and with target cells, using neurotransmitters, such as acetylcholine nitric oxide and serotonin, as well as neuropeptides such as neuropeptide Y, substance P and vasoactive intestinal peptide. Collectively, these messengers assist in controlling gastrointestinal homeostasis and motility^{293,294} (FIG. 1b).

> benzalkonium chloride treatment, levels of substance P and vasoactive intestinal polypeptide (the two main neuropeptides driving MNNG-induced carcinogenesis) decrease, leading to an inflamed and acidic gastric environment, which reduces the effectiveness of MNNG52,54. Correspondingly, benzalkonium chloride-induced denervation of the rat oesophagus⁵¹ and distal colon⁵⁵ is associated with megaoesophagus and megacolon, with the latter condition also hindering colorectal tumorigenesis in humans⁵⁶. By contrast, a small increased risk of developing oesophageal cancer has been observed in several cohorts of patients with megaoesophagus⁵⁷. Given that Munari et al. ascribed this risk to prolonged contact with food, microbial overgrowth and increased nitrate levels⁵⁷, it is questionable whether the absence of nerves is really accountable for the discrepant gastrointestinal cancer risks rather than the influence of the distended gut wall or obstructed luminal content on the carcinogenic process. To accommodate for this aspect, Zhao et al. used a mouse gastric model wherein they performed denervation after completion of the cancer induction protocol. The finding that vagotomy also inhibited gastric tumorigenesis in this model suggests that this is a nerve-driven inhibitory effect²³.

Before these denervation studies can be translated to the patient, it is important to fully understand the neuroanatomy and biology during carcinogenesis, which is currently far from being fully established. The manner (surgical versus chemical), location, period (short term versus long term) and timing (before or during cancer onset or progression) of the employed interference represent fundamental denominators for the experimental outcomes and could account for the discrepancies that have been described. For instance, although it seems that, especially in the human situation, denervation before the onset of carcinogenesis (for example, treatment of ulcers) inhibits tumour formation, Zhao et al. demonstrated equally positive effects of denervation before or after the establishment of gastric cancer on tumour incidence and progression in mice²³. Moreover, the various techniques described have limitations due to their incomplete denervation, subsequent nerve regeneration or unintended secondary effects on other members of the TME. In effect, the observation that parasympathetic but not sympathetic denervation reduces intestinal carcinogenesis in the ApcMin/+ mouse model might be explained by the difference in complete parasympathetic versus partial sympathetic denervation of the small intestine35,58,59.

Modulation of neural factors

Several neurotransmitters, neurotrophic factors and their respective receptors have been shown to regulate intestinal epithelial growth and proliferation and are commonly overexpressed in gastrointestinal cancer tissue¹⁵. As a consequence, multiple in vitro and in vivo studies have applied a variety of techniques, such as knockdown, overexpression, blocking or (ant)agonizing, to assess the influence of neurotrophin and neurotransmitter signalling on gastrointestinal carcinogenesis. Neurotrophic factors are defined as endogenous molecules that regulate survival, growth and morphological plasticity of neurons. Considering their name, neurotrophic molecules were initially thought to be uniquely related to the nervous system. However, since the mid-1900s, it has been shown that other cells, including cancer cells, also possess the machinery to produce, secrete and respond to neural factors60. Interestingly, different receptor subtypes have been identified on a variety of cell types within the gastrointestinal tract. However, the implication in gastrointestinal carcinogenesis is restricted to the muscarinic M₃R receptor and, to a lesser extent, the M₁R receptor as well as the α_2 -adrenoceptor, β_2 -adrenoceptor and β₃-adrenoceptor for neurotransmitters and the Trk and p75NTR receptors for neurotrophins 16,61. A comprehensive overview of the main neurotrophic factors and neurotransmitters involved in gastrointestinal carcinogenesis is provided elsewhere^{15,62}; however, within these studies, the origin of these messengers was often not defined. In the following section, we elaborate on the neuronal cell-derived messengers and their respective receptors on cancer cells.

Cholinergic signalling in gastrointestinal cancers. During gastric tumorigenesis, cholinergic nerve density increases with tumour progression and both mouse and human oesophageal, gastric and colon cancer cells upregulate the expression of $M_3R^{23,63-67}$ (FIG. 2). Binding of acetylcholine to M_3R activates intracellular EGFR–ERK–AKT signalling in gastric and colon cancer cells in vitro 68,69 and in vivo (azoxymethane-induced CRC mouse model) 70 . M_3R receptor activation also induces ligand-independent Wnt signalling through YAP in vivo (MNU gastric mouse model), with a concomitant upregulation of, amongst other Wnt targets, CD44 and Lgr5 as well as nuclear translocation of β -catenin. Both pathways trigger proliferation and expansion of the gastrointestinal epithelium $^{63,68-72}$. In addition, activating the M_3R -AMPK–MACC1 and matrix metalloproteinase (MMP) pathway by acetylcholine promotes

Box 2 | Perineural invasion in gastrointestinal cancers

One of the key factors affecting the aggressiveness of a cancer cell is its ability to disseminate and migrate towards remote tissue sites. Although lymph and blood vessels are considered the primary routes for cancer cells to migrate, metastatic spread via nerve fibres was identified in the mid-1800s and termed perineural invasion (PNI). To date, PNI has been recognized as an important hallmark and a prognostic feature for different cancer types. Even though it is well established that this process comprises neoplastic spreading via nerves, a universal definition for PNI is still lacking.

Originally, PNI was thought to be a passive process, being defined as "silent extensions of malignant cells along the nerve sheath" by Mohs and Lathrop in 1952 (REF.²⁹⁵). Later on, Batsakis broadly defined PNI as "tumour cell touching and invasion in, around and through nerves"296. This definition covers all of the histopathological characteristics that have been observed for PNI, yet might vary according to subjective interpretations. The following suggestion by Veness to cite PNI "only when tumour cells are able to invade the perineurium" seems too stringent given that the nerve sheath consists of three connective tissue layers: epineurium, perineurium and endoneurium²⁹⁷. Consequently, to further limit variable interpretations of this definition, different papers (Fagan et al.²⁹⁸, Bockman et al.²⁹⁹ and Nagakawa et al.³⁰⁰) have described that 'at least 33% of the nerve circumference should be surrounded by tumour cells' to classify it as PNI. Liebiq and colleagues then advocated to define PNI as tumour cell touching and invasion in, around and through nerves, with tumour cells in any of the three nerve sheath layers, or involving at least 33% of the nerve circumference³⁰¹. Importantly, it has become apparent that PNI is not a simple, passive process, but rather an active process involving reciprocal communication between nerves and tumour cells via paracrine signalling. Such a mechanism was first described by Ayala et al. using mouse dorsal root ganglia and prostate cancer cells co-cultured in a Matrigel matrix302.

For gastrointestinal cancers specifically, the prevalence of PNI varies from 4% to 76%, depending on the specific gastrointestinal site and the definition that has been used^{18,19,22,303,304}. Nevertheless, it has been identified as a prognostic factor associated with poor prognostic outcomes in a variety of gastrointestinal cancer types. For oesophageal cancer, PNI was found to be an independent predictor for shorter disease-free and disease-specific survival^{17,18}. Similarly, high levels of PNI have been observed in gastric cancer, corresponding with disease progression and also predicting gastric cancer recurrence in patients who underwent curative resection 19,20. For colorectal cancer, PNI has been characterized as a prognostic marker associated with a worse clinical outcome because of increased local recurrences and shorter 5-year disease-free survival. Its prognostic value reportedly compares with that of other well-established markers such as differentiation grade and depth of invasion^{21,22}. Moreover, work by Duchalais et al. provided a mechanistic understanding of how cancer cells adhere to nerves: with tumour epithelial cells preferentially and directly adhering to enteric neurons via N-cadherin and L1CAM, predominantly at the invasive front, and faithfully following the neural trajectory³⁰⁵.

These findings underscore that PNI should be respected as a high-risk feature and supports its implementation in the standardized reporting criteria of (gastrointestinal) cancers. However, determination of the PNI pattern can be challenging in clinical practice due to potential subjective pathological interpretation among certified pathologists and the requirement of deeper sections for immunohistochemistry. Thus, the identification of proper neural-related biomarkers for diagnostic purposes would be desirable 306-308.

gastric and colon cancer migration and invasion in vitro⁷³ and in vivo^{74,75}. Progression towards a cancerous gastrointestinal epithelium reinforces bidirectional crosstalk as human gastric and colon (cancer) cells can synthesize and release acetylcholine, which subsequently stimulates neighbouring nerve fibres and axonogenesis^{63–65,69,76,77}.

Adrenergic signalling in gastrointestinal cancers. Given that adrenergic stimulation, for example, via α₂-adrenergicinduced EGFR-MEK-ERK signalling, in the gastrointestinal epithelium supports cell migration and wound healing^{73,78}, it is not surprising that dysregulation of these signalling cues is involved in the pathogenesis of gastrointestinal cancers (FIG. 3). Moreover, adrenergic signals and receptors are commonly overexpressed within cancer tissues, further suggesting that adrenergic signals take part in gastric⁷⁹ and colorectal⁸⁰ tumorigenesis. According to several in vitro and in vivo studies^{79–86}, the carcinogenic influences are mostly mediated by β_2 -adrenoceptor and α_2 -adrenoceptor, attenuating critical cell functions such as apoptosis and immune responses, inducing epithelial-mesenchymal transition, and promoting metastatic and invading capacities. Both in vitro^{79,84} and in vivo⁷⁹ studies have shown that this process is mediated via activation of the VEGF-MMP and STAT3-ERK-MAPK pathways. For instance, noradrenaline promotes cell survival through AMPK-dependent autophagy in gastric cancer cell lines⁸⁵. Similarly, β_3 -signalling enhances the survival of gastric cancer xenografts, whereas pharmacological blocking of the β_2 -adrenoceptor promotes apoptosis of these xenografts (propranolol induced)79 and of CRC cells in vitro86. In agreement, elevated levels of the β_2 -adrenoceptor have been identified as a clinically significant prognostic marker for CRC in humans80. Notably, the studies described mainly focus on extrinsic innervation and no knowledge is available on the ENS.

Targeting neurotrophin signalling. As mentioned earlier, acetylcholine can immediately fuel a reciprocal nerve-cancer communication by activating M3R on epithelial cells. This process stimulates nerve growth factor (NGF) production, which subsequently targets its respective TrkA receptor on nerves and triggers cholinergic neurite growth⁶³. Functional blocking or inhibition of either M₃R or TrkA receptors reduces epithelial cell proliferation and tumour innervation in mouse gastric cancer models, thereby emphasizing the contribution of neurotrophic signals to the pathogenesis of gastric cancer⁶³. Likewise, oesophageal carcinogenesis can be enhanced by neurotrophic factors and their receptors. Human oesophageal cancer tissues overexpressing NGF are characterized by the presence of nerve bundles and neuropeptide-immunoreactive nerve fibres expressing the TrkA receptor 17,87. Moreover, oesophageal cancer cells undergo apoptosis upon silencing of the nonspecific neurotrophin receptor p75NTR (REF.88). However, in CRC, this receptor is often silenced in human tissues to counteract its tumour suppressive effects, that is, inhibiting proliferation and promoting apoptosis in vitro^{89,90}.

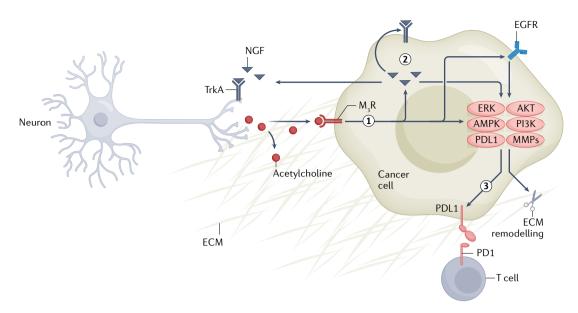


Fig. 2 | Cholinergic signalling in gastrointestinal cancers. Gastrointestinal cancer cells can upregulate the expression of muscarinic M_3R receptors. (1) By binding to these receptors, acetylcholine activates intracellular EGFR–ERK–AKT signalling as well as matrix metalloproteinases (MMPs), which promotes cancer cell invasion and migration. (2) M_3R activation can also stimulate nerve growth factor (NGF) production, which subsequently targets its respective TrkA receptor on nerves and triggers cholinergic neurite growth. (3) In addition, cholinergic signalling induces the expression of PDL1 ligands in colon cancer cells. ECM, extracellular matrix.

Similar to acetylcholine, noradrenaline also directly nourishes bidirectional crosstalk as its binding to the β_3 -adrenoceptor increases the levels of brain-derived neurotrophic factor (BDNF), which consequently stimulates axonogenesis and tumour growth and/or progression via its native NTRK2 (also known as TrkB) receptor on nerve cells^{79,91-93}. BDNF-TrkB signalling can also instantaneously modify cancer cell characteristics, favouring malignant progression in vitro^{94,95}. This aspect is achieved by transactivation of the EGF pathway on cancer cells, thereby promoting the proliferation of small and large intestinal cancer cell lines94,95, a mechanism that confers resistance to EGFR inhibitors in CRC96,97. Moreover, this pathway activates ERK and AKT signalling92,98, which leads to the upregulation of anti-apoptotic proteins, such as BCL-2, in vitro. Such survival effect is abolished by BDNF knockdown in CRC cell lines99. These data might justify studying the benefits of therapeutic co-administration of TrkB and EGFR inhibitors in patients with CRC.

Taken together, these studies emphasize that a variety of neural signalling mechanisms fuel the interaction between the nervous system and intestinal epithelium. However, even though some mechanistic insights have established how cancer cells regulate nerve recruitment, we need to better understand which cells secrete which neural factors and which receptor is targeted. Interestingly, this nerve dependence in cancer follows a long-reported nerve dependence during tissue regeneration and tissue remodelling, and common cellular and molecular mechanisms have been described. Here, outgrowth and infiltration of nerves have also been observed, and the release of transferrin¹⁰⁰, neuregulin¹⁰¹ and the neurotransmitter substance P¹⁰² by nerve endings can rescue the regeneration capacity in

several experimental degeneration models; factors for which also an oncogenic potential has been described. Although the concept of neoneurogenesis during cancer development and/or progression is discussed in several papers, this aspect remains mainly a 'black box' in gastrointestinal cancers. It has been described in vivo that neural progenitor cells from the brain escape the blood-brain barrier to colonize prostate cancer²⁸ and that cancer stem cells have the potential to differentiate into neural-like cell populations both in vitro and in vivo^{25,26}. However, these findings need more in-depth investigation, particularly in the context of gastrointestinal cancers. In fact, as the cancerous gastrointestinal tract contains cells with neurogenic potential¹⁰³, future studies need to define whether they contribute to neoneurogenesis in gastrointestinal cancers.

Neural cells, the TME and hallmarks of cancer

In 2000, Hanahan and Weinberg introduced the hallmarks of cancer, a rationalizing set of six acquired biological properties that govern neoplastic transformation¹⁰⁴. A decade later, they revised this list, resulting in ten hallmarks: "sustaining proliferative signalling", "evading growth suppressors", "resisting cell death", "enabling replicative immortality", "inducing angiogenesis", "activating invasion and metastasis", "tumor-promoting inflammation", "genome instability and mutation", "deregulating cellular energetics" and "avoiding immune destruction"6. As reasoned earlier, the interaction between nerves and their environment clearly influences the survival, growth and dissemination of tumour cells. Based on these findings, Senga and Grose recommended appending "neuronal signalling" to the hallmarks, in addition to "dedifferentiation and transdifferentiation", "epigenetic dysregulation" and "altered microbiome"

(discussed later)¹⁰⁵. Although limited data are available on how the nervous system influences cancer hallmarks with respect to gastrointestinal cancers, in the following sections, we discuss studies that provide evidence on the role of neural signalling in this context (FIGS. 2–4).

Angiogenesis and the neuroendothelial unit

In 1971, Folkman reported angiogenesis as an important trait of tumours as it enables the supply of necessary nutrients to ensure tumour cell survival¹⁰⁶. This tumour-associated neovasculature was initially thought to be vital only for aggressive and rapidly growing tumours. Yet, nowadays, angiogenesis has a full-standing status within the hallmarks of cancer as it also contributes to the initial cancer stages⁶. Neuroangiogenesis refers to the process wherein nerves bundle along blood vessels, which is key for organ development and wound healing but also seems to contribute to the carcinogenic process¹⁰⁷⁻¹¹⁰. Within this partnership, endothelial cells are partially guided by neural signals and vice versa; that is, angiogenic factors promote neural growth^{79,111,112}. During prostate cancer, it is well established that adrenergic cues (such as noradrenaline), induce an 'angiogenic switch' in endothelial cells, thereby tuning the initiation and patterning of angiogenesis in vivo¹¹³. Such mechanism is not well defined for gastrointestinal cancers yet is supported by the fact that activation of either β-adrenergic receptors or exogenous administration of (nor)adrenaline upregulate levels of angiogenic factors such as VEGF, MMPs and IL-8 in CRC and gastric cancer both in vivo and in vitro — an effect that is abolished by blocking the β_2 -adrenoceptor^{79,114}. Similarly, the inhibition of β_1 -adrenergic receptors suppresses endothelial cell proliferation via inhibition of the glycolytic flux, which limits tumour formation of CRC cells orthotopically injected in the caecum of immunocompromised mice111. Again, this current knowledge is restricted to extrinsic innervation; however, exogenously added serotonin could also exert pro-angiogenic effects by activating endothelial cells directly or by inhibiting expression of the angiogenic inhibitor MMP12 in tumour-infiltrating macrophages within colon cancer allografts¹¹⁶. By contrast, dopamine has an antiangiogenic effect by binding D2 receptors on endothelial cells, leading to the suppression of VEGFR2 phosphorylation and downstream MAPK and FAK signalling in colon and gastric cancer-bearing rodents, thereby inhibiting endothelial cell proliferation and migration whilst promoting tumour cell apoptosis 117-120. Thus, specific studies of the ENS in this regard are needed.

Activating invasion and metastasis

Structural and compositional changes of the ECM scaffold within the TME have a leading role in tumour growth and metastasis⁴. Initially, the ECM provides a physical barrier that limits tumour development and prevents immune cell infiltration^{4,121}. Several neurotrophic factors, neurotransmitters and neuropeptides

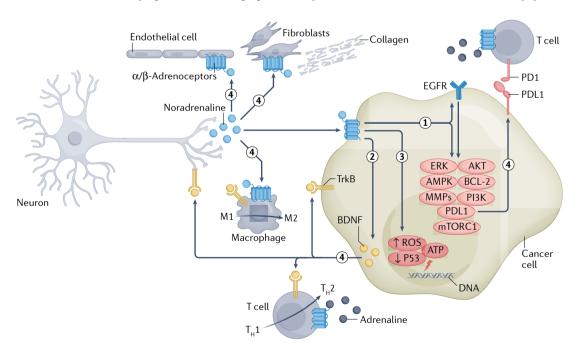


Fig. 3 | Adrenergic signalling in gastrointestinal cancers. (1) Signalling via α -adrenoceptors and β -adrenoceptors activates the VEGF-matrix metalloproteinase (MMP) and STAT3-ERK-MAPK pathways, attenuating apoptosis and immune responses, inducing epithelial-mesenchymal transition and promoting metastatic capacities of the tumour. (2) β_3 -Adrenoceptor activation increases the levels of brain-derived neurotrophic factor (BDNF), which stimulates axonogenesis. BDNF-TrkB signalling transactivates the EGF pathway on cancer cells. (3) Adrenergic signalling stimulates the degradation of p53, thereby leading to the accumulation of DNA damage in cancer cells. Upregulation of BCL-2 inhibits their apoptosis. (4) Moreover, β -adrenergic signalling increases type I collagen fibres via activation of cancer-associated fibroblasts, induces an angiogenic switch in endothelial cells, mediates the transformation of M1 (pro-inflammatory) to M2 (anti-inflammatory) macrophages, shifts cytokine production from T helper 1 (T_H 1) to T_H 2 pattern, controls lymphocyte trafficking, and blocks effector CD8+T cell activation through PDL1 upregulation. ROS, reactive oxygen species.

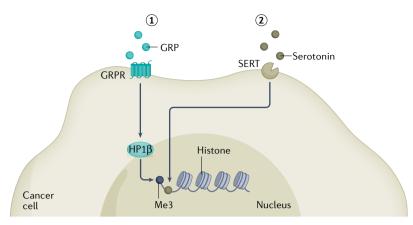


Fig. 4 | Neural-related factors alter the epigenome in gastrointestinal cancer cells. (1) Gastric-releasing peptide (GRP) upregulates the heterochromatin protein 1β (HP1 β) in colon cancer cells. HP1 β mediates chromatin condensation and gene silencing stabilization. (2) Serotonylation of the glutamine residue on histone 3 H3Q5 stabilizes H3K4 methylation. GRPR, gastric-releasing peptide receptor.

upregulated during carcinogenesis can introduce changes within this scaffold in favour of tumour progression and spreading, in particular by directly activating MMPs or indirectly triggering TME cells that actively produce and remodel the ECM (FIGS. 2 and 3).

In accordance with its overexpression in human gastric cancer and CRC cells and tissues^{23,63,122}, expression levels of M₂R have been shown to correlate with cancer stage as well as lymph node metastasis in gastric cancer⁶⁴. Activation of M₃R on cancer cells in vitro results in the upregulation of several MMPs, including MMP1, MMP7 and MMP10, which facilitate CRC cell migration and invasion; this effect can be reversed by blocking MMP1 activation^{74,75,122-124}. Additionally, MMP2, MMP7 and MMP9 levels are increased in human gastric cancer and oesophageal cancer tissues125-127 but their potential regulation by neural factors is currently not established. Furthermore, (nor)adrenaline can also stimulate migratory capacities of gastric cancer xenografts⁷⁹ and colon cancer cells in vitro, respectively, which can be inhibited by the β -adrenergic blocker propranolol¹²⁸. Interestingly, dependent on which receptor NGF activates, contradictory effects on ECM remodelling have been reported. TrkA activation facilitates CRC cell migration and invasion in vitro via MAP-ERK signalling and enhances MMP2 and MMP9 activity¹²⁹. By contrast, NGF signalling via p75NTR suppresses gastric cancer cell metastasis by attenuating urokinase-type plasminogen activator (a cell motility factor) and MMP9 levels, whilst increasing levels of the tissue inhibitor of matrix metalloproteinase 1 (TIMP1)130. This finding is reflected by upregulated p75NTR levels in non-metastatic compared with metastatic human gastric cancer tissues¹³⁰. During gastric and colon cancer, the BDNF-TrkB pathway promotes invasion and suppresses anoikis in vitro and in vivo (intravenously injected), presumably by boosting the epithelial-mesenchymal transition 94,95,131. Finally, in gastric xenografts, inhibition of TrkB receptors proved sufficient to abrogate this effect131.

Besides MMPs, noradrenaline can cause an increase in type I collagen fibres via activation of cancer-associated

fibroblasts (CAFs)2,132. According to evidence in experimental models, this step favours tumour progression, in part by supporting migration of blood vessels and nerve fibres and paving the way for tumour cells to invade^{4,133}. CAFs are one of the most dominant members of the TME, representing a heterogeneous group of activated fibroblasts that are involved in several hallmarks of cancer. Their extensive functions in gastrointestinal cancers are comprehensively reviewed elsewhere 134-136. Importantly, in addition to CAFs, enteric neurons themselves are also capable of synthetizing and secreting ECM molecules as first evidenced by the Goldstein group^{137,138}. In 2021, Vaes et al. also uncovered that mature enteric neurons can secrete ECM molecules, such as Nidogen 1 and Fibulin 2, thereby promoting colorectal carcinogenesis¹³⁹. Furthermore, myenteric denervation by benzalkonium chloride administration has been shown to be associated with an increased frequency of reticular and elastic fibres within the non-cancerous gastric mucosa, whereas it shifts the fibrillary component towards more elastic fibres in gastroadenocarcinomas (benzalkonium chloride plus MNNG rat model)¹⁴⁰. Such observation correlates with increased expression of elastin in human colorectal tumour tissues and cancer cells¹⁴¹. This finding suggests that, in this situation, elastic fibres have protective effects and that degradation of these fibres, as observed in non-denervated gastric adenocarcinomas in MNNG-treated rats, supports aggressive tumour growth 140,142.

Genome instability and epigenetics

Tumour formation represents a multistep process driven by (epi)genomic alterations in non-neoplastic cells6. It has been described that various neurotransmitters can influence the cellular genome mainly via pathways involved in DNA damage and repair. For instance, via its β_1 -adrenoceptor, noradrenaline protects DNA in CRC cell lines from oxidative stress^{111,143,144}. Similarly, serotonin protects the colonic epithelium against carcinogen-induced DNA damage in tumorigenic mouse models¹⁴⁵. On the other hand, adrenaline has been shown to stimulate degradation of p53 and the production of reactive oxygen species via β_2 -adrenergic receptors in vitro, thereby leading to DNA damage in stem cells and cancer cells 146-149. Next to the genome, the cellular epigenome can also be regulated by several neuropeptides and neurotransmitters such as gastric-releasing peptide (GRP) and serotonin (FIG. 4). In cultured human colon cancer cells, GRP-induced signalling enhances the expression of the heterochromatin protein 1β (HP1 β)^{150,151}. Even though HP1 β is a known epigenomic reader and modulator with critical effects on chromatin structure¹⁵², the exact consequences on heterochromatin status in gastrointestinal cancers are currently unknown and warrant further investigation¹⁵⁰. Nevertheless, antagonizing GRP-induced signals in CRC cells in vitro reduces HP1β levels and the concomitant invasive characteristics as well as cell growth via EGF signalling^{153–156}. Importantly, when combined with 5-fluorouracil, an anti-metabolic drug widely used in cancer chemotherapy, GRP antagonists synergistically inhibit CRC cell growth in vitro¹⁵⁷. Serotonin has been shown to bind the glutamine residue on histone 3 (H3Q5) next to the critical lysine residue (H3K4). The H3K4 residue represents a major methylation site that is known for its global effects on gene expression in normal and cancer cells¹⁵⁸. This so-called 'serotonylation' of Q5 stabilizes H3K4 methylation¹⁵⁸. Moreover, serotonylation can influence the functioning of proteins such as the mTORC1 oncoprotein. Inhibiting this serotonylation leads to diminished cancer cell proliferation in vitro and reduced tumour size in an ectopic CRC mouse model¹⁵⁹. Again, with most data being derived from in vitro experiments, it cannot be deduced where the neuronal messengers are derived from.

Reprogramming energy metabolism

To fuel their own growth and survival, cancer cells adapt their metabolism to comply with the high energetic demand of carcinogenic processes. Oncogenic mutations and subsequent mitochondrial dysfunction render tumour cells, including gastrointestinal cancer cells, dependent on glutamine metabolism¹⁶⁰⁻¹⁶⁴. In 2021, Rabben et al. uncovered that vagal innervation of gastric tumours maintains glutaminolysis in mice, whereas vagotomy re-established energy production by oxidative phosphorylation¹⁶⁰. This innervation-induced metabolic reprogramming effect is mediated by modifying functions of metabolic regulators such as HIF1A and SIRT1 (REF. 165). Besides, adrenergic signalling affects cancer cell metabolism as it has been implicated in mitochondrial respiration of (colon) cancer cell lines that express the β_1 -adrenoceptor. More specifically, nebivolol, a common β_1 -receptor blocker, inhibited mitochondrial respiration and subsequent ATP synthesis in several cancer cell types (including colon cancer) in vitro by upregulating ATPase inhibitory factor 1 (IF1) levels and impairing the phosphorylation of components of respiratory complexes I and V. Additionally, in vivo, when colon cancer cells were injected subcutaneously in nude mice, nebivolol impaired energy production and proliferation of colon cancer cells, while enhancing their apoptotic rate¹¹¹, thereby pinpointing towards the glycolytic-inducing capacities of noradrenaline.

Inflammation and immune evasion

Even though the neuroimmune crosstalk is increasingly recognized as a crucial regulator of digestive function and gastrointestinal homeostasis 166, mechanistic understanding of neuroimmune interactions in the context of gastrointestinal cancers is limited. Although, so far, the focus has been on innate macrophages as well as on adaptive T cells, a role for neural signalling to other types of immune cells is also likely as these cells express a variety of neurotransmitter receptors that interfere with their ability to attack tumour cells 167-169. However, here, we only focus on established neuroimmune interactions during gastrointestinal carcinogenesis.

Neural signals and innate immune cells. For breast, prostate and pancreatic cancer, extensive evidence depicts the opposing effects of sympathetic (enhancive) and parasympathetic (suppressive) innervation on the recruitment of tumour-associated macrophages (TAMs)¹⁷⁰.

However, this process is less well established for gastrointestinal cancers. Chronic restraint stress^{79,85} and unresolved inflammation represent an important risk factor for the development of gastric cancer, causing immune cell infiltration and subsequent promotion of epithelial proliferation. As a result, the TME of patients with gastric cancer is usually characterized by high levels of inflammation and recruited TAMs^{171,172}. Importantly, stress and infection-induced inflammation activate β-adrenergic signalling, to which TAMs and muscularis macrophages can respond. Thus, sympathetic signalling modulates macrophage infiltration turning the TME into a tumour-promoting environment, an aspect observed in different cancer types¹⁷³. β-Adrenoceptor signalling also results in suppressed immune activity, potentially via transformation of M1 (pro-inflammatory) to M2 (anti-inflammatory) macrophages, and the subsequent production of polyamines 166,174-177. This process represents a plausible mechanism given the observation that the presence of polarized M2 macrophages (antiinflammatory) correlates with tumour progression in patients with gastric cancer $(n = 113)^{178}$ and CRC $(n = 30)^{178}$ (REFS. 179,180), n = 205 (REF. 181)). Still, the role of TAMs in CRC is rather conflicting, partly dictated by their position within the TME. In fact, TAMs are considered oncogenic because they can promote cancer cell invasion and metastasis while suppressing antitumour immune responses. At the same time, their absence can induce similar effects, probably through a compensatory mechanism. Given that the presence of TAMs within tumour stroma negatively correlates with survival of patients with CRC and their position at the tumour front has the opposite effect, it could well be that their location within the TME determines their eventual antitumorigenic or protumorigenic capacity¹⁸².

Neural signs and adaptive immune cells. Infiltration of CD8+ effector T cells (or cytotoxic T lymphocytes) in the TME is associated with improved survival for patients in many types of cancer, including gastrointestinal cancers $^{183-185}$. In human CRC tissues (n = 39), perineural invasion was associated with decreased effector memory T cells¹⁸⁶. This finding is supported by the observation that β₂-adrenergic-mediated signalling controls lymphocyte trafficking in mice and blocks effector CD8+ T cell activation both in vivo 187,188 and in vitro 181. Such an effect can be reversed by blocking β_2 -adrenergic signalling, resulting in T cell activation and tumour shrinkage in mice^{168,189,190}. Interestingly, Tavazoie et al. observed that liver X receptor (LXR) agonists reduce the abundance of innate, myeloid-derived suppressor cells, which triggers CD8+ T cell responses and tumour regression, in both mouse and humans with colon cancer¹⁹¹. In human gastric cancer, increased CD8+ T cell density combined with elevated PDL1 expression correlates with metastatic and more advanced disease stages, which, according to research from other cancer types, can be induced by tumour innervation192-194. Adrenergic signalling, for instance, is associated with an increase in PDL1 expression in pancreatic cancer cells in vitro¹⁹⁵ and nerve fibres themselves can produce PDL1 in prostate cancer¹⁹⁶. Interestingly, work published as a preprint by Kuol et al.

suggests that expression of PDL1 ligands in CRC cell lines can be induced by cholinergic signalling 197 . Based on these data, a combined treatment effect of neuromodulators and immune-checkpoint inhibitors warrants future investigation. Nerves have similar tumour-supporting effects via T helper cells. Hou et al. revealed a shift from T helper 1 ($T_{\rm H}1$) to $T_{\rm H}2$ cytokine production in a mouse stress model of CRC 198 , whereas suppression of β -adrenergic signalling redirected the balance towards the $T_{\rm H}1$ side 199 . Similarly, Mitsui et al. showed that truncal vagotomy in mice limited $T_{\rm H}1$ and $T_{\rm H}2$ cytokine levels within the small intestine 200 .

Although it is apparent from the studies highlighted already that neural signalling can influence cancer hallmarks, there is still a largely undiscovered field of investigation with huge potential to better understand gastrointestinal cancer pathogenesis and to identify novel targets for proper gastrointestinal cancer management. As it is already known that various subtypes of immune cells have different effects on cancer hallmarks²⁰¹, future studies should also investigate the influence of different neuronal subtypes on cancer hallmarks and further distinguish local and systemic influences. In experimental animal models of melanoma and pancreatic cancer, it is described that sensory neurons are also involved in tumour initiation and progression, whereas, to our knowledge, this is not studied in other gastrointestinal cancers 187,202. Interesting, but not discussed within this Review, is the role of stress on gastrointestinal cancers, especially as we know that stress hormones can also affect cancer hallmarks as discussed earlier. For more information on this matter, we would like to refer to other reviews covering this topic 146,188.

Finally, we would like to address that, despite an increasing body of literature showing that glial cells are vital for nervous system function and homeostasis²⁰³, glial cells are not specifically considered in the neuronal signalling hallmark. Enteric glia are the non-neuronal cells within the ENS that not only provide structural support

Box 3 | The gut microbiota

The complex ecological system of microorganisms colonizing our digestive system—the gut microbiota—has a long-standing reputation of preserving our intestinal health^{232,309}. As early as 1901, enriched levels of bacteria within the gastrointestinal tract were discovered³¹⁰; thereafter, the presence of viruses, fungi and archaea was confirmed, with increased density whilst moving along the gastrointestinal tract³¹¹. Together, these species, both commensal and pathogenic, account for more than 100 trillion microorganisms, forming the largest reservoir of microorganisms communal to humans. This reservoir has co-evolved with its host to establish a sophisticated and mutually favourable relationship.

The gut microbiota is mostly established at birth but its composition changes swiftly over the first couple of years of life, reaching a diverse, adult-like microbial composition at ~2.5 years of age³¹². In humans, this system comprises three primary phylae, Bacillota (synonym Firmicutes), Bacteroidota (synonym Bacteroidetes) and Actinomycetota (synonym Actinobacteria)³¹³, and offers benefits to the host by strengthening intestinal epithelial barrier integrity³¹⁴, regulating host immunity³¹⁵ and protecting against pathogens³¹⁶. Despite its relatively constant composition, gut microbiota can be influenced and/or disrupted by environmental factors, including diet, lifestyle and antibiotic use. This disruption can lead to an imbalance between intestinal microorganisms, or 'dysbiosis', which impairs intestinal homeostasis and could set the stage for the development of several intestinal diseases such as inflammatory bowel disease and cancer²³².

to neurons but also regulate several tasks important for gut function such as intestinal motility and epithelial barrier integrity^{103,204,205}. Enteric glia are active partners in ENS activity and possess the required machinery to integrate and transmit information along enteric neural circuits^{206,207}. Although the exact role of enteric glia is not clear, the ENS is involved in several mucosal functions, including the maintenance of epithelial integrity and the gastrointestinal stem cell niche in the so-called neural-glial-epithelial unit^{208,209}. During gastrointestinal carcinogenesis, these neuroglial networks exhibit structural abnormalities, with a denser and more branched network towards cancer cells as well as a changed cellular subtype distribution with increased levels of neuroprotective messengers like PGE2, TGFβ and galanin, as observed in vitro and in mouse and human tissues²¹⁰⁻²¹³. Additionally, when activated by colon cancer cells, enteric glial cells, at least in vitro, stimulate stem cell expansion and tumour formation via the EGFR-ERK pathway, highlighting the reciprocal communication within the TME²¹⁰. Also, depletion of GFAP+ enteric glia prior to the induction of CRC in mice (azoxymethane-dextran sodium sulfate and $Apc^{Min/+}$) markedly reduces tumour burden (about 80-90% and 30%, respectively), whereas GFAP+ glial cell depletion after tumour formation does not affect tumour growth or number²¹⁴. This finding supports the notion that enteric glial cells mainly contribute to the initial phases of colorectal carcinogenesis — a finding that is further evidenced by Baghdadi et al., who revealed that GFAP+ enteric glia regulate the regeneration of the intestinal stem cell niche via WNT signalling^{215,216}. Together, even though there is much to uncover, also in the context of their possible contribution to neoneurogenesis²¹⁷, these data pinpoint to the addition of glial cells to the cancer hallmarks.

Neural relationship with the microbiota

Together with the enteric nervous, immune and endocrine systems, the multifaceted intestinal microbial community (BOX 3) orchestrates intestinal responses to pathophysiological challenges^{3,218,219}. Even though thorough experimental evidence linking the nervous system, microbiota and gastrointestinal cancers is currently lacking, the findings discussed next suggest functional associations between these three components, as the ENS and microbial community are both altered during gastrointestinal cancers and reciprocally communicate and modulate each other's composition and functioning during health and disease.

Using a *sox10* mutant zebrafish line, a Hirschsprung model characterized by absence of the ENS, Rolig et al. described profound alterations in the microbiota with an excess of pro-inflammatory microorganisms and a lack of anti-inflammatory lineages²¹⁸. Vice versa, the microbiota influences the structural organization of enteric neuron–glial networks as well as ENS and gut functioning (for example, intestinal permeability and ion transport) in a region-specific fashion^{220–224}. However, whether these are direct or indirect effects remains to be investigated. Notwithstanding possible links with gut innervation, the importance of a diverse yet balanced microbial

community and the fact that imbalances herein might have deleterious consequences are evidenced by various experimental models²²⁵. Disruptions of the commensal microbial community in mice have been shown to be associated with the development of (gastrointestinal) diseases, including gastrointestinal cancer^{226–228}. In this regard, the gut microbiota has been proposed as a novel hallmark of cancer¹⁰⁵.

Microbiota influence intestinal health by producing a diverse metabolite repertoire from dietary products within the gut, which, amongst other functions, strengthen the intestinal epithelial barrier and immune defence mechanisms²²⁹. However, upon intestinal dysfunction and/or carcinogenic transformation, the intestinal epithelial barrier gets compromised, providing microorganisms with the opportunity to infiltrate the internal environment, thereby setting the stage for disease progression²³⁰⁻²³³. One of the best-characterized commensal microorganisms residing in the gastric and duodenal lumen is *H. pylori*^{226,234}. However, this strain, alone or in combination with other aetiological factors, also represents one of the main risk factors for the development of gastric cancer and potentially increases the risk of CRC²³⁵. Infection with H. pylori can affect the release of different neurotransmitters, such as vagal acetylcholine, which influences ENS morphologically (for example, neuronal and axonal degeneration) and functionally (for example, changes in neuropeptide levels such as vasoactive intestinal peptide and c-fos) as well as the composition of the gut microbiota^{236,237}. Likewise, during Citrobacter rodentium infection in DMH-induced and ApcMin/+ mouse models of CRC, adrenergic signalling via the β_2 -adrenoceptor leads to recruitment of ChaT+T cells, thereby promoting colonic tumour development²³⁸. Additionally, gut microbiota have the ability to influence CRC susceptibility and progression by producing protumorigenic, neuroactive metabolites and modifying cancer hallmarks (for example, inflammation and genome instability) as evidenced by human-derived microorganism screenings^{239,240}. This phenomenon is further reflected by the observation that the gut microbiome of patients with CRC displays prominent differences compared with healthy individuals, including reduced butyrate-producing bacterial lineages and enriched levels of pathogenic bacteria such as Escherichia coli, Salmonella and Shigella. Such bacteria have been shown to strongly adhere to colonic epithelial cells and induce DNA damage in vitro (rat cell lines) and in vivo (CRC mouse models)3,241-243. Patients with CRC also display an altered virome (such as presence of cytomegalovirus) and mycobiome (for example, abundance of fungal Malassezia), when compared with healthy individuals, but no link with neural messengers has been established yet²²⁸.

Neural stemness and signatures in cancers

Cancer biology represents an intriguing process that seems to combine the fundamentals of embryonic development and organogenesis. Embryonic stem cells have the ability to differentiate into all three germ layers. However, their native identity is neural as they differentiate into a neural lineage (ectoderm) in vitro in the

absence of external cues, that is, under minimal culturing conditions^{244,245}. It is arguable that the neural-default state of embryonic stem cells goes along with their tumorigenic ability because it is well established that embryonic stem cells are capable of forming 2D colonies and 3D teratomas, that is, tumours, when propagated in vitro and in vivo, respectively²⁴⁶. Analogue observations have been described for neural precursor cells. With increased potential for tumorigenesis in terms of the required number of injected cells and tumour size, when compared with embryonic stem cells neural precursor cells could be regarded as 'potent' tumorigenic cells^{247,248}. Some derivatives of the neural precursor lineage, like neural crest cells, also display potent disseminating capacities as they can undergo an epithelial-mesenchymal transition and migrate away through xeno-environments to their destinations²⁴⁹. Interestingly, both embryonic stem cells and neural precursor cells lose their tumorigenicity when forced to differentiate by retinoic acid in vitro²⁴⁸. In most common gastrointestinal cancer types, cancer cells are primarily restricted to an epithelial lineage identity vet they still express potency markers such as MYC, OCT4 and SOX2 (REF. 250). Key effectors crucial for neural stemness (for example, nervous system polycomb 1 and Musashi 1) have also been shown to be important for colon and gastric cancer cells²⁵¹⁻²⁵⁴. Advances in multi-omics have highlighted that neural-related genes are more prone to DNA hypermethylation and, subsequently, are more often downregulated in human gastrointestinal cancers²⁵⁵⁻²⁵⁸. Although this finding seems counterintuitive, these downregulated pathways are related to neuron formation and functions^{255–258}. In this regard, it is tempting to suggest that cancer cells hijack neural reprogramming pathways to promote their tumorigenicity and potency. As described earlier, gastrointestinal cancer stem cells are able to generate a neuron-like phenotype, mimicking neuronal cells both in vitro and in vivo, as indicated by the protein expression of neural markers such as Tuj1 (REF.26). However, functional neuronal properties, such as action potential firings, have not been evaluated and are needed to further identify the functional role of these cells during gastrointestinal carcinogenesis. Similar to embryonic stem cells and neural precursor cells, gastrointestinal cancer cell lines treated with retinoic acid to stimulate neuronal differentiation have less tumorigenic capacities^{259,260}. Altogether, these findings propose a connection between neural identity and pluripotency as well as tumorigenicity.

Therapeutic potential

In view of the importance of the nervous system and neural signalling pathways in gastrointestinal cancer development, progression and dissemination, the modulation of neural–cancer crosstalk as well as of neural and neural stemness pathways in cancer cells is gaining more ground in the approach against gastrointestinal cancers. These strategies mainly comprise antagonizing parasympathetic and sympathetic activity and modulation of other neurotrophic signalling mechanisms, though they are not yet aimed at targeting the ENS.

Targeting sympathetic signals

Blocking sympathetic β-adrenergic receptors has shown promise in lowering the recurrence and long-term mortality of several (gastrointestinal) cancer types^{261,262}, but failed to reduce short-term mortality in patients with oesophageal cancer^{263,264}. Moreover, a meta-analysis of the observational studies in CRC have shown no improvement in overall survival (HR 0.90, 95% CI 0.93-1.10)²⁶⁵. Based on the involvement of β -adrenergic signalling in gastrointestinal carcinogenesis and the ability of β-blockers to enhance radiation efficacy in colon tumours in vivo^{266,267}, intervention-designed clinical trials in patients with gastric cancer and CRC are currently being conducted. Such trials assess the potential benefits of these beta-blockers either preoperatively (gastrointestinal cancer²⁶⁸), in combination with chemotherapy and/or radiotherapy (gastric cancer²⁶⁹, oesophageal adenocarcinoma²⁷⁰), or perioperatively with a prostaglandin inhibitor, aiming to ameliorate stress-inflammatory responses (CRC²⁷¹).

Targeting parasympathetic signalling

Inhibition of the acetylcholine pathway (botulinum toxin injection) weakens colonic tumour growth and invasiveness and sensitizes gastric cancer cells to chemotherapy. This process results in prolonged survival of mice with gastric cancer^{64,69,272,273}. As it has been shown that surgical denervation and botulinum toxin suppresses gastric cancer and recurrence in mice and humans^{23,31}, botulinum toxin was tested in a pilot phase II trial consisting of six patients with gastric cancer²⁷⁴. Despite promising safety, its application to treat gastric cancer still needs broader investigations¹⁶⁰.

Targeting neurotrophic receptors

Owing to the well-described role of neurotrophins and their Trk receptors in tumorigenesis, they represent possible targets in the treatment of gastrointestinal cancers. Blocking NGF–TrkA or BDNF–TrkB signalling reduces innervation and the size of gastric tumours in genetic mice models⁶³ and suppresses gastric cancer growth in vitro and in vivo¹³¹. In this respect, larotrectinib and entrectinib, two potent small-molecule inhibitors of Trk, have been approved by the FDA to treat Trk fusion-positive tumours²⁷⁵. Clinical trials and development of second-generation Trk inhibitors are also ongoing to overcome resistance issues (phase I^{276–278} and phase II^{279,280}).

Neural stem cells as drug transporters

Neural stem cells are known for their tropism towards cancer cells in vitro and in vivo²⁸¹. Consequently, they have been used as vehicles to specifically express

anticancer drugs and prodrug-activating enzymes at the tumour site, thereby reducing potential adverse effects^{282,283}. This approach is under phase I clinical trials for brain cancer^{284,285} but has not yet been explored in the context of gastrointestinal cancers.

Conclusions

The multifaceted characteristics of carcinogenesis mirror the essential processes that drive the development, growth and survival of multicellular organisms. Within these processes, cells are in close and permanent contact with their neighbourhood, with every cell, even cancer cells, expressing similar messengers and receptors to communicate. During carcinogenesis, cancer cells utilize nearby and remote resources to serve their increasing energy demand. It has been established that neurons reside within the TME and communicate with cancer cells as well as with other TME members. Nerves not only provide a 'railway' for dissemination but are also deployed by cancer cells to constantly send out a variety of signals that favour their growth and survival. As a consequence, manipulating the nervous system shows promise in the treatment of gastrointestinal cancers. However, within the reciprocal neural-cancer crosstalk, many enigmas still have to be unravelled before proper therapies can be designed. This situation is complicated by the fact that several messenger molecules have dual roles, being either protumorigenic or antitumorigenic, depending on the cell type, receptor and intracellular targets they influence as well as the tissue wherein they exert their actions. In addition, the complex gastrointestinal environment in which several other tissue components, such as the immune system and intestinal microbiota, are implicated, complicates the development of neural-oriented treatment strategies for gastrointestinal cancers. Finally, the findings in this field are mainly established using in vitro and in vivo assays, whereas translation to the human situation is often missing. Nerves are identified in the tumour stroma of patients with gastrointestinal cancer and affect patient outcomes but in-depth molecular profiling of tumour-associated neurons in humans is lacking. This research might be hindered as neurons represent a rare intestinal cell population that is difficult to capture, and long-lived culture potential is restricted. Altogether, these aspects emphasize the need to adopt technologies and tools from the neuroscience field and to establish strong collaborations between neuroscientists and cancer biologists. Further in-depth basic and translational research is warranted to understand the role of neurons in gastrointestinal cancers and to identify the best therapeutic targets.

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- Sung, H. et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J. Clin. 71, 209–249 (2021).
- Anderson, N. M. & Simon, M. C. The tumor microenvironment. *Curr. Biol.* 30, R921–R925 (2020).
- Quante, M., Varga, J., Wang, T. C. & Greten, F. R. The gastrointestinal tumor microenvironment. Gastroenterology 145, 63–78 (2013).
- Zahalka, A. H. & Frenette, P. S. Nerves in cancer. Nat. Rev. Cancer 20, 143–157 (2020).
 Prominent review highlighting the importance of nerves within the TME in a variety of cancer types.
 Mancino, M., Ametller, E., Cascon, P. & Almendro, V.
- Mancino, M., Ametiler, E., Gascon, P. & Almendro, The neuronal influence on tumor progression. *Biochim. Biophys. Acta* 1816, 105–118 (2011).
- Hanahan, D. & Weinberg, R. A. Hallmarks of cancer: the next generation. *Cell* 144, 646–674 (2011).
- Hanahan, D. Hallmarks of cancer: new dimensions. Cancer Discov. 12, 31–46 (2022).
 Timely revisited view on the hallmarks of cancer, building on the original hallmarks.
- Furness, J. B., Callaghan, B. P., Rivera, L. R. & Cho, H. J. The enteric nervous system and gastrointestinal innervation: integrated local and central control. *Adv. Exp. Med. Biol.* 817, 39–71 (2014).
- Langley, J. N. The autonomic nervous system. *Brain* 26, 1–26 (1903).

- Furness, J. B. *The Enteric Nervous System* 1st edn, 288 (Blackwell Publishing, 2006).
- Bon-Frauches, A. C. & Boesmans, W. The enteric nervous system: the hub in a star network. *Nat. Rev. Gastroenterol. Hepatol.* 17, 717–718 (2020).
- Rao, M. & Gershon, M. D. The bowel and beyond: the enteric nervous system in neurological disorders. Nat. Rev. Gastroenterol. Hepatol. 13, 517–528 (2016).
- Niesler, B., Kuerten, S., Demir, I. E. & Schafer, K. H. Disorders of the enteric nervous system - a holistic view. Nat. Rev. Gastroenterol. Hepatol. 18, 393–410 (2021).
- Holland, A. M., Bon-Frauches, A. C., Keszthelyi, D., Melotte, V. & Boesmans, W. The enteric nervous system in gastrointestinal disease etiology. *Cell Mol. Life Sci.* 78, 4713–4733 (2021).
- 15. Rademakers, G. et al. The role of enteric neurons in the development and progression of colorectal cancer. *Biochim. Biophys. Acta* 1868, 420–434 (2017). The first review summarizing the importance of enteric neurons in colorectal carcinogenesis, focusing on the link between the ENS, inflammation and CRC
- Schledwitz, A., Xie, G. & Raufman, J. P. Exploiting unique features of the gut-brain interface to combat gastrointestinal cancer. *J. Clin. Invest.* https://doi.org/ 10.1172/JCI143776 (2021).
- Griffin, N. et al. Clinicopathological significance of nerves in esophageal cancer. *Am. J. Pathol.* 190, 1921–1930 (2020).
- Xu, G. et al. Prognosis and progression of ESCC patients with perineural invasion. Sci. Rep. 7, 43828 (2017).
- Deng, J. et al. Prognostic value of perineural invasion in gastric cancer: a systematic review and meta-analysis. PLoS ONE 9, e88907 (2014).
- Tianhang, L., Guoen, F., Jianwei, B. & Liye, M. The effect of perineural invasion on overall survival in patients with gastric carcinoma. *J. Gastrointest. Surg.* 12, 1263–1264 (2008).
- 21. Knijn, N., Mogk, S. C., Teerenstra, S., Simmer, F. & Nagtegaal, I. D. Perineural invasion is a strong prognostic factor in colorectal cancer: a systematic review. Am. J. Surg. Pathol. 40, 103–112 (2016). Systematic review depicting the prominence of perineural invasion by summarizing all evidence for its prognostic value for CRC.
- Liebig, C. et al. Perineural invasion is an independent predictor of outcome in colorectal cancer. *J. Clin.* Oncol. 27, 5131–5137 (2009).
- 23. Zhao, C. M. et al. Denervation suppresses gastric tumorigenesis. Sci. Transl. Med. https://doi.org/10.1126/scitranslmed.3009569 (2014). Leading research article uncovering the functional importance of parasympathetic nerves in gastric cancer and their highly valuable potential as therapeutic targets. This article paved the way to gain more insights into the role of the nervous
- system in (gastric) cancer.
 24. Albo, D. et al. Neurogenesis in colorectal cancer is a marker of aggressive tumor behavior and poor outcomes. *Cancer* 117, 4834–4845 (2011).
- Mauffrey, P. et al. Progenitors from the central nervous system drive neurogenesis in cancer. *Nature* 569, 672–678 (2019).
- Lu, R. et al. Neurons generated from carcinoma stem cells support cancer progression. *Signal. Transduct. Target. Ther.* 2, 16036 (2017).
- Weinstein, V. A., Druckerman, L. J., Lyons, A. S. & Colp, R. Gastroenterostomy and vagotomy in the treatment of duodenal ulcer. *Ann. Surg.* 141, 482–487 (1955).
- Kraft, R. O., Myers, J., Overton, S. & Fry, W. J. Vagotomy and the gastric ulcer. *Am. J. Surg.* 121, 122–128 (1971).
- Calmels, S., Bereziat, J. C., Ohshima, H. & Bartsch, H. Bacterial formation of N-nitroso compounds from administered precursors in the rat stomach after omeprazole-induced achlorhydria. *Carcinogenesis* 12, 435–439 (1991).
- Moller, H., Nissen, A. & Mosbech, J. Use of cimetidine and other peptic ulcer drugs in Denmark 1977-1990 with analysis of the risk of gastric cancer among cimetidine users. *Gut* 33, 1166–1169 (1992).
 Rabben, H. L., Zhao, C. M., Hayakawa, Y. Wang, T. C.
- Rabben, H. L., Zhao, C. M., Hayakawa, Y., Wang, T. O. & Chen, D. Vagotomy and gastric tumorigenesis. *Curr. Neuropharmacol.* 14, 967–972 (2016).
- Tatsuta, M., Iishi, H., Yamamura, H., Baba, M. & Taniguchi, H. Effects of bilateral and unilateral vagotomy on gastric carcinogenesis induced by N-methyl-N'-nitro-N-nitrosoguanidine in Wistar rats. Int. J. Cancer 42, 414–418 (1988).

- Kim, S. J. & Choi, C. W. Common locations of gastric cancer: review of research from the endoscopic submucosal dissection era. J. Korean Med. Sci. 34, e231 (2019)
- Cassaro, M. et al. Topographic patterns of intestinal metaplasia and gastric cancer. *Am. J. Gastroenterol.* 95, 1431–1438 (2000).
- Liu, V. et al. Extrinsic intestinal denervation modulates tumor development in the small intestine of Apc^{Min/+} mice. J. Exp. Clin. Cancer Res. 34, 39 (2015).
- Nelson, R. L., Briley, S., Vaz, O. P. & Abcarian, H. The effect of vagotomy and pyloroplasty on colorectal tumor induction in the rat. *J. Surg. Oncol.* 51, 281–286 (1992).
- Bayón Lara, A. M. et al. Colonic carcinogenesis in vagotomyzed rats. Rev. Esp. Enferm. Dig. 93, 576–586 (2001).
- Caygill, C. P., Hill, M. J., Hall, C. N., Kirkham, J. S. & Northfield, T. C. Increased risk of cancer at multiple sites after gastric surgery for peptic ulcer. *Gut* 28, 924–928 (1987).
- Bundred, N. J. et al. Gastric surgery and the risk of subsequent colorectal cancer. *Br. J. Surg.* 72, 618–619 (1985).
- Watt, P. C., Patterson, C. C. & Kennedy, T. L. Late mortality after vagotomy and drainage for duodenal ulcer. *Br. Med. J.* 288, 1335–1338 (1984).
 Tatsuta, M., lishi, H., Baba, M. & Taniguchi, H.
- Tatsuta, M., lishi, H., Baba, M. & Taniguchi, H. Inhibitions by 6-hydroxydopamine and neostigmine singly or together of gastric carcinogenesis induced by N-methyl-N'-nitro-N-nitrosoguanidine in Wistar rats. Int. J. Cancer 51, 767–771 (1992).
- Tatsuta, M., Iishi, H., Baba, M. & Taniguchi, H. Inhibition of azoxymethane-induced experimental colon carcinogenesis in Wistar rats by 6-hydroxydopamine. *Int. J. Cancer* 50, 298–301 (1992)
- Sadighparvar, S., Darband, S. G., Ghaderi-Pakdel, F., Mihanfar, A. & Majidinia, M. Parasympathetic, but not sympathetic denervation, suppressed colorectal cancer progression. Eur. J. Pharmacol. 913, 174626 (2021).
- Miyato, H., Kitayama, J., Ishigami, H., Kaisaki, S. & Nagawa, H. Loss of sympathetic nerve fibers around intratumoral arterioles reflects malignant potential of gastric cancer. *Ann. Surg. Oncol.* 18, 2281–2288 (2011).
- Gao, J. & Liu, S. G. Role of sympathetic and parasympathetic nerves in the development of gastric cancer through antagonism. *Chin. Med. J.* 134, 908–909 (2021).
- Bae, G. E. et al. Lower sympathetic nervous system density and beta-adrenoreceptor expression are involved in gastric cancer progression. *Anticancer Res.* 39, 231–236 (2019).
- Borresen, T., Henriksen, O. B. & Christensen, N. J. Decreased norepinephrine concentration in normal tissue neighboring a malignant tumor. *Cancer Res.* 40, 4320–4321 (1980).
- Garcia, S. B., Minto, S. B., Marques, I. D. S. & Kannen, V. Myenteric denervation of the gut with benzalkonium chloride: a review of forty years of an experimental model. *Can. J. Gastroenterol. Hepatol.* 2019, 3562492 (2019).
- Zucoloto, S. et al. Effect of chemical ablation of myenteric neurones on intestinal cell proliferation. *Cell Tissue Kinet.* 21, 213–219 (1988).
- Romanello, L. M. F. Effect of destruction of myenteric neurons of rat stomach by benzalkonium chloride on gastric acid secretion and gastrin release. *Gut* 32, 594 (1991).
- Febronio, L. H., Britto-Garcia, S., de Oliveira, J. S. & Zucoloto, S. Megaesophagus in rats. Res. Exp. Med. 197, 109–115 (1997).
- Polli-Lopes, A. C., Zucoloto, S., de Queiros Cunha, F., da Silva Figueiredo, L. A. & Garcia, S. B. Myenteric denervation reduces the incidence of gastric tumors in rats. Cancer Lett. 190, 45–50 (2003).
- Vespucio, M. V. et al. Intrinsic denervation of the colon is associated with a decrease of some colonic preneoplastic markers in rats treated with a chemical carcinogen. *Braz. J. Med. Biol. Res.* 41, 311–317 (2008).
- Estofolete, C. F., Zucoloto, S., Oliani, S. M., Polli-Lopes, A. C. & Gil, C. D. Myenteric denervation downregulates galectin-1 and -3 expression in gastric carcinogenesis. *Dig. Dis. Sci.* 56, 1637–1644 (2011).
- Sato, A. et al. Pathophysiology of aganglionic colon and anorectum: an experimental study on aganglionosis produced by a new method in the rat. *J. Pediatr. Surg.* 13, 399–435 (1978).

- Garcia, S. B., Oliveira, J. S. M., Pinto, L. Z., Muccillo, G. & Zucoloto, S. The relationship between megacolon and carcinoma of the colon: an experimental approach. *Carcinogenesis* 17, 1777–1779 (1996).
- Munari, F. F. et al. The relationship between esophageal cancer, chagasic megaesophagus and HPV: myths, tales or reality? *Histol. Histopathol.* 33, 1135–1149 (2018).
- Kennedy, M. F., Tutton, P. J. & Barkla, D. H. Adrenergic factors involved in the control of crypt cell proliferation in jejunum and descending colon of mouse. Clin. Exp. Pharmacol. Physiol. 10, 577–586 (1983).
- Berthoud, H. R., Carlson, N. R. & Powley, T. L. Topography of efferent vagal innervation of the rat gastrointestinal tract. *Am. J. Physiol.* 260, R200–R207 (1991).
- Aloe, L. Rita Levi-Montalcini and the discovery of NGF, the first nerve cell growth factor. *Arch. Ital. Biol.* 149, 175–181 (2011).
- Liu, S. M. Neurotrophic factors in enteric physiology and pathophysiology. *Neurogastroent. Motil.* https://doi.org/10.1111/nmo.13446 (2018).
- Schonkeren, S. L., Thijssen, M. S., Vees, N., Boesmans, W. & Melotte, V. The emerging role of nerves and glia in colorectal cancer. *Cancers* https://doi.org/10.3390/cancers13010152 (2021).
 Havakawa, Y. et al. Nerve growth factor promotes
- Hayakawa, Y. et al. Nerve growth factor promotes gastric tumorigenesis through aberrant cholinergic signaling. Cancer Cell 31, 21–34 (2017).
 Prominent research paper describing that cholinergic signalling affects gastric carcinogenesis in vitro and in vivo.
- Wang, L. et al. Muscarinic receptor M3 mediates cell proliferation induced by acetylcholine and contributes to apoptosis in gastric cancer. *Tumour Biol.* 37, 2105–2117 (2016).
- Cheng, K. et al. Acetylcholine release by human colon cancer cells mediates autocrine stimulation of cell proliferation. Am. J. Physiol. Castrointest. Liver Physiol. 295, G591–C597 (2008).
- Yang, W. L. & Frucht, H. Cholinergic receptor up-regulates COX-2 expression and prostaglandin E(2) production in colon cancer cells. *Carcinogenesis* 21, 1789–1793 (2000).
- Frucht, H., Gazdar, A. F., Park, J. A., Oie, H. & Jensen, R. T. Characterization of functional receptors for gastrointestinal hormones on human colon cancer cells. *Cancer Res.* 52, 1114 (1992).
- Cheng, K. R., Zimniak, P. & Raufman, J. P. Transactivation of the epidermal growth factor receptor mediates cholinergic agonist-induced proliferation of h508 human colon cancer cells. Cancer Res. 63, 6744–6750 (2003).
- Yu, H. et al. Acetylcholine acts through M3 muscarinic receptor to activate the EGFR signaling and promotes gastric cancer cell proliferation. Sci. Rep. 7, 40802–40802 (2017).
 Raufman. J. P. et al. Genetic ablation of M3
- Rauman, J. P. et al. Genetic ablation of M3 muscarinic receptors attenuates murine colon epithelial cell proliferation and neoplasia. *Cancer Res.* 68, 3573–3578 (2008).
- Cai, J., Maitra, A., Anders, R. A., Taketo, M. M. & Pan, D. β-Catenin destruction complex-independent regulation of Hippo-YAP signaling by APC in intestinal tumorigenesis. *Genes Dev.* 29, 1493–1506 (2015).
- Rosenbluh, J. et al. β-Catenin-driven cancers require a YAP1 transcriptional complex for survival and tumorigenesis. *Cell* 151, 1457–1473 (2012).
 Schaak, S. et al. Alpha, adrenoceptors regulate
- Schaak, S. et al. Alpha₂ adrenoceptors regulate proliferation of human intestinal epithelial cells. Gut 47, 242–250 (2000).
- Peng, Z., Heath, J., Drachenberg, C., Raufman, J. P. & Xie, G. Cholinergic muscarinic receptor activation augments murine intestinal epithelial cell proliferation and tumorigenesis. *BMC Cancer* 13, 204 (2013).
- Xie, G., Cheng, K., Shant, J. & Raufman, J. P. Acetylcholine-induced activation of M3 muscarinic receptors stimulates robust matrix metalloproteinase gene expression in human colon cancer cells. Am. J. Physiol. Gastrointest. Liver Physiol. 296, G755–G763 (2009).
- Zhong, L. R., Estes, S., Artinian, L. & Rehder, V. Acetylcholine elongates neuronal growth cone filopodia via activation of nicotinic acetylcholine receptors. *Dev. Neurobiol.* 73, 487–501 (2013)
- Jobling, P. et al. Nerve-cancer cell cross-talk: a novel promoter of tumor progression. *Cancer Res.* 75, 1777–1781 (2015).
- 78. Buffin-Meyer, B. et al. EGF receptor transactivation and P13-kinase mediate stimulation of ERK by α_{2A} -adrenoreceptor in intestinal epithelial cells: a role in

REVIEWS

- wound healing. *Eur. J. Pharmacol.* **574**, 85–93 (2007).
- Zhang, X. et al. Chronic stress promotes gastric cancer progression and metastasis: an essential role for ADRB2. *Cell Death Dis.* 10, 788 (2019).
- Ogawa, H. et al. Prognostic significance of β2-adrenergic receptor expression in patients with surgically resected colorectal cancer. *Int. J. Clin. Oncol.* 25, 1137–1144 (2020).
- 81. Shi, M. et al. Catecholamine up-regulates MMP-7 expression by activating AP-1 and STAT3 in gastric cancer. *Mol. Cancer* **9**, 269 (2010).
- Shan, T. et al. Novel regulatory program for norepinephrine-induced epithelial-mesenchymal transition in gastric adenocarcinoma cell lines. *Cancer Sci.* 105, 847–856 (2014).
- Lu, Y. J. et al. Isoprenaline induces epithelialmesenchymal transition in gastric cancer cells. *Mol. Cell Biochem.* 408, 1–13 (2015).
- Njeim, R. & Eid, A. Alpha-2c adrenergic receptor promotes the malignant phenotype of colon cancer cells. FASEB J. 32, 695.5 (2018).
- Zhi, X. et al. Adrenergic modulation of AMPKdependent autophagy by chronic stress enhances cell proliferation and survival in gastric cancer. Int. J. Oncol. 54, 1625–1638 (2019).
- cancer. Int. J. Oncol. 54, 1625–1638 (2019).
 86. Chin, C. C. et al. Selective β2-AR blockage suppresses colorectal cancer growth through regulation of EGFR-Akt/ERK1/2 signaling, G1-phase arrest, and apoptosis. J. Cell Physiol. 231, 459–472 (2016).
- Griffin, N. et al. The neurotrophic tyrosine kinase receptor 1 (TirkA) is overexpressed in oesophageal squamous cell carcinoma. *Pathology* 53, 470–477 (2021).
- Ökumura, T. et al. The biological role of the low-affinity p75 neurotrophin receptor in esophageal squamous cell carcinoma. Clin. Cancer Res. 12, 5096–5103 (2006).
- Chen, H. et al. NGFR increases the chemosensitivity of colorectal cancer cells by enhancing the apoptotic and autophagic effects of 5-fluorouracil via the activation of \$100A9. Front. Oncol. 11, 652081 (2021).
- Yang, Z. et al. Epigenetic inactivation and tumorsuppressor behavior of NGFR in human colorectal cancer. Mol. Cancer Res. 13, 107–119 (2015).
- Allen, J. K. et al. Sustained adrenergic signaling promotes intratumoral innervation through BDNF induction. *Cancer Res.* 78, 3233–3242 (2018).
- Akil, H., Perraud, A., Melin, C., Jauberteau, M. O. & Mathonnet, M. Fine-tuning roles of endogenous brain-derived neurotrophic factor, TrkB and sortilin in colorectal cancer cell survival. *PLoS ONE* 6, e25097 (2011).
- Perrone, M. G., Notarnicola, M., Caruso, M. G., Tutino, V. & Scilimati, A. Upregulation of beta3adrenergic receptor mRNA in human colon cancer: a preliminary study. *Oncology* 75, 224–229 (200
- a preliminary study. Oncology 75, 224–229 (2008).
 Douma, S. et al. Suppression of anoikis and induction of metastasis by the neurotrophic receptor TrkB. Nature 430, 1034–1039 (2004).
- Fujikawa, H. et al. High TrkB expression levels are associated with poor prognosis and EMT induction in colorectal cancer cells. J. Gastroenterol. 47, 775–784 (2012).
- Meng, L. et al. Targeting the BDNF/TrkB pathway for the treatment of tumors. *Oncol. Lett.* 17, 2031–2039 (2019).
- de Farias, C. B. et al. BDNF/TrkB signaling protects HT-29 human colon cancer cells from EGFR inhibition. Biochem. Biophys. Res. Commun. 425, 328–332 (2012).
- Huang, S. M. et al. Brain-derived neurotrophic factor regulates cell motility in human colon cancer. *Endocr. Relat. Cancer* 22, 455–464 (2015).
- Yang, X., Martin, T. A. & Jiang, W. G. Biological influence of brain-derived neurotrophic factor (BDNF) on colon cancer cells. Exp. Ther. Med. 6, 1475–1481 (2013).
- 100. Mescher, A. L., Connell, E., Hsu, C., Patel, C. & Overton, B. Transferrin is necessary and sufficient for the neural effect on growth in amphibian limb regeneration blastemas. *Dev. Growth Differ.* 39, 677–684 (1997).
- Wang, L., Marchionni, M. A. & Tassava, R. A. Cloning and neuronal expression of a type III newt neuregulin and rescue of denervated, nerve-dependent newt limb blastemas by rhGGF2. J. Neurobiol. 43, 150–158 (2000).
- 102. Smith, M. J., Globus, M. & Vethamany-Globus, S. Nerve extracts and substance P activate the phosphatidylinositol signaling pathway and

- mitogenesis in newt forelimb regenerates. *Dev. Biol.* **167**, 239–251 (1995).

 103. Boesmans, W. et al. Development, diversity, and
- 103. Boesmans, W. et al. Development, diversity, and neurogenic capacity of enteric glia. Front. Cell Dev. Biol. 9, 775102 (2021).
- 04. Hanahan, D. & Weinberg, R. A. The hallmarks of cancer. Cell 100, 57–70 (2000). Landmark paper describing the original hallmarks of cancer, that is, the acquired, functional and biological capacities that govern neoplastic transformation.
- 105. Senga, S. S. & Grose, R. P. Hallmarks of cancer-the new testament. *Open Biol.* https://doi.org/10.1098/ rsob.200358 (2021).
 - New perspective to add neuronal signalling to the hallmarks of cancer.
- 106. Folkman, J. Tumor angiogenesis: therapeutic implications. *N. Engl. J. Med.* **285**, 1182–1186 (1971).
- 107. Wang, W. et al. Nerves in the tumor microenvironment: origin and effects. Front. Cell Dev. Biol. 8, 601738 (2020).
- 108. Klagsbrun, M. & Eichmann, A. A role for axon guidance receptors and ligands in blood vessel development and tumor angiogenesis. Cytokine Growth Factor. Rev. 16, 535–548 (2005).
- 109. Gysler, S. M. & Drapkin, R. Tumor innervation: peripheral nerves take control of the tumor microenvironment. J. Clin. Invest. https://doi. org/10.1172/JC1147276 (2021).
- 110. Ekstrand, A. J. et al. Deletion of neuropeptide Y (NPY) 2 receptor in mice results in blockage of NPY-induced angiogenesis and delayed wound healing. *Proc. Natl Acad. Sci. USA* **100**, 6033–6038 (2003).
- Nuevo-Tapioles, C. et al. Coordinate beta-adrenergic inhibition of mitochondrial activity and angiogenesis arrest tumor growth. Nat. Commun. 11, 3606 (2020)
- 112. Chen, S. H. et al. Perineural invasion of cancer: a complex crosstalk between cells and molecules in the perineural niche. Am. J. Cancer Res. 9, 1–21 (2019).
- 113. Zahalka, A. H. et al. Adrenergic nerves activate an angio-metabolic switch in prostate cancer. *Science* 358, 321–326 (2017).
 - One of the first studies to indicate that the nervous system affects the angiometabolic switch in prostate cancer cells.
- 114. Liu, J. et al. The effect of chronic stress on anti-angiogenesis of sunitinib in colorectal cancer models. *Psychoneuroendocrinology* 52, 130–142 (2015)
- Zamani, A. & Qu, Z. C. Serotonin activates angiogenic phosphorylation signaling in human endothelial cells. FEBS Lett. 586, 2360–2365 (2012).
- Nocito, A. et al. Serotonin regulates macrophagemediated angiogenesis in a mouse model of colon cancer allografts. *Cancer Res.* 68, 5152–5158 (2008).
- Chakroborty, D. et al. Depleted dopamine in gastric cancer tissues: dopamine treatment retards growth of gastric cancer by inhibiting angiogenesis. *Clin. Cancer Res.* 10, 4349–4356 (2004).
- Sarkar, C. et al. Dopamine in vivo inhibits VEGF-induced phosphorylation of VEGFR-2, MAPK, and focal adhesion kinase in endothelial cells. *Am. J. Physiol. Heart Circ. Physiol.* 287, H1554–H1560 (2004).
- 119. Sarkar, C., Chakroborty, D., Chowdhury, U. R., Dasgupta, P. S. & Basu, S. Dopamine increases the efficacy of anticancer drugs in breast and colon cancer preclinical models. *Clin. Cancer Res.* 14, 2502–2510 (2008).
- Basu, S. et al. The neurotransmitter dopamine inhibits angiogenesis induced by vascular permeability factor/ vascular endothelial growth factor. *Nat. Med.* 7, 559–574 (2001).
- Joyce, J. A. & Fearon, D. T. T cell exclusion, immune privilege, and the tumor microenvironment. *Science* 348, 74–80 (2015).
- Hering, N. A. et al. Blockage of cholinergic signaling via muscarinic acetylcholine receptor 3 inhibits tumor growth in human colorectal adenocarcinoma. *Cancers* 13, 3220 (2021).
- Belo, A. et al. Múscarinic receptor agonists stimulate human colon cancer cell migration and invasion. Am. J. Physiol. Gastrointest. Liver Physiol. 300, G749–G750 (2011)
- G749–G760 (2011).
 124. Raufman, J. P. et al. Muscarinic receptor agonists stimulate matrix metalloproteinase 1-dependent invasion of human colon cancer cells. *Biochem. Biophys. Pag. Commun.* 415, 319–324 (2011).
- Biophys. Res. Commun. 415, 319–324 (2011). 25. Sawayama, H., Ishimoto, T. & Baba, H. Microenvironment in the pathogenesis of gastric cancer metastasis. J. Cancer Metastasis Treat. 4, 10 (2018).

- 126. Samantaray, S., Sharma, R., Chattopadhyaya, T. K., Gupta, S. D. & Ralhan, R. Increased expression of MMP-2 and MMP-9 in esophageal squamous cell carcinoma. *J. Cancer Res. Clin. Oncol.* 130, 37–44 (2004).
- 127. Li, Y. et al. Overexpression of MMP-2 and MMP-9 in esophageal squamous cell carcinoma. *Dis. Esophagus* 22, 664–667 (2009).
- 128. Masur, K., Niggemann, B., Zanker, K. S. & Entschladen, F. Norepinephrine-induced migration of SW 480 colon carcinoma cells is inhibited by beta-blockers. *Cancer Res.* 61, 2866–2869 (2001).
 129. Lei, Y. et al. Nerve growth factor orchestrates NGAL
- 129. Lei, Y. et al. Nerve growth factor orchestrates NGAI and matrix metalloproteinases activity to promote colorectal cancer metastasis. *Clin. Transl. Oncol.* https://doi.org/10.1007/s12094-021-02666-x (2021).
- Jin, H. F. et al. P75 neurotrophin receptor inhibits invasion and metastasis of gastric cancer. *Mol. Cancer Res.* 5, 423–433 (2007).
- Okugawa, Y. et al. Brain-derived neurotrophic factor/ tropomyosin-related kinase B pathway in gastric cancer. Br. J. Cancer 108, 121–130 (2013).
- 132. Szpunar, M. J., Burke, K. A., Dawes, R. P., Brown, E. B. & Madden, K. S. The antidepressant desipramine and a2-adrenergic receptor activation promote breast tumor progression in association with altered collagen structure. *Cancer Prev. Res.* 6, 1262–1272 (2013).
- 133. Ganguly, D. et al. Cancer-associated fibroblasts: versatile players in the tumor microenvironment. Cancers https://doi.org/10.3390/cancers12092652 (2020).
- 134. Sahai, E. et al. A framework for advancing our understanding of cancer-associated fibroblasts. *Nat. Rev. Cancer* 20, 174–186 (2020).
- 135. Ping, O. R. et al. Cancer-associated fibroblasts: overview, progress, challenges, and directions. *Cancer Gene Ther.* 28, 984–999 (2021).
 136. Kobayashi, H. et al. Cancer-associated fibroblasts in
- 136. Kobayashi, H. et al. Cancer-associated fibroblasts in gastrointestinal cancer. *Nat. Rev. Gastro Hepat.* 16, 282–295 (2019).
- Akbareian, S. E. et al. Enteric neural crest-derived cells promote their migration by modifying their microenvironment through tenascin-C production. *Dev. Biol.* 382, 446–456 (2013).
- 138. Nagy, N. et al. Collagen 18 and agrin are secreted by neural crest cells to remodel their microenvironment and regulate their migration during enteric nervous system development. *Development* https://doi.org/ 10.1242/dev.160317 (2018).
- 139. Vaes, N. et al. Loss of enteric neuronal Ndrg4 promotes colorectal cancer via increased release of Nid1 and Fbln2. EMBO Rep. https://doi.org/ 10.15252/embr.202051913 (2021).
 - Research paper addressing the role of enteric neural-derived ECM proteins in CRC.
- 140. Estofolete, C. F. et al. Effects of myenteric denervation on extracellular matrix fibers and mast cell distribution in normal stomach and gastric lesions. *Cancer Cell Int.* 10, 18 (2010).
- Li, J. et al. Elastin is a key factor of tumor development in colorectal cancer. *BMC Cancer* https://doi.org/ 10.1186/s12885-020-6686-x (2020).
- 10.1186/s12885-020-6686-x (2020).
 142. Wang, Y., Song, E. C. & Resnick, M. B. in *Tumor Microenvironment: Extracellular Matrix Components Part B* (ed. Birbrair, A.) 1–16 (Springer International Publishing, 2020).
- 143. Patel, P. R. et al. Norepinephrine reduces reactive oxygen species (ROS) and DNA damage in ovarian surface epithelial cells. *J. Bioanal. Biomed.* 7, 75–80 (2015).
- 144. Rizzi, É. et al. β1-Adrenergic blockers exert antioxidant effects, reduce matrix metalloproteinase activity, and improve renovascular hypertension-induced cardiac hypertrophy. Free Radic. Biol. Med. 73, 308–317 (2014)
- 145. Šakita, J. Y. et al. Serotonin synthesis protects the mouse colonic crypt from DNA damage and colorectal tumorigenesis. J. Pathol. 249, 102–113 (2019).
- 146. Eckerling, A., Ricon-Becker, I., Sorski, L., Sandbank, E. & Ben-Eliyahu, S. Stress and cancer: mechanisms, significance and future directions. *Nat. Rev. Cancer* 21, 727–785 (2021).
- 21, 767–785 (2021).

 147. Hara, M. R. et al. A stress response pathway regulates DNA damage through β2-adrenoreceptors and β-arrestin-1. *Nature* 477, 349–353 (2011).
- 148. Sun, F. et al. Adrenergic DNA damage of embryonic pluripotent cells via β2 receptor signalling. *Sci. Rep.* 5, 15950 (2015).
- 149. Lamboy-Caraballo, R. et al. Norepinephrine-induced DNA damage in ovarian cancer cells. *Int. J. Mol. Sci.* https://doi.org/10.3390/ijms21062250 (2020).

- Dialynas, G. K., Vitalini, M. W. & Wallrath, L. L. Linking heterochromatin protein 1 (HP1) to cancer progression. *Mutat. Res.* 647, 13–20 (2008).
- 151. Ruginis, T., Taglia, L., Matusiak, D., Lee, B. S. & Benya, R. V. Consequence of gastrin-releasing peptide receptor activation in a human colon cancer cell line: a proteomic approach. *J. Proteome Res.* 5, 1460–1468 (2006).
- 152. Zeng, W., Ball, A. R. Jr. & Yokomori, K. HP1: heterochromatin binding proteins working the genome. *Epigenetics* 5, 287–292 (2010).
 153. Cornelio, D. B. Roesler, R. & Schwartsmann, G.
- 153. Cornelio, D. B., Roesler, R. & Schwartsmann, G. Gastrin-releasing peptide receptor as a molecular target in experimental anticancer therapy. *Ann. Oncol.* 18, 1457–1466 (2007).
- 154. Tell, R. et al. Gastrin-releasing peptide signaling alters colon cancer invasiveness via heterochromatin protein 1Hsß. Am. J. Pathol. 178, 672–678 (2011).
- 155. Schwartsmann, G. et al. A phase I trial of the bombesin/gastrin-releasing peptide (BN/GRP) antagonist RC3095 in patients with advanced solid malignancies. *Invest. New Drugs* 24, 403–412 (2006).
- Brunetto de Farias, C. et al. BDNF/TrkB content and interaction with gastrin-releasing peptide receptor blockade in colorectal cancer. *Oncology* 79, 430–439 (2010).
- 157. Rick, F. G. et al. Combination of gastrin-releasing peptide antagonist with cytotoxic agents produces synergistic inhibition of growth of human experimental colon cancers. Cell Cycle 11, 2518–2525 (2012).
- 158. Zhao, S. et al. Histone H3Q5 serotonylation stabilizes H3K4 methylation and potentiates its readout. *Proc. Natl Acad. Sci. USA* https://doi.org/10.1073/ pnas.2016742118 (2021).
- 159. Ye, D. et al. Targeting SERT promotes tryptophan metabolism: mechanisms and implications in colon cancer treatment. J. Exp. Clin. Cancer Res. 40, 173 (2021)
- Rabben, H. L. et al. Neural signaling modulates metabolism of gastric cancer. iScience 24, 102091 (2021).
 - Important study identifying a metabolic signature in gastric cancer as well as the reversibility of this metabolic reprogramming by vagotomy.
- 161. Song, Z., Wei, B., Lu, C., Li, P. & Chen, L. Glutaminase sustains cell survival via the regulation of glycolysis and glutaminolysis in colorectal cancer. *Oncol. Lett.* 14, 3117–3123 (2017).
- 162. Hao, Y. et al. Oncogenic PIK3CA mutations reprogram glutamine metabolism in colorectal cancer. *Nat. Commun.* 7, 11971 (2016).
- Oie, S. et al. Targeting glutamine-addiction and overcoming CDK4/6 inhibitor resistance in human esophageal squamous cell carcinoma. *Nat. Commun.* 10, 1296 (2019).
- 164. Zhao, Y. et al. Colorectal cancers utilize glutamine as an anaplerotic substrate of the TCA cycle in vivo. Sci. Rep. 9, 19180 (2019).
- 165. Shi, M., Liu, D., Yang, Z. & Guo, N. Central and peripheral nervous systems: master controllers in cancer metastasis. *Cancer Metastasis Rev.* 32, 603–621 (2013).
- 166. Jacobson, A., Yang, D., Vella, M. & Chiu, I. M. The intestinal neuro-immune axis: crosstalk between neurons, immune cells, and microbes. *Mucosal Immunol.* 14, 555–565 (2021).
- 167. Hodo, T. W., de Aquino, M. T. P., Shimamoto, A. & Shanker, A. Critical neurotransmitters in the neuroimmune network. Front. Immunol. 11, 1869 (2020)
- 168. Qiao, G. et al. β-Adrenergic signaling blocks murine CD8* T-cell metabolic reprogramming during activation: a mechanism for immunosuppression by adrenergic stress. Cancer Immunol. Immunother. 68, 11–22 (2019).
- 169. Salmon, H., Remark, R., Gnjatic, S. & Merad, M. Host tissue determinants of tumour immunity. Nat. Rev. Cancer 19, 215–227 (2019).
- 170. Wang, H. et al. Role of the nervous system in cancers: a review. *Cell Death Discov.* **7**, 76 (2021).
- 171. Rihawi, K. et al. Tumor-associated macrophages and inflammatory microenvironment in gastric cancer: novel translational implications. *Int. J. Mol. Sci.* 22, 3805 (2021).
- 172. Kitamura, T., Qian, B. Z. & Pollard, J. W. Immune cell promotion of metastasis. *Nat. Rev. Immunol.* **15**, 73–86 (2015).
- 173. Kuol, N., Stojanovska, L., Apostolopoulos, V. & Nurgali, K. Crosstalk between cancer and the neuro-immune system. *J. Neuroimmunol.* **315**, 15–23 (2018).

- 174. Qin, J.-F. et al. Adrenergic receptor β2 activation by stress promotes breast cancer progression through macrophages M2 polarization in tumor microenvironment. BMB Rep. 48, 295–300 (2015).
- 175. Lamkin, D. M. et al. β-Adrenergic-stimulated macrophages: comprehensive localization in the M1-M2 spectrum. *Brain Behav. Immun.* 57, 338–346 (2016).
- Gabanyi, I. et al. Neuro-immune interactions drive tissue programming in intestinal macrophages. *Cell* 164, 378–391 (2016).
- Matheis, F. et al. Adrenergic signaling in muscularis macrophages limits infection-induced neuronal loss. *Cell* 180, 64–78.e16 (2020).
- 178. Park, J. Y. et al. Polarized CD163⁺ tumor-associated macrophages are associated with increased angiogenesis and CXCL12 expression in gastric cancer. Clin. Res. Hepatol. Gastroenterol. 40, 357–365 (2016).
- Nakayama, Y. et al. Relationships between tumor-associated macrophages and clinicopathological factors in patients with colorectal cancer. *Anticancer Res* 22, 4291–4296 (2002).
- 180. Herrera, M. et al. Cancer-associated fibroblast and M2 macrophage markers together predict outcome in colorectal cancer patients. *Cancer Sci.* 104, 437–444 (2013).
- Koelzer, V. H. et al. Phenotyping of tumor-associated macrophages in colorectal cancer: impact on single cell invasion (tumor budding) and clinicopathological outcome. *Oncoimmunology* https://doi.org/10.1080/2 162402X.2015.1106677 (2016).
- 182. Zhong, X. M., Chen, B. & Yang, Z. W. The role of tumor-associated macrophages in colorectal carcinoma progression. *Cell Physiol. Biochem.* 45, 356–365 (2018).
- Pages, F. et al. International validation of the consensus immunoscore for the classification of colon cancer: a prognostic and accuracy study. *Lancet* 391, 2128–2139 (2018).
- 184. Zhang, Z. et al. Development of a prognostic signature for esophageal cancer based on nine immune related genes. *BMC Cancer* 21, 113 (2021).
- genes. *BMC Cancer* **21**, 113 (2021). 185. Zhang, N. et al. Prognostic role of tumor-infiltrating lymphocytes in gastric cancer: a meta-analysis and experimental validation. *Arch. Med. Sci.* **16**, 1092–1103 (2020).
- Pages, F. et al. Effector memory T cells, early metastasis, and survival in colorectal cancer. N. Engl. J. Med. 353, 2654–2666 (2005).
- 187. Costa, P. A. C. et al. Chemogenetic modulation of sensory neurons reveals their regulating role in melanoma progression. *Acta Neuropathol. Commun.* 9, 183 (2021).
- Dai, S. et al. Chronic stress promotes cancer development. Front. Oncol. 10, 1492 (2020)
- 189. Bucsek, M. J. et al. β-Adrenergic signaling in mice housed at standard temperatures suppresses an effector phenotype in CD8* T cells and undermines checkpoint inhibitor therapy. Cancer Res. 77, 5639–5651 (2017).
- 190. Nakai, A., Hayano, Y., Furuta, F., Noda, M. & Suzuki, K. Control of lymphocyte egress from lymph nodes through β2-adrenergic receptors. J. Exp. Med. 211, 2583–2598 (2014).
- Tavazoie, M. F. et al. LXR/ApoE activation restricts innate immune suppression in cancer. *Cell* 172, 825–840.e18 (2018).
- 192. Guo, W. et al. Prognostic value of PD-L1 in esophageal squamous cell carcinoma: a meta-analysis. *Oncotarget* 9, 13920–13933 (2018).
- 193. Li, Y. et al. The prognostic and clinicopathological roles of PD-L1 expression in colorectal cancer: a systematic review and meta-analysis. *Front. Pharmacol.* 10, 139 (2019).
- 194. Gu, L. et al. PD-L1 and gastric cancer prognosis: a systematic review and meta-analysis. PLoS ONE 12, e0182692 (2017).
- 195. Wang, L. et al. Immune sculpting of norepinephrine on MHC-I, B7-I, IDO and B7-H1 expression and regulation of proliferation and invasion in pancreatic carcinoma cells. *PLoS ONE* 7, e45491 (2012).
- 196. Mo, R. J. et al. Expression of PD-L1 in tumorassociated nerves correlates with reduced CD8+ tumor-associated lymphocytes and poor prognosis in prostate cancer. *Int. J. Cancer* **144**, 3099–3110 (2019).
- 197. Kuol, N. et al. Cholinergic signalling influences the expression of immune checkpoint inhibitors, PD-L1 and PD-L2, in human colorectal cancer tissues and cell lines. Preprint at Res. Sq. https://doi.org/10.21203/ rs.3.rs-665274/V1 (2021).

- 198. Hou, N. et al. A novel chronic stress-induced shift in the Th1 to Th2 response promotes colon cancer growth. *Biochem. Biophys. Res. Commun.* 439, 471–476 (2013).
- 199. Qiao, G. X. et al. Chronic adrenergic stress contributes to metabolic dysfunction and an exhausted phenotype in T cells in the tumor microenvironment. Cancer Immunol. Res. 9, 651–664 (2021).
- Mitsui, T. et al. Truncal vagotomy temporarily decreases the pro- and anti-inflammatory cytokine levels in the small intestine. Surg. Today 44, 1123–1127 (2014).
 Costa, A. C., Santos, J. M. O., Gil da Costa, R. M. &
- Costa, A. C., Santos, J. M. O., Gil da Costa, R. M. & Medeiros, R. Impact of immune cells on the hallmarks of cancer: a literature review. *Crit. Rev. Oncol. Hematol.* 168, 103541 (2021).
- 202. Saloman, J. L. et al. Ablation of sensory neurons in a genetic model of pancreatic ductal adenocarcinoma slows initiation and progression of cancer. *Proc. Natl Acad. Sci. USA* 113, 3078–3085 (2016).
- Barres, B. A. The mystery and magic of glia: a perspective on their roles in health and disease. *Neuron* 60, 430–440 (2008).
 Seguella, L. & Gulbransen, B. D. Enteric glial biology,
- Seguella, L. & Gulbransen, B. D. Enteric glial biology intercellular signalling and roles in gastrointestinal disease. Nat. Rev. Gastroenterol. Hepatol. 18, 571–587 (2021).
 - Prominent review discussing enteric glial heterogeneity and their functional roles in gastrointestinal pathophysiology.
- Rosenberg, H. J. & Rao, M. Enteric glia in homeostasis and disease: from fundamental biology to human pathology. iScience 24, 102863 (2021).
- Boesmans, W. et al. Structurally defined signaling in neuro-glia units in the enteric nervous system. *Glia* 67, 1167–1178 (2019).
- Ahmadzai, M. M., Seguella, L. & Gulbransen, B. D. Circuit-specific enteric glia regulate intestinal motor neurocircuits. *Proc. Natl Acad. Sci. USA* https://doi.org/ 10.1073/pnas.2025938118 (2021).
- 208. Vergnolle, N. & Cirillo, C. Neurons and glia in the enteric nervous system and epithelial barrier function. *Physiology* 33, 269–280 (2018).
- 209. Neunlist, M. et al. The digestive neuronal-glial-epithelial unit: a new actor in gut health and disease. Nat. Rev. Gastroenterol. Hepatol. 10, 90–100 (2013). Renowned review summarizing the importance of the ENS for maintenance of intestinal epithelial barrier integrity, thereby putting forward the digestive 'neuronal-glial-epithelial unit'.
- 210. Vales, S. et al. Tumor cells hijack enteric glia to activate colon cancer stem cells and stimulate tumorigenesis. EBioMedicine 49, 172–188 (2019). This paper is one of the first functional studies describing glia as part of the TME.
- Neunlist, M. et al. Enteric glia inhibit intestinal epithelial cell proliferation partly through a TGF-beta1dependent pathway. Am. J. Physiol. Gastrointest. Liver Physiol. 292, G231–G241 (2007).
- Liver Physiol. 292, G231–G241 (2007).
 212. Liu, Y. A. et al. 3-D imaging, illustration, and quantitation of enteric glial network in transparent human colon mucosa. Neurogastroenterol. Motil. 25, e324–e338 (2013).
- 213. Godlewski, J. & Kmiec, Z. Colorectal cancer invasion and atrophy of the enteric nervous system: potential feedback and impact on cancer progression. *Int. J. Mol. Sci.* https://doi.org/10.3390/ijms21093391 (2020).
- 214. Yuan, R. et al. Enteric glia play a critical role in promoting the development of colorectal cancer. Front. Oncol. https://doi.org/10.3389/fonc.2020 595892 (2020).
- 215. Baghdadi, M. B. et al. Enteric glial cell heterogeneity regulates intestinal stem cell niches. *Cell Stem Cell* 29, 86–100.e6 (2022).
- Progatzky, F. & Pachnis, V. Enteric glia bring fresh WNT to the intestinal stem cell niche. *Cell Stem Cell* 29, 3–4 (2022).
- Boesmans, W. et al. Imaging neuron-glia interactions in the enteric nervous system. Front. Cell Neurosci. https://doi.org/10.3389/fncel.2013.00183 (2013).
- Rolig, A. S. et al. The enteric nervous system promotes intestinal health by constraining microbiota composition. *PLoS Biol.* https://doi.org/10.1371/ journal.pbio.2000689 (2017).
- 219. Ge, Y. et al. Gut microbiota influence tumor development and Alter interactions with the human immune system. J. Exp. Clin. Cancer Res. 40, 42 (2021).
- De Vadder, F. et al. Gut microbiota regulates maturation of the adult enteric nervous system via enteric serotonin networks. *Proc. Natl Acad. Sci. USA* 115, 6458–6463 (2018).

REVIEWS

- Collins, J., Borojevic, R., Verdu, E. F., Huizinga, J. D. & Ratcliffe, E. M. Intestinal microbiota influence the early postnatal development of the enteric nervous system. *Neurogastroent. Motil.* 26, 98–107 (2014).
- 222. Vicentini, F. A. et al. Intestinal microbiota shapes gut physiology and regulates enteric neurons and glia. *Microbiome* 9, 210 (2021).
- 223. Kabouridis, P. S. et al. Microbiota controls the homeostasis of glial cells in the gut lamina propria. *Neuron* 85, 289–295 (2015).
- 224. Obata, Y. et al. Neuronal programming by microbiota regulates intestinal physiology. *Nature* **578**, 284–289 (2020).
- Sepich-Poore, G. D. et al. The microbiome and human cancer. Science https://doi.org/10.1126/science. abc4552 (2021).
 - A detailed overview of the known links between the microbiome and various types of cancer, including the microbiome–cancer–immune crosstalk.
- Lofgren, J. L. et al. Lack of commensal flora in Helicobacter pylori-infected INS-GAS mice reduces gastritis and delays intraepithelial neoplasia. Gastroenterology 140, 210–220 (2011).
- Zhan, Y. et al. Gut microbiota protects against gastrointestinal tumorigenesis caused by epithelial injury. *Cancer Res.* 73, 7199–7210 (2013).
- 228. Wong, S. H. & Yu, J. Gut microbiota in colorectal cancer: mechanisms of action and clinical applications. *Nat. Rev. Gastroenterol. Hepatol.* 16, 690–704 (2019).
 - Summary of the evidence linking CRC to the gut microbiome, with specific attention to the clinical applicability of this knowledge.
- Takiishi, T., Fenero, C. I. M. & Camara, N. O. S. Intestinal barrier and gut microbiota: shaping our immune responses throughout life. *Tissue Barriers* https://doi.org/10.1080/21688370.2017.1373208 (2017).
- Karin, M. & Clevers, H. Reparative inflammation takes charge of tissue regeneration. *Nature* **529**, 307–315 (2016).
- Zhou, J. & Boutros, M. JNK-dependent intestinal barrier failure disrupts host-microbe homeostasis during tumorigenesis. *Proc. Natl Acad. Sci. USA* 117, 9401–9412 (2020).
- 232. Lynch, S. V. & Pedersen, O. The human intestinal microbiome in health and disease. *N. Engl. J. Med.* **375**, 2369–2379 (2016).
- Ha, C. W. Y. et al. Translocation of viable gut microbiota to mesenteric adipose drives formation of creeping fat in humans. *Cell* 183, 666–683.e17 (2020)
- 234. Piscione, M., Mazzone, M., Di Marcantonio, M. C., Muraro, R. & Mincione, G. Eradication of helicobacter pylori and gastric cancer: a controversial relationship. *Front. Microbiol.* **12**, 630852 (2021).
- 235. Butt, J. & Epplein, M. Helicobacter pylori and colorectal cancer — a bacterium going abroad? *PLoS Pathog*. https://doi.org/10.1371/journal. ppat.1007861 (2019).
- Sticlaru, L. et al. Dangerous liaison: helicobacter pylori, ganglionitis, and myenteric gastric neurons: a histopathological study. *Anal. Cell Pathol.* https://doi.org/10.1155/2019/3085181 (2019).
- 237. Gorle, N., Bauwens, E., Haesebrouck, F., Smet, A. & Vandenbroucke, R. E. Helicobacter and the potential role in neurological disorders: there is more than Helicobacter pylori. *Front. Immunol.* https://doi.org/10.3389/fimmu.2020.584165 (2021).
- Ramirez, V. T. et al. T-cell derived acetylcholine aids host defenses during enteric bacterial infection with Citrobacter rodentium. *PLoS Pathog.* 15, e1007719 (2010)
- Colosimo, D. A. et al. Mapping interactions of microbial metabolites with human G-protein-coupled receptors. *Cell Host Microbe* 26, 273–282.e7 (2019).
- Chen, H. W. et al. A forward chemical genetic screen reveals gut microbiota metabolites that modulate host physiology. *Cell* 177, 1217–1231.e18 (2019).
- Choudhry, H. The microbiome and its implications in cancer immunotherapy. *Molecules* https://doi.org/ 10.3390/molecules26010206 (2021).
- 242. Wang, T. T. et al. Structural segregation of gut microbiota between colorectal cancer patients and healthy volunteers. *ISME J.* 6, 320–329 (2012).
- 243. Arthur, J. C. et al. Intestinal inflammation targets cancer-inducing activity of the microbiota. *Science* 338, 120–123 (2012).
 244. Tropepe, V. et al. Direct neural fate specification from
- 244. Tropepe, V. et al. Direct neural fate specification from embryonic stem cells: a primitive mammalian neural stem cell stage acquired through a default mechanism. *Neuron* 30, 65–78 (2001).

- 245. Smukler, S. R., Runciman, S. B., Xu, S. & van der Kooy, D. Embryonic stem cells assume a primitive neural stem cell fate in the absence of extrinsic influences. *J. Cell Biol.* 172, 79–90 (2006).
- Przyborski, S. A. Differentiation of human embryonic stem cells after transplantation in immune-deficient mice. Stem Cell 23, 1242–1250 (2005).
- 247. Germain, N. D. et al. Teratocarcinoma formation in embryonic stem cell-derived neural progenitor hippocampal transplants. *Cell Transpl.* 21, 1603–1611 (2012).
 248. Xu. L. et al. Neural stemness contributes to cell
- 248. Xu, L. et al. Neural stemness contributes to ce tumorigenicity. *Cell Biosci.* 11, 21 (2021).
- 249. Nagy, N. & Goldstein, A. M. Enteric nervous system development: a crest cell's journey from neural tube to colon. Semin. Cell Dev. Biol. 66, 94–106 (2017).
- to colon. Semin. Cell Dev. Biol. 66, 94–106 (2017).
 250. Munro, M. J., Wickremesekera, S. K., Peng, L., Tan, S. T. & Itinteang, T. Cancer stem cells in colorectal cancer: a review. J. Clin. Pathol. 71, 110–116 (2018).
- Sakakibara, S. et al. Mouse-musashi-1, a neural RNA-binding protein highly enriched in the mammalian CNS stem cell. *Dev. Biol.* 176, 230–242 (1996).
- 252. Ji, G. et al. PCGF1 promotes epigenetic activation of stemness markers and colorectal cancer stem cell enrichment. *Cell Death Dis.* 12, 633 (2021).
- 253. Guan, A. et al. MiR-330-3p inhibits gastric cancer progression through targeting MSI1. *Am. J. Transl. Res.* **8**, 4802–4811 (2016).
- 254. Gao, C. et al. Downregulation of Msi1 suppresses the growth of human colon cancer by targeting p21cip1. *Int. J. Oncol.* 46, 732–740 (2015).
- 255. Yang, W. et al. Identification of hub genes and therapeutic drugs in esophageal squamous cell carcinoma based on integrated bioinformatics strategy. Cancer Cell Int. 19, 142 (2019).
- 256. Yang, Y. et al. Identification of regulatory role of DNA methylation in colon cancer gene expression via systematic bioinformatics analysis. *Medicine* 96, e8487 (2017).
- 257. Fan, J. et al. Genome-wide DNA methylation profiles of low- and high-grade adenoma reveals potential biomarkers for early detection of colorectal carcinoma. *Clin. Epigenetics* 12, 56 (2020).
- Clin. Epigenetics 12, 56 (2020).
 258. Rademakers, G. et al. Identification of DNA methylation markers for early detection of CRC indicates a role for nervous system-related genes in CRC. Clin. Epigenetics 13, 80 (2021).
- Zhang, Z. et al. Similarity in gene-regulatory networks suggests that cancer cells share characteristics of embryonic neural cells. J. Biol. Chem. 292, 12842–12859 (2017).
 - One of the earliest reports that highlight neural stemness in cancer cells.
- 260. Lei, A. et al. EZH2 regulates protein stability via recruiting USP7 to mediate neuronal gene expression in cancer cells. Front. Genet. 10, 422 (2019).
- 261. Chang, P. Y. et al. Propranolol reduces cancer risk: a population-based cohort study. *Medicine* **94**, e1097 (2015).
- 262. Ahl, R. et al. Effects of beta-blocker therapy on mortality after elective colon cancer surgery: a Swedish nationwide cohort study. BMJ Open 10, e036164 (2020).
- 263. Reda, S. et al. Pre-operative beta-blocker therapy does not affect short-term mortality after esophageal resection for cancer. BMC Surg. 20, 333 (2020).
- 264. Fiala, O. et al. Incidental use of beta-blockers is associated with outcome of metastatic colorectal cancer patients treated with bevacizumab-based therapy: a single-institution retrospective analysis of 514 patients. Cancers https://doi.org/10.3390/ cancers11121856 (2019).
- Deng, Y. et al. Effects of antihypertensive drugs use on risk and prognosis of colorectal cancer: a meta-analysis of 37 observational studies. Front. Pharmacol. 12, 670657 (2021).
- 266. Liao, X. et al. Effects of propranolol in combination with radiation on apoptosis and survival of gastric cancer cells in vitro. *Radiat. Oncol.* 5, 98 (2010).
- MacDonald, C. R. et al. Adrenergic receptor signaling regulates the response of tumors to ionizing radiation. *Radiat. Res.* 191, 585–589 (2019).
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT03245554 (2018).
- 269. US National Library of Medicine. *ClinicalTrials.gov* https://clinicaltrials.gov/ct2/show/NCT04005365 (2019).
- 270. US National Library of Medicine. *ClinicalTrials.gov* https://clinicaltrials.gov/ct2/show/NCT04682158 (2021).
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT03919461 (2019).

- 272. Felton, J., Hu, S. & Raufman, J. P. Targeting M3 muscarinic receptors for colon cancer therapy. *Curr. Mol. Pharmacol.* 11, 184–190 (2018).
- 273. Ali, O., Tolaymat, M., Hu, S., Xie, G. & Raufman, J. P. Overcoming obstacles to targeting muscarinic receptor signaling in colorectal cancer. *Int. J. Mol. Sci.* https://doi.org/10.3390/ijms22020716 (2021).
- 274. US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT01822210 (2018)
- 275. Jin, W. Roles of TrkC signaling in the regulation of tumorigenicity and metastasis of cancer. *Cancers* https://doi.org/10.3390/cancers12010147 (2020).
- 276. US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT03556228 (2022).
- 277. US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT02650401 (2022).
 278. US National Library of Medicine. ClinicalTrials.gov
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT02219711 (2020).
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT02568267 (2022).
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT02576431 (2022).
- Aboody, K. S., Najbauer, J. & Danks, M. K. Stem and progenitor cell-mediated tumor selective gene therapy. *Gene Ther.* 15, 739–752 (2008).
- 282. Park, G. T., Kim, S. U. & Choi, K. C. Anti-proliferative effect of engineered neural stem cells expressing cytosine deaminase and interferon-beta against lymph node-derived metastatic colorectal adenocarcinoma in cellular and xenograft mouse models. *Cancer Res. Treat.* 49, 79–91 (2017).
- 283. Yi, B. R. et al. Suppression of the growth of human colorectal cancer cells by therapeutic stem cells expressing cytosine deaminase and interferon-beta via their tumor-tropic effect in cellular and xenograft mouse models. *Mol. Oncol.* 7, 543–554 (2013).
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT02015819 (2021).
- US National Library of Medicine. ClinicalTrials.gov https://clinicaltrials.gov/ct2/show/NCT03072134 (2022).
- 286. Sharma, M. & Flood, P. M. in *Neuroprotection* Ch. 4 https://doi.org/10.5772/intechopen.81343 (IntechOpen, 2019).
- 287. Costa, M., Brookes, S. J. & Hennig, G. W. Anatomy and physiology of the enteric nervous system. *Gut* **47** (Suppl. 4), iv15–iv19 (2000).
- 288. Brookes, S., Chen, N., Humenick, A., Spencer, N. J. & Costa, M. Extrinsic sensory innervation of the gut: structure and function. *Adv. Exp. Med. Biol.* 891, 63–69 (2016).
- Beyak, M. J. et al. in *Physiology of the Gastrointestinal Tract* 4th edn (ed. Johnsond, L. R.) 685–725 (Academic Press, 2006).
- 290. Ratcliffe, E. M. Molecular development of the extrinsic sensory innervation of the gastrointestinal tract. *Auton. Neurosci.* **161**, 1–5 (2011).
- Brierley, S. M. & Linden, D. R. Neuroplasticity and dysfunction after gastrointestinal inflammation. *Nat. Rev. Gastroenterol. Hepatol.* 11, 611–627 (2014).
- 292. Furness, J. B. Novel gut afferents: Intrinsic afferent neurons and intestinofugal neurons. *Auton. Neurosci.* 125, 81–85 (2006).
- 293. Saffrey, M. J. in *Encyclopedia of Neuroscience* (ed. Squire, L. R.) 1097–1102 (Academic Press, 2009).
- 294. Campbell, I. Gut motility and its control. *Anaesth. Intensive Care Med.* **13**, 59–61 (2012).
- 295. Mohs, F. E. & Lathrop, T. G. Modes of spread of cancer of skin. *AMA Arch. Derm. Syphilol.* **66**, 427–439 (1952).
- 296. Batsakis, J. G. Nerves and neurotropic carcinomas. *Ann. Otol. Rhinol. Laryngol.* **94**, 426–427 (1985).
- Veness, M. J. Perineural spread in head and neck skin cancer. *Australas. J. Dermatol.* 41, 117–119 (2000).
 Fagan, J. J. et al. Perineural invasion in squamous cell
- carcinoma of the head and neck. *Arch. Otolaryngol. Head Neck Surg.* **124**, 637–640 (1998).
- Bockman, D. E., Buchler, M. & Beger, H. G. Interaction of pancreatic ductal carcinoma with nerves leads to nerve damage. *Gastroenterology* 107, 219–230 (1994).
 Nagakawa, T. et al. Patterns of neural and plexus
- Nagakawa, I. et al. Patterns of neural and plexus invasion of human pancreatic cancer and experimental cancer. *Int. J. Pancreatol.* 10, 113–119 (1991).
- Liebig, C., Ayala, G., Wilks, J. A., Berger, D. H. & Albo, D. Perineural invasion in cancer: a review of the literature. *Cancer* 115, 3379–3391 (2009).
- 302. Ayala, G. E. et al. In vitro dorsal root ganglia and human prostate cell line interaction: redefining

- perineural invasion in prostate cancer. *Prostate* **49**, 213–223 (2001).
- 303. Mirkin, K. A. et al. Impact of perineural invasion on survival in node negative colon cancer. *Cancer Biol. Ther.* **18**, 740–745 (2017).
- 304. Hu, G., Li, L. & Hu, K. Clinical implications of perineural invasion in patients with colorectal cancer. *Medicine* 99. e 19860 (2020).
- Medicine 99, e19860 (2020).
 305. Duchalais, E. et al. Colorectal cancer cells adhere to and migrate along the neurons of the enteric nervous system. Cell Mol. Gastroenterol. Hepatol. 5, 31–49 (2018)
- 306. Bakst, R. L. et al. Perineural invasion and perineural tumor spread in head and neck cancer. *Int. J. Radiat. Oncol. Biol. Phys.* **103**, 1109–1124 (2019).
- 307. Chi, A. C., Katabi, N., Chen, H. S. & Cheng, Y. L. Interobserver variation among pathologists in evaluating perineural invasion for oral squamous cell carcinoma. *Head. Neck Pathol.* **10**, 451–464 (2016).
- 308. Schmitd, L. B., Scanlon, C. S. & D'Silva, N. J. Perineural invasion in head and neck cancer. *J. Dent. Res.* **97**, 742–750 (2018).
- 309. Thursby, E. & Juge, N. Introduction to the human gut microbiota. *Biochem. J.* 474, 1823–1836 (2017).
- Metchnikoff, E. The wilde medal and lecture of the manchester literary and philosophical society. *Br. Med. J.* 1, 1027–1028 (1901).

- Hyland, N. P. & Cryan, J. F. Microbe-host interactions: Influence of the gut microbiota on the enteric nervous system. *Dev. Biol.* 417, 182–187 (2016).
 Rodriguez, J. M. et al. The composition of the
- Rodriguez, J. M. et al. The composition of the gut microbiota throughout life, with an emphasis on early life. *Microb. Ecol. Health Dis.* 26, 26050 (2015).
- 313. Gagniere, J. et al. Gut microbiota imbalance and colorectal cancer. *World J. Gastroenterol.* **22**, 501–518 (2016).
- Natividad, J. M. M. & Verdu, E. F. Modulation of intestinal barrier by intestinal microbiota: pathological and therapeutic implications. *Pharmacol. Res.* 69, 42–51 (2013).
- Gensollen, T., Iyer, S. S., Kasper, D. L. & Blumberg, R. S. How colonization by microbiota in early life shapes the immune system. *Science* 352, 539–544 (2016).
- Baumler, A. J. & Sperandio, V. Interactions between the microbiota and pathogenic bacteria in the gut. *Nature* 535, 85–93 (2016).

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