



Published in final edited form as:

Anesthesiology. 2017 July ; 127(1): 58–69. doi:10.1097/ALN.0000000000001671.

Neurophysiologic Correlates of Ketamine Sedation and Anesthesia: a High-Density Electroencephalography Study in Healthy Volunteers

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Conflicts of interest: the authors declare no conflicts of interest

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Abstract

Background—Previous studies have demonstrated inconsistent neurophysiologic effects of ketamine, although discrepant findings might relate to differences in doses studied, brain regions analyzed, co-administration of other anesthetic medications, and resolution of the electroencephalograph. The objective of this study was to characterize the dose-dependent effects of ketamine on cortical oscillations and functional connectivity.

Methods—Ten healthy human volunteers were recruited for study participation. Data were recorded using a 128-channel electroencephalograph during baseline consciousness, subanesthetic dosing (0.5 mg/kg over 40 minutes), anesthetic dosing (1.5 mg/kg bolus), and recovery. No other sedative or anesthetic medications were administered. Spectrograms, topomaps, and functional connectivity (weighted and directed phase lag index) were computed and analyzed.

Results—Frontal theta bandwidth power (decibels, dB) increased most dramatically during ketamine anesthesia (mean power \pm standard deviation [SD], 4.25 ± 1.90 dB) compared to baseline (0.64 ± 0.28 dB), subanesthetic (0.60 ± 0.30 dB), and recovery (0.68 ± 0.41 dB) states; $P < 0.001$. Gamma power was also increased during ketamine anesthesia. Weighted phase lag index demonstrated theta phase locking within anterior regions (0.2349 ± 0.1170 , $P < 0.001$) and between anterior and posterior regions (0.2159 ± 0.1538 , $P < 0.01$) during ketamine anesthesia. Alpha power gradually decreased with subanesthetic ketamine, and anterior-to-posterior directed connectivity was maximally reduced (0.0282 ± 0.0772) during ketamine anesthesia compared to all other states; $P < 0.05$.

Conclusions—Ketamine anesthesia correlates most clearly with distinct changes in the theta bandwidth, including increased power and functional connectivity. Anterior-to-posterior connectivity in the alpha bandwidth becomes maximally depressed with anesthetic ketamine administration, suggesting a dose-dependent effect.

Introduction

Ketamine is a general anesthetic with unique features at the molecular, neural, and behavioral levels. Unlike the intravenous and inhaled anesthetics in common clinical use, ketamine is thought to work primarily by antagonizing N-methyl-D-aspartate (NMDA) receptors¹⁻⁴ and hyperpolarization-activated, cyclic-nucleotide gated (HCN)-1 channels,^{5,6} without strong agonist effects on synaptic gamma-aminobutyric acid (GABA) receptors. Furthermore, electroencephalographic characteristics of ketamine anesthesia are distinct from those associated with anesthetics that act primarily via GABA receptor agonism. For example, coherent frontal alpha oscillations are observed with propofol-induced unconsciousness,^{7,8} which are believed to result from thalamocortical hypersynchrony⁹ and which possibly inhibit corticocortical communication.⁸ These same patterns are noted with ether-based volatile anesthetics during surgical levels of unconsciousness.¹⁰ Ketamine anesthesia, however, is not associated with these electroencephalographic

characteristics.^{11,12} During anesthetic induction with ketamine, an increase in gamma power has been consistently reported,^{10,12,13} and when ketamine is administered in the presence of propofol or ether-based volatile anesthetics, there is a power shift to the beta bandwidth.^{14,15} These effects may be due to anti-NMDA-mediated disinhibition of pyramidal neurons¹⁶ and HCN-1 channel inhibition.¹⁵

Previous studies¹¹⁻¹³ have examined electroencephalographic effects of ketamine but the precise neurophysiologic correlates remain unclear for a number of reasons. For example, ketamine has often been studied in the presence of other concomitantly administered sedative and anesthetic medications,^{13,14,17} potentially altering or obfuscating neurophysiologic signatures specific to ketamine. Additionally, electroencephalogram studies to date have been constrained by either regionally limited analyses or low-resolution acquisition.¹¹⁻¹³ Studies have also focused on either subanesthetic^{18,19} or anesthetic dosing,¹¹⁻¹³ without being able to directly compare dose-dependent electroencephalographic characteristics. Lastly, there is a dearth of information regarding recovery from ketamine-induced anesthesia, as studies have tended to focus on the beginning of surgical cases, with GABAergic anesthetics often co-administered or administered directly after ketamine induction. Improving the understanding of ketamine-specific effects on the electroencephalogram may provide insights into the neuroscientific correlates of consciousness, particularly given the unique molecular, neural, and phenomenological features of ketamine that differ from other anesthetics. Thus, in this study, we used spectral and connectivity analyses of high-density electroencephalographic recordings to characterize neurophysiologic changes associated with ketamine as a single agent during subanesthetic administration, anesthetic dosing, and a recovery period. We hypothesized that ketamine would induce distinct, dose-dependent effects on spectral and functional connectivity patterns that would distinguish between these arousal states.

Materials and Methods

This study was approved by the University of Michigan Medical School Institutional Review Board (HUM00061087), and written informed consent was obtained from all participants prior to the study. All study procedures were conducted at the University of Michigan Medical School. Ten volunteers were recruited from September 2015 through January 2016 using recruitment flyers posted throughout the medical school and University Hospital of the University of Michigan. Phone screening was conducted by a member of the research team to review inclusion and exclusion criteria prior to enrollment.

Study Population

Participants were considered eligible if they were American Society of Anesthesiologists (ASA) Class 1 physical status, between the ages of 20-40, had a body mass index (BMI) <30, and had no predictors of a difficult airway. Candidates were excluded from participation if they had cardiovascular disease, cardiac conduction abnormalities, hypertension, obstructive sleep apnea, asthma, ongoing respiratory illness, gastroesophageal reflux, history of drug use (or positive drug screen prior to experiment), family history of problems with anesthesia, neurologic disorders, psychiatric disorders, or current pregnancy.

The number of volunteers recruited was based on previous studies that investigated neurophysiologic correlates of anesthetic-induced unconsciousness.^{7,20}

Anesthetic Protocol

Experiments were conducted between the hours of 8:00 a.m. and 1:00 p.m. for all participants except for one (Participant ID #2, studied between 4:00 p.m. and 7:00 p.m.). Prior to initiation of the experiment, a full medical and anesthetic history was obtained, and physical examination was performed. All participants fasted from food and drink for 8 hours prior to the experiment. Peripheral intravenous catheters were placed and ASA standard monitors were applied prior to drug administration. At least two anesthesiologists were present for the full duration of the experiment. All participants underwent the following stepwise protocol, with electroencephalogram data collection throughout each period:

1. A 5-minute eye-closed resting period prior to ketamine administration (baseline condition)
2. Subanesthetic ketamine infusion (0.5 mg/kg) over 40 minutes (subanesthetic condition) with eyes closed, followed by 8 mg ondansetron for nausea and vomiting prophylaxis
3. Break for completion of questionnaire (data not reported here)
4. Anesthetic (1.5 mg/kg) bolus dose (anesthetic condition) with eyes closed
5. Recovery period (recovery condition) with eyes closed

Participants were instructed to keep their eyes closed throughout each of these recording periods. Rather than using target-controlled infusions, we chose dosing strategies that were directly relevant to clinical care for either depression (0.5 mg/kg over 40 minutes) or anesthetic induction (1.5 mg/kg bolus) in order to enhance the translational relevance of this study. Participants were monitored through both loss of consciousness (LOC) and return of consciousness (ROC) using a previously described protocol;²⁰ we acknowledge that our definition of LOC relates to consciousness of the environment and does not exclude the possibility of endogenous experiences such as dreams or hallucinations. In brief, volunteers held an object in each hand that makes a sound when squeezed. An audio loop then started playing that instructed participants, in a random manner, to squeeze their right or left hand. This audio loop would deliver a command every 30 seconds, and the loop played before, during, and after anesthetic ketamine bolus administration. LOC was marked when subjects ceased to respond to two audio commands in a row. A scopolamine patch (1.5 mg) was also placed after ROC for further nausea and vomiting prophylaxis. Participants were then evaluated and monitored after the conclusion of the experiment. Additional antiemetics were administered if needed. Participants were discharged once they were awake, alert, and responsive, had no significant nausea or vomiting, were able to ambulate with minimal assistance, and had a responsible adult to accompany them home. Follow-up calls were made to patients both the evening after the experiment and the next day.

Electroencephalography Analysis

Data Acquisition—Electroencephalogram data were acquired with 128-channel Hydrocel Nets (Electrical Geodesics, Inc., Eugene, OR), Netamps 400 amplifiers, and Netstation 4.5 software. The electroencephalogram was digitized continuously at 500Hz with a vertex reference. Per manufacturer recommendations, channel impedances were kept below 50 k Ω , and the net was wrapped with gauze to optimize contact between the electrodes and scalp.

Spectral Processing and Analysis—Data processing was performed with Chronux (<http://chronux.org>)^{21,22} and custom MATLAB (MathWorks, Natick, MA) scripts and toolboxes. All visual inspection was performed by one of the investigators (TB) trained in dense electroencephalography review. After each recording session, the data were band-pass filtered at 0.5 to 55 Hz [*eegiltnew*, *firfilt plugin*, *zero-phase*, *Hamming-windowed FIR filter*, *3301 points (6.6s)*, *0.5 Hz transition band*, *0.5 and 55 passband edges*, *-6 dB cutoff frequencies: 0.25 and 55.25*] to decrease the influence of low-frequency drift and high-frequency artifacts. Electrodes on the lowest parts of the face and head were removed, leaving 98 remaining channels. Bad channels were then detected and removed using visual inspection and the *rejchan* and *clean_rawdata* functions, with 86 to 91 channels retained. The channels were then average-referenced to the mean of the voltage across all remaining channels. Conditions of interest – the baseline eyes-closed period (5 minutes), subanesthetic infusion (0 to 38min), anesthetic bolus dose (from LOC to 5 minutes after LOC, 3.8 minutes for one volunteer), and recovery period (5 minutes, collected 8 to 15 minutes after ROC) – were extracted from the data for each participant. These specific time epochs were chosen to (1) include data relatively preserved from artifact, and (2) represent stable neurophysiologic periods for each of the four experimental conditions. Large artifacts were removed after visual inspection and the *rejcont* function. Remaining periods of continuous data were segmented into 3-second epochs. Data epochs with remaining large artifacts were removed via a combination of visual inspection and the *rej Kurt*, *rejtrend*, and *eegthresh* functions. For each participant, the remaining epochs from the four periods of interest were submitted to independent component analysis (*runica* function, *infomax*, extended). Independent components representing eyeblink, lateral eye, muscle, facial electromyography, focal channel noise, and focal trial noise were removed from all epochs using visual inspection and the following component classification plugins: SASICA,²³ IC-MARC,²⁴ and ADJUST.²⁵ All remaining epochs for each period and each participant were then visually inspected again to remove any remaining epochs with excessive artifacts. For the four conditions (Baseline, Subanesthetic, Anesthetic, Recovery), the mean remaining trial counts were 54 (SD=17), 574 (SD=48), 66 (SD=8), and 73 (SD=11), respectively. The previously removed channels in each dataset were interpolated to the 98 channel subset. Spectral power was computed with multitaper spectral analyses (*mtspecgramc* function; time window: 3s, overlap: .5s, number of tapers: 3, time-bandwidth product: 5, spectral resolution: .25 Hz). The median absolute power ($10 \cdot \log_{10}[\mu\text{V}^2/\text{Hz}]$) was calculated for each of the four experimental periods at each of five frequency bands (delta: 1-4Hz, theta: 4-8Hz, alpha: 8-13Hz, beta: 13-30Hz, gamma: 30-48Hz) for all 98 channels, and then for eight frontal channels centered on the Fz site (see figure, Supplemental Digital Content 1). The final data presented in the spectrograms are the sequential, 3-second epochs (grand-averaged across all participants) that remained after the data cleaning steps described above were implemented.

Data were temporally sequential, but not necessarily contiguous given that some epochs were removed after cleaning and artifact removal. Images presented are log-transformed ($10 \cdot \log_{10}$ transform of the grand average of the single-subject data). Recovery period data for one participant were unavailable and therefore recovery period calculations were completed for only nine participants.

Topographic Analysis—The topographic maps of spectral power for each experimental condition and electroencephalogram bandwidth were constructed using the `topoplot` function in the MATLAB toolbox EEGLAB.²⁶

Connectivity Processing and Analysis—The selected continuous data epochs from the spectral analyses were used for connectivity analysis without channel interpolation or independent component analysis (ICA); this was done to preserve the phase information of the original signals.^{27,28} Additionally, eight occipital channels were removed due to excessive artifact presence, which was deemed necessary for connectivity analyses, leaving 90 remaining channels. The functional connectivity between electroencephalogram channels was examined across all experimental conditions. The undirected and directed connectivity were measured with debiased weighted Phase Lag Index (wPLI)²⁹ and directed Phase Lag Index (dPLI),³⁰ respectively. The wPLI is a measure of how the instantaneous phases of two electroencephalogram signals are phase-locked to each other. If the instantaneous phase of one signal is consistently ahead or behind of the other signal, the phases are considered locked, and $wPLI = 1$. However, if the signals randomly alternate between a phase lead and a phase lag relationship, there is no phase locking and $wPLI = 0$. The directionality of functional connectivity between two electroencephalogram signals was determined with the asymmetry of the phase lead/lag relationship. If the instantaneous phase of one signal consistently leads that of the other signal, $dPLI = 1$, and with the inverse case, $dPLI = -1$. If there is no a bias in the phase lead/lag relationship, then $dPLI = 0$. In practice, both measures are robust to the volume conduction problem of scalp electroencephalography. Electroencephalogram signals were divided into one-minute long epochs with 50% overlap, which was further divided into 2 second non-overlapping sub-epochs. For each sub-epoch, the cross-spectral density was estimated using the multitaper method, with time-bandwidth product $TW = 2$ and number of tapers $K = 3$,^{21,22} and from these repetitions the averaged wPLI and dPLI values at variable frequencies were estimated using a custom-written function adapted from the Fieldtrip toolbox.³¹ To remove the bias of the measures for a given electroencephalogram data set, the shuffled-data method was used. A series ($N = 20$) of shuffled signal pairs were generated for each pair of electroencephalogram signals, and the wPLI and dPLI measures were calculated with the shuffled data, the mean of which was subtracted from the raw wPLI and dPLI values as the final estimation of undirected and directed connectivity.³² Final baseline and recovery time epochs were chosen as 2-minute segments selected in the middle of each respective recording. Subanesthetic epochs were selected as two sequential 2-minute epochs (subanes-1, subanes-2) that were representative of the observed spectral power during the subanesthetic infusion period. Anesthetic epochs (anes-1, anes-2) were chosen and displayed in the same manner.

Statistical Analysis

We tested for differences between the experimental periods (Baseline, Subanesthetic, Anesthetic, Recovery) by entering the single-subject electroencephalogram-derived values into a linear mixed model (LMM) analysis in SPSS 22. LMM analysis allowed for statistical comparisons in the event of partially missing data from a given participant (as was the case with missing recovery period data from one volunteer, noted above). Each experimental period served as the fixed factor with participant data serving as a correlated random effect. We used restricted maximum likelihood estimation and diagonal covariance structure with heterogeneous variances and zero correlation between elements. Direct pairwise comparisons were adjusted using Bonferroni's method. We did not model repeated covariance effects and a *P*-value less than 0.05 was considered statistically significant. The average of median connectivity values was reported for connectivity data, as has been previously described methodologically.⁷ All variability estimates are in standard deviation (SD).

Results

All study participants successfully completed subanesthetic and anesthetic dosing protocols; 6/10 (60%) experienced nausea and vomiting after ketamine anesthesia and 5/10 (50%) required additional antiemetic treatment. One volunteer (1/10, 10%) briefly required a chin lift and jaw thrust to maintain airway patency shortly after LOC. No adverse clinical events were otherwise noted. All 10 participants experienced LOC after the 1.5 mg/kg ketamine bolus, and the mean time between bolus administration and LOC was 1.23 (\pm 0.35) minutes and LOC lasted an average of 10.68 (\pm 3.51) minutes.

Dose-Dependent Global Effects of Ketamine – Spectral and Topographic Analysis

Prominent changes were apparent across theta, alpha, and gamma bandwidths. There was a marked increase in theta power during ketamine anesthesia (figs. 1, 2, and Supplemental Digital Content 2 for individual participants), and this power localized to both frontal and posterior channels (fig. 2). There was a reduction in posterior alpha power from baseline through anesthetic dosing and no increase or anteriorization of alpha power was noted with either ketamine dose (fig. 2). Increased gamma power was also apparent during the anesthetic epoch compared to other states (figs. 1, 2).

Frontal Channel Cluster – Ketamine-Induced Changes across Brain States

Frontal channels are of particular interest given their accessibility and use in the operating room. During the subanesthetic infusion, a gradual dissipation of alpha power was noted compared to baseline (mean power \pm standard deviation [SD], decibels [dB] – 0.46 \pm 0.34 dB and 0.88 \pm 0.80 dB, respectively), and alpha power remained decreased during the periods of ketamine anesthesia (0.49 \pm 0.22 dB) and recovery (0.34 \pm 0.38 Db), $F(3,26)=3.679$, $P<0.05$ (fig. 3A and B). With anesthetic dosing, there was a significant increase in theta bandwidth power (4.25 \pm 1.90 dB) compared to baseline (0.64 \pm 0.28 dB), subanesthetic (0.60 \pm 0.30 dB), and recovery (0.68 \pm 0.41 dB) periods, $F(3,26)=41.01$, $P<0.001$ (fig. 3A and B, Table 1). Theta power was not otherwise significantly modulated during non-anesthetic periods, and relative increases in theta power during ketamine anesthesia were higher than that of any other bandwidth (Table 1).

Functional Connectivity Analyses

Given the significant increases in theta power and decreases in alpha power with progressive ketamine dosing, we undertook connectivity analyses in these bandwidths across each condition. Reduced functional connectivity in the alpha bandwidth has also been previously demonstrated with anesthetic ketamine dosing,¹¹ further motivating our choice of bandwidths for analysis. Although we noted marked increases in gamma bandwidth power, electromyographic artifact might contribute to gamma activity despite our data cleaning.^{33,34} Thus, we did not examine gamma connectivity in this study. Furthermore, subanesthetic ketamine has been shown to increase gamma power in other paradigms,^{35,36} whereas increased theta power seems to correlate specifically to the anesthetized state.^{19,37,38}

Weighted Phase Lag Index – Theta Phase-Locking during Ketamine

Anesthesia—Figure 4 presents the global and regional wPLI changes across each experimental condition. Figure 4A shows the average median wPLI between anterior and posterior regions in the time and frequency domains (see Supplemental Digital Content 3, for individual participants). Alpha wPLI was dominant at baseline and increased during the initial phase of the subanesthetic infusion (fig. 4A). During the anesthetic period, however, wPLI connectivity strength shifted to the theta bandwidth (fig. 4A). Figure 4B presents the topographic changes of global wPLI networks for the theta and alpha bandwidths across experimental conditions. Within the anterior region, theta wPLI was highest during the anesthetic period (mean wPLI \pm SD; 0.2349 ± 0.1170) compared to all other states; $F(5,44)=7.996$, $P<0.001$ (fig. 3B). Theta wPLI was also highest during ketamine anesthesia between anterior and posterior regions (0.2159 ± 0.1538); $F(5,44)=4.192$, $P<0.01$ (fig. 3B). Alpha wPLI within the anterior region remained relatively unchanged throughout each study condition ($F(5,44)=0.697$, $P=0.628$), although significant changes were noted between anterior and posterior regions, with mean wPLI lowest (0.0833 ± 0.1083) during the anesthetic period; $F(5,44)=3.147$, $P<0.05$ (fig. 4B).

Directed Phase Lag Index – Directional Connectivity Changes during

Ketamine Anesthesia—Figure 5 demonstrates how the directionality of functional connectivity changed across the experimental conditions. Figure 5A shows the average median dPLI between anterior and posterior electrodes in the time and frequency domains (see Supplemental Digital Content 3, for individual participants). An anterior-to-posterior directionality (the positive dPLIs denoted with red color) was observed in the alpha bandwidth during the baseline and subanesthetic periods. Theta shows a consistently opposite directionality from posterior-to-anterior regions (the negative dPLI denoted with blue color). Figure 5B shows the frequency-specific and state-specific topographies of electroencephalographic directionality. Alpha dPLI was maximally reduced between anterior and posterior channels during ketamine anesthesia (mean dPLI \pm SD; 0.0282 ± 0.0772); $F(5,44)=2.484$, $P<0.05$ (fig. 4B). Alpha dPLI was also examined between prefrontal and frontal channels, and was again maximally reduced during ketamine anesthesia (-0.0224 ± 0.0400); $F(5,44)=4.137$, $P<0.01$ (fig. 4B). Compared to baseline (0.0427 ± 0.0717), direct pairwise comparisons (with Bonferroni correction) revealed a statistically significant reduction in alpha dPLI only during each of the anesthetic epochs (Anes-1, -0.0170 ± 0.0481 , $P<0.05$; Anes-2, -0.0224 ± 0.0400 , $P<0.05$). Thus, an overall maximally reduced

anterior-to-posterior connectivity pattern was demonstrated with alpha dPLI during ketamine anesthesia among these channel regions. Although a posterior-to-anterior pattern was noted with theta dPLI during ketamine anesthesia (fig. 5B), these associations did not reach statistically significant differences between anterior and posterior channels ($F(5,44)=0.177$, $P=0.970$) or for prefrontal and frontal channels ($F(5, 44)=0.768$, $P=0.578$).

Discussion

This study characterized ketamine-induced electroencephalographic changes during subanesthetic, anesthetic, and recovery periods (relative to baseline) in healthy volunteers using high-density electroencephalography. During subanesthetic ketamine administration, spectral power gradually (and modestly) shifts from the alpha bandwidth to the theta bandwidth, with anterior-to-posterior connectivity (as measured by alpha dPLI) maintained. During ketamine anesthesia, however, the power shifts dramatically to the theta bandwidth, theta wPLI increases in anterior and posterior regions, and anterior-to-posterior alpha connectivity (as measured by alpha dPLI) is significantly reduced. Upon recovery, these connectivity patterns return to near baseline levels in each bandwidth. Ketamine anesthesia was also associated with increased gamma and delta power, as previously reported,^{10,12,13} and frontal gamma power remained slightly elevated compared to baseline.

Our results demonstrate increased theta power correlating with ketamine anesthesia. Of note, increased theta oscillatory activity during ketamine anesthesia was first documented more than five decades ago. In the seminal human study that first examined pharmacologic effects of ketamine in 1965, Domino et al. remarked, “Characteristic thetalike waves were recorded at dosage levels which produced coma; these were distributed over most of the cerebral hemisphere.”³⁷ Increased theta activity has since been documented during anesthetic ketamine administration.^{13,15,38} A strictly receptor- and channel-based explanation for this theta resonance may be difficult to describe, because ketamine has a considerably diverse array of molecular targets.^{1,2,5,6,39-42} A network-level consideration of the phenomenon might provide insight regarding the consequences for information processing. In the awake state, theta and alpha frequencies interact in a circular, reciprocating pattern of “information flow” between anterior and posterior brain regions.^{43,44} Although oscillatory amplitude and phase are theoretically independent, amplitude (and thus, power) may associate with phase in order to facilitate interactions with other neuronal populations within a network.⁴⁵ As alpha power and anterior-to-posterior connectivity becomes maximally depressed during ketamine anesthesia, increased theta power and phase-locking between anterior and posterior regions may represent large-scale network disequilibrium. This is, however, a speculative interpretation and additional studies are needed to further explore the neurobiologic relevance of such connectivity patterns. From a clinical perspective, the distinct appearance of theta power during anesthesia may be informative for clinicians who routinely administer ketamine. The appearance of increasing theta power could alert clinicians to anesthetic ketamine dosing, which may be particularly helpful in settings where subanesthetic dosing is desired (as in the treatment of depression).^{46,47} As the global and frontal spectrograms in this study both illustrated marked increases in theta power during ketamine anesthesia, the use of frontal electroencephalogram channels alone may be sufficient to monitor for this theta signature.

It is notable that ketamine anesthesia is associated with a maximal depression of anterior-to-posterior connectivity in the alpha bandwidth. Inhibition of frontal-to-parietal connectivity (with certain measures) has correlated with general anesthesia in humans across a diverse range of anesthetics,^{12,32,48,49} and our group has previously demonstrated reduced alpha directional connectivity during ketamine anesthesia with low-resolution EEG.¹¹ Recently, disruptions in NMDA- and AMPA-mediated frontoparietal connectivity patterns have been demonstrated during subanesthetic ketamine administration in healthy volunteers,³⁵ who remained conscious during these experiments. This raises the possibility that disrupted frontal-to-parietal connectivity may not strictly correlate with anesthetic-induced unconsciousness. An alternative explanation, however, is that such connectivity was not maximally depressed during these subanesthetic infusion periods. Findings from our study demonstrate that alpha directional connectivity remained relatively unchanged during subanesthetic dosing, decreased dramatically during anesthetic ketamine administration, and returned to baseline levels at recovery. The connectograms (figs. 4A, 5A) demonstrate significant functional connectivity in the alpha bandwidth during baseline and subanesthetic states—despite a gradual reduction of power—and this connectivity diminishes significantly during ketamine anesthesia. Thus, there may be a dose-dependent relationship between connectivity and loss of consciousness. In fact, Untergerhrer et al. illustrated dynamic fluctuations in functional connectivity patterns, whereby maximum functional connectivity occurs during varying time intervals on a continuous scale, with surrogates of information transfer times being fastest during unconsciousness.⁵⁰ Accordingly, connectivity is a dynamic process and may need to cross certain real-time, continuous thresholds to correlate with varying levels of consciousness.

Multiple and unique strengths of this study are worth highlighting. No other sedative, opioid, or hypnotic medications were administered; thus, electroencephalographic effects of ketamine were not confounded by co-administration of other psychoactive medications. Additionally, an advanced data cleaning process was used for spectral analysis. In addition to visual inspection, data from each time epoch underwent independent component analysis blind-source separation, and independent components representing eye blink, eye muscle, facial muscle, channel-noise, and single-trial artifact were removed. Analytic comparisons were also made across multiple states – we were able to compare effects of ketamine before administration, at both subanesthetic and anesthetic doses, and during recovery. High-density electroencephalography was utilized, allowing for high-resolution and multi-regional analyses. Lastly, functional connectivity based on phase relationships was also assessed with the use of wPLI and dPLI. To our knowledge, this is the first study to combine all of these methodologic strengths for the electroencephalographic analysis of ketamine in humans (Table 2).

There are significant limitations to the study as well. Administering the subanesthetic ketamine infusion immediately prior to anesthetic dosing may influence neurophysiologic patterns observed during the anesthetic period. Our spectral data demonstrate a gradual reduction in alpha bandwidth power and increased theta power during the subanesthetic infusion. Whether this preceding subanesthetic exposure modulates subsequent neurophysiology and network connectivity during anesthetic dosing is unclear. Additionally, although a 128-channel electroencephalogram system was used, 86 to 91 channels were

ultimately retained for spectral analyses after removal of artifact and bad channels. These artifact removal strategies may also yield spectral patterns that differ from a real-world, clinical setting. For example, ocular artifact removal may have attenuated changes in delta power that might otherwise be present.⁵¹ As we wanted to optimize electroencephalogram data for connectivity analyses, we did not employ the same artifact-removal strategies (e.g., ICA, visual inspection) as the spectral analyses, because these may interfere with phase synchronization data.^{27,28} Thus, spectral and connectivity data results emerged from different data processing steps. The administration of both ondansetron and scopolamine could potentially have modulated central neurophysiology, but we regarded this as unlikely and both of these medications were ultimately warranted for participant comfort. Participants were instructed to keep their eyes closed during each experimental period, but sporadic eye opening was noted throughout recording sessions. This could have impacted certain epochs of the neurophysiologic data acquisition and analysis.

There are also significant limitations to consider regarding connectivity analyses. Although connectivity measures (in this case, wPLI and dPLI) are used as a surrogate for functional interactions across the cortex, this is merely an assumption. Our recent data in nonhuman primates confirm a breakdown of corticocortical information transfer during ketamine anesthesia,⁵² but different connectivity measures or experimental conditions can yield disparate results.⁴³ Phase lag index can also be associated with significant inter-subject variability, particularly given the complex neural processes associated with these phase-based methods; as a result, PLI is often not amenable to statistical averaging or traditional artifact reduction strategies.⁵³ In terms of the general interpretation of directional connectivity results, it must be kept in mind that these measures reflect a large spatial scale and a long temporal scale, with an unclear relationship to the underlying neural spiking networks. As an intuitive example, the Dow Jones Industrial Average samples the performance of only a select number of publically-traded companies. As a single value that rises or falls, it does not capture the rich dynamics and widespread stock exchange on the trading room floor. Similarly, directed connectivity measures sample features that might reflect large-scale cortical interactions but they do not capture the neuronal dynamics and widespread information exchange in cortical and subcortical systems. With that being said, changes in directed connectivity measures and surrogates of information transfer observed in the brain during anesthetic-induced unconsciousness have been shown to reflect fundamental network properties.⁴⁵

In summary, this study utilized high-resolution electroencephalographic data to characterize ketamine-induced changes across multiple doses and associated levels of consciousness. Notably, increased theta power and phase locking occur between anterior and posterior regions during ketamine anesthesia, returning to baseline upon recovery. Anterior-to-posterior connectivity in the alpha bandwidth was maximally suppressed during ketamine anesthesia and this connectivity is also restored to baseline levels during recovery. This constellation of power and connectivity results (increased theta power, as seen during rapid eye movement sleep, coupled with loss of anterior-to-posterior alpha connectivity, as seen during anesthesia) might contribute to the unique qualities of ketamine anesthesia.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

The authors would like to thank Yumeng Li, M.S. (Graduate Student Research Assistant, Center for Statistical Consultation and Research, University of Michigan, Ann Arbor, Michigan) for statistical consultation.

Supported by the National Institutes of Health, Bethesda, Maryland, Grants T32GM103730 (Dr. Vlisides-Trainee, Dr. Mashour-Principal Investigator) and R01GM111293 (Dr. Mashour), and the Department of Anesthesiology, University of Michigan, Ann Arbor, Michigan.

References

1. Anis NA, Berry SC, Burton NR, Lodge D. The dissociative anaesthetics, ketamine and phencyclidine, selectively reduce excitation of central mammalian neurones by N-methyl-aspartate. *Br J Pharmacol.* 1983; 79:565–75. [PubMed: 6317114]
2. Thomson AM, West DC, Lodge D. An N-methylaspartate receptor-mediated synapse in rat cerebral cortex: a site of action of ketamine? *Nature.* 1985; 313:479–81. [PubMed: 2982106]
3. Yamamura T, Harada K, Okamura A, Kemmotsu O. Is the site of action of ketamine anesthesia the N-methyl-D-aspartate receptor? *Anesthesiology.* 1990; 72:704–10. [PubMed: 1969718]
4. Petrenko AB, Yamakura T, Fujiwara N, Askalany AR, Baba H, Sakimura K. Reduced sensitivity to ketamine and pentobarbital in mice lacking the N-methyl-D-aspartate receptor GluReceptor1 subunit. *Anesth Analg.* 2004; 99:1136–40. [PubMed: 15385364]
5. Chen X, Shu S, Bayliss DA. HCN1 channel subunits are a molecular substrate for hypnotic actions of ketamine. *J Neurosci.* 2009; 29:600–9. [PubMed: 19158287]
6. Zhou C, Douglas JE, Kumar NN, Shu S, Bayliss DA, Chen X. Forebrain HCN1 channels contribute to hypnotic actions of ketamine. *Anesthesiology.* 2013; 118:785–95. [PubMed: 23377220]
7. Purdon PL, Pierce ET, Mukamel EA, Prerau MJ, Walsh JL, Wong KF, Salazar-Gomez AF, Harrell PG, Sampson AL, Cimenser A, Ching S, Kopell NJ, Tavares-Stoeckel C, Habeeb K, Merhar R, Brown EN. Electroencephalogram signatures of loss and recovery of consciousness from propofol. *Proc Natl Acad Sci U S A.* 2013; 110:E1142–51. [PubMed: 23487781]
8. Lewis LD, Weiner VS, Mukamel EA, Donoghue JA, Eskandar EN, Madsen JR, Anderson WS, Hochberg LR, Cash SS, Brown EN, Purdon PL. Rapid fragmentation of neuronal networks at the onset of propofol-induced unconsciousness. *Proc Natl Acad Sci U S A.* 2012; 109:E3377–86. [PubMed: 23129622]
9. Vijayan S, Ching S, Purdon PL, Brown EN, Kopell NJ. Thalamocortical mechanisms for the anteriorization of alpha rhythms during propofol-induced unconsciousness. *J Neurosci.* 2013; 33:11070–5. [PubMed: 23825412]
10. Purdon PL, Sampson A, Pavone KJ, Brown EN. Clinical Electroencephalography for Anesthesiologists: Part I: Background and Basic Signatures. *Anesthesiology.* 2015; 123:937–60. [PubMed: 26275092]
11. Blain-Moraes S, Lee U, Ku S, Noh G, Mashour GA. Electroencephalographic effects of ketamine on power, cross-frequency coupling, and connectivity in the alpha bandwidth. *Front Syst Neurosci.* 2014; 8:114. [PubMed: 25071473]
12. Lee U, Ku S, Noh G, Baek S, Choi B, Mashour GA. Disruption of frontal-parietal communication by ketamine, propofol, and sevoflurane. *Anesthesiology.* 2013; 118:1264–75. [PubMed: 23695090]
13. Akeju O, Song AH, Hamilos AE, Pavone KJ, Flores FJ, Brown EN, Purdon PL. Electroencephalogram signatures of ketamine anesthesia-induced unconsciousness. *Clin Neurophysiol.* 2016; 127:2414–22. [PubMed: 27178861]
14. Tsuda N, Hayashi K, Hagihira S, Sawa T. Ketamine, an NMDA-antagonist, increases the oscillatory frequencies of alpha-peaks on the electroencephalographic power spectrum. *Acta Anaesthesiol Scand.* 2007; 51:472–81. [PubMed: 17378787]

15. Bojak I, Day HC, Liley DT. Ketamine, Propofol, and the EEG: A neural field analysis of HCN1-mediated interactions. *Front Comput Neurosci*. 2013; 7:22. [PubMed: 23576979]
16. Olney JW, Newcomer JW, Farber NB. NMDA receptor hypofunction model of schizophrenia. *J Psychiatr Res*. 1999; 33:523–33. [PubMed: 10628529]
17. Kurehara K, Asano N, Iwata T, Yamaguchi A, Kawano Y, Furuya H. The influence of ketamine on the bispectral index, the spectral edge frequency 90 and the frequency bands power during propofol anesthesia. *Masui*. 1999; 48:611–6. [PubMed: 10402812]
18. Shaw AD, Saxena N, L EJ, Hall JE, Singh KD, Muthukumaraswamy SD. Ketamine amplifies induced gamma frequency oscillations in the human cerebral cortex. *Eur Neuropsychopharmacol*. 2015; 25:1136–46. [PubMed: 26123243]
19. Kochs E, Scharein E, Mollenberg O, Bromm B, Schulte am Esch J. Analgesic efficacy of low-dose ketamine. Somatosensory-evoked responses in relation to subjective pain ratings. *Anesthesiology*. 1996; 85:304–14. [PubMed: 8712446]
20. Blain-Moraes S, Tarnal V, Vanini G, Alexander A, Rosen D, Shortal B, Janke E, Mashour GA. Neurophysiological correlates of sevoflurane-induced unconsciousness. *Anesthesiology*. 2015; 122:307–16. [PubMed: 25296108]
21. Bokil H, Andrews P, Kulkarni JE, Mehta S, Mitra PP. Chronux: a platform for analyzing neural signals. *J Neurosci Methods*. 2010; 192:146–51. [PubMed: 20637804]
22. Mitra, PP., Bokil, H. *Observed Brain Dynamics*. New York: Oxford University Press; 2008.
23. Chaumon M, Bishop DV, Busch NA. A practical guide to the selection of independent components of the electroencephalogram for artifact correction. *J Neurosci Methods*. 2015; 250:47–63. [PubMed: 25791012]
24. Frolich L, Andersen TS, Morup M. Classification of independent components of EEG into multiple artifact classes. *Psychophysiology*. 2015; 52:32–45. [PubMed: 25048104]
25. Mognon A, Jovicich J, Bruzzone L, Buiatti M. ADJUST: An automatic EEG artifact detector based on the joint use of spatial and temporal features. *Psychophysiology*. 2011; 48:229–40. [PubMed: 20636297]
26. Delorme A, Makeig S. EEGLAB: an open source toolbox for analysis of single-trial EEG dynamics including independent component analysis. *J Neurosci Methods*. 2004; 134:9–21. [PubMed: 15102499]
27. Meinecke FC, Ziehe A, Kurths J, Muller KR. Measuring phase synchronization of superimposed signals. *Phys Rev Lett*. 2005; 94:084102. [PubMed: 15783894]
28. Nolte G, Meinecke FC, Ziehe A, Muller KR. Identifying interactions in mixed and noisy complex systems. *Phys Rev E Stat Nonlin Soft Matter Phys*. 2006; 73:051913. [PubMed: 16802973]
29. Vinck M, Oostenveld R, van Wingerden M, Battaglia F, Pennartz CM. An improved index of phase-synchronization for electrophysiological data in the presence of volume-conduction, noise and sample-size bias. *Neuroimage*. 2011; 55:1548–65. [PubMed: 21276857]
30. Stam CJ, van Straaten EC. Go with the flow: use of a directed phase lag index (dPLI) to characterize patterns of phase relations in a large-scale model of brain dynamics. *Neuroimage*. 2012; 62:1415–28. [PubMed: 22634858]
31. Oostenveld R, Fries P, Maris E, Schoffelen JM. FieldTrip: Open source software for advanced analysis of MEG, EEG, and invasive electrophysiological data. *Comput Intell Neurosci*. 2011; 2011:156869. [PubMed: 21253357]
32. Papan A, Kugiumtzis D, Larsson PG. Reducing the bias of causality measures. *Phys Rev E Stat Nonlin Soft Matter Phys*. 2011; 83:036207. [PubMed: 21517575]
33. Whitham EM, Pope KJ, Fitzgibbon SP, Lewis T, Clark CR, Loveless S, Broberg M, Wallace A, DeLosAngeles D, Lillie P, Hardy A, Fronsco R, Pulbrook A, Willoughby JO. Scalp electrical recording during paralysis: quantitative evidence that EEG frequencies above 20 Hz are contaminated by EMG. *Clin Neurophysiol*. 2007; 118:1877–88. [PubMed: 17574912]
34. Whitham EM, Lewis T, Pope KJ, Fitzgibbon SP, Clark CR, Loveless S, DeLosAngeles D, Wallace AK, Broberg M, Willoughby JO. Thinking activates EMG in scalp electrical recordings. *Clin Neurophysiol*. 2008; 119:1166–75. [PubMed: 18329954]

35. Muthukumaraswamy SD, Shaw AD, Jackson LE, Hall J, Moran R, Saxena N. Evidence that subanesthetic doses of ketamine cause sustained disruptions of NMDA and AMPA-mediated frontoparietal connectivity in humans. *J Neurosci*. 2015; 35:11694–706. [PubMed: 26290246]
36. Rivolta D, Heidegger T, Scheller B, Sauer A, Schaum M, Birkner K, Singer W, Wibral M, Uhlhaas PJ. Ketamine dysregulates the amplitude and connectivity of high-frequency oscillations in cortical-subcortical networks in humans: evidence from resting-state magnetoencephalography-recordings. *Schizophr Bull*. 2015; 41:1105–14. [PubMed: 25987642]
37. Domino EF, Chodoff P, Corssen G. Pharmacologic effects of CI-581, a new dissociative anesthetic, in man. *Clin Pharmacol Ther*. 1965; 6:279–91. [PubMed: 14296024]
38. Schuttler J, Stanski DR, White PF, Trevor AJ, Horai Y, Verotta D, Sheiner LB. Pharmacodynamic modeling of the EEG effects of ketamine and its enantiomers in man. *J Pharmacokinet Biopharm*. 1987; 15:241–53. [PubMed: 3668802]
39. Yamakage M, Hirshman CA, Croxton TL. Inhibitory effects of thiopental, ketamine, and propofol on voltage-dependent Ca²⁺ channels in porcine tracheal smooth muscle cells. *Anesthesiology*. 1995; 83:1274–82. [PubMed: 8533920]
40. Hayashi Y, Kawaji K, Sun L, Zhang X, Koyano K, Yokoyama T, Kohsaka S, Inoue K, Nakanishi H. Microglial Ca(2+)-activated K(+) channels are possible molecular targets for the analgesic effects of S-ketamine on neuropathic pain. *J Neurosci*. 2011; 31:17370–82. [PubMed: 22131399]
41. Yamakura T, Chavez-Noriega LE, Harris RA. Subunit-dependent inhibition of human neuronal nicotinic acetylcholine receptors and other ligand-gated ion channels by dissociative anesthetics ketamine and dizocilpine. *Anesthesiology*. 2000; 92:1144–53. [PubMed: 10754635]
42. Cai YC, Ma L, Fan GH, Zhao J, Jiang LZ, Pei G. Activation of N-methyl-D-aspartate receptor attenuates acute responsiveness of delta-opioid receptors. *Mol Pharmacol*. 1997; 51:583–7. [PubMed: 9106622]
43. Hillebrand A, Tewarie P, van Dellen E, Yu M, Carbo EW, Douw L, Gouw AA, van Straaten EC, Stam CJ. Direction of information flow in large-scale resting-state networks is frequency-dependent. *Proc Natl Acad Sci U S A*. 2016; 113:3867–72. [PubMed: 27001844]
44. Meier J, Zhou X, Hillebrand A, Tewarie P, Stam CJ, Miegheem PV. The epidemic spreading model and the direction of information flow in brain networks. *Neuroimage*. 2017; 152:639–46. [PubMed: 28179163]
45. Moon JY, Lee U, Blain-Moraes S, Mashour GA. General relationship of global topology, local dynamics, and directionality in large-scale brain networks. *PLoS Comput Biol*. 2015; 11:e1004225. [PubMed: 25874700]
46. Singh JB, Fedgchin M, Daly EJ, De Boer P, Cooper K, Lim P, Pinter C, Murrrough JW, Sanacora G, Shelton RC, Kurian B, Winokur A, Fava M, Manji H, Drevets WC, Van Nueten L. A double-blind, randomized, placebo-controlled, dose-frequency study of intravenous ketamine in patients with treatment-resistant depression. *Am J Psychiatry*. 2016; 173:816–26. [PubMed: 27056608]
47. Newton A, Fitton L. Intravenous ketamine for adult procedural sedation in the emergency department: a prospective cohort study. *Emerg Med J*. 2008; 25:498–501. [PubMed: 18660398]
48. Imas OA, Ropella KM, Ward BD, Wood JD, Hudetz AG. Volatile anesthetics disrupt frontal-posterior recurrent information transfer at gamma frequencies in rat. *Neurosci Lett*. 2005; 387:145–50. [PubMed: 16019145]
49. Boly M, Moran R, Murphy M, Boveroux P, Bruno MA, Noirhomme Q, Ledoux D, Bonhomme V, Bricchant JF, Tononi G, Laureys S, Friston K. Connectivity changes underlying spectral EEG changes during propofol-induced loss of consciousness. *J Neurosci*. 2012; 32:7082–90. [PubMed: 22593076]
50. Untergerher G, Jordan D, Kochs EF, Ilg R, Schneider G. Fronto-parietal connectivity is a non-static phenomenon with characteristic changes during unconsciousness. *PLoS One*. 2014; 9:e87498. [PubMed: 24475298]
51. Waterman D, Woestenburg JC, Elton M, Hofman W, Kok A. Removal of ocular artifacts from the REM sleep EEG. *Sleep*. 1992; 15:371–5. [PubMed: 1519014]
52. Schroeder KE, Irwin ZT, Gaidica M, Bentley JN, Patil PG, Mashour GA, Chestek CA. Disruption of corticocortical information transfer during ketamine anesthesia in the primate brain. *Neuroimage*. 2016; 134:459–65. [PubMed: 27095309]

53. Lau TM, Gwin JT, McDowell KG, Ferris DP. Weighted phase lag index stability as an artifact resistant measure to detect cognitive EEG activity during locomotion. *J Neuroeng Rehabil.* 2012; 9:47. [PubMed: 22828128]

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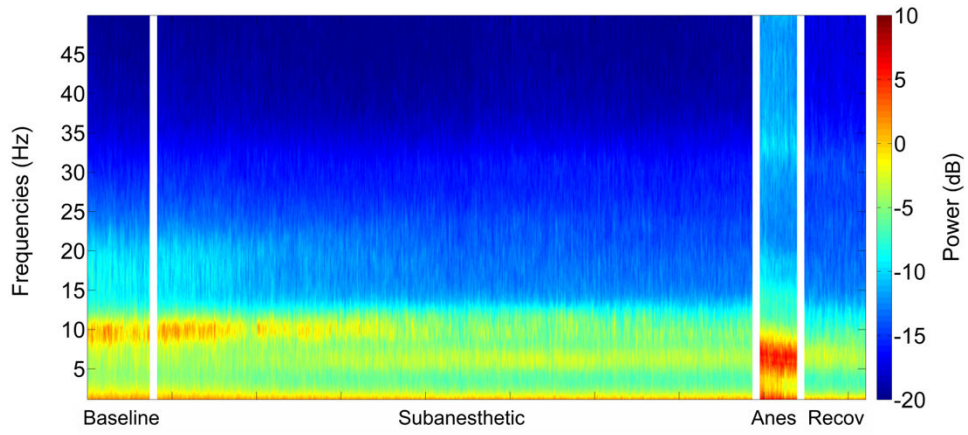


Fig. 1. Group-level absolute power spectrogram (.05 to 50 Hz bandpass filter). There is a marked increase in theta bandwidth power during ketamine anesthesia. Anes = anesthetic period, Recov = recovery period, dB = decibels.

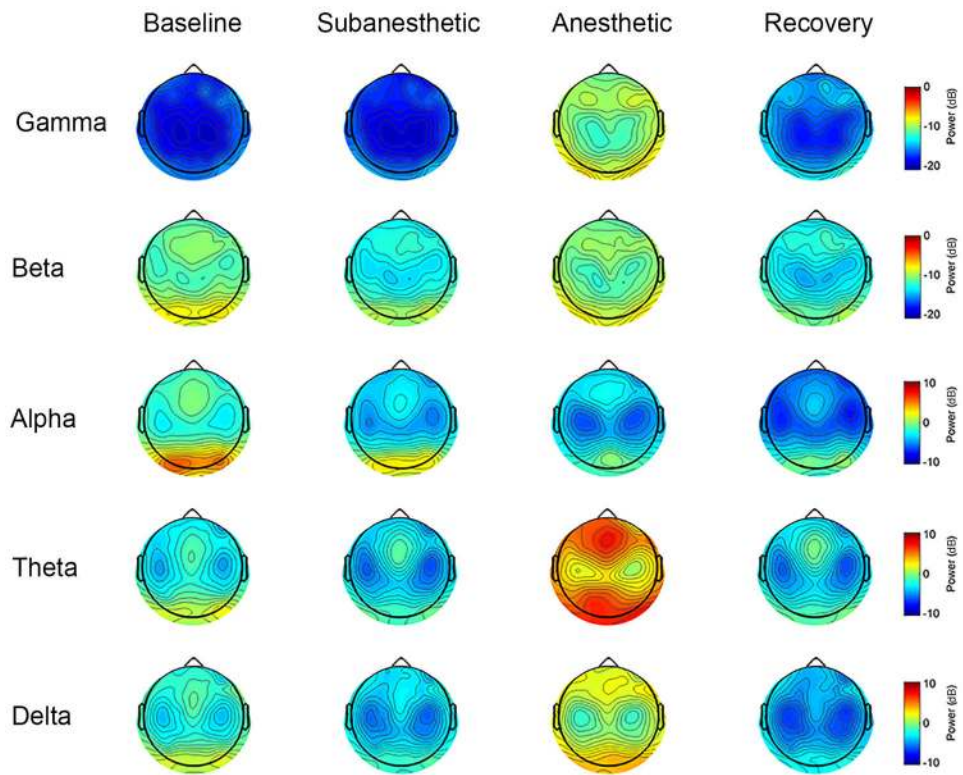


Fig. 2. Topomaps from retained trials for each condition. During ketamine anesthesia, theta power increases in frontal and posterior channel clusters. Posterior alpha power decreases sequentially during subanesthetic and anesthetic ketamine dosing, and no anteriorization of alpha power is noted during anesthetic dosing [delta (1-4 Hz), theta (4-8 Hz), alpha 1 (8-13 Hz), beta (13-30 Hz), gamma (30-48 Hz)].

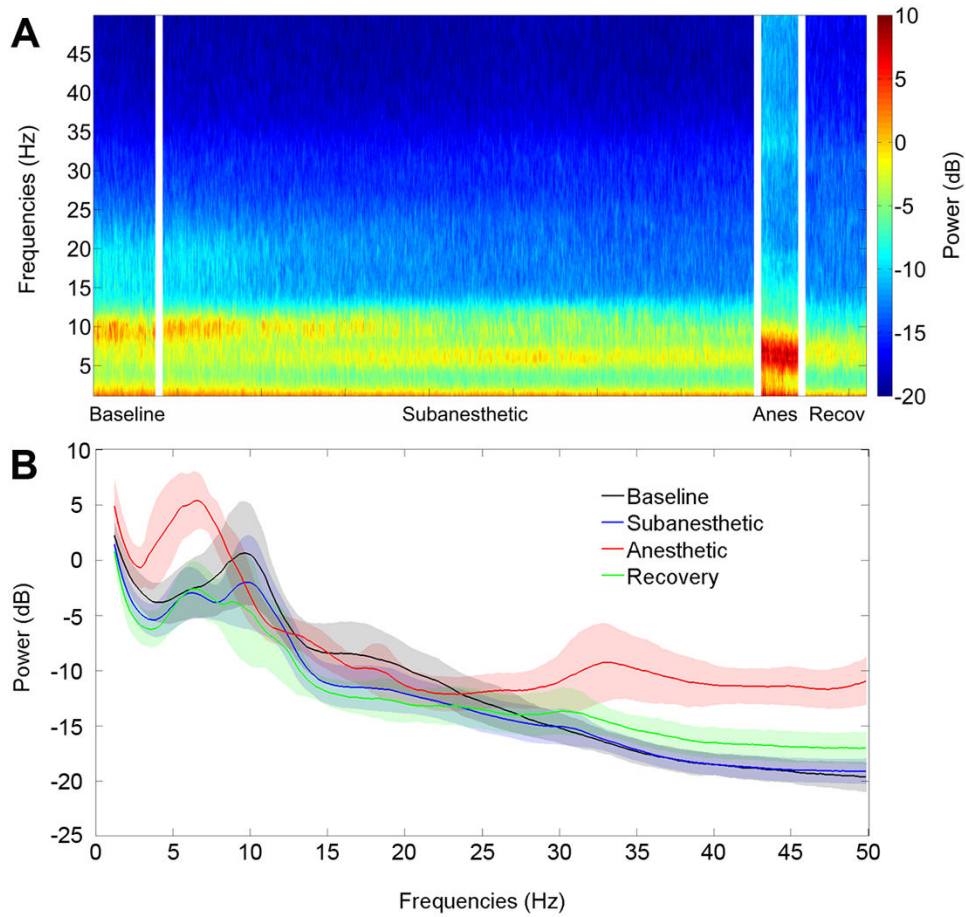


Fig. 3. Frontal channel, group-level spectrogram and line graph (see figure, Supplemental Digital Content 1, for electrode placement location). (A) Spectrogram depicts marked increase in frontal channel theta power during the anesthetic period. Increased gamma bandwidth power compared to baseline is noted as well. (B) Line graph demonstrates a shift to theta bandwidth power during ketamine anesthesia. Shaded regions represent ± 1 SD. Anes = anesthetic period, Recov = recovery period, dB = decibels.

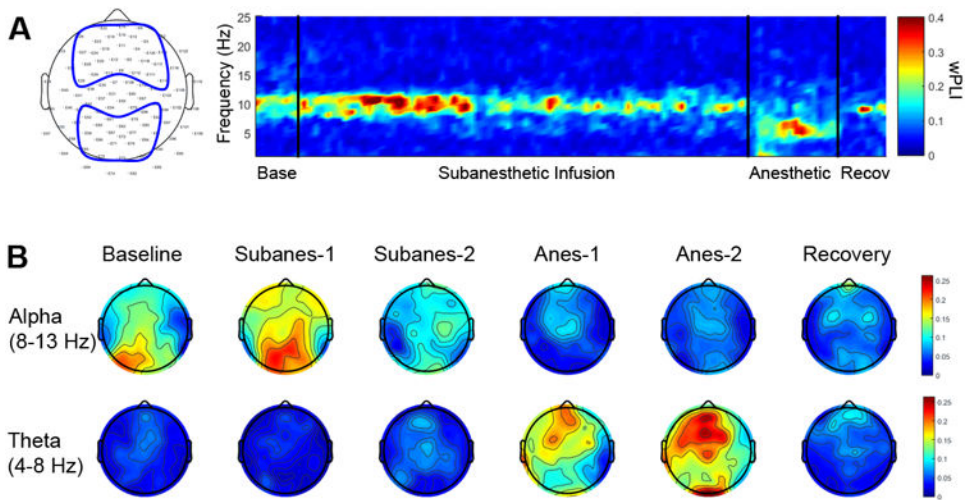


Fig. 4. Weighted functional connectivity changes, as assessed by weighted Phase Lag Index (wPLI). (A) Group level connectogram of the mean wPLI between anterior and posterior regions. The vertical lines in black separate the waking, subanesthetic, anesthetic, and recovery periods. The time periods during subanesthetic and anesthetic recordings were rescaled – horizontal axis indicates number of epochs (1-min long with 50% overlapping). (B) Scalp topography of the mean wPLI at alpha (8-13 Hz) and theta (4-8 Hz) for the six periods studied. Base = baseline period, Recov = recovery period, wPLI = weighted Phase Lag Index, Subanes-1 = first subanesthetic period examined, Subanes-2 = second subanesthetic period examined, Anes-1 = first anesthetic period examined, Anes-2 = second anesthetic period examined.

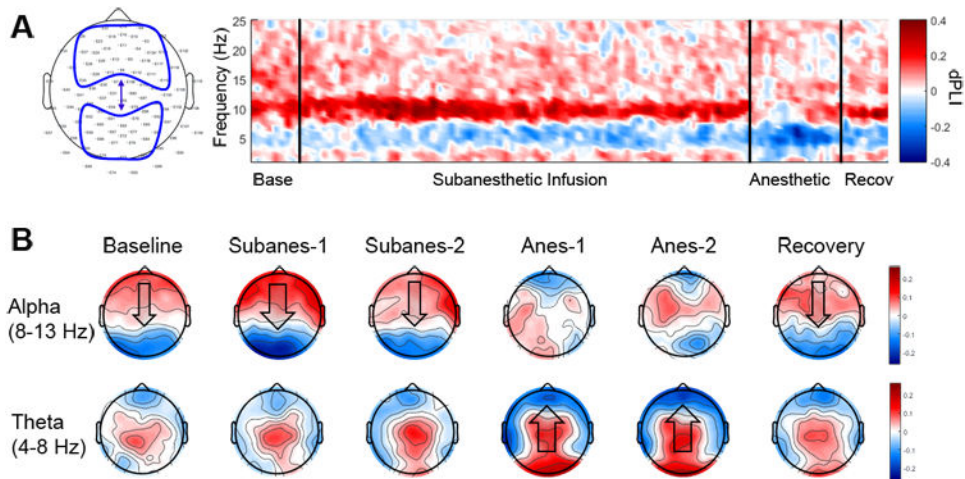


Fig. 5. Directional functional connectivity changes, as assessed by directed Phase Lag Index (dPLI), across all conditions in the alpha and theta bandwidths. (A) Group-level connectogram of the mean dPLI between anterior and posterior regions. The vertical lines in black separate the baseline, subanesthetic, anesthetic, and recovery periods. Time on the horizontal axis was rescaled; epochs are 1-minute long with 50% overlap. (B) Scalp topography of the mean dPLI (in each channel and with all other channels) at alpha and theta for the six studied conditions. Base = baseline period, Recov = recovery period, Subanes-1 = first subanesthetic period examined, Subanes-2 = second subanesthetic period examined, Anes-1 = first anesthetic period examined, Anes-2 = second anesthetic period examined.

Table 1
Mean Power Values Across Ketamine Doses and Electroencephalogram Bandwidths – Frontal Channels

Bandwidth	Baseline	Subanesthetic Dose	Anesthetic Dose	Recovery
Gamma (>30 Hz) <i>mean power, dB (\pmSD)</i>	0.02 (\pm 0.00)	0.02 (\pm 0.00)	0.09 (\pm 0.06)	0.03 (\pm 0.01)
Beta (13-30 Hz) <i>mean power, dB (\pmSD)</i>	0.09 (\pm 0.04)	0.06 (\pm 0.02)	0.08 (\pm 0.03)	0.06 (\pm 0.02)
Alpha (8-13 Hz) <i>mean power, dB (\pmSD)</i>	0.88 (\pm 0.80)	0.46 (\pm 0.34)	0.49 (\pm 0.22)	0.34 (\pm 0.38)
Theta (4-8 Hz) <i>mean power, dB (\pmSD)</i>	0.64 (\pm 0.28)	0.60 (\pm 0.30)	4.25 (\pm 1.90)	0.68 (\pm 0.41)
Delta (1-4 Hz) <i>mean power, dB (\pmSD)</i>	0.72 (\pm 0.23)	0.46 (\pm 0.13)	1.39 (\pm 0.70)	0.36 (\pm 0.12)

Absolute power values across conditions and frequency bandwidths in frontal channels. Linear mixed modeling (LMM) analysis used for statistical comparisons, $P < 0.001$ for all bandwidths except Alpha ($P < 0.05$); dB = decibels, SD = standard deviation.

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Table 2
Related Human Studies Examining the Effects of Ketamine on EEG and MEG Recordings

Study	High-Density Data Acquisition	Subanesthetic Dosing	Anesthetic Dosing	Recovery Period	Ketamine Only	Connectivity/Coherence Analysis
Domino et al. 1965 ³⁷		•	•	•	•	
Schuttler et al. 1987 ³⁸		•	•	•	•	
Kochs et al. 1996 ¹⁹		•	•	•	•	
Lee et al. 2013 ¹²			•		•	•
Blain-Moraes et al. 2014 ¹¹			•		•	•
Muthukumaraswamy et al. 2015 ³⁵	•	•		•	•	•
Rivolta et al. 2015 ³⁶	•	•			•	•
Akeju et al. 2016 ¹³			•			•
Vlisides et al. 2017	•	•	•	•	•	•

• = presence of the specified methodological consideration for each given study. EEG = electroencephalogram, MEG = magnetoencephalogram