

Neuroprotective effects of histone deacetylase inhibitors in brain ischemia

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Stroke resulting from cerebral ischemia or haemorrhage is a common cause of the death of neuronal cells and neurological dysfunction in humans. Finding therapeutics to improve outcome from brain ischemic injury has proved to be challenging. The efficacy of neuroprotective compounds identified in experimental brain ischemia models thus far have failed to successfully translate in clinical human trials. Recent experimental evidence indicates that inhibition of zinc-dependent histone deacetylases can protect neuronal and oligodendroglial cells from the damaging effects of ischemic insult, which may contribute to improved functional outcome. In this review we briefly highlight the current data supporting a beneficial role of histone deacetylation in experimental brain ischemia. We also discuss the molecular mechanism of neuroprotection.

Key words: brain ischemia, histone deacetylase inhibitors, neuroprotection, neuroregeneration

INTRODUCTION

Posttranslational (epigenetic) modifications of histones have profound effects on gene expression by modifying the accessibility of genes to the transcriptional machinery. One type of modification is a coordinated process carried out by two classes of enzyme - histone acetyltransferases (HATs) and histone deacetylases (HDACs). HDACs regulate gene expression by removing the acetyl groups of select lysine residues in the conserved tails of core histone proteins, to shift them to a positive charge, and enhancing the interaction with the negatively charged DNA. This is a reversible reaction, in balance with HATs adding acetyl groups. In general, increased histone acetylation provides a more open chromatin structure correlating with gene transcriptional activity, whereas deacetylation (decreased acetylation) is associated with repression of gene transcription (Strahl and Allis 2000, Saha and Pahan 2006, Hildman et al. 2007). Therefore, the balance of these two enzymes is a key element in the regulation of the expression of specific

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sets of genes. Disruption of the balance between HAT and HDAC activities leads to disequilibrium in acetylation which may have critical impact on cellular functioning (Struhl 1998).

A significant amount of data highlights the cellspecific aspect of HDAC inhibition, which seems to impair DNA repair in cancerous cells (Minucci and Pelicci 2006, Duvic and Vu 2007, Lawless et al. 2009, Kristensen et al. 2009), and displays strong protective properties in a broad range of neuropathologies. In agreement with this, treatment with various HDAC inhibitors has emerged as attractive therapeutic approach in in vitro and in vivo models of neurotoxicity and neurodegeneration as well as in acute brain ischemia (Ferrante et al. 2003, Kazantsev and Thompson 2008, Chuang et al. 2009, Langley et al. 2009, Du and Jiao 2011, Gräff et al. 2012). HDACi could possibly correct aberrant acetylation patterns and ameliorate disease state. In this review we discuss some recent data supporting the role for histone deacetylase inhibitors as treatment options for brain ischemia.

HISTONE DEACETYLASES

Histone deacetylases (HDACs) are evolutionary conservative enzymes involved in the epigenetic regula-

tions of gene expression and protein functions. To date, 11 potential HDACs have been identified in mammalian cells and all of them are expressed in adult rodent brain. They are organized into four different subclasses (I-IV) based on function and DNA sequence similarity (Thiagalingam et al. 2003). Generally speaking, class I HDACs display an extensive presence in nuclei. In addition to well-established nuclear localization, the expression of class I HDAC isoforms has also been observed in cytoplasmic domains (Baltan et al. 2011a). The class I HDACs (HDAC 1, 2, 3 and 8) are constitutively nuclear proteins, whereas class II enzymes, further divided into class IIa and IIb, comprised of HDACs 4, 5, 6, 7, 9, and 10, can shuttle between the nucleus and cytoplasm (Khochbin et al. 2001, Wang and Yang 2001, Xu et al. 2007). Class IV, currently consist of one member, HDAC11, with little known of its function. The enzymes belonging to class I, II and III are categorized as "zinc-dependent". The sirtuins, referred to as class III HDACs (Baur et al. 2012) are structurally and enzymatically distinct NAD-dependent enzymes. Sirtuins have been discussed in recent reviews (Baur et al. 2012, Houtkooper et al. 2012).

Although deacetylation of histones and compaction of chromatin is a common mechanism of Class I and II HDACs, some individual isoforms can also target many other non-histone protein, including transcription factors, nuclear hormone receptors, molecular chaperons, inflammation mediators, signal transduction mediators and cytoskeletal elements (Glozak et al. 2005, Spange et al. 2009, Ocker 2010).

Biological functions of individual HDACs have been difficult to define due to the lack of isoform-specific inhibitors. This obstacle is related to the high sequence homology within the catalytically active sites of HDACs (Bieliauskas and Pflum 2008). Nevertheless, the use of non-specific pharmacological inhibitors indicate the implication of histone deacetylases in the whole array of biological processes including, among others, regulation of developmental program, neurogenesis, apoptosis, synaptogenesis, neurite outgrowth (Bolger and Yao 2005, Minucci and Pelicci 2006, Akhtar et al. 2009, Brunmeir et al. 2009, Chuang et al. 2009, Liu et al. 2012a).

HISTONE DEACETYLASE INHIBITORS

Histone deacetylase inhibitors (HDACi) are a heterogeneous group of agents that inhibit HDACs and

promote posttranslational acetylation of lysine residues within nuclear and cytoplasmic proteins, which may alter their activity and function. In particular, HDACs inhibition can have a profound effect on the acetylation status of histone proteins within chromatin, resulting in the augmented expression of genes relevant to protection from an ischemic insult. However, inhibiting deacetylation not only affects chromatin structure. HDACis equally promote the acetylation of non-histone proteins which can determine the interactions, localization and stability of these proteins (Glozak et al. 2005).

All HDAC inhibitors function by chelating the zinc ion at the deacetylase active site. Structurally, HDAC inhibitors can be grouped into diverse classes: hydroxamic acid dervatives, aliphatic acids, benzamides, electrophilic ketanes, cyclic peptides, and a few substances not assignable to these groups (Miller et al. 2003, Langley et al. 2005, Marks and Breslow 2007, Xu et al. 2007, Murphy et al. 2014). These inhibitors exhibit some HDAC isoform selectivity. For example, selective inhibitors have been developed for HDAC6, however, in many cases these compounds exhibit a high degree of lipophilicity and are difficult to synthesize, rendering them more useful as tools for research (Dallavalle et al. 2012).

Classical, non-selective HDACis, such as the hydroxamic acid-based suberoylanilide hydroxamic acid (SAHA, vorinostat) and trichostatin A (TSA), a fungal antibiotic, inhibit most of class I, II and IV HDACs (Yoshida and Horinouchi 1999). The benzamide MS-275 and also a small chain fatty acid – valproic acid (VPA), have a narrower range of target. VPA inhibits selectively class I HDACs (Göttlicher et al. 2001, Phiel et al. 2001) and to a lesser extent class II, but not HDAC6 or 10 (Gurvich et al. 2004), while MS-275 are selective towards only a subset of class I HDACs (Khan et al. 2008, Murphy et al. 2014).

Other fatty acid derivatives, sodium butyrate (SB) and 4-phenyl butyrate (4PB) inhibit class I and II histone deacetalases. However, isoform-specific inhibition of HDACs still remains a challenging task (Khan et al. 2008, Thomas 2009).

It is very likely that the non-specificity of deacetylase inhibitors is responsible for the opposing effect noted in distinct type of cells. As is becoming apparent, HDACs inhibition appears to be mainly protective for neurons, and yet, deadly to cancerous cells. Similarly, the different effect of HDACi is also observed in cells that contribute to inflammatory pathways, where treatment results in pro- or anti-inflammatory stimulation (Halili et al. 2009). One of the explanations of such discrepancy involves the particular function of individual HDAC isoforms in activation of different transcription factors and then expression of different sets of genes. For instance, NFkB which is a target of HDACs, can trigger transcription survival as well as apoptotic gene products (Graham and Gibson 2005). It these circumstances only the synthesis of isoform specific inhibitors will make possible to decipher the effects of HDAC inhibition on gene transcription not only *via* increased histone acetylation but also on acetylation of non-histone proteins (Bieliauskas and Pflum 2008, Thomas 2009).

HDACI-INDUCED NEUROPROTECTION

Treatment with various HDAC inhibitors (HDACi) has emerged as an attractive therapeutic approach for acute injury and neurodegeneration in the last decade (Langley et al. 2008, 2009, Haberland et al. 2009, Uo et al. 2009, Selvi et al. 2010). The attractiveness of these agents increased by the finding that HDAC inhibitors can also enhance neuronal plasticity and memory and thereby may contribute to improved functional recovery (Hockly et al. 2003, Fischer et al. 2007, Vecsey et al. 2007). These properties bring about increased interest to use them as emerging tools for therapeutic interventions in the context of post-acute stroke. Since then, many HDACi have been tested in experimental stroke models. One of the first tested molecule was valproic acid (VPA) (Ren et al. 2004). It is worthy to note that VPA for the last 40 years has been commonly employed as both an antiepileptic and mood stabilizer (Emrich et al. 1980, Gurvich and Klein 2002). The efficacy of VPA in diverse form of epilepsy and bipolar disorders, as well as neuropathic pain (Covington 1998) suggests that VPA acts through multiple central nervous system targets. It occurred that in addition to the mention above functions, VPA administered after induction of focal ischemia resulted in a significant reduction of infarct volume and neurological deficit scores caused by middle cerebral artery occlusion (MCAO). The mechanism of neuroprotection implicated thus far include the elevation of histone 3 (H3) acetylation, increased expression of the prosurvival heat-shock protein HSP-70 and diminished activation of pro-death caspase 3 in injured brain

hemisphere. Of note, earlier study *in vitro* also clearly demonstrated that VPA can promote neuronal survival in culture subjected to glutamate-induced neurotoxicity (Hashimoto et al. 2002) or to oxygen-glucose deprivation injury (Reckling 2003).

During the last 10 years the considerable research activity has been focused on potential roles for a number of zinc-dependent HDAC inhibitors (such as VPA, TSA, SAHA, SB) used in different models of brain ischemia. The majority of studies employing focal ischemia, including transient (Ren et al. 2004, Yildrim et al. 2008, Wang et al. 2012) or permanent (Faraco et al. 2006, Kim et al. 2007, 2009, Langley et al. 2008, Liu et al. 2012b, Wang et al. 2012, Liesz et al. 2013) middle cerebral artery occlusion (MCAO).

Although permanent MCAO produces a more severe and rapid brain infarction, the basic effects of HDACi treatment presented by reduced brain infarction, suppression neuroinflammation in the ischemic region, and amelioration of neurological deficits do not depends on the severity of brain pathology. HDACi also attenuate MCAO-induced disruption of brain-blood barrier and decrease edema, at least partly by inhibiting NfκB activation as well MMP9 expression and activity (Wang et al. 2011). In addition, administration of HDAC inhibitors occurred to be effective whether given pre- or post-injury. Nevertheless, the highest beneficial effects were most evident when VPA or SB was administered at least 3 h after the ischemic onset. This time window, together with the long-lasting neurological improvement suggest that HDAC inhibition might have utility in treating acute stroke.

We have found only one published report describing the effect of HDAC inhibitor (VPA) in the experimental intracerebral haemorrhage (Sinn et al. 2007). The obtained results show suppression of hematoma expansion during the acute phase, probably by decreased levels of endogenous proteolytic enzymes – tPA and MMP9. The notable reduction of the hematoma which is an important factor of neurological deterioration (Brott et al. 1997) leads probably to the observed behavioral improvement.

All these findings show that targeting global HDAC activity is sufficient to protect neurons from ischemia-induced death (Drummond et al. 2005, Wang et al. 2012). In addition, relevant studies show that HDACi may induce sprouting of dendrites, an increase number of synapses and reinstate access to long term memories (Fischer et al. 2007).

It is clear that in addition to the damaging effect of ischemia in grey matter, an important component of stroke pathology is white matter injury. Compared to our understanding of the mechanism of ischemia-induced neuronal death and protection in grey matter, white matter pathophysiology remains relatively elusive, as does the development of potential protective agents for oligodendrocyte damage. Injury to white matter and simultaneous loss of oligodendrocytes is a key to the impaired brain function associated with a variety of pathological conditions, including stroke. Thus, it is logical to state that the efficient therapeutic compound should preserve both, grey and white matter. Recent in vitro and in vivo studies show that administration of HDACi (such as VPA, SB, TSA, SAHA and MS-375) after ischemic insult promotes functional recovery of axons, reduces loss of oligodendrocytes and preserves white matter architecture (Baltan et al. 2011b, 2013, Liu et al. 2012b, Kim and Chuang 2014, Murphy et al. 2014). This protective effects correlate with reduced excitotoxicity, maintenance of ATP, preservation of axonal mitochondria and oligodendrocytes during OGD deprivation in a pure white matter tract of optic nerve (Baltan et al. 2011b, 2013).

POTENTIAL MECHANISM OF PROTECTION

Potential explanation for the beneficial effects of HDACs inhibition following brain ischemia are multifold and due to the published results influence diverse array of targets to maintain neuronal function and survival and preserve white matter in response to injury. One of the well-described modes of protective action of HDACi after ischemia is associated with the reduction of neuroinflammation. It is increasingly recognized that cerebral inflammation mediated by activated microglia and infiltrating leukocytes, including monocytes/macrophages, plays a key role in focal ischemia-induced neurodegeneration by releasing proinflammatory factors (Zheng and Yenari 2004). In support of the crucial role of neuroinflammation in the white matter injury is that macrophages/microglia are located in the damaged axonal bundles after ischemic injury (Moxon-Emre and Schlichter 2010). It is postulated that HDACs inhibitor-mediated suppression of astrocytic and microglial activation, cytokine production and down regulating pro-inflammatory factors, is

likely to be important mechanism in decrease inflammation and secondary damage after stroke (Kim et al. 2007, Xu et al. 2007). An important mechanism of cerebroprotective effect induced by HDACi is the increase in interleukin-10 production, which restricted postischemic cerebral inflammatory gene up-regulation (Liesz et al. 2013). In addition, a promising therapeutic action of HDAC inhibitors may be also mediated by modifying the activity of other transcription factors such as erythroid 2-related factor. Nrf2 may represent one critical example of survival-promoting transcription factor (Martin-Montalvo et al. 2011). Activation of Nrf2 pathway has been shown to increase the resistance to oxidative stress and metabolic insult in culture, and to ischemic stroke *in vivo* (Son et al. 2010).

In general, acetylation of histone proteins with gene promoters and regulatory region, as well as transcription factors, can increase the expression of multiple genes which protein products contribute to neuroprotection, plasticity and memory (Fig. 1).

A significant amount of data show that the administration of HDACi after stroke was correlated with upregulation of heat-shock protein, HSP70, a probable viable target for neuroprotection (Ren et al. 2004, Faraco et al. 2006, Kim et al. 2007), in addition to being a molecular chaperone assisting in proper protein folding (Rajdev et al. 2000, Hoehn et al. 2001). Importantly, VPA-mediated neuroprotection against glutamate-induced cell death was lost if HSP70 induction was blocked. Anti-apoptotic effect of HSP70 may involve multiple mechanism, such as inhibition of cytochrom c dependent activation of death-promoting caspase 3 and its downstream effectors, suppression the activity of apoptosis inducing factor (AIF) (Pandey et al. 2000, Ravagnan et al. 2001), enhanced expression of antiapoptotic Bcl2 protein and suppression microglia/monocyte activation following experimental stroke (Yenari et al. 2005). Furthermore, inhibitors of HDACs may compensate MCAO-induced deficiency of pro-survival phospho-Akt and phospho-ERK in the ischemic hemisphere and raise significantly the level of brain-derived neurotrophic factor (BDNF) and glial-derived neurotrophic factor (GDNF) expression. Of note, increased expression of neurotrophins by HDACi treatment were found to be involved in the postischemic neuroprotection and neuronal restorative effects (Fukumoto et al. 2001, Chen et al. 2006, Hasan et al. 2013). Another target regulated positively by HDACi after ischemia is gelsolin, a protein involved in actin filament organization and

by this route contributed to neuroprotection from ischemic injury (Yidrim et al. 2008).

Inhibition of HDACs enhances resistance to death of neurons not only by positive regulation of pro-survival agents but also by blocking the action of transcription factors that regulate apoptosis. The tumor suppressor, p53, has been implicated as a key regulator of apoptosis in acute and chronic neurologic insults (Morrison et al. 2003). In line with this, several studies reported downregulation of p53 in injured ischemic hemisphere. In addition, inhibitors of class I/II HDACs protected cultured cortical neurons from p53-dependent cell death by suppression of p53-dependent expression of PUMA, a critical signaling intermediate linking p53 to Bax activation. Thereby, HDAC inhibitors block Bax-dependent cell death (Uo et al. 2009, Brochier et al. 2013). However, MS 275, in contrast to other HDAC inhibitors, does not directly affect p53 acetylation, or its stability and activity. If the target for MS-275 is not p53, the drug could modulate neuronal survival by modifying the activity of other transcription factors (Murphy et al. 2014).

In addition to transcription events, non-transcriptional events may also play a role in protective action of HDACi. For example, the selective inhibition of HDAC6 increases alfa-tubulin acetylation which may underlie neurite outgrowth without altering H4 acetylation and increase the vesicular transport. It is suggested that this effect might be independent on transcription (Riveccio et al. 2009).

Taken altogether the above findings allow us to state that irrespective of the precise mechanisms involved, HDACi have potential in preventing the consequences of acute ischemic injury.

HDACI-INDUCED REGENERATION

It is clear that successful stroke treatment also need to promote plasticity and repair mechanism(s) in the post-acute phase. The reduced infarct volume and

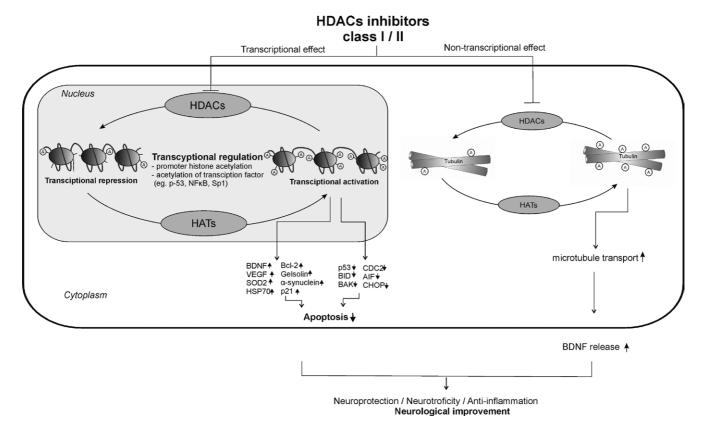


Fig.1. The cellular effects of HDACs inhibitors after stroke. Inhibition of HDAC activity has transcriptional and non-transcriptional effects. Acetylation of histone proteins within gene promoters as well as transcriptional factors can increase the expression of multiple genes which contribute to neuroprotection, plasticity and learning /memory. Acetylation of transcription factors may also decrease DNA binding and gene expression of some pro-death genes (e.g. p-53) and reduce apoptotic processes. Non-transcriptional effects of HDAC inhibitors also play critical role in neuroprotection and repair after stroke; hyperacetylation and stabilization of microtubule proteins increases vesicular transport and release of BDNF.

improved neurological performance may suggest the replacement of new neurons which contributes to the self-repair system of cerebral ischemic injury. A notable previous finding is that experimental stroke increased cell proliferation and neurogenesis in neurogenic areas - subventricular zone (SVZ) and dentate gyrus (DG) as well as in non-neurogenic regions (striatum and cerebral cortex) (Jin et al. 2001, Arvidsson et al. 2002, Parent et al. 2002, Zhang et al. 2004, Yamashita et al. 2006, Ziemka-Nałęcz and Zalewska 2012). Subsequently, a number of study reported that VPA and TSA induced neurite outgrowth and neurogenesis in vitro without an ischemic insult (Hao et al. 2004, Di Daniel et al. 2005, Yamauchi et al. 2007). Additionally, it was found that neural progenitor cells cultured in the presence of histone deacetylase inhibitors - VPA, TSA or SB, differentiated into neurons (Liu et al. 2012a), presumably by inducing the neurogenic transcription factor Neuro D (Hsieh et al. 2004). Interesting recent data indicate that VPA promotes neural differentiation via the increased expression of pro-neural genes Ngn1, Math1, and P15 associated with the increased acetylation of H4 (Yu et al. 2009).

Although there have been many in vivo and in vitro studies on the role of HDAC inhibitors as neuroprotective agents, very little work has been done to define their involvement in postischemic neuronal regeneration. First evidence showing stimulation of neurogenesis in the ischemic brain after administration of histone deacetylase inhibitors has been provided by Kim and coworkers (2009). Consistently, Liu and others (2012b) found that treatment with VPA enhances the number of neuroblasts in the SVZ as well as in ischemic boundary zone. It was also discovered that administration of SB or TSA, expanded the population of proliferating cells in the injured brain areas and restored the loss of neurons. The authors indicate that generation of new cells depends on the activation of BDNF-TrkB signaling and then activation of cAMPresponse element binding protein (CREB). CREB was identified by Chiaramello and colleagues (2007) as a part of molecular mechanism underlying BDNF-TrkB induced migration, differentiation and survival of SVZ neuroblasts. Moreover, its phosphorylated form appeared to be a prominent transcription factor involved in the expression of numerous neuroprotective, neurotrophic, and anti-inflammatory protein molecules (Chiu et al. 2013). The presented data are consistent

with findings demonstrating that HDACs inhibition by VPA and TSA leads to neuronal regeneration in cultures subjected to oxygen and glucose deprivation (OGD) and reoxygenation.

HDACi (TSA and SB) not only counteract ischemia-induced loss of oligodendrocytes in the injured hemisphere but also promote differentiation of oligodendroglial progenitors into mature cells. This process was associated with elevation of myelinated axonal density in the peri-infarct region (Liu et al. 2012b, Kim and Chuang 2014). The HDAC isoforms involved in the protective effects on oligodendrocytes after ischemic stroke remain unidentified. Paradoxically, it was reported that HDAC1 and HDAC2 activity are required to regulate oligodendrocyte differentiation (Ye et al. 2009), and that HDAC1 and HDA2 functions are critical for myelination and survival of Schwann cells in peripheral nervous system (Jacob et al. 2011). These results do not necessarily negate the importance of HDAC1 and HDAC2 inhibition in mediating the effects reported above, because of the different experimental conditions employed ischemic stroke versus normal, nonpathological conditions of the cited study. Future investigations exploring the role of specific isoforms involved in the pathophysiology of white matter injury and the beneficial effect of HDAC inhibitors will provide crucial information for therapeutic interventions. Although it is not at all clear if the notable amplifications of neurogenesis post-stroke contributes to subsequent restoration of function, the ability of HDACi to promote migration and neuronal differentiation could be exploited therapeutically (Kahle and Bix 2013).

Concurrently, VPA treatment enhanced postischemic angiogenesis by increasing microvessel density, facilitating endothelial cell proliferation, and up-regulating rate of cerebral blood flow (rCBF) in the ipsilateral cortex. This may contribute to the long-term functional outcome. These events may be associated with up-regulation of hypoxia inducible factor 1 alfa (HIF1alfa) and its downstream proangiogenic vascular endothelial growth factor (VEGF) as well as extracellular matrix metalloproteinases MMP2/9 (Sun et al. 2003, Shimotake et al. 2010, Wang et al. 2012). In addition, newly-generated vessels provide additional neurotrophic support to concurrent neurogenesis and synaptogenesis, and this ultimately may lead to functional recovery.

EFFECT OF HDACI IN NEONATAL HYPOXIA-ISCHEMIA

Despite ever growing information highlighting the beneficial role of HDACi after stroke in adult brains only a few available reports were addressed upon their effect in injured immature brains (Kabakus et al. 2005, Fleiss et al. 2012, George et al. 2013). These experiments were performed with the use of different experimental models, thus it is not possible to make the explicit conclusion related to the effect of HDACi. The data reported by Kabakus and coauthors (2005) showed only the tendency towards regression of cerebral infarct after VPA administration to 7 day-old pups subjected to hypoxia-ischemia (H-I). Nevertheless, the preliminary study performed by our group on the same model demonstrate beneficial effects SB expressed by reduced volume of brain lesion. Furthermore, SB treatment enhanced significantly the number of neuroblasts and progenitors of oligodendrocytes (Ziemka-Nalecz et al., unpublished data presented on 9th FENS Forum of Neuroscience).

Fleiss and colleagues (2012) examined the neuropathological and functional effects of the TSA in a model of neonatal lipopolysacharide-sensitized hypoxic-ischemic brain injury (LPS/HI) in mice. One striking feature of their results is that TSA diminished grey and white matter damage only in females. The neuroprotection was associated with increased acetylation of histone H4 and correlates with improved long-term learning. Interestingly, the beneficial effect of TSA was not connected neither with caspase-3 activation nor up-regulation of heat shock protein HSP70 and gelsolin, implicated in the neuroprotection observed in adult animals. Moreover, none of the inflammatory mechanisms assessed that are known to mediate neuroprotection by HDACi in adults correlated with improved outcome in TSA-treated neonatal females. TSA did not result in decrease the total number of microglia cells or NFkB-mediated reductions in cytokine expressions. TSA either did not impair oligodendrocyte maturation, which increases the possible clinical relevance of this strategy. Therefore it appears that TSA exerts neuroprotection via mechanism unique to neonates, yet unknown. This is consistent with the fact that many aspects of the evolving post-ischemic brain injury differ in the immature brain and thereby the efficacy of neuroprotectants can differ between adults and neonates (Cheng et al. 1997, Zhu et al.

2005). It is also possible that LPS sensitization may antagonize TSA efficacy in this model.

George and coworkers (2013) investigated long-term treatment of VPA and TSA upon stroke-injured immature mice (12 days old), using permanent ligation of the carotid artery. Presented data demonstrate that chronic, two weeks HDACi treatment did not modify the severity of brain atrophy assessed two months after the insult. In contrast to data reported by Fleiss and others (2012) the sex-related differences were not observed. Significant increase of neurogenesis in dentate gyrus with both TSA and VPA treatment were noted in the injured as well as uninjured animals but only at specific time point after administration of BrdU – marker of proliferation. It may mean that the high level of plasticity and neurogenesis cannot be up-regulated any further by manipulations with HDACi. In agreement with this statement, Foti and coauthors (2013) using neurosphere assays to determine neural stem cells (NSC) regulation by HDACi found that clinically used HDACi like VPA and TSA can perturb postnatal neurogenesis. It supports generally accepted view that HDAC activity is essential for oligodendrocyte differentiation in the developing rodent brain (Shen et al. 2005, Lyssiotis et al. 2007, Ye et al. 2009). Therefore, blockage of HDAC activity by inhibitors or ablation of HDAC1 and HDAC2 genes during the early postnatal stage, when myelination forms, prevents oligodendrocyte differentiation and leads to hypomyelination (Romm et al. 2005, Lyssiotis et al. 2007, Shen and Casacia-Bonnefil 2008, Ye et al. 2009). However, the effect of HDACi on suppression of oligodendrocytes differentiation is transient and only takes place during the first postnatal week (Shen et al. 2005). In addition, a temporary defined treatment regimen avoids this toxic effect (Langley et al. 2008).

The results obtained with HDAC inhibitors indicate that the functions of these agents in the nervous system injury are diverse and depend greatly on the developmental stage. For example, there is evidence that HDAC2 is critical for adult neurogenesis, but not required for embryonic neurogenesis (Jawerka et al. 2010). Which of HDAC among 11 members are targets for neuroprotection is unknown at present. Equally unknown is whether therapeutic efficacy can be obtained by targeting a single HDAC. *In vitro* studies showing that HDAC6-selective inhibition can protect against oxidative stress-induced neuronal death (Kozikowski et al. 2007) may provide some sugges-

tions that targeting a single HDAC isoform may indeed be neuroprotective in stroke. However, it should be also noted that SB, inhibitor of the class I HDAC, which does not inhibit HDAC6, occurred to be protective in the same *in vitro* oxidative stress model (Langley et al. 2008). Most current HDACs inhibitors are broad-spectrum inhibitors, and their use does not take into account the possible stage-and context-specific expression of different HDACs. Although considerable progress has been made in elucidating the effects of HDACi in brain ischemia, this area are still in early stage of discovery.

CONCLUSIONS

Based on the combined findings it may be hypothesized that HDACi provide a suitable option for brain in the clinical manifestations of stroke to facilitate successful translation of experimental ischemia research to clinical trials. However, considering the beneficial effects of HDACi in experimental stroke studies commonly use in rodents, it is worthy to underline that efficacy of drug in young, healthy, in majority male animals appears to be very poor predictor of clinical outcome. No studies assessed the neuroprotective effectiveness of HDAC inhibitors in "aged" animals, while the aged persons usually suffer from stroke.

In addition, not all currently available HDAC inhibitors are well tolerated when administered chronically. For instance, TSA have basal toxicity and prolonged treatment at high doses induces neuronal death in vitro and in vivo, so compromising their neuroprotective effect (Boutillier et al. 2002, Jeong et al. 2003). Also the other HDAC inhibitor, valproic acid, despite that is well tolerated in children and adolescents may have detrimental effects on postnatal neural development, which have not been fully explored. This agent is a potent teratogen in both humans and mouse models (Gurvich and Klein 2002). The teratogenic activity is associated with neural tube closure defects (Nau et al. 1991). It became clear that nontoxic analogues of these HDACi, targeting specific isoforms will help treat the injury of the nervous system.

Conclusion: The neuroprotective and regenerative actions of histone deacetylase inhibitors indicate that they may be considered as therapeutic agents after brain ischemia. However, new, nontoxic analogues targeting individual HDAC isoforms are still needed.

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REFERENCES

- Akhtar MW, Raingo J, Nelson ED, Montgomery RL, Olson EN, Kavalali ET, Monteggia LM (2009) Histone deacety-lases 1 and 2 form a developmental switch that controls excitatory synapse maturation and function. J Neurosci 29: 8288–8297.
- Arvidsson A, Collin T, Kirik D, Kokaia Z, Lindvall O (2002) Neuronal replacement from endogenous precursors in the adult brain after stroke. Nat Med 8: 963–970.
- Baltan S, Bachleda A, Morrison RS, Murphy SP (2011a) Expression of histone deacetylases in cellular compartments of the mouse brain and the effects of ischemia. Transl Stroke Res 2: 411–423.
- Baltan S, Murphy SP, Danilov CA, Bachleda A, Morrison RS (2011b) Histone deacetylase inhibitors preserve white matter structure and function during ischemia by conserving ATP and reducing excitotoxicity. J Neurosci 31: 3990–3999.
- Baltan S, Morrison RS, Murphy SP (2013) Novel protective effects of histone deacetylase inhibition on stroke and white matter ischemic injury. Neurotherapeutics 10: 798–807.
- Baur JA, Ungvari Z, Minor RK, Le Couteur DG, de Cabo R (2012) Are sirtuins viable targets for improving health-span and lifespan? Nat Rev Drug Discov 11: 443–461.
- Bieliauskas AV, Pflum MK (2008) Isoform-selective histone deacetylase inhibitors. Chem Soc Rev 37: 1402–1413.
- Bolger TA, Yao TP (2005) Intracellular trafficking of histone deacetylase 4 regulates neuronal cell death. J Neurosci 25: 9544–9553.
- Boutillier AL, Trinh E, Loeffler JP (2002) Constitutive repression of E2F1 transcriptional activity through HDAC proteins is essential for neuronal survival. Ann N Y Acad Sci 973: 438–442.
- Brochier C, Dennis G, Rivieccio MA, McLaughlin K, Coppola G, Ratan RR, Langley B (2013) Specific acetylation of p53 by HDAC inhibition prevents DNA damage-induced apoptosis in neurons. J Neurosci 33: 8621–8632.
- Brott T, Broderick J, Kothari R, Barsan W, Tomsick T, Sauerbeck L, Spilker J, Duldner J, Khoury J (1997) Early hemorrhage growth in patients with intracerebral hemorrhage. Stroke 28: 1–5.

- Brunmeir R, Lagger S, Seiser C (2009) Histone deacetylase HDAC1/HDAC2-controlled embryonic development and cell differentiation. Int J Dev Biol 53: 275–289.
- Chen PS, Peng GS, Li G, Yang S, Wu X, Wang CC, Wilson B, Lu RB, Gean PW, Chuang DM, Hong JS (2006) Valproate protects dopaminergic neurons in midbrain neuron/glia cultures by stimulating the release of neurotrophic factors from astrocytes. Mol Psychiatry 11: 1116–1125.
- Cheng Y, Gidday JM, Yan Q, Shah AR, Holtzman DM (1997) Marked age-dependent neuroprotection by brain-derived neurotrophic factor against neonatal hypoxic-ischemic brain injury. Ann Neurol 41: 521–529.
- Chiaramello S, Dalmasso G, Bezin L, Marcel D, Jourdan F, Peretto P, Fasolo A, De Marchis S (2007) BDNF/ TrkB interaction regulates migration of SVZ precursor cells via PI3-K and MAP-K signalling pathways. Eur J Neurosci 26: 1780–1790.
- Chiu SP, Wu MJ, Chen PY, Ho YR, Tai MH, Ho CT, Yen JH (2013) Neurotrophic action of 5-hydroxylated polymethoxyflavones: 5-demethylnobiletin and gardenin A stimulate neuritogenesis in PC12 cells. J Agric Food Chem 61: 9453–9463.
- Chuang DM, Leng Y, Marinova Z, Kim HJ, Chiu CT (2009) Multiple roles of HDAC inhibition in neurodegenerative conditions. Trends Neurosci 32: 591–601.
- Covington EC (1998) Anticonvulsants for neuropathic pain and detoxification. Cleve Clin J Med 65 Suppl 1: SI21–9; discussion SI45–7.
- Dallavalle S, Pisano C, Zunino F (2012) Development and therapeutic impact of HDAC6-selective inhibitors. Biochem Pharmacol 84: 756–765.
- Di Daniel E, Mudge AW, Maycox PR (2005) Comparative analysis of the effects of four mood stabilizers in SH-SY5Y cells and in primary neurons. Bipolar Disord 7: 33–41.
- Drummond DC, Noble CO, Kirpotin DB, Guo Z, Scott GK, Benz CC (2005) Clinical development of histone deacety-lase inhibitors as anticancer agents. Annu Rev Pharmacol Toxicol 45: 495–528.
- Du G, Jiao R (2011) To prevent neurodegeneration: HDAC6 uses different strategies for different challenges. Commun Integr Biol 4: 139–142.
- Duvic M, Vu J (2007) Update on the treatment of cutaneous T-cell lymphoma (CTCL): Focus on vorinostat. Biologics 1: 377–392.
- Emrich HM, von Zerssen D, Kissling W, Möller HJ, Windorfer A (1980) Effect of sodium valproate on mania. The GABA-hypothesis of affective disorders. Arch Psychiatr Nervenkr 229: 1–16.

- Faraco G, Pancani T, Formentini L, Mascagni P, Fossati G, Leoni F, Moroni F, Chiarugi A (2006) Pharmacological inhibition of histone deacetylases by suberoylanilide hydroxamic acid specifically alters gene expression and reduces ischemic injury in the mouse brain. Mol Pharmacol 70: 1876–1884.
- Ferrante RJ, Kubilus JK, Lee J, Ryu H, Beesen A, Zucker B, Smith K, Kowall NW, Ratan RR, Luthi-Carter R, Hersch SM (2003) Histone deacetylase inhibition by sodium butyrate chemotherapy ameliorates the neurodegenerative phenotype in Huntington's disease mice. J Neurosci 23: 9418–9427.
- Fischer A, Sananbenesi F, Wang X, Dobbin M, Tsai LH (2007) Recovery of learning and memory is associated with chromatin remodelling. Nature 447: 178–182.
- Fleiss B, Nilsson MK, Blomgren K, Mallard C (2012) Neuroprotection by the histone deacetylase inhibitor trichostatin A in a model of lipopolysaccharide-sensitised neonatal hypoxic-ischaemic brain injury. J Neuroinflammation 9: 70.
- Foti SB, Chou A, Moll AD, Roskams AJ (2013) HDAC inhibitors dysregulate neural stem cell activity in the postnatal mouse brain. Int J Dev Neurosci 31: 434–447.
- Fukumoto T, Morinobu S, Okamoto Y, Kagaya A, Yamawaki S (2001) Chronic lithium treatment increases the expression of brain-derived neurotrophic factor in the rat brain. Psychopharmacology (Berl) 158: 100–106.
- George S, Kadam SD, Irving ND, Markowitz GJ, Raja S, Kwan A, Tu Y, Chen H, Rohde C, Smith DR, Comi AM (2013) Impact of trichostatin A and sodium valproate treatment on post-stroke neurogenesis and behavioral outcomes in immature mice. Front Cell Neurosci 7: 123.
- Glozak MA, Sengupta N, Zhang X, Seto E (2005) Acetylation and deacetylation of non-histone proteins. Gene 363: 15–23.
- Göttlicher M, Minucci S, Zhu P, Krämer OH, Schimpf A, Giavara S, Sleeman JP, Lo Coco F, Nervi C, Pelicci PG, Heinzel T (2001) Valproic acid defines a novel class of HDAC inhibitors inducing differentiation of transformed cells. EMBO J 20: 6969–6978.
- Gräff J, Rei D, Guan JS, Wang WY, Seo J, Hennig KM, Nieland TJ, Fass DM, Kao PF, Kahn M, Su SC, Samiei A, Joseph N, Haggarty SJ, Delalle I, Tsai LH (2012) An epigenetic blockade of cognitive functions in the neurodegenerating brain. Nature 483: 222–226.
- Graham B, Gibson SB (2005) The two faces of NFkappaB in cell survival responses. Cell Cycle 4: 1342–1345.
- Gurvich N, Klein PS (2002) Lithium and valproic acid: parallels and contrasts in diverse signaling contexts. Pharmacol Ther 96: 45–66.

- Gurvich N, Tsygankova OM, Meinkoth JL, Klein PS (2004) Histone deacetylase is a target of valproic acid-mediated cellular differentiation. Cancer Res 64: 1079–1086.
- Haberland M, Montgomery RL, Olson EN (2009) The many roles of histone deacetylases in development and physiology: implications for disease and therapy. Nat Rev Genet 10: 32–42.
- Halili MA, Andrews MR, Sweet MJ, Fairlie DP (2009) Histone deacetylase inhibitors in inflammatory disease. Curr Top Med Chem 9: 309–319.
- Hao Y, Creson T, Zhang L, Li P, Du F, Yuan P, Gould TD, Manji HK, Chen G (2004) Mood stabilizer valproate promotes ERK pathway-dependent cortical neuronal growth and neurogenesis. J Neurosci 24: 6590–6599.
- Hasan MR, Kim JH, Kim YJ, Kwon KJ, Shin CY, Kim HY, Han SH, Choi DH, Lee J (2013) Effect of HDAC inhibitors on neuroprotection and neurite outgrowth in primary rat cortical neurons following ischemic insult. Neurochem Res 38: 1921–1934.
- Hashimoto R, Hough C, Nakazawa T, Yamamoto T, Chuang DM (2002) Lithium protection against glutamate excitotoxicity in rat cerebral cortical neurons: involvement of NMDA receptor inhibition possibly by decreasing NR2B tyrosine phosphorylation. J Neurochem 80: 589–597.
- Hildmann C, Riester D, Schwienhorst A (2007) Histone deacetylases an important class of cellular regulators with a variety of functions. Appl Microbiol Biotechnol 75: 487–497.
- Hockly E, Richon VM, Woodman B, Smith DL, Zhou X, Rosa E, Sathasivam K, Ghazi-Noori S, Mahal A, Lowden PA, Steffan JS, Marsh JL, Thompson LM, Lewis CM, Marks PA, Bates GP (2003) Suberoylanilide hydroxamic acid, a histone deacetylase inhibitor, ameliorates motor deficits in a mouse model of Huntington's disease. Proc Natl Acad Sci U S A 100: 2041–2046.
- Hoehn B, Ringer TM, Xu L, Giffard RG, Sapolsky RM, Steinberg GK, Yenari MA (2001) Overexpression of HSP72 after induction of experimental stroke protects neurons from ischemic damage. J Cereb Blood Flow Metab 21: 1303–1309.
- Houtkooper RH, Pirinen E, Auwerx J (2012) Sirtuins as regulators of metabolism and healthspan. Nat Rev Mol Cell Biol 13: 225–238.
- Hsieh J, Nakashima K, Kuwabara T, Mejia E, Gage FH (2004) Histone deacetylase inhibition-mediated neuronal differentiation of multipotent adult neural progenitor cells. Proc Natl Acad Sci U S A 101: 16659–16664.
- Jacob C, Christen CN, Pereira JA, Somandin C, Baggiolini A, Lötscher P, Ozçelik M, Tricaud N, Meijer D, Yamaguchi

- T, Matthias P, Suter U (2011) HDAC1 and HDAC2 control the transcriptional program of myelination and the survival of Schwann cells. Nat Neurosci 14: 429–436.
- Jawerka M, Colak D, Dimou L, Spiller C, Lagger S, Montgomery RL, Olson EN, Wurst W, Göttlicher M, Götz M (2010) The specific role of histone deacetylase 2 in adult neurogenesis. Neuron Glia Biol 6: 93–107.
- Jeong MR, Hashimoto R, Senatorov VV, Fujimaki K, Ren M, Lee MS, Chuang DM (2003) Valproic acid, a mood stabilizer and anticonvulsant, protects rat cerebral cortical neurons from spontaneous cell death: a role of histone deacetylase inhibition. FEBS Lett 542: 74–78.
- Jin K, Minami M, Lan JQ, Mao XO, Batteur S, Simon RP, Greenberg DA (2001) Neurogenesis in the dentate subgranular zone and rostral subventricular zone after focal cerebral ischemia in the rat. Proc Natl Acad Sci U S A 98: 4710–4715.
- Kabakus N, Ay I, Aysun S, Söylemezoglu F, Ozcan A, Celasun B (2005) Protective effects of valproic acid against hypoxic-ischemic brain injury in neonatal rats. J Child Neurol 20: 582–587.
- Kahle MP, Bix GJ (2013) Neuronal restoration following ischemic stroke: influences, barriers, and therapeutic potential. Neurorehabil Neural Repair 27: 469–478.
- Kazantsev AG, Thompson LM (2008) Therapeutic application of histone deacetylase inhibitors for central nervous system disorders. Nat Rev Drug Discov 7: 854–868.
- Khan N, Jeffers M, Kumar S, Hackett C, Boldog F, Khramtsov N, Qian X, Mills E, Berghs SC, Carey N, Finn PW, Collins LS, Tumber A, Ritchie JW, Jensen PB, Lichenstein HS, Sehested M (2008) Determination of the class and isoform selectivity of small-molecule histone deacetylase inhibitors. Biochem J 409: 581–589.
- Khochbin S, Verdel A, Lemercier C, Seigneurin-Berny D (2001) Functional significance of histone deacetylase diversity. Curr Opin Genet Dev 11: 162–166.
- Kim HJ, Chuang DM (2014) HDAC inhibitors mitigate ischemia-induced oligodendrocyte damage: potential roles of oligodendrogenesis, VEGF, and anti-inflammation. Am J Transl Res 6: 206–223.
- Kim HJ, Rowe M, Ren M, Hong JS, Chen PS, Chuang DM (2007) Histone deacetylase inhibitors exhibit anti-inflammatory and neuroprotective effects in a rat permanent ischemic model of stroke: multiple mechanisms of action. J Pharmacol Exp Ther 321: 892–901.
- Kim HJ, Leeds P, Chuang DM (2009) The HDAC inhibitor, sodium butyrate, stimulates neurogenesis in the ischemic brain. J Neurochem 110: 1226–1240.

- Kozikowski AP, Chen Y, Gaysin A, Chen B, D'Annibale MA, Suto CM, Langley BC (2007) Functional differences in epigenetic modulators-superiority of mercaptoacetamide-based histone deacetylase inhibitors relative to hydroxamates in cortical neuron neuroprotection studies. J Med Chem 50: 3054–3061.
- Kristensen LS, Nielsen HM, Hansen LL (2009) Epigenetics and cancer treatment. Eur J Pharmacol 625: 131–142.
- Langley B, Gensert JM, Beal MF, Ratan RR (2005) Remodeling chromatin and stress resistance in the central nervous system: histone deacetylase inhibitors as novel and broadly effective neuroprotective agents. Curr Drug Targets CNS Neurol Disord 4: 41–50.
- Langley B, D'Annibale MA, Suh K, Ayoub I, Tolhurst A, Bastan B, Yang L, Ko B, Fisher M, Cho S, Beal MF, Ratan RR (2008) Pulse inhibition of histone deacetylases induces complete resistance to oxidative death in cortical neurons without toxicity and reveals a role for cytoplasmic p21(waf1/cip1) in cell cycle-independent neuroprotection. J Neurosci 28: 163–176.
- Langley B, Brochier C, Rivieccio MA (2009) Targeting histone deacetylases as a multifaceted approach to treat the diverse outcomes of stroke. Stroke 40: 2899–2905.
- Lawless MW, Norris S, O'Byrne KJ, Gray SG (2009) Targeting histone deacetylases for the treatment of disease. J Cell Mol Med 13: 826–852.
- Liesz A, Zhou W, Na S-Y, Hammerling GJ, Garbi N, Karcher S, Mracsko E, Backs J, Rivest S, Veltkamp R (2013) Boosting Regulatory T Cells Limits Neuroinflammation in Permanent Cortical Stroke. J Neurosci 33: 17350–17362.
- Liu H, Wu H, Wang Y, Wang Y, Wu X, Ju S, Wang X (2012a) Inhibition of class II histone deacetylase blocks proliferation and promotes neuronal differentiation of the embryonic rat neural progenitor cells. Acta Neurobiol Exp (Wars) 72: 365–376.
- Liu XS, Chopp M, Kassis H, Jia LF, Hozeska-Solgot A, Zhang RL, Chen C, Cui YS, Zhang ZG (2012b) Valproic acid increases white matter repair and neurogenesis after stroke. Neuroscience 220: 313–321.
- Lyssiotis CA, Walker J, Wu C, Kondo T, Schultz PG, Wu X (2007) Inhibition of histone deacetylase activity induces developmental plasticity in oligodendrocyte precursor cells. Proc Natl Acad Sci U S A 104: 14982–14987.
- Marks PA, Breslow R (2007) Dimethyl sulfoxide to vorinostat: development of this histone deacetylase inhibitor as an anticancer drug. Nat Biotechnol 25: 84–90.
- Martín-Montalvo A, Villalba JM, Navas P, de Cabo R (2011) NRF2, cancer and calorie restriction. Oncogene 30: 505–520.

- Miller TA, Witter DJ, Belvedere S (2003). Histone deacety-lase inhibitors. J Med Chem 46: 5097–5116.
- Minucci S, Pelicci PG (2006) Histone deacetylase inhibitors and the promise of epigenetic (and more) treatments for cancer. Nat Rev Cancer 6: 38–51.
- Morrison RS, Kinoshita Y, Johnson MD, Guo W, Garden GA (2003) p53-dependent cell death signaling in neurons. Neurochem Res 28: 15–27.
- Moxon-Emre I, Schlichter LC (2010) Evolution of inflammation and white matter injury in a model of transient focal ischemia. J Neuropathol Exp Neurol 69: 1–15.
- Murphy SP, Lee RJ, McClean ME, Pemberton HE, Uo T, Morrison RS, Bastian C, Baltan S (2014) MS-275, a Class I histone deacetylase inhibitor, protects the p-53-deficient mouse against ischemic injury. J Neurochem 129: 509–515.
- Nau H, Hauck RS, Ehlers K (1991) Valproic acid-induced neural tube defects in mouse and human: aspects of chirality, alternative drug development, pharmacokinetics and possible mechanisms. Pharmacol Toxicol 69: 310–321.
- Ocker M (2010) Deacetylase inhibitors focus on non-histone targets and effects. World J Biol Chem 1: 55–61.
- Pandey P, Saleh A, Nakazawa A Kumar S, Srinivasula SM, Kumar V, Weichselbaum R, Nalin C, Alnemri ES, Kufe D, Kharbanda S (2000) Negative regulation of cytochrome c-mediated oligomerization of Apaf-1 and activation of procaspase-9 by heat shock protein 90. EMBO J 19: 4310–4322.
- Parent JM, Vexler ZS, Gong C, Derugin N, Ferriero DM (2002) Rat forebrain neurogenesis and striatal neuronal replacement after focal stroke. Ann Neurol 52: 802–813.
- Phiel CJ, Zhang F, Huang EY, Guenther MG, Lazar MA, Klein PS (2001) Histone deacetylase is a direct target of valproic acid, a potent anticonvulsant, mood stabilizer, and teratogen. J Biol Chem 276: 36734–36741.
- Rajdev S, Hara K, Kokubo Y, Mestril R, Dillmann W, Weinstein PR, Sharp FR (2000) Mice overexpressing rat heat shock protein 70 are protected against cerebral infarction. Ann Neurol 47: 782–791.
- Ravagnan L, Gurbuxani S, Susin SA, Maisse C, Daugas E, Zamzami N, Mak T, Jäättelä M, Penninger JM, Garrido C, Kroemer G (2001) Heat-shock protein 70 antagonizes apoptosis-inducing factor. Nat Cell Biol 3: 839–843.
- Reckling JC (2003) Neuroprotective effects of anticonvulsants in rat hippocampal slice culture exposed to oxygen/glucose deprivation. Neurosci Lett 335: 167–170.
- Ren M, Leng Y, Jeong M, Leeds PR, Chuang DM (2004) Valproic acid reduces brain damage induced by transient

- focal cerebral ischemia in rats: potential roles of histone deacetylase inhibition and heat shock protein induction. J Neurochem 89: 1358–1367.
- Rivieccio MA, Brochier C, Willis DE, Walker BA,
 D'Annibale MA, McLaughlin K, Siddiq A, Kozikowski
 AP, Jaffrey SR, Twiss JL, Ratan RR, Langley B (2009)
 HDAC6 is a target for protection and regeneration following injury in the nervous system. Proc Natl Acad Sci U S A 106: 19599–19604.
- Romm E, Nielsen JA, Kim JG, Hudson LD (2005) Myt1 family recruits histone deacetylase to regulate neural transcription. J Neurochem 93: 1444–1453.
- Saha RN, Pahan K (2006) HATs and HDACs in neurodegeneration: a tale of disconcerted acetylation homeostasis. Cell Death Differ 13: 539–550.
- Selvi BR, Cassel JC, Kundu TK, Boutillier AL (2010) Tuning acetylation levels with HAT activators: therapeutic strategy in neurodegenerative diseases. Biochim Biophys Acta 1799: 840–853.
- Shen S, Casaccia-Bonnefil P (2008) Post-translational modifications of nucleosomal histones in oligodendrocyte lineage cells in development and disease. J Mol Neurosci 35: 13–22.
- Shen S, Li J, Casaccia-Bonnefil P (2005) Histone modifications affect timing of oligodendrocyte progenitor differentiation in the developing rat brain. J Cell Biol 169: 577–589.
- Shimotake J, Derugin N, Wendland M, Vexler ZS, Ferriero DM (2010) Vascular endothelial growth factor receptor-2 inhibition promotes cell death and limits endothelial cell proliferation in a neonatal rodent model of stroke. Stroke 41: 343–349.
- Sinn DI, Kim SJ, Chu K, Jung KH, Lee ST, Song EC, Kim JM, Park DK, Kun Lee S, Kim M, Roh JK (2007) Valproic acid-mediated neuroprotection in intracerebral hemorrhage via histone deacetylase inhibition and transcriptional activation. Neurobiol Dis 26: 464–472.
- Son TG, Camandola S, Arumugam TV, Cutler RG, Telljohann RS, Mughal MR, Moore TA, Luo W, Yu QS, Johnson DA, Johnson JA, Greig NH, Mattson MP (2010) Plumbagin, a novel Nrf2/ARE activator, protects against cerebral ischemia. J Neurochem 112:1316–1326.
- Spange S, Wagner T, Heinzel T, Krämer OH (2009) Acetylation of non-histone proteins modulates cellular signalling at multiple levels. Int J Biochem Cell Biol 41: 185–198.
- Strahl BD, Allis CD (2000) The language of covalent histone modifications. Nature 403: 41–45.
- Struhl K (1998) Histone acetylation and transcriptional regulatory mechanisms. Genes Dev 12: 599–606

- Sun Y, Jin K, Xie L, Childs J, Mao XO, Logvinova A, Greenberg DA (2003) VEGF-induced neuroprotection, neurogenesis, and angiogenesis after focal cerebral ischemia. J Clin Invest 111: 1843–1851.
- Thiagalingam S, Cheng KH, Lee HJ, Mineva N, Thiagalingam A, Ponte JF (2003) Histone deacetylases: unique players in shaping the epigenetic histone code. Ann N Y Acad Sci 983: 84–100.
- Thomas EA (2009) Focal nature of neurological disorders necessitates isotype-selective histone deacetylase (HDAC) inhibitors. Mol Neurobiol 40: 33–45.
- Uo T, Veenstra TD, Morrison RS (2009) Histone deacetylase inhibitors prevent p53-dependent and p53-independent Bax-mediated neuronal apoptosis through two distinct mechanisms. J Neurosci 29: 2824–2832.
- Vecsey CG, Hawk JD, Lattal KM, Stein JM, Fabian SA, Attner MA, Cabrera SM, McDonough CB, Brindle PK, Abel T, Wood MA (2007) Histone deacetylase inhibitors enhance memory and synaptic plasticity via CREB:CBP-dependent transcriptional activation. J Neurosci 27: 6128–6140.
- Wang AH, Yang XJ (2001) Histone deacetylase 4 possesses intrinsic nuclear import and export signals. Mol Cell Biol 21: 5992–6005.
- Wang Z, Leng Y, Tsai LK, Leeds P, Chuang DM (2011) Valproic acid attenuates blood-brain barrier disruption in a rat model of transient focal cerebral ischemia: the roles of HDAC and MMP-9 inhibition. J Cereb Blood Flow Metab 31: 52–57.
- Wang Z, Tsai LK, Munasinghe J, Leng Y, Fessler EB, Chibane F, Leeds P, Chuang DM (2012) Chronic valproate treatment enhances postischemic angiogenesis and promotes functional recovery in a rat model of ischemic stroke. Stroke 43: 2430–2436.
- Xu WS, Parmigiani RB, Marks PA (2007) Histone deacety-lase inhibitors: molecular mechanisms of action. Oncogene 26: 5541–5552.
- Yamashita T, Ninomiya M, Hernández Acosta P, García-Verdugo JM, Sunabori T, Sakaguchi M, Adachi K, Kojima T, Hirota Y, Kawase T, Araki N, Abe K, Okano H, Sawamoto K (2006) Subventricular zone-derived neuroblasts migrate and differentiate into mature neurons in the post-stroke adult striatum. J Neurosci 26: 6627–6636.
- Yamauchi J, Miyamoto Y, Murabe M, Fujiwara Y, Sanbe A, Fujita Y, Murase S, Tanoue A (2007) Gadd45a, the gene induced by the mood stabilizer valproic acid, regulates neurite outgrowth through JNK and the substrate paxillin in N1E-115 neuroblastoma cells. Exp Cell Res 313: 1886–1896.

- Ye F, Chen Y, Hoang T, Montgomery RL, Zhao XH, Bu H, Hu T, Taketo MM, van Es JH, Clevers H, Hsieh J, Bassel-Duby R, Olson EN, Lu QR (2009) HDAC1 and HDAC2 regulate oligodendrocyte differentiation by disrupting the beta-catenin-TCF interaction. Nat Neurosci 12: 829–838.
- Yenari MA, Liu J, Zheng Z, Vexler ZS, Lee JE, Giffard RG (2005) Antiapoptotic and anti-inflammatory mechanisms of heat-shock protein protection. Ann N Y Acad Sci 1053: 74–83.
- Yildirim F, Gertz K, Kronenberg G, Harms C, Fink KB, Meisel A, Endres M (2008) Inhibition of histone deacetylation protects wildtype but not gelsolin-deficient mice from ischemic brain injury. Exp Neurol 210: 531–542.
- Yoshida M, Horinouchi S (1999) Trichostatin and leptomycin. Inhibition of histone deacetylation and signal-dependent nuclear export. Ann N Y Acad Sci 886: 23–36.
- Yu IT, Park JY, Kim SH, Lee JS, Kim YS, Son H (2009) Valproic acid promotes neuronal differentiation by induc-

- tion of proneural factors in association with H4 acetylation. Neuropharmacology 56: 473–480.
- Zhang R, Zhang Z, Wang L, Wang Y, Gousev A, Zhang L, Ho KL, Morshead C, Chopp M (2004) Activated neural stem cells contribute to stroke-induced neurogenesis and neuroblast migration toward the infarct boundary in adult rats. J Cereb Blood Flow Metab 24: 441–448.
- Zheng Z, Yenari MA (2004) Post-ischemic inflammation: molecular mechanisms and therapeutic implications. Neurol Res 26: 884–892.
- Zhu C, Wang X, Xu F, Bahr BA, Shibata M, Uchiyama Y, Hagberg H, Blomgren K (2005) The influence of age on apoptotic and other mechanisms of cell death after cerebral hypoxia-ischemia. Cell Death Differ 12: 162–176.
- Ziemka-Nałęcz M, Zalewska T (2012) Endogenous neurogenesis induced by ischemic brain injury or neurodegenerative diseases in adults. Acta Neurobiol Exp (Wars) 72: 309–324.